DOI: 10.1002/jmv.25432

RESEARCH ARTICLE

WILEY MEDICAL VIROLOGY

Prevalence and characteristics of acute respiratory virus infections in pediatric cancer patients

Nadia Soudani^{1,2,3} | Miguela A. Caniza^{4,5} | Aia Assaf-Casals^{2,6} | Rouba Shaker^{2,6} | Mireille Lteif^{2,6} | Yin Su⁷ | Li Tang⁷ | Imad Akel^{2,6} | Samar Muwakkit^{6,8} | Ahmad Chmaisse^{1,2} | Maysam Homsi⁵ | Ghassan Dbaibo^{2,6} | Hassan Zaraket^{1,2}

¹Department of Experimental Pathology, Immunology and Microbiology, American University of Beirut Faculty of Medicine, Beirut, Lebanon

²Center for Infectious Diseases Research, American University of Beirut Faculty of Medicine, Beirut, Lebanon

³Department of Biology, Faculty of Sciences, EDST, Lebanese University, Lebanon

⁴Department of Infectious Diseases, St Jude Children's Research Hospital, Memphis, Tennessee

⁵Department of Global Pediatric Medicine, St Jude Children's Research Hospital, Memphis, Tennessee

⁶Department of Pediatrics and Adolescent Medicine, American University of Beirut Faculty of Medicine, Beirut, Lebanon

⁷Department of Biostatistics, St Jude Children's Research Hospital, Memphis, Tennessee

⁸Children's Cancer Center of Lebanon, American University of Beirut, Beirut, Lebanon

Correspondence

Ghassan Dbaibo and Hassan Zaraket, American University of Beirut, PO Box 11-0236, Riad El-Solh, Beirut, Lebanon. Email: hz34@aub.edu.lb; gdbaibo@aub.edu.lb

Funding information

American University of Beirut Faculty of Medicine; Children's Infection Defense Center (CIDC) fund, St. Jude Children's Research Hospital, Grant/Award Number: N/A

Abstract

Background: Patients with pediatric cancer have a higher risk of morbidity and mortality because of respiratory viral infections than other patient populations.

Objectives: To investigate the causative viruses of respiratory infections and their burden among patients with pediatric cancer in Lebanon.

Study design: Nasopharyngeal swabs along with clinical and demographic data were collected from patients with pediatric cancer presenting febrile episodes with upper respiratory tract symptoms. Total nucleic acid was extracted from specimens followed by the real-time PCR analysis targeting 14 respiratory viruses to estimate the frequency of infections.

Results: We obtained 89 nasopharyngeal swabs from patients with pediatric cancer (mean age, 5.8 ± 4.2 years). Real-time PCR confirmed viral infection in 77 swabs (86.5%). Among these, 151 respiratory viruses were detected. Several viruses cocirculated within the same period; respiratory syncytial virus (RSV) being the most common (45.45%), followed by parainfluenza virus (PIV; 26%), influenza type B (26%), human metapneumovirus (24.6%), and human coronavirus (HCoV; 24.6%). Coinfections were detected in 55% of the subjects, and most of them involved RSV with one or more other viruses. A strong correlation was found between PIV, Flu (influenza of any type), RSV, and HCoV with the incidence of coinfections. RSV was associated with lower respiratory tract infections, nasal congestion, bronchitis, and bacteremia. HCoV was associated with bronchiolitis; rhinovirus was associated with hospital admission. **Conclusion:** Patients with pediatric cancer have a high burden of respiratory viral infections and a high incidence of coinfections. Molecular diagnostics can improve management of febrile episodes and reduce antibiotic use.

KEYWORDS

coinfection, patients with pediatric cancer, prevalence, real-time PCR, respiratory tract infections, virus infections

1 | INTRODUCTION

Immunocompromised patients, such as those with cancer and hematopoietic stem cell transplantation, have a higher risk for respiratory infections,¹ and single or mixed respiratory viruses are frequently detected in those with acute respiratory symptoms.²⁻⁶ Defects in innate and adaptive immunity³ coupled with damage in the mucosal membrane and frequent exposure to a healthcare environment contribute to increased morbidity^{7,8} and mortality of respiratory infections in these patients.^{2,3,7,9-11} In healthy children, respiratory viruses are usually confined to the upper respiratory tract; in immunocompromised patients, progression to the lower respiratory tract is a more frequent and feared complication.^{2,12,13} Despite advances in cancer therapy and outcomes during the last decade, respiratory viral infections and complications are frequent barriers to the success of antineoplastic treatment.^{11,12,14}

Respiratory infections are major causes of febrile episodes in patients with pediatric cancer.³ These patients often are initiated on broad-spectrum antibiotics to cover serious bacterial diseases, leading to unnecessary increased exposure to antibiotics and the potential emergence of antibiotic resistance.^{3,15,16} Accurate respiratory viral diagnosis and early access to treatment can improve outcomes, allow the prompt initiation of infection control measures, and limit antibiotic use.¹⁰ Molecular diagnostic assays for respiratory virus detection and identification are becoming increasingly popular because they outperform traditional viral detection methods, such as antigen detection and cell culture-based assays in terms of speed, efficiency, specificity, and sensitivity.^{10,12} Altogether, these facts highlight the need for surveillance studies that utilize molecular diagnostic tools to elucidate the role of viral pathogens during respiratory infections, their risk factors, and outcomes.^{12,17-19}

Epidemiological studies of respiratory viral infections in patients with pediatric cancer in low-resource settings are scarce.^{3,11,17-21} The main purpose of this study was to screen for viral etiologic agents and associated risk factors and complications during respiratory infections in patients with pediatric cancer in Lebanon to address the joint need for surveillance studies using molecular diagnostic tools and additional studies of respiratory infections in developing countries.

2 | MATERIALS AND METHODS

2.1 | Study design and data collection

Between October 2014 and December 2015, nasopharyngeal swabs were collected from cancer patients with acute respiratory tract infection (ARTI) at the Children's Cancer Center in Lebanon. Patients were considered eligible for this study if they were patients with pediatric cancer having febrile episodes with upper respiratory tract symptoms. The inclusion criteria were as follows: age <18 years, received cancer treatment within the last three months, fever ≥38°C within the previous 72 hours, and having one or more of the following symptoms: cough, sore throat, nasal congestion, rhinorrhea, or respiratory distress. The following data were obtained from enrolled participants: age, sex, demographics, use of influenza vaccines and antivirals, date of onset of symptoms, and hospital admission and length of stay. Medical charts were reviewed to determine health complications, bacterial infection, and absolute neutrophil and lymphocyte counts. Another chart review was performed one month after the initial febrile illness to monitor potential complications. The project was approved by the institutional review boards at the American University of Beirut and St. Jude Children's Research Hospital. Parental informed consent and participant assent, when applicable, was obtained.

2.2 | Sample collection and screening

Nasopharyngeal swabs from each patient were collected by health care providers, preserved in virus transport media, and transported to the laboratory for further analysis. Viral nucleic acid was extracted by using PureLink Viral RNA/DNA mini Kit (Invitrogen, Carlsbad, CA). The AgPath-ID One-Step RT-PCR kit (Applied Biosystems, Austin, TX) was used to screen extracted RNA samples for 14 respiratory viruses: human metapneumovirus (HMPV), respiratory syncytial virus (RSV), influenza A virus (Flu A), influenza B virus (Flu B), rhinovirus (RhV), adenovirus (AdV), parainfluenza viruses 1 to 4 parainfluenza virus (PIV1-4), and human coronaviruses (HCoV-HKU1, HCoV-229E, HCoV-OC43, and HCoV-NL63). The sequences of primers and probes were obtained from the Center for Disease Control and Prevention (CDC). All runs were performed in the presence of a notemplate control (NTC) and positive control for each target. Extraction controls were screened to exclude cross-contamination during extraction. Flu A-positive samples were further subtyped via real-time PCR using the CDC-established protocol. RSV-positive samples were subtyped by conventional PCR followed by 1.5% gel electrophoresis using RSV-A and RSV-B primers specific for the G gene hypervariable region.^{22,23}

2.3 | Statistical analysis

The univariate regression analysis was performed to determine the association between viral mono- and coinfections with variables and outcomes, including demographics, hospital/ICU admission, lower respiratory tract infection (LRTI), and other clinical symptoms, such as bronchitis, fever, mechanical ventilation, nasal congestion, respiratory distress, vomiting, neutropenia (absolute neutrophil count [ANC] < 1500 cells/µL), and lymphopenia (absolute lymphocyte count < 2000 cells/ μ L). The χ 2 test and odds ratio were computed to test the association between the categorical variables. All variables that were statistically associated with severe outcomes in the univariate models were included in multivariate logistic regression using the backward selection method; a significance level of 0.10 or less was required for a covariate to stay in the model. Then, odds ratio estimates with P values for tested variables were monitored after adjusting for age and sex. Similarly, the correlation analysis between variables was also included, and correlation coefficients

MEDICAL VIROLOGY -WILEY

3 | RESULTS

3.1 | Characteristics of the study population

During the 14-month study period, 89 febrile episodes were recorded in 67 individual patients. The median age of patients was 4.5 years (IQR 3-8 years), and 54% of the patients were male (Table 1); 31.5% of the patients had solid tumors, and 68.5% had liquid tumors (Table 1). The most prevalent respiratory symptoms in this population were cough (84.3%), rhinorrhea (85.4%), and nasal congestion (74.2%). In our sample population, 33.3% of the children were admitted to the hospital.

significant. Statistical analysis was performed using SAS 9.4 software.

3.2 | Prevalence and seasonal distribution of respiratory viruses

A total of 151 respiratory viruses were detected in 86.5% (77/89) nasal swabs obtained from 67 patients with pediatric cancer presenting fever. Most patients (55%, n = 49) had coinfections with two or more viruses; 31.5% (n = 28) had monoinfections (ie, a one-virus infection). According to the chart review, none of the patients had coinfection with bacteria. RSV was the most common virus in the 77 febrile episodes with at least one detected virus, followed by PIV, Flu B, and HMPV. The most prevalent viruses in the 28 monoinfections were HMPV, RSV, and RhV, whereas the most commonly detected viruses in the 49 coinfections were RSV, FluB, and PIV3 (Figure 1). Respiratory viral infections were detected mainly during the winter season (December to March) and, to a lesser extent, during the spring (Figure 2). Sporadic infections were detected during the summer and fall seasons. Several respiratory viruses were cocirculating during the same period.

The influenza vaccination rate was relatively low: only 44.7% (n = 30) of the patients with febrile episodes who were eligible to receive the vaccine (ie, age > 6 months; n = 67) did. Of this vaccineeligible group, 22% (n = 15) had Flu A and 28% (n = 19) had Flu B. In the vaccinated group, 43.3% (n = 13) had influenza A and/or B infection; 16.7% (n = 5) had both influenza A and B.

3.3 | Risk factors and clinical outcomes of ARTI with viral etiology

The univariate analysis was used to assess the association between demographic variables, clinical findings, and respiratory viruses with ARTI (Table 2). Children younger than 2 years were at a significantly higher risk of developing respiratory distress than were those aged 2 to 6 years (P < 0.01, OR, 0.077 [CL 0.015-0.400]) or those older than 6 years (P = 0.0053, OR, 0.086 [CL 0.015-0.482]). Neutropenia (ANC < 1500 cells/µL) was identified as a risk factor for hospital admission (P = 0.0192, OR, 3.625 [CL 1.234-10.65]). No statistically

significant association was observed with respect to sex, cancer type and treatment, lymphopenia, or antiviral drug administration and the tested variables. The type of cancer, receiving an anticancer drug, neutropenia, and lymphopenia was not associated with an increased risk of coinfection. In addition, the presence of viral coinfection did not seem to correlate with any of the recorded clinical symptoms (Table 2).

Next, the association of the detected viruses with complications and coinfection was assessed. To account for the small sample size, genotypes or subtypes of the same virus were grouped together (eg, influenza virus for Flu A and B). RhV was significantly associated with hospital admission (P = 0.023, OR, 4.343 [CL 1.214-15.532]), whereas RSV was significantly associated with LRTI (P = 0.0493, OR, 4.316 [CL 1.005-18.544]) and nasal congestion (P=0.0387, 5.357 [1.091-26.300]). HCoV was significantly associated with bronchiolitis (P = 0.049, OR, 12.223 [CL 1.002-149.166]), whereas PIV and HMPV were significantly associated with nasal congestion (P = 0.01, OR, 0.205 [CL 0.057-0.728]). PIV, influenza virus, RSV, and HCoV were significantly associated with coinfection (P < 0.01, OR, 8.522 [CL 1.754- 41.411]; P = 0.0039, OR, 6.158 [CL 1.792-21.159]; P < 0.01, OR, 10.679 [CL 3.075-37.079]; P=0.0283, OR, 10.741 [CL 1.287-89.605], respectively). Among these, RSV had the strongest correlation with coinfection (Table 2).

Variables were further analyzed via multivariate logistic regression analysis with the selection method of backward elimination, whereby a *P* value of 0.1 was required for a covariate to stay in the model. Children aged 2 to 6 years and those older than 6 years had lower odds (94.8%, and 95.2%, respectively) of having respiratory distress than those younger than 2 years (P < 0.01). The odds of having a coinfection was 27.3 times greater (P < 0.001, OR, 27.3 [CL 2.8-268]) in patients with influenza than in those without influenza. Furthermore, the patients with RSV infections were 70.7 times more likely to have a coinfection (P < 0.001, OR, 70.7 [CL7.4-678]) than

TABLE 1 Baseline characteristics of enrolled pediatric cancer

 patients with ARTI

Patient characteri	stics	n (N = 89)	%
Age, y	0-2	15	16.9
	2-6	41	46.1
	>6	33	37.1
Sex	Male	48	54
	Female	41	46
Clinical findings	Fever	89	100.0
	Bacterial coinfection	1	1.1
	Pneumonia	4	4.5
	Respiratory distress	19	21.3
	Cough	75	84.3
	Rhinorrhea	76	85.4
	Nasal congestion	66	74.2
	Sore throat	15	16.9
	Vomiting	10	11.2
	Diarrhea	5	5.6
Tumor type	Solid tumor	28	31.5
	Liquid tumor	61	68.5

Abbreviation: ARTI, acute respiratory tract infection

1194 WILEY - MEDICAL VIROLOGY

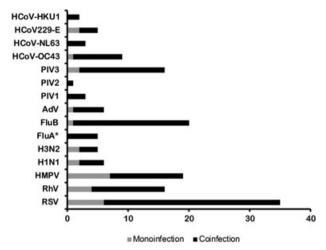


FIGURE 1 Viruses detected in the respiratory specimens. The frequency of respiratory viruses detected among cancer patients with monoinfections (n = 28) or coinfections (n = 49; 123 detected viruses among coinfections). (FluA*: influenza A viruses that were not subtyped). AdV, adenovirus; Flu A, influenza A virus, Flu B, influenza B virus, HCoV, human coronaviruses; HMPV, human metapneumovirus; PIV 1 to 4, parainfluenza viruses; RhV, rhinovirus; RSV, respiratory syncytial virus

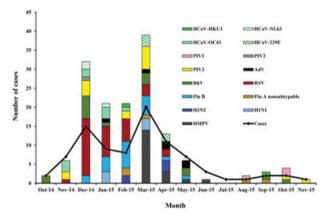


FIGURE 2 Monthly distribution of the ARTI cases and the detected viruses. Respiratory viruses were detected throughout the year, with a peak in winter. AdV, adenovirus; ARTI, acute respiratory tract infection; Flu A, influenza A virus; Flu B, influenza B virus; HCoV, human coronaviruses; HMPV, human metapneumovirus; PIV 1 to 4, parainfluenza viruses; RhV, rhinovirus; RSV, respiratory syncytial virus

those with the other tested viruses. PIV remained significantly associated with coinfection (P < 0.001, OR, 41.5 [CL 3.3-514.5]). Infection with RSV remained significantly associated with nasal congestion (P = 0.046). The odds of having nasal congestion were 5.1 (CL [1.03-25.3]) times greater for patients with RSV than for those without RSV infection. After adjusting for age and sex, the estimates and significance did not change.

The correlation analysis showed that RSV (r = 0.5, P < 0.0001), influenza virus (r = 0.37, P < 0.0001), PIV (r = 0.36, P < 0.001), and HCoV (r = 0.32, P < 0.001) were significantly associated with coinfection. AdV (r = 0.23, P = 0.0648) showed a moderate relationship

with coinfection. Neutropenia was positively associated with RhV (r = 0.34, P = 0.0053), but PIV had a negative association (r = -0.21, P = 0.095). RSV was positively correlated with HCoV (r = 0.29, P = 0.0161) but negatively correlated with HMPV (r = -0.39, P < 0.01; Figure 3A). Hospital admission and neutropenia were significantly correlated (r = 0.4, P < 0.001), as were hospital admission and RhV (r = 0.29, P = 0.018). Diarrhea was significantly associated with HCoV (r = 0.49, P < 0.0001) and with RSV (r = 0.26, and P = 0.0312). Nasal congestion was also significantly correlated with RSV (r = 0.27, P = 0.0256). HCoV (r = 0.22, and P = 0.0681) had a moderate relationship with respiratory distress (Figure 3B).

The follow-up data revealed that recurrent fever was significantly related to the previous RSV infection (r = 0.33, P < 0.01) and moderately correlated with solid tumors (r = 0.22, P = 0.0821). Rhinorrhea was positively related to the previous RhV infection (r = 0.25, P = 0.040) but negatively correlated with lymphopenia (r = -0.28, P = 0.025). Both bronchitis (r = 0.26, P = 0.031) and bacteremia (r = 0.26, P = 0.0312) were significantly related to the prior RSV infection during the follow-up period. LRTI was significantly associated with RSV (r = 0.25, P = 0.0384) but moderately associated with coinfections (r = 0.21, P = 0.0901). Moreover, positive correlations were observed between bronchiolitis and previous HCoV (r = 0.29, P = 0.016) and RSV infections (r = 0.26, P = 0.031), whereas URTI was moderately correlated with the previous PIV infection (r = 0.22, P = 0.074; Figure 3C).

3.4 | Repeated detection of respiratory viruses and RSV genotyping

During the study period, 17 of the 67 patients (25.4%) had repeated febrile episodes, which often led to the detection of the same virus (es) associated with the first episode (Table 3). The median time between repeated febrile episodes with a confirmed viral infection was 61 days (range 10-196 days). Repeated detection of the same virus was noted in nine patients. RSV was most frequently detected (6/17) in the repeated episodes. Using genotype-specific PCR to detect RSV showed that three of these paired samples had RSVA; for the remaining three paired specimens, a genotype could not be determined in at least one of the specimens. We also detected one case for each of the repeated episodes of HCoV-OC43, Flu B, RhV, PIV3, AdV, and HMPV.

4 | DISCUSSION

This study demonstrates that respiratory viruses are a leading cause of acute respiratory infections in pediatric cancer patients and should be considered an important etiology of febrile neutropenia and hospital admissions. The data showed that viral infections were associated with 86.5% of the febrile episodes in patients with pediatric cancer. Detection of a respiratory viral infection in this population has multiple implications, including persistence of infection and disease reactivation. These findings also highlight the urgent

I									\$	MED	ICAI	L VIROLO	GI	**ILI	
	C	<u>م</u>	0.8	0.8	0.6	0.5	0.5	0.4	0.5916	0.9	0.99	0.8		0.008	0.004
	Coinfection	OR [95% CI]	0.8 [0.19-3.372]	0.788 [0.176-3.526]	0.762 [0.29-2.004]	1.719 [0.417-7.082]	1.944 [0.303- 12.469]	0.633 [0.216-1.855]	1.314 [0.485-3.559]	1.123 [0.305-4.136]		1.167 [0.329-4.131]		8.522 [1.754- 41.411]	6.158 [1.792- 21.159]
		۹.	0.2	0.8	0.4	0.96	0.97	0.4	0.83	0.5	0.99	0.5		0.6	0.4
	LRTI	OR [95% CI]	0.258 [0.031- 2.125]	1.333 [0.220- 8.099]	1.920 [0.488- 7.549]			1.778 [0.440- 7.191]	20.861 [0.218, 3.398]	2.000 [0.227- 17.633]		0.465 [0.053- 4.062]		1.451 [0.328- 6.420]	0.463 [0.090- 2.390]
	c	۹.	0.29	0.44	0.34	0.94	0.76	0.18	0.0192	0.24	0.99	0.61		0.8	0.39
	Hospital admission	OR [95% CI]	2.526 [0.457- 13.964]	2 [0.342- 11.703]	0.599 [0.209-1.719]	1.055 [0.237-4.697]	0.75 [0.116- 4.856]	2.146 [0.708-6.507]	3.625 [1.234- 10.65]	2.647 [0.519- 13.492]		0.691 [0.164- 2.915]		0.856 [0.255-2.869]	1.598 [0.548-4.66]
	c	۹.	0.95	0.87	0.65	0.96	0.97	0.87	0.73	0.45	0.99	0.96		0.7	0.94
		P OR [95% CI]	0.002	0.005 0.818 [0.066- 10.196]	0.173 0.567 [0.049- 6.568]	0.341	0.833	0.669 1.222 [0.104- 14.339]	0.605 0.648 [0.056, 7.529]	0.327 0.385 [0.032-4.658]	0.984	0.92		0.583 1.633 [0.138- 19.294]	0.877
	Respiratory distress	OR [95% CI]	0.077 [0.015-0.400]	0.086 [0.015-0.482]	0.437 [0.133-1.438]	2.857[0.329- 24.790]	1.277 [0.132- 12.320]	0.756 [0.209-2.729]	0.736 [0.231, 2.345]	0.500 [0.125- 1.999]		1.077 [0.253-4.578]		0.675 [0.166-2.748]	0.909 [0.272-3.039]
	ć	e.	0.35	0.46	0.75	0.49	0.977	0.89	0.32	0.34	0.99	0.69		0.01	0.13
	Nasal congestion	OR [95% CI]	0.347 [0.038-3.178]	0.422 [0.043-4.165]	0.828 [0.255-2.691]	0.469 [0.053-4.123]		0.912 [0.243-3.421]	1.929 [0.527-7.061]	0.350 [0.041-3.015]		1.395 [0.269-7.246]		0.205 [0.057-0.728]	0.395 [0.118-1.319]
		<u>م</u>	0.03*	0.26	0.78	0.56	0.967	0.09**	0.453	0.41	0.985	0.9		0.92	0.08
	Fever	OR [95% CI]	0.15 [0.026- 0.85]	0.395 [0.079- 1.962]	1.2 [0.344- 4.188]	1.913 [0.216- 16.932]		3.077 [0.844- 11.212]		0.533 [0.118- 2.408]		0.9 [0.17-4.76]		1.077 [0.253- 4.578]	0.147 [0.018- 1.224]
		Patient characteristics	2-6 y vs 0-2 y	>6 y vs 0-2 y	Sex (female vs male)	Treatment in last 3 mo	Anticancer drug	Tumor (Solid vs liquid)	Neutropenia (Yes vs no)	Lymphopenia	Antiviral drug	Family fever in last 8 d	Virus detected	Parainfluenza virus	Influenza virus

TABLE 2 Risk factors and clinical outcomes of patients with respiratory infections

1195

	_
1	nea
;+;	Ē
č	5
ç	٧
Ц	ų
5	
2	ť
H	-

	Clinical Outcomes												
Patient characteristics	Fever OR [95% CI] P	Nasal congestion OR [95% CI]	ط	Respiratory distress OR [95% CI]	ď	Bronchiolitis OR [95% CI]	ط	Hospital admission OR [95% CI]	ط	LRTI OR [95% CI] 1	d	Coinfection OR [95% CI]	٩
Adenovirus	0.97	0.780 [0.075-8.129]	0.84	1.067 [0.103- 11.032]	0.957		0.98		0.97		0.97		0.97
Rhinovirus	1.5 0.59 [0.343- 6.561]	3.805 [0.451- 32.122]	0.22	1.556 [0.407-5.949]	0.518	0.518 2.167 [0.181- 25.903]	0.54	4.343 [1.214- 15.532]	0.02	1.045 ([0.194- 5.628]	0.96	2.089 [0.573-7.612]	0.3
Respiratory syncytial 6.167 virus [1.486 25.58	6.167 0.01 [1.486- 25.586]	. 5.357 [1.091- 26.300]	0.04	1.684 [0.543-5.227]	0.367		0.95	2.24 [0.786-6.385]	0.13	4.316 ([1.005- 18.544]	0.049	0.049 10.679 [3.075- 37.079]	0.0002
Human coronavirus	1.958 0.38 [0.434- 8.833]	i 1.227 [0.233-6.453]	0.81	3.409 [0.877- 13.251]	0.08	0.08 12.223 [1.002- 149.166]	0.049	0.049 1.814 [0.485-6.779]	0.38	2.625 ([0.560- 12.309]	0.2	10.741 [1.287- 89.605]	0.03
Human metapneumovirus	0.242 0.19 [0.029- 2.043]	0.205 [0.057-0.728]	0.01	1.083 [0.294-3.989]	0.904		0.95	0.408 [0.102-1.634]	0.21	0.311 ([0.036- 2.667]	0.3	1.481 [0.468-4.687]	0.5
*Statistically significant values (P < 0.05) are indicated in bold. **Borderline values ($P \gtrsim 0.05$) are presented in bold italics.	llues (P < 0.05) are ir .05) are presented ii	idicated in bold. n bold italics.											

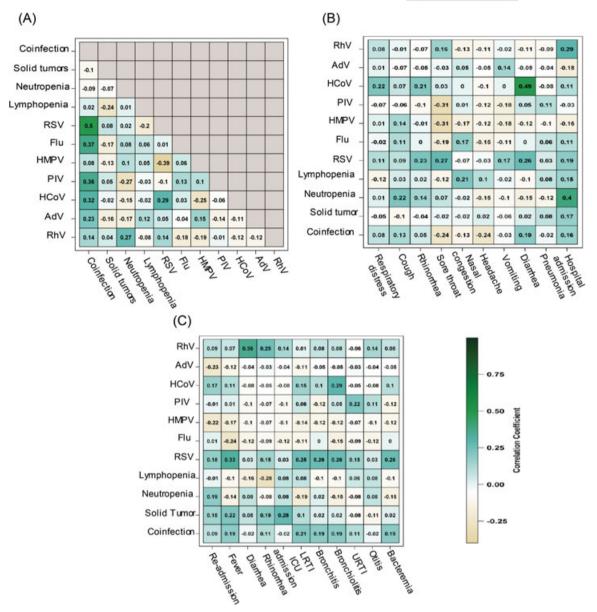


FIGURE 3 Heat maps representing (A) correlation matrix of infections (n = 67), (B) correlation between infections and events (n = 67), and (C) correlation between infections and events in follow-up (n = 67). Negative correlations are shown in yellow, and positive correlations are shown in green. Neutropenia was defined as absolute neutrophil count < 1500 cells/µL, and lymphopenia was defined as lymphocyte count < 2000µL. AdV, adenovirus; Flu, influenza; HCoV, human coronaviruses; HMPV, human metapneumovirus; PIV 1 to 4, parainfluenza viruses; RhV, rhinovirus; RSV, respiratory syncytial virus

need for vaccines and effective drugs against the commonly circulating viral pathogens, which will limit unnecessary administration of antivirals and antibacterials, and best management of isolation precautions of infected patients.

A substantial subset (55%) of our study population had coinfections with multiple viruses. We did not find any association between coinfections and clinical symptoms/outcome or having neutropenia or lymphopenia in our patients. Consistent with our findings, Torres et al¹⁸ found no difference in clinical outcome in children with febrile neutropenia having mixed respiratory viral infections and those with monoinfections. Similar findings were reported by Rotzén-Östlund et al²⁴ who found that immunocompetent children with mixed or

monoinfections had a similar duration of hospitalization, risk of pediatric ICU admission, and need for oxygen. However, other studies in immunocompetent children found that coinfection was associated with more-severe lower respiratory tract infection and a higher risk of hospital admission than monoinfection was.^{25,26} A meta-analysis of 21 studies involving 4280 immunocompetent patients found no evidence of increased severity as a result of coinfections compared with monoinfections.²⁷ Therefore, the clinical implications of respiratory viral coinfections need further investigation.

Most studies investigating respiratory viral infections in immunocompromised children were carried out in developed countries. In Sweden, respiratory viral infections were detected in 45% of cases of

TABLE 3 Viruses detected among patients with multiple febrile episodes

	Virus(es) detect	ed in each sampl	le			
Patient ID#	1	2	3	4	5	Interval between specimen collection (days)
1	RhV	*				
	RSVA	HCoV-OC43				73
	HCoV-229E	RSV				82
	PIV3	PIV1				196
10	HCoV-OC43	RSV				
	RSVA	HCoV-OC43				16
12	RSVA	RhV	PIV3			
	RSVA					12
13	RSVA	RhV				
	RSVA	FluB	FluA			51
17	RSVA	RhV	FluB	PIV2	PIV3	
	RhV	PIV3				80
18	RSVA	PIV3				
	HMPV					72
	HMPV	RhV	HCoV-229E			11
						22
21	RSVA	RhV				
		FluB	H1N1			10
	RSVA	FluB	H3N2			40
23	RSVA					
	HCoV-229E					89
26	RSV**	AdV				
	AdV					133
28	RSV	FluB	HCoV-OC43			
	HMPV	RSV				65
31	RSV	FluB	PIV3			
	HCoV-229E					57
32	RSV	FluB	FluA	HCoV-OC43	HCoV-HKU1	
	RSVB					50
35						
						43
40	HMPV	Flu B	PIV3			
-						127
42	 HMPV	PIV3				
	AdV	FluB				70
45	HMPV					
	FluB					37
62	HMPV					<u>.</u>
52	HCoV-HEKU1	 RhV	 FluA			76

Abbreviations: AdV, adenovirus; Flu A, influenza A virus; Flu B, influenza B virus; HCoV, human coronaviruses; HMPV, human metapneumovirus; PIV 1 to 4, parainfluenza viruses; RhV, rhinovirus; RSV, respiratory syncytial virus

*No additional virus was detected in this specimen.

**Subtype could not be determined.

febrile neutropenia in children with mixed malignancies.¹⁹ A similar rate of respiratory viral infections in children with leukemia or mixed malignancies was reported in the USA. Germany, Finland, and Chile.^{2,18,28,29} In contrast, in Spain, respiratory viral infections were detected in only 12% of patients with cancer, and all viral infections were monoinfections.¹¹ In the mentioned studies, RSV, RhV, and Flu A were the most commonly detected viruses.^{2,3,11,18,19,29} In our study group, RSV was the most prevalent virus, followed by PIV, and Flu B, with HMPV being the most frequent virus in monoinfections and RSV detected predominantly in coinfections. The higher rate of virus detection in our study population in Lebanon could be attributed to the use of real-time PCR, the higher number of screened viruses, and including respiratory symptoms as an inclusion criterion. Some studies identified respiratory viruses via one or more of the following techniques: clinical symptoms and serology testing, virus culture, fluorescent antibody detection, and PCR for limited viral targets.^{2,11,18,19,28,29} Nonetheless, even when compared with studies that utilized PCR as a diagnostic tool to screen a comparable number of viral targets (10 or more) in patients with cancer, this study has a higher incidence of virus detection.^{3,17-19} This higher incidence of viral infection might be attributed to the cultural interaction that includes close contact, thus promoting transmission.^{30,31}

This study reflects the importance of using molecular diagnostic assays in clinical practice for better diagnosis and improving patient care.³²⁻³⁴ It is important to note that mutations in the region of the detection primers could result in reduced detection rates by PCR. Therefore, continuous monitoring of the circulating virus strains and updating of the detection primers to capture any novel, variant strains is necessary.

Accurate information on the frequency of respiratory viruses and their burden allows for accurate recommendations in prioritizing drug and vaccine development. For instance, children receiving chemotherapy developed life-threating complications when infected with RSV.^{11,35} Previous studies in patients with pediatric cancer reported that RSV can be associated with pneumonia and neutropenia leading to severe life-threatening complications.^{11,17,35,36} Mucositis was very common in patients with neutropenia, and this might be one of the predictive factors of the severity of respiratory viral infections, especially those because of RSV.^{11,17,37} The data showed that RSV was significantly associated with LRTI, fever, bronchiolitis, nasal congestion, and diarrhea but did not increase the risk of ICU admission. Similar to what was previously reported, the data suggests that encountering RSV infection in patients with pediatric cancer might add to their disease burden and affect their response to the treatment.^{35,38-40} Moreover, this study revealed a high prevalence of RSV in coinfections and in repeated infections in patients with pediatric cancer, highlighting the urgent need for the development of RSV antivirals and vaccines. Currently, RSV vaccines under development are in phase III clinical trials, constituting a step in the right direction.41-43 In infants at high-risk (preterm or having chronic illness), RSV neutralizing monoclonal antibodies, including the FDAapproved palivizumab, are effective for prophylaxis but remain very expensive in developing countries.44-46 The emergence of RSV

variants that are resistant to palivizumab is a concern that highlights the need for more therapeutic options.^{47,48} Prophylaxis should be utilized in patients with cancer, especially in those with the highest risk for infection (in this study, younger children), to reduce potential complications.

In patients with cancer, influenza infections are usually accompanied by bacteremia and delays in chemotherapy.^{49,50} A study conducted by Mendoza Sánchez et al¹¹ showed that 40% of immunocompromised children (cancer and HIV; age \leq 14 years) infected with influenza required hospitalization. Such outcomes would be effectively reduced by annual vaccinations, which are now recommended for patients and their health care providers. In this study, infection with influenza A and/or B was detected in 26.9% (24/89) of the cases. The vaccination rate among the children \geq 6 months (recommended age for the vaccine) with febrile episodes was 44.7%, which is considered to be low for this high-risk population. Therefore, emphasizing the importance of attaining universal vaccination in this population and their caregivers is likely to improve patient outcomes by preventing influenza infection.

The duration of shedding and frequency of recurrent virus detections have not been thoroughly investigated in immunocompromised children.^{19,35} In the current study, 17 of the 67 patients had repeated detection of respiratory viruses in subsequent febrile episodes, and half of these patients had repeated detection of the same virus. RSV was the most common virus detected repeatedly in subsequent febrile episodes. It was not possible to rule out whether these infections were persistent or repeated infections because this study only included patients in whom a febrile episode developed rather than monitoring all patients on a routine basis. Hall et al³⁵ reported prolonged RSV shedding that persisted for more than 20 days in some cases.³⁵ Martin et al⁵¹ reported prolonged shedding of respiratory viruses in samples collected at least 7 days apart: the viruses involved in prolonged shedding included HMPV, RSV, AdV, HCoV, RhV, and PIV. A study by Soderman et al¹⁹ suggested that the shedding time of Flu, HMPV, RSV, and PIV is limited, and virus clearance was evident at a median follow-up time of 28 days. In contrast, the same RhV genotypes were detected from subsequent follow-up samples after 12 to 51 days, and some RhV-positive patients reported the appearance of respiratory symptoms 6 days or more before fever onset, indicating the possibility of prolonged shedding. Longitudinal studies whereby patients are screened on a weekly basis until no detection of a given virus is confirmed are required to determine the frequency of repeated infections vs persistent infections among children with cancer and their impact on patient outcomes. A molecular approach using sequencing of the sequentially detected viruses should allow a better understanding of the extent of virus evolution in these patients.

The current study had several limitations, including the heterogeneity of cancer types in the studied population and the absence of a control group of patients with asymptomatic cancer. Another limitation was the lack of follow-up sampling to monitor the status of the detected respiratory virus and differentiate between prolonged vs persistent shedding. Moreover, human bocavirus and enteroviruses were not Y-MEDICAL VIROLOGY

analyzed, which could have further increased the overall detection rate. Despite the mentioned limitations, screening for respiratory viruses, and their related clinical manifestations is of great importance, especially in scarcely studied populations and particularly in developing countries, to guide better patient management and to develop evidence-based infection control measures.

5 | CONCLUSION

Respiratory infections can lead to serious complications and might become life-threatening, particularly in the context of weakened immune systems of patients with cancer. Few studies have investigated respiratory infections in patients with cancer, especially in children. In this study, we detected a high incidence of respiratory viral infections among children with cancer in Lebanon via real-time PCR. The results demonstrate the usefulness of real-time PCR in diagnosis and, therefore, in guiding proper clinical management and infection control. Preventing respiratory viral infections in immunocompromised patients, including those with cancer, is critical for protecting these patients, especially given the absence of effective vaccines and antiviral drugs for most of these viruses.

FUNDING

This project was supported by the Children's Infection Defense Center fund, ALSAC, and the American University of Beirut Faculty of Medicine seed fund.

CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

ORCID

Hassan Zaraket D http://orcid.org/0000-0003-3807-6409

REFERENCES

- Corti M, Palmero D, Eiguchi K. Respiratory infections in immunocompromised patients. *Curr Opin Pulm Med.* 2009;15(3):209-217. https://doi.org/10.1097/MCP.0b013e328329bd2c
- Hakim H, Dallas R, Zhou Y, et al. Acute respiratory infections in children and adolescents with acute lymphoblastic leukemia. *Cancer*. 2016;122(5):798-805. https://doi.org/10.1002/cncr.29833
- Benites ECA, Cabrini DP, Silva ACB, et al. Acute respiratory viral infections in pediatric cancer patients undergoing chemotherapy. *J Pediatr (Rio J)*. 2014;90(4):370-376. https://doi.org/10.1016/j.jped. 2014.01.006
- Fazekas T, Eickhoff P, Rauch M, et al. Prevalence and clinical course of viral upper respiratory tract infections in immunocompromised pediatric patients with malignancies or after hematopoietic stem cell transplantation. J Pediatr Hematol Oncol. 2012;34(6):442-449. https:// doi.org/10.1097/MPH.0b013e3182580bc8
- Perlin E, Bang KM, Shah A, et al. The impact of pulmonary infections on the survival of lung cancer patients. *Cancer*. 1990;66(3):593-596.

- Kohno S, Kola H, Oka M, et al. The pattern of respiratory infection in patients with lung cancer. *Tohoku J Exp Med*. 1994;173(4):405-411.
- 7. Lustberg MB. Management of neutropenia in cancer patients. *Clin Adv Hematol Oncol HO*. 2012;10(12):825-826.
- Rolston KVI. Infections in cancer patients with solid tumors: a review. Infect Dis Ther. 2017;6(1):69-83. https://doi.org/10.1007/s40121-017-0146-1
- Aronchick JM. Pulmonary infections in cancer and bone marrow transplant patients. Semin Roentgenol. 2000;35(2):140-151. https:// doi.org/10.1053/ro.2000.6152
- Godbole G, Gant V. Respiratory tract infections in the immunocompromised. Curr Opin Pulm Med. 2013;19(3):244-250. https://doi.org/ 10.1097/MCP.0b013e32835f82a9
- Mendoza Sánchez MC, Ruiz-Contreras J, Vivanco L, et al. Respiratory virus infections in children with cancer or HIV infection. J Pediatr Hematol Oncol. 2006;28(3):154-159. https://doi.org/10.1097/01.mph. 0000210061.96075.8e
- von Lilienfeld-Toal M, Berger A, Christopeit M, et al. Community acquired respiratory virus infections in cancer patients—guideline on diagnosis and management by the infectious diseases working party of the German society for haematology and medical oncology. *Eur J Cancer.* 2016;67(Suppl C):200-212. https://doi.org/10.1016/j.ejca. 2016.08.015
- Hussain Z, Ansari JA, Salman M, Khan EA, Asghar J. An evaluation of acute respiratory infection surveillance systems in Gilgit-Baltistan Pakistan. JPMA J Pak Med Assoc. 2016;66(6):682-687.
- Dizon DS, Krilov L, Cohen E, et al. Clinical cancer advances 2016: annual report on progress against cancer from the American society of clinical oncology. J Clin Oncol. 2016;34(9):987-1011. https://doi. org/10.1200/JCO.2015.65.8427
- Akinosoglou KS, Karkoulias K, Marangos M. Infectious complications in patients with lung cancer. Eur Rev Med Pharmacol Sci. 2013;17(1):8-18.
- Freifeld AG, Bow EJ, Sepkowitz KA, et al. Clinical practice guideline for the use of antimicrobial agents in neutropenic patients with cancer: 2010 update by the infectious diseases society of America. *Clin Infect Dis.* 2011;52(4):e56-e93. https://doi.org/10.1093/cid/cir073
- Christensen MS, Nielsen LP, Hasle H. Few but severe viral infections in children with cancer: a prospective RT-PCR and PCR-based 12month study. *Pediatr Blood Cancer*. 2005;45(7):945-951. https://doi. org/10.1002/pbc.20469
- Torres JP, De la Maza V, Kors L, et al. Respiratory viral infections and coinfections in children with cancer, fever and neutropenia: clinical outcome of infections caused by different respiratory viruses. *Pediatr Infect Dis J.* 2016;35(9):949-954. https://doi.org/ 10.1097/INF.00000000001209
- Söderman M, Rhedin S, Tolfvenstam T, et al. Frequent respiratory viral infections in children with febrile neutropenia - a prospective follow-up study. *PLoS One*. 2016;11(6):e0157398. https://doi.org/10. 1371/journal.pone.0157398
- Vliora C, Syridou G, Papaevangelou V. Viral respiratory infections in children receiving chemotherapy or undergoing stem cell transplantation. *Clin Microbiol Open Access*. 2014;3(1):137. https://doi.org/10. 4172/2327-5073.1000137
- Meerhoff TJ, Simaku A, Ulqinaku D, et al. Surveillance for severe acute respiratory infections (SARI) in hospitals in the WHO European region - an exploratory analysis of risk factors for a severe outcome in influenza-positive SARI cases. BMC Infect Dis. 2015;15:1. https:// doi.org/10.1186/s12879-014-0722-x
- Peret TC, Golub JA, Anderson LJ, Hall CB, Schnabel KC. Circulation patterns of genetically distinct group A and B strains of human respiratory syncytial virus in a community. J Gen Virol. 1998;79(Pt 9):2221-2229. https://doi.org/10.1099/0022-1317-79-9-2221
- 23. Sato M, Saito R, Sakai T, et al. Molecular epidemiology of respiratory syncytial virus infections among children with acute respiratory

MEDICAL VIROLOGY - WILEY

symptoms in a community over three seasons. J Clin Microbiol. 2005;43(1):36-40. https://doi.org/10.1128/JCM.43.1.36-40.2005

- Rotzén-Östlund M, Eriksson M, Tiveljung Lindell A, Allander T, Zweygberg Wirgart B, Grillner L. Children with multiple viral respiratory infections are older than those with single viruses. *Acta Paediatr.* 2014;103(1):100-104. https://doi.org/10.1111/apa.12440
- Aberle JH, Aberle SW, Pracher E, Hutter H-P, Kundi M, Popow-Kraupp T. Single versus dual respiratory virus infections in hospitalized infants: impact on clinical course of disease and interferon-gamma response. *Pediatr Infect Dis J.* 2005;24(7):605-610.
- Cilla G, Oñate E, Perez-Yarza EG, Montes M, Vicente D, Perez-Trallero E. Viruses in community-acquired pneumonia in children aged less than 3 years old: high rate of viral coinfection. J Med Virol. 2008;80(10):1843-1849. https://doi.org/10.1002/jmv.21271
- Asner SA, Science ME, Tran D, Smieja M, Merglen A, Mertz D. Clinical disease severity of respiratory viral co-infection versus single viral infection: a systematic review and meta-analysis. *PLoS One.* 2014;9(6):e99392. https://doi.org/10.1371/journal.pone.0099392
- Katsimpardi K, Papadakis V, Pangalis A, et al. Infections in a pediatric patient cohort with acute lymphoblastic leukemia during the entire course of treatment. *Support Care Cancer.* 2006;14(3):277-284. https://doi.org/10.1007/s00520-005-0884-6
- Koskenvuo M, Möttönen M, Rahiala J, et al. Respiratory viral infections in children with leukemia. *Pediatr Infect Dis J.* 2008;27(11):974-980. https://doi.org/10.1097/INF.0b013e31817b0799
- Mossong J, Hens N, Jit M, et al. Social contacts and mixing patterns relevant to the spread of infectious diseases. *PLoS Med.* 2008;5(3): e74. https://doi.org/10.1371/journal.pmed.0050074
- Stein ML, Van steenbergen JE, Buskens V, et al. Comparison of contact patterns relevant for transmission of respiratory pathogens in Thailand and the Netherlands using respondent-driven sampling. *PLoS One*. 2014;9(11):e113711. https://doi.org/10.1371/journal.pone.0113711
- Rhedin S, Lindstrand A, Rotzen-Ostlund M, et al. Clinical utility of PCR for common viruses in acute respiratory illness. *Pediatrics*. 2014;133(3):e538-e545. https://doi.org/10.1542/peds.2013-3042
- Vallières E, Renaud C. Clinical and economical impact of multiplex respiratory virus assays. *Diagn Microbiol Infect Dis.* 2013;76(3):255-261. https://doi.org/10.1016/j.diagmicrobio.2013.03.008
- 34. van Elden LJR, van Kraaij MGJ, Nijhuis M, et al. Polymerase chain reaction is more sensitive than viral culture and antigen testing for the detection of respiratory viruses in adults with hematological cancer and pneumonia. *Clin Infect Dis.* 2002;34(2):177-183. https:// doi.org/10.1086/338238
- Hall CB, Powell KR, MacDonald NE, et al. Respiratory syncytial viral infection in children with compromised immune function. N Engl J Med. 1986;315(2):77-81. https://doi.org/10.1056/NEJM198607103150201
- Whimbey E, Englund JA, Couch RB. Community respiratory virus infections in immunocompromised patients with cancer. *Am J Med.* 1997;102(3A):10-18. discussion 25-26
- Anaissie EJ, Mahfouz TH, Aslan T, et al. The natural history of respiratory syncytial virus infection in cancer and transplant patients: implications for management. *Blood*. 2004;103(5):1611-1617. https:// doi.org/10.1182/blood-2003-05-1425
- El Saleeby CM, Somes GW, DeVincenzo JP, Gaur AH. Risk factors for severe respiratory syncytial virus disease in children with cancer: the importance of lymphopenia and young age. *Pediatrics*. 2008; 121(2):235-243. https://doi.org/10.1542/peds.2007-1102

- Salzer W, Dinndorf P, Dreyer Z, Hilden J, Reaman GH. Analysis of infectious complications in infants with acute lymphoblastic leukemia treated on the Children's Cancer Group Protocol 1953: a report from the Children's Oncology Group. J Pediatr Hematol Oncol. 2009;31(6):398-405. https://doi.org/10.1097/MPH.0b013e3181a6dec0
- 40. Torres HA, Aguilera EA, Mattiuzzi GN, et al. Characteristics and outcome of respiratory syncytial virus infection in patients with leukemia. *Haematologica*. 2007;92(9):1216-1223.
- Wu H, Pfarr DS, Johnson S, et al. Development of motavizumab, an ultra-potent antibody for the prevention of respiratory syncytial virus infection in the upper and lower respiratory tract. J Mol Biol. 2007;368(3):652-665. https://doi.org/10.1016/j.jmb.2007.02.024
- 42. Weisman LE. Motavizumab, a second-generation humanized mAb for the prevention of respiratory syncytial virus infection in high-risk populations. *Curr Opin Mol Ther.* 2009;11(2):208-218.
- Neuzil KM. Progress toward a respiratory syncytial virus vaccine. Clin Vaccine Immunol. 2016;23(3):186-188. https://doi.org/10.1128/CVI. 00037-16
- 44. Turner T, Kopp B, Paul G, Hayes jr D, Thompson R, Landgrave L. Respiratory syncytial virus: current and emerging treatment options. *Clin Outcomes Res CEOR*. 2014;6:217-225. https://doi.org/10.2147/ CEOR.S60710
- Anderson LJ, Dormitzer PR, Nokes DJ, Rappuoli R, Roca A, Graham BS. Strategic priorities for respiratory syncytial virus (RSV) vaccine development. *Vaccine*. 2013;31(Suppl 2):B209-B215. https://doi.org/ 10.1016/j.vaccine.2012.11.106
- Olchanski N, Hansen RN, Pope E, et al. Palivizumab prophylaxis for respiratory syncytial virus: examining the evidence around value. *Open Forum Infect Dis.* 2018;5(3). https://doi.org/10.1093/ofid/ofy031
- Adams O, Bonzel L, Kovacevic A, Mayatepek E, Hoehn T, Vogel M. Palivizumab-resistant human respiratory syncytial virus infection in infancy. *Clin Infect Dis.* 2010;51(2):185-188. https://doi.org/10.1086/ 653534
- Zhu Q, Patel NK, McAuliffe JM, et al. Natural polymorphisms and resistance-associated mutations in the fusion protein of respiratory syncytial virus (RSV): effects on RSV susceptibility to palivizumab. J Infect Dis. 2012;205(4):635-638. https://doi.org/10.1093/infdis/ jir790
- Joseph C, Togawa Y, Shindo N. Bacterial and viral infections associated with influenza. *Influenza Other Respir Viruses*. 2013;7(Suppl 2):105-113. https://doi.org/10.1111/irv.12089
- Ljungman P, Gleaves CA, Meyers JD. Respiratory virus infection in immunocompromised patients. *Bone Marrow Transplant*. 1989;4(1):35-40.
- Martin ET, Fairchok MP, Stednick ZJ, Kuypers J, Englund JA. Epidemiology of multiple respiratory viruses in childcare attendees. *J Infect Dis.* 2013;207(6):982-989. https://doi.org/10.1093/infdis/ jis934

How to cite this article: Soudani N, Caniza MA, Assaf-Casals A, et al. Prevalence and characteristics of acute respiratory virus infections in pediatric cancer patients. *J Med Virol.* 2019;91:1191-1201. https://doi.org/10.1002/jmv.25432