

RESEARCH ARTICLE

Lower Mutation Frequency of BCP/Precore Regions in e Antigen-Negative Chronic HBV-Infected Children instead of Adults Patients

Yong Huang^{1,2}, Haijun Deng¹, Xuefeng Shan³, Xuyang Gong¹, Xiaosong Li¹, Zen Tu¹, Quanxin Long^{1,4*}, Ailong Huang^{1,4*}

1 The Second Affiliated Hospital and the Key Laboratory of Molecular Biology of Infectious Diseases of the Chinese Ministry of Education, Chongqing Medical University, Yuzhong, Chongqing, China, **2** Department of Clinical Laboratory, Second Affiliated Hospital, Chongqing Medical University, Yuzhong, Chongqing, China, **3** The Department of Pharmacy, the First Affiliated Hospital, Chongqing Medical University, Yuzhong, Chongqing, China, **4** Collaborative Innovation Center for diagnosis and treatment of infectious diseases, Hangzhou, China

* longquanxin@gmail.com (QL); ahuang1964@163.com (AH)



OPEN ACCESS

Citation: Huang Y, Deng H, Shan X, Gong X, Li X, Tu Z, et al. (2015) Lower Mutation Frequency of BCP/Precore Regions in e Antigen-Negative Chronic HBV-Infected Children instead of Adults Patients. PLoS ONE 10(3): e0120733. doi:10.1371/journal.pone.0120733

Academic Editor: Isabelle A Chemin, CRCL-INSERM, FRANCE

Received: November 1, 2014

Accepted: February 6, 2015

Published: March 30, 2015

Copyright: © 2015 Huang et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: Accession numbers of BCP/precure regions sequences from child patients with CHB were from KP309449 to KP309693, and that of adult patients with CHB were from KP309694 to KP309785.

Funding: This study was funded in full by National Science and Technology Major Project (2013ZX10002002), Chongqing Science and Technology Commission (cstc2013jcyjC10002), and National Natural Science Foundation of China (81301394). The funders had no role in study design,

Abstract

To describe the Hepatitis B e antigen(HBeAg) seroconversion related mutation profiles of the basal core promoter(BCP)/precure regions in e antigen seroconverted child patients, a cohort of 245 child patients with CHB and a control patients group of 92 adult patients with CHB were recruited. The mutation frequencies of six nucleotides or nucleotide combinations including nucleotide (nt)1896, nt1762/1764, nt1752, nt1846, nt1899 and nt1753 showed significant differences between HBeAg positive and HBeAg-negative child patients groups. The frequencies of these HBeAg seroconversion-related mutations were significantly lower in HBeAg-negative children with CHB than in HBeAg-negative adults with CHB, especially for the mutation G1896A (41.1% vs 91.7%, $P<0.001$), and the average number of BCP/precure region mutations in samples from HBeAg-negative child patients was also obviously lower than in HBeAg-negative adult patients(3.62 ± 3.03 vs 4.89 ± 2.09 , $P<0.001$), suggesting less impact of mutations in the BCP/precure region on HBeAg seroconversion in child patients than adult patients.

Introduction

Chronic hepatitis B virus(HBV) infection(CHB) leads to a continuum of clinical outcomes, ranging from an asymptomatic carrier state to chronic active hepatitis, cirrhosis, and hepatocellular carcinoma[1]. Although the prevalence of HBV infection in children and adolescents decreased dramatically due to the universal HBV vaccination plan recommended by the World Health Organization(WHO) beginning in 1991[2, 3], more than 240 million people remain chronic liver infected, representing a serious global health problem[4].

The natural history of CHB can be divided into four phases: the immune tolerant phase, the immune active phase, the inactive phase and reactivation[5–7]. hepatitis B e antigen (HBeAg)

data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

seroconversion, defined as the loss of HBeAg followed by gain of anti-HBe antibodies, is an important hallmark of HBV natural infection history[8], suggesting the transition from an active phase to an inactive phase. Previous studies have demonstrated that HBeAg seroconversion in HBeAg-positive CHB patients is accompanied by improved clinical outcomes, and current clinical management guidelines have adopted HBeAg seroconversion as an appropriate treatment end point for CHB patients[9–11].

HBeAg seroconversion can be defined as a balance point in the battle between HBV and the host immune response. G1896A and the A1762T and G1764A double mutation are the most frequent isolated mutations in the precore and basal core promoter (BCP) regions of the HBV genome isolated from HBeAg-negative patients. The mutation G1896A creates a premature stop codon (28th codon) that prevents the production of HBeAg [12, 13], and the A1762T and G1764A double mutation in the BCP region can reduce the synthesis of HBeAg and enhance viral replication [14]. These mutation in BCP/precore regions were significantly associated with aggressive hepatitis and advanced liver disease[15].

Most HBeAg seroconversion factors were largely studied in adult patients[16, 17], the relationship between HBeAg seroconversion and BCP/precore region mutations in infants and children with CHB was less addressed[18, 19]. One previous study revealed that spontaneous HBeAg seroconversion usually occurred after puberty, with approximately 90% of children <15 years of age still HbeAg positive[20], suggesting possible difference between natural seroconversion rates in child and adult patients. From a population age perspective, sequences from HBeAg-negative child patients is much closer to seroconversion time point than that from HBeAg-negative adult patients, and sequences may more truly reflect HBeAg seroconversion-related feature. In this study, we performed a cross-sectional study to analyze the mutation profiles of the BCP/precore regions in a large number of well-defined children patients and adult control patients.

Materials and Methods

Subjects

A total of 337 serum samples were obtained from the Children's Hospital of Chongqing Medical University, the first affiliated hospital of Chongqing Medical University, the second affiliated hospital of Chongqing Medical University, Guangzhou Women and Children's Medical Center and Guangdong Women and Children's Hospital between June 2011 and September 2013 and stored at -70°C until further testing. In detail, all sera from HBeAg negative child CHB patients admitted between June 2011 and September 2013(110 samples) were collected, 90 of these samples were successfully amplified. Another 155 HBeAg positive child patients and 92 adult patients were recruit as controls.

All of the patients, including 245 children and 92 adults with chronic HBV infection, were positive for hepatitis B surface antigen (HBsAg) but negative for anti-HBs, anti-HCV(Hepatitis C virus), anti-HDV(Hepatitis D virus)and anti-HIV(Human Immunodeficiency Virus) antibodies. The Chongqing Medical University ethics committee approved the study, and written informed consent for participation in this study was obtained from all of the adult patients or the caretakers on behalf of the children enrolled in this study.

Serological Investigations and HBV DNA Quantification

HBsAg, anti-HBs, HBeAg, anti-HBe and anti-HBc levels were determined using enzyme-linked immunosorbent assay(ELISA, KeHua) or radioimmunoassay(RIA) kits(Roche). The liver function profile was determined using an auto-analyzer (Hitachi 7170, Tokyo, Japan). HBV viral load

was assessed by real-time polymerase chain reaction (PCR) using a fluorescence quantitative (FQ)-PCR kit for HBV (DaAn Gene Co., China) according to the manufacturer's instructions.

Amplification and Sequencing of BCP/precure regions of the HBV genome

Serum HBV DNA extraction, BCP/precure region(nt1653-1959) amplifications and sequencing followed previous descriptions[21]. Accession numbers of BCP/precure regions sequences from child patients with CHB were from KP309449 to KP309693, and that of adult patients with CHB were from KP309694 to KP309785.

Mutations and Genotype Analysis

Mutations of the BCP/precure regions were determined by aligning the sequences of the clinical samples with their corresponding consensus sequences for genotype B or genotype C. Consensus sequences for the different genotypes of HBV were calculated using the Basic Local Alignment Search Tool(BLAST) software with 2600 complete HBV genome sequence from GenBank. The complete S gene (nt65 to 870) of 337 samples was amplified by nested PCR as previously described[22] and then analyzed using the National Center for Biotechnology Information (NCBI) genotyping tool.

Statistical Analysis

The chi-squared test and Fisher's exact test were used to determine the differences between groups for categorical variables such as frequency of mutation. The Wilcoxon-Mann-Whitney test was used for continuous variables, such as alanine aminotransferase (ALT) level, DNA load, and the number of nucleotide mutations. Logistic linear regression analysis was used to assess the correlation between the number of nucleotide mutations and the HBeAg-negative subjects ratio. A P value of < 0.05 was considered statistically significant. All of the tests were analyzed using the Statistical Package for Social Science(SPSS) version 17.0 software.

Results

Clinical information of samples

A total of 245 children with CHB and 92 adults with CHB were enrolled in this cross-sectional study. The percentage of HBeAg-negative samples in the children and adult patients was 36.7% and 52.1%, respectively. The average age of the HBeAg seroconverted patients was older than the average age of the HBeAg-positive patients in both groups (7.93±3.82 vs 5.66±3.94 in children, 42.48±9.05 vs 35.05±8.10 in adults, P < 0.001 for both), suggesting that HBeAg seroconversion is positively correlated with age(Table 1). The serum HBV DNA titers of the HBeAg-

Table 1. The clinical information of chronic hepatitis B patients.

	HBeAg(+) Children N = 155	HBeAg(-) Children N = 90	P value	HBeAg(+) adult N = 44	HBeAg(-) adult N = 48	P value
Sex: Male/Female	92/63	57/33	0.632	31/13	35/13	0.976
Age (years)	5.66±3.94	7.93±3.82	<0.001	35.05±8.10	42.48±9.05	<0.001
ALT (IU/L)	73.27±148.88	60.17±111.63	0.436	82.9±86.94	81.3±98.2	0.834
HBV DNA(log10 copies/ml)	8.02±1.16	4.25±1.88	<0.001	8.04±0.95	5.99±1.49	<0.001
Genotype: B/C	116/39	75/15	0.166	27/3	25/6	0.504

doi:10.1371/journal.pone.0120733.t001

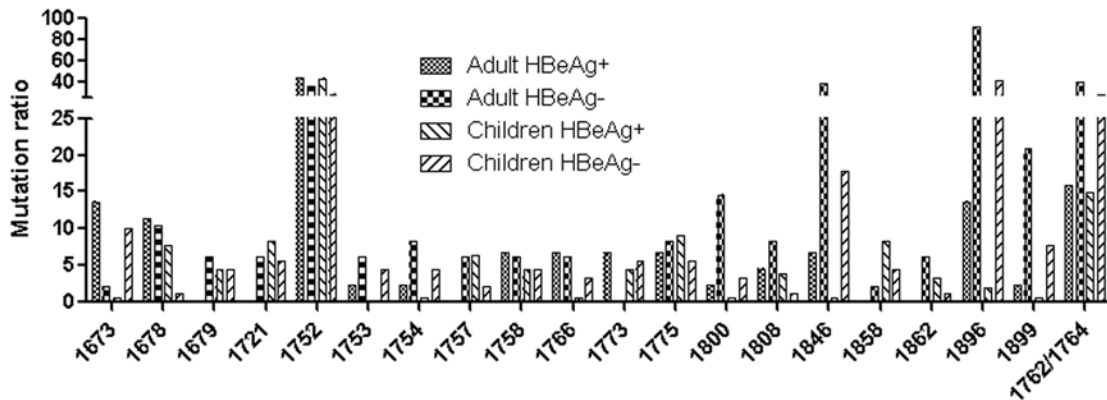


Fig 1. Mutation sites with >5% ratio in BCP/precure region.

doi:10.1371/journal.pone.0120733.g001

negative patients were obviously lower than that of antigen-positive patients. There were no significant differences in virus genotype, gender or ALT levels between HBeAg-negative and-positive child or adult patients.

Mutation sites correlated with HBeAg seroconversion

As shown in Fig 1, there were 20 nucleotides with mutation rates higher than 5%. Among them, six nucleotide or nucleotide combinations, including nt1752, nt1753, nt1762/1764, nt1846, nt1896, and nt1899, showed a significant difference between the HBeAg-positive and-negative child patient groups (Table 2). Moreover, only four sites or site combinations (nt1846, nt1896, nt1899 and nt1762/1764) showed a significant difference in the mutation ratio between the HBeAg-positive and-negative adult patient groups. Notably, the frequency of the G1896A mutation in the HBeAg-negative child patients was significantly lower than in the HBeAg-negative adult patients (41.1% vs 91.7%, $P < 0.001$), and similar results were also observed for other BCP/precure mutations such as A1846T (17.8% vs 37.5%, $P = 0.010$) and the G1899A substitution (7.8% vs 20.8%, $P = 0.026$). The mutation frequencies for nt1762/1764, nt1752 and nt1753 in HBeAg-negative child patients were also lower than in e negative-adult patients, although the difference lacked statistical significance.

Table 2. Mutation sites that correlated with the HBeAg status in children patients and adult patients.

Mutation sites	Children patients, n = 245		P_1 value	Adult patients, n = 92		P_2 value	P_3 value
	HBeAg +, n = 155	HBeAg-, n = 90		HBeAg +, n = 44	HBeAg-, n = 48		
1752	42.6%, 66/155	27.8%, 25/90	0.029	43.2%, 19/44	35.4%, 17/48	0.446	0.353
1753	0.0%, 0/155	4.4%, 4/90	0.034	2.3%, 1/44	6.3%, 3/48	0.342	0.958
1846	0.6%, 1/155	17.8%, 16/90	<0.001	6.8%, 3/44	37.5%, 18/48	<0.001	0.010
1896	1.9%, 3/155	41.1%, 37/90	<0.001	8.2%, 6/44	91.7%, 44/48	<0.001	<0.001
1899	0.6%, 1/155	7.8%, 7/90	0.008	2.3%, 1/44	20.8%, 10/48	0.006	0.026
1762/1764	14.8%, 23/155	26.7%, 24/90	0.023	15.9%, 7/44	39.5%, 19/48	0.012	0.119

P_1 represented the statistics difference of certain site mutation ratio between the HBeAg positive children patients and HBeAg negative children patients.

P_2 represented the statistics difference of certain site mutation ratio between HBeAg positive adult patient and HBeAg negative adult patients.

P_3 represented the statistics difference of certain site mutation ratio between HBeAg negative children patients and HBeAg negative adult patients.

doi:10.1371/journal.pone.0120733.t002

Table 3. The mutation profiles/ frequencies of HBeAg seroconversion related mutation nucleotides site in different HBV genotypes.

Mutation sites	Genotype B	Genotype C	Children patients		P_1 value	Adult patients		P_2 value
			B (n = 191)	C (n = 54)		B (n = 66)	C (n = 26)	
1752	A1752G	A1752G	46.1%	5.6%	0.000	53.0%	3.8%	0.000
1753	T1753V	T1753V	0.5%	5.6%	0.035	1.5%	11.5%	0.120
1846	A1846T	A1846T	7.9%	3.7%	0.449	23.4%	23.1%	0.971
1896	G1896A	G1896A	18.3%	9.3%	0.112	57.6%	46.2%	0.322
1899	G1899A	G1899A	2.6%	5.6%	0.523	10.6%	15.4%	0.780
1762/1764	A1762T/G1764A	A1762T/G1764A	15.7%	31.5%	0.009	24.2%	43.5%	0.081

P_1 represented the statistics difference of certain site mutation ratio between the genotype B and genotype C in children patients. P_2 represented the statistics difference of certain site mutation ratio between the genotype B and genotype C in adult patients.

doi:10.1371/journal.pone.0120733.t003

Several HBeAg seroconversion-related mutation sites were also significantly correlated with the HBV genotype (Table 3). The frequency of the A1752G mutation was higher in genotype B than genotype C HBV (46.1% vs 5.6% in children, 53.0% vs 3.8% in adults, $P < 0.001$ for both). In comparison, the frequencies of the A1762T/G1764A double mutation and T1753V mutation increased in genotype C infected children patients ($P < 0.05$).

Combined mutation profiles of BCP/precure regions in HBeAg seroconverted child and adult patients

There were 127 combined mutation types in the child and adult patients with CHB; among them, 10 mutation types showed a differential distribution between HBeAg-negative and HBeAg-positive samples (Table 4). The ratio of three combined mutations (G1896A/A1762T/G1764A, G1896A/A1846T and G1896A/A1752G) was obviously higher in HBeAg-negative patients than in the HBeAg-positive patient groups, suggesting that these combined mutation types could distinguish the HBeAg seroconverted patients with CHB. Notably, the higher

Table 4. Combined mutations profiles of BCP/precure in the HBeAg seroconversion children and adult patients.

combined mutation types	children patients		P_1 value	adult patients		P_2 value	P_3 value
	HBeAg+, n = 155	HBeAg-, n = 90		HBeAg+, n = 44	HBeAg-, n = 48		
1896/1846	0	12.2	0.000	4.5	35.4	0.000	0.001
1896/1899	0	4.6	0.017	0	18.8	0.008	0.015
1896/1762/1764	0.6	12.2	0.000	2.3	35.4	0.000	0.001
1896/1752	1.3	12.2	0.001	4.5	35.4	0.000	0.001
1899/1762/1764	0	5.9	0.006	0	12.5	0.045	0.269
1899/1896/1846	0	3.4	0.099	0	12.5	0.045	0.086
1899/1896/1762/1764	0	3.4	0.049	0	10.4	0.082	0.189
1752/1896/1762/1764	0.6	4.4	0.062	0	18.8	0.008	0.015
1762/1764/1846/1899/1896	0	2.3	0.134	0	6.3	0.272	0.467
1896/1846/1762/1764	0	2.3	0.134	2.3	14.6	0.085	0.015

Mutation ratio was the ratio of the number of specific linkage mutation type to the total sample number. P_1 value, P_1 represented the statistics difference of certain linkage mutation ratio between the HBeAg positive children patients and HBeAg negative children patients. P_2 represented the statistics difference of certain linkage mutation ratio between HBeAg positive adult patient and HBeAg negative adult patients. P_3 represented the statistics difference of certain linkage mutation ratio between HBeAg negative children patients and HBeAg negative adult patients.

doi:10.1371/journal.pone.0120733.t004

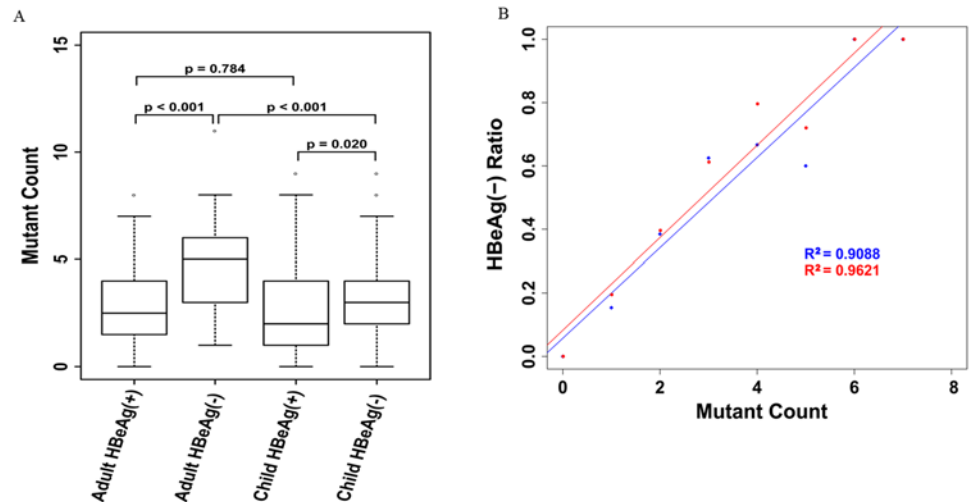


Fig 2. The relationship between number of mutation in BCP/precure region and e antigen seroconversion status. A. Average number of mutations nucleotides in BCP/precure region in children and adult patients with CHB, Average number of mutations in BCP/precure regions of HBeAg negative patients were obviously higher than in HBeAg positive patients either in adult and children patients group; Average number of mutations in BCP/precure regions of adult HBeAg negative patients was also statistical higher than that of children HBeAg negative patients. B. Correlations between the number of mutant sites in BCP/precure and HBeAg seroconversion. red line represented the correlation between BCP/precure mutation count and HBeAg negative ratio in children patients, $P < 0.001$; blue line represented the correlation between precure/core mutation count and HBeAg negative ratio in adult patients, $P < 0.001$.

doi:10.1371/journal.pone.0120733.g002

mutation ratio in the adult patients than in the child patients for these combined mutations (G1896A/A1846T: 35.4% vs 12.2%, $P < 0.001$; G1896A/G1899A: 18.8% vs 4.6%, $P = 0.015$; G1896A/A1762T/G1764A: 35.4% vs 12.2%, $P < 0.001$) implied that combined mutations in the BCP/precure regions were less correlated with HBeAg seroconversion in child patients than adult patients.

Correlation analysis of BCP/precure region mutation numbers and HBeAg serological status

In the present study, the average number of mutations in the BCP/precure regions of the HBeAg-negative subjects was significantly higher than in the HBeAg-positive subjects in both the child and adult patient groups (3.62 ± 3.03 vs 2.88 ± 1.94 , $P = 0.020$ in child patients; 4.89 ± 2.09 vs 2.91 ± 1.88 , $P < 0.001$ in adult patients) (Fig. 2a), directly proving that the number of mutations was positively related to HBeAg seroconversion. In addition, the average number of mutations in this region in HBeAg-negative child patients was also obviously lower than in HBeAg-negative adult patients (3.62 ± 3.03 vs 4.89 ± 2.09 , $P < 0.001$). Samples were also grouped according to the number of mutations in the BCP/precure region, and the HBeAg seroconversion ratio was calculated for each group. A significant positive correlation between the number of mutations and HBeAg seroconversion was observed in our samples ($R^2 = 0.9088$ in children, $R^2 = 0.9621$ in adults, $P < 0.05$ for both) (Fig. 2b), and the relevance level in adult patients was also higher than in children, confirming that the aforementioned BCP/precure mutations are less important for children who HBeAg seroconvert.

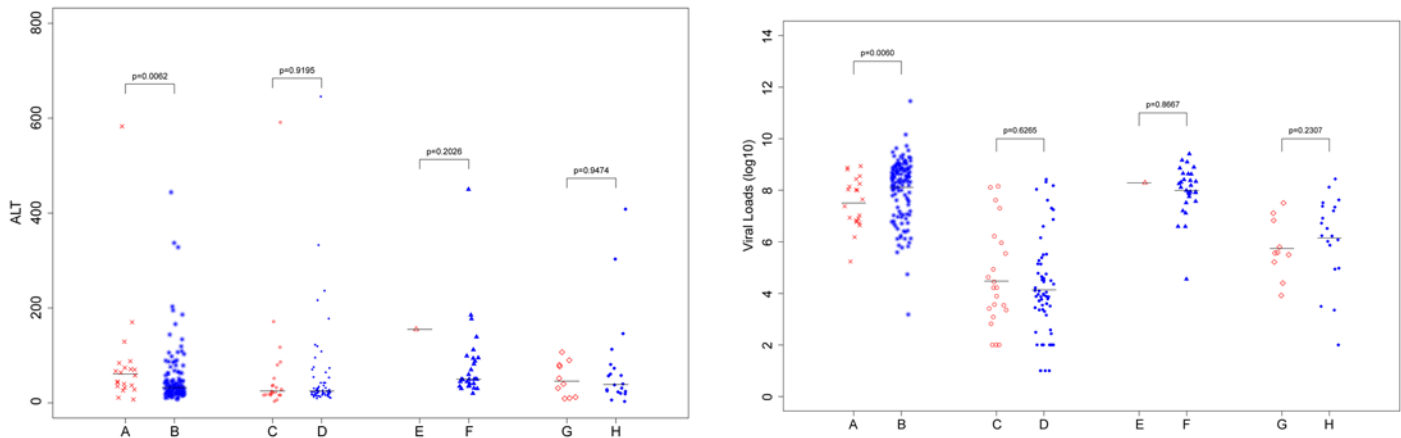


Fig 3. The association analysis between A1762T/G1764A double mutation and viral load, ALT level in different chronic HBV patients. A. children HBeAg positive patients containing A1762T/G1764A double mutation; B. children HBeAg positive patients not containing double mutation; C. children HBeAg negative patients containing double mutation; D. children HBeAg negative patients not containing double mutation; E. adult HBeAg positive patients containing double mutation; F. adult HBeAg positive patients not containing double mutation; G. adult HBeAg negative patients containing double mutation; H. adult HBeAg negative patients not containing double mutation.

doi:10.1371/journal.pone.0120733.g003

The association of HBV BCP/precure mutations with clinical presentation in chronic HBV patients

The association between 20 hotspot mutation sites and the patients' clinical presentations, including viral DNA titers, ALT levels and ages, was analyzed. Most mutations had no correlation with the clinical phenotypes, except the A1762T/G1764A double mutation, which was significantly correlated with a low viral DNA load and high ALT levels in HBeAg-positive child patients (viral load $\log_{10}8.10 \pm 1.17$ vs $\log_{10}7.51 \pm 0.97$, $P = 0.006$ and ALT 59.3 ± 114.7 vs 129.7 ± 242.4 , $P = 0.006$ for wildtype group vs double mutation group, respectively), but this correlation was not present in the HBeAg-negative child patient group (Fig. 3). In HBeAg-positive adult CHB patients, the A1762T/G1764A double mutation was significantly correlated with a low viral load ($\log_{10}7.39 \pm 1.36$ vs $\log_{10}6.15 \pm 1.59$, wild type group vs double mutation group, $P = 0.0077$) but not with ALT level (89.6 ± 101.8 vs 317.5 ± 229.8 , wild type group vs double mutation group, respectively, $P = 0.0681$), which differs from previous findings[23]. This difference is most likely due to the small sample size of our HBeAg-positive adult patients(Fig. 3).

Discussion

Two mutations in BCP/precure region were frequently associated with HBeAg seroconversion. One variation is a G-to-A mutation at nt1896, which creates a premature stop codon and then abolishes the synthesis of HBeAg[24]. The other is a two-nucleotide substitution, T1762A and G1764A[25], and transfection studies have shown that the T1762A and G1764A double mutation decreases the level of pre-C mRNA by 50% to 70%, leading to reduced HBeAg synthesis [26, 27]. In these six HBeAg seroconversion statistics-related sites, G1896A and the A1762T/G1764A double mutation had obviously higher mutation ratios in HBeAg-negative patients, which is in accordance with previous studies[12, 28]. In our study, the nt A1846T mutation ratio in HBeAg-negative child and adult patients was also significantly higher than in HBeAg-positive child and adult patients (17.8% vs 0.6% , $P < 0.01$, 37.5% vs 6.8% , $P < 0.01$ for children and adults, respectively), providing a clue that A1846T is closely associated with HBeAg seroconversion. The wild-type HBV sequence at position 1846 is T in most common genotypes, but it is A in the genotypes B and C that are prevalent in China(29). A1846T mutation is silent

at the amino acid level, and previous studies have observed that this mutation is associated with the progression of age-dependent disease (liver cirrhosis and hepatocellular carcinoma) as well as severe liver disease (fulminant hepatitis and acute-on-chronic liver failure, ACLF) [29, 30]; however, the relationship between this mutation and HBeAg seroconversion needs further experimental validation. Although nt1753 mutation has been reported as a rarity [31], significantly higher mutation ratio have also been observed in HBeAg-negative child patients. Combined mutations at nt1753, nt1762, nt1764, and nt1766 contribute to lower levels of HBeAg expression than double mutations at nt1762 and nt1764 [32], which suggests that nt1753 mutation is most likely related to lower HBeAg expression levels. Although the G1899A mutation only changes the glycine at codon 29 into aspartic acid and is always accompanied by the G1896A mutation, a previous study revealed that approximately 50% of the HBeAg negative variants contained this combined mutation [33]. Interestingly, the frequency of the A1752G mutation was significantly higher in HBeAg-positive child patients (42.6% vs 27.8%, $P < 0.05$) but was negatively correlated with HBeAg seroconversion. This statistically significant difference was not observed in adult patients. And the older patients possessed higher HBeAg seroconversion ratio, consistent with previous reports [20].

Due to limitations in sample size for cross-sectional studies, previous studies have rarely attempted to analyze the mutation frequency discrepancy between HBeAg negative child patients and HBeAg negative adult patients [34]. It is worth noting that the mutation ratio of G1896A is obviously lower in HBeAg-negative child (41.1%) than in HBeAg-negative adult patients (91.7%) ($P < 0.001$). Similar features were also observed for other sites located in the BCP/precure regions (G1899A and A1846T, $P < 0.05$) (Table 2). These difference suggest the possibility that mutations in the BCP/precure region have less impact on HBeAg seroconversion in child patients than in adult patients. In addition to mutations in the BCP/precure region, deletions in core regions may also have an impact on HBeAg expression. Previous studies have demonstrated that core gene mutants appear during the HBeAg clearance phase [35], and a long-term, large-scale cohort study carried out in children also showed the core mutations that most strongly signified HBeAg seroconversion within 1 year, and the core deletion mutations disappeared after HBeAg seroconversion [36]. Deletions in core regions instead of the G1896A and A1762T/G1764A double mutation were observed in early seroconversion processes in a longitudinal study of infants (our own unpublished data), which also supports the idea that HBeAg seroconversion does not solely rely on BCP/precure mutations. Mutation or deletions in core regions were apt to occur due to host immune stress because core particles are the major carriers of T/B cell epitopes [37], but the fitness cost of core protein silencing was higher due to its essential roles in virus nucleocapsid assembly [38]. In combination with the lower mutation frequency of the BCP/precure regions in HBeAg-negative child patients, we hypothesize that deletions in core regions that were screened by the host immune response were the major reason for the initial phase of HBeAg seroconversions, but due to its high fitness cost, BCP/precure mutations were substituted for core region mutations in the latter phase of HBeAg seroconversion. Sequential nucleotide acid variations in the core regions and BCP/precure regions may explain HBeAg seroconversion.

The relationship between the A1762T/G1764A double mutation and the level of viral DNA replication level is unclear, and studies that employed the transfection of human hepatoma cell lines with clinical HBV viral samples may not truly reflect the true *in vivo* status [26, 39–43]. In our cross-sectional study, BCP double mutations were associated with a lower viral load and an elevated ALT level in HBeAg-positive child patients. There were high levels of viral replication during the immune tolerance phase, while the HBV DNA levels fluctuated and progressively decreased in the immunoreactive phase. The HBeAg remained positive until cleared by the immune system at the end of the second phase. HBeAg has the potential to preferentially deplete

inflammatory HBeAg- and HBcAg-specific Th1 cells that are necessary for viral clearance, therefore promoting HBV persistence[44]. An A1762T/G1764A double mutation in the BCP/precure region would reduce the levels of HBeAg by inhibiting precure mRNA expression[45], leading to the transition from the immune tolerance to the immune reactive phase, followed by lower DNA viral loads and elevated ALT levels. The reasons for the lower viral loads with A1762T/G1764A double mutations in HBeAg-positive patients were more likely related to the decreased HBeAg synthesis and alterations in the complex delicate balance between the virus and the host immune system, but the alterations cannot simply be attributed to reduced viral replication mediated by the double mutation.

Conclusions

The mutation ratios of nt1752, nt1753, nt1762/1764, nt1846, nt1896 and nt1899 were significantly different between the HBeAg-positive and -negative child patients groups, and the frequencies of these mutations in HBeAg-negative child patients were significantly lower than in HBeAg-negative adult patients, implying that the role of BCP/precure mutations is less important in the early phases of HBeAg seroconversion. The number of mutations in the BCP/precure regions was also positively correlated with the HBeAg negative ratio. The correlation analysis between the mutations and the clinical features revealed only that the nt1762/1764 double mutation was associated with lower viral loads and higher ALT levels in HBeAg-positive children with CHB.

Author Contributions

Conceived and designed the experiments: QL AH. Performed the experiments: YH XS XG XL ZT. Analyzed the data: YH HD QL AH. Contributed reagents/materials/analysis tools: HD ZT. Wrote the paper: YH QL AH.

References

1. Dienstag JL. Hepatitis B virus infection. *N Engl J Med*. 2008; 359(14):1486–500. Epub 2008/10/04. doi: [10.1056/NEJMra0801644](https://doi.org/10.1056/NEJMra0801644) 359/14/1486 [pii]. PMID: [18832247](https://pubmed.ncbi.nlm.nih.gov/18832247/)
2. Zanetti AR, Van Damme P, Shouval D. The global impact of vaccination against hepatitis B: a historical overview. *Vaccine*. 2008; 26(49):6266–73. Epub 2008/10/14. doi: [10.1016/j.vaccine.2008.09.056](https://doi.org/10.1016/j.vaccine.2008.09.056) S0264-410X(08)01277-2 [pii]. PMID: [18848855](https://pubmed.ncbi.nlm.nih.gov/18848855/)
3. Yong-Ping Yan H-XS, Zhao-Hua Ji, Zhong-Jun Shao, Zhong-Shu Pu. Epidemiology of Hepatitis B Virus Infection in China: Current Status and Challenges. *Journal of Clinical and Translational Hepatology* 2014; 2:7. doi: [10.14218/JCTH.2013.00027](https://doi.org/10.14218/JCTH.2013.00027)
4. Lavanchy D. Hepatitis B virus epidemiology, disease burden, treatment, and current and emerging prevention and control measures. *J Viral Hepat*. 2004; 11(2):97–107. Epub 2004/03/05. 487 [pii]. PMID: [14996343](https://pubmed.ncbi.nlm.nih.gov/14996343/)
5. McMahon BJ. The natural history of chronic hepatitis B virus infection. *Hepatology*. 2009; 49(5 Suppl): S45–55. Epub 2009/04/29. doi: [10.1002/hep.22898](https://doi.org/10.1002/hep.22898) PMID: [19399792](https://pubmed.ncbi.nlm.nih.gov/19399792/)
6. Hoofnagle JH, Doo E, Liang TJ, Fleischer R, Lok AS. Management of hepatitis B: summary of a clinical research workshop. *Hepatology*. 2007; 45(4):1056–75. Epub 2007/03/30. doi: [10.1002/hep.21627](https://doi.org/10.1002/hep.21627) PMID: [17393513](https://pubmed.ncbi.nlm.nih.gov/17393513/)
7. Lok AS, Heathcote EJ, Hoofnagle JH. Management of hepatitis B: 2000—summary of a workshop. *Gastroenterology*. 2001; 120(7):1828–53. Epub 2001/05/29. S0016508501827657 [pii]. PMID: [11375963](https://pubmed.ncbi.nlm.nih.gov/11375963/)
8. Liaw YF, Lau GK, Kao JH, Gan E. Hepatitis B e antigen seroconversion: a critical event in chronic hepatitis B virus infection. *Dig Dis Sci*. 2010; 55(10):2727–34. Epub 2010/03/20. doi: [10.1007/s10620-010-1179-4](https://doi.org/10.1007/s10620-010-1179-4) PMID: [20238245](https://pubmed.ncbi.nlm.nih.gov/20238245/)
9. EASL Clinical Practice Guidelines: management of chronic hepatitis B. *J Hepatol*. 2009; 50(2):227–42. Epub 2008/12/05. [10.1016/j.jhep.2008](https://doi.org/10.1016/j.jhep.2008). doi: [10.001S0168-8278\(08\)00637-5](https://doi.org/10.001S0168-8278(08)00637-5) [pii]. PMID: [19054588](https://pubmed.ncbi.nlm.nih.gov/19054588/)

10. Liaw YF, Leung N, Kao JH, Piratvisuth T, Gane E, Han KH, et al. Asian-Pacific consensus statement on the management of chronic hepatitis B: a 2008 update. *Hepatology*. 2008; 2(3):263–83. Epub 2009/08/12. doi: [10.1007/s12072-008-9080-3](https://doi.org/10.1007/s12072-008-9080-3) PMID: [19669255](https://pubmed.ncbi.nlm.nih.gov/19669255/)
11. Lok AS, McMahon BJ. Chronic hepatitis B: update 2009. *Hepatology*. 2009; 50(3):661–2. Epub 2009/08/29. doi: [10.1002/hep.23190](https://doi.org/10.1002/hep.23190) PMID: [19714720](https://pubmed.ncbi.nlm.nih.gov/19714720/)
12. Carman WF, Jacyna MR, Hadziyannis S, Karayiannis P, McGarvey MJ, Makris A, et al. Mutation preventing formation of hepatitis B e antigen in patients with chronic hepatitis B infection. *Lancet*. 1989; 2-(8663):588–91. Epub 1989/09/09. S0140-6736(89)90713-7 [pii]. PMID: [2570285](https://pubmed.ncbi.nlm.nih.gov/2570285/)
13. Okamoto H, Yotsumoto S, Akahane Y, Yamanaka T, Miyazaki Y, Sugai Y, et al. Hepatitis B viruses with precore region defects prevail in persistently infected hosts along with seroconversion to the antibody against e antigen. *J Virol*. 1990; 64(3):1298–303. Epub 1990/03/01. PMID: [2304145](https://pubmed.ncbi.nlm.nih.gov/2304145/)
14. Song BC, Cui XJ, Kim HU, Cho YK. Sequential accumulation of the basal core promoter and the pre-core mutations in the progression of hepatitis B virus-related chronic liver disease. *Intervirology*. 2006; 49(5):266–73. Epub 2006/05/23. 93456 [pii] doi: [10.1159/000093456](https://doi.org/10.1159/000093456) PMID: [16714855](https://pubmed.ncbi.nlm.nih.gov/16714855/)
15. Kitab B, Essaid El Feydi A, Afifi R, Trepo C, Benazzouz M, Essamri W, et al. Variability in the precore and core promoter regions of HBV strains in Morocco: characterization and impact on liver disease progression. *PLoS One*. 2012; 7(8):e42891. Epub 2012/08/21. doi: [10.1371/journal.pone.0042891](https://doi.org/10.1371/journal.pone.0042891) PONE-D-12-12954 [pii]. PMID: [22905181](https://pubmed.ncbi.nlm.nih.gov/22905181/)
16. Nie H, Evans AA, London WT, Block TM, Ren XD. Quantitative dynamics of hepatitis B basal core promoter and precore mutants before and after HBeAg seroconversion. *J Hepatol*. 2012; 56(4):795–802. Epub 2011/12/17. doi: [10.1016/j.jhep.2011.11.012](https://doi.org/10.1016/j.jhep.2011.11.012) S0168-8278(11)00863-4 [pii]. PMID: [22173170](https://pubmed.ncbi.nlm.nih.gov/22173170/)
17. Yang HC, Chen CL, Shen YC, Peng CY, Liu CJ, Tseng TC, et al. Distinct evolution and predictive value of hepatitis B virus precore and basal core promoter mutations in interferon-induced hepatitis B e antigen seroconversion. *Hepatology*. 2013; 57(3):934–43. Epub 2012/11/01. doi: [10.1002/hep.26121](https://doi.org/10.1002/hep.26121) PMID: [23112104](https://pubmed.ncbi.nlm.nih.gov/23112104/)
18. Ni YH, Chang MH, Chen PJ, Tsai KS, Hsu HY, Chen HL, et al. Viremia profiles in children with chronic hepatitis B virus infection and spontaneous e antigen seroconversion. *Gastroenterology*. 2007; 132-(7):2340–5. Epub 2007/06/16. S0016-5085(07)00637-3 [pii] doi: [10.1053/j.gastro.2007.03.111](https://doi.org/10.1053/j.gastro.2007.03.111) PMID: [17570209](https://pubmed.ncbi.nlm.nih.gov/17570209/)
19. Kang HS, Kang KS, Song BC. Precore and core promoter mutations of the hepatitis B virus gene in chronic genotype C-infected children. *J Korean Med Sci*. 2011; 26(4):546–50. Epub 2011/04/07. doi: [10.3346/jkms.2011.26.4.546](https://doi.org/10.3346/jkms.2011.26.4.546) PMID: [21468263](https://pubmed.ncbi.nlm.nih.gov/21468263/)
20. Chu CM, Liaw YF. Chronic hepatitis B virus infection acquired in childhood: special emphasis on prognostic and therapeutic implication of delayed HBeAg seroconversion. *J Viral Hepat*. 2007; 14-(3):147–52. Epub 2007/02/20. JVH810 [pii] doi: [10.1111/j.1365-2893.2006.00810.x](https://doi.org/10.1111/j.1365-2893.2006.00810.x) PMID: [17305879](https://pubmed.ncbi.nlm.nih.gov/17305879/)
21. Chauhan R, Kazim SN, Bhattacharjee J, Sakhuja P, Sarin SK. Basal core promoter, precore region mutations of HBV and their association with e antigen, genotype, and severity of liver disease in patients with chronic hepatitis B in India. *J Med Virol*. 2006; 78(8):1047–54. Epub 2006/06/22. doi: [10.1002/jmv.20661](https://doi.org/10.1002/jmv.20661) PMID: [16789012](https://pubmed.ncbi.nlm.nih.gov/16789012/)
22. Lai MW, Lin TY, Tsao KC, Huang CG, Hsiao MJ, Liang KH, et al. Increased seroprevalence of HBV DNA with mutations in the s gene among individuals greater than 18 years old after complete vaccination. *Gastroenterology*. 2012; 143(2):400–7. Epub 2012/05/15. doi: [10.1053/j.gastro.2012.05.002](https://doi.org/10.1053/j.gastro.2012.05.002) S0016-5085(12)00682-8 [pii]. PMID: [22580098](https://pubmed.ncbi.nlm.nih.gov/22580098/)
23. Fang ZL, Sabin CA, Dong BQ, Wei SC, Chen QY, Fang KX, et al. The association of HBV core promoter double mutations (A1762T and G1764A) with viral load differs between HBeAg positive and anti-HBe positive individuals: a longitudinal analysis. *J Hepatol*. 2009; 50(2):273–80. Epub 2008/12/17. doi: [10.1016/j.jhep.2008.09.014](https://doi.org/10.1016/j.jhep.2008.09.014) S0168-8278(08)00714-9 [pii]. PMID: [19070921](https://pubmed.ncbi.nlm.nih.gov/19070921/)
24. Akahane Y, Yamanaka T, Suzuki H, Sugai Y, Tsuda F, Yotsumoto S, et al. Chronic active hepatitis with hepatitis B virus DNA and antibody against e antigen in the serum. Disturbed synthesis and secretion of e antigen from hepatocytes due to a point mutation in the precore region. *Gastroenterology*. 1990; 99(4):1113–9. Epub 1990/10/01. S0016508590003523 [pii]. PMID: [2394332](https://pubmed.ncbi.nlm.nih.gov/2394332/)
25. Kurosaki M, Enomoto N, Asahina Y, Sakuma I, Ikeda T, Tozuka S, et al. Mutations in the core promoter region of hepatitis B virus in patients with chronic hepatitis B. *J Med Virol*. 1996; 49(2):115–23. Epub 1996/06/01. doi: [10.1002/\(SICI\)1096-9071\(199606\)49:2<115::AID-JMV8>3.0.CO;2-8](https://doi.org/10.1002/(SICI)1096-9071(199606)49:2<115::AID-JMV8>3.0.CO;2-8) [pii]. PMID: [8991934](https://pubmed.ncbi.nlm.nih.gov/8991934/)
26. Buckwold VE, Xu Z, Chen M, Yen TS, Ou JH. Effects of a naturally occurring mutation in the hepatitis B virus basal core promoter on precore gene expression and viral replication. *J Virol*. 1996; 70-(9):5845–51. Epub 1996/09/01. PMID: [8709203](https://pubmed.ncbi.nlm.nih.gov/8709203/)
27. Gunther S, Piwon N, Will H. Wild-type levels of pregenomic RNA and replication but reduced pre-C RNA and e-antigen synthesis of hepatitis B virus with C(1653)→T, A(1762)→T and G(1764)→A mutations in the core promoter. *J Gen Virol*. 1998; 79 (Pt 2):375–80. Epub 1998/02/24. PMID: [9472623](https://pubmed.ncbi.nlm.nih.gov/9472623/)

28. Lok AS, Akarca U, Greene S. Mutations in the pre-core region of hepatitis B virus serve to enhance the stability of the secondary structure of the pre-genome encapsidation signal. *Proc Natl Acad Sci U S A*. 1994; 91(9):4077–81. Epub 1994/04/26. PMID: [8171038](#)
29. Yan T, Li K, Li F, Su H, Mu J, Tong S, et al. T1846 and A/G1913 are associated with acute on chronic liver failure in patients infected with hepatitis B virus genotypes B and C. *J Med Virol*. 2011; 83(6):996–1004. Epub 2011/04/20. doi: [10.1002/jmv.22067](#) PMID: [21503912](#)
30. Yin J, Xie J, Liu S, Zhang H, Han L, Lu W, et al. Association between the various mutations in viral core promoter region to different stages of hepatitis B, ranging of asymptomatic carrier state to hepatocellular carcinoma. *Am J Gastroenterol*. 2011; 106(1):81–92. Epub 2010/10/21. doi: [10.1038/ajg.2010.399](#) [ajg2010399 \[pii\]](#) PMID: [20959817](#)
31. Chen CH, Lee CM, Lu SN, Changchien CS, Eng HL, Huang CM, et al. Clinical significance of hepatitis B virus (HBV) genotypes and precore and core promoter mutations affecting HBV e antigen expression in Taiwan. *J Clin Microbiol*. 2005; 43(12):6000–6. Epub 2005/12/08. doi: [10.1128/JCM.43.12.6000-6006.2005](#) PMID: [16333089](#)
32. Parekh S, Zoulim F, Ahn SH, Tsai A, Li J, Kawai S, et al. Genome replication, virion secretion, and e antigen expression of naturally occurring hepatitis B virus core promoter mutants. *J Virol*. 2003; 77(12):6601–12. Epub 2003/05/28. PMID: [12767980](#)
33. Yuan TT, Faruqi A, Shih JW, Shih C. The mechanism of natural occurrence of two closely linked HBV precore predominant mutations. *Virology*. 1995; 211(1):144–56. Epub 1995/08/01. S0042-6822(85)71387-6 [pii] doi: [10.1006/viro.1995.1387](#) PMID: [7645207](#)
34. Nordin M, Ingman M, Lindqvist B, Kidd-Ljunggren K. Variability in the precore and core promoter region of the hepatitis B virus genome. *J Med Virol*. 2014; 86(3):437–45. Epub 2013/11/20. doi: [10.1002/jmv.23839](#) PMID: [24249691](#)
35. Bozkaya H, Ayola B, Lok AS. High rate of mutations in the hepatitis B core gene during the immune clearance phase of chronic hepatitis B virus infection. *Hepatology*. 1996; 24(1):32–7. Epub 1996/07/01. S0270913996003023 [pii] doi: [10.1002/hep.510240107](#) PMID: [8707278](#)
36. Ni YH, Chang MH, Hsu HY, Chen HL. Long-term follow-up study of core gene deletion mutants in children with chronic hepatitis B virus infection. *Hepatology*. 2000; 32(1):124–8. Epub 2000/06/28. S0270-9139(00)44477-0 [pii] doi: [10.1053/jhep.2000.8529](#) PMID: [10869299](#)
37. Pumpens P, Grens E. HBV core particles as a carrier for B cell/T cell epitopes. *Intervirology*. 2001; 44(2–3):98–114. Epub 2001/08/18. 50037 [pii] 50037. PMID: [11805444](#)
38. Birnbaum F, Nassal M. Hepatitis B virus nucleocapsid assembly: primary structure requirements in the core protein. *J Virol*. 1990; 64(7):3319–30. Epub 1990/07/01. PMID: [2191149](#)
39. Pang A, Yuen MF, Yuan HJ, Lai CL, Kwong YL. Real-time quantification of hepatitis B virus core-promoter and pre-core mutants during hepatitis E antigen seroconversion. *J Hepatol*. 2004; 40(6):1008–17. Epub 2004/05/26. doi: [10.1016/j.jhep.2004.02.024](#) S0168827804000984 [pii]. PMID: [15158343](#)
40. Yoo BC, Park JW, Kim HJ, Lee DH, Cha YJ, Park SM. Precore and core promoter mutations of hepatitis B virus and hepatitis B e antigen-negative chronic hepatitis B in Korea. *J Hepatol*. 2003; 38(1):98–103. Epub 2002/12/14. S0168827802003495 [pii]. PMID: [12480566](#)
41. Jammeh S, Tavner F, Watson R, Thomas HC, Karayiannis P. Effect of basal core promoter and pre-core mutations on hepatitis B virus replication. *J Gen Virol*. 2008; 89(Pt 4):901–9. Epub 2008/03/18. doi: [10.1099/vir.0.83468-0](#) 89/4/901 [pii]. PMID: [18343830](#)
42. Chun YK, Kim JY, Woo HJ, Oh SM, Kang I, Ha J, et al. No significant correlation exists between core promoter mutations, viral replication, and liver damage in chronic hepatitis B infection. *Hepatology*. 2000; 32(5):1154–62. Epub 2000/10/26. S0270913900950651 [pii] doi: [10.1053/jhep.2000.19623](#) PMID: [11050069](#)
43. Chen WN, Oon CJ. Mutations and deletions in core promoter and precore stop codon in relation to viral replication and liver damage in Singaporean hepatitis B virus carriers. *Eur J Clin Invest*. 2000; 30(9):787–92. Epub 2000/09/21. eci723 [pii]. PMID: [10998078](#)
44. Milich DR, Chen MK, Hughes JL, Jones JE. The secreted hepatitis B precore antigen can modulate the immune response to the nucleocapsid: a mechanism for persistence. *J Immunol*. 1998; 160(4):2013–21. Epub 1998/02/20. PMID: [9469465](#)
45. Okamoto H, Tsuda F, Akahane Y, Sugai Y, Yoshida M, Moriyama K, et al. Hepatitis B virus with mutations in the core promoter for an e antigen-negative phenotype in carriers with antibody to e antigen. *J Virol*. 1994; 68(12):8102–10. Epub 1994/12/01. PMID: [7966600](#)