The risk of new-onset cancer associated with HFE C282Y and H63D mutations: evidence from 87,028 participants

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Abstract

To investigate the association between mutation of HFE (the principal pathogenic gene in hereditary haemochromatosis) and risk of cancer, we conducted a meta-analysis of all available case–control or cohort studies relating to two missense mutations, C282Y and H63D mutations. Eligible studies were identified by searching databases including PubMed, Embase and the ISI Web of Knowledge. Overall and subgroup analyses were performed and odds ratios (ORs) combined with 95% confidence intervals (Cls) were applied to evaluate the association between C282Y mutation, H63D mutation and cancer risk. Sensitivity and cumulative analyses were used to evaluate the stability of the results. A total of 36 eligible studies were included, comprising 13,680 cases and 73,348 controls. C282Y was significantly associated with elevated cancer risk in a recessive genetic model (OR: 1.991, 95% Cl: 1.448–2.737). On subgroup analysis stratified by cancer type, statistically significantly increased cancer risks were found for breast cancer, colorectal cancer and hepatocellular carcinoma in a recessive model. When stratified by territory, a significantly increased visk of cancer was found in Oceanic populations in a recessive model and in Asian populations in an allele model and dominant model. H63D mutation did not significantly increase overall cancer risk in any genetic model. However, when, stratified by territory, an increased cancer risk was found in the Asian population in an allele and dominant. C282Y but not H63D mutation was related to elevated cancer risk. Further large-scale studies considering gene–environment interactions and functional research should be conducted to further investigate this association.

Keywords: hereditary haemochromatosis • HFE • mutation • C282Y • H63D • meta-analysis • cancer

Introduction

Hereditary haemochromatosis is an autosomal recessive disease, the principal pathogenic gene of which is HFE [1, 2]. The condition is characterized by a disorder of intestinal iron absorption that causes progressive accumulation of iron in organs including the liver, heart and pancreas, leading to their dysfunction [3]. An important pathogenic mechanism may the catalytic activity of iron in the formation of hydroxyl radicals. Iron may also suppress host defence cell activity and promote cancer cell proliferation. It is increasingly reported that two mutations in HFE – C282Y (rs1800562G>A) and H63D (rs1799945 C>G) – are associated with an increased risk of cancers, including hepatocellular [4, 5], breast [6], colorectal [7] and prostate cancer [8], as well as others [9–12]. However, some other studies

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have shown no association between haemochromatosis genotype and neoplasia [13–16]. This controversy warrants further studies.

In 1996, C282Y and H63D were shown to be related to altered iron status [17]. The damage caused by iron overload is associated with oxidative stress, and several studies have demonstrated iron overload to be correlated with carcinogenesis [18]. A number of studies have investigated the association between C282Y and H63D and an increased cancer risk. However, the studies have been underpowered and the findings have proved somewhat controversial. For, a meta-analysis in 2010 by Jin *et al.* [4] found a significant association between C282Y and H63D and hepatocellular carcinoma. However, they included a cross-sectional [19]. Moreover, there are now a number of other studies reported [14, 20–23]. In 2013, Chen *et al.* reported a significant association between C282Y and colorectal cancer. They only used a recessive model and classified all those from the United States as Caucasians [24]. In the same year, Liu *et al.*

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reported similar findings. They classified those from the United States and Brazil as being Europeans [25].

In our study, we have employed cumulative analysis, which has not been previously used. To the best of our knowledge, this is the most comprehensive meta-analysis of C282Y and H63D HFE mutations and the risk of cancer. We included 36 studies, comprising 13,680 cases and 73,348 controls. The malignancies studied were principally hepatocellular, breast, colorectal and prostate carcinomas and acute leukaemia.

Materials and methods

Study identification and selection criteria

We searched PubMed, Embase, the ISI Web of Knowledge, the Chinese Biomedical database and the China National Knowledge Infrastructure to identify relevant studies, from which only case–control and cohort studies published between December 1995 and May 2014 were selected. The terms 'Case–Control Studies or Cohort Studies', 'Neoplasms or Carcinoma', 'Alleles or SNP or Genetic Variation or Mutation or Polymorphism' and 'Haemochromatosis or HFE or C282Y or H63D' were combined. The reference lists and related articles were also scrutinized to identify additional studies.

This study was performed according to the Newcastle–Ottawa Scale (NOS) for meta-analysis of observational studies [26]. The NOS uses a star system (range, 0-9 stars) for evaluating the quality of such studies, allowing a mean value of included studies to be calculated. Articles were selected if they met all of the following criteria: (*i*) the study was a case–control study or cohort study concerning the association between the haemochromatosis genotype C282Y or H63D and risk of cancer; (*ii*) the articles provided data on the distribution of the alleles, the size of the sample and number of controls, the exact number of each genotype or other information to aid the calculations; (*iii*) neoplasms were diagnosed by histopathological biopsy and the controls were free from cancer; and (*iv*) the publication language was English or Chinese. The control group included in our study were hospitalized controls or randomly selected from a pool of eligible participants matched to the index case by age, sex and township of residence.

Data extraction

Two authors (Yang-fan Lv and Xian Chang) extracted information independently from the selected studies. The results were compared and collated, and contradictions were resolved by discussion or by consultation with the corresponding author of the study in question. The data extracted were: first author name; title of article; publication year; country where study was performed; territory of participants; HFE mutation type; precise size of case and control groups; and distribution of genotypes in both case and control groups.

Statistical methods

The control groups of all of the included articles were tested for Hardy– Weinberg equilibrium [27]. The strength of the association between HFE genotypes and cancer risk was measured by the odds ratio (OR) with 95% confidence intervals (CIs). $P_{OB} < 0.05$ was regarded as statistically significant. Subgroup meta-analyses were performed according to cancer type and territory for both C282Y and H63D, independently. The chi-squared test and l^2 statistic were used to evaluate heterogeneity [28]. P-values less than 0.10 indicated heterogeneity among studies and a random-effects model was used to estimate the pooled OR. Otherwise, a fixed-effects model was used. Sensitivity analysis was performed to evaluate the impact of the studies and the stability of the results. To investigate the dynamic trend of the association between HFE mutation and cancer risk, cumulative analysis was performed according to year of publication and sample size [29]. Furthermore, Begg's test [30] and Egger's test [31] were performed to assess the publication bias of the literature [30, 32]. P < 0.05 was considered statistically significant. All statistical tests were performed with STATA 12.0 software [31]. Finally, to adjust for multiple comparisons, the Bonferroni method were applied (see Tables S1 and S2).

Results

Eligible studies

One hundred and twenty-nine studies were found concerning the association between HFE mutation and cancer risk. Following a review of all articles according to the criteria (shown in Fig. 1), 36 eligible studies were included in our pooled analysis. Among these, 33 [7, 8, 10, 12–16, 20–23, 33–53] were concerned with C282Y, 30 [6–10, 12–14, 16, 20, 22, 23, 38–55] with H63D and 27 [7, 8, 10, 12–16, 20, 22, 23, 38–53] with both C282Y and H63D. The principal characteristics of the studies concerning C282Y and H63D are listed in Tables 1 and 2. It should be noted that one study [56] was excluded because it did not provide sufficient data of the distribution of genotypes in both case and control groups.

Meta-analysis results

C282Y

The principal findings for C282Y came from 37 data sets from 33 studies, comprising 7487 cases and 59,324 controls (Table 1). Six studies concerned breast cancer [8, 33, 34, 36, 41, 51], nine colorectal cancer [7, 15, 16, 33, 36, 40, 45, 46, 48], thirteen hepatocellular carcinoma [13, 14, 20-23, 38, 39, 43, 49, 50, 52, 53] and eight studies included six other types of cancer [8, 10, 12, 35, 37, 42, 44, 47] including basal cell carcinoma, cervical cancer, prostatic carcinoma, pancreatic carcinoma, acute leukaemia and ovarian carcinoma. Twenty-seven studies were European [7, 8, 12-15, 20, 21, 33, 35, 37, 38, 41–53], three Oceanian [36, 39, 45], four North American [10, 16, 34, 40] and two Asian [22, 23]. Overall, a significantly elevated cancer risk was found according to a recessive genetic model [57] (OR: 1.991, 95% CI: 1.448-2.737) and an allele model [53] (OR: 1.116, 95% CI: 1.024-1.217) (Fig. 2), whereas no statistically significant difference was found in a dominant model [57] (OR: 1.088, 95% CI: 0.992-1.193). Moderate heterogeneity was detected in the dominant model ($P_{\rm h} = 0.004$, $l^2 = 42.3\%$) and the allele model ($P_{\rm h} = 0.003$,

 $l^2 = 43.1\%$), but there was zero heterogeneity in the recessive model ($P_h = 0.632$, $l^2 = 0.0\%$).

On subgroup analysis stratified by cancer type (Table 3), statistically significantly elevated cancer risk was detected in a recessive model for breast cancer (OR: 2.143, 95% CI: 1.24-3.697), hepatocellular carcinoma (OR: 3.642, 95% CI: 1.454-9.122) and colorectal carcinoma (OR: 1.692, 95% CI: 1.041-2.750). The other cancer types showed no significantly increased risk. On subgroup analysis stratified by territory (Table 3), significantly increased risk of cancer was demonstrated in the Oceanian study population in a recessive model (OR: 2.558, 95% CI: 1.657-3.949), in the Asian population in an allele model (OR: 6.975, 95% CI: 1.315-36.999) with significant heterogeneity and in the Asian population in the dominant model (OR: 5.622, 95% CI: 1.014-31.178). No increased cancer risk was found in either European or North American study populations in any genetic model. Heterogeneity was not observed or was slight in all studies. except in an Asian population using an allele model ($P_{\rm h} = 0.106$, $l^2 = 61.7\%$).

H63D

The results for H63D are comprised of 33 data sets extracted from 30 studies with 6193 cases and 14,024 controls (listed in Table 2). Twelve studies were concerned with hepatocellular carcinoma [13, 14, 20, 22, 23, 38, 39, 43, 45, 49, 50, 52, 53], two with acute leukaemia [12, 47], seven with colorectal cancer [7, 16, 40, 45, 46, 48], five with breast cancer [6, 8, 41, 51, 55] and seven with other neoplasms

including glioma [54], prostatic cancer [8], cervical cancer [42], pancreatic cancer [44], ovarian cancer [10], endometrial cancer [10] and gastric carcinoma [9]. Twenty-two studies were European [7–9, 12– 14, 20, 38, 41–44, 46–54], four were North American [10, 16, 40], three were Oceanian [39, 45], three were Asian [6, 22, 23] and one was South American [55]. Overall, unlike C282Y, no significant increase in cancer risk was found in any genetic model (Table 4). No heterogeneity ($P_{\rm h} = 0.754$, $f^2 = 0.0\%$) was found in the recessive model (Fig. 3); the other two models showed significant heterogeneity (dominant – $P_{\rm h} = 0.002$, $f^2 = 46.7\%$; allele – $P_{\rm h} = 0.002$, $f^2 = 47.2\%$).

Subgroup meta-analysis was performed according to cancer type and territory. For cancer type, elevated cancer risk was detected in a dominant model for 'others', with moderate heterogeneity ($P_{\rm h} = 0.048$, $f^2 = 52.7\%$). Given that 'others' included several types of cancer and that heterogeneity was significant, this result should be viewed with caution. No significantly elevated cancer risk was detected in any other genetic model, suggesting that H63D is not associated with these types of cancer. For territory, increased cancer risk was found in the Asian study population in a dominant model (OR: 2.066, 95% CI: 1.280–3.334, $P_{\rm h} = 0.946$) and an allele model (OR: 1.880, 95% CI: 1.248–2.832, $P_{\rm h} = 0.868$), both with no heterogeneity ($f^2 = 0.0\%$). In the European, North American, Oceanian and South American populations, no significantly elevated cancer risk was detected in any genetic model.



Table 1 Main chara	cteristics	of all case-control c	or cohort studies inclu	uded in H63D and ca	ncer risk							
						Sample size	Case			Control		
First author	Year	study design	Country	I erritory	cancer type	Case/control	cc	CW	MM	CC	CW	ΜM
Beckman	1999	Case-control	Sweden	European	Breast	165/294	-	25	139	4	35	255
Altes	1999	Case-control	France	European	Colorectal	73/76	0	5	68	0	9	70
Beckman	1999	Case-control	Sweden	European	Colorectal	173/294	2	21	150	4	35	255
Gimferrer	1999	Case-control	Spain	European	AL	36/106	0	ი	33	0	9	100
Racchi	1999	Case-control	Italy	European	Hepatocellular	15/130	0	S	12	0	11	119
Beckman	2000	Case-control	Sweden	European	Hepatocellular	54/294		10	43	-	38	255
Parkkila	2001	Case-control	Finland	European	AL	18/102	0	0	18	0	10	92
Fargion	2001	Case-control	Italy	European	Hepatocellular	81/128	0	7	74	0	2	126
Campo S	2001	Case-control	Italy	European	Hepatocellular	23/304	0	0	23	0		303
Lauret	2002	Case-control	Spain	European	Hepatocellular	77/359	0	12	65	0	22	337
Boige	2003	Case-control	France	European	Hepatocellular	133/100	0	7	126	-	9	93
Cauza	2003	Case-control	Australia	Oceanican	Hepatocellular	162/671	5	18	139	5	63	603
Shaheen	2003	Case-control	United States	North American	Colorectal	475/833	0	44	431	S	68	762
Hellerbrand	2003	Case-control	Germany	European	Hepatocellular	137/233	0	17	120	0	10	223
van der	2003	Case-control	Netherlands	European	Colorectal	191/573	0	16	175	S	38	532
Kallianpur	2004	Cohort	United States	North American	Breast	41/129	5	10	26	7	15	107
Abraham	2005	Case-control	Germany	European	Breast	566/649	2	59	505	-	71	577
McGlynn	2005	Case-control	United States	North American	Colorectal	635/650	5	70	560	S	76	571
Robinson	2005	Case-control	United Kingdom	European	Colorectal	327/322	2	50	275	4	39	279
Shi	2005	Case-control	China	Asian	Hepatocellular	56/60	9	с	47	0		59
Festa	2005	Case-control	Sweden	European	Basal cell	241/259	2	17	222	-	22	236
Syrjakoski	2006	Cohort	Finland	European	Prostatic	843/480	6	55	677	с	45	432
Syrjakoski	2006	Cohort	Finland	European	Breast	116/480	-	2	110	S	45	432

Table 1. Continued												
-			-	:		Sample size	Case			Control		
First author	Year	Study design	Country	l erritory	Cancer type	Case/control	CC	CW	MM	CC	CW	MM
Cardoso	2006	Case-control	Portugal	European	Cervical	150/91	0	14	136		5	85
Kondrashova	2006	Case-control	Russia	European	Breast	100/260	0	2	98	0	17	243
Ropero	2007	Case-control	Spain	European	Hepatocellular	196/181	-	12	183	0	23	158
Yonal	2007	Case-control	Turkey	Asian	Hepatocellular	19/251	0	0	19	2	2	247
Hucl	2007	Case-control	Germany	European	Pancreatic	117/428	-	7	109		30	397
Nahon	2008	Cohort	France	European	Hepatocellular	103/198	0	12	91	0	18	180
Ezzikouri	2008	Case-control	France	European	Hepatocellular	96/222	0	2	94	0	с	219
Shi	2009	Case-control	Australia	Oceanica	Colorectal	85/3079	0	16	69	16	424	2639
Shi	2009	Case-control	Polish	European	Colorectal	75/1622	0		74	2	123	1497
Osborne	2010	Cohort	Australia	Oceanican	Colorectal	620/28,414	10	80	530	193	3882	24,339
Osborne	2010	Cohort	Australia	Oceanican	Breast	664/16,399	6	06	565	06	2263	14,046
Gannon	2011	Cohort	Canada	North American	Ovarian	354/80	2	32	320	0	2	78
Ekblom	2012	Cohort	Sweden	European	Colorectal	211/400	2	27	182		47	352
Rodriguez-Lopez	2013	Case-control	Spain	European	AL	59/173	0	2	57	0	16	157
Total						7487/59,324						
C indicates C282Y m	utant and V	W indicates wild-typ	oe respectively, AL in	dicates acute leukaen	nia.							

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Table 2 Main charat	cteristics c	of all case-control c	or cohort studies inclu	ded in H63D and can	cer risk							
			c			Sample size	Case			Control		
First author	Year	stuay aesign	country	l erritory	cancer type	Case/control	Ŧ	MH	MM	Ŧ	MH	MN
Racci	1999	Case-control	Italy	European	Hepatocellular	12/130	0	e	6	e	42	85
Gimferrer	1999	Case-control	Spain	European	AL	36/106	2	=	23	2	28	76
Altes	1999	Case-control	France	European	Colorectal	110/100	9	36	68	2	28	70
Beckman	2000	Case-control	Sweden	European	Hepatocellular	54/294	0	17	37	9	59	229
Campo S	2001	Case-control	Italy	European	Hepatocellular	23/304	-	9	16	12	06	202
Martinez	2001	Case-control	Italy	European	Gliomas	174/144	9	56	112	2	32	110
Lauret	2002	Case-control	Spain	European	Hepatocellular	77/359	0	25	52	33	92	234
Boige	2003	Case-control	France	European	Hepatocellular	133/100	0	41	92		40	59
Cauza	2003	Case-control	Australia	Oceanican	Hepatocellular	162/671	с	31	128	6	133	529
Shaheen	2003	Case-control	United States	North American	Colorectal	475/833	10	88	377	12	135	686
Hellerbrand	2003	Case-control	Germany	European	Hepatocellular	137/233	2	27	108	4	52	177
Abraham	2005	Case-control	Germany	European	Breast	571/646	12	138	421	16	173	457
McGlynn	2005	Case-control	United States	North American	Colorectal	662/650	13	164	485	15	146	489
Robinson	2005	Case-control	United Kingdom	European	Colorectal	327/322	ø	83	236	8	73	241
Shi	2005	Case-control	China	Asian	Hepatocellular	56/60	2	4	50	. 	ę	56
Gunel-Ozcan	2006	Case-control	Turkey	Asian	Breast	88/100	0	39	49		26	73
Syrjakoski	2006	Cohort	Finland	European	Prostatic	843/480	17	177	649	7	88	385
Syrjakoski	2006	Cohort	Finland	European	Breast	116/480	6	26	89	7	88	385
Cardoso	2006	Case-control	Portugal	European	Cervical	185/135	9	43	136	9	46	85
Kondrashova	2006	Case-control	Russica	European	Breast	99/260	2	30	67	5	75	180
Yonal	2007	Case-control	Turkey	Asian	Hepatocellular	19/251	2	9	÷	4	61	186
Hucl	2007	Case-control	Germany	European	Pancreatic	158/549	с	46	109	œ	144	397
Ropero	2007	Case-control	Spain	European	Hepatocellular	196/181	6	85	102	5	52	124

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Table 2. Continued												
-			-	F		Sample size	Case			Control		
First author	Year	Study design	Country	l erritory	Cancer type	Case/control	王	MH	MM	王	MH	MN
Ezzikouri	2008	Case-control	France	European	Hepatocellular	96/226	ę	34	59	2	60	160
Nahon	2008	Cohort	France	European	Hepatocellular	103/198	0	28	75	0	49	149
Shi	2009	Case-control	Australia	Oceanican	Colorectal	78/2614	-	18	59	63	732	1819
Shi	2009	Case-control	Australia	Oceanican	Colorectal	70/1605	4	15	51	40	402	1163
Batschauer	2011	Case-control	Brazil	South American	Breast	68/85	9	13	49	ę	25	57
Gannon	2011	Cohort	Canada	North American	Ovarian	354/80	8	92	254	с	17	60
Gannon	2011	Cohort	Canada	North American	Endometrial	111/80	4	36	71	co	17	60
Ekblom	2012	Cohort	Sweden	European	Colorectal	218/414	2	42	171	13	96	305
Agudo	2013	Case-control	Spain	European	Gastric	323/1158	÷	82	230	23	249	885
Rodriguez-Lopez	2013	Case-control	Spain	European	AL	59/179	-	6	49	5	60	114
Total						6193/14,024						
H indicates H63D mu	Itant and M	/ indicates wild-type	e respectively. AL indi	icates acute leukaemi	- -							

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Publication bias and sensitivity analysis

For C282Y, funnel plots and Begg's and Egger's test were performed to analyse for publication bias in all three genetic models. The shapes of the funnel plots (Fig. 4) appeared symmetrical, indicating no statistically significantly publication bias for the association between C282Y and risk of cancer. This was in agreement with the results from Begg's and Egger's tests (Table 3). Similarly, there was no evidence of publication bias for H63D (Table 4). All of these results indicate that the findings of our study were robust.

Sensitivity analysis [58] was conducted to determine the publication bias and influence of each study on the pooled OR by sequentially omitting individual studies from the analysis. The series of pooled ORs with 95% CIs lies not far from the midline for the C282Y mutation, which means that the statistical findings were not materially altered by the elimination of any study in the recessive model (Fig. 5). Thus, the possible positive association between C282Y and cancer risk was stable, especially for breast cancer, colorectal cancer and hepatocellular carcinoma.

Similar results were achieved in the sensitivity analysis for H63D mutation, confirming the stability of our findings for H63D.

Cumulative analysis

Cumulative meta-analysis [29] was performed by sorting studies by chronological order and sample size. This allows the stability of the research findings over time to be explored. As shown in Figure 6,



Fig. 2 Forest plot (fixed-effects model) showed C282Y was associated with increased cancer risk in an allele model. Each study is shown by the point estimate of the OR (the size of the square is proportional to the weight of each study) and 95% CI for the OR (extending lines).

lable 3 Pooled analy	ysis of association	of C282Y and cancer risk								
		Dominant model			Recessive model			Allele model		
	Case/control	(CC+CW) versus WW			CC versus (CW+WW)			C versus W		
		OR	μ	μ	OR	μ	ß	OR	Ph	ß
Total	7487/59,324	1.088 (0.992–1.193)	0.004	42.30%	1.991 (1.448–2.737)	0.811	0.00%	1.116 (1.024–1.217)	0.003	43.10%
Cancer type										
Breast	1652/18,211	1.046 (0.884–1.236)	0.031	59.40%	2.143 (1.242–3.697)	0.673	0.00%	1.091 (0.934–1.274)	0.025	61.20%
Colorectal	2865/36,263	1.062 (0.927–1.216)	0.77	0.00%	1.692 (1.041–2.750)	0.523	0.00%	1.073 (0.946–1.219)	0.852	%00.0
Hepatocellular	1152/3131	1.574 (1.217–2.036)	0.016	51.40%	3.642 (1.454–9.122)	0.568	0.00%	1.608 (1.263–2.049)	0.01	54.10%
Others	1818/1719	0.874 (0.662–1.152	0.252	22.30%	1.546 (0.593-4.031)	0.724	0.00%	0.920 (0.709–1.194)	0.324	13.60%
Territory										
European	4376/8758	1.057 (0.921–1.213)	0.01	42.90%	1.255 (0.702–2.244)	0.831	%00.0	1.059 (0.929–1.207)	0.026	37.70%
Oceanican	1531/48,563	1.083 (0.937–1.251)	0.46	0.00%	2.558 (1.657–3.949)	0.795	%00.0	1.142 (1.000–1.305)	0.373	4.00%
Asian	75/311	5.622 (1.014–31.178)	0.261	20.90%	6.647 (0.807–54.756)	0.402	0.00%	6.975 (1.315–36.999)	0.106	61.70%
North American	1505/1692	1.166 (0.917–1.482)	0.029	66.80%	1.682 (0.721–3.923)	0.572	%00.0	1.183 (0.944–1.482)	0.017	70.40%
Begg		P = 0.367			<i>P</i> = 0.216			<i>P</i> = 0.425		
Egger		<i>P</i> = 0.217			P = 0.100			P = 0.334		
$P_{\rm h}$: test for heterogen ${\cal P}$: the percentage of t C indicates C282Y mu	eity, OR: odds rati total variation acro latant and W indica:	o, CI: confidence interval. ss studies that is a result tes wild-type respectively.	of heteroge	eneity rather	than chance.					

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Table 4 Pooled analy	sis of association	of H63D and cancer risk								
		Dominant model			Recessive model			Allele model		
	Case/Control	(HH+HM) versus WW			HH <i>versus</i> (HW+WW)			H versus W		
		OR	Ph	P	OR	ĥ	P	OR	Ph	ß
Total	6193/14,024	1.107 (1.025–1.196)	0.002	46.70%	1.215 (0.966–1.528)	0.754	0.00%	1.095 (1.023-1.172)	0.002	47.20%
Cancer type										
Breast	942/1571	1.014 (0.841–1.221)	0.072	53.50%	0.996 (0.555–1.788)	0.629	0.00%	1.010 (0.857–1.191)	0.2	33.20%
Colorectal	1940/6538	1.065 (0.929–1.221)	0.339	11.90%	1.152 (0.781–1.699)	0.496	0.00%	1.064 (0.942–1.202)	0.252	23.20%
Hepatocellular	1068/3003	1.169 (0.988–1.383)	0.051	43.90%	1.447 (0.828–2.529)	0.321	12.90%	1.126 (0.971–1.306)	0.017	52.20%
AL	95/285	0.681 (0.395–1.175)	0.013	83.80%	1.447 (0.333–6.289)	0.279	14.70%	0.785 (0.486–1.268)	0.01	84.80%
Others	2148/2627	1.212 (1.048–1.402)	0.048	52.70%	1.278 (0.844–1.934)	0.719	0.00%	1.191 (1.047–1.355)	0.053	51.70%
Territory										
European	4050/6995	1.089 (0.992–1.195)	0.001	55.70%	1.162 (0.872–1.549)	0.783	0.00%	1.074 (0.989–1.167)	0.001	57.10%
Oceanican	310/4890	0.907 (0.685–1.200)	0.654	0.00%	1.590 (0.742–3.405)	0.411	0.00%	0.960 (0.748–1.232)	0.464	0.00%
Asian	163/411	2.066 (1.280–3.334)	0.946	0.00%	3.147 (0.853–11.612)	0.268	24.00%	1.880 (1.248–2.832)	0.868	0.00%
North American	1602/1643	1.187 (1.001–1.408)	0.683	0.00%	0.986 (0.603–1.611)	0.669	0.00%	1.147 (0.984–1.336)	0.697	0.00%
South American	68/85	0.789 (0.393–1.584)			2.645 (0.636–10.994)			1.010 (0.564–1.809)		
Begg		P = 0.963			P = 0.466			P = 0.963		
Egger		P = 0.987			P = 0.526			P = 0.995		

 $P_{\rm h}$: test for heterogeneity, OR: odds ratio, CI: confidence interval. \hat{F} : the percentage of total variation across studies that is a result of heterogeneity rather than chance. H indicates H63D mutant and W indicates wild-type respectively.

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Racchi Gimferrer Altes Beckman Campo S Martinez Lauret Shaheen Hellerbrand Boige	Year	Cancer	OR (95% CI)	Weight
Gimferrer Altes Beckman Campo S Martinez Lauret Shaheen Hellerbrand Boige	1000	Honotocollular		0.59
Altes Beckman Campo S Martinez Lauret Shaheen Hellerbrand Boige	1999		1.40 (0.07, 29.83)	0.58
Aites Beckman Campo S Martinez Lauret Shaheen Hellerbrand Boige	1999	AL	3.06 (0.41, 22.35)	1.52
Geckman Campo S Martinez Lauret Shaheen Hellerbrand Boige	1999	Colorectal		1.99
Campo S Martinez Lauret Shaheen Hellerbrand Boige	2000	Hepatocellular	0.41 (0.02, 7.33)	0.63
Martinez Lauret Shaheen Hellerbrand Boige	2001	Hepatocellular		1.21
Lauret Shaheen Hellerbrand Boige	2001	Gliomas	2.54 (0.50, 12.76)	2.01
Shaheen Hellerbrand Boige	2002	Hepatocellular	0.06 (0.00, 1.04)	0.67
Hellerbrand Boige	2003	Colorectal	1.47 (0.63, 3.43)	7.32
Boige	2003	Hepatocellular	0.85 (0.15, 4.69)	1.79
Causa	2003	Hepatocellular	0.25 (0.01, 6.16)	0.51
Cauza	2003	Hepatocellular	1.39 (0.37, 5.19)	3.02
Robinson	2005	Colorectal	0.98 (0.36, 2.66)	5.33
McGlynn	2005	Colorectal	0.85 (0.40, 1.80)	9.32
Abraham	2005	Breast	0.85 (0.40, 1.80)	9.16
Shi	2005	Hepatocellular	2.19 (0.19, 24.79)	0.89
Gunel–Ozcan	2006	Breast	0.37 (0.02, 9.32)	0.51
Syrjakoski	2006	Breast	0.59 (0.07, 4.82)	1.19
Kondrashova	2006	Breast	1.05 (0.20, 5.51)	1.91
Syrjakoski	2006	Prostatic	1.39 (0.57, 3.38)	6.67
Cardoso	2006	Cervical	0.73 (0.23, 2.32)	3.94
Yonal	2007	Hepatocellular	7.26 (1.24, 42.53)	1.68
Hucl	2007	Pancreatic	1.31 (0.34, 4.99)	2.93
Ropero	2007	Hepatocellular	1.69 (0.56, 5.15)	4.24
Ezzikouri	2008	Hepatocellular	3.55 (0.58, 21.59)	1.61
Shi	2009	Colorectal -	0.53 (0.07, 3.84)	1.33
Shi	2009	Colorectal	2.37 (0.82, 6.82)	4.70
Batschauer	2011	Breast	2.65 (0.64, 10.99)	2.59
Gannon	2011	Ovarian	0.59 (0.15, 2.29)	2.88
Gannon	2011	Endometrial	0.96 (0.21, 4.41)	2.26
Ekblom	2012	Colorectal	0.72 (0.25, 2.06)	4.81
Rodriguez–Lopez	2013	AL —	• 0.60 (0.07, 5.24)	1.12
Agudo	2013	Gastric	1.74 (0.84, 3.60)	9.87
Nahon	2008	Hepatocellular	(Excluded)	0.00
Overall (I–square	d = 0.0%	p = 0.754)	1.22 (0.97, 1.53)	100.00
		I .00381	1 1 262	

Fig. 3 Forest plot (fixed-effects model) indicated H63D was not associated with increased cancer risk in a recessive model. Each study is shown by the point estimate of the OR combined with 95% CI for the OR. % weight represents the weight of each study.

there is a tendency towards a positive association between C282Y and cancer risk with time. Simultaneously, 95% CIs became narrower, indicating improved precision and accuracy. Increasing sample sizes also narrowed the 95% CIs; the implications being similar.

Discussion

In this compound study, we performed a meta-analysis of the association between mutations of the HFE gene and risk of cancer including 36 eligible case-control or cohort studies. Thirty-three studies concerned the C282Y mutation, with 7487 cases and 59,324 controls. C282Y was found to increase the risk of cancer

twofold in the recessive model and 1.1-fold in the allele mode. On stratified analysis by cancer type, a statistically significant increase was found for breast cancer, colorectal cancer and hepatocellular carcinoma in the recessive model, in accordance with the studies of Jin *et al.* [4], Chen *et al.* [24] and Liu *et al.* [25]. These results suggest that the C282Y/C282Y genotype is associated with a twofold elevated risk for breast cancer, a 1.7-fold elevated risk of colorectal, and a 3.6-fold increased risk of hepatocellular cancer. There is insufficient evidence to conclude that it is a risk factor for other types of cancer. Subgroup analysis stratified by territory showed that the C282Y mutation was associated with a 2.6-fold increased risk of cancer in Oceanian populations in a recessive model and by 6.9-fold in Asian populations in an allele model. These findings suggest that the living



Fig. 4 Funnel plot illustrating publication bias (recessive model of C282Y polymorphism).



Fig. 5 Analysis of the influence of summary odds ratio coefficients on the association between C282Y mutation and cancer risk in the recessive model.

environment, genetic background and dietary habits are candidate factors that influence the risk of cancer because of HFE mutations. This is the most comprehensive study reported to date, evaluating the association between HFE genotype and overall cancer risk, with stratification based on territory.

H63D, another missense mutation of the *HFE* gene, was investigated in thirty studies with 6193 cases and 14,024 controls. We found that H63D did not increase the overall cancer risk or the risk of particular types of cancer on subgroup analysis, with ORs only slightly over 1 in all genetic models. However, the result of 'others' showed H63D increased cancer risk 1.2-fold in both dominant and allele models. Given that 'others' included several types of cancer, and that the heterogeneity in both model was moderate, we advise that these findings should be viewed with caution. Our results indicated that H63D is a weak or irrelevant factor in the development of cancer. However, in the Asian study population, H63D was found to be related to elevated cancer risk in both a dominant by twofold and an allele model by 1.9-fold, suggesting a possible role for genetic background, diet and lifestyle, and environmental conditions.

Generally, it could be concluded from our study that the C282Y mutation, especially the C82Y/C282Y genotype, is a risk factor for cancer. The association between C282Y and breast, colorectal and hepatocellular carcinoma was statistically significant. However, H63D was not a distinct risk factor or only a weak one. It is well known that HFE is an atypical major histocompatibility complex class I molecule, affecting iron load and immune function through its interaction with β 2 microglobulin (β 2 m) and the TfRs (TfR1 and TfR2) [59, 60]. Generally, normal HFE associates with β2 m, transits to the membrane, and binds with TfRs. When combining with TfR1, HFE competes with transferrin to limit the rate of iron uptake. promoting a homoeostatic level of iron load. However, when forming a complex with TfR2, it stimulates the secretion of Hepcidin, thus suppressing the iron export protein ferroportin and promoting cells to retain iron intracellularly. All these finding indicated that HFE plays vital role in iron homoeostasis regulation [61]. Expectedly, mutations in HFE cause the disruption of HFE function, leading to iron overload. Specifically, C282Y polymorphism cannot interact with $\beta 2$ m, preventing its surface translocation and variant H63D translocates to the cell surface but fails to participate in the interactions with the TfR1, which might promote the interaction with TfR2 in hepatocytes, causing a systemic increase in hepcidin and suppression of ferroportin [59, 62].

The mechanism of the damage caused by excess iron might be related to the creation of free radicals during the Fenton reaction, leading to the formation of reactive oxygen species (ROSs). It is known that ROSs can cause lipid peroxidation, protein modification, and DNA and RNA mutations, thus resulting in dysregulation of normal cell functioning, pathological states and cell death [63, 64]. Specifically, intracellular iron overload leads to cell cycle arrest at the G1/S stage by affecting the expression of certain cyclins and protein kinases. Reactive oxygen species can react with DNA, causing damage, mutation, oncogene activation or inactivation of cancer suppressor genes. In addition, hydroxyl radicals may cause apoptosis [65] because of their effects on mitochondrial and lysosomal membranes.

As suggested by the American Association for the Study of Liver Diseases, phlebotomy is the principle treatment for hereditary haemochromatosis, being an effective method for maintaining serum ferritin levels. Thus, a number of the cases included in our study had probably undergone phlebotomy, which would have reduced their serum ferritin levels and might have reduced their susceptibility to cancer. This may have affected the results of our study.

Our study has limitations. First, our meta-analysis was based on unadjusted related data, and any confounding factors could not be controlled for because most of the included studies did not provide any relevant data. Second, the sample sizes of several of the studies might not have been large enough to detect any possible risks associated with the HFE mutations. This is most likely to have applied to the results concerning Oceanian and Asian populations. Third, because

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Author	Year	Cancer				OR (95% CI)
Beckman	1999	Breast 🗲	•			0.44 (0.05, 3.99)
Beckman	1999	Colorectal	 •			0.66 (0.17, 2.56)
Altes	1999	Colorectal				1.99 (1.45, 2.74)
Gimferrer	1999	AL				1.99 (1.45, 2.74)
Racchi	1999	Hepatocellular				1.99 (1.45, 2.74)
Beckman	2000	Hepatocellular		+	-	0.99 (0.29, 3.34)
Parkkila	2001	AL				1.99 (1.45, 2.74)
Fargion	2001	Hepatocellular		│		1.99 (1.45, 2.74)
Campo S	2001	Hepatocellular				1.99 (1.45, 2.74)
Lauret	2002	Hepatocellular				1.99 (1.45, 2.74)
Boige	2003	Hepatocellular				0.83 (0.27, 2.60)
van der	2003	Colorectal	+			0.77 (0.27, 2.21)
Cauza	2003	Hepatocellular	_	+ +	_	1.57 (0.70, 3.52)
Shaheen	2003	Colorectal	_	+ •	1	1.38 (0.63, 3.01)
Hellerbrand	2003	Hepatocellular				1.99 (1.45, 2.74)
Kallianpur	2004	Breast		↓ → ─	•	1.63 (0.85, 3.14)
Shi	2005	Hepatocellular		↓	-	1.82 (0.96, 3.44)
Festa	2005	Basal cell		├ →	-	1.84 (0.99, 3.41)
Robinson	2005	Colorectal		+		1.58 (0.88, 2.82)
McGlynn	2005	Colorectal		↓ →		1.59 (0.93, 2.73)
Abraham	2005	Breast		↓ →		1.62 (0.96, 2.74)
Cardoso	2006	cervical		+		1.54 (0.91, 2.58)
Syrjakoski	2006	prostatic		↓ →		1.56 (0.96, 2.53)
Syrjakoski	2006	Breast		↓ → →		1.55 (0.97, 2.49)
Kondrashova	2006	Breast				1.99 (1.45, 2.74)
Ropero	2007	Hepatocellular		—		1.57 (0.98, 2.50)
Yonal	2007	Hepatocellular		→		1.59 (1.00, 2.52)
Hucl	2007	Pancreatic		 →→		1.62 (1.03, 2.56)
Nahon	2008	Hepatocellular		→		1.99 (1.45, 2.74)
Ezzikouri	2008	Hepatocellular				1.99 (1.45, 2.74)
Shi	2009	Colorectal				1.66 (1.06, 2.60)
Shi	2009	Colorectal		 →→		1.64 (1.05, 2.56)
Osborne	2010	Breast				1.86 (1.28, 2.70)
Osborne	2010	Colorectal				1.98 (1.43, 2.73)
Gannon	2011	Ovarian		→		1.97 (1.43, 2.71)
Ekblom	2012	Colorectal		→		1.99 (1.45, 2.74)
Rodriguez–Lopez	2013	AL				1.99 (1.45, 2.74)
						-1
		.049		1		20.4

Fig. 6 Forest plots for cumulative meta-analysis of the association between C282Y and cancer risk in the recessive model (year of publication).

cancer is a complex disease with a multifactorial aetiology, genegene and gene-environment interactions should be evaluated; however, we did not address this in our study. Last, most of the studies included in our meta-analysis were concerned with breast cancer, colorectal cancer or hepatocellular carcinoma; those concerning several other types of cancer were simply combined together as 'others'. As a consequence, our findings with these studies might not be precise. We hope to address this in future studies.

In conclusion, this is a comprehensive meta-analysis concerning HFE gene mutation (C282Y and H63D) and overall cancer risk. The C282Y mutation was associated with increased overall cancer susceptibility, especially for hepatocellular carcinoma, breast cancer and colorectal cancer, whereas the H63D mutation produced non-significant results for these three types of cancer. The effect of territory on the association between HFE mutation and cancer could be a factor in susceptibility. Further well-designed epidemiological studies of cancer types and territory and large-scale studies concerning gene-gene or gene-environment interactions should be conducted to clarify the association. The molecular mechanism of how C282Y increases cancer risk also merits further study, to aid understanding of the role of HFE gene mutation in carcinogenesis.

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Conflicts of interest

The authors disclose no potential conflicts of interest.

Supporting information

Additional Supporting Information may be found in the online version of this article:

Table S1 Summary odds ratios for C282Y.

Table S2 Summary odds ratios for H63D.

References

- Kanwar P, Kowdley KV. Metal storage disorders: Wilson disease and hemochromatosis. *Med Clin North Am.* 2014; 98: 87–102.
- Kanwar P, Kowdley KV. Diagnosis and treatment of hereditary hemochromatosis: an update. *Expert Rev Gastroenterol Hepatol.* 2013; 7: 517–30.
- Nichols GM, Bacon BR. Hereditary hemochromatosis: pathogenesis and clinical features of a common disease. Am J Gastroenterol. 1989: 84: 851–62.
- Jin F, Qu LS, Shen XZ. Association between C282Y and H63D mutations of the HFE gene with hepatocellular carcinoma in European populations: a meta-analysis. J Exp Clin Cancer Res. 2010; 29: 18.
- Kew MC. Hepatic iron overload and hepatocellular carcinoma. *Liver Cancer.* 2014; 3: 31–40.
- Gunel-Ozcan A, Alyilmaz-Bekmez S, Guler EN, et al. HFE H63D mutation frequency shows an increase in Turkish women with breast cancer. BMC Cancer. 2006; 6: 37.
- Robinson JP, Johnson VL, Rogers PA, et al. Evidence for an association between compound heterozygosity for germ line mutations in the hemochromatosis (HFE) gene and increased risk of colorectal cancer. Cancer Epidemiol Biomarkers Prev. 2005; 14: 1460–3.
- Syrjakoski K, Fredriksson H, Ikonen T, et al. Hemochromatosis gene mutations among Finnish male breast and prostate cancer patients. Int J Cancer. 2006; 118: 518–20.
- Agudo A, Bonet C, Sala N, et al. Hemochromatosis (HFE) gene mutations and risk of gastric cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC) study. Carcinogenesis. 2013; 34: 1244–50.
- Gannon PO, Medelci S, Le Page C, et al. Impact of hemochromatosis gene (HFE) mutations on epithelial ovarian cancer risk and prognosis. Int J Cancer. 2011; 128: 2326–34.

- Pflugmacher R, Schleicher P, Schroeder RJ, et al. Maintained pain reduction in five patients with multiple myeloma 12 months after treatment of the involved cervical vertebrae with vertebroplasty. Acta Radiol. 2006; 47: 823–9.
- Rodriguez-Lopez R, Donoso M, Fernandez-Cavada M, et al. Diagnostic utility of HFE variants in Spanish patients: association with HLA alleles and role in susceptibility to acute lymphoblastic leukemia. *Gene.* 2013; 514: 31–5.
- 13. **Boige V, Castera L, de Roux N**, *et al.* Lack of association between HFE gene mutations and hepatocellular carcinoma in patients with cirrhosis. *Gut.* 2003; 52: 1178–81.
- 14. **Campo S, Restuccia T, Villari D**, *et al*. Analysis of haemochromatosis gene mutations in a population from the Mediterranean Basin. *Liver*. 2001; 21: 233–6.
- van der A D, van der Hel O, Roest M, et al. Heterozygosity for the Cys282Tyr mutation in the HFE gene and the risk of colorectal cancer (Netherlands). Cancer Causes Control. 2003; 14: 541–5.
- McGlynn KA, Sakoda LC, Hu Y, et al. Hemochromatosis gene mutations and distal adenomatous colorectal polyps. *Cancer Epidemiol Biomarkers Prev.* 2005; 14: 158–63.
- Jouanolle AM, Fergelot P, Gandon G, et al. A candidate gene for hemochromatosis: frequency of the C282Y and H63D mutations. *Hum Genet.* 1997; 100: 544–7.
- Fonseca-Nunes A, Jakszyn P, Agudo A. Iron and cancer risk–a systematic review and meta-analysis of the epidemiological evidence. *Cancer Epidemiol Biomarkers Prev.* 2014; 23: 12–31.
- Willis G, Bardsley V, Fellows IW, et al. Hepatocellular carcinoma and the penetrance of HFE C282Y mutations: a cross sectional study. *BMC Gastroenterol.* 2005; 5: 17.
- Racchi O, Mangerini R, Rapezzi D, et al. Mutations of the HFE gene and the risk of hepatocellular carcinoma. *Blood Cells Mol Dis.* 1999; 25: 350–3.

- Fargion S, Stazi MA, Fracanzani AL, et al. Mutations in the HFE gene and their interaction with exogenous risk factors in hepatocellular carcinoma. Blood Cells Mol Dis. 2001; 27: 505–11.
- Shi WJ, Chen H, Zhou B, et al. [Association of mutations of HFE gene and hepatocellular carcinoma following chronic hepatitis B]. Zhonghua Gan Zang Bing Za Zhi. 2005; 13: 682–4.
- Yonal O, Hatirnaz O, Akyuz F, et al. HFE gene mutation, chronic liver disease, and iron overload In Turkey. *Dig Dis Sci.* 2007; 52: 3298–302.
- Chen W, Zhao H, Li T, et al. HFE gene C282Y variant is associated with colorectal cancer in Caucasians: a meta-analysis. *Tumour Biol.* 2013; 34: 2255–9.
- Liu X, Lv C, Luan X, et al. C282Y polymorphism in the HFE gene is associated with risk of breast cancer. *Tumour Biol.* 2013; 34: 2759–64.
- Wells G, Shea B, O'Connell D, et al. The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses. Ottawa, ON: Ottawa Health Research Institute; 1999.
- Chen JJ, Duan T, Single R, et al. Hardy-Weinberg testing of a single homozygous genotype. *Genetics*. 2005; 170: 1439–42.
- Lau J, Ioannidis JP, Schmid CH. Quantitative synthesis in systematic reviews. Ann Intern Med. 1997; 127: 820–6.
- Leimu R, Koricheva J. Cumulative metaanalysis: a new tool for detection of temporal trends and publication bias in ecology. *Proc Biol Sci.* 2004; 271: 1961–6.
- Seagroatt V, Stratton I. Bias in meta-analysis detected by a simple, graphical test. Test had 10% false positive rate. *BMJ.* 1998; 316: 470.
- Sterne JA, Egger M, Smith GD. Systematic reviews in health care: investigating and dealing with publication and other biases in meta-analysis. *BMJ*. 2001; 323: 101–5.

- Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics.* 1994; 50: 1088–101.
- Beckman LE, Van Landeghem GF, Sikstrom C, et al. Interaction between haemochromatosis and transferrin receptor genes in different neoplastic disorders. *Carcinogene*sis. 1999; 20: 1231–3.
- Kallianpur AR, Hall LD, Yadav M, et al. Increased prevalence of the HFE C282Y hemochromatosis allele in women with breast cancer. *Cancer Epidemiol Biomarkers Prev.* 2004; 13: 205–12.
- Festa F, Kumar R, Sanyal S, et al. Basal cell carcinoma and variants in genes coding for immune response, DNA repair, folate and iron metabolism. *Mutat Res.* 2005; 574: 105–11.
- Osborne NJ, Gurrin LC, Allen KJ, et al. HFE C282Y homozygotes are at increased risk of breast and colorectal cancer. *Hepatology*. 2010; 51: 1311–8.
- Parkkila S, Niemela O, Savolainen ER, et al. HFE mutations do not account for transfusional iron overload in patients with acute myeloid leukemia. *Transfusion*. 2001; 41: 828–31.
- Beckman LE, Hagerstrand I, Stenling R, et al. Interaction between haemochromatosis and transferrin receptor genes in hepatocellular carcinoma. Oncology. 2000; 59: 317–22.
- Cauza E, Peck-Radosavljevic M, Ulrich-Pur H, et al. Mutations of the HFE gene in patients with hepatocellular carcinoma. Am J Gastroenterol. 2003; 98: 442–7.
- Shaheen NJ, Silverman LM, Keku T, et al. Association between hemochromatosis (HFE) gene mutation carrier status and the risk of colon cancer. J Natl Cancer Inst. 2003; 95: 154–9.
- Abraham BK, Justenhoven C, Pesch B, et al. Investigation of genetic variants of genes of the hemochromatosis pathway and their role in breast cancer. Cancer Epidemiol Biomarkers Prev. 2005; 14: 1102–7.
- Cardoso CS, Araujo HC, Cruz E, et al. Haemochromatosis gene (HFE) mutations in viral-associated neoplasia: linkage to cervi-

cal cancer. *Biochem Biophys Res Commun.* 2006; 341: 232–8.

- Ropero P, Briceno O, Lopez-Alonso G, et al. The H63D mutation in the HFE gene is related to the risk of hepatocellular carcinoma. Rev Esp Enferm Dig. 2007; 99: 376– 81.
- Hucl T, Kylanpaa-Back ML, Witt H, et al. HFE genotypes in patients with chronic pancreatitis and pancreatic adenocarcinoma. *Genet Med.* 2007; 9: 479–83.
- Shi Z, Johnstone D, Talseth-Palmer BA, et al. Haemochromatosis HFE gene polymorphisms as potential modifiers of hereditary nonpolyposis colorectal cancer risk and onset age. Int J Cancer. 2009; 125: 78–83.
- Ekblom K, Marklund SL, Palmqvist R, et al. Iron biomarkers in plasma, HFE genotypes, and the risk for colorectal cancer in a prospective setting. *Dis Colon Rectum*. 2012; 55: 337–44.
- Gimferrer E, Nomdedeu J, Gich I, et al. Prevalence of hemochromatosis related HFE gene mutations in patients with acute myeloid leukemia. *Leuk Res.* 1999; 23: 597–8.
- Altes A, Gimferrer E, Capella G, et al. Colorectal cancer and HFE gene mutations. *Haematologica*. 1999; 84: 479–80.
- Lauret E, Rodriguez M, Gonzalez S, et al. HFE gene mutations in alcoholic and virusrelated cirrhotic patients with hepatocellular carcinoma. Am J Gastroenterol. 2002; 97: 1016–21.
- Hellerbrand C, Poppl A, Hartmann A, et al. HFE C282Y heterozygosity in hepatocellular carcinoma: evidence for an increased prevalence. *Clin Gastroenterol Hepatol.* 2003; 1: 279–84.
- Kondrashova TV, Neriishi K, Ban S, et al. Frequency of hemochromatosis gene (HFE) mutations in Russian healthy women and patients with estrogen-dependent cancers. *Biochim Biophys Acta*. 2006; 1762: 59–65.
- Nahon P, Sutton A, Rufat P, et al. Liver iron, HFE gene mutations, and hepatocellular carcinoma occurrence in patients with cirrhosis. Gastroenterology. 2008; 134: 102–10.
- 53. Ezzikouri S, El Feydi AE, El Kihal L, et al. Prevalence of common HFE and SERPINA1

mutations in patients with hepatocellular carcinoma in a Moroccan population. *Arch Med Res.* 2008; 39: 236–41.

- Martinez di Montemuros F, Tavazzi D, Salsano E, et al. High frequency of the H63D mutation of the hemochromatosis gene (HFE) in malignant gliomas. *Neurology*. 2001; 57: 1342.
- Batschauer AP, Cruz NG, Oliveira VC, et al. HFE, MTHFR, and FGFR4 genes polymorphisms and breast cancer in Brazilian women. *Mol Cell Biochem*. 2011; 357: 247–53.
- Asberg A, Thorstensen K, Irgens WO, et al. Cancer risk in HFE C282Y homozygotes: results from the HUNT 2 study. Scand J Gastroenterol. 2013; 48: 189–95.
- Lewis CM, Knight J. Introduction to genetic association studies. *Cold Spring Harb Protoc*. 2012; 2012: 297–306.
- Copas J, Shi JO. Meta-analysis, funnel plots and sensitivity analysis. *Biostatistics*. 2000; 1: 247–62.
- Weston C, Connor J. Evidence for the influence of the iron regulatory MHC class I molecule HFE on tumor progression in experimental models and clinical populations. *Transl Oncol.* 2014: 6: 6.
- 60. **Maja V.** Molecular basis of HFE–hemochromatosis. *Front Pharmacol.* 2014; 5: 42.
- Almeida SFD, Carvalho IF, Cardoso CS, et al. HFE cross-talks with the MHC class I antigen presentation pathway. *Blood.* 2005; 3: 971–7.
- Parkkila S, Niemela O, Britton RS, et al. Molecular aspects of iron absorption and HFE expression. *Gastroenterology.* 2001; 6: 1489–96.
- Santos MA, Marques SM, Chaves S. Hydroxypyridinones as "privileged" chelating structures for the design of medicinal drugs. *Coord Chem Rev.* 2012; 256: 240–59.
- Weinreb O, Mandel S, Youdim MB, et al. Targeting dysregulation of brain iron homeostasis in Parkinson's disease by iron chelators. *Free Radic Biol Med.* 2013; 62: 52–64.
- Valko M, Rhodes CJ, Moncol J, et al. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact.* 2006; 160: 1–40.