

Current Advances in PD-1/PD-L1 Blockade in Recurrent Epithelial Ovarian Cancer

Yuedi Zhang^{1,2†}, Qiulin Cui^{1†}, Manman Xu¹, Duo Liu¹, Shuzhong Yao^{1*} and Ming Chen^{1*}

¹ Department of Gynecology, The First Affiliated Hospital of Sun Yat-sen University, Guangzhou, China, ² School of Medicine, Sun Yat-sen University, Guangzhou, China

OPEN ACCESS

Edited by:

Stergios Boussios, King's College London, United Kingdom

Reviewed by:

AJ Robert McGray, University at Buffalo, United States Nicholas Pavlidis, University of Ioannina, Greece

*Correspondence:

Ming Chen chenm229@mail.sysu.edu.cn Shuzhong Yao yszlfy@163.com

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

Received: 22 March 2022 Accepted: 30 May 2022 Published: 27 June 2022

Citation:

Zhang Y, Cui Q, Xu M, Liu D, Yao S and Chen M (2022) Current Advances in PD-1/PD-L1 Blockade in Recurrent Epithelial Ovarian Cancer. Front. Immunol. 13:901772. doi: 10.3389/fimmu.2022.901772 Immunotherapies have revolutionized the treatment of a variety of cancers. Epithelial ovarian cancer is the most lethal gynecologic malignancy, and the rate of advanced tumor progression or recurrence is as high as 80%. Current salvage strategies for patients with recurrent ovarian cancer are rarely curative. Recurrent ovarian cancer is a "cold tumor", predominantly due to a lack of tumor antigens and an immunosuppressive tumor microenvironment. In trials testing programmed death-1 (PD-1)/programmed death ligand 1 (PD-L1) blockade as a monotherapy, the response rate was only 8.0-22.2%. In this review, we illustrate the status of cold tumors in ovarian cancer and summarize the existing clinical trials investigating PD-1/PD-L1 blockade in recurrent ovarian cancer. Increasing numbers of immunotherapy combination trials have been set up to improve the response rate of EOC. The current preclinical and clinical development of immunotherapy combination therapy to convert an immune cold tumor into a hot tumor and their underlying mechanisms are also reviewed. The combination of anti-PD-1/PD-L1 with other immunomodulatory drugs or therapies, such as chemotherapy, antiangiogenic therapies, poly (ADP-ribose) polymerase inhibitors, adoptive cell therapy, and oncolytic therapy, could be beneficial. Further efforts are merited to transfer these results to a broader clinical application.

Keywords: programmed death-1 (PD-1), programmed death ligand 1 (PD-L1), immunotherapy, recurrent ovarian cancer, cold tumor

INTRODUCTION

Epithelial ovarian cancer (EOC) is a common gynecological cancer and the most lethal gynecological malignancy among women (1). Most patients are diagnosed with advanced stages of this disease (stage III-IV), and the rate of tumor progression or recurrence is as high as 80% (2). The standard treatment for EOC consists of primary debulking surgery (PDS) and platinum plus taxane chemotherapy. Patients with apparently unresectable tumors by imaging study or laparoscopic inspection or patients with low performance status (PS) or medical complications often receive neoadjuvant chemotherapy (NACT), followed by interval debulking surgery (IDS) and subsequent chemotherapy as an alternative choice (3). However, the recurrence of advanced EOC is as high as 70%, with very short overall survival (OS) (4, 5). Therefore, current strategies for

1

recurrent EOC are rarely curative. New treatment strategies are in demand (6, 7). Immunotherapy will be an attractive treatment in the future.

Checkpoint inhibitor immunotherapy (CPI) has undergone impressive development in recent years. Patients with melanoma, bladder cancer, non-small-cell lung cancer, kidney cancer and colon cancer have reached remarkable long-term survival via immunotherapy (8). Among the different types of cancer immunotherapy, immune checkpoint blockade has had the most promising impact. Several antibodies have been approved for therapeutically targeting the programmed cell death 1 (PD1)-PD1 ligand 1 (PD-L1) axis, T-cell immunoglobulin mucin-3 (TIM-3), or cytotoxic T lymphocyte (CTL)-associated antigen-4 (CTLA-4) (9, 10). Moreover, a large number of monoclonal antibodies (MABs) or small molecules targeting other putative immune checkpoints (such as LAG3, TIGIT, B7H3, CD39, CD73 and adenosine A2A receptor), disrupting negative regulation between tumor cells and T cells, or myeloid cells and T cells, are in clinical and preclinical development (11). Among them, anti-PD-1/PD-L1 antibodies have been the most well studied and evaluated. PD-L1 (B7-H1, CD274) is a 290 aa type I transmembrane protein encoded by the Cd274 gene on mouse chromosome 19 and human chromosome (12). Blocking the interaction between PD-1 and PD-L1 could reverse and/or prevent the exhaustion of tumor-specific T lymphocytes, promoting the surveillance and destruction of tumor cells (Figure 1). Many studies have revealed that recurrent EOC is a "cold tumor" (13) that always has an immunosuppressive tumor microenvironment (TME) with few tumor-associated antigens (TAAs) or tumor-specific antigens (TSAs, or neoantigens), resulting in insufficient recognition and eradication of cancer by the immune system. Therefore, only a small subpopulation of EOC patients benefit from PD-1/PD-L1 blockade (14). There is a great demand to improve the efficacy of PD-1/PD-L1 inhibitors in recurrent EOC patients.

In this review, we briefly introduce the characteristics of the TME in EOC and then provide an update of the results of clinical trials of PD-1/PD-L1 blockade therapy in recurrent EOC. Furthermore, the advancement of therapeutic strategies to turn cold tumors into hot tumors will be discussed.

LITERATURE REVIEW

A comprehensive literature review was conducted using PubMed, Scopus, Web of Science and other sources to identify articles exploring the development of immunotherapy in recurrent EOC, according to the published guidance on narrative reviews (15). We used search terms of "immunotherapy", "gynecological cancer", "recurrent ovarian cancer", "refractory ovarian cancer", "PD-1/PD-L1 inhibitor", and "blockade" in different combinations. Search terms were used as key words and as MeSH terms to maximize the output from the literature. Only available full-text articles in English published until September 2021 were included. Additional exclusion criteria were studies on case reports, commentaries and studies not concerning PD-1/PD-L1 inhibitors. The reference lists of the selected articles were reviewed to identify additional articles meeting the eligibility criteria. The database search resulted in 4,179 articles. Finally, after assessing the full-text articles for eligibility and screening of the reference lists, a total of 141 full-text articles were included in the present review (**Figure 2**).

COLD TUMORS: TME OF EOC

EOC is classified as a cold tumor and is one of the most difficult malignancies to treat, with an extremely high recurrence rate and a low survival rate. EOC is categorized into four phases based on the International Federation of Gynecology and Obstetrics (FIGO) staging criteria. Early-stage (FIGO stage I-II) EOC can be successfully cured by conventional therapies, including surgery and chemotherapy. In contrast, most advanced EOC patients die predominantly due to relapse and chemoresistance (16). Immunotherapy has opened a new era of cancer treatment, achieving satisfactory efficacy across many tumor types. However, immunotherapy for recurrent EOC has not been approved by the US Food and Drug Administration (FDA) due to the lower efficacy of immunotherapy for this cold tumor. The reason mainly lies in two aspects: 1) a lack of tumor antigens and 2) an immunosuppressive TME.

Tumor antigens have two primary categories: TAAs (relatively restricted to tumor cells) and TSAs (unique to tumor cells) (17). TAAs are antigens present on tumor cells at elevated levels but are also expressed on normal cells at a lower level. TAA peptides fused with human leukocyte antigens (HLAs) can be identified by T somatic cells. In EOC, more than thirty TAAs have been reported to date (18, 19). Research has shown that in the epithelial tissues of the respiratory and reproductive organs, ovarian TAAs are weakly expressed, but in tumor cells, they are always overexpressed (20). CA125 and human epididymis protein 4 (HE4) are two typical TAAs in ovarian tumors, although the diagnostic discrimination value of these markers remains suboptimal due to low sensitivity for early-stage disease (21). In contrast, TSAs, also called tumor neoantigens, can be presented by cancer cells. They contain mutated amino acid sequences generated from genomic perturbations. Such accumulation and preservation of mutations is an important mechanism for cancer development. These mutant peptides presented by HLA class I or class II molecules can prime T cells. However, the mutational burden and the potential of neoantigen expression differ according to cancer types. EOC has a lower tumor mutational burden (TMB) with a mean value of 5.3 mutations/Mb, while the TMB-high category is defined as >10 mutations/Mb. A lack of both TSA and TAA indicates the poor immunogenicity of ovarian tumors. Moreover, a solid tumor comprises heterogeneous populations of tumor cells, in which cancer stem cells (CSCs) can further hamper antigen presentation by reducing the expression of HLA class I molecules (22, 23). CSCs have more potential to escape cytotoxic T-cell killing than other tumor cell populations, providing another resistance against immunotherapy for EOC.



In addition to poor immunogenicity, an immunosuppressive TME also contributes to immunotherapy resistance. Cold tumors have a noninflamed TME. Its characteristics can be summarized as low infiltration of CD8+ T cells and CD4+ T cells as well as increased expression of regulatory T cells (Tregs) and PD-L1+ myeloid cells (24, 25). In cold tumors, immunosuppressive cells, including cancer-associated fibroblasts (CAFs), Tregs and myeloid-derived suppressor cells (MDSCs), are highly active, while the immune response executors CTLs are inactive (Figure 3) (26, 27). The lack of effector T cells on a neoplasm leads to a limited patient response to immunotherapy (28). Due to being trapped in the stroma of the tumor or within the peritumoral tissue, CTLs cannot reach the central area (29). Moreover, CAFs also prohibit the immersion of CTLs into the neoplasm and promote a cumulative process of Treg and MDSC into tumors at the center of cancer (30). This status also inhibits the recruitment of dendritic cells (DCs) (31). Furthermore, multiple cell populations, such as MDSCs, macrophages and CSCs, secrete immunosuppressive cytokines, such as transforming growth factor- β (TGF- β), interleukin (IL)-10 and IL-4. These factors can inhibit T-/natural killer (NK) cell activation (32, 33). Nonetheless, a number of abnormalities inside the solid tumor, including abnormal vessels, hypoxia, and an acidic pH, can modulate the interactions among tumor cells, stromal cells, and immune cells and further suppress the immune environment during cancer initiation, progression, and metastasis (34).

Based on the presence or absence of tumor-infiltrating lymphocytes (TIL) and PD-L1 expression in the TME, one immune response classification into four groups has been proposed. The stratification includes type I (PD-L1 positive with TILs driving adaptive immune resistance), type II (PD-L1 negative with no TIL indicating immune ignorance), type III (PD-L1 positive with no TIL indicating intrinsic induction), and type IV (PD-L1 negative with TIL indicating the role of other suppressors in promoting immune tolerance) (28, 35). In advanced melanoma, for which immunotherapy data are the most mature, approximately 38% of patients present with a type I tumor microenvironment and are considered the group that largely benefits from single-agent anti-PD-1/L1 blockade (36). Webb et al. adopted this classification in various histotypes of EOC and found it an important determinant of clinical response (Table 1) (37). In HGSC, the Type I pattern (PD-L1⁺/CD8⁺) was most common, followed by the Type IV pattern and the Type II and III patterns. In contrast, the Type IV pattern was more common in the other histological subtypes of EOC. Among all the patients who received standard chemotherapy, the Type I cases of HGSC had the best prognosis (HR 0.2191), suggesting that the immunological benefits associated with adaptive resistance could extend to conventional treatments as well. Moreover, the status of TIL infiltration and PD-L1 expression, or the transition between different types, could be modulated by treatments, such as chemotherapy. Hao et al. calculated an immunological score based on the transcriptomics of 2203 advanced EOC samples from a public dataset. The immune



score system selected 69 marker genes and 7 antigen-presenting genes for specific immune cell subtypes. Increased immune scores and cytotoxic activity were found in postchemotherapy tumors compared with their correlated prechemotherapy samples. Patients with high immune scores had a better survival and response to immunotherapy than those with low scores (38). Therefore, the conversion of cold tumors to hot tumors is feasible if EOC has been properly treated. The combined use of immunotherapies and other treatments is promising.

POSSIBLE THERAPEUTIC APPROACHES IN EOC THAT COULD TRANSFORM A COLD INTO A HOT TUMOR

Currently, six PD-1/PD-L1 blockades have been approved by the FDA, including antibodies targeting PD-1 (nivolumab,

pembrolizumab, and cemiplimab) or PD-L1 (atezolizumab, avelumab, and durvalumab) (**Table 2**). Recent clinical trials observed satisfactory survival results of PD-1/PD-L1 inhibitors in melanoma, non-small-cell lung cancer (NSCLC), head and neck cancer, and cutaneous squamous cell carcinoma (39–42). In contrast, in many recurrent solid tumors, the traditional treatment is dismal. For example, metastatic castration-resistant prostate cancer (mCRPC) is resistant to androgen deprivation therapy and is related to poor prognosis and limited therapeutic options (43). Recurrent EOC is the same case. However, in these patients, PD-1/PD-L1 inhibitors still do not seem to provide ideal results (44–46). As a single treatment, immune checkpoint inhibition (ICI) has a low response rate of only 10–35% (47).

Like most solid tumors at the advanced stage, recurrent or refractory EOC is always a cold tumor with a mild response to ICI immunotherapies and remains the most fatal gynecological cancer (48). **Table 3** summarizes the clinical studies investigating



migrate from the edge toward the center of tumor, making the core of tumor immunologically hot.

PD-1/PD-L1 inhibitors in recurrent EOC (49–63). In trials testing immune CPIs as monotherapy, the observed recurrent EOC response rate was 8.0-22.2%. As far as concerned, the KEYNOTE-100 study of phase II was the largest study of a single-agent immune checkpoint for recurrent EOC (51). In this study, 376 patients with recurrent EOC were enrolled. Pembrolizumab (200 mg) was given intravenously every 3 weeks. The overall response rate (ORR) was 8.0%, and the median progression-free survival (PFS) was 2.1 months.

To improve response rates, different combinations with anti-PD1-/PD-L1, including chemotherapy to release tumor antigens, antiangiogenic drugs to accelerate T-cell movement into the tumor, and improved lymph node effector T-cell priming and activation by anti-CTLA-4, DNA damage agents such as poly (adenosine diphosphate-ribose) polymerase inhibitors (PARPis) or vaccines to enhance T-cell reactivity against neoantigens, have been used (**Figure 4**) (64). Here, we summarize the latest developments in ICI combinations for recurrent EOC treatment. Furthermore, novel strategies that potentially strengthen immunotherapies for ovarian tumors are discussed.

Chemotherapy

Studies in recent decades have demonstrated that chemotherapy has distinct effects on antitumor immunity (65). Immunogenic cell death might be promoted, and the immunogenicity of tumor cells could be increased under the effects of chemotherapy on tumor immunity. However, as the amount of neoantigen is very low, such an impact on the immunogenicity brought by chemotherapy might be insignificant (66).

TABLE 1 | The immune subtype distribution in different histotypes of ovarian cancer [based on Webb et al. (37)].

Histotype	Percentage of different tumor microenvironment type (%)					
	Type I: Adaptive immune resistance TIL &PD-L1 (+)	Type II: Immunological ignorance TIL &PD-L1 (-)	Type III: Intrinsic induction TIL (-) & PD-L1 (+)	Type IV: Tolerance TIL (+) & PD-L1 (-)		
High grade serous cancer	57.4	5.1	0	37.4		
Low grade serous cancer	0	9.1	0	90.9		
Clear cell cancer	16.2	30.2	0	53.5		
Endometrioid cancer	22.4	14.4	1.6	61.6		
Mucinous cancer	26.7	16.7	0	56.7		

TILs, tumor-infiltrating lymphocytes; PD-L1, programmed death ligand 1.

TABLE 2 | Six PD-1/PD-L1 inhibitors approved by the Food and Drug Administration (FDA).

Drug	Inhibited immune checkpoint
Atezolizumab	PD-L1
Avelumab	PD-L1
Durvalumab	PD-L1
Cemiplimab	PD-1
Nivolumab	PD-1
Pembrolizumab	PD-1

PD-1, programmed death-1 receptor; PD-L1, programmed death ligand 1.

PD-1/PD-L1 regimens	Trial number	Phase	Treatment	Numbers of OC patients	Enrolled patients	Survival results	Reference
Single regimen	NCT00729664	1	BMS-936559	17	Recurrent EOC	PR:6% (1/17; duration 1.3+ months.); SD:18% (3/17; duration 24+ weeks.)	(49)
	UMIN000005714	2	Nivolumab	20	Platinum-resistant EOC	CR: 10% (2/20, duration 11+ months); PR: 5% (1/20, duration 11+ months); SD: 30% (6/20, duration 11+ months)	(50)
	NCT02674061/ KEYNOTE-100	2	Pembrolizumab	376	Recurrent EOC	ORR: 8.0%; Median PFS: 2.1 months	(51)
	NCT02054806/ KEYNOTE-028	1b	Pembrolizumab	26	PD-L1–positive advanced metastatic ovarian cancer	ORR: 11.5%; Median PFS: 1.9 months; Median OS: 13.8 months	(52)
	NCT01772004/ JAVELIN Solid Tumor Trial	1b	Avelumab	125	Recurrent or refractory EOC	ORR: 9.6%; Median PFS: 2.6 months; Median OS: 11.2 months	(53)
	NCT01375842	1	Atezolizumab	12	Recurrent EOC	ORR: 22.2%; Median PFS: 2.9 months; Median OS:11.3 months	(54)
Combined with CTLA-4	NCT02498600/ NRG GY003	2	Arm 1: Nivolumab; Arm 2: Nivolumab + Ipilimumab	100	Persistent or recurrent EOC; PFI< 6 months	ORR: 12.2% vs 31.4%; Median PFS: 2 vs 3.9 months; Median OS: 21.8 vs. 28.8 months	(55)
Combined with chemotherapy	NCT02580058/ JAVELIN Ovarian 200	3	Arm 1: Avelumab; Arm 2: PLD; Arm 3: Avelumab + PLD	566	Platinum-resistant/ refractory EOC	ORR: 4% vs. 4% vs. 13%; DCR: 33% VS. 49% VS. 57%; Median PFS: 1.9 vs. 3.5 vs. 3.7 months; Median OS: 11.8vs. 13.1 vs. 15.7 months	(56)
	NCT02865811	2	Pembrolizumab and pegylated liposomal doxorubicin	26	Platinum- resistant/ refractory EOC	CBR: 52.2%; ORR: 26.1%	(57)
	NCT02440425	2	Weekly paclitaxel + pembrolizumab	37	Recurrent EOC platinum- resistant EOC	ORR: 51.4%; DCR: 86.50%; Median PFS: 7.6 months; Median OS:13.4 months	(58)
Combined with PARPi	NCT02657889/ TOPACIO/ KEYNOTE-162	1/2	Pembrolizumab + Niraparib	60	Recurrent EOC	ORR: 18%; DCR: 65%; Median PFS: 3.4 months	(59)
	NCT02484404	2	Arm 1: Durvalumab + Olaparib; Arm 2: Durvalumab + Cediranib	19	Recurrent or metastatic EOC	ORR: 17% vs. 50%	(60)
	NCT02734004/ MEDIOLA	1/2	Olaparib× 4w, then Olaparib + Durvalumab	34	gBRCAm platinum- sensitive relapsed	ORR: 63%; DCR: 81%	(61)
Combined with VEGFi ± chemotherapy	NCT02853318	2	Pembrolizumab + Bevacizumab + Oral metronomic cyclophosphamide	40	Recurrent platinum- sensitive/resistant/ refractory	ORR: 47.5%; Median PFS:10 months	(62)
	NCT02873962	2	Bevacizumab + Nivolumab	38	Recurrent platinum- sensitive/resistant	ORR: 28.9%; DCR: 34.2%; Median PES: 8.1 months	(63)

PR, partial response; SD, stable disease; CR, complete response; ORR, objective response rate; PFS, progression-free survival; OS, overall survival; CBR, clinical benefit rate; DCR, duration of response.



Immunogenic dead tumor cells generated from chemotherapy release TSA and/or TAA along with damageassociated molecular patterns (DAMPs), which actively stimulate innate immune receptors and recruit immune cells into the TME (67). For example, cancer cell lines treated with anthracyclines could expose calreticulin on their surface during their immunogenic cell death. Such changes help DCs engulf cancer cells and promote tumor antigen presentation and tumorspecific CTL responses. Similarly, a standard dose of gemcitabine could increase the levels of class I or class II MHC molecules, thus augmenting the immune system response (68). Taxane and cyclophosphamide deplete MDSCs and Treg cells, respectively, reducing the inhibition of T-cell infiltration into tumors (69, 70). Dying tumor cells can also stimulate a Toll-like receptor (TLR) 3 dependent, cancer cell-autonomous type I interferon reaction and subsequently produce C-X-C chemokine ligand (CXCL) 10, leading memory T cells to the tumor bed (71, 72).

Meanwhile, chemotherapeutic drugs can induce a general immune response via off-target effects (73). 5-Fluorouracil could deplete tumor MDSCs, while cyclophosphamide could reduce Treg cells and promote the translocation of immune-stimulatory bacteria (74–76). Joalland et al. reported that the adoptive transfer of immune effector cells, consisting of allogeneic human V γ 9V δ 2T lymphocytes, is an effective goal for tracing and determining the spread of tumors in organisms. The results showed that chemotherapy and immunotherapy could be used together to improve the antitumor impacts (77). The impact between chemotherapy and immunotherapy is mutual. In fact, the immune status inside tumors also affects the efficacy of chemotherapy. Hot tumors are most likely more susceptible to chemotherapy than cold tumors (78). For example, tumorassociated macrophages (TAMs) are closely related to the progression and chemoresistance of EOC. The main tumorpromoting function of M2-like TAMs directly induces the invasion ability and chemoresistance of EOC cells by triggering its prosurvival signaling pathway (79). Paclitaxel and carboplatin have been the standard chemotherapy regimens for EOC for decades, both of which can induce apoptosis in tumor cells.

In summary, chemotherapy has an inhibitory effect on malignant tumors and participates in human immune system adjustment. Obvious survival benefits from the combination of platinum chemotherapy with pembrolizumab have been observed in a recent trial of lung cancer (80). The success of anti-PD-1 therapy combined with chemotherapy in the treatment of metastatic NSCLC demonstrated the advantages of this dual approach. It is hopeful that chemotherapy could improve the immunotherapy effects in EOC via tumor immune microenvironment modification. A preclinical model of EOC confirmed that the number of activated T cells and DCs and the expression of cytotoxic factors can be increased by combining chemotherapy (paclitaxel + carboplatin) and immunotherapy (anti-IL-10, 2' 3'-cGAMP + anti-PD-L1) (81). To evaluate the impact of ICI on follow-up therapy, Liu conducted a retrospective study that recruited 79 patients with recurrent EOC. This result suggested that ICI could improve the subsequent chemotherapy outcome. The most commonly used cytotoxic drugs included taxane, platinum, and pegylated

liposomal doxorubicin. The median OS of 18.3 (95% CI, 11.8-22.7) months from initiation of first treatment after ICI is promising compared to the inferior outcomes of other recent studies (82). In another clinical trial, Wehnham et al. evaluated 37 patients with platinum-resistant EOC treated with weekly paclitaxel (80 mg/m²) and pembrolizumab (200 mg q3 w); the ORR was 51%, and the 6-month PFS was 52% (58). Lee's phase II trial showed that the ORR and median PFS of the combination of pembrolizumab with pegylated liposomal doxorubicin in platinum-resistant EOC could be improved compared to PLD or anti-PD-1/PD-L1 agents alone in the past (57). The study observed 12 patients who achieved clinical benefit with a clinical benefit response (CBR) of 52.2% (95% confidence interval (CI), 30.6-73.2%). There were five partial responses (PRs) (21.7%) and one complete remission (CR) (4.3%), and the ORR was 26.1%. Six patients had stable disease (SD) lasting at least twenty-four weeks. However, there was no significant correlation between PD-L1 staining and objective response, PD-L1 staining and TIL score, or TIL score and objective response. To date, the largest trial with reported data in platinum-resistant ovarian cancer is the JAVELIN Ovarian 200 trial, a three-arm randomized phase III trial that enrolled 566 patients. Their results showed that neither avelumab plus pegylated liposomal doxorubicin (PLD) nor avelumab alone significantly improved progression-free survival or OS versus PLD. However, retrospective subgroup analysis including 442 tumor samples revealed that patients with tumors expressing PD-L1 may benefit from avelumab + PLD. PD-L1+ patients who received avelumab combination therapy had a trend toward prolonged PFS and OS (PFS: 3.7 vs. 3 months, P = 0.0143; OS: 15.7 vs. 13.1 months, P = 0.2082) (56).

These studies suggest that the synergy of cytotoxic therapy and PD-1/PD-L1 blockade may be beneficial and feasible in the treatment of EOC in selected patients. Biomarker analysis including PD-L1 and CD8 expression might predict benefit with chemotherapy and immunotherapy treatment in ovarian cancer.

Antiangiogenic Therapies

Vascular abnormality is one distinguishing feature of many solid tumors, including EOC. Angiogenesis and immunosuppression have a close connection (83). Angiogenesis facilitates immune escape, which is involved in nonpathological tissue repair as a physiological mechanism promoting cancer development (84). Many proangiogenic molecules can suppress the immune system, such as antigen presentation, T-cell trafficking, T-cell priming and T-cell tumor infiltration (85). Thus, antiangiogenic drugs would be helpful in improving the antitumor immune response. The interactions of anti-angiogenesis and immune systems include 1) direct effects when binding to their cognate receptors expressed by immune cells; 2) indirect effects when they induce changes in endothelial cell protein expression; and 3) indirect physical effects by promoting vascular normalization or reducing angiogenesis (86).

In advanced-stage melanoma, vascular endothelial growth factor (VEGF) is elevated, which is associated with negative immune effects, such as impaired dendritic cell function. It was

also linked with both elevated and decreased T helper 2 (Th2) cytokines. These effects were found to result in suppression of effective antitumour immunity (87). Patients with higher levels of serum VEGF were found to have worse OS with ipilimumab treatment, which provided a rationale for targeting VEGF (88). Compared with vaccination alone, antiangiogenic therapy combined with vaccination or adoptive cell transfer increased T cells in the tumor, thereby enhancing anticancer activity (89). Moreover, the normalization of blood vessels was related to the increase in CD4+ T helper type 1-cell (Th1) cell accumulation and anticancer activity (90). Nevertheless, VEGF can deplete T cells by upregulating the expression of PD-1 on T cells (91). Cytotoxic effects were observed in the colorectal cancer model CT26 mice treated with PD-1 blockade and anti-VEGF drugs. The tumor growth reduction rate of the experimental group was as high as 75% compared with that of the control group. In the same way, in mice with tumors derived from colon cancer C26 cell injection, the combined use of anti-VEGFR2 and anti-PD-1 MABs enhanced the effects of tumor growth inhibition (92). In EOC, Zhang et al. found that PD-L1 and two angiogenesisrelated proteins (VEGF and Semaphorin4D) were overexpressed in cisplatin-resistant EOC patients. In vitro studies demonstrated the synergistic effects of atezolizumab and bevacizumab in inhibiting the proliferation and metastasis of cisplatin-resistant EOC cell lines (93). This result provides an additional methodology for the treatment of cisplatin-resistant recurrent EOC.

Liu et al. conducted a phase 2 study to evaluate the activity of combined nivolumab and bevacizumab in women with relapsed ovarian cancer. The ORR was 40.0% (19.1%-64.0%) in platinumsensitive participants and 16.7% (95% CI 3.6%-41.4%) in platinum-resistant participants. Thus, it is possible that bevacizumab activity is higher in platinum-sensitive patients. Interestingly, better response rates were observed in patients with PD-L1-negative tumors in this study, suggesting that PD-L1 expression unreliable in predicting the response to immunotherapy activity (59). Similarly, Zsiros et al. reported one open-label, single-arm phase 2 cohort study with 40 recurrent EOC patients enrolled. Thirty patients (75.0%) had platinum-resistant disease, and 10 (25.0%) had platinumsensitive disease (63). The regimen was intravenous pembrolizumab 200 mg, bevacizumab 15 mg/kg every 3 weeks, cyclophosphamide 50 mg orally once daily at the same time. The ORRs in platinum-resistant and platinum-sensitive patients were 43.3% and 60.0%, respectively, with a total ORR of 47.5%. When patients were analyzed based on the number of prior chemotherapy lines, patients with no more than 3 prior chemotherapy lines had a significantly longer PFS. This result revealed that earlier introduction of immunotherapy in the EOC may lead to greater PFS benefit. Moreover, several ongoing trials are carried out to evaluate the combined effects of antiangiogenic and anti-PD-1/PD-L1 agents. In the recent GOG 3015/ENGOT OV-39/IMagyn050 trial, patients with FIGO stage III and macroscopic residual disease who underwent primary major surgery or those with stage IV disease were enrolled in the trial. The participants received chemotherapeutic strategies of paclitaxel, carboplatin and bevacizumab, then with maintenance

of bevacizumab. Subsequently, they were randomized to receive an atezolizumab 1200 mg dose every 3 weeks or placebo during the chemotherapy and maintenance phases. A total of 1301 patients were recruited. In patients receiving atezolizumab vs. placebo, the median PFS was 19.5 vs. 18.4 months (P = 0.28) in the intention-to-treat population and 20.8 vs. 18.5 months (P = 0.038) in the PD-L1-positive population. In the midterm (immature) OS results, atezolizumab had no obvious effect. However, the effects of combined immunotherapy and bevacizumab in EOC still need to be further explored (94).

PARPi

PARPis are one of the most representative new therapies for EOC, targeting the DNA repair fragility of tumor cells. In homologous recombination deficiency (HRD) tumors, doublestrand breaks cannot be accurately repaired in the presence of PARPis (95). Tumor cells are destroyed in the accumulation of unrepaired DNA damage. Among the PARP inhibitors, olaparib, niraparib, and rucaparib trap PARP approximately 100-fold more efficiently than veliparib, while talazoparib appears to be the most potent PARP trapper investigated thus far. Increased PARP trapping is associated with high myelosuppression, which results in variation of the recommended doses across PARP inhibitors (96). Since PARPis could increase the burden of neoantigens and activate DNA damage-associated innate immune pathways in tumor cells, the strategies of combining PARPis and checkpoint inhibition will probably be feasible. This idea is supported by Jiao's study (97). They found that PD-L1 overexpression in breast cancer cell lines and animal models after PARPi treatment, as PARPi, inactivated GSK3 β. The overexpression of PD-L1 reduced the effects of PARPi. The application of PD-L1 blockade again improved the T-cell killing effects of PARPi. This combined therapy significantly improved the therapeutic effect in vivo compared to using each drug separately. Appleton's research examined the effectiveness of using three-dimensional (3D) ellipsoids to detect patientspecific immune-related activities in EOC (98). They found that pembrolizumab increased the secretion of cytokines in a T-cell-dependent way, enhanced the cytotoxicity of olaparib, and reduced the viability of the spheroids. The combination of durvalumab and olaparib could kill cancer cells through a synergistic effect. This work demonstrates the effectiveness of ICIs in combination with PARPis in a preclinical setting.

Several phase II trials have already yielded encouraging results of PARPi and immunotherapy combinations in recurrent EOC patients. The MEDIOLA trial demonstrated promising antitumour activity and safety of the combination of olaparib and durvalumab compared to monotherapy studies (62). The study was carried out on 34 germline breast cancer susceptibility gene (BRCA)-mutated platinum-sensitive recurrent EOC patients. They found that the ORR and 12week DCR of the combination of olaparib and durvalumab in these patients were 72% and 81%, respectively. The median PFS was 8.2 months (95% CI 4.6–11.8), and the median OS was 21.5 months (95% CI 16.2–25.7). TOPACIO/KEYNOTE-162 was a single-arm phase 1/2 clinical trial that enrolled patients regardless of BRCA gene status. Patients with advanced triplenegative breast cancer (TNBC) or recurrent EOC received niraparib and pembrolizumab. The ORR of the EOC cohort (n =62 patients) was 18% (90% CI, 11-29%), and the disease control rate (DCR) was 65% (90% CI, 54-75%) (60). Note that PD-L1 expression was not related to the treatment effects. Participants with only one prior cytotoxic therapy had a better response rate. The most common treatment-related adverse events (AEs) were fatigue, nausea, iron deficiency anemia and severe constipation. In addition, Färkkilä et al. found that mutation feature 3 reflected defective homologous recombination DNA repair and had a better immune score (99). The results of immunogenomic analysis of tumor samples from the TOPACIO study indicated that a positive immune score could predict the efficacy of the niraparib/ pembrolizumab combination. Exhausted CD8+ T cells could be predictive of the response to the niraparib/pembrolizumab combination (100). The clinical data strongly support the strategy of dual targeting PARP and PD-1.

Adoptive Cell Therapy

The elimination of tumor cells depends largely on T cells. In recent years, T-cell adoptive immunotherapy has been developing rapidly. Adoptive T-cell therapy includes the autologous or allogeneic transplantation of TILs or genetically modified T cells with chimeric antigen receptors (CARs) or tumor-specific T-cell receptors (101). This personalized strategy is particularly promising for cancer patients who do not respond well to conventional therapies.

Adoptive cell therapy with TILs has been proven effective in many cancers, especially malignant melanoma, with ORRs ranging from 40-70% (102-104). TIL therapy demonstrates the effects of eliminating solid tumors with high mutational burden malignancies such as melanoma. Studies of melanoma have found that TILs are now considered an abundant source of effector T cells, which exhibit tumor recognition. The TILs isolated from the patient's tumor tissue are reinfused after in vitro activation and expansion, which represents the ultimate personalized therapy. In recent decades, TIL-based adoptive cell therapy in EOC has been extensively studied. Many studies suggest that the presence of the preferential intraepithelial location of TILs is a good predictor for EOC. Aoki et al. published the first trial of TIL therapy using TILs expanded in IL-2 for advanced or recurrent EOC cases in 1991 (105). Five out of 7 patients showed clinical benefits after receiving TILs without chemotherapy. Meanwhile, 9 out of 10 patients showed clinical benefits after receiving chemotherapy before TIL infusion. This study demonstrated the effects of TIL therapy in EOC for the first time. Another study in Japan evaluated the clinical benefits of TIL infusion with combination chemotherapy for EOC patients after primary debulking surgery. Patients receiving TILs had better OS than those whose TILs did not expand to sufficient numbers. Their results suggested that the adoptive transfer of TILs probably induced immunoactivation of cellular immunity and enhanced natural killer activity in EOC (106). One pilot study enrolled six patients with progressive platinum-resistant metastatic EOC. The participants received chemotherapy, followed by TIL infusion and intravenous IL-2. The treatment was well tolerated, and the toxicities were durable. Four patients

had SD for three months, and two patients had SD for five months, with five patients with the target lesion size decreasing. Antitumor reactivity was observed in TIL infusion products from five patients, but no antitumor reactivity was detectable in peripheral blood lymphocytes collected after treatment (107). In addition, some experiments showed the fusion of CPI and adoptive T-cell transfer (ACT) in EOC patients. In one trial, six patients with advanced metastatic high-grade serous EOC were treated with immunotherapy including ipilimumab followed by surgery to obtain TILs and infusion of REP-TILs (TILs expanded in the rapid expansion protocol), low-dose IL-2 and nivolumab. One patient had PR, and the other five had SD for 12 months (108).

CAR-T-cell therapy is another representative of adoptive Tcell immunotherapy. It has the advantages of high specificity, not being limited by MHC, and long-term efficacy after infusion. CAR-T cells are a combination of single-chain variable fragments (scFvs) created by gene transduction that recognize tumor-associated antigens and activation motifs of T cells (109). CAR-T cells have been shown to have a good therapeutic effect on hematologic cancer (110). Recently, Xu et al. described a method that involved a bispecific CAR-T targeting two different B-cell maturation antigen (BCMA) epitopes, combining two camel-derived antigen-binding domains in the construction of a single CAR in patients with multiple myeloma. They achieved the highest CR rate of 76% (13/17) (111). Compared to blood tumors, solid tumors respond poorly to CAR-T-cell therapy. However, several studies with small samples have confirmed the effects of CAR-T cells in EOC. The surface antigens targeted by CARs are mainly proteins and glycolipids (112). In CAR-T-cell therapy for recurrent EOC, the most common target antigens consist of Mucin-16 (MUC-16), mesothelin, human epidermal growth factor receptor-2 (HER-2) and folate receptor-alpha (FR- α). MUC-16 is overexpressed in more than 80% of EOCs (113). In vitro experiments showed that MUC-16-CAR-T cells have specific killing effects on MUC-16⁺ EOC cells. Intravenous or intraperitoneal administration of MUC-16-CAR-T cells can inhibit the growth and development of cancer cells and mouse tumor-bearing solid models (114). Mesothelin is a glycoprotein anchored to the plasma membrane according to the phosphatidylinositol region (GPI). It also rises in the pleura, peritoneum, pericardium, mesothelium and EOC cells (115). Beatty et al. reported two malignant pleural mesothelioma patients who received adoptive transfer of mRNA CAR-T cells that target mesothelin (CARTmeso cells) (116). After intravenous injection, CAR T cells existed temporarily in the blood and moved to cancer regions. The treatment demonstrated clinical and laboratory benefits of antitumor activity and no overt off-tumor on-target toxicity. Oberg developed the bispecific antibody [(HER-2)2xCD16] in the tribody format, which could redirect CD16-expressing $\gamma\delta$ T lymphocytes and NK cells to the tumor-associated cell surface antigen HER-2 to improve their cytotoxic antitumor activity. In HER2-expressing cancer cells, compared with trastuzumab, tribody[(HER-2) 2xcd16] was better at inducing $\gamma\delta$ T-cell- and NK-cell-mediated lysis, including pancreatic ductal adenocarcinoma (PDAC), breast

cancer and EOC (117). However, most CAR-T-cell treatments in EOC are still in the experimental stage. The central issue hindering development was that CAR-T cells could not persist and expand in vivo for more than a few days (118). Efforts to overcome this problem by increasing the dose have failed and have greatly restricted the application of CAR-T cells in clinical practice (119). To obtain a durable clinical response to cell-based gene therapy, permanent transgene expression is indispensable. Murine gamma retroviruses and lentiviruses are two vector systems applied in clinical gene therapy that may provide longterm CAR transgene expression (120). Recent data showed that the expression levels of T-cell coinhibitory receptors, such as PD-1 and Tim-3, were upregulated on CAR-T cells (121). The PD-L1/PD-1 pathway was able to directly inactivate CD28 signaling in CAR-T cells and therefore inhibit CAR-T-cell function (122). Taken together, PD-1/PD-L1 inhibitors and CAR-T therapy might exert synergistic antitumor effects.

Compared to CARs binding unprocessed tumor surface antigens without MHC processing, TCR therapy addresses both tumor intracellular and surface antigenic peptides embedded in MHC. TCRs target special antigens to boost the efficacy of anticancer immunotherapy (123). Many TAAs could be used as targets for immunotherapy. These include the cancer testis antigens (e.g., New York Esophageal Squamous Cell Carcinoma-1 (NY-ESO-1)), differentiation antigens (e.g., Melanoma antigen family A-3 (MAGE-A3)), overexpressed oncogenes (e.g., Wilms' Tumor antigen 1 (WT1)), tumor suppressor genes (e.g., Tumor Protein P53(TP53)); and TAAs that are organ-specific antigens that are transiently expressed. Moreover, oncogenic viruses, such as HPV, EBV, and HBV, are promisingly targeting antigens, which could induce eradication to virusinduced cancer cells. TCRs have several structural advantages, such as more subunits in their receptor structure, greater immunoreceptor tyrosine-based activation motifs (ITAMs), less dependence on antigens and more costimulatory receptors (such as CD3, CD4, and CD28) (124). NY-ESO-1 is a cancer testis antigen and a promising target for immunotherapy that is overexpressed in approximately 40% of non-small cell lung, ovarian, and melanoma cancers and in greater than 70% of certain sarcoma subtypes (125, 126). It has been found to be able to trigger spontaneous humoral and cellular immune responses. NY-ESO-1-directed vaccination has obtained some promising clinical results in early phase I/II studies, and adoptive cell therapy has resulted in partial responses (PRs) in 60% of NY-ESO-1-expressing sarcomas in small phase I trials (127). Somaiah et al. conducted a first-in-human, open-label phase I trial to examine the clinical safety and preliminary efficacy of LV305 in patients with advanced sarcoma or other solid tumors expressing NY-ESO-1 (128). Overexpression of NY-ESO-1 in EOC relates to poor clinical outcomes (129). A phase I, openlabel study assessed the safety and in vivo immunogenicity of synthetic overlapping long peptides (OLPs) from NY-ESO-1 in 28 relapsed EOCs. They found that OLP could rapidly induce consistent integrated immune responses with CD8+ and CD4+ antibodies. However, the induction of NY-ESO-1 antibody and NY- ESO-1 CD8+ and CD4+ T-cell responses was similar in

patients with and without NY-ESO-1 tumor expression. Somaiah et al. conducted a first-in-human, open-label phase I trial to examine the clinical safety and preliminary efficacy of LV305 in patients with advanced sarcoma or other solid tumors expressing NY-ESO-1 (128). LV305 is a novel lentiviral-based cancer vaccine that induces the expression of NY-ESO-1 cancer testis antigen in dendritic cells. Patients were enrolled with sarcoma (n = 24), ovarian (n = 8), melanoma (n = 6), and lung cancer (n=1). All the participants received LV305 (1 mL per dose) intradermally once every 3 weeks. The disease control rate was 56.4% in all patients and 62.5% in sarcoma patients. A post hoc exploratory analysis revealed significantly longer OS in patients with increased T-cell clonality in posttreatment peripheral blood mononuclear cells among all patients (P = 0.0116)), indicating that LV305-induced clonal expansion of T-cell anti-NY-ESO-1 responses may be associated with OS. Further development of LV305 is planned in combination with G305 as the combination therapy "CMB305." G305 is a cancer vaccine composed of a fulllength recombinant NY-ESO-1 protein and the synthetic TLR4 agonist glucopyranosyl lipid An in a stable emulsion (130). Studies of CMB305 alone or in combination with the anti-PD-L1 checkpoint inhibitor atezolizumab are ongoing (NCT02609984, NCT02387125).

Although clinical data on the combination of ICB and ACT are currently limited in EOC, the field of immune therapy has the potential to reach better cancer care.

Vaccine-Based Therapy

Ovarian cancer cells affect the immune system by coordinating the molecular signal network exposed or secreted on the surface, causing T-cell exhaustion and DC dysfunction (131). DC dysfunction may still prevent the complete recovery of immune function, although treatment with immune CPIs can reverse tumor-induced T-cell exhaustion. This problem can be solved by inoculation with functional antigen-presenting cells (APCs).

DCs carry out antigen-specific immune responses, phagocytosis, processing and presentation of foreign and pathogen-related antigens on their cell surface (132). Therefore, DCs hold great potential for becoming effective cancer vaccines in the adjuvant setting. Tumor antigens include tumor cell lysate and tumor-associated peptides/ proteins. DCs present tumor-associated antigens to activate T cells by MHCI/II molecules (133). DC vaccines are regarded as a promising strategy. A phase III trial is currently determining the effectiveness of an RNA-loaded autologous DC vaccine as an adjuvant vaccination in treating uveal melanoma (NCT01983748). Although the DC vaccine is generally safe and has extremely low toxicity, it has shown limited clinical benefits thus far and rarely achieves an objective clinical response of no more than 15% in most indications (134). One explanation for the low response rate is that patients with large tumor burdens have an immunosuppressive TME and have immune tolerance to their tumors. Therefore, the number of clinical trials of the combination of DC vaccines with other drugs has increased dramatically.

In one trial, autologous DCs pulsed with HOCl-oxidized autologous tumor lysate (OCDC vaccine) were obtained from patients as a personalized vaccine. Patients received 5 dose injections of DC vaccine every two weeks (\sim 5-10*10⁶ DCs/ dose), followed by maintenance treatment every month until disease progression or vaccine supply exhaustion. Five patients with recurrent EOC (aged 48-63 years) were included in the clinical study. Two subjects (S2 and S3) had no evidence of disease (NED) after recurrence. Three participants (S1, S4 and S5) were admitted to the trial with radiographically measurable tumors. Two of them progressed, and the other one showed a mixed response according to the solid tumor efficacy evaluation specification (RECIST). The durable PFS times were 36 months and 44 months. All the data showed the efficacy of DC vaccine treatment in EOC patients (135).

Rodney Rocconi and colleagues reported another vaccine strategy (136). Whole tumor lysates prepared at primary or interval debulking surgery were used to create tumor-specific EOC vaccines. After randomization, patients received maintenance vaccination (gemogenovatucel-T) or placebo after complete clinical remission. The subgroup analysis showed that BRCA wild-type EOC patients had an outstandingly longer recurrence-free survival than the control group (HR 0.51, 90% CI 0.30-0.88; P=0.020). The 1-year and 2-year OS of vaccinated BRCA wild-type disease patients were significantly longer than those of unvaccinated patients. Autologous tumor vaccines could be a new and personalized maintenance strategy for patients with EOC. This strategy is especially necessary for the overwhelming majority of BRCA wild-type EOC patients. However, the recurrence-free survival rate was not significantly improved (11.5 months in the vaccine group vs. 8.4 months in the placebo group; P=0.078). Nonetheless, one multicenter clinical trial VITALIA will begin (NCT 03905902). In the study, during the induction period, EOC patients will receive a DC vaccine named DCVAC/OVCA or placebo combined with platinumbased chemotherapy, regardless of whether bevacizumab was used. In the maintenance period, patients will be injected with the DC vaccine combined with bevacizumab and PARPi or only supportive treatment (137).

Kadam et al. applied an anti-PD-1 plus CCL21-DC tumor lysate vaccine in a murine lung cancer model. The expression of perforin and granzyme B in the TME, as well as TIL activity, were increased. Mice with treatment-induced tumor eradication developed immunological memory, leading to cancer recurrence-free survival (138). Since PD-1/PD-L1 blockade will increase the amplitude of therapeutic cancer vaccine-mediated activated T-cell responses in the TME, therapeutic cancer vaccines combined with PD-1/PD-L1 immune checkpoint blockade therapy could be a reasonable approach with the potential for cancer-free survival (139).

Oncolytic Therapy

An oncolytic virus (OV) is a virus that exists naturally or can be obtained by recombination. The virus eliminates cancer cells and targets tumor blood vessels through oncolytic action, direct action or immune-mediated action to selectively kill cancer cells and related stromal cells. Clinical trials of viruses in cancer treatment are needed to advance this field (140). In addition to direct and nearby antitumor activity, OVs can induce more powerful, systemic and durable antitumor immunity (141). Tumor cells undergoing apoptosis cause an effective antitumor immune response, releasing TAAs and additional DAMPs. More importantly, the treatment exerts therapeutic effects not only at the local site but also at distant tumor sites. The OVs must retain their immune stimulation ability without toxic factors to be used for treatment. Many viruses have been developed and evaluated in clinical research, including adenovirus, poxvirus, measles virus, herpes simplex virus type1 (HSV-1), poliovirus, Coxsackie virus, Newcastle disease virus (NDV) and reovirus (142).

Havunen established oncolytic adenoviruses coding for human IL-2, tumor necrosis factor- α (TNF- α), or both. According to the results of the study, they found that these viruses showed strong antitumor effects on human tumors in severe combined immune deficiency (SCID) mice. The treated animals were protected from tumor recurrence, which indicated a memory response. They considered that cytokine-armed oncolytic adenoviruses could change the TME to synergize with adoptive cell therapy (143). Santos cultivated fresh tumor cultures ex vivo originating from patients with advanced EOC and confirmed that the oncolytic adenovirus Ad5/3-E2F-D24hTNFa-IRES-hIL2 was adapted to enhance antitumor TIL responsiveness by reconnecting to the ovarian TME (144). Another OV, granulocyte macrophage colony stimulating factor (GM-CSF)-coding oncolytic adenovirus, was given to patients with solid tumors refractory to standard therapies in one clinical trial. Among the 37 EOC patients, treatment was generally well tolerated. Intestinal problems were the most common adverse events. No grade 4 or grade 5 adverse events were found in 12 patients. Only grade 3 adverse events were reported. The whole cancer control rates were 56% and 55% with imaging (RECIST1.1 or PET criteria) and CA125 detection, respectively (145).

Although the expression of PD-L1 is very low in many solid tumors, recent data have shown that oncogenic viruses can induce chronic inflammation and secrete cytokines such as IFN- γ , which upregulate PD-L1 expression in the TME as a means of self-protection (146–148). Therefore, expanding the application of OV and PD-1/PD-L1 inhibitors would be inspiring. Liu et al. developed a vaccinia virus expressing CXCL1, which could enhance T-cell infiltration into the tumor and upregulate the expression of PD-L1. Combined treatment of OV with anti-PD-L1 finally elicited systemic and potent antitumour immunity in tumor models in B6 mice. The combination therapy led to over 40% cure in aggressive models of peritoneal carcinomatosis from colon and ovarian cancers (142).

In summary, adenoviruses could generate new T-cell responses against tumor antigens and neoantigens in preclinical models and clinical trials. Recent preclinical data have shown that the combination of OV and PD-1/PD-L1 inhibitors could work synergistically to exert cytotoxicity to

cancer cells, eliminate immunosuppressive cells and elicit more potent and sustained systemic antitumor immunity. Therefore, expanding the successful application of oncolytic therapy would be significant in recurrent EOC.

T-Cell Immunomodulators

The therapeutic effects of cytokines such as IL-2, IL-7, IL-12, IL-15, IL-21, GM-CSF and interferon- α (IFN- α) have been evaluated in many clinical trials. They regulate the function of tumor-killing T cells in direct or indirect ways (149). One main reason that patients do not benefit from PD-L1 blockade is the

absence of preexisting tumor inflammation (150). Therefore, a strategy to sensitize the response to PD-L1 blockade could include pretreatment with an immunotherapy that induces intratumoral T-cell infiltration. Mice lacking IL2 die from lethal T-cell-driven autoimmunity due to reduced functional Tregs, indicating that IL2 signaling is essential for excessive Tcell activation (151).

IL-2 is a cytokine produced by activated CD4+T cells that activates the proliferation of CD8+T cells and CD4+ Tregs (152). Despite its contradictory effects, IL-2 treatment received FDA approval in 1998. Aldesleukin, recombinant IL-2, achieved an ORR of 16% and a CR rate of 6% in patients with advanced melanoma in the first trial (153). One phase I/II study evaluated the safety and feasibility of autologous DCs and IL-2 in EOC patients with minimal residual disease. The enrolled patients received two subcutaneous injections of DCs derived from autologous monocytes pulsed with autologous tumor lysate and keyhole limpet hemocyanin (KLH) every 4 weeks, 4 months after initial debulking and chemotherapy (154). Then, a low dose of IL-2 (200 mIU) as an immune adjuvant was given for 14 consecutive days. Among the 10 patients, 3 showed CR after DC vaccination for 83, 80.9 and 38.2 months and no disease recurrence. One patient with SD had a tumor that completely disappeared after DC vaccination and continued for 50.8 months without recurrence. Alternative IL-2 structures are under study to improve the effects and attenuate toxicities. NKTR-214 (bempegaldesleukin) is a drug of recombinant IL-2 coupled with 6 releasable polyethylene glycol (PEG) chains. By PEGylating IL-2, the binding ability of the IL2Ra receptor is reduced, and the half-life is longer. Its affinity for Tregs is also reduced (155). Several phase I/II and III trials tested NKTR-214 as a single treatment or combined immune checkpoint blockade in melanoma (NCT03635983, NCT03138889). The results of the phase I/II PIVOT-02 study of NKTR-214 combined with nivolumab (NCT02983045) showed that in 11 patients with advanced melanoma, the DCR was 91%, the ORR was 64%, and the toxicity was mild (156).

Drerup found that IL2 receptor β -selective IL2/anti-IL2 complexes (IL2c) could preferentially stimulate effector T cells in an orthotopic mouse ID8 aggressive ovarian cancer model (157). IL2c obviously lowered the tumor microenvironmental CD8+/Treg ratio and induced a fragile Treg phenotype. The combination of anti-PD-L1 antibody and IL2c could generate complete tumor regression and protective immune memory, which could not be achieved by either monotherapy. These data support the application of IL2c combined with other immunotherapies to enhance antitumor immunity.

IL-15 can effectively promote the activation and proliferation of CD8+T and NK cells. It did not cause capillary leak syndrome in the preclinical model and had acceptable toxicity in the IL-15 subcutaneous injection test (158). *In vivo*, IL-15 has a short halflife time and lacks potent transpresentation by IL-15 receptor α (IL-15R α). N-803 is an IL-15 superagonist developed by ImmunityBio that combines IL-15 with an activating mutation, an IL-15R α sushi domain for trans-presentation, and IgG1-Fc for increased half-life. Experiments showed that N-803 could increase IFN γ -induced CXCL10 secretion and target killing after increasing exposure time (159). Unfortunately, no significant benefits of IL-15 have been observed in clinical studies until now (160). However, some studies found that IL-7 and IL-15 might be better than IL-2 in adoptive cell therapy (161).

CONCLUSIONS

Although changing a cold tumor into a hot tumor remains a great challenge, an increasing number of pioneer studies focusing on the mechanisms of the immunosuppressive microenvironment and PD-1/PD-L1 blockade combination treatment have shed light on a brilliant future of EOC treatment. Our review provides an overview of valuable evidence and significant therapeutic strategies for PD-1/PD-L1 blockade in recurrent EOC. Many studies have demonstrated that PD-1/PD-L1 blockade therapy is a safe treatment in patients with EOC, but monotherapy showed only modest efficacy in recurrent EOC. This may demonstrate the limited immunity and immunosuppressive microenvironment in EOC. Overall, recurrent EOC is a cold tumor. Its conversion into an inflamed tumor will require a prior combination of therapies to induce immune infiltration, followed by immune checkpoint modulators to remove the breaks. To enhance immunity in EOC, combining a priming therapy to increase T-cell responses, including DC-based vaccination, would be rational. Moreover, according to the preclinical and clinical data, immunotherapy combined with chemotherapeutics, antiangiogenic drugs or DNA demethylating

REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global Cancer Statistics 2018: Globocan Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin (2018) 68 (6):394–424. doi: 10.3322/caac.21492
- 2. Arora T, Mullangi S, Lekkala MR. Ovarian Cancer. In: *Statpearls*. Treasure Island (FL: StatPearls Publishing LLC (2021).
- Moschetta M, Boussios S, Rassy E, Samartzis EP, Funingana G, Uccello M. Neoadjuvant Treatment for Newly Diagnosed Advanced Ovarian Cancer: Where do We Stand and Where are We Going? *Ann Transl Med* (2020) 8 (24):1710. doi: 10.21037/atm-20-1683
- Pignata S, CC S, Du Bois A, Harter P, Heitz F. Treatment of Recurrent Ovarian Cancer. Ann Oncol (2017) 28(Suppl 8):viii51-6. doi: 10.1093/ annonc/mdx441
- 5. McMeekin DS, Tillmanns T, Chaudry T, Gold M, Johnson G, Walker J, et al. Timing Isn't Everything: An Analysis of When to Start Salvage

agents can stimulate anticancer immunity by various mechanisms and hence may be beneficial in patients with cold EOC.

Another tremendous challenge impeding the application of immunotherapies in recurrent EOC is the lack of comprehensive knowledge of the cancer-immune interaction parameters. How to use standardized methods to measure the individual immunity parameters is lacking. Additionally, accurate algorithms of treatment depending on different tumor or TME characteristics should be developed to select adequate immunotherapy strategies. To solve these problems, the development of biomarkers to determine these therapies effectively is critical, and biomarker-guided clinical trials will be necessary to tailor these approaches to EOC patients. Tumor genomic profiling with immune profiling will help to provide a more comprehensive understanding of an individual patient's tumor and select the accurate treatment.

AUTHOR CONTRIBUTIONS

SY and MC designed the manuscript. MC, YZ, and QC wrote the manuscript. MC, YZ, MX, and DL drew the figures and tables. SY mainly revised the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This study was funded by the National Natural Science Foundation of China 81602261; CSCO Cancer Research Foundation (Y-sy2018-120); Beijing Kanghua Foundation (KH-2021-LLZX-052). The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

ACKNOWLEDGMENTS

We apologize to those colleagues whose important work could not be cited due to space constraints.

Chemotherapy in Ovarian Cancer. *Gynecol Oncol* (2004) 95(1):157-64. doi: 10.1016/j.ygyno.2004.07.008

- Kuroki L, Guntupalli SR. Treatment of Epithelial Ovarian Cancer. BMJ (2020) 371:m3773. doi: 10.1136/bmj.m3773
- Coleman RL, Spirtos NM, Enserro D, Herzog TJ, Sabbatini P, Armstrong DK, et al. Secondary Surgical Cytoreduction for Recurrent Ovarian Cancer. N Engl J Med (2019) 381(20):1929–39. doi: 10.1056/ NEJMoa1902626
- Haslam A, Prasad V. Estimation of the Percentage of Us Patients With Cancer Who are Eligible for and Respond to Checkpoint Inhibitor Immunotherapy Drugs. JAMA Netw Open (2019) 2(5):e192535. doi: 10.1001/jamanetworkopen.2019.2535
- Mortezaee K. Immune Escape: A Critical Hallmark in Solid Tumors. Life Sci (2020) 258:118110. doi: 10.1016/j.lfs.2020.118110
- Bonnefoy N, Olive D, Vanhove B. [Next Generation of Anti-Immune Checkpoints Antibodies]. Med Sci (Paris) (2019) 35(12):966-74. doi: 10.1051/medsci/2019193

- Kalbasi A, Ribas A. Tumour-Intrinsic Resistance to Immune Checkpoint Blockade. Nat Rev Immunol (2020) 20(1):25–39. doi: 10.1038/s41577-019-0218-4
- Necchi A, Anichini A, Raggi D, Briganti A, Massa S, Lucianò R, et al. Pembrolizumab as Neoadjuvant Therapy Before Radical Cystectomy in Patients With Muscle-Invasive Urothelial Bladder Carcinoma (Pure-01): An Open-Label, Single-Arm, Phase Ii Study. J Clin Oncol (2018) 36 (34):3353–60. doi: 10.1200/jco.18.01148
- Bonaventura P, Shekarian T, Alcazer V, Valladeau-Guilemond J, Valsesia-Wittmann S, Amigorena S, et al. Cold Tumors: A Therapeutic Challenge for Immunotherapy. *Front Immunol* (2019) 10:168. doi: 10.3389/ fimmu.2019.00168
- Liu YL, Zamarin D. Combination Immune Checkpoint Blockade Strategies to Maximize Immune Response in Gynecological Cancers. *Curr Oncol Rep* (2018) 20(12):94. doi: 10.1007/s11912-018-0740-8
- Gasparyan AY, Ayvazyan L, Blackmore H, Kitas GD. Writing a Narrative Biomedical Review: Considerations for Authors, Peer Reviewers, and Editors. *Rheumatol Int* (2011) 31(11):1409–17. doi: 10.1007/s00296-011-1999-3
- Jayson GC, Kohn EC, Kitchener HC, Ledermann JA. Ovarian Cancer. Lancet (2014) 384(9951):1376–88. doi: 10.1016/s0140-6736(13)62146-7
- Finn OJ. Human Tumor Antigens Yesterday, Today, and Tomorrow. Cancer Immunol Res (2017) 5(5):347–54. doi: 10.1158/2326-6066.Cir-17-0112
- Badgwell D, Bast RCJr. Early Detection of Ovarian Cancer. Dis Markers (2007) 23(5-6):397–410. doi: 10.1155/2007/309382
- Bast RCJr., Badgwell D, Lu Z, Marquez R, Rosen D, Liu J, et al. New Tumor Markers: Ca125 and Beyond. *Int J Gynecol Cancer* (2005) 15 Suppl:3, 274– 81. doi: 10.1111/j.1525-1438.2005.00441.x
- Drapkin R, von Horsten HH, Lin Y, Mok SC, Crum CP, Welch WR, et al. Human Epididymis Protein 4 (He4) is a Secreted Glycoprotein That Is Overexpressed by Serous and Endometrioid Ovarian Carcinomas. *Cancer Res* (2005) 65(6):2162–9. doi: 10.1158/0008-5472.Can-04-3924
- Cramer DW, Bast RCJr., Berg CD, Diamandis EP, Godwin AK, Hartge P, et al. Ovarian Cancer Biomarker Performance in Prostate, Lung, Colorectal, and Ovarian Cancer Screening Trial Specimens. *Cancer Prev Res (Phila)* (2011) 4(3):365–74. doi: 10.1158/1940-6207.Capr-10-0195
- Tallerico R, Conti L, Lanzardo S, Sottile R, Garofalo C, Wagner AK, et al. Nk Cells Control Breast Cancer and Related Cancer Stem Cell Hematological Spread. Oncoimmunology (2017) 6(3):e1284718. doi: 10.1080/ 2162402x.2017.1284718
- Di Tomaso T, Mazzoleni S, Wang E, Sovena G, Clavenna D, Franzin A, et al. Immunobiological Characterization of Cancer Stem Cells Isolated From Glioblastoma Patients. *Clin Cancer Res* (2010) 16(3):800–13. doi: 10.1158/ 1078-0432.Ccr-09-2730
- 24. Abiko K, Mandai M, Hamanishi J, Yoshioka Y, Matsumura N, Baba T, et al. Pd-L1 on Tumor Cells is Induced in Ascites and Promotes Peritoneal Dissemination of Ovarian Cancer Through Ctl Dysfunction. *Clin Cancer Res* (2013) 19(6):1363–74. doi: 10.1158/1078-0432.Ccr-12-2199
- Siska PJ, Singer K, Evert K, Renner K, Kreutz M. The Immunological Warburg Effect: Can a Metabolic-Tumor-Stroma Score (Mets) Guide Cancer Immunotherapy? *Immunol Rev* (2020) 295(1):187–202. doi: 10.1111/ imr.12846
- 26. Mortezaee K. Hypoxia Induces Core-to-Edge Transition of Progressive Tumoral Cells: A Critical Review on Differential Yet Corroborative Roles for Hif-1α and Hif-2α. *Life Sci* (2020) 242:117145. doi: 10.1016/ j.lfs.2019.117145
- Najafi M, Mortezaee K, Majidpoor J. Stromal Reprogramming: A Target for Tumor Therapy. Life Sci (2019) 239:117049. doi: 10.1016/j.lfs.2019.117049
- 28. Weed DT, Zilio S, Reis IM, Sargi Z, Abouyared M, Gomez-Fernandez CR, et al. The Reversal of Immune Exclusion Mediated by Tadalafil and an Anti-Tumor Vaccine Also Induces Pdl1 Upregulation in Recurrent Head and Neck Squamous Cell Carcinoma: Interim Analysis of a Phase I Clinical Trial. *Front Immunol* (2019) 10:1206. doi: 10.3389/fimmu.2019.01206
- Galon J, Mlecnik B, Bindea G, Angell HK, Berger A, Lagorce C, et al. Towards the Introduction of the 'Immunoscore' in the Classification of Malignant Tumours. J Pathol (2014) 232(2):199–209. doi: 10.1002/path.4287
- Mlecnik B, Van den Eynde M, Bindea G, Church SE, Vasaturo A, Fredriksen T, et al. Comprehensive Intrametastatic Immune Quantification and Major

Impact of Immunoscore on Survival. J Natl Cancer Inst (2018) 110(4):438. doi: 10.1093/jnci/djx123

- Palucka K. And Banchereau J: Cancer Immunotherapy via Dendritic Cells. Nat Rev Cancer (2012) 12(4):265–77. doi: 10.1038/nrc3258
- 32. Todaro M, Alea MP, Di Stefano AB, Cammareri P, Vermeulen L, Iovino F, et al. Colon Cancer Stem Cells Dictate Tumor Growth and Resist Cell Death by Production of Interleukin-4. *Cell Stem Cell* (2007) 1(4):389–402. doi: 10.1016/j.stem.2007.08.001
- 33. Schatton T, Schütte U, Frank NY, Zhan Q, Hoerning A, Robles SC, et al. Modulation of T-Cell Activation by Malignant Melanoma Initiating Cells. *Cancer Res* (2010) 70(2):697–708. doi: 10.1158/0008-5472.Can-09-1592
- 34. Taleb M, Atabakhshi-Kashi M, Wang Y, Rezvani Alanagh H, Farhadi Sabet Z, Li F, et al. Bifunctional Therapeutic Peptide Assembled Nanoparticles Exerting Improved Activities of Tumor Vessel Normalization and Immune Checkpoint Inhibition. *Adv Healthc Mater* (2021) 10(12):e2100051. doi: 10.1002/adhm.202100051
- Teng MW, Ngiow SF, Ribas A, Smyth MJ. Classifying Cancers Based on T-Cell Infiltration and Pd-L1. *Cancer Res* (2015) 75(11):2139–45. doi: 10.1158/ 0008-5472.Can-15-0255
- Taube JM, Klein A, Brahmer JR, Xu H, Pan X, Kim JH, et al. Association of Pd-1, Pd-1 Ligands, and Other Features of the Tumor Immune Microenvironment With Response to Anti-Pd-1 Therapy. *Clin Cancer Res* (2014) 20(19):5064–74. doi: 10.1158/1078-0432.Ccr-13-3271
- Webb JR, Milne K, Kroeger DR, Nelson BH. Pd-L1 Expression is Associated With Tumor-Infiltrating T Cells and Favorable Prognosis in High-Grade Serous Ovarian Cancer. *Gynecol Oncol* (2016) 141(2):293–302. doi: 10.1016/ j.ygyno.2016.03.008
- Hao D, Liu J, Chen M, Li J, Wang L, Li X, et al. Immunogenomic Analyses of Advanced Serous Ovarian Cancer Reveal Immune Score is a Strong Prognostic Factor and an Indicator of Chemosensitivity. *Clin Cancer Res* (2018) 24(15):3560–71. doi: 10.1158/1078-0432.Ccr-17-3862
- Franklin C, Livingstone E, Roesch A, Schilling B, Schadendorf D. Immunotherapy in Melanoma: Recent Advances and Future Directions. *Eur J Surg Oncol* (2017) 43(3):604–11. doi: 10.1016/j.ejso.2016.07.145
- Majem M, Juan O, Insa A, Reguart N, Trigo JM, Carcereny E, et al. Seom Clinical Guidelines for the Treatment of Non-Small Cell Lung Cancer (2018). *Clin Transl Oncol* (2019) 21(1):3–17. doi: 10.1007/s12094-018-1978-1
- 41. Cohen EEW, Bell RB, Bifulco CB, Burtness B, Gillison ML, Harrington KJ, et al. The Society for Immunotherapy of Cancer Consensus Statement on Immunotherapy for the Treatment of Squamous Cell Carcinoma of the Head and Neck (Hnscc). J Immunother Cancer (2019) 7(1):184. doi: 10.1186/s40425-019-0662-5
- 42. Ferris RL, Licitra L, Fayette J, Even C, Blumenschein GJr., Harrington KJ, et al. Nivolumab in Patients With Recurrent or Metastatic Squamous Cell Carcinoma of the Head and Neck: Efficacy and Safety in Checkmate 141 by Prior Cetuximab Use. *Clin Cancer Res* (2019) 25(17):5221–30. doi: 10.1158/ 1078-0432.Ccr-18-3944
- Ghose A, Moschetta M, Pappas-Gogos G, Sheriff M, Boussios S. Genetic Aberrations of DNA Repair Pathways in Prostate Cancer: Translation to the Clinic. Int J Mol Sci (2021) 22(18):9783. doi: 10.3390/ijms22189783
- 44. Handa S, Hans B, Goel S, Bashorun HO, Dovey Z, Tewari A. Immunotherapy in Prostate Cancer: Current State and Future Perspectives. *Ther Adv Urol* (2020) 12:1756287220951404. doi: 10.1177/ 1756287220951404
- Fan CA, Reader J, Roque DM. Review of Immune Therapies Targeting Ovarian Cancer. Curr Treat Options Oncol (2018) 19(12):74. doi: 10.1007/ s11864-018-0584-3
- Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, et al. Understanding the Tumor Immune Microenvironment (Time) for Effective Therapy. *Nat Med* (2018) 24(5):541–50. doi: 10.1038/s41591-018-0014-x
- Majidpoor J, Mortezaee K. The Efficacy of Pd-1/Pd-L1 Blockade in Cold Cancers and Future Perspectives. *Clin Immunol* (2021) 226:108707. doi: 10.1016/j.clim.2021.108707
- 48. Hendry S, Salgado R, Gevaert T, Russell PA, John T, Thapa B, et al. Assessing Tumor-Infiltrating Lymphocytes in Solid Tumors: A Practical Review for Pathologists and Proposal for a Standardized Method From the International Immuno-Oncology Biomarkers Working Group: Part 2: Tils

in Melanoma, Gastrointestinal Tract Carcinomas, non-Small Cell Lung Carcinoma and Mesothelioma, Endometrial and Ovarian Carcinomas, Squamous Cell Carcinoma of the Head and Neck, Genitourinary Carcinomas, and Primary Brain Tumors. *Adv Anat Pathol* (2017) 24 (6):311–35. doi: 10.1097/pap.00000000000161

- Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, et al. Safety and Activity of Anti-Pd-L1 Antibody in Patients With Advanced Cancer. N Engl J Med (2012) 366(26):2455-65. doi: 10.1056/ NEJMoa1200694
- Hamanishi J, Mandai M, Ikeda T, Minami M, Kawaguchi A, Murayama T, et al. Safety and Antitumor Activity of Anti-Pd-1 Antibody, Nivolumab, in Patients With Platinum-Resistant Ovarian Cancer. J Clin Oncol (2015) 33 (34):4015–22. doi: 10.1200/jco.2015.62.3397
- Matulonis UA, Shapira-Frommer R, Santin AD, Lisyanskaya AS, Pignata S, Vergote I, et al. Antitumor Activity and Safety of Pembrolizumab in Patients With Advanced Recurrent Ovarian Cancer: Results From the Phase Ii Keynote-100 Study. Ann Oncol (2019) 30(7):1080–7. doi: 10.1093/annonc/mdz135
- Varga A, Piha-Paul S, Ott PA, Mehnert JM, Berton-Rigaud D, Morosky A, et al. Pembrolizumab in Patients With Programmed Death Ligand 1-Positive Advanced Ovarian Cancer: Analysis of Keynote-028. *Gynecol Oncol* (2019) 152(2):243–50. doi: 10.1016/j.ygyno.2018.11.017
- 53. Disis ML, Taylor MH, Kelly K, Beck JT, Gordon M, Moore KM, et al. Efficacy and Safety of Avelumab for Patients With Recurrent or Refractory Ovarian Cancer: Phase 1b Results From the Javelin Solid Tumor Trial. *JAMA Oncol* (2019) 5(3):393–401. doi: 10.1001/jamaoncol.2018.6258
- 54. Liu JF, Gordon M, Veneris J, Braiteh F, Balmanoukian A, Eder JP, et al. Safety, Clinical Activity and Biomarker Assessments of Atezolizumab From a Phase I Study in Advanced/Recurrent Ovarian and Uterine Cancers. *Gynecol Oncol* (2019) 154(2):314–22. doi: 10.1016/j.ygyno.2019.05.021
- 55. Zamarin D, Burger RA, Sill MW, Powell DJJr., Lankes HA, Feldman MD, et al. Randomized Phase Ii Trial of Nivolumab Versus Nivolumab and Ipilimumab for Recurrent or Persistent Ovarian Cancer: An Nrg Oncology Study. J Clin Oncol (2020) 38(16):1814–23. doi: 10.1200/jco.19.02059
- 56. Pujade-Lauraine E, Fujiwara K, Ledermann JA, Oza AM, Kristeleit R, Ray-Coquard IL, et al. Avelumab Alone or in Combination With Chemotherapy Versus Chemotherapy Alone in Platinum-Resistant or Platinum-Refractory Ovarian Cancer (Javelin Ovarian 200): An Open-Label, Three-Arm, Randomised, Phase 3 Study. *Lancet Oncol* (2021) 22(7):1034–46. doi: 10.1016/s1470-2045(21)00216-3
- Lee EK, Xiong N, Cheng SC, Barry WT, Penson RT, Konstantinopoulos PA, et al. Combined Pembrolizumab and Pegylated Liposomal Doxorubicin in Platinum Resistant Ovarian Cancer: A Phase 2 Clinical Trial. *Gynecol Oncol* (2020) 159(1):72–8. doi: 10.1016/j.ygyno.2020.07.028
- Wenham RM, Apte SM, Shahzad MM, Lee JK, Dorman D, Chon HS. Phase Ii Trial of Dose Dense (Weekly) Paclitaxel With Pembrolizumab (Mk-3475) in Platinum-Resistant Recurrent Ovarian Cancer. J Clin Oncol (2016) 34 (Suppl 15):TPS5612–TPS5612. doi: 10.1200/JCO.2016.34.15_suppl.TPS5612
- Liu JF, Herold C, Gray KP, Penson RT, Horowitz N, Konstantinopoulos PA, et al. Assessment of Combined Nivolumab and Bevacizumab in Relapsed Ovarian Cancer: A Phase 2 Clinical Trial. *JAMA Oncol* (2019) 5(12):1731–8. doi: 10.1001/jamaoncol.2019.3343
- 60. Konstantinopoulos PA, Waggoner S, Vidal GA, Mita M, Moroney JW, Holloway R, et al. Single-Arm Phases 1 and 2 Trial of Niraparib in Combination With Pembrolizumab in Patients With Recurrent Platinum-Resistant Ovarian Carcinoma. JAMA Oncol (2019) 5(8):1141–9. doi: 10.1001/jamaoncol.2019.1048
- 61. Lee JM, Cimino-Mathews A, Peer CJ, Zimmer A, Lipkowitz S, Annunziata CM, et al. Safety and Clinical Activity of the Programmed Death-Ligand 1 Inhibitor Durvalumab in Combination With Poly (Adp-Ribose) Polymerase Inhibitor Olaparib or Vascular Endothelial Growth Factor Receptor 1-3 Inhibitor Cediranib in Women's Cancers: A Dose-Escalation, Phase I Study. *J Clin Oncol* (2017) 35(19):2193–202. doi: 10.1200/JCO.2016.72.1340
- Domchek SM, Postel-Vinay S, Im SA, Park YH, Delord JP, Italiano A, et al. Olaparib and Durvalumab in Patients With Germline Brca-Mutated Metastatic Breast Cancer (Mediola): An Open-Label, Multicentre, Phase 1/ 2, Basket Study. *Lancet Oncol* (2020) 21(9):1155–64. doi: 10.1016/s1470-2045(20)30324-7

- 63. Zsiros E, Lynam S, Attwood KM, Wang C, Chilakapati S, Gomez EC, et al. Efficacy and Safety of Pembrolizumab in Combination With Bevacizumab and Oral Metronomic Cyclophosphamide in the Treatment of Recurrent Ovarian Cancer: A Phase 2 Nonrandomized Clinical Trial. *JAMA Oncol* (2021) 7(1):78–85. doi: 10.1001/jamaoncol.2020.5945
- Edwards SC, Hoevenaar WHM, Coffelt SB. Emerging Immunotherapies for Metastasis. Br J Cancer (2021) 124(1):37–48. doi: 10.1038/s41416-020-01160-5
- Bracci L, Schiavoni G, Sistigu A, Belardelli F. Immune-Based Mechanisms of Cytotoxic Chemotherapy: Implications for the Design of Novel and Rationale-Based Combined Treatments Against Cancer. *Cell Death Differ* (2014) 21(1):15–25. doi: 10.1038/cdd.2013.67
- 66. McGranahan N, Furness AJ, Rosenthal R, Ramskov S, Lyngaa R, Saini SK, et al. Clonal Neoantigens Elicit T Cell Immunoreactivity and Sensitivity to Immune Checkpoint Blockade. *Science* (2016) 351(6280):1463–9. doi: 10.1126/science.aaf1490
- Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunogenic Cell Death in Cancer and Infectious Disease. *Nat Rev Immunol* (2017) 17(2):97– 111. doi: 10.1038/nri.2016.107
- Galluzzi L, Senovilla L, Zitvogel L, Kroemer G. The Secret Ally: Immunostimulation by Anticancer Drugs. Nat Rev Drug Discovery (2012) 11(3):215–33. doi: 10.1038/nrd3626
- McDonnell AM, Lesterhuis WJ, Khong A, Nowak AK, Lake RA, Currie AJ, et al. Tumor-Infiltrating Dendritic Cells Exhibit Defective Cross-Presentation of Tumor Antigens, But is Reversed by Chemotherapy. *Eur J Immunol* (2015) 45(1):49–59. doi: 10.1002/eji.201444722
- Zheng Y, Dou Y, Duan L, Cong C, Gao A, Lai Q, et al. Using Chemo-Drugs or Irradiation to Break Immune Tolerance and Facilitate Immunotherapy in Solid Cancer. *Cell Immunol* (2015) 294(1):54–9. doi: 10.1016/ j.cellimm.2015.02.003
- Sistigu A, Yamazaki T, Vacchelli E, Chaba K, Enot DP, Adam J, et al. Cancer Cell-Autonomous Contribution of Type I Interferon Signaling to the Efficacy of Chemotherapy. *Nat Med* (2014) 20(11):1301–9. doi: 10.1038/nm.3708
- Vacchelli E, Ma Y, Baracco EE, Sistigu A, Enot DP, Pietrocola F, et al. Chemotherapy-Induced Antitumor Immunity Requires Formyl Peptide Receptor 1. Science (2015) 350(6263):972–8. doi: 10.1126/science.aad0779
- Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunological Effects of Conventional Chemotherapy and Targeted Anticancer Agents. *Cancer Cell* (2015) 28(6):690–714. doi: 10.1016/j.ccell.2015.10.012
- Vincent J, Mignot G, Chalmin F, Ladoire S, Bruchard M, Chevriaux A, et al. 5-Fluorouracil Selectively Kills Tumor-Associated Myeloid-Derived Suppressor Cells Resulting in Enhanced T Cell-Dependent Antitumor Immunity. *Cancer Res* (2010) 70(8):3052–61. doi: 10.1158/0008-5472.Can-09-3690
- 75. Ghiringhelli F, Menard C, Puig PE, Ladoire S, Roux S, Martin F, et al. Metronomic Cyclophosphamide Regimen Selectively Depletes Cd4+Cd25+ Regulatory T Cells and Restores T and Nk Effector Functions in End Stage Cancer Patients. *Cancer Immunol Immunother* (2007) 56(5):641–8. doi: 10.1007/s00262-006-0225-8
- 76. Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillère R, Hannani D, et al. The Intestinal Microbiota Modulates the Anticancer Immune Effects of Cyclophosphamide. *Science* (2013) 342(6161):971–6. doi: 10.1126/ science.1240537
- 77. Joalland N, Lafrance L, Oullier T, Marionneau-Lambot S, Loussouarn D, Jarry U, et al. Combined Chemotherapy and Allogeneic Human Vγ9vδ2 T Lymphocyte-Immunotherapies Efficiently Control the Development of Human Epithelial Ovarian Cancer Cells In Vivo. Oncoimmunology (2019) 8(11):e1649971. doi: 10.1080/2162402x.2019.1649971
- Anitei MG, Zeitoun G, Mlecnik B, Marliot F, Haicheur N, Todosi AM, et al. Prognostic and Predictive Values of the Immunoscore in Patients With Rectal Cancer. *Clin Cancer Res* (2014) 20(7):1891–9. doi: 10.1158/1078-0432.Ccr-13-2830
- Nowak M, Klink M. The Role of Tumor-Associated Macrophages in the Progression and Chemoresistance of Ovarian Cancer. *Cells* (2020) 9(5):1299. doi: 10.3390/cells9051299
- 80. Gandhi L, Rodríguez-Abreu D, Gadgeel S, Esteban E, Felip E, De Angelis F, et al. Pembrolizumab Plus Chemotherapy in Metastatic non-Small-Cell

Lung Cancer. N Engl J Med (2018) 378(22):2078–92. doi: 10.1056/ NEJMoa1801005

- Hartl CA, Bertschi A, Puerto RB, Andresen C, Cheney EM, Mittendorf EA, et al. Combination Therapy Targeting Both Innate and Adaptive Immunity Improves Survival in a Pre-Clinical Model of Ovarian Cancer. *J Immunother Cancer* (2019) 7(1):199. doi: 10.1186/s40425-019-0654-5
- Liu YL, Zhou Q, Iasonos A, Emengo VN, Friedman C, Konner JA, et al. Subsequent Therapies and Survival After Immunotherapy in Recurrent Ovarian Cancer. *Gynecol Oncol* (2019) 155(1):51–7. doi: 10.1016/ j.ygyno.2019.08.006
- Motz GT, Coukos G. The Parallel Lives of Angiogenesis and Immunosuppression: Cancer and Other Tales. *Nat Rev Immunol* (2011) 11(10):702–11. doi: 10.1038/ nri3064
- Jain RK. Normalization of Tumor Vasculature: An Emerging Concept in Antiangiogenic Therapy. Science (2005) 307(5706):58–62. doi: 10.1126/ science.1104819
- Chen DS, Mellman I. Oncology Meets Immunology: The Cancer-Immunity Cycle. *Immunity* (2013) 39(1):1–10. doi: 10.1016/j.immuni.2013.07.012
- Galon J, Bruni D. Approaches to Treat Immune Hot, Altered and Cold Tumours With Combination Immunotherapies. *Nat Rev Drug Discovery* (2019) 18(3):197–218. doi: 10.1038/s41573-018-0007-y
- Revythis A, Shah S, Kutka M, Moschetta M, Ozturk MA, Pappas-Gogos G, et al. Unraveling the Wide Spectrum of Melanoma Biomarkers. *Diagn* (*Basel*) (2021) 11(8):1341. doi: 10.3390/diagnostics11081341
- Yuan J, Zhou J, Dong Z, Tandon S, Kuk D, Panageas KS, et al. Pretreatment Serum Vegf is Associated With Clinical Response and Overall Survival in Advanced Melanoma Patients Treated With Ipilimumab. *Cancer Immunol Res* (2014) 2(2):127–32. doi: 10.1158/2326-6066.Cir-13-0163
- Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, et al. Vascular Normalizing Doses of Antiangiogenic Treatment Reprogram the Immunosuppressive Tumor Microenvironment and Enhance Immunotherapy. *Proc Natl Acad Sci U S A* (2012) 109(43):17561–6. doi: 10.1073/pnas.1215397109
- Tian L, Goldstein A, Wang H, Ching Lo H, Sun Kim I, Welte T, et al. Mutual Regulation of Tumour Vessel Normalization and Immunostimulatory Reprogramming. *Nature* (2017) 544(7649):250–4. doi: 10.1038/nature21724
- Kim CG, Jang M, Kim Y, Leem G, Kim KH, Lee H, et al. Vegf-A Drives Tox-Dependent T Cell Exhaustion in Anti-Pd-1-Resistant Microsatellite Stable Colorectal Cancers. *Sci Immunol* (2019) 4(41):eaay0555. doi: 10.1126/ sciimmunol.aay0555
- 92. Yasuda S, Sho M, Yamato I, Yoshiji H, Wakatsuki K, Nishiwada S, et al. Simultaneous Blockade of Programmed Death 1 and Vascular Endothelial Growth Factor Receptor 2 (Vegfr2) Induces Synergistic Anti-Tumour Effect In Vivo. Clin Exp Immunol (2013) 172(3):500–6. doi: 10.1111/ cei.12069
- 93. Zhang L, Chen Y, Li F, Bao L, Liu W. Atezolizumab and Bevacizumab Attenuate Cisplatin Resistant Ovarian Cancer Cells Progression Synergistically via Suppressing Epithelial-Mesenchymal Transition. Front Immunol (2019) 10:867. doi: 10.3389/fimmu.2019.00867
- 94. Moore KN, Bookman M, Sehouli J, Miller A, Anderson C, Scambia G, et al. Atezolizumab, Bevacizumab, and Chemotherapy for Newly Diagnosed Stage Iii or Iv Ovarian Cancer: Placebo-Controlled Randomized Phase Iii Trial (Imagyn050/Gog 3015/Engot-Ov39). J Clin Oncol (2021) 39(17):1842–55. doi: 10.1200/jco.21.00306
- 95. Lin Q, Liu W, Xu S, Shang H, Li J, Guo Y, et al. Parp Inhibitors as Maintenance Therapy in Newly Diagnosed Advanced Ovarian Cancer: A Meta-Analysis. BJOG (2021) 128(3):485–93. doi: 10.1111/1471-0528.16411
- 96. Boussios S, Karihtala P, Moschetta M, Abson C, Karathanasi A, Zakynthinakis-Kyriakou N, et al. Veliparib in Ovarian Cancer: A New Synthetically Lethal Therapeutic Approach. *Invest New Drugs* (2020) 38 (1):181–93. doi: 10.1007/s10637-019-00867-4
- 97. Jiao S, Xia W, Yamaguchi H, Wei Y, Chen MK, Hsu JM, et al. Parp Inhibitor Upregulates Pd-L1 Expression and Enhances Cancer-Associated Immunosuppression. *Clin Cancer Res* (2017) 23(14):3711–20. doi: 10.1158/1078-0432.Ccr-16-3215
- Appleton KM, Elrod AK, Lassahn KA, Shuford S, Holmes LM, DesRochers TM. Pd-1/Pd-L1 Checkpoint Inhibitors in Combination With Olaparib Display Antitumor Activity in Ovarian Cancer Patient-Derived Three-

Dimensional Spheroid Cultures. *Cancer Immunol Immunother* (2021) 70 (3):843–56. doi: 10.1007/s00262-021-02849-z

- Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, et al. Signatures of Mutational Processes in Human Cancer. *Nature* (2013) 500(7463):415–21. doi: 10.1038/nature12477
- 100. Färkkilä A, Gulhan DC, Casado J, Jacobson CA, Nguyen H, Kochupurakkal B, et al. Immunogenomic Profiling Determines Responses to Combined Parp and Pd-1 Inhibition in Ovarian Cancer. *Nat Commun* (2020) 11(1):1459. doi: 10.1038/s41467-020-15315-8
- 101. Zacharakis N, Chinnasamy H, Black M, Xu H, Lu YC, Zheng Z, et al. Immune Recognition of Somatic Mutations Leading to Complete Durable Regression in Metastatic Breast Cancer. *Nat Med* (2018) 24(6):724–30. doi: 10.1038/s41591-018-0040-8
- 102. Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, et al. Durable Complete Responses in Heavily Pretreated Patients With Metastatic Melanoma Using T-Cell Transfer Immunotherapy. *Clin Cancer Res* (2011) 17(13):4550–7. doi: 10.1158/1078-0432.Ccr-11-0116
- 103. Andersen R, Donia M, Ellebaek E, Borch TH, Kongsted P, Iversen TZ, et al. Long-Lasting Complete Responses in Patients With Metastatic Melanoma After Adoptive Cell Therapy With Tumor-Infiltrating Lymphocytes and an Attenuated Il2 Regimen. *Clin Cancer Res* (2016) 22(15):3734–45. doi: 10.1158/1078-0432.Ccr-15-1879
- 104. Besser MJ, Shapira-Frommer R, Itzhaki O, Treves AJ, Zippel DB, Levy D, et al. Adoptive Transfer of Tumor-Infiltrating Lymphocytes in Patients With Metastatic Melanoma: Intent-To-Treat Analysis and Efficacy After Failure to Prior Immunotherapies. *Clin Cancer Res* (2013) 19(17):4792–800. doi: 10.1158/1078-0432.Ccr-13-0380
- 105. Aoki Y, Takakuwa K, Kodama S, Tanaka K, Takahashi M, Tokunaga A, et al. Use of Adoptive Transfer of Tumor-Infiltrating Lymphocytes Alone or in Combination With Cisplatin-Containing Chemotherapy in Patients With Epithelial Ovarian Cancer. *Cancer Res* (1991) 51(7):1934–9.
- 106. Ikarashi H, Fujita K, Takakuwa K, Kodama S, Tokunaga A, Takahashi T, et al. Immunomodulation in Patients With Epithelial Ovarian Cancer After Adoptive Transfer of Tumor-Infiltrating Lymphocytes. *Cancer Res* (1994) 54 (1):190–6.
- 107. Pedersen M, Westergaard MCW, Milne K, Nielsen M, Borch TH, Poulsen LG, et al. Adoptive Cell Therapy With Tumor-Infiltrating Lymphocytes in Patients With Metastatic Ovarian Cancer: A Pilot Study. *Oncoimmunology* (2018) 7(12):e1502905. doi: 10.1080/2162402x.2018.1502905
- 108. Kverneland AH, Pedersen M, Westergaard MCW, Nielsen M, Borch TH, Olsen LR, et al. Adoptive Cell Therapy in Combination With Checkpoint Inhibitors in Ovarian Cancer. *Oncotarget* (2020) 11(22):2092–105. doi: 10.18632/oncotarget.27604
- 109. Morgan RA, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case Report of a Serious Adverse Event Following the Administration of T Cells Transduced With a Chimeric Antigen Receptor Recognizing Erbb2. *Mol Ther* (2010) 18(4):843–51. doi: 10.1038/mt.2010.24
- 110. Lee DW, Kochenderfer JN, Stetler-Stevenson M, Cui YK, Delbrook C, Feldman SA, et al. T Cells Expressing Cd19 Chimeric Antigen Receptors for Acute Lymphoblastic Leukaemia in Children and Young Adults: A Phase 1 Dose-Escalation Trial. *Lancet* (2015) 385(9967):517–28. doi: 10.1016/ s0140-6736(14)61403-3
- 111. Xu J, Chen LJ, Yang SS, Sun Y, Wu W, Liu YF, et al. Exploratory Trial of a Biepitopic Car T-Targeting B Cell Maturation Antigen in Relapsed/ Refractory Multiple Myeloma. *Proc Natl Acad Sci USA* (2019) 116 (19):9543–51. doi: 10.1073/pnas.1819745116
- 112. Sadelain M, Brentjens R, Rivière I. The Promise and Potential Pitfalls of Chimeric Antigen Receptors. *Curr Opin Immunol* (2009) 21(2):215–23. doi: 10.1016/j.coi.2009.02.009
- 113. Haridas D, Ponnusamy MP, Chugh S, Lakshmanan I, Seshacharyulu P, Batra SK. Muc16: Molecular Analysis and its Functional Implications in Benign and Malignant Conditions. *FASEB J* (2014) 28(10):4183–99. doi: 10.1096/fj.14-257352
- 114. Chekmasova AA, Rao TD, Nikhamin Y, Park KJ, Levine DA, Spriggs DR, et al. Successful Eradication of Established Peritoneal Ovarian Tumors in Scid-Beige Mice Following Adoptive Transfer of T Cells Genetically Targeted to the Muc16 Antigen. *Clin Cancer Res* (2010) 16(14):3594–606. doi: 10.1158/1078-0432.Ccr-10-0192

- 115. Hassan R, Thomas A, Alewine C, Le DT, Jaffee EM, Pastan I. Mesothelin Immunotherapy for Cancer: Ready for Prime Time? J Clin Oncol (2016) 34 (34):4171–9. doi: 10.1200/jco.2016.68.3672
- 116. Beatty GL, Haas AR, Maus MV, Torigian DA, Soulen MC, Plesa G, et al. Mesothelin-Specific Chimeric Antigen Receptor Mrna-Engineered T Cells Induce Anti-Tumor Activity in Solid Malignancies. *Cancer Immunol Res* (2014) 2(2):112–20. doi: 10.1158/2326-6066.Cir-13-0170
- 117. Oberg HH, Kellner C, Gonnermann D, Sebens S, Bauerschlag D, Gramatzki M, et al. Tribody [(Her2)(2)Xcd16] Is More Effective Than Trastuzumab in Enhancing γδ T Cell and Natural Killer Cell Cytotoxicity Against Her2-Expressing Cancer Cells. *Front Immunol* (2018) 9:814. doi: 10.3389/fimmu.2018.00814
- Stone JD, Kranz DM. Role of T Cell Receptor Affinity in the Efficacy and Specificity of Adoptive T Cell Therapies. *Front Immunol* (2013) 4:244. doi: 10.3389/fimmu.2013.00244
- 119. Mensali N, Myhre MR, Dillard P, Pollmann S, Gaudernack G, Kvalheim G, et al. Preclinical Assessment of Transiently Tcr Redirected T Cells for Solid Tumour Immunotherapy. *Cancer Immunol Immunother* (2019) 68(8):1235– 43. doi: 10.1007/s00262-019-02356-2
- 120. Cooray S, Howe SJ, Thrasher AJ. Retrovirus and Lentivirus Vector Design and Methods of Cell Conditioning. *Methods Enzymol* (2012) 507:29–57. doi: 10.1016/b978-0-12-386509-0.00003-x
- 121. Zolov SN, Rietberg SP, Bonifant CL. Programmed Cell Death Protein 1 Activation Preferentially Inhibits Cd28.Car-T Cells. *Cytotherapy* (2018) 20 (10):1259–66. doi: 10.1016/j.jcyt.2018.07.005
- 122. Hui E, Cheung J, Zhu J, Su X, Taylor MJ, Wallweber HA, et al. T Cell Costimulatory Receptor Cd28 Is a Primary Target for Pd-1-Mediated Inhibition. *Science* (2017) 355(6332):1428–33. doi: 10.1126/science.aaf1292
- 123. Zhao Q, Jiang Y, Xiang S, Kaboli PJ, Shen J, Zhao Y, et al. Engineered Tcr-T Cell Immunotherapy in Anticancer Precision Medicine: Pros and Cons. *Front Immunol* (2021) 12:658753. doi: 10.3389/fimmu.2021.658753
- Harris DT, Kranz DM. Adoptive T Cell Therapies: A Comparison of T Cell Receptors and Chimeric Antigen Receptors. *Trends Pharmacol Sci* (2016) 37 (3):220–30. doi: 10.1016/j.tips.2015.11.004
- 125. Albershardt TC, Campbell DJ, Parsons AJ, Slough MM, Ter Meulen J, Berglund P. Lv305, a Dendritic Cell-Targeting Integration-Deficient Zvex (Tm)-Based Lentiviral Vector Encoding Ny-Eso-1, Induces Potent Anti-Tumor Immune Response. *Mol Ther Oncolytics* (2016) 3:16010. doi: 10.1038/ mto.2016.10
- 126. Park TS, Groh EM, Patel K, Kerkar SP, Lee CC, Rosenberg SA. Expression of Mage-a and Ny-Eso-1 in Primary and Metastatic Cancers. J Immunother (2016) 39(1):1–7. doi: 10.1097/cji.00000000000101
- 127. Thomas R, Al-Khadairi G, Roelands J, Hendrickx W, Dermime S, Bedognetti D, et al. Ny-Eso-1 Based Immunotherapy of Cancer: Current Perspectives. Front Immunol (2018) 9:947. doi: 10.3389/fimmu.2018.00947
- 128. Somaiah N, Block MS, Kim JW, Shapiro GI, Do KT, Hwu P, et al. First-In-Class, First-in-Human Study Evaluating Lv305, a Dendritic-Cell Tropic Lentiviral Vector, in Sarcoma and Other Solid Tumors Expressing Ny-Eso-1. Clin Cancer Res (2019) 25(19):5808–17. doi: 10.1158/1078-0432.Ccr-19-1025
- 129. Szender JB, Papanicolau-Sengos A, Eng KH, Miliotto AJ, Lugade AA, Gnjatic S, et al. Ny-Eso-1 Expression Predicts an Aggressive Phenotype of Ovarian Cancer. *Gynecol Oncol* (2017) 145(3):420–5. doi: 10.1016/j.ygyno. 2017.03.509
- 130. Mahipal A, Ejadi S, Gnjatic S, Kim-Schulze S, Lu H, Ter Meulen JH, et al. First-In-Human Phase 1 Dose-Escalating Trial of G305 in Patients With Advanced Solid Tumors Expressing Ny-Eso-1. *Cancer Immunol Immunother* (2019) 68(7):1211–22. doi: 10.1007/s00262-019-02331-x
- 131. Huang RY, Francois A, McGray AR, Miliotto A, Odunsi K. Compensatory Upregulation of Pd-1, Lag-3, and Ctla-4 Limits the Efficacy of Single-Agent Checkpoint Blockade in Metastatic Ovarian Cancer. Oncoimmunology (2017) 6(1):e1249561. doi: 10.1080/2162402x.2016.1249561
- Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ, et al. Immunobiology of Dendritic Cells. Annu Rev Immunol (2000) 18:767–811. doi: 10.1146/annurev.immunol.18.1.767
- Chiang CL, Balint K, Coukos G, Kandalaft LE. Potential Approaches for More Successful Dendritic Cell-Based Immunotherapy. *Expert Opin Biol Ther* (2015) 15(4):569–82. doi: 10.1517/14712598.2015.1000298

- 134. Boudewijns S, Westdorp H, Koornstra RH, Aarntzen EH, Schreibelt G, Creemers JH, et al. Immune-Related Adverse Events of Dendritic Cell Vaccination Correlate With Immunologic and Clinical Outcome in Stage Iii and Iv Melanoma Patients. J Immunother (2016) 39(6):241–8. doi: 10.1097/cji.000000000000127
- 135. Chiang CL, Kandalaft LE, Tanyi J, Hagemann AR, Motz GT, Svoronos N, et al. A Dendritic Cell Vaccine Pulsed With Autologous Hypochlorous Acid-Oxidized Ovarian Cancer Lysate Primes Effective Broad Antitumor Immunity: From Bench to Bedside. *Clin Cancer Res* (2013) 19(17):4801– 15. doi: 10.1158/1078-0432.Ccr-13-1185
- 136. Rocconi RP, Grosen EA, Ghamande SA, Chan JK, Barve MA, Oh J, et al. Gemogenovatucel-T (Vigil) Immunotherapy as Maintenance in Frontline Stage Iii/Iv Ovarian Cancer (Vital): A Randomised, Double-Blind, Placebo-Controlled, Phase 2b Trial. *Lancet Oncol* (2020) 21(12):1661–72. doi: 10.1016/s1470-2045(20)30533-7
- SOTIO. Sotio's Dcvac/Ovca Significantly Improves Survival in Patients With Recurrent Ovarian Cancer, 2019. In: SOTIO Biotech (2019).
- Kadam P, Sharma S. Pd-1 Immune Checkpoint Blockade Promotes Therapeutic Cancer Vaccine to Eradicate Lung Cancer. Vaccines (Basel) (2020) 8(2):317. doi: 10.3390/vaccines8020317
- 139. Mougel A, Terme M, Tanchot C. Therapeutic Cancer Vaccine and Combinations With Antiangiogenic Therapies and Immune Checkpoint Blockade. Front Immunol (2019) 10:467. doi: 10.3389/ fimmu.2019.00467
- 140. García-Martínez E, Smith M, Buqué A, Aranda F, de la Peña FA, Ivars A, et al. Trial Watch: Immunostimulation With Recombinant Cytokines for Cancer Therapy. Oncoimmunology (2018) 7(6):e1433982. doi: 10.1080/ 2162402x.2018.1433982
- 141. Kaufman HL, Kohlhapp FJ, Zloza A. Oncolytic Viruses: A New Class of Immunotherapy Drugs. Nat Rev Drug Discovery (2015) 14(9):642–62. doi: 10.1038/nrd4663
- 142. Liu Z, Ravindranathan R, Kalinski P, Guo ZS, Bartlett DL. Rational Combination of Oncolytic Vaccinia Virus and Pd-L1 Blockade Works Synergistically to Enhance Therapeutic Efficacy. *Nat Commun* (2017) 8:14754. doi: 10.1038/ncomms14754
- 143. Havunen R, Siurala M, Sorsa S, Grönberg-Vähä-Koskela S, Behr M, Tähtinen S, et al. Oncolytic Adenoviruses Armed With Tumor Necrosis Factor Alpha and Interleukin-2 Enable Successful Adoptive Cell Therapy. *Mol Ther Oncolytics* (2017) 4:77–86. doi: 10.1016/j.omto. 2016.12.004
- 144. Santos JM, Heiniö C, Cervera-Carrascon V, Quixabeira DCA, Siurala M, Havunen R, et al. Oncolytic Adenovirus Shapes the Ovarian Tumor Microenvironment for Potent Tumor-Infiltrating Lymphocyte Tumor Reactivity. J Immunother Cancer (2020) 8(1):e000188. doi: 10.1136/jitc-2019-000188
- 145. Kanerva A, Koski A, Liikanen I, Oksanen M, Joensuu T, Hemminki O, et al. Case-Control Estimation of the Impact of Oncolytic Adenovirus on the Survival of Patients With Refractory Solid Tumors. *Mol Ther* (2015) 23 (2):321–9. doi: 10.1038/mt.2014.218
- 146. Green MR, Rodig S, Juszczynski P, Ouyang J, Sinha P, O'Donnell E, et al. Constitutive Ap-1 Activity and Ebv Infection Induce Pd-L1 in Hodgkin Lymphomas and Posttransplant Lymphoproliferative Disorders: Implications for Targeted Therapy. *Clin Cancer Res* (2012) 18(6):1611–8. doi: 10.1158/1078-0432.Ccr-11-1942
- 147. Abiko K, Matsumura N, Hamanishi J, Horikawa N, Murakami R, Yamaguchi K, et al. Ifn-γ From Lymphocytes Induces Pd-L1 Expression and Promotes Progression of Ovarian Cancer. Br J Cancer (2015) 112(9):1501–9. doi: 10.1038/bjc.2015.101
- 148. Chen L, Han X. Anti-Pd-1/Pd-L1 Therapy of Human Cancer: Past, Present, and Future. J Clin Invest (2015) 125(9):3384–91. doi: 10.1172/jci80011
- 149. Zhang S, Rabkin SD. The Discovery and Development of Oncolytic Viruses: Are They the Future of Cancer Immunotherapy? *Expert Opin Drug Discovery* (2021) 16(4):391–410. doi: 10.1080/17460441.2021.1850689
- 150. Ribas A, Hu-Lieskovan S. What Does Pd-L1 Positive or Negative Mean? J Exp Med (2016) 213(13):2835–40. doi: 10.1084/jem.20161462
- 151. Sadlack B, Merz H, Schorle H, Schimpl A, Feller AC, Horak I. Ulcerative Colitis-Like Disease in Mice With a Disrupted Interleukin-2 Gene. *Cell* (1993) 75(2):253–61. doi: 10.1016/0092-8674(93)80067-0

- Boyman O, Sprent J. The Role of Interleukin-2 During Homeostasis and Activation of the Immune System. *Nat Rev Immunol* (2012) 12(3):180–90. doi: 10.1038/nri3156
- 153. Atkins MB, Lotze MT, Dutcher JP, Fisher RI, Weiss G, Margolin K, et al. High-Dose Recombinant Interleukin 2 Therapy for Patients With Metastatic Melanoma: Analysis of 270 Patients Treated Between 1985 and 1993. J Clin Oncol (1999) 17(7):2105–16. doi: 10.1200/jco.1999.17.7.2105
- 154. Baek S, Kim YM, Kim SB, Kim CS, Kwon SW, Kim Y, et al. Therapeutic Dc Vaccination With Il-2 as a Consolidation Therapy for Ovarian Cancer Patients: A Phase I/Ii Trial. *Cell Mol Immunol* (2015) 12(1):87–95. doi: 10.1038/cmi.2014.40
- 155. Doberstein SK. Bempegaldesleukin (Nktr-214): A Cd-122-Biased Il-2 Receptor Agonist for Cancer Immunotherapy. Expert Opin Biol Ther (2019) 19(12):1223–8. doi: 10.1080/14712598.2019.1685489
- 156. Ascierto PA, Flaherty K, Goff S. Emerging Strategies in Systemic Therapy for the Treatment of Melanoma. *Am Soc Clin Oncol Educ Book* (2018) 38:751–8. doi: 10.1200/edbk_199047
- 157. Drerup JM, Deng Y, Pandeswara SL, Padrón ÁS, Reyes RM, Zhang X, et al. Cd122-Selective Il2 Complexes Reduce Immunosuppression, Promote Treg Fragility, and Sensitize Tumor Response to Pd-L1 Blockade. *Cancer Res* (2020) 80(22):5063–75. doi: 10.1158/0008-5472.Can-20-0002
- 158. Waldmann TA, Lugli E, Roederer M, Perera LP, Smedley JV, Macallister RP, et al. Safety (Toxicity), Pharmacokinetics, Immunogenicity, and Impact on Elements of the Normal Immune System of Recombinant Human Il-15 in Rhesus Macaques. *Blood* (2011) 117(18):4787–95. doi: 10.1182/blood-2010-10-311456
- 159. Van der Meer JMR, Maas RJA, Guldevall K, Klarenaar K, de Jonge P, Evert JSH, et al. Il-15 Superagonist N-803 Improves Ifnγ Production and Killing of

Leukemia and Ovarian Cancer Cells by Cd34(+) Progenitor-Derived Nk Cells. *Cancer Immunol Immunother* (2021) 70(5):1305–21. doi: 10.1007/ s00262-020-02749-8

- 160. Margolin K, Morishima C, Velcheti V, Miller JS, Lee SM, Silk AW, et al. Phase I Trial of Alt-803, a Novel Recombinant Il15 Complex, in Patients With Advanced Solid Tumors. *Clin Cancer Res* (2018) 24(22):5552–61. doi: 10.1158/1078-0432.Ccr-18-0945
- 161. Kaneko S, Mastaglio S, Bondanza A, Ponzoni M, Sanvito F, Aldrighetti L, et al. Il-7 and Il-15 Allow the Generation of Suicide Gene-Modified Alloreactive Self-Renewing Central Memory Human T Lymphocytes. *Blood* (2009) 113(5):1006–15. doi: 10.1182/blood-2008-05-156059

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zhang, Cui, Xu, Liu, Yao and Chen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

	RV	Continued		
aloosa		NDV	newcastle disease virus	
ACT	adoptive T-cell transfer	NED	no evidence of disease	
BCMA	B-cell maturation antigen	NK	natural killer	
BRCA	Breast Cancer Susceptibility Gene	NYESO-1	New York Esophageal Squamous Cell Carcinoma-1	
CAF	cancer-associated fibroblast	ORR	overall response rate	
CAR	chimeric antigen receptor	OS	overall survival	
CBR	clinical benefit response	OV	oncolytic virus	
CPI	checkpoint inhibitor	PARPi	poly (adenosine diphosphate-ribose) polymerase inhibitor	
CSC	cancer stem cell	PD-1	programmed death protein 1	
CTLA-4	cytotoxic T lymphocyte associated antigen-4	PDAC	pancreatic ductal adenocarcinoma	
CTL	cytotoxic T lymphocyte	PD-L1	programmed death ligand 1	
CXCL	C-X-C chemokine ligand	PFS	progression-free survival	
DAMP	damage-associated molecular pattern	PR	partial response	
DC	dendritic cell	REP-TILs	TILs expanded in the rapid expansion protocol	
DCR	disease control rate	SCID	severe combined immune deficiency	
EOC	epithelial ovarian cancer	SD	stable disease	
FDA	Food and Drug Administration	TAA	tumor-associated antigen	
FR-α	folate receptor-alpha	TGF-β	growth factor-β	
GM-CSF	granulocyte macrophage colony stimulating factor	Th1	T helper type 1-cell	
GPI	phosphatidylinositol region	TIGIT	T cell immunoglobulin and ITIM domain	
HER-2	human epidermal growth factor receptor-2	TILs	tumor-infiltrating lymphocytes	
HLA	human leukocyte antigen	TIM-3	T-cell immunoglobulin mucin-3	
HSV-1	herpes simplex virus type 1	TLR	toll-like receptor	
IFN-α	interferon-α	TMB	tumor mutational burden	
IL	interleukin	TME	tumor microenvironment	
LAG3	Lymphocyte activation gene-3	TNF-α	tumor necrosis factor-α	
MAB	monoclonal antibody	TP53	Tumor Protein P53	
MAGE-A3	Melanoma antigen family A-3	Treg	regulatory T cells	
MDSC	myeloid-derived suppressor cell	TSA	tumor-specific antigen	
MUC-16	Mucin-16	VEGF	vascular endothelial growth factor	
		– WT1	Wilms'Tumor antigen 1	

(Continued)