

https://doi.org/10.3346/jkms.2017.32.5.757 • *J Korean Med Sci 2017; 32: 757-763*



Effect of Coenzyme Q10 on Radiation Nephropathy in Rats

Yongkan Ki,¹ Wontaek Kim,¹ Yong Ho Kim,¹ Donghyun Kim,¹ Jin Sook Bae,¹ Dahl Park,¹ Hosang Jeon,² Ju Hye Lee,² Jayoung Lee,² and Jiho Nam²

¹Department of Radiation Oncology, Biomedical Research Institute, Pusan National University Hospital and Pusan National University School of Medicine, Busan, Korea; ²Department of Radiation Oncology, Pusan National University Yangsan Hospital, Yangsan, Korea

Received: 10 November 2016 Accepted: 5 February 2017

Address for Correspondence:
Wontaek Kim, MD
Department of Radiation Oncology, Biomedical Research
Institute, Pusan National University Hospital and Pusan National
University School of Medicine, 179 Gudeok-ro, Seo-gu, Busan
49241, Republic of Korea
E-mail: rokwt@hanmail.net

Funding: This study was supported by Biomedical Research Institute & Busan Cancer Center Grant (2014–03), Pusan National University Hospital. The kidney is one of the most radiosensitive organs in the abdominal cavity and is the dose-limiting structure in cancer patients receiving abdominal or total body irradiation. In the present study, the effect of coenzyme Q10 (CoQ10) on radiation nephropathy was evaluated in rats. A total of 72 rats were equally randomized into 4 groups: Control, CoQ10, irradiation with 10 Gy (RT)+placebo, or RT+CoQ10. The 2 RT groups received single 10 Gy of abdominal irradiation. The 2 CoQ10 groups were supplemented daily with 1 mL of soybean oil containing 10 mg/kg of CoQ10. The RT+placebo and control groups received same dose of soybean oil. After 24 weeks, laboratory and histopathologic findings were compared. The 2 RT groups showed significant increases in blood urea nitrogen (BUN) and creatinine levels and significant pathologic changes such as glomerulosclerosis and tubulointerstitial fibrosis. CoQ10 supplementation resulted in significant reductions of BUN and creatinine levels compared with the RT+placebo group (P < 0.001 and P = 0.038, respectively). CoQ10 treatment significantly attenuated glomerular and tubular changes of irradiated kidney in semiquantitative analysis (P < 0.001 for both). Administration of CoQ10 can alleviate the radiation-induced nephropathy.

Keywords: Coenzyme Q10; Radiation Therapy; Total Body Irradiation; Radiation Nephropathy; Rats

INTRODUCTION

The kidney is known as one of the most radiosensitive organs in the abdominal cavity and is the dose-limiting structure for abdominal irradiation (1). Furthermore, over the past decades, total body irradiation (TBI) has emerged as a potential cause of renal impairment after receiving bone marrow transplantation (BMT) (2,3). Radiation nephropathy is characterized by a slow progressive reduction of renal function, and its clinical symptoms such as hypertension, azotemia, and anemia start after 6 to 12 months after irradiation (4). The radiation-induced kidney damage could progress to chronic renal failure, which was reported to develop in more than 20% of patients receiving TBI (5,6).

Coenzyme Q10 (CoQ10) is a fat-soluble, vitamin-like substance also called ubiquinone. CoQ10 is found everywhere in the body, and the highest concentrations are found in vital organs with high-energy turnover such as brain, heart, liver, muscle, and kidney (7). CoQ10 is an indispensable compound in the respiratory chain of the inner mitochondrial membrane and acts as essential and powerful antioxidants, scavenging free radicals, and inhibiting lipid peroxidation (8). The antioxidant effect of CoQ10 also enhance the availability of other antioxidants such as vitamin C, vitamin E, and beta-carotene (9).

The anti-oxidative and anti-inflammatory potential of CoQ10 was reported to be beneficial for the nephropathy induced by cisplatin or cyclosporine (10,11). However, there has been no previous report of treatment of CoQ10 for radiation nephropathy. The aim of this study was to identify the effects of CoQ10 on radiation nephropathy in rats.

MATERIALS AND METHODS

Animals

A total of 72 adult male Sprague-Dawley rats aged 3–4 months, weighing 300–330 g, were used. The animals were housed in wire cages at a constant temperature of $22^{\circ}\text{C} \pm 2^{\circ}\text{C}$. They were maintained on a 12-hour dark-light cycle with free access to standard laboratory chow and ultraviolet (UV)-sterilized water. After adaptation period of 1 week, all rats were randomly divided into the 4 groups of 18 animals each: Control, CoQ10, irradiation with 10 Gy (RT)+placebo, or RT+CoQ10.

Irradiation

All rats were anesthetized with an intraperitoneal injection of 100 mg/kg ketamine hydrochloride. The RT+placebo and RT+ CoQ10 groups received a single 10 Gy of whole abdominal irradiation using a 6-MV photon beam. The dose of 10 Gy was giv-

en based on the previous report that the significant renal function decline occurred 6 months after irradiation with a single dose of 10 Gy and higher in rats (12). Six rats at a time were restrained by the tail in a prone position on a 1 cm-thick acrylic plate and covered by another plate with the same thickness. Radiation was administered simultaneously to the rats at a 3 cm depth through anterior-posterior and posterior-anterior fields using a linear accelerator (Clinac iX, Varian Medical System, Inc., Palo Alto, CA, USA). The radiation dose rate was 1.06 Gy per minute.

Drugs and laboratory test

The control and RT+placebo group received daily intraperitoneal injection of 1 mL of soybean oil for 24 weeks after irradiation. The CoQ10 and RT+CoQ10 group received the same volume of soybean oil containing 10 mg/kg of CoQ10 using the same protocol. Body weights were checked daily. On the last experimental day, the animals were anesthetized by 100 mg/kg ketamine hydrochloride and blood samples of them were taken from trunk vessels for measurement of blood urea nitrogen (BUN) and creatinine levels.

Histopathologic evaluation

All rats were sacrificed by cervical dislocation after blood collection and underwent laparotomy immediately, and right kidneys of each animal were removed. The kidneys were cut in half perpendicular to the long axis at the level of the renal pelvis. Tissue samples were then fixed in 10% buffered formalin for 24 hours and embedded in paraffin wax. The sections were cut at 5 μm and stained with hematoxylin and eosin (H & E), periodic acid Schiff (PAS), and Masson's trichrome stains.

The severity of glomerulosclerosis was assessed using a semi-quantitative scoring method that was used widely in previous reports (13). One hundred glomeruli per animal on PAS stained sections (\times 400) were selected randomly and graded from 0 to 4: grade 0, normal; grade 1, sclerotic area up to 25% of glomerular tuft; grade 2, sclerotic area 25%–50%; grade 3, sclerotic area 50%–75%; and grade 4, sclerotic area 75%–100%. The glomerulosclerosis index (GSI) was calculated as follows:

([1 \times number of glomeruli in grade 1]+[2 \times number of glomeruli in grade 2]+[3 \times number of glomeruli in grade 3]+[4 \times number of glomeruli in grade 4])/total number of glomeruli examined (100)

The degree of tubulointerstitial fibrosis was also evaluated by a semiquantitative analysis from previous study (14). Twenty fields per section, randomly selected, were assessed on Masson's trichrome stained section (\times 100) and graded as follows: 0, no fibrosis; 1, fibrosis in up to 10% of field; grade 2, fibrosis in 10%–25% of field; grade 3, fibrosis in 25%–50% of field; grade 4, fibrosis in 50%–100% of field. The tubulointerstitial fibrosis index (TIF) was calculated as follows:

($[1 \times number of fields in grade 1]+[2 \times number of fields in grade 2]+[3 \times number of fields in grade 3]+[4 \times number of fields in grade 4])/total number of fields examined (20)$

All histopathological evaluation was performed in a blind method.

Statistical analysis

Mean differences between the 4 groups were assessed by oneway analysis of variance followed by Bonferroni's multiple comparison test. Statistical analyses were performed using SPSS, version 22.0 (IBM Corp., Armonk, NY, USA).

Ethics statement

The experimental protocol was reviewed and approved by the Institutional Animal Care and Use Committee at Pusan National University Hospital (No. PNUH 2014-065).

RESULTS

Laboratory findings and body weights

The levels of BUN and creatinine showed significant differences between the 4 groups (P < 0.001). Rats received 10 Gy of whole abdominal irradiation represented significant increases in BUN and creatinine levels, compared to the control group (P < 0.001 for both). CoQ10 administration resulted in a significant reduction of BUN levels in irradiated rats (P < 0.001). The creatinine level was also significantly decreased in the RT+CoQ10 group, compared with RT+placebo group (P = 0.038). The laboratory results were exhibited in Table 1.

The control and CoQ10 groups showed steady weight gain and the average increases of body weight were 194 g and 197 g, respectively. All the irradiated rats developed diarrhea and did not gain weight for about 1 week since irradiation. Mean weight gain of the RT+placebo and RT+CoQ10 group were 164 g and 176 g, respectively. The RT+placebo and RT+CoQ10 groups showed significant differences in mean weight change compared with the control group (P < 0.001 and P = 0.028, respectively). There were no significant differences between the control and CoQ10 groups with laboratory findings and weight changes.

Table 1. Effect of CoQ10 and radiation on renal function in rats

Variables	Control (n = 18)	CoQ10 (n = 18)	RT+placebo (n = 18)	RT+CoQ10 (n = 18)
BUN, mg/dL	22.3 ± 4.8*	21.8 ± 4.5*	$36.0 \pm 8.4^{\dagger}$	$27.5 \pm 3.2^{\star,\dagger}$
Creatinine, mg/dL	$0.46 \pm 0.10^*$	$0.44 \pm 0.09^*$	$0.65\pm0.08^\dagger$	$0.57 \pm 0.07^{*,\dagger}$

Data represent means \pm SD.

 $\mbox{CoQ10} = \mbox{coenzyme}$ Q10, RT = irradiation with 10 Gy, BUN = blood urea nitrogen, SD = standard deviation.

*P < 0.05, compared with the RT+placebo group; †P < 0.05, compared with the control group.

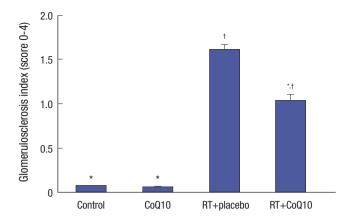


Fig. 1. GSI in rats. Data represent means \pm SD (n=18 per group). GSI = glomerulosclerosis index, CoQ10 = coenzyme Q10, RT = irradiation with 10 Gy, SD = standard deviation.

*P < 0.05, compared with the RT+placebo group; †P < 0.05, compared with the control group.

Histopathologic results

The kidney sections of the control and CoQ10 groups showed no obvious glomerular change and the mean GSI of them were measured as 0.08 ± 0.02 and 0.07 ± 0.02 , respectively. Glomerular injury such as mesangiolysis and glomerulosclerosis were observed in the 2 RT groups. The mean GSI were 1.61 ± 0.26 in the RT+placebo group and 1.03 ± 0.29 in the RT+CoQ10. CoQ10 treatment significantly decreased the mean GSI of irradiated kidney (P < 0.001). The comparisons of glomerular changes of rat kidneys were displayed in Figs. 1 and 2.

There were no tubulointerstitial fibrosis in the control and CoQ10 groups and their mean TIF were calculated as 0.06 ± 0.04 and 0.05 ± 0.04 , respectively. The sections of irradiated rats showed significant tubulointerstitial fibrosis. The mean TIF of RT+placebo and RT+CoQ10 groups were 1.43 ± 0.16 and 0.55 ± 0.17 , respectively. The RT+CoQ10 group showed a significantly lower fibrosis grade than RT+placebo group (P < 0.001). The comparisons of fibrotic changes of the kidneys from rats were represented in Figs. 3 and 4.

DISCUSSION

In the present study, we demonstrated that daily supplementation with 10 mg/kg of CoQ10 has beneficial effects in preventing radiation-induced nephropathy in experimental rats. The abdominal irradiation with single dose of 10 Gy increased the BUN and creatinine levels known as serum markers of kidney function, and CoQ10 administration reduced the degrees of elevations of them. CoQ10 also alleviated the pathologic changes in kidney such as glomeulosclerosis and tubulointerstitial fibrosis resulting from whole abdominal irradiation.

The tolerance dose of kidney had been thought to be 20 to 25 Gy in 3 to 5 weeks (15,16). In more recent studies, however, the reasonable threshold dose of radiation nephropathy was report-

ed to be lower than 15 Gy with conventional fractionation (5). The radiation nephritis also can occur even after irradiation with a single dose of 5 Gy (17). The improvements in radiation techniques such as intensity-modulated radiation therapy and volumetric arc radiation therapy have resulted in decreasing the radiation dose of kidney and the incidence of radiation nephropathy. Radiation-induced renal damage, nevertheless, cannot be prevented thoroughly because of the low tolerance of kidney. Radiation-induced renal injury is still considered as the principal late toxicity of abdominal irradiation or TBI associated with an inevitable radiation exposure of kidney (1-3).

Radiation nephropathy is known as typical chronic radiation damage characterized by progressive glomerulosclerosis and tubulointerstitial fibrosis, resulting in ultimate renal failure (4,18). Several studies showed that the glomerular and tubular fibrotic changes occurred most significantly 24 weeks after irradiation (14,19). The chronic progressive nature of radiation-induced renal damage has been reported without any recovery (20). Although there were variable experimental trials to manage the radiation nephropathy, such as angiotensin-converting-enzyme (ACE) inhibitor, amifostine and L-carnitine, only ACE inhibitor was considered to have a clear benefit of mitigating nephropathy without severe adverse effect (21-23). ACE inhibitor may slow but not prevent the progression of radiation nephropathy by controlling blood pressure or reducing proteinuria (23).

Radiation damage to cells results from the direct and indirect actions on deoxyribonucleic acid (DNA) molecules. About twothirds of the damage induced by photon is occurred through the indirect effect. The free radicals, produced by radiolysis of water, indirectly interact with DNA molecules and cause the critical cell damages. Free radicals have unpaired electrons, resulting in highly reactive and short-lived characteristics. These reactive oxygen species play a significant role in the oxidative damage of DNA by radiation therapy (24). The development and progression of radiation nephropathy, especially, was known to be associated chronic oxidative stress (25). The oxidative stress of irradiation can be a cause of the fibrogenesis (26). Radiationinduced lipid peroxidation, that is an important cause of cell damage and destruction, stimulates fibrogenic cytokines, and drives the radiation-induced fibrosis (27). Antioxidants such as CoQ10 have a positive role in preventing the fibrosis by their abilities to scavenge free radicals and inhibit lipid peroxidation (28). In addition, CoO10 has anti-inflammatory potentials reducing the pro-inflammatory cytokines in inflammatory process (29). These anti-oxidative and anti-inflammatory effects of CoQ10 have been used in treatments of various clinical conditions with oxidative stress and damage such as cardiovascular diseases, neurodegenerative diseases, diabetes, and migraines (30).

Emulsified and oil-based preparations was reported to be helpful to improve the absorption rate of CoQ10 (31). A wide

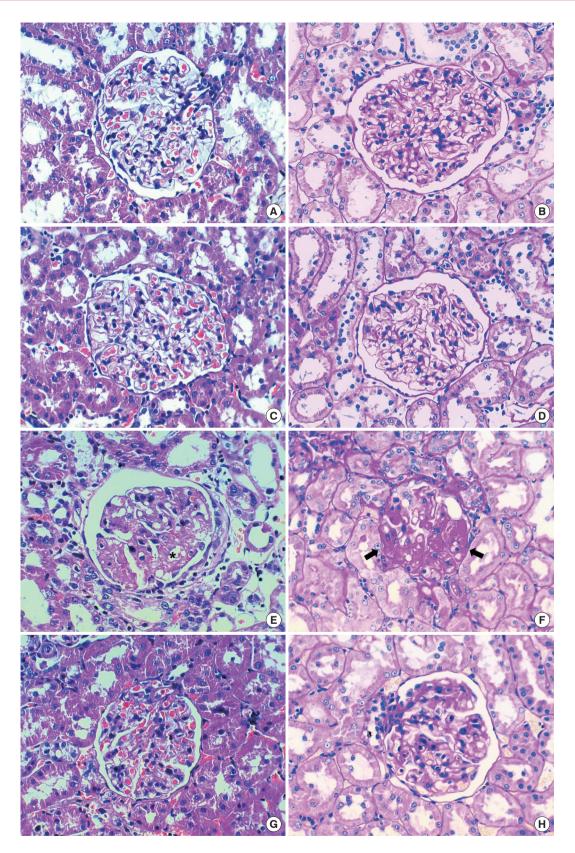


Fig. 2. Histopatholologic features of rat kidneys. (A, B) Control kidney. (C, D) CoQ10 group, exhibiting normal glomeruli. (E, F) RT+placebo group, showing mesangiolysis (asterisk) and glomerulosclerosis (arrow). (G, H) RT+CoQ10 group, representing marked reduction of glomerulosclerosis compared to RT+placebo group (A, C, E, G: H & E; B, D, F, H: PAS, \times 400).

 $CoQ10 = coenzyme\ Q10,\ RT = irradiation\ with\ 10\ Gy,\ H\ \&\ E = hematoxylin\ and\ eosin,\ PAS = periodic\ acid\ Schiff.$

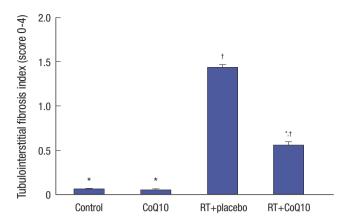


Fig. 3. Tubulointerstitial fibrosis index in rats. Data represent means \pm SD (n = 18 per group).

CoQ10 = coenzyme Q10, RT = irradiation with 10 Gy, SD = standard deviation. $^*P < 0.05$, compared with the RT+placebo group; $^\dagger P < 0.05$, compared with the control group.

range of doses (60-300 mg/day) of CoQ10 are generally used clinically, and a dose up to 2,400 mg/day was used safely for neurodegenerative disease (30). The administration of CoQ10 was well-tolerated in rats with daily dose of 1,200 mg/kg (32). CoQ10 is known to be safe and well-tolerable, and consumption of high dose CoQ10 over long periods has excellent safety reports (30,33). In the present study, 10 mg/kg of CoQ10 was administered to rats based on the previous experimental researches that showed the positive effect of the same dose of CoQ10 on the drug-induced oxidative stress and nephrotoxicity (10,11). There were no deaths or CoQ10-related toxicity findings during the experimental period. Supplemental CoQ10 increases plasma CoQ10 concentrations, but may not accumulate in tissues. Furthermore, because the half-life of CoQ10 in plasma is only about 33 hours, steady administration of CoQ10 is needed for chronic oxidative diseases (34).

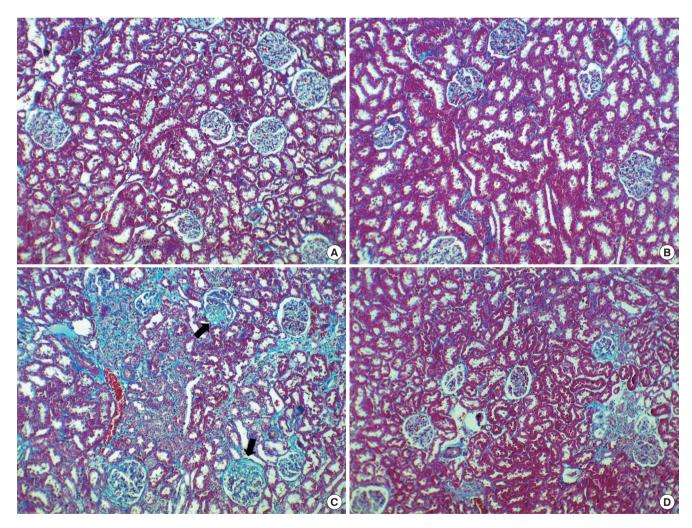


Fig. 4. Histopathologic features of rat kidneys. (A) Control kidney. (B) CoQ10 group, exhibiting no fibrosis. (C) RT+placebo group, showing tubulointerstitial fibrosis and glomeru-losclerosis (arrow). (D) RT+CoQ10 group, representing marked alleviation of fibrosis compared to RT+placebo group (Masson's trichrome, × 100). CoQ10 = coenzyme Q10. RT = irradiation with 10 Gy.

In conclusion, the administration of CoO10 improved the radiation-induced nephritis of rats in both laboratory and histopathologic findings. This result indicated that steady supplementation of CoO10 after abdominal or TBI could ameliorate radiation-induced nephrotoxicity. Further studies will be required to clarify the protective mechanism against radiation-induced nephrotoxicity and confirm the feasibility of CoO10 supplementation for preventing radiation nephropathy in patients.

DISCLOSURE

The authors have no potential conflicts of interest to disclose.

AUTHOR CONTRIBUTION

Conceptualization: Ki Y, Kim W. Data curation: Ki Y, Nam J. Formal analysis: Ki Y, Park D, Jeon H. Funding acquisition: Ki Y. Investigation: Ki Y, Kim YH, Kim D, Bae JS, Lee J, Lee JH. Writing original draft: Ki Y, Kim D. Writing - review & editing: Kim W.

ORICD

Yongkan Ki http://orcid.org/0000-0003-0757-8211 Wontaek Kim http://orcid.org/0000-0001-8626-7344 Yong Ho Kim http://orcid.org/0000-0002-4362-8426 Donghyun Kim http://orcid.org/0000-0002-8062-4619 Jin Sook Bae http://orcid.org/0000-0001-6304-9862 Dahl Park http://orcid.org/0000-0001-5143-3920 Hosang Jeon http://orcid.org/0000-0003-3960-3469 Ju Hye Lee http://orcid.org/0000-0002-6758-6822 Jayoung Lee http://orcid.org/0000-0001-6226-8799 Jiho Nam http://orcid.org/0000-0001-8495-0166

REFERENCES

- 1. Dawson LA, Kavanagh BD, Paulino AC, Das SK, Miften M, Li XA, Pan C, Ten Haken RK, Schultheiss TE. Radiation-associated kidney injury. Int J Radiat Oncol Biol Phys 2010; 76: S108-15.
- 2. Cohen EP. Radiation nephropathy after bone marrow transplantation. Kidney Int 2000; 58: 903-18.
- 3. Kang SH, Hwang HS, Park HS, Sun IO, Choi SR, Chung BH, Choi BS, Yang CW, Kim YS, Min CK, et al. Changes in renal function after different tandem hematopoietic stem-cell transplantation approaches in patients with multiple myeloma. J Korean Med Sci 2011; 26: 1310-5.
- 4. Cohen EP, Robbins ME. Radiation nephropathy. Semin Nephrol 2003; 23: 486-99
- 5. Cassady JR. Clinical radiation nephropathy. Int J Radiat Oncol Biol Phys 1995; 31: 1249-56.
- 6. Park J, Choi EK, Kim JH, Lee SW, Song SY, Yoon SM, Kim YS, Kim SS, Park JH, Park J, et al. Effects of total body irradiation-based conditioning on allogeneic stem cell transplantation for pediatric acute leukemia: a singleinstitution study. Radiat Oncol J 2014; 32: 198-207.

- 7. Mizuno K, Tanaka M, Nozaki S, Mizuma H, Ataka S, Tahara T, Sugino T, Shirai T, Kajimoto Y, Kuratsune H, et al. Antifatigue effects of coenzyme Q10 during physical fatigue. Nutrition 2008; 24: 293-9.
- 8. Kim J, Seok YM, Jung KJ, Park KM. Reactive oxygen species/oxidative stress contributes to progression of kidney fibrosis following transient ischemic injury in mice. Am J Physiol Renal Physiol 2009; 297: F461-70.
- 9. Shekelle P, Morton S, Hardy ML. Effect of supplemental antioxidants vitamin C, vitamin E, and coenzyme Q10 for the prevention and treatment of cardiovascular disease. Evid Rep Technol Assess (Summ) 2003: 1-3.
- 10. Fouad AA, Al-Sultan AI, Refaie SM, Yacoubi MT. Coenzyme Q10 treatment ameliorates acute cisplatin nephrotoxicity in mice. Toxicology 2010; 274:
- 11. Ishikawa A, Homma Y. Beneficial effect of ubiquinol, the reduced form of coenzyme Q10, on cyclosporine nephrotoxicity. Int Braz J Urol 2012; 38:
- 12. Jongejan HT, van der Kogel AJ, Provoost AP, Molenaar JC. Radiation nephropathy in young and adult rats. Int J Radiat Oncol Biol Phys 1987; 13: 225-32
- 13. Maric C, Sandberg K, Hinojosa-Laborde C. Glomerulosclerosis and tubulointerstitial fibrosis are attenuated with 17beta-estradiol in the aging Dahl salt sensitive rat. J Am Soc Nephrol 2004; 15: 1546-56.
- 14. Kanter M, Topcu-Tarladacalisir Y, Uzal C. Role of amifostine on acute and late radiation nephrotoxicity: a histopathological study. In Vivo 2011; 25:
- 15. Luxton RW. Radiation nephritis. QJ Med 1953; 22: 215-42.
- 16. Krochak RJ, Baker DG. Radiation nephritis. Clinical manifestations and pathophysiologic mechanisms. Urology 1986; 27: 389-93.
- 17. Moulder JE. Post-irradiation approaches to treatment of radiation injuries in the context of radiological terrorism and radiation accidents: a review. Int I Radiat Biol 2004: 80: 3-10.
- 18. Robbins ME, Bonsib SM. Radiation nephropathy: a review. Scanning Microsc 1995; 9: 535-60.
- 19. Kaldir M, Cosar-Alas R, Cermik TF, Yurut-Caloglu V, Saynak M, Altaner S, Caloglu M, Kocak Z, Tokatli F, Türe M, et al. Amifostine use in radiationinduced kidney damage. Preclinical evaluation with scintigraphic and histopathologic parameters. Strahlenther Onkol 2008; 184: 370-5.
- 20. Yildiz F, Atahan IL, Tuncel M, Konan A. The influence of dose per fraction on the pathogenesis of radiation nephropathy. Australas Radiol 1998; 42: 347-53.
- 21. Cosar R, Yurut-Caloglu V, Eskiocak S, Ozen A, Altaner S, Ibis K, Turan N, Denizli B, Uzal C, Saynak M, et al. Radiation-induced chronic oxidative renal damage can be reduced by amifostine. Med Oncol 2012; 29: 768-75.
- 22. Yurut-Caloglu V, Caloglu M, Deniz-Yalta T, Aktoz T, Nurlu D, Kilic-Durankus N, Arda E, Turkkan G, İnci O. Radiation-induced acute kidney toxicity: protective effect of L-carnitine versus amifostine. Int J of Radiat Res 2015; 13:317-24.
- 23. Moulder JE, Fish BL, Cohen EP. Treatment of radiation nephropathy with ACE inhibitors and AII type-1 and type-2 receptor antagonists. Curr Pharm Des 2007; 13: 1317-25.
- 24. Gillies NE. Effects of radiations on cells. Br Med J (Clin Res Ed) 1987; 295: 1390-1
- 25. Robbins ME, Zhao W, Davis CS, Toyokuni S, Bonsib SM. Radiation-induced kidney injury: a role for chronic oxidative stress? Micron 2002; 33: 133-41.
- 26. Zhao W, Spitz DR, Oberley LW, Robbins ME. Redox modulation of the pro-fibrogenic mediator plasminogen activator inhibitor-1 following ion-

- izing radiation. Cancer Res 2001; 61: 5537-43.
- 27. Yarnold J, Brotons MC. Pathogenetic mechanisms in radiation fibrosis. *Radiother Oncol* 2010; 97: 149-61.
- 28. Lefaix JL, Delanian S, Leplat JJ, Tricaud Y, Martin M, Nimrod A, Baillet F, Daburon F. Successful treatment of radiation-induced fibrosis using Cu/Zn-SOD and Mn-SOD: an experimental study. *Int J Radiat Oncol Biol Phys* 1996; 35: 305-12.
- 29. Schmelzer C, Lindner I, Rimbach G, Niklowitz P, Menke T, Döring F. Functions of coenzyme Q10 in inflammation and gene expression. *Biofactors* 2008: 32: 179-83
- 30. Littarru GP, Tiano L. Clinical aspects of coenzyme Q10: an update. Nutri-

- tion 2010; 26: 250-4.
- 31. Lyon W, Van den Brink O, Pepe S, Wowk M, Marasco S, Rosenfeldt FL. Similar therapeutic serum levels attained with emulsified and oil-based preparations of coenzyme Q10. *Asia Pac J Clin Nutr* 2001; 10: 212-5.
- 32. Williams KD, Maneke JD, AbdelHameed M, Hall RL, Palmer TE, Kitano M, Hidaka T. 52-Week oral gavage chronic toxicity study with ubiquinone in rats with a 4-week recovery. *J Agric Food Chem* 1999; 47: 3756-63.
- Hidaka T, Fujii K, Funahashi I, Fukutomi N, Hosoe K. Safety assessment of coenzyme Q10 (CoQ10). Biofactors 2008; 32: 199-208.
- 34. Overvad K, Diamant B, Holm L, Holmer G, Mortensen SA, Stender S. Coenzyme Q10 in health and disease. *Eur J Clin Nutr* 1999; 53: 764-70.