

Multiscale analysis and optimal glioma therapeutic candidate discovery using the CANDO platform

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Abstract

Glioma is a highly malignant brain tumor with limited treatment options. We employed the Computational Analysis of Novel Drug Opportunities (CANDO) platform for multiscale therapeutic discovery to predict new glioma therapies. We began by computing interaction scores between extensive libraries of drugs/compounds and proteins to generate “interaction signatures” that model compound behavior on a proteomic scale. Compounds with signatures most similar to those of drugs approved for a given indication were considered potential treatments. These compounds were further ranked by degree of consensus in corresponding similarity lists. We benchmarked performance by measuring the recovery of approved drugs in these similarity and consensus lists at various cutoffs, using multiple metrics and comparing results to random controls and performance across all indications. Compounds ranked highly by consensus but not previously associated with the indication of interest were considered new predictions. Our benchmarking results showed that CANDO improved accuracy in identifying glioma-associated drugs across all cutoffs compared to random

33 controls. Our predictions, supported by literature-based analysis, identified 23
34 potential glioma treatments, including approved drugs like vitamin D, taxanes,
35 vinca alkaloids, topoisomerase inhibitors, and folic acid, as well as investigational
36 compounds such as ginsenosides, chrysin, resiniferatoxin, and cryptotanshinone.
37 Further functional annotation-based analysis of the top targets with the strongest
38 interactions to these predictions identified Vitamin D3 receptor, thyroid hormone
39 receptor, acetylcholinesterase, cyclin-dependent kinase 2, tubulin alpha chain,
40 dihydrofolate reductase, and thymidylate synthase. These findings indicate that
41 CANDO's multitarget, multiscale framework is effective in identifying glioma
42 drug candidates thereby informing new strategies for improving treatment.

43 **Keywords:** glioma, multiscale drug discovery, computational drug repurposing,
44 translational bioinformatics, deep learning, systems biology

45 List of Abbreviations

CANDO	Computational Analysis of Novel Drug Opportunities
BBB	Blood-brain barrier
P-gp	P-glycoprotein
ADME	Absorption, distribution, metabolism, and excretion
BANDOCK	Bioanalytical docking protocol
AF2	AlphaFold2
CTD	Comparative Toxicogenomics Database
MeSH	Medical Subject Headings
ECFP4	Extended Connectivity Fingerprints with a diameter of 4
RMSD	Root-mean-square deviation
IA	Indication accuracy
AIA	Average indication accuracy
nIA	New indication accuracy
NDCG	Normalized discounted cumulative gain
nNDCG	New NDCG
PI3K	Phosphatidylinositol-3'-kinase
mTOR	Mammalian target of rapamycin
JAK	Janus kinase
STAT	Signal transducer and activator of transcription
STAT3	Signal transducer and activator of transcription 3
AChE	Acetylcholinesterase
THRB	Thyroid hormone receptor beta
CDK1	Cyclin-dependent kinase 1
DHFR	Dihydrofolate reductase
TYMS	Thymidylate synthase
CNS	Central nervous system
GBM	Glioblastoma multiforme
TUBA1C	Tubulin alpha-1C chain

LGG	Low-grade gliomas
TOPI	Type I topoisomerases
TOPII	Type II topoisomerases
CPT-11	Irinotecan
SN-38	7-Ethyl-10-hydroxycamptothecin
FA	Folic acid
FR	Folate receptor
TRP	Transient vanilloid receptor
NIH	National Institutes of Health
NCATS	NIH Clinical and Translational Sciences
NLM	NIH National Library of Medicine
NIST	National Institute of Standards of Technology
CCR	Center for Computational Research

46 1 Introduction

47 Glioma is one of the most aggressive and fatal forms of malignant brain tumors, partic-
48 ularly prevalent among the elderly, with high rates of occurrence and mortality [1, 2].
49 Currently, chemotherapy is the primary treatment for glioma due to its aggressive
50 progression, various pathologies, and the challenges associated with complete surgi-
51 cal removal [3, 4]. However, the effectiveness of chemotherapy is significantly limited
52 by factors such as the selective permeability of the blood-brain barrier (BBB), neu-
53 rotoxicity, and inadequate drug delivery to the tumor site [5–9]. Furthermore, the
54 ATP-dependent efflux transporter, P-glycoprotein (P-gp), located on the BBB, con-
55 tributes to the removal of chemotherapeutic agents [10]. A substantial proportion
56 of patients with glioma (about 90%) experience tumor recurrence in the local area
57 after initial treatment [11]. Unfortunately, effective therapeutic options for recurrent
58 glioma are lacking. As a result, there is an urgent need to advance our understanding
59 of the molecular pathology of glioma, identify new therapeutic targets, and develop
60 innovative treatment strategies. A major challenge in modern medicine is the limited
61 availability of new glioma drugs that can cross the BBB [12–14].

62 The process of drug discovery aims to identify chemical compounds with thera-
63 peutic potential for treating human diseases. Despite substantial advances, the success
64 rate for the introduction of new drugs to the market has declined, with the aver-
65 age drug discovery pipeline now exceeding 12 years and costing more than 2 billion
66 dollars [15, 16]. Computational approaches, such as virtual high-throughput screen-
67 ing, are increasingly being used to identify potential lead compounds by simulating
68 and evaluating the binding affinity of numerous compounds against a target [17–20].
69 Challenges such as the vast combinatorial space of binding poses [21, 22] and ligand
70 conformations [23, 24], coupled with the complex dynamics of these systems [25], limit
71 the effectiveness of traditional virtual screening in reliably producing effective leads.
72 Some computational methods stand out for their efficiency, accuracy, comprehensive
73 assessment of interaction spaces, and broad exploration of chemical space, helping to
74 address the limitations of conventional approaches [26–33]. Although many computa-
75 tional screenings focus on a single protein target, drugs in humans interact with various

76 biological targets through processes such as absorption, distribution, metabolism, and
77 excretion (ADME), which influences their efficacy and safety [31–37]. Considering
78 drug interactions on a proteomic scale could yield more accurate predictions of bioac-
79 tivity and safety by accounting for both primary and secondary targets, essential for
80 optimizing therapeutic impact and minimizing toxicity.

81 We developed the Computational Analysis of Novel Drug Repurposing Opportu-
82 nities (CANDO) platform for multitarget drug discovery, repurposing, and design,
83 aiming to address the limitations of traditional single target, single disease approaches
84 [38–53]. CANDO exploits the fact that drugs approved for human use achieve thera-
85 peutic effects and optimal ADME through interactions with multiple biological targets,
86 and that off-target interactions are modulated to minimize adverse drug reactions.
87 CANDO capitalizes on this inherent multitargeting property of small molecules by con-
88 structing interaction signatures that reflect drug/compound behaviors across various
89 biological scales. The platform predicts putative drug candidates for every indica-
90 tion/disease by comparing and ranking these interaction signatures in an all-against-all
91 manner, with the hypothesis that drugs/compounds with similar interaction signatures
92 are more likely to display similar biological behavior. The platform is benchmarked by
93 evaluating the recovery of known drug-indication associations in these ranked lists of
94 interaction signatures within specified cutoffs. CANDO therefore deepens our under-
95 standing of small molecule therapeutics and their effects on proteins, pathways, and
96 various diseases by leveraging vast multiscale biomedical data on biological systems
97 and the phenotypic impact of their modulation. In addition to rigorous benchmarking
98 [38–53], CANDO and/or its components have been extensively validated prospectively
99 in the context of more than a dozen indications [38, 41, 47, 50–52, 54–64]. Herein, we
100 describe the use of CANDO to predict novel drug candidates for glioma treatment.

101 2 Methods

102 2.1 Applying the CANDO platform for glioma drug discovery 103 overview

104 We developed a pipeline within the CANDO platform to identify potential drug
105 candidates for glioma (Figure 1). Our approach is based on the hypothesis that
106 drugs/compounds with similar interactions across entire proteomes (“interaction sig-
107 natures”) are more likely to share therapeutic effects. Signatures were generated by
108 calculating interaction scores between every drug/compound and a comprehensive
109 library of proteins to capture the proteome-wide behaviors of a compound. Com-
110 pounds with interaction signatures closely matching those of drugs approved for glioma
111 were identified as potential treatments. We benchmarked performance by measuring
112 how frequently known drugs for a given indication were retrieved at various cutoffs
113 in ranked lists of predictions. Next, we compared our glioma specific results against
114 random controls, as well as across all indications. The novel predictions for glioma
115 were then corroborated through literature-based analysis to identify the highest con-
116 fidence drug candidates. Finally, we conducted a consensus analysis of proteins with
117 the strongest interactions to these novel glioma drug candidates which was further
118 corroborated using protein functional annotations.

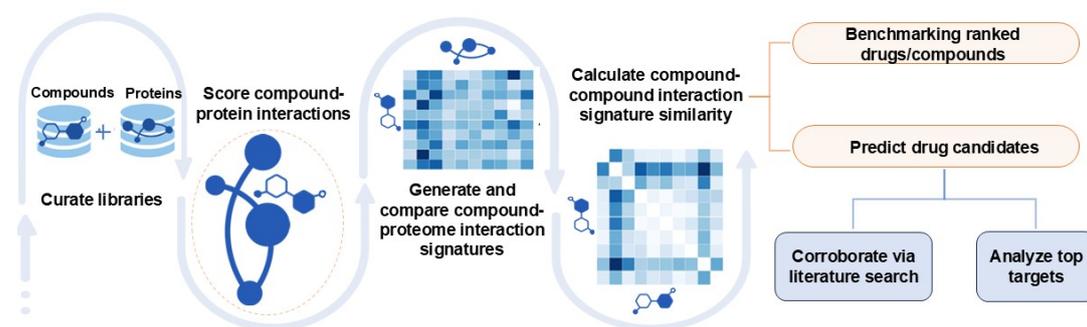


Fig. 1: Overview of pipeline for generating novel putative drug candidates for glioma within the CANDO multiscale drug discovery platform. Interaction scores between every protein and drug/compound in the corresponding libraries were calculated using our bioanalytical docking protocol (BANDOCK) [38–53]. This resulted in a compound-proteome interaction signature for each drug/compound describing its functional behavior. Interaction signature similarity was then calculated by comparing pairs of drug-proteome interaction signatures in an all-against-all manner. These interaction signature similarities were sorted and ranked for all drugs approved for an indication and used to benchmark performance and generate putative drug candidates. Benchmarking was conducted by measuring how approved drugs were recovered at various cutoffs. We performed a literature-based analysis to corroborate the glioma drug candidates for their potential to treat this disease. Finally, we identified the protein targets with the strongest interactions to these candidates and further corroborated them using protein functional annotations. The CANDO platform successfully identified multiple candidates demonstrating significant anti-glioma potential, offering a promising avenue to address the current lack of effective treatments for this disease.

119 2.2 Curating compound/protein libraries and indication 120 mapping

121 Our drug/compound library, sourced mainly from DrugBank [65], comprises 2,449
122 approved drugs and 10,741 experimental or investigational compounds, totaling 13,457
123 molecules. The “Homo sapiens AlphaFold2” (AF2) protein library was curated follow-
124 ing the application of the AlphaFold2 structure prediction program [66] to the Homo
125 sapiens proteome yielding 20,295 proteins used for this study. The Comparative Toxi-
126 cogenomics Database (CTD) was used to map the 2,449 approved drugs to 22,771
127 drug-indication associations based on DrugBank identifiers for drugs and compounds,
128 and Medical Subject Headings (MeSH) terms for approved/associated indications

129 [67, 68]. Benchmarking, which uses a leave-one-out approach (section 2.5), was carried
130 out on indications with at least two approved drugs, yielding a drug-indication map-
131 ping consisting of 1,595 indications and 13,226 associations. There were 35 associations
132 in our drug-indication mapping for the indication glioma (MeSH identifier: D005910).

133 2.3 Scoring compound-protein interactions and generating 134 interaction signatures

135 Interaction scores between each compound and protein were computed using our in-
136 house bioanalytical docking protocol (BANDOCK); these scores serve as a proxy for
137 binding strength/probability [38, 39, 41, 43, 46, 48]. Binding site prediction was first
138 performed using the COACH algorithm from the I-TASSER suite (version 5.1) [69].
139 COACH utilizes a library of protein structures bound to ligands, determined through
140 x-ray diffraction, to predict the binding sites and corresponding ligands for each pro-
141 tein based on structural and sequential similarity [70]. BANDOCK then calculates
142 interaction scores by comparing the COACH predicted ligands to the query compound,
143 using similarity between their Extended Connectivity Fingerprints with a diameter of
144 4 (ECFP4), generated via RDKit [71]. The chemical similarity score is quantified using
145 the Sorenson-Dice coefficient [72], which reflects the similarity between the query com-
146 pound and the predicted ligand. The highest chemical similarity score is multiplied
147 by the corresponding COACH binding site confidence score to assign an interaction
148 score between a compound and a protein by BANDOCK [38, 39, 41, 43, 46, 48]. BAN-
149 DOCK is applied between every compound and all proteins in the library, producing
150 compound-proteome interaction signatures describing (compound) behavior.

151 2.4 Calculating ranked compound similarity lists

152 CANDO calculates all-against-all similarities between compound-proteome interaction
153 signatures to compute drug repurposing accuracy and predict drug candidates [46].
154 We employed cosine distance for similarity calculations instead of the usual root-mean-
155 square deviation (RMSD) [53] as it enhanced computational speed while maintaining
156 performance. This process was repeated iteratively for all compound pairs in the
157 library, producing a ranked similarity list for each compound.

158 2.5 Benchmarking

159 Compounds are ranked by the number of times they appear in the similarity lists of the
160 associated drugs above a certain cutoff, resulting in a consensus list. We benchmarked
161 the performance of CANDO by evaluating the recovery of known/approved drugs
162 within similarity lists and aggregated consensus lists across various cutoffs using multi-
163 ple metrics. The consensus lists classify/rank compounds according to their consensus
164 scores, which reflect how frequently they appear in multiple similarity lists corre-
165 sponding to all approved drugs for an indication. As mentioned above (section 2.2),
166 we used drug-indication mappings from the Comparative Toxicogenomics Database
167 (CTD) [73] to determine the ranking of approved drugs within specific cutoffs (e.g.,
168 top 10, 25, 50, 100) in the similarity and consensus lists of drugs for a given indi-
169 cation with at least two approved drugs [38–53]. Benchmarking performance for all

170 indications, including glioma, was compared to a random control that calculated the
171 probability of correctly selecting an approved drug for an indication by chance, using
172 a hypergeometric distribution [51, 74].

173 CANDO calculates the following metrics developed in-house to assess performance:
174 indication accuracy, average indication accuracy, new indication accuracy, and new
175 average indication accuracy. Indication accuracy (IA) is the percentage of cases in
176 which at least one approved drug for a given indication appears within a specified
177 rank cutoff in the similarity list of another drug associated with that same indication.
178 Averaging the IA values for all indications with at least two approved drugs produces
179 the average indication accuracy (AIA). New indication accuracy (nIA) captures the
180 frequency with which approved drugs for a given indication appear within particu-
181 lar cutoffs in the consensus list for that indication. The nIA is averaged across all
182 indications to yield the new average indication accuracy (nAIA) metric.

183 CANDO also calculates the normalized discounted cumulative gain (NDCG) met-
184 ric, an evaluation measure commonly used in information retrieval to assess the
185 relevance of ranked items based on their positions [75, 76], to evaluate our predictions.
186 In CANDO, NDCG evaluates how effectively a given pipeline prioritizes approved
187 drugs for a specific indication within its similarity lists at specified cutoffs. The NDCG
188 score ranges from 0 to 1, with 1 indicating a perfect ranking [51]. Similarly, the
189 new NDCG (nNDCG) metric assesses the recovery of approved drugs across specified
190 cutoffs in the consensus list for an indication.

191 2.6 Generating drug predictions and corroborating them using 192 literature searches

193 The CANDO platform was applied to predict novel putative therapeutics for glioma
194 (MeSH identifier: D005910) which had 35 known associations in our drug-indication
195 mapping (section 2.2). As described above, drugs/compounds with interaction sig-
196 natures similar to those of drugs associated with glioma were ranked. Next, their
197 frequency, or consensus, among the similarity lists was used to identify the top 100
198 novel drug candidates for glioma. We conducted a literature review using search terms
199 that consisted of the name of each putative drug candidate and “glioma” in Google
200 Scholar and PubMed. We categorized the candidates as follows: *high-corroboration* for
201 drugs supported by two or more studies showing positive glioma treatment results;
202 *low-corroboration* for drugs targeting glioma-related pathways or supported by a sin-
203 gle positive study but lacking confirmation; and *no data found* when no data was
204 present to arrive at any conclusion regarding corroboration.

205 2.7 Analyzing top targets and associated pathways for glioma

206 We used our in-house top targets protocol to identify the proteins with the
207 strongest interactions with each putative drug candidate that was classified as
208 high-corroboration above. Interaction scores were calculated as described previously
209 (section 2.3) using the BANDOCK protocol, where higher scores (maximum of 1.0)
210 indicate stronger predicted interactions. We then conducted a literature search on

211 Google Scholar and PubMed with the names of the putative drug candidates and pro-
212 teins to find corroborative evidence supporting the target rationale used by CANDO
213 in generating predictions. We used this information to analyze whether the top targets
214 of the putative drug candidates overlap with proteins in biochemical pathways linked
215 to glioma.

216 **2.8 Assessing corroboration between protein functional** 217 **annotations and predicted top targets**

218 We curated three protein libraries, or “gold standards”, from UniProt [77], GeneCards
219 [78], and a comprehensive literature search to serve as references for evaluating the
220 target predictions for putative glioma treatment candidates. The literature search,
221 data presented in Table 3, focused on identifying targets implicated in glioma from
222 the top targets analysis for high-corroboration putative drug candidates generated
223 by the CANDO platform. The benchmarks assessed the overlap between the gold
224 standard libraries and the top protein targets predicted by CANDO for the top 24
225 drug candidate predictions. This assessment was repeated with a random set of 24
226 drug predictions, and the bottom 24 drug predictions as controls. The bottom 24 drug
227 predictions were filtered to include only compounds with at least five heavy atoms
228 to maintain meaningful molecular complexity. To quantify the alignment between the
229 gold standards and the predictions, we employed three key metrics: (A) frequency
230 distribution, (B) percentage overlap, and (C) the Jaccard coefficient, a commonly
231 used metric for assessing similarity across datasets [79, 80]. The Jaccard coefficient
232 calculates the ratio of the intersection and union of two groups and is defined as:

$$J(A, B) = \frac{|A \cap B|}{|A \cup B|}$$

233 In this context, A represents proteins annotated with glioma-related functions, and B
234 represents the top protein targets predicted by CANDO. A high Jaccard coefficient
235 indicates that CANDO accurately identifies protein targets that are independently
236 corroborated by functional annotation libraries.

237 Additionally, we compared the Jaccard coefficient across glioma and other disease
238 indications by using the top predicted targets from the top 24 or top 100 drug can-
239 didate predictions for glioma as one group, and functional protein annotations from
240 UniProt as the other group. Selected indications included cancer indications (e.g.,
241 metastatic melanoma, non-small cell lung cancer, acute myeloid leukemia) and non-
242 cancer diseases (e.g., Alzheimer’s disease, rheumatoid arthritis, asthma). Additionally,
243 we analyzed functional annotations for protein targets in the UniProt database to
244 assess their association with glioma and other disease indications. The Jaccard coeffi-
245 cient was computed separately for glioma-related targets and targets associated with
246 other diseases, including cancer indications (e.g., metastatic melanoma, non-small cell
247 lung cancer, acute myeloid leukemia) and non-cancer diseases (e.g., Alzheimer’s dis-
248 ease, rheumatoid arthritis, asthma). The comparison involved the predicted targets
249 from the top 24 and top 100 drug candidate predictions to evaluate performance
250 differences across indications.

251 3 Results

252 In summary, the results of this study provided strong evidence for the utility of the
253 CANDO platform in identifying putative drug candidates for glioma. The multitarget
254 approach enabled precise ranking and identification of compounds based on their
255 interaction signatures across the human proteome for treating glioma. The drug candidates
256 exhibited high interaction signature similarity to those of established glioma
257 treatments and were observed to target critical pathways associated with glioma patho-
258 genesis. Benchmarking and comparison with random controls affirmed the robustness
259 of the platform, indicating a high degree of predictive accuracy.

260 3.1 Benchmarking performance

261 Figure 2 illustrates the benchmarking performance of the CANDO platform for glioma
262 relative to all indications and random controls for both the similarity and consensus
263 lists (section 2.5). The approved drug library returned 35 associated drugs for glioma.
264 Figure 2A shows the AIA and nAIA metrics for all 1,595 indications with at least
265 two approved compounds. AIA ranged from 22% to 44% at the top 10, 25, 50, and
266 100 cutoffs, outperforming random control accuracies, which ranged from 4% to 26%.
267 CANDO achieved nAIA values ranging from 9% to 26% across the same cutoffs,
268 outperforming the random control for nAIA. The NDCG and nNDCG metrics for all
269 indications, presented in Figure 2B, further validate this performance. NDCG values
270 ranged from 0.044 to 0.059, while nNDCG values varied from 0.049 to 0.083, both
271 exceeding the random control for NDCG.

272 Figure 2C focuses on glioma-specific benchmarking, where CANDO demonstrated
273 enhanced IA values across all cutoffs compared to controls. The IA for glioma, evalu-
274 ated using similarity lists, ranged from 20% to 60% across the top 10 to top 100 cutoffs,
275 with a notable top 10 IA of 20%, which is nearly seven times the nIA of 3%. Figure 2D
276 presents the NDCG and nNDCG values for glioma-specific predictions. Glioma has
277 the same NDCG values at the top 10 and top 100 cutoffs, which are both 0.023, while
278 nNDCG values varied from 0.008 to 0.022. In comparison, random controls produced
279 substantially lower NDCG/nNDCG values. The IA/AIA metrics, applied to similarity
280 lists, and the nIA/nAIA metric, specific to consensus lists, collectively demonstrated
281 the robustness of the CANDO platform in leveraging interaction signature similarity
282 and consensus frequency to identify potential drug candidates effectively.

283 3.2 Identifying drug candidates

284 We used the CANDO platform to predict potential drug candidates for glioma (section
285 2.6). The 24 most compelling high-corroboration predictions based on ranking metrics
286 from the platform and literature analysis are shown in Table 2. The list of all the
287 top 100 putative drug candidates is given in **Supplementary Materials**. The top
288 ranked drug candidates were Vitamin D compounds: calcifediol, ergocalciferol, and
289 cholecalciferol. Additional drug candidates for glioma included taxanes (cabazitaxel),
290 vinca alkaloids (vinflunine), and topoisomerase inhibitors (topotecan).

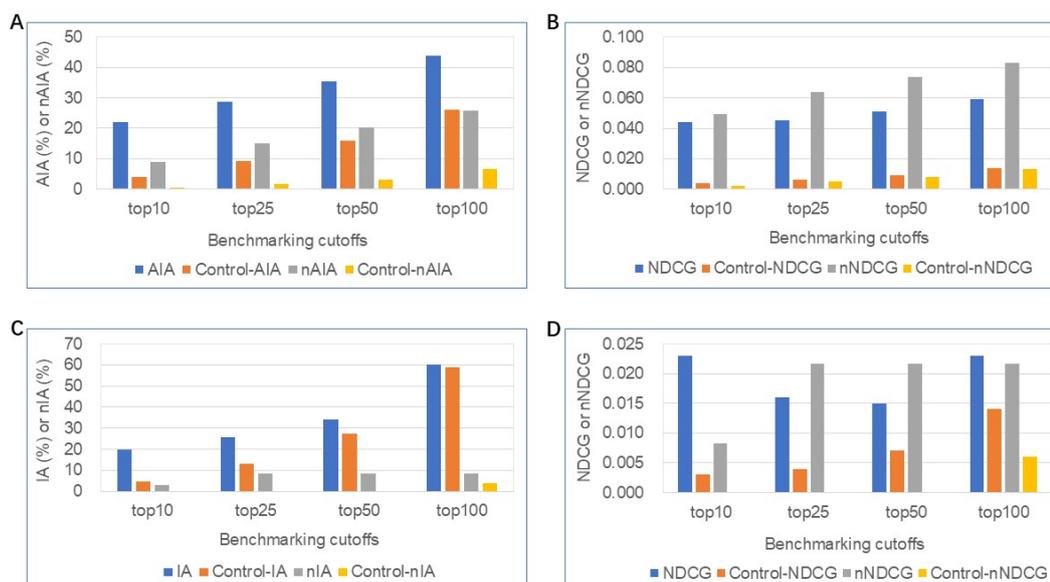


Fig. 2: Benchmarking performance of the CANDO platform for glioma relative to all indications and random controls. Performance was evaluated using (A) average indication accuracy (AIA)/new average indication accuracy (nAIA), as well as the normalized discounted cumulative gain (NDCG)/new normalized discounted cumulative gain (nNDCG) metrics across all indications (B). AIA and nAIA at the top 10, 25, 50, and 100 cutoffs, ranging from 22% to 44% and 9% to 26%, respectively, significantly outperform random controls; NDCG and nNDCG metrics, also significantly higher than random controls. In panels C/D, the indication accuracy (IA) and new indication accuracy (nIA) metrics, along with NDCG and nNDCG, were evaluated specifically for glioma. IA ranged from 20% to 60% at the top 10 to top 100 cutoffs, outperforming random controls; NDCG/nNDCG metrics were also higher than random controls. The results indicated that CANDO consistently outperforms random controls in identifying and prioritizing relevant compounds across all indications and glioma-specific predictions.

291 3.3 Analyzing targets and pathways related to glioma

292 The information considered when selecting putative drug candidates for novel treat-
293 ment included the top (i.e., strongest interaction) protein targets predicted by
294 CANDO, protein and pathway interactions corroborated using the literature, and stud-
295 ies of small molecules in the treatment of glioma observed in the literature 2.7. The
296 top targets predicted by CANDO are outlined in Table 3 and encompass Vitamin D3
297 receptor, thyroid hormone receptor, acetylcholinesterase, cyclin-dependent kinase 2,
298 tubulin alpha chain, dihydrofolate reductase, and thymidylate synthase. Among these,

Table 2: Predicted drug candidates for glioma using CANDO platform that were corroborated using literature analysis. The names of the 24 high-corroboration drug candidates (section 3.2), along with their ranks, consensus/average scores, and probability values are listed. The consensus score represents the number of drug–drug interaction signature similarity lists in which a compound appeared within a particular cutoff. The probability estimates the likelihood of a particular ranked compound appearing by chance, with lower values indicating a better outcome. The overall ranking of a potential drug is determined first by its consensus score and then by its average rank (section 2.6). The best ranked compounds in this consensus list are considered to be the top predictions for an indication. Vitamin D includes a group of compounds such as calcifediol, ergocalciferol, and cholecalciferol, which are ranked as the top three predictions with highest consensus score. This analysis indicates that the signature similarity pipeline within the CANDO platform can generate putative drug candidates for glioma.

Drug rank	Consensus score	Average score	Probability	Drug name	Drug rank	Consensus score	Average score	Probability	Drug name
1	4	28.5	2.68E-05	Calcifediol	51	3	35.3	4.22E-04	Ergosterol
2	4	30.2	2.68E-05	Ergocalciferol	55	3	38.3	4.22E-04	Vinblastine
3	4	37.5	2.68E-05	Cholecalciferol	56	3	38.3	4.22E-04	Lanosterol
6	3	9.0	4.22E-04	Cabazitaxel	63	3	44.0	4.22E-04	Brivaracetam
10	3	10.3	4.22E-04	Docetaxel	64	3	44.3	4.22E-04	Loperamide
25	3	22.0	4.22E-04	Calcitriol	65	3	44.3	4.22E-04	Ginsenosides
26	3	22.3	4.22E-04	Tacalcitol	66	3	47.0	4.22E-04	Gimatecan
29	3	23.0	4.22E-04	Vinflunine	73	3	49.7	4.22E-04	Ortaxel
31	3	23.3	4.22E-04	Vinorelbine	74	3	50.0	4.22E-04	Resiniferatoxin
32	3	23.3	4.22E-04	Folic acid	78	3	51.7	4.22E-04	Irinotecan
49	3	32.7	4.22E-04	Topotecan	80	3	52.7	4.22E-04	Chrysin
50	3	33.0	4.22E-04	Calcipotriol	95	3	66.0	4.22E-04	Cryptotanshinone

299 the strongest interaction was observed between the Vitamin D3 receptor and calcife-
300 diol, with a BANDOCK score of 0.850. Figure 3 highlights various important related
301 pathways implicated in the pathogenesis of glioma, including phosphatidylinositol-
302 3'-kinase (PI3K)/Akt, mammalian target of rapamycin (mTOR), and Janus kinase
303 (JAK)/signal transducer and activator of transcription (STAT) pathways.

304 Insert figure 3 here...

305 3.4 Determining overlap between protein functional 306 annotations and CANDO predicted top targets

307 Figure 4A illustrates the frequency distribution of overlaps between our three gold
308 standard protein libraries and the top protein targets predicted by CANDO. For all
309 gold standards, targets of the top 24 drug candidates showed the highest proportion
310 of overlaps within the highest ranked bin (1-20). In contrast, targets from the ran-
311 dom 24 drug candidates and bottom 24 drug candidates exhibited a comparatively

312 uniform distribution across the 5 bins. Figure 4B presents the cumulative percentage
313 overlap as a function of rank cutoff for the predicted targets across the gold stan-
314 dard libraries. The targets from the top 24 drug candidate predictions demonstrated
315 a near-saturation of overlap at lower rank cutoffs (e.g., 80% overlap by rank 20 for
316 Table 3 and UniProt), emphasizing their strong alignment with gold standard targets.
317 In contrast, the targets from the random 24 and bottom 24 drug candidate predictions
318 exhibited a slower increase in overlap percentage, with cumulative overlaps remain-
319 ing below 10% even at a rank cutoff of 100 for Table 2 and GeneCards. As shown in
320 Figure 4C, the Jaccard coefficient values further corroborate the findings from the fre-
321 quency distribution and overlap percentage analyses. Across all libraries, the Jaccard
322 coefficient for the top protein targets from the top 24 drug candidate predictions was
323 consistently higher compared to those derived from the random 24 or bottom 24 drug
324 candidate predictions.

325 We found that the Jaccard coefficient for top (rank ≤ 10) predicted targets of the
326 top 24 and top 100 drug candidate predictions for glioma was higher when compared
327 to UniProt glioma protein functional annotations (Figure 5). In contrast, the Jaccard
328 coefficient was lower when comparing glioma targets to protein functional annotations
329 for other indications. Indications demonstrating a lower Jaccard coefficient include
330 other cancer indications such as non-small cell lung cancer and metastatic melanoma,
331 as well as non-cancer diseases like Alzheimer’s disease and rheumatoid arthritis. This
332 result suggests that the top predicted glioma targets identified by CANDO are more
333 functionally relevant to glioma-related gold standard protein targets than those of
334 other indications, highlighting the effectiveness of the pipeline in identifying mean-
335 ingful targets. When compared to the broader rank distribution shown in the earlier line
336 plot (Figure 4), the rank ≤ 10 results highlight the ability of the pipeline to capture
337 high-confidence and/or known targets for glioma. This trend underscores the utility
338 of using stringent rank cutoffs to identify highly specific target overlaps, particularly
339 for glioma.

340 Insert figure 4 here...

341 Insert figure 5 here...

342 Insert table 3 here...

343 4 Discussion

344 CANDO identified potential glioma treatments that included drugs approved for
345 other indications such as vitamin D (calcifediol), taxanes (cabazitaxel and docetaxel),
346 vinca alkaloids (vinblastine and vinflunine), topoisomerase inhibitors (topotecan and
347 irinotecan), and folic acid. Additionally, investigational compounds like ginsenosides,
348 brivaracetam, chrysin, resiniferatoxin, and cryptotanshinone were also identified as
349 promising drug candidates (Table 2). Literature-based analysis was conducted to
350 corroborate these potential drugs and compounds for glioma, examining supporting
351 evidence for their targets and pathways (Table 3 and Figure 3). The top drug candi-
352 dates generated via the interactomic signature pipeline of CANDO may be exerting
353 their therapeutic effects by impacting multiple pathways implicated in glioma. We
354 examined the top drugs/compounds and targets predicted by CANDO in further

355 detail, comparing and contrasting to what is known about their relevance to glioma
356 in the literature; a detailed description follows below.

357 **4.1 Vitamin D3 receptor metabolites**

358 Vitamins may have a role in the etiopathogenesis of central nervous system (CNS)
359 cancers [108]. Vitamin D comprises a group of fat-soluble steroids, with vitamin
360 D3 (cholecalciferol) and vitamin D2 (ergocalciferol) being the most significant [109].
361 Calcifediol (25-hydroxyvitamin D3), is the precursor for calcitriol, the active form
362 of vitamin D [110]. Recent research suggests that the levels of the progenitor of
363 calcitriol correlate with progression of glioma [111–114]. Cholecalciferol has shown
364 promise in glioma treatment, especially glioblastoma multiforme (GBM), due to its
365 ability to regulate cell cycle biomarkers and enhance the anti-tumor immune response
366 [85, 115]. Studies indicate that vitamin D analogs, including ergocalciferol, could
367 modulate biomarkers involved in cell cycle regulation and apoptosis in glioblastoma
368 [115]. Cell cycle arrest is one of the most well-studied mechanisms accounting for the
369 anti-tumor activity of vitamin D in gliomas. Vitamin D has been shown to induce anti-
370 glioma effects through cell cycle arrest and the phosphoinositide 3-kinase (PI3K)/Akt
371 pathway [87].

372 **4.2 Taxanes**

373 Taxanes are a class of diterpenes commonly used as chemotherapy agents, mainly
374 including cabazitaxel, docetaxel and paclitaxel [116–118]. Cabazitaxel is a second-
375 generation semisynthetic taxane. Contrary to other taxane compounds, cabazitaxel is
376 a poor substrate for P-gp efflux pump; therefore, it easily penetrates across the BBB
377 [119, 120]. Cabazitaxel shows a significant inhibitory effect on glioma [121, 122]. Other
378 studies have reported that cabazitaxel exerts its anti-proliferative effects on cancer
379 cells by binding to tubulin [123]. One study indicates that tubulin alpha-1C chain
380 (TUBA1C) may potentially regulate the pathogenesis of glioma through Janus kinase
381 (JAK)/signal transducer and activator of transcription (STAT) (JAK-STAT) pathway
382 [124]. Docetaxel, a taxane-class anti-mitotic agent, demonstrates the ability to induce
383 cell apoptosis in glioma and shows substantial inhibitory activity against tumor growth
384 [125]. Furthermore, it is recognized as one of the leading drug candidates for brain
385 tumor therapy [126]. In our study, both cabazitaxel and docetaxel are predicted to
386 strongly interact with TUBA1C, with predicted interaction scores of 0.716 and 0.790,
387 respectively (Table 3).

388 **4.3 Vinca alkaloids**

389 Vinca alkaloids are a class of chemotherapy agents with anti-mitotic and anti-
390 microtubule properties, including compounds such as vinflunine, vinorelbine, vin-
391 blastine, and vincristine [127–129]. Vinflunine, a fluorinated vinca alkaloid, disrupts
392 microtubule dynamics, a process essential for cell division, and has shown potential
393 for glioma treatment [130, 131]. Vinorelbine, a semi-synthetic vinca alkaloid, is an
394 anti-mitotic chemotherapy drug used to treat various cancers, including breast cancer,
395 non-small cell lung cancer, and glioma [132]. Its antitumor effect arises from its ability

396 to inhibit mitosis by interacting with tubulin [133]. In 2000, a pilot study of weekly vin-
397 blastine in patients with recurrent low-grade gliomas (LGG) yielded promising results
398 [134, 135]. Compared to vinflunine and vinorelbine, vinblastine demonstrated a higher
399 interaction score with the TUBA1C target (Table 3).

400 4.4 Topoisomerase I inhibitors

401 Topoisomerase inhibitors are chemical compounds that block the action of topoisom-
402 erases, which are broken into two broad subtypes: type I topoisomerases (TopI) and
403 type II topoisomerases (TopII) [136, 137]. TopI inhibitors, like topotecan, are water-
404 soluble camptothecin analogs that have shown cytotoxicity toward a variety of tumor
405 types [138]. Topotecan can pass through the BBB and exhibits significant activity
406 in treating glioblastoma multiforme [139, 140]. Additionally, it has been observed to
407 induce cell cycle arrest at the G0/G1 and S phases [90, 141]. Irinotecan (CPT-11), a
408 prodrug of 7-Ethyl-10-hydroxycamptothecin (SN-38), shows some antitumor activity
409 in patients with recurrent glioblastoma multiforme, with response rates of 0 to 17%
410 in several trials [142, 143]. Gimatecan is a lipophilic oral camptothecin analogue with
411 preclinical activity in glioma models [144].

412 4.5 Folic acid

413 Folic acid (FA) targets the folate receptor (FR), which is overexpressed on the cell
414 surface of various cancer cells [145, 146]. Folate supplementation, particularly at high
415 doses, has been suggested to have cytotoxic effects on glioma cells, making it a poten-
416 tial candidate for further exploration in glioma therapies [147]. In addition, utilizing
417 lidocaine liposomes modified with folic acid has been demonstrated to suppress the
418 proliferation and motility of glioma cells [148]. One clinical research study explored
419 the role of folate-related compounds, such as L-methylfolate, in combination thera-
420 pies for glioma, showing potential epigenetic modifications and enhanced sensitivity
421 to standard treatments like temozolomide [149]. Zhao, et al. [101] hypothesized that
422 inhibition of dihydrofolate reductase/thymidylate synthase might modulate the cell
423 sensitivity of glioma cells to temozolomide through the mTOR signaling pathway.
424 DHFR and TYMS are key metabolic enzymes in the folic acid signaling pathway, with
425 high predicted interaction scores of 0.755 and 0.688, respectively, to folic acid in this
426 study (Table 3).

427 4.6 Other drug candidates and key target interactions

428 Ginsenosides, active components found in Panax ginseng, show potential in glioma
429 treatment due to their various therapeutic properties, including anticancer and neu-
430 roprotective effects [150, 151]. Additionally, ginsenoside has been shown to inhibit the
431 growth of human glioma U251 cells, promoting apoptosis and affecting key signaling
432 pathways involved in cell survival and death [152]. Additionally, compounds such as
433 brivaracetam, which lack enzyme-inducing activity on the cytochrome system, could
434 be considered promising candidates for addressing brain tumor-related epilepsy [153].
435 Chrysin, an active natural bioflavonoid, is predicted to target cyclin-dependent kinase
436 1 with an interaction score of 0.655 (Table 3), and has been proven to protect against

437 carcinogenesis [107]. Cyclin-dependent kinase is the target for glioma cell cycle arrest
438 at G2 and M phases [154]. Chrysin exerts anticancer activity in glioblastoma cell
439 lines possibly via the ERK/Nrf2 signaling pathway [155]. Resiniferatoxin, a naturally
440 occurring irritant tricyclic diterpene which combines structural features of the phorbol
441 ester tumor promoters and of capsaicin [156]. It activates transient vanilloid receptor
442 (TRP), which was previously associated primarily with cardiovascular and neuronal
443 regulation, but might also present avenues for exploration in glioma pathogenesis [157].
444 We observed evidence of interaction between resiniferatoxin and the thyroid hormone
445 receptor beta target (Table 3 and Figure 3). Cryptotanshinone is one of the main rep-
446 resentative components isolated from the roots of *Salvia miltiorrhiza*. Lu, et al. [158]
447 indicated that cryptotanshinone can inhibit human glioma cell proliferation.

448 As shown in Table 3 and Figure 3, we predicted interactions between topotecan,
449 irinotecan, and cryptotanshinone and acetylcholinesterase (AChE), a newly recognized
450 marker for glioma (Table 3). One study reported that irinotecan or its metabolites
451 directly interact with AChE, inhibiting the conversion of acetylcholine to choline,
452 which leads to an accumulation of acetylcholine and subsequent cholinergic syndrome
453 symptoms [159]. Bioinformatic analysis has shown that AChE is connected to proteins
454 in the PI3K/Akt pathway, which promotes anti-apoptotic and proliferative effects in
455 brain tumors [94]. There is limited evidence of interactions between topotecan, irinote-
456 can, cryptotanshinone, and AChE; our study therefore provides predictive evidence of
457 these interactions. Additionally, predictions of topotecan and resiniferatoxin targeting
458 thyroid hormone receptor beta are novel to this study. The thyroid hormone recep-
459 tor influences glioma progression by regulating the PI3K/Akt signaling pathway [92].
460 Therefore, candidates targeting the PI3K/Akt pathway may hold promise for glioma
461 treatment.

462 Our protein list, derived from predictions, highlights glioma-relevant targets but is
463 inherently incomplete, similar to databases like UniProt or GeneCards, as each cap-
464 tures only a partial view of glioma biology. While this list serves as one of the curated
465 gold standards for our analysis, incorporating known treatment targets in future stud-
466 ies could provide a more comprehensive benchmark. Limitations of this study include
467 the arbitrary rank cutoffs, which may exclude moderately ranked targets that over-
468 lap meaningfully with gold standard libraries, and the use of the Jaccard coefficient,
469 a binary metric that overlooks relative ranks or prediction scores. Additionally, our
470 focus on glioma leaves the robustness of this approach across other indications under-
471 explored, particularly for diseases with fewer validated targets. Finally, the analyses
472 may bias toward frequently predicted top targets, underrepresenting less common
473 targets with potential therapeutic value. To address these limitations, future stud-
474 ies will integrate score-based cutoffs, and consider a broader range of rank and score
475 distributions.

476 5 Conclusions

477 We utilized our CANDO platform to explore potential novel treatments and their
478 associated protein targets for glioma. By integrating a combination of computa-
479 tionally generated and experimentally observed data from benchmarking, prediction,

480 corroboration of putative drug candidates using literature-based searches, top protein
481 target analysis, and protein functional annotation, we identified promising treat-
482 ments for glioma, including Vitamin D, taxanes, vinca alkaloids and topoisomerase
483 inhibitors. Additionally, we highlighted several protein targets and related path-
484 ways linked to glioma, including Vitamin D3 receptor, thyroid hormone receptor,
485 acetylcholinesterase, cyclin-dependent kinase 2, tubulin alpha chain, dihydrofolate
486 reductase and thymidylate synthase. This study offers insights into the potential mech-
487 anisms underlying glioma and demonstrates the potential of the CANDO platform in
488 identifying effective treatments against this disease.

489 **Acknowledgements.** The authors would like to acknowledge the Center for Com-
490 putational Research (CCR) at University at Buffalo for computational support. We
491 would also like to thank all members of the Samudrala Computational Biology Group.

492 **Declarations**

- 493 • **Funding:** This work was supported in part by a National Institutes of Health (NIH)
494 Director’s Pioneer Award (DP1OD006779), a NIH Clinical and Translational Sci-
495 ences (NCATS) Award (UL1TR001412), a NIH National Library of Medicine (NLM)
496 T15 Award (T15LM012495), a NIH NLM R25 Award (R25LM014213), a NIH
497 NCATS ASPIRE Design Challenge Award, a NIH NCATS ASPIRE Reduction-to-
498 Practice Award, a National Institute of Standards of Technology (NIST) Award
499 (60NANB22D168), a NIDA Mentored Research Scientist Development Award
500 (K01DA056690), and startup funds from the Department of Biomedical Informatics
501 at the University at Buffalo.
- 502 • **Competing interests:** The funders had no role in the design of the study; in the
503 collection, analyses, or interpretation of data; in the writing of the manuscript, or
504 in the decision to publish the results. The authors have formed multiple startups
505 that seek to commercialise the outputs of the CANDO platform.
- 506 • **Ethics approval and consent to participate:** Not applicable
- 507 • **Consent for publication:** Not applicable
- 508 • **Data availability:** Data including compound-protein interaction matrices are avail-
509 able upon request.
- 510 • **Materials availability:** Not applicable
- 511 • **Code availability:** Not applicable
- 512 • **Author contribution:** SM.X. helped conceive the research project and pipelines,
513 conducted all analyses, and drafted the manuscript. W.M., A.E., M.V.N., and YK.H.
514 helped data analyses. Z.F. and R.S. conceived of the research design, approach, and
515 methods, supervised the overall project, and edited the manuscript. All authors
516 have read and agreed to the published version of the manuscript.

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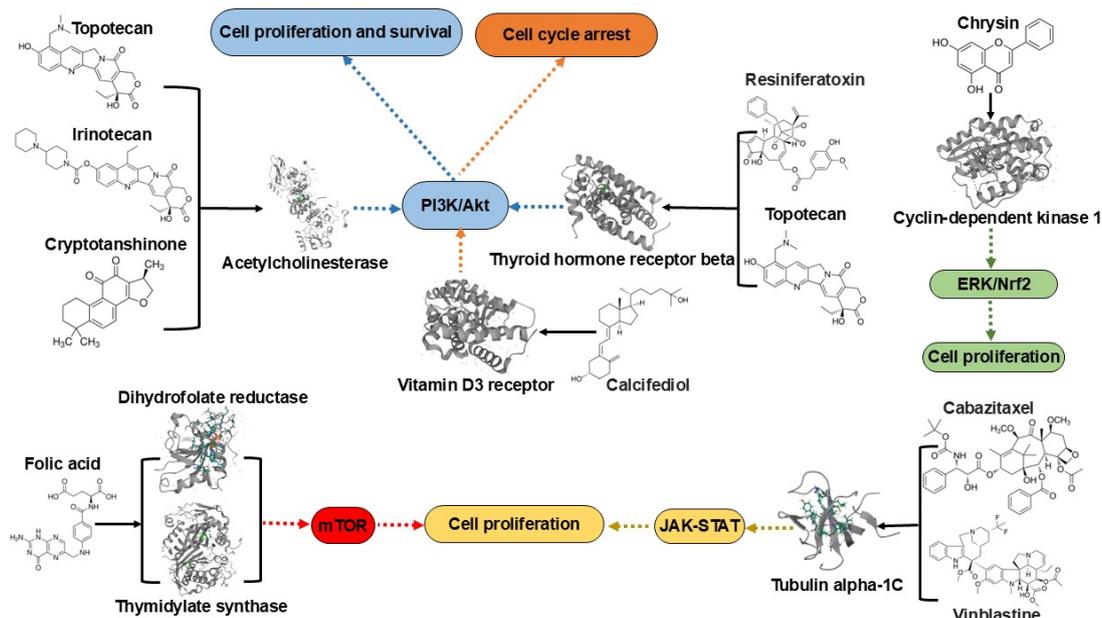


Fig. 3: Downstream pathways of top targets for putative drugs for glioma treatment predicted by CANDO. The top targets for putative drugs for glioma are those with the strongest interactions as predicted by our CANDO platform (Table 3) and verified by a functional annotation search (section 3.4). The phosphatidylinositol-3'-kinase (PI3K)/Akt, mammalian target of rapamycin (mTOR), Janus kinase (JAK)/signal transducer and activator of transcription 3 (STAT3) pathways play important roles in the biology of malignant gliomas [81–84]. Topotecan, irinotecan, and cryptotanshinone all interact with acetylcholinesterase (ACHE), ranking 8th (topotecan and irinotecan) and 1st (cryptotanshinone), respectively. Compounds resiniferatoxin and topotecan strongly interact with the thyroid hormone receptor beta (THRB). The targets ACHE and THRB both influence glioma cell proliferation and survival by regulating the PI3K/Akt signaling pathway (blue). Cell cycle arrest is one of the most well-studied mechanisms accounting for the anti-tumor activity of vitamin D in gliomas. The compound chrysin interacts with cyclin-dependent kinase 1 (CDK1), targeting glioma cell proliferation via the ERK/Nrf2 signaling pathway (green). Dihydrofolate reductase (DHFR) and thymidylate synthase (TYMS) are key targets of folic acid, modulating glioma cell proliferation through the mTOR signaling pathway (red). Taxanes (e.g., cabazitaxel) and vinca alkaloids (e.g., vinblastine) interact with tubulin alpha-1C, influencing glioma through the JAK-STAT pathway (yellow). Our study allows for comprehensive mechanistic understanding of drug candidate behavior across multiple scales, showcasing the CANDO platform's capability to accurately identify novel drug candidates and their mechanisms through a multifaceted strategy.

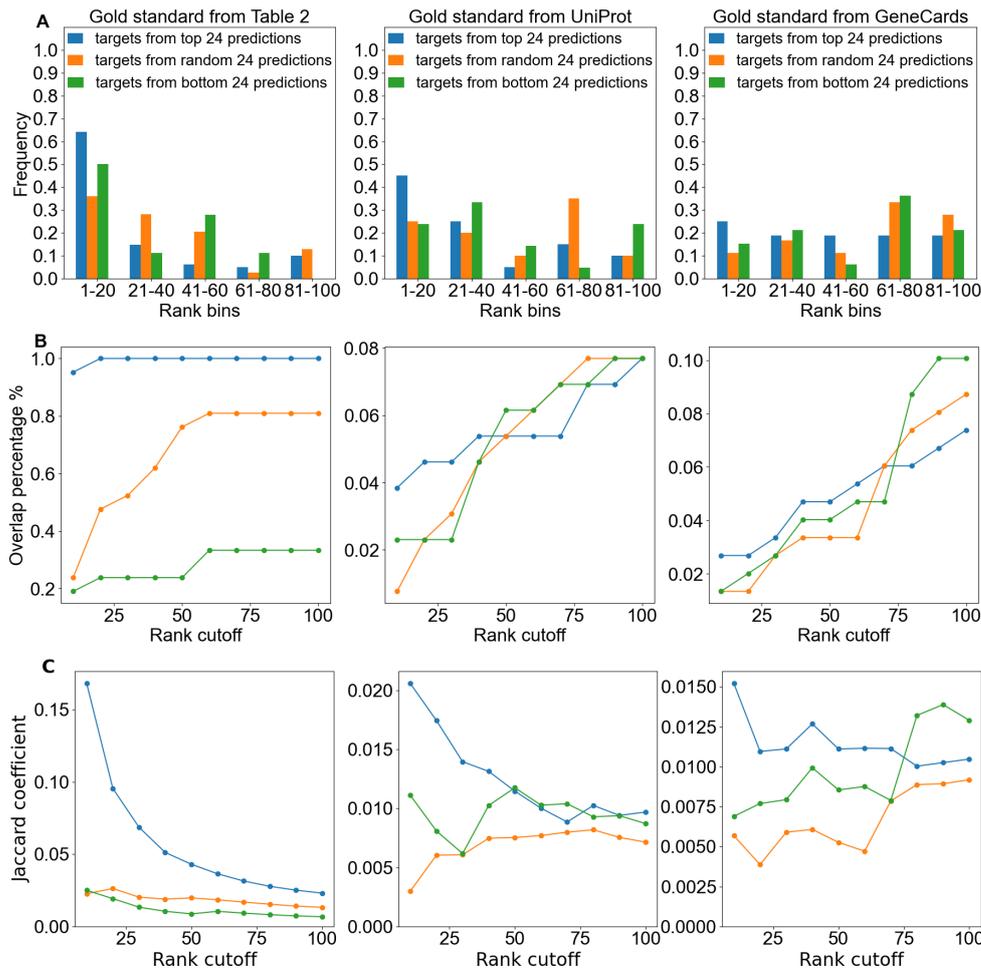


Fig. 4: Overlap between protein functional annotations and CANDO predicted top targets across gold standard libraries. This figure compares the overlap between CANDO predicted targets and gold standard annotations (Table 3, UniProt, and GeneCards) for glioma-related proteins. A) Frequency distributions show the proportion of predicted targets that overlap with gold standard proteins within rank bins for top 24 drug candidate predictions (blue), random 24 drug candidate predictions (orange), and bottom 24 drug candidate predictions (green). B) The line graphs show the percentage of gold standard proteins that overlap with prediction targets as a function of rank cutoff (from 10 to 100). C) Targets from the top 24 drug candidate predictions demonstrate a higher Jaccard coefficient compared to from random 24 and bottom 24 drug candidate predictions across all gold standards. The Jaccard coefficient quantifies the similarity between CANDO predicted targets and gold standard targets. Each column corresponds to a different gold standard: Table 3 (left), UniProt (center), and GeneCards (right). Results demonstrate targets from the top 24 drug candidate predictions generally reflect a stronger signal compared to that of random or bottom drug candidate predictions, highlighting the predictive accuracy of the top candidates.

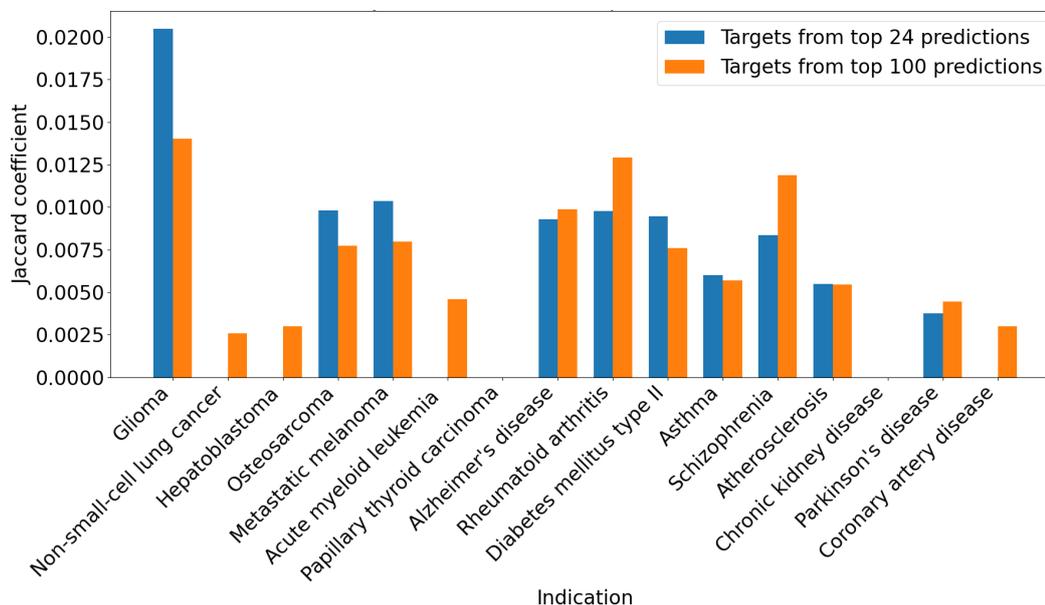


Fig. 5: Overlap between protein functional annotations and CANDO predicted top targets across indications. This bar chart compares the overlap between targets with the strongest predicted interactions to our top drug candidates and existing protein annotations across glioma and other indications, using the Jaccard coefficient (vertical axes). The Jaccard coefficient quantifies the overlap between protein functional annotations (from UniProt) and CANDO-predicted drug targets. Two comparisons are made: the overlap with top 24 drug candidate predictions (blue) and the overlap with top 100 drug candidate predictions (orange). A higher coefficient indicates stronger alignment between the predicted and known targets. The horizontal axis lists the various indications, including cancers (glioma, hepatoblastoma, metastatic melanoma) and non-cancer conditions (diabetes mellitus type II, coronary artery disease, and chronic kidney disease). Results show that the Jaccard coefficient for glioma is notably higher than that of other indications, highlighting the effectiveness of the CANDO platform in identifying glioma-related protein targets.

Table 3: Top targets analysis for high-corroboration putative drug candidates for glioma generated by the CANDO platform. The name of the high-corroboration drug candidates, their predicted ranks based on consensus scoring (section 2.6), the rank of the target from the top targets analysis (section 2.7), the target UniProt identifier, the predicted interaction score between these predictions and targets, the target name, and the evidence we found for the target being implicated in glioma, are listed. Higher scores indicate a higher likelihood of interaction. From this analysis, we highlighted Vitamin D3 receptor, thyroid hormone receptor, acetylcholinesterase, cyclin-dependent kinase 1, tubulin alpha chain, dihydrofolate reductase and thymidylate synthase as the most promising targets for glioma.

Drug rank	Drug name	Target rank	Target identifier	Score	Target name	Evidence
1	Calcifediol	1	P11473	0.850		Yuan, et al. (2023) [85]
2	Ergocalciferol	32	P11473	0.605	Vitamin D3 receptor	Diesel, et al. (2005) [86]
3	Cholecalciferol	3	P11473	0.740		Sze-Ching Lo, et al. (2022) [87]
6	Cabazitaxel	13	P68363	0.725	Tubulin alpha-1B chain	
		19	Q9BQE3	0.716	Tubulin alpha-1C chain	
10	Docetaxel	13	P68363	0.800	Tubulin alpha-1B chain	
		19	Q9BQE3	0.790	Tubulin alpha-1C chain	
73	Ortataxel	13	P68363	0.580	Tubulin alpha-1B chain	
		19	Q9BQE3	0.572	Tubulin alpha-1C chain	Liu, et al. (2018) [88]
29	Vinflunine	2	P68363	0.672	Tubulin alpha-1B chain	
		18	Q9BQE3	0.633	Tubulin alpha-1C chain	Hu, et al. (2022) [89]
31	Vinorelbine	2	P68363	0.669	Tubulin alpha-1B chain	
		18	Q9BQE3	0.631	Tubulin alpha-1C chain	
55	Vinblastine	2	P68363	0.765	Tubulin alpha-1B chain	
		18	Q9BQE3	0.721	Tubulin alpha-1C chain	
		2	P11387	0.565	DNA topoisomerase I	
		3	P06276	0.373	Cholinesterase	
49	Topotecan	4	P10828	0.371	Thyroid hormone receptor beta	
		5	P02768	0.370	Albumin	
		8	P22303	0.359	Acetylcholinesterase	Zhang, et al. (2013) [90]
		9	P27487	0.358	Dipeptidyl peptidase 4	Ma, et al. (2020) [91]
		97	O00763	0.301	Acetyl-CoA carboxylase 2	Zhang, et al. (2021) [92]
					Li, et al. (2023) [93]	
		2	P11387	0.459	DNA topoisomerase I	Obukhova, et al. (2021) [94]
		3	P27487	0.392	Dipeptidyl peptidase 4	Tsuji, et al. (2024) [95]
66	Gimatecan	4	P14174	0.389	Macrophage migration inhibitory factor	Jones, et al. (2017) [96]
		7	P15559	0.373	NAD(P)H dehydrogenase quinone 1	Guo, et al. (2017) [97]
		10	O00763	0.364	Acetyl-CoA carboxylase 2	Lei, et al. (2020) [98]
		2	P11387	0.466	DNA topoisomerase I	
		8	P22303	0.390	Acetylcholinesterase	
78	Irinotecan	37	O00763	0.324	Acetyl-CoA carboxylase 2	
		47	P15559	0.317	NAD(P)H dehydrogenase quinone 1	
		1	P48449	0.960	Lanosterol synthase	
56	Lanosterol	9	Q9Y6A2	0.437	Cholesterol 24-hydroxylase	Nguyen, et al. (2023) [99]
		10	P02768	0.435	Albumin	Han, et al. (2020) [100]
65	Ginsenosides	1	P48449	0.610	Lanosterol synthase	Li, et al. (2023) [93]
		4	P02768	0.503	Albumin	
		1	P00374	0.755	Dihydrofolate reductase	Zhao, et al. (2019) [101]
32	Folic acid	3	Q04609	0.723	Glutamate carboxypeptidase 2	Kunikowska, et al. (2022) [102]
		6	P04818	0.688	Thymidylate synthase	Ding, et al. (2015) [103]
63	Brivaracetam	1	P24941	0.390	Cyclin-dependent kinase 2	Liu, et al. (2022) [104]
		2	Q9Y6A2	0.522	Cholesterol 24-hydroxylase	Han, et al. (2020) [100]
64	Loperamide	5	Q92769	0.467	Histone deacetylase 2	Was, et al. (2019) [105]
		24	O00763	0.440	Acetyl-CoA carboxylase 2	Jones, et al. (2017) [96]
74	Resiniferatoxin	2	P10828	0.425	Thyroid hormone receptor beta	Zhang, et al. (2021) [92]
		8	Q96GR4	0.402	Palmitoyltransferase ZDHHC12	Lu, et al. (2022) [106]
80	Chrysin	7	P06493	0.655	Cyclin-dependent kinase 1	Jiang, et al. (2022) [107]
95	Cryptotanshinone	1	P22303	0.473	Acetylcholinesterase	Obukhova2021, et al. (2021) [94]
		7	P15559	0.362	NAD(P)H dehydrogenase quinone 1	Lei, et al. (2020) [98]