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### Review

# Diagnostic approach in acute hypoxemic respiratory failure

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### ABSTRACT

Acute hypoxemic respiratory failure (AHRF) is the leading cause of intensive care unit (ICU) admissions. Of patients with AHRF, 40 %–50 % will require invasive mechanical ventilation during their stay in the ICU, and 30 %–80 % will meet the Berlin Criteria for Acute Respiratory Distress Syndrome (ARDS). Rapid identification of the underlying cause of AHRF is necessary before initiating targeted treatment. Almost 10 % of patients with ARDS have no identified classic risk factors however, and the precise cause of AHRF may not be identified in up to 15 % of patients, particularly in cases of immunosuppression. In these patients, a multidisciplinary, comprehensive, and hierarchical diagnostic work-up is mandatory, including a detailed history and physical examination, chest computed tomography, extensive microbiological investigations, bronchoalveolar lavage fluid cytological analysis, immunological tests, and investigation of the possible involvement of pneumotoxic drugs.

### Introduction

Acute hypoxemic respiratory failure (AHRF) is the leading cause of intensive care unit (ICU) admissions. In the multinational LUNG SAFE study,[1] 10% of patients admitted to the ICU met the criteria for acute respiratory distress syndrome (ARDS). Of patients with AHRF, 40 %-50 % will require invasive mechanical ventilation during their stay in the ICU<sup>[2,3]</sup> and 30 %-80% will meet the Berlin Criteria for ARDS. [1,3,4] While symptomatic supportive care of patients with AHRF is required immediately (i.e., admission to the ICU, administration of appropriate ventilatory support, management of associated organ failures), rapid identification of the underlying cause of AHRF is necessary before initiating targeted treatment, and in order to improve outcomes. [5] A diagnosis of ARDS requires that the AHRF is not exclusively or primarily related to cardiogenic pulmonary edema or fluid overload, atelectasis or lung collapse, pleural effusion, or pulmonary embolism. [6] The first step in the diagnostic approach to AHRF is therefore to investigate these conditions during initial assessments. The most common causes of AHRF reported in the FLORALI trial[3] were community-acquired

pneumonia (63.5%), hospital-acquired pneumonia (11.9%), extrapulmonary sepsis (5.2%), and aspiration and near drowning (1.9%). Less common risk factors for ARDS reported in the LUNG SAFE study[1] included non-cardiogenic shock (7.5%), and trauma (4.2%). These should therefore be prioritized for screening so that the cause of AHRF can be rapidly identified in the majority of cases. In rare cases, however, the cause of AHRF remains elusive after an initial diagnostic work-up. [7,8] This occurs more commonly in immunosuppressed patients, where the precise cause of AHRF cannot be identified in up to 15% of cases.<sup>[9]</sup> The management of patients in whom the cause of AHRF cannot be identified is a challenge for the intensivist, who must attempt both diagnosis and specific treatment without a clear diagnosis. Although the prognoses of patients with unidentified causes of AHRF remain a subject of debate, [10-12] it is imperative that a comprehensive diagnostic work-up is performed to ensure that curable conditions are not overlooked. This review summarizes the essential elements of the initial diagnostic work-up, then details the main conditions that should be investigated in the setting of AHRF when no ARDS risk factor has been identified. The review does not consider

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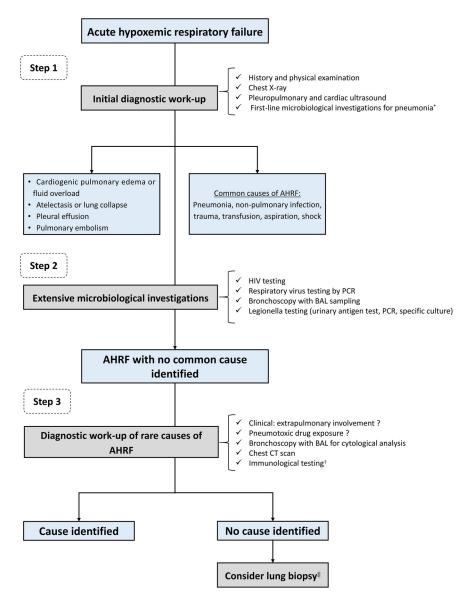


Figure 1. Diagnostic approach for application in patients with AHRF.

AAN: Anti-neutrophil cytoplasmic antibody; BAL: Bronchoalveolar lavage; CCP: Cyclic citrullinated peptide; CT: Computed tomography; ENA: Extractable nuclear antigen; GBM: Glomerular basement membrane; GM-CSF: Granulocyte-macrophage colony-stimulating factor; HIV: Human immunodeficiency virus; PCR: Polymerase chain reaction.

airway, vascular, neuromuscular, or chest wall diseases that lead to AHRF.

## **Initial Diagnostic Work-Up**

A pragmatic diagnostic approach in cases of AHRF is summarized in Figure 1. The first step is to rule out cardiogenic pulmonary edema or fluid overload, atelectasis or lung collapse, pleural effusion, and pulmonary embolism – which are at the forefront of acute respiratory distress. In most cases, a thorough medical history focusing on cardiovascular and thromboembolic comorbidities, a complete physical examination, chest X-ray, and cardiac and pleuropulmonary ultrasound [13,14] are sufficient

to diagnose these conditions. After these conditions have been investigated and ruled out, the second step is to look for the most common causes of AHRF, such as pneumonia, non-pulmonary infection, trauma, transfusion, aspiration, and shock. [6] These causes can be rapidly identified via history taking and physical examination, medical record review, and laboratory tests. In a large multicenter study conducted in France, no common risk factor could be identified in 7.5% of ARDS patients. [11] Strikingly, this figure was confirmed in an ancillary analysis of the LUNG SAFE study, in which approximately 8% of ARDS patients had no ARDS risk factor identified at the time of ARDS recognition. [12] The etiologies of this subgroup can be considered rare causes of AHRF. These patients without identified common

<sup>\*</sup>First-line microbiological investigations for pneumonia include respiratory sample and blood culture, pneumococcal and legionella urinary antigen analysis, and respiratory virus testing via PCR.

<sup>†</sup>Immunological testing includes AAN, anti-ENA panel, rheumatoid factor, anti-CCP antibodies, ANCA, myositis-specific antibodies, and depending on the clinical context, anti-GBM, and anti-GM-CSF antibodies.

<sup>‡</sup>If the patient is invasively ventilated.

Table 1
Main non-infectious causes of AHRF with no common risk factors for ARDS identified.

Rare causes of ARDS	Main entities	Associated radiological patterns	BAL findings	Key element of the diagnostic work-up	Highlighted references
Immune	CTDs: Idiopathic inflammatory myositis (ASS, dermatomyositis with anti-MDA-5 antibodies), SLE, rheumatoid arthritis, systemic sclerosis, and mixed CTD Small-vessel vasculitides: microscopic polyangiitis, granulomatosis with polyangiitis, anti-GBM antibody disease	Alveolar consolidations, ground glass opacities with/ without signs of lung fibrosis*  Ground glass opacities sparing the lung periphery, alveolar consolidation Nodules/excavations	Non-specific  DAH <sup>†</sup> Lymphocytic alveolitis DAH <sup>†</sup>	Immunological testing: AAN, anti-ENA panel, rheumatoid factor, anti-CCP antibodies, ANCA, myositis-specific antibodies with/without anti-GBM, and anti-GM-CSF antibodies	[34]
Drug induced	Amiodarone methotrexate (Methotrexate), bleomycin (Bleomycin) Targeted therapy, immune-checkpoint inhibitors	Ground glass or alveolar opacities with/without signs of lung fibrosis*	Lymphocytes >15 % Eosinophils >15 % Neutrophilic alveolitis	Medical history https://www.pneumotox. com/drug/index/	Pneumotox® [64,66]
Malignant	Lymphangitic carcinomatosis Lepidic adenocarcinoma Hematological malignancies (AML)	Mediastinal lymphadenopathies and/or lung nodules or masses Alveolar diffuses opacities	Lymphocytes >15 % Tumoral cells Rare presence of blasts	Histopathology  Bone marrow exam	[10] [73]
Idiopathic	Acute exacerbations of idiopathic pulmonary fibrosis or other underlying ILD	Alveolar consolidations, ground glass opacities and signs of lung fibrosis*	Neutrophils alveolitis (>30 %)	Medical history, CT-scan pattern	[75]
	Acute hypereosinophilic pneumonia	Alveolar opacities and pleural effusion	Eosinophils >25 %	Eosinophilia (blood sample, BAL fluid)	
	Cryptogenic OP	Inverted halo sign, patchy, migrating infiltrates and alveolar consolidations Centrilobular nodules, mosaic	Mixed alveolitis with >15 % lymphocytes	CT-scan pattern, BAL fluid findings	
	Acute hypersensitivity pneumonitis Acute interstitial pneumonia	air-trapping, and upper lobe distribution Bilateral patchy ground-glass opacities with/without bilateral consolidations, patchy ground-glass opacities	Lymphocytes >15 % Neutrophilic alveolitis (>30 %)	-	

<sup>\*</sup> Signs of lung fibrosis: reticulations, traction bronchiectasis, honeycombing.

causes of AHRF require a very specific etiological diagnostic approach, which we describe in detail below.

## **Rare Causes of AHRF**

It is impossible to provide an exhaustive list of non-infectious rare causes of AHRF, but pragmatically they can be divided into four categories; immune, drug-induced, malignant, and idiopathic. A pragmatic list of the main entities, based on expert opinion, [8,11,12] is provided in Table 1. Identifying situations that suggest a rare cause of AHRF is crucial, even if a common cause appears to have been identified after the initial diagnostic work-up. This is especially true because two causes may be associated, for example, an infection may occur in addition to a flare-up of an immune disorder. [15]

Several clinical features may cause clinicians to suspect rare causes of AHRF, including a previous history of immune disease or exposure to a known pneumotoxic drug, a subacute presentation (i.e., onset of respiratory symptoms >7 days but <30 days prior to presentation), the presence of extrapulmonary manifestations, or an unresolved pneumonia with adequate antibiotic treatment and no microbiological documentation.<sup>[8,16]</sup> Importantly, the onset of respiratory symptoms in <7 days, although less suggestive, should not lead clinicians to rule out an unusual cause. Bronchoalveolar lavage (BAL) fluid analysis and chest computed tomography (CT), although often non-specific, can prompt suspicion of a rare cause of AHRF. Certain CT pat-

terns are suggestive of a rare cause of AHRF, including diffuse or patchy ground-glass lung infiltrates, signs of underlying interstitial lung disease (ILD) (e.g., subpleural reticulations, traction bronchiectasis), or pulmonary nodules with or without cavitation. In the absence of clinical, radiological, or BAL cytology evidence suggesting a rare cause of AHRF, comprehensive microbiological investigations are necessary, particularly if symptoms started <7 days before and the patient became febrile.

# Comprehensive microbiological investigations

Microbiological documentation is lacking in 40%-60% of cases of severe community-acquired pneumonia, and should therefore not be the only criterion for ruling out the diagnosis.[17-19] Rapid improvement of AHRF with antibiotic therapy strongly supports an infectious etiology, and may justify delaying further microbiological investigations. However, if there is no improvement despite appropriate antibiotic therapy, further investigation is warranted. The presence of immunosuppression is a key factor guiding additional microbiological testing. Currently, it seems appropriate to perform the following tests in all patients regardless of their immunosuppression status: Legionella testing, human immunodeficiency virus testing, respiratory virus testing by polymerase chain reaction (PCR), and bronchoscopy with BAL sampling. New molecular tools such as the BioFire® FilmArray® Pneumonia Panel (Biomérieux, Marcy l'Etoile, France) may be useful, [20,21] particularly when antibi-

<sup>†</sup> Bloody BAL fluid (i.e., alveolar red blood cells) and/or hemosiderin-laden macrophages on Perls staining with a Golde score >100 or >60 % alveolar siderophages. AAN: Antinuclear antibodies; AHRF: Acute hypoxemic respiratory failure; AML: Acute myeloid leukemia; ANCA: Antineutrophil cytoplasmic antibodies; ARDS: Acute respiratory distress syndrome; ASS: Antisynthetase syndrome;; BAL: Bronchoalveolar lavage; CCP: Cyclic citrullinated peptide; CT: Computed tomography; CTD: Connective tissue disease; DAD: Diffuse alveolar damage; DAH: Diffuse alveolar hemorrhage; ENA: Extractable nuclear antigen; GBM: Glomerular basement membrane; GM-CSF: Granulocyte-macrophage colony-stimulating factor; ILD: Interstitial lung disease.; MDA-5: Anti-melanoma differentiation-associated protein 5; NSIP: Non-specific interstitial pneumonia; OP: Organizing pneumonia; SLE: Systemic lupus erythematosus; UIP: Usual interstitial pneumonia.

otic therapy has been used prior to conventional microbiological culture. Aggressive testing for Legionella is essential, using a combination of assessments including repeated urinary antigen tests at intervals ≥24 h, and specific respiratory sample culture. Legionella PCR (specific or multiplex PCR), although not recommended, [22] can reportedly yield better diagnostic results than urinary antigen testing, [23] though it should be combined with other tests. Pathogens known to be difficult to culture, such as atypical bacteria (e.g., Mycoplasma pneumoniae, Chlamydia pneumoniae, Chlamydia psittaci, Coxiella burnetii, Leptospira spp.) should also be investigated via PCR or specific serological tests, depending on the time elapsed since the onset of symptoms. Non-respiratory viruses (e.g., Hantavirus, viral hemorrhagic fevers) can also cause pneumonia, but these are often associated with multisystemic manifestations. [24] In such cases, PCR or serology may also be helpful. In immunosuppressed patients, the search for pathogens should be more comprehensive, and tailored to the type of immunosuppression. [25] Fungal (e.g., pneumocystosis, aspergillosis, emerging invasive fungal infections) and viral (e.g., cytomegalovirus, herpes simplex virus) evaluations need to be comprehensive, incorporate the analysis of chest CT patterns, and potentially include bronchoscopy, BAL analysis, tests for specific biomarkers, and PCR tests. Lastly, metagenomics, an emerging unbiased pan-pathogen molecular approach, may also be of use in these challenging diagnostic situations. [26-28]

### Immune causes

Pulmonary involvement in immune diseases may present as AHRF requiring ICU admission. Typically, there are associated suggestive extrapulmonary manifestations (e.g., skin, kidney, joint, muscle), but some patients may present with isolated pulmonary involvement.

# Connective tissue diseases (CTDs)

CTDs are the most common autoimmune disorders seen in the ICU, particularly in cases of AHRF. [8,29-31] The term rapidly progressive interstitial lung disease (RP-ILD), the definition of which is not universally agreed upon, is often used to describe AHRF associated with CTD.[32] In such patients, the pulmonary manifestations of idiopathic inflammatory myopathies (IIMs), a group of rare CTDs characterized by varying degrees of muscle inflammation, [33] are emblematic due to their frequency during disease progression and the severity of respiratory impairment.[34] The two most commonly implicated subtypes of IIMs are antisynthetase syndrome and dermatomyositis with antimelanoma differentiation-associated protein 5 (MDA-5) antibodies. Antisynthetase syndrome has been characterized as the association of inflammatory myositis, ILD, arthritis, Raynaud's phenomenon, and mechanic's hands with the presence of antiaminoacyl-tRNA-synthetase (ARS) antibodies.[35] Other symptoms overlapping with Sjögren's syndrome and systemic sclerosis are also quite commonly reported. A total of eight ARS antibodies have been identified. Although anti-JO1 antibodies are the most common, forms with anti-PL7, anti-PL12, and anti-EJ antibodies appear to be associated with greater respiratory severity. [35-38] In addition to ARS antibodies, anti-MDA-5 antibodies were discovered in 2005 in patients with clinically amyopathic dermatomyositis (CADM), thus they were initially

termed anti-CADM antibodies.[39] Dermatomyositis with anti-MDA-5 antibodies is most commonly characterized by respiratory involvement, skin involvement, Raynaud's syndrome, and joint involvement.[40,41] Muscle involvement is variable and often mild when present, justifying the term amyopathic dermatomyositis. Lung involvement is sometimes isolated, making diagnosis more difficult. The reputation of the disease is based on the frequency and severity of lung involvement.<sup>[34,42,43]</sup> Vuillard et al., [34] analyzed a multicenter cohort of patients with anti-synthetase or anti-MDA-5 antibodies presenting with AHRF requiring ICU admission, and reported that the latter had dramatically higher in-hospital mortality than the former (29 % vs. 84%) despite aggressive management. In cases requiring extracorporeal membrane oxygenation, mortality can reach 100% without emergency lung transplantation. [42] Pulmonary involvement in other IIMs is rarer, more chronic, and less aggressive, and therefore much less common in ICU patients. [33] Other CTDs that occasionally present with ILD leading to AHRF include rheumatoid arthritis, scleroderma, primary Sjögren's syndrome, and mixed CTD.[31,44] Systemic lupus erythematosus with or without antiphospholipid syndrome – can cause severe and abrupt AHRF associated with diffuse alveolar hemorrhage (DAH) related to pulmonary capillaritis, which is accompanied by acute glomerulonephritis in more than two-thirds of cases and can be the inaugural manifestation of the disease. [31,45] Similarly, antiphospholipid syndrome may present as AHRF, particularly as DAH in cases of catastrophic syndrome<sup>[46]</sup> Other CTDs, including rheumatoid arthritis, systemic sclerosis, and mixed CTD, can also present as AHRF, but this is much rarer and often occurs in the setting of underlying ILD. In this situation, a thorough assessment is essential to rule out alternative diagnoses, particularly infections, before concluding a disease flare-up.

### Vasculitides

AHRF requiring ICU admission may complicate or, in up to 90% of cases, reveal small-vessel vasculitides, [47,48] mostly granulomatosis with polyangiitis and microscopic polyangiitis. Patients may present with isolated acute respiratory failure (approximately one-third of cases), typically related to DAH, or with a pulmonary-renal syndrome (approximately one-third of cases), defined as the association of DAH with rapidly progressive glomerulonephritis. Eosinophilic granulomatosis with polyangiitis can present as a severe exacerbation of asthma, sometimes associated with pulmonary infiltrates. [49] Early identification of patients with vasculitis flares is necessary to initiate targeted treatment and prevent worsening of acute respiratory failure and renal function deterioration, which can lead to renal death. Anti-neutrophil cytoplasmic antibody (ANCA) testing, using high-quality proteinase 3 and myeloperoxidase ANCA immunoassays as opposed to indirect immunofluorescence, is now recommended as the preferred initial screening method and has high diagnostic accuracy in selected patients. [49,50]

Anti-glomerular basement membrane (GBM) disease is a rare cause of DAH, and typically presents as a pulmonary-renal syndrome. Circulating anti-GBM antibodies can be detected with high specificity by enzyme-linked immunosorbent assays in >90% of patients. Notably, however, clinicians should be aware that negative test results may be due to all the antibodies being fixed to their target (lung and kidney), or to a lack of sensitivity. In the latter clinical setting, renal biopsy can be per-

formed, and typically indicates extracapillary glomerulonephritis with linear immunoglobulin G deposits along the GBM. In patients with anti-GBM disease, urgent immunosuppression and plasma exchange are required to prevent permanent loss of renal function. [51] It remains uncertain whether patients with ANCA-associated vasculitis benefit from plasma exchange. [52,53]

### Other immune diseases

Less commonly, autoinflammatory diseases can present as AHRF. Although predominantly seen in pediatrics, it is important to be aware of situations that can occur in adults in the ICU. Severe forms of Still's disease may require admission to intensive care, and pulmonary involvement is reported in 75% of patients, typically with fever, often associated with cardiac involvement and hemophagocytosis.<sup>[54]</sup> A transient skin rash, arthralgias, odynophagia, and markedly elevated ferritin levels contribute to the formulation of the diagnosis. [55] COPA syndrome, a recently described monogenic interferon-related autoinflammatory syndrome, is mainly associated with DAH, ILD, and arthritis. Forms with isolated pulmonary involvement have been described, and adult-onset has been documented in a few patients.<sup>[56]</sup> Positive interferon assessment is a disease marker. Myelodysplastic/myeloproliferative neoplasms are increasingly recognized for their immunological manifestations, and can be associated with pulmonary involvement<sup>[57]</sup> Prominent examples include non-Langerhans histiocytosis<sup>[58,59]</sup> and the recently described vacuoles, E1 enzyme, X-linked, autoinflammatory, somatic (VEXAS) syndrome, [60,61] which can present with AHRF requiring ICU admission. Diagnosis is based on a complete blood count, assessment for extrapulmonary manifestations, and bone marrow biopsy with next-generation sequencing myeloid panel analyses.

Lastly, pulmonary alveolar proteinosis (PAP) is a disease characterized by abnormal accumulation of surfactant in the alveoli, and alveolar macrophage dysfunction. PAP results in progressive dyspnea of insidious onset. Primary PAP is characterized by the disruption of granulocyte-macrophage colonystimulating factor (GM-CSF) signaling. Autoimmune PAP is caused by elevated levels of GM-CSF autoantibodies, and accounts for >90 % of all cases. [62,63]

# Drug-induced causes

Extensive lists of drugs have been reported as causes of druginduced pneumonias that can lead to AHRF. To our knowledge, no studies have reported the prevalence of drug-related causes in patients admitted to the ICU with AHRF, but several studies have addressed this issue in patients with ARDS. Dhokarh et al. [64] reported that in a retrospective cohort of 514 ARDS patients, 9.5 % could be classified as having drug-induced ARDS. In a study conducted by Gibelin et al., [11] 26 % of patients with ARDS and no common risk factor identified - 7.5 % of the whole cohort - were ultimately diagnosed with drug-induced ARDS, with chemotherapeutic agents and amiodarone being the most common culprits. Anan et al.[65] reported that in a retrospective cohort of 197 patients with ARDS, 13.7% could be classified as being drug-induced. In fact, all drugs can be associated with pulmonary adverse effects, with a wide spectrum of clinical and radiological patterns (e.g., chronic ILD, ARDS, DAH, eosinophilic pneumonia), making diagnosis particularly challenging for clinicians. The drug-induced respiratory disease website (pneumotox.com) can be a useful tool in this regard. Patients at higher risk of pulmonary toxicity include those of older age, those with pre-existing lung diseases including ILD and lung cancer, and lung surgery patients. [66] The main drugs involved are amiodarone, bleomycin (Bleomycin), methotrexate (Methotrexate), mechanistic target of rapamycin inhibitors, cyclophosphamide and other alkylating agents, and anti-CD20 therapies. [66] Thoracic radiotherapy is also a classic cause of acute lung injury that can lead to AHRF, particularly in the presence of underlying ILD. [67]

Immune-checkpoint inhibitors, which represent the latest breakthrough in oncology, have recently been reported to cause immune-related adverse events. [68] These include several pulmonary complications such as sarcoid-like granulomatosis, pleural effusion, exacerbation of obstructive lung disease, and most notably, checkpoint inhibitor pneumonitis (CIP). [68] Of these complications, CIP is the most widely recognized, and is of serious concern due to its high morbidity and mortality rates. The incidence of CIP in clinical trials has been reported to be <6 %. This claim has been challenged by real-world data, however, which have indicated higher rates ranging from 10% to 20%. Several factors have been shown to increase the risk of CIP, including the use of programmed cell death protein (PD-1) or programmed cell death protein ligand agents (vs. cytotoxic T-lymphocyte-associated antigen 4 [CTLA-4] inhibitors), combination immunotherapy (CTLA-4 and PD-1 combination vs. monotherapy), radiotherapy, and the organ of tumor origin (e.g., non-small cell lung cancer vs. renal cell or other cancer types). [68,69] The clinical presentation of CIP includes dyspnea, cough, and hypoxemia, either at rest or on exertion. These symptoms may appear soon after treatment initiation, typically within days to weeks. The reported timeframes vary between study cohorts, but the reported median onset is approximately 2-3 months after starting on immune-checkpoint inhibitors. The most common radiographic patterns found are ground-glass opacities, organizing pneumonitis, and hypersensitivity pneumonitis.[69]

# Malignant causes

Several cancers can be revealed as a result of AHRF, and fulfill the clinical and radiological criteria for ARDS, [11] including mainly solid cancers with lymphangitic carcinomatosis and lung adenocarcinomas. Mediastinal/hilar lymphadenopathies and/or pulmonary nodules or masses on chest CT may help to raise clinical suspicion, and guide the diagnostic strategy. Lepidic adenocarcinoma is a well-differentiated adenocarcinoma that develops along intact alveolar septa without stromal, pleural, or vascular invasion, [70] and its clinical and radiological presentation may mimic pneumonia.<sup>[71]</sup> Cytological analysis of respiratory samples (sputum or BAL) enables diagnosis. Small cell neuroendocrine carcinoma is a rare but well-known cause of AHRF in the ICU.[72] Among the hematological malignancies, acute myeloid leukemia is most commonly associated with pulmonary involvement, which may be secondary to pulmonary leukostasis, pulmonary leukemic infiltration, and acute lysis pneumopathy. [25,73] In-hospital mortality is particularly high in these patients, approaching 50% according to a multicenter, international, prospective study. [74]

### Idiopathic diseases

Acute interstitial pneumonia refers to an idiopathic form of AHRF characterized histologically by the presence of diffuse alveolar damage in the absence of exposure to ARDS risk factors, and no other identified cause, including CTD and drug exposure. [75,76] It affects patients of all ages and both sexes, and is often preceded by a flu-like illness. The duration of symptoms in the original series ranged from 2 days to 11 days. [76] Physical findings are non-specific and do not include the digital clubbing seen in patients with acute exacerbations of idiopathic pulmonary fibrosis/usual interstitial pneumonia, which are chronic forms of idiopathic lung disease. Treatment is based on supportive care and high-dose intravenous steroids, although the level of evidence is low. [76] Mortality has ranged from 50% to 100% in previously reported series. [76] Cryptogenic organizing pneumonia is a subacute idiopathic interstitial pneumonia<sup>[77]</sup> characterized by patchy and often migratory multifocal alveolar consolidations and a rapid response to steroids, but frequent relapses. Acute eosinophilic pneumonia can have an abrupt and severe onset, presenting with ARDS criteria, and may be idiopathic, although identifiable causes such as smoking and inhalation exposures, medications, and infections have been reported. [78] Marked BAL fluid eosinophilia (>25%) is a key feature of its diagnosis. Peripheral blood eosinophilia may be absent in the early stages of the disease, and a marked increase occurs after corticosteroids are initiated, peaking 4-5 days thereafter. [79] A rapid clinical response is typically achieved within 24-48 h of corticosteroid initiation.

## Pragmatic diagnostic approach for rare causes of AHRF

When faced with a patient with AHRF without an apparent cause after an initial diagnostic work-up, it is appropriate to perform a comprehensive microbiological evaluation (Fiure 1), guided by possible underlying immunosuppression, bronchoscopic BAL analysis, and high-resolution chest CT. BAL is a high-risk procedure in non-intubated patients, with the potential for secondary deterioration. A non-invasive strategy has been shown to have comparable diagnostic performance to an invasive approach in onco-hematological patients with AHRF.[80] In non-intubated patients, BAL should only be considered after a well-executed non-invasive strategy. If the diagnostic work-up remains negative at this stage, a comprehensive search for the main causes of ARDS with no risk factor is required, using a multidisciplinary approach. First, a detailed history and physical examination are essential to look for possible extra-respiratory symptoms, signs, and exposure to pneumotoxic drugs. Second, a comprehensive immunological assessment is required, including antinuclear antibodies, anti-extractable nuclear antigen panel, rheumatoid factor, anti-cyclic citrullinated peptide (CCP) antibodies, ANCA, myositis-specific antibodies and - depending on the clinical context - anti-GBM and anti-GM-CSF antibodies. The presence of a macroscopically bloody BAL in the absence of bronchial hemorrhage is associated with DAH in >80 % of cases, [81,82] and together with the presence of extrapulmonary symptoms (e.g., arthralgias, renal involvement), it is highly suggestive of an immune cause. [83] If a diagnosis remains elusive a lung biopsy approach may be considered, [7,14] but this strategy is only feasible in invasively ventilated patients. Previous ob-

servational series have reported high diagnostic performance of open lung biopsy in selected patients.[10,84-86] Specifically, in a meta-analysis conducted by Libby et al., [85] open lung biopsy was associated with a high diagnostic performance of 84% in ARDS patients, with resulting changes in patient management in 73% of cases. Other studies have reported lower diagnostic performance, however, [87] and the potential complications of the procedure are substantial, with reported complication rates of 25 %-35 %. [84,86] These reported complications are mainly related to air leaks, which can sometimes be fatal. The specificity of histological findings is also questionable. Therefore, the decision to perform a diagnostic open lung biopsy should only be considered in a multidisciplinary context, after the failure of a comprehensive and well-conducted non-invasive approach in the face of ARDS with no identified risk factors and unfavorable evolution, most often after a trial of high-dose corticosteroids. [14] Notably, of the 2813 ARDS patients managed in the LUNG SAFE study, only 11 (0.4%) underwent open lung biopsy, indicating that the vast majority of ARDS cases are managed without a lung biopsy. [1] The roles of less invasive techniques such as cryobiopsies and transbronchial biopsies remain to be determined in the ICU setting.

### **Conclusions**

Rapid identification of the cause of AHRF is essential in order to facilitate the initiation of targeted treatment as soon as possible. Patients with AHRF in whom no common cause has been identified should rapidly undergo a comprehensive and hierarchical diagnostic work-up aimed at categorizing the underlying disease as immune, drug-induced, malignant, or idiopathic. A multidisciplinary approach involving intensivists and organ specialists is required to determine which patients may benefit from a particular intervention such as immunosuppression, removal of pneumotoxic drugs, or corticosteroid therapy.

### **CRediT Authorship Contribution Statement**

**Pierre Bay:** Writing – review & editing, Writing – original draft, Conceptualization. **Nicolas de Prost:** Writing – review & editing, Validation, Supervision, Conceptualization.

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### **Ethics Statement**

Not applicable.

## **Conflict of Interest**

N.D.P. has served as an advisor or speaker for Moderna and AstraZeneca.

### **Data Availability**

The data sets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

### References

- [1] Bellani G, Laffey JG, Pham T, Fan E, Brochard L, Esteban A, et al. Epidemiology, patterns of care, and mortality for patients with acute respiratory distress syndrome in intensive care units in 50 countries. JAMA 2016;315(8):788–800. doi:10.1001/jama.2016.0291.
- [2] Frat JP, Quenot JP, Badie J, Coudroy R, Guitton C, Ehrmann S, et al. Effect of high-flow nasal cannula oxygen vs standard oxygen therapy on mortality in patients with respiratory failure due to COVID-19: the SOHO-COVID randomized clinical trial. JAMA 2022;328(12):1212–22. doi:10.1001/jama.2022.15613.
- [3] Frat JP, Thille AW, Mercat A, Girault C, Ragot S, Perbet S, et al. High-flow oxygen through nasal cannula in acute hypoxemic respiratory failure. N Engl J Med 2015;372(23):2185–96. doi:10.1056/NEJMoa1503326.
- [4] Villar J, Mora-Ordoñez JM, Soler JA, Mosteiro F, Vidal A, Ambrós A, et al. The PANDORA study: prevalence and outcome of acute hypoxemic respiratory failure in the pre-COVID-19 era. Crit Care Explor 2022;4(5):e0684. doi:10.1097/CCE.00000000000000684.
- [5] Thompson BT, Chambers RC, Liu KD. Acute respiratory distress syndrome. N Engl J Med 2017;377(6):562–72. doi:10.1056/NEJMra1608077.
- [6] Matthay MA, Arabi Y, Arroliga AC, Bernard G, Bersten AD, Brochard LJ, et al. A new global definition of acute respiratory distress syndrome. Am J Respir Crit Care Med 2023;209(1):37–47. doi:10.1164/rccm.202303-0558WS.
- [7] Papazian L, Calfee CS, Chiumello D, Luyt CE, Meyer NJ, Sekiguchi H, et al. Diagnostic workup for ARDS patients. Intensive Care Med 2016;42(5):674–85. doi:10.1007/s00134-016-4324-5.
- [8] Gibelin A, Parrot A, Fartoukh M, de Prost N. Rare respiratory diseases in the ICU: when to suspect them and specific approaches. Curr Opin Crit Care 2019;25(1):29–36. doi:10.1097/MCC.0000000000000572.
- [9] Coudroy R, Frat JP, Ehrmann S, Pène F, Decavèle M, Terzi N, et al. High-flow nasal oxygen alone or alternating with non-invasive ventilation in critically ill immunocompromised patients with acute respiratory failure: a randomised controlled trial. Lancet Respir Med 2022;10(7):641–9. doi:10.1016/S2213-2600(22)00096-0.
- [10] Papazian L, Doddoli C, Chetaille B, Gernez Y, Thirion X, Roch A, et al. A contributive result of open-lung biopsy improves survival in acute respiratory distress syndrome patients. Crit Care Med 2007;35(3):755–62. doi:10.1097/01.CCM.0000257325.88144.30.
- [11] Gibelin A, Parrot A, Maitre B, Brun-Buisson C, Mekontso Dessap A, Far-toukh M, et al. Acute respiratory distress syndrome mimickers lacking common risk factors of the Berlin definition. Intensive Care Med 2016;42(2):164–72. doi:10.1007/s00134-015-4064-y.
- [12] de Prost N, Pham T, Carteaux G, Mekontso Dessap A, Brun-Buisson C, Fan E, et al. Etiologies, diagnostic work-up and outcomes of acute respiratory distress syndrome with no common risk factor: a prospective multicenter study. Ann Intensive Care 2017;7(1):69. doi:10.1186/s13613-017-0281-6.
- [13] Smit MR, Mayo PH, Mongodi S. Lung ultrasound for diagnosis and management of ARDS. Intensive Care Med 2024;50(7):1143–5. doi:10.1007/s00134-024-07422-7.
- [14] Bos LDJ, de Grooth HJ, Tuinman PR. A structured diagnostic algorithm for patients with ARDS. Crit Care 2023;27(1):94. doi:10.1186/s13054-023-04368-y.
- [15] Torres-Aguilar H, Sosa-Luis SA, SR Aguilar-Ruiz. Infections as triggers of flares in systemic autoimmune diseases: novel innate immunity mechanisms. Curr Opin Rheumatol 2019;31(5):525–31. doi:10.1097/BOR.00000000000000630.
- [16] Guérin C, Thompson T, Brower R. The ten diseases that look like ARDS. Intensive Care Med 2015;41(6):1099–102. doi:10.1007/s00134-014-3608-x.
- [17] Dequin PF, Meziani F, Quenot JP, Kamel T, Ricard JD, Badie J, et al. Hydrocortisone in severe community-acquired pneumonia. N Engl J Med 2023;388(21):1931–41. doi:10.1056/NEJMoa2215145.
- [18] Meduri GU, Shih MC, Bridges L, Martin TJ, El-Solh A, Seam N, et al. Low-dose methylprednisolone treatment in critically ill patients with severe community-acquired pneumonia. Intensive Care Med 2022;48(8):1009–23. doi:10.1007/s00134-022-06684-3.
- [19] Torres A, Cilloniz C, Niederman MS, Menéndez R, Chalmers JD, Wunderink RG, et al. Pneumonia. Nat Rev Dis Primers 2021;7(1):1–28. doi:10.1038/s41572-021-00259-0.
- [20] Edin A, Eilers H, Allard A. Evaluation of the Biofire Filmarray Pneumonia panel plus for lower respiratory tract infections. Infect Dis (Lond) 2020;52(7):479–88. doi:10.1080/23744235.2020.1755053.
- [21] Dessajan J, Timsit JF. Impact of multiplex PCR in the therapeutic management of severe bacterial pneumonia. Antibiotics (Basel) 2024;13(1):95. doi:10.3390/antibiotics13010095.
- [22] CDC. Investigating Legionnaires' disease. Methods for legionella testing and specimen collection. 2024. Available from: https://www.cdc.gov/investigatelegionella/php/public-health-strategy/legionella-testing.html. Accessed July 22, 2024
- [23] Avni T, Bieber A, Green H, Steinmetz T, Leibovici L, Paul M. Diagnostic accuracy of PCR alone and compared to urinary antigen testing for detection of Legionella spp.: a systematic review. J Clin Microbiol 2016;54(2):401–11. doi:10.1128/JCM.02675-15.

- [24] Avšič-Županc T, Saksida A, Korva M. Hantavirus infections. Clin Microbiol Infect 2019;21:e6–16. doi:10.1111/1469-0691.12291.
- [25] Azoulay E, Mokart D, Kouatchet A, Demoule A, Lemiale V. Acute respiratory failure in immunocompromised adults. Lancet Respir Med 2019;7(2):173–86. doi:10.1016/S2213-2600(18)30345-X.
- [26] Gu W, Deng X, Lee M, Sucu YD, Arevalo S, Stryke D, et al. Rapid pathogen detection by metagenomic next-generation sequencing of infected body fluids. Nat Med 2021;27(1):115–24. doi:10.1038/s41591-020-1105-z.
- [27] Peng JM, Du B, Qin HY, Wang Q, Shi Y. Metagenomic next-generation sequencing for the diagnosis of suspected pneumonia in immunocompromised patients. J Infect 2021;82(4):22–7. doi:10.1016/j.jinf.2021.01.029.
- [28] Neyton LPA, Langelier CR, Calfee CS. Metagenomic sequencing in the ICU for precision diagnosis of critical infectious illnesses. Crit Care 2023;27(1):90. doi:10.1186/s13054-023-04365-1.
- [29] Larcher R, Pineton de Chambrun M, Garnier F, Rubenstein E, Carr J, Charbit J, et al. One-year outcome of critically ill patients with systemic rheumatic disease: a multi-center cohort study. Chest 2020;158(3):1017–26. doi:10.1016/j.chest.2020.03.050.
- [30] Bay P, Lebreton G, Mathian A, Demondion P, Desnos C, Chommeloux J, et al. Outcomes of severe systemic rheumatic disease patients requiring extracorporeal membrane oxygenation. Ann Intensive Care 2021;11(1):29. doi:10.1186/s13613-021-00819-3.
- [31] Zafrani L, Lemiale V, Lapidus N, Lorillon G, Schlemmer B, Azoulay E. Acute respiratory failure in critically ill patients with interstitial lung disease. PLoS One 2014;9(8):e104897. doi:10.1371/journal.pone.0104897.
- [32] Johnson SR, Bernstein EJ, Bolster MB, Chung JH, Danoff SK, George MD, et al. 2023 American College of Rheumatology (ACR)/American College of Chest Physicians (CHEST) guideline for the screening and monitoring of interstitial lung disease in people with systemic autoimmune rheumatic diseases. Arthritis Rheumatol 2024;76(8):1201–13. doi:10.1002/art.42860.
- [33] Lundberg IE, Fujimoto M, Vencovsky J, Aggarwal R, Holmqvist M, Christopher-Stine L, et al. Idiopathic inflammatory myopathies. Nat Rev Dis Primers 2021;7(1):86. doi:10.1038/s41572-021-00321-x.
- [34] Vuillard C, Pineton de Chambrun M, de Prost N, Guérin C, Schmidt M, Dargent A, et al. Clinical features and outcome of patients with acute respiratory failure revealing anti-synthetase or anti-MDA-5 dermato-pulmonary syndrome: a French multicenter retrospective study. Ann Intensive Care 2018;8(1):87. doi:10.1186/s13613-018-0433-3.
- [35] Hervier B, Devilliers H, Stanciu R, Meyer A, Uzunhan Y, Masseau A, et al. Hierarchical cluster and survival analyses of antisynthetase syndrome: phenotype and outcome are correlated with anti-tRNA synthetase antibody specificity. Autoimmun Rev 2012;12(2):210–17. doi:10.1016/j.autrev.2012.06.006.
- [36] Liu Y, Liu X, Xie M, Chen Z, He J, Wang Z, et al. Clinical characteristics of patients with anti-EJ antisynthetase syndrome associated interstitial lung disease and literature review. Respir Med 2020;165:105920. doi:10.1016/j.rmed.2020.105920.
- [37] Pinal-Fernandez I, Casal-Dominguez M, Huapaya JA, Albayda J, Paik JJ, Johnson C, et al. A longitudinal cohort study of the anti-synthetase syndrome: increased severity of interstitial lung disease in black patients and patients with anti-PL7 and anti-PL12 autoantibodies. Rheumatology (Oxford) 2017;56(6):999–1007. doi:10.1093/rheumatology/kex021.
- [38] Pineton de Chambrun M, Hervier B, Chauveau S, Tandjaoui-Lambiotte Y, Combes A, Uzunhan Y. Tofacitinib in antisynthetase syndrome-related rapidly progressive interstitial lung disease. Rheumatology (Oxford) 2020;59(12):e142–3. doi:10.1093/rheumatology/keaa323.
- [39] Ye S, Chen XX, Lu XY, Wu MF, Deng Y, Huang WQ, et al. Adult clinically amyopathic dermatomyositis with rapid progressive interstitial lung disease: a retrospective cohort study. Clin Rheumatol 2007;26(10):1647–54. doi:10.1007/s10067-007-0562-9.
- [40] Allenbach Y, Uzunhan Y, Toquet S, Leroux G, Gallay L, Marquet A, et al. Different phenotypes in dermatomyositis associated with anti-MDA5 antibody: study of 121 cases. Neurology 2020;95(1):e70–8. doi:10.1212/WNL.0000000000009727.
- [41] Lu X, Peng Q, Wang G. Anti-MDA5 antibody-positive dermatomyositis: pathogenesis and clinical progress. Nat Rev Rheumatol 2024;20(1):48–62. doi:10.1038/s41584-023-01054-9.
- [42] Bay P, de Chambrun MP, Roux A, Bunel V, Combes A, Biet DI, et al. Extracorporeal life support allows lung transplant in anti-MDA5+ rapidly progressive-interstitial lung disease. Eur Respir J 2022;59(5):2102968. doi:10.1183/13993003.02968-2021.
- [43] Bay P, de Chambrun MP, Rothstein V, Mahevas M, De Prost N, Roux A, et al. Efficacy of plasma exchange in patients with anti-MDA5 rapidly progressive interstitial lung disease. J Autoimmun 2022;133:102941. doi:10.1016/j.jaut.2022.102941.
- [44] Lhote R, Grenier P, Haroche J, Miyara M, Boussouar S, Mathian A, et al. Characterization of interstitial lung disease associated with anti-ribonucleoprotein antibodies. J Clin Rheumatol 2020;26(8):327–33. doi:10.1097/RHU.0000000000001127.
- [45] Triboulet F, Guérin E, Boussouar S, Hékimian G, Pha M, Rouvier P, et al. Systemic lupus erythematosus-related acute lung disease. Lupus 2023;32(9):1117–22. doi:10.1177/09612033231188034.
- [46] Pineton de Chambrun M, Larcher R, Pène F, Argaud L, Mayaux J, Jamme M, et al. In-hospital mortality-associated factors in patients with thrombotic antiphospholipid syndrome requiring ICU admission. Chest 2020;157(5):1158–66. doi:10.1016/j.chest.2019.11.010.
- [47] Demiselle J, Auchabie J, Beloncle F, Gatault P, Grangé S, Du Cheyron D, et al. Patients with ANCA-associated vasculitis admitted to the intensive care unit with acute vasculitis manifestations: a retrospective and comparative multicentric study. Ann Intensive Care 2017;7(1):39. doi:10.1186/s13613-017-0262-9.
- [48] Kimmoun A, Baux E, Das V, Terzi N, Talec P, Asfar P, et al. Outcomes of patients admitted to intensive care units for acute manifestation of small-

- vessel vasculitis: a multicenter, retrospective study. Crit Care 2016;20:27. doi:10.1186/s13054-016-1189-5.
- [49] Emmi G, Bettiol A, Gelain E, Bajema IM, Berti A, Burns S, et al. Evidence-based guideline for the diagnosis and management of eosinophilic granulomatosis with polyangiitis. Nat Rev Rheumatol 2023;19(6):378–93. doi:10.1038/s41584-023-00958-w.
- [50] Bossuyt X, Cohen Tervaert JW, Arimura Y, Blockmans D, Flores-Suárez LF, Guillevin L, et al. Position paper: revised 2017 international consensus on testing of ANCAs in granulomatosis with polyangiitis and microscopic polyangiitis. Nat Rev Rheumatol 2017;13(11):683–92. doi:10.1038/nrrheum.2017.140.
- [51] Henderson SR, Salama AD. Diagnostic and management challenges in Goodpasture's (anti-glomerular basement membrane) disease. Nephrol Dial Transplant 2018;33(2):196–202. doi:10.1093/ndt/gfx057.
- [52] Walsh M, Merkel PA, Peh CA, Szpirt WM, Puéchal X, Fujimoto S, et al. Plasma exchange and glucocorticoids in severe ANCA-associated vasculitis. N Engl J Med 2020;382(7):622–31. doi:10.1056/NEJMoa1803537.
- [53] Fussner LA, Flores-Suárez LF, Cartin-Ceba R, Specks U, Cox PG, Jayne DRW, et al. Alveolar hemorrhage in antineutrophil cytoplasmic antibody-associated vasculitis: results of an international randomized controlled trial (PEXIVAS). Am J Respir Crit Care Med 2024;209(9):1141–51. doi:10.1164/rccm.202308-14260C.
- [54] Néel A, Wahbi A, Tessoulin B, Boileau J, Carpentier D, Decaux O, et al. Diagnostic and management of life-threatening adult-onset Still disease: a French nation-wide multicenter study and systematic literature review. Crit Care 2018;22(1):88. doi:10.1186/s13054-018-2012-2.
- [55] Giacomelli R, Ruscitti P, Shoenfeld Y. A comprehensive review on adult onset Still's disease. J Autoimmun 2018;93:24–36. doi:10.1016/j.jaut.2018.07.018.
- [56] Frémond ML, Nathan N. COPA syndrome, 5 years after: where are we? Joint Bone Spine 2021;88(2):105070. doi:10.1016/j.jbspin.2020.09.002.
- [57] Hochman MJ, DeZern AE. Myelodysplastic syndrome and autoimmune disorders: two sides of the same coin? Lancet Haematol 2022;9(7):e523–34. doi:10.1016/S2352-3026(22)00138-7.
- [58] Moyon Q, Boussouar S, Maksud P, Emile JF, Charlotte F, Aladjidi N, et al. Lung involvement in Destombes-Rosai-Dorfman disease: clinical and radiological features and response to the MEK inhibitor cobimetinib. Chest 2020;157(2):323–33. doi:10.1016/j.chest.2019.09.036.
- [59] Wang JN, Wang FD, Sun J, Liang ZY, Li J, Zhou DB, et al. Pulmonary manifestations of Erdheim-Chester disease: clinical characteristics, outcomes and comparison with Langerhans cell histiocytosis. Br J Haematol 2021;194(6):1024–33. doi:10.1111/bjh.17712.
- [60] Borie R, Debray MP, Guedon AF, Mekinian A, Terriou L, Lacombe V, et al. Pleuropulmonary manifestations of vacuoles, E1 enzyme, X-linked, autoinflammatory, somatic (VEXAS) syndrome. Chest 2023;163(3):575–85. doi:10.1016/j.chest.2022. 10.011
- [61] Beck DB, Ferrada MA, Sikora KA, Ombrello AK, Collins JC, Pei W, et al. Somatic mutations in UBA1 and severe adult-onset autoinflammatory disease. N Engl J Med 2020;383(27):2628–38. doi:10.1056/NEJMoa2026834.
- [62] Trapnell BC, Whitsett JA, Nakata K. Pulmonary alveolar proteinosis. N Engl J Med 2003;349(26):2527–39. doi:10.1056/NEJMra023226.
- [63] Trapnell BC, Nakata K, Bonella F, Campo I, Griese M, Hamilton J, et al. Pulmonary alveolar proteinosis. Nat Rev Dis Primers 2019;5(1):16. doi:10.1038/s41572-019-0066-3.
- [64] Dhokarh R, Li G, Schmickl CN, Kashyap R, Assudani J, Limper AH, et al. Drug-associated acute lung injury: a population-based cohort study. Chest 2012;142(4):845–50. doi:10.1378/chest.11-2103.
- [65] Anan K, Ichikado K, Kawamura K, Johkoh T, Fujimoto K, Suga M. Clinical characteristics and prognosis of drug-associated acute respiratory distress syndrome compared with non-drug-associated acute respiratory distress syndrome: a single-centre retrospective study in Japan. BMJ Open 2017;7(11):e015330. doi:10.1136/bmjopen-2016-015330.
- [66] Spagnolo P, Bonniaud P, Rossi G, Sverzellati N, Cottin V. Druginduced interstitial lung disease. Eur Respir J 2022;60(4):2102776. doi:10.1183/13993003.02776-2021.
- [67] Goodman CD, Nijman SFM, Senan S, Nossent EJ, Ryerson CJ, Dhaliwal I, et al. A primer on interstitial lung disease and thoracic radiation. J Thorac Oncol 2020;15(6):902–13. doi:10.1016/j.jtho.2020.02.005.
- [68] Ghanbar MI, Suresh K. Pulmonary toxicity of immune checkpoint immunotherapy. J Clin Invest 2024;134(2):e170503. doi:10.1172/JCI170503.

- [69] Delaunay M, Cadranel J, Lusque A, Meyer N, Gounant V, Moro-Sibilot D, et al. Immune-checkpoint inhibitors associated with interstitial lung disease in cancer patients. Eur Respir J 2017;50(2):1700050. doi:10.1183/13993003.00050-2017.
- [70] Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisinger KR, Yatabe Y, et al. International association for the study of lung cancer/American Thoracic Society/European Respiratory Society international multidisciplinary classification of lung adenocarcinoma. J Thorac Oncol 2011;6(2):244–85. doi:10.1097/JTO.0b013e318206a221.
- [71] Elabbadi A, Fajac A, Antoine M, Cadranel J, Voiriot G, Fartoukh M, et al. A severe COVID-19 pneumonia revealing a lepidic adenocarcinoma: a diagnostic challenge during the pandemic period. Am J Respir Crit Care Med 2022;205(3):357–9. doi:10.1164/rccm.202104-0826IM.
- [72] Huang J, Kwon DH, Brondfield S. Outcomes of patients with small cell lung cancer admitted to the intensive care unit. J Clin Oncol 2023;41(16\_suppl):e20620. doi:10.1200/JCO.2023.41.16\_suppl.e20620.
- [73] Moreau AS, Lengline E, Seguin A, Lemiale V, Canet E, Raffoux E, et al. Respiratory events at the earliest phase of acute myeloid leukemia. Leuk Lymphoma 2014;55(11):2556–63. doi:10.3109/10428194.2014.887709.
- [74] Secreto C, Chean D, van de Louw A, Kouatchet A, Bauer P, Cerrano M, et al. Characteristics and outcomes of patients with acute myeloid leukemia admitted to intensive care unit with acute respiratory failure: a post-hoc analysis of a prospective multicenter study. Ann Intensive Care 2023;13(1):79. doi:10.1186/s13613-023-01172-3.
- [75] Travis WD, Costabel U, Hansell DM, King TE Jr, Lynch DA, Nicholson AG, et al. An Official American Thoracic Society/European Respiratory Society Statement: update of the international multidisciplinary classification of the idiopathic interstitial pneumonias. Am J Respir Crit Care Med 2013;188(6):733–48. doi:10.1164/rccm.201308-1483ST.
- [76] Mukhopadhyay S, Parambil JG. Acute interstitial pneumonia (AIP): relationship to Hamman-Rich syndrome, diffuse alveolar damage (DAD), and acute respiratory distress syndrome (ARDS). Semin Respir Crit Care Med 2012;33(5):476–85. doi:10.1055/s-0032-1325158.
- [77] King TE, Lee JS. Cryptogenic organizing pneumonia. N Engl J Med 2022;386(11):1058–69. doi:10.1056/NEJMra2116777.
- [78] De Giacomi F, Vassallo R, Yi ES, Ryu JH. Acute eosinophilic pneumonia. Causes, diagnosis, and management. Am J Respir Crit Care Med 2018;197(6):728–36. doi:10.1164/rccm.201710-1967CI.
- [79] Bay P, Groh M, Gaillet A, Schmidt M, Luyt CE, Combes A, et al. Extracorporeal membrane oxygenation for refractory acute eosinophilic pneumonia. J Crit Care 2024;79:154437. doi:10.1016/j.jcrc.2023.154437.
- [80] Azoulay E, Mokart D, Lambert J, Lemiale V, Rabbat A, Kouatchet A, et al. Diagnostic strategy for hematology and oncology patients with acute respiratory failure: randomized controlled trial. Am J Respir Crit Care Med 2010;182(8):1038–46. doi:10.1164/rccm.201001-0018OC.
- [81] de Prost N, Parrot A, Cuquemelle E, Picard C, Antoine M, Fleury-Feith J, et al. Diffuse alveolar hemorrhage in immunocompetent patients: etiologies and prognosis revisited. Respir Med. 2012;106(7):1021–32. doi:10.1016/j.rmed.2012.03.015.
- [82] de Prost N, Parrot A, Picard C, Ancel PY, Mayaud C, Fartoukh M, et al. Diffuse alveolar haemorrhage: factors associated with in-hospital and long-term mortality. Eur Respir J 2010;35(6):1303–11. doi:10.1183/09031936.00075309.
- [83] de Prost N, Parrot A, Cuquemelle E, Picard C, Cadranel J. Immune diffuse alveolar hemorrhage: a retrospective assessment of a diagnostic scale. Lung 2013;191(5):559–63. doi:10.1007/s00408-013-9491-3.
- [84] Philipponnet C, Cassagnes L, Pereira B, Kemeny JL, Devouassoux-Shisheboran M, Lautrette A, et al. Diagnostic yield and therapeutic impact of open lung biopsy in the critically ill patient. PLoS One 2018;13(5):e0196795. doi:10.1371/jourpal.pose.0196795
- [85] Libby LJ, Gelbman BD, Altorki NK, Christos PJ, Libby DM. Surgical lung biopsy in adult respiratory distress syndrome: a meta-analysis. Ann Thorac Surg 2014;98(4):1254–60. doi:10.1016/j.athoracsur.2014.05.029.
- [86] Guerin C, Bayle F, Leray V, Debord S, Stoian A, Yonis H, et al. Open lung biopsy in nonresolving ARDS frequently identifies diffuse alveolar damage regardless of the severity stage and may have implications for patient management. Intensive Care Med 2015;41(2):387. doi:10.1007/s00134-015-3657-9.
- [87] Papazian L, Thomas P, Bregeon F, Garbe L, Zandotti C, Saux P, et al. Openlung biopsy in patients with acute respiratory distress syndrome. Anesthesiology 1998;88(4):935–44. doi:10.1097/00000542-199804000-00013.