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Review article

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Cholesterol: The driving force behind the remodeling of tumor microenvironment in colorectal cancer

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ABSTRACT

Essential membrane components and metabolites with a wide range of biological roles are both produced by cholesterol metabolism. Cell-intrinsic and cell-extrinsic stimuli alter cholesterol metabolism in the tumor microenvironment (TME), which in turn encourages colorectal carcinogenesis. Metabolites produced from cholesterol play intricate roles in promoting the development of colorectal cancer (CRC) and stifling immunological responses. By altering the extracellular matrix of the main tumor, redesigning its immunological environment, and altering its mechanical stiffness, cholesterol can encourage the epithelial-mesenchymal transition of the primary tumor, opening up a pathway for tumor metastasis. Its functions in TME remodeling and therapeutic techniques designed to block cholesterol metabolism, and discuss how combining these strategies with already available anti-CRC medicines can have combined effects and open up new therapeutic avenues.

1. Introduction

In 2020 CRC accounted for 18 % of all cancer-related deaths and remained the second leading cause [1]. In 2040 there will likely be 28.4 million new instances of CRC worldwide, up 47 % from 2020 [2]. In the past few decades most research in the field of CRC has been focused on the tumor cells themselves. Bidirectional interactions between cancer cells and their surroundings, which result in remodeling of the tumor microenvironment (TME), are now understood to be essential for cancer growth and metastasis. Paracrine and juxtacrine signaling mediates and sustainably develops these linkages by secreting tumor-derived factors (TDFs) and tumor-derived exosomes [3]. Different strategies, including extracellular matrix (ECM) remodeling, metabolic reprogramming, immunosuppression, immune cell fatigue, and stromal cell activation can be used to establish TME remodeling, which facilitates the progression and

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distant colonization of tumor cells [3,4].

Cholesterol, a vital component of plasma and membrane lipids [5], supports tumor cells biomechanically and promotes their growth [6], invasion [7], metastasis [8], and drug resistance [9] by modulating TME remodeling. Although the idea of focusing on cholesterol metabolism to treat cancer has been the subject of extensive clinical research, the advantages are minimal, necessitating a thorough understanding of cholesterol metabolism in tumorigenesis and progression of CRC [10]. Emerging evidence suggests that cholesterol can modulate tumor biology by preventing tumor antigens from being identified by antigen presenting cells [11], reducing the manifestation of costimulatory and major histocompatibility complex class I components in dendritic cells [12], and impairing T cell antigen presentation. Fundamentally, cholesterol decreases the development of dendritic cells that invade tumors [13]. Cholesterol damages the T cell receptor's structure and lowers immunodetection during T cell priming and activation. Notably however, it also encourages T cell receptor clustering and relative signal transmission [14]. Lastly, cholesterol reduces granule-dependent cytotoxicity when T lymphocytes are killing cancer cells. Cholesterol and PGE2 can boost immunological checkpoint expression, increase the activity of immuneosuppressive cells, and encourage the release of immunosuppressive cytokines [15]. Erik et al. [16]reported that a reduced tumor response to endocrine treatments is a risk factor for colorectal cancers and hypercholesterolemia. More recently investigations have shown that cholesterol is one of the key regulators of TME remodeling in CRC.

In this review we summarize the most recent research on TME remodeling caused by cholesterol, with a focus on the role of several related derivatives, enzymes, and transcription factors in the initiation of CRC. We discuss potential cholesterol biomarkers for predicting metastases. Targeting cholesterol metabolism in the management and treatment of metastatic CRC is also discussed, as are potential ramifications and significant difficulties.

2. Various components involved in TME remodeling

The TME is an integral part of cancer. The concept of a complex tumor environment that supports tumor growth and metastatic dispersion has replaced the tumor cell-centered perspective of cancer development, as a result of the realization that the TME is fundamental to the evolution of cancer [17]. It is now widely acknowledged that stromal cells and the ECM, which together make up the main component of the TME, closely interact with cancer cells rather than acting independently [18]. Similarly, bone marrow-derived (stromal) cells are enlisted to create collections of purportedly normal tissue to form tumorigenic microenvironments (Table 1). There are two main ways that colorectalcancer cells can communicate with other cells and TME elements. One is through contact-dependent processes with other cells or the ECM, and the other is through contact-independent mechanisms via soluble

Table 1

morecular and central components promoting rails remote	Molecular a	nd	cellular	com	ponents	promoting	TME	remodeling.
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TME-Remodeling		Cancer	Underlying Mechanisms	References
Molecules		Туре		nererenceo
Tumor- derived	GM-CSF	PDA	Tumor cell-derived GM-CSF can orchestrate excessive Gr-1 ⁺ CD11b ⁺ imma ture myeloid cells	
	CCL21	melanoma	and CD8 ⁺ T cells	[21]
	IL-25	melanoma	CCL21 and IL-25 can induce MDSCs to accumulate in the TME, thereby forming an in	[22]
	IL-1β	gastric	flammatory microenvironment that is conducive to early tumor proliferation	[23,24]
		cancer	IL-1 β , CCL2, and PGE ₂ can attract adjacent peripheral fibers and neural pro genitor cells,	
	CCL2	gastric	which can guide cancer cell dissemination	[25]
	DOD	cancer	TGF-1 is more easily expressed when miR-142-3p and miR-506-3p are present	
	PGE ₂ miP 142	CPC		
	3n	CINC		
	miR-506-			
	3p			
Stroma- derived	Pdcd1	Melanoma	Increased protein geranylgeranylation induced by mevalonate metabolism is signaled by	[26]
	SREBP2	HCC	increased Pdcd1 expression in tumors, which is dependent on SREBP activity	[27]
	MMP8	CRC	In CD8 ⁺ T cells SREBP2 signaling was more potent, resulting in more active cholesterol up	[28]
	MMP9	CRC	take that provides the building blocks for quicker proliferation	[29]
	CSF-1	lung cancer	MMP8 and MMP9 can induce VEGF production, promoting metastasis	[30]
	TLR2	LLC	CSF-1 signaling is originally used to draw macrophages to tumor locations	
	MyD88		TLR2 and MyD88 can promote cholesterol activity and an immunosuppres sive macro phage	
	E4.885	m	phenotype	5013
Lipid- derived	FABP5	Tregs	Loss of cristae structure and poor lipid metabolism are all signs of mitochon drial alterations	[31]
	YBD1	breast	IIIUUCCU Dy FADF5 The endoplasmic reticulum stress sensor YBD1 can be activated by elevated cholesterol	[5]
	ADI 1	cancer	The endoptasine refeaturin seess sensor ADI I can be activated by crevated endoesteror	[0]
	2B4	carcinoma	2B4 transcription can suppress CD8 ⁺ T cell antitumor activity and transform the TME into an	[32]
			immunosuppressive phenotype	
Cytokine- derived	CD36	CRC	CD36-mediated absorption of fatty acids reduces generation of cytotoxic cytokines and	[44]
			diminished antitumor activity	
	TGF-β1	CRC	TGF - $\beta 1$ expression can promote differentiation of N1 neutrophils into N2 neutrophils via the	[41]
			miR-142-3p/miR-506-3p-TGF-β1 axis.	
	IFN	CRC	IFN-induced cholesterol 25-hydroxylase (CH25H) and type I interferon (IFN) receptor	[38]
			expression encourage CRC lung spread.	

chemicals such as cytokines, lipid mediators, and growth factors [18,19]. This procedure encourages tumor cell invasion, survival, and growth within the TME [20].

2.1. Primary tumor-derived components

Soluble compounds released by primary tumors [33] and disseminated CTCs [34] are examples of components generated from primary tumors. TME reshaping depends on soluble chemicals called TDFs, which primary tumors secrete [35]. Numerous studies indicate that TDFs encourage TME remodeling by promoting immune evasion and cholesterol-induced macrophage induction. Cholesterol 25-hydroxylase (CH25H), a gene that controls the production of 25HC, is reportedly repressed by activating transcription factor-3 (ATF3) [36]. ATF3-CH25H axis stimulation of trogocytosis in intratumoral cytotoxic T lymphocytes reduced anti-tumor immunity, promoting tumor development. Moreover, in pancreatic ductal adenocarcinoma (PDA), tumor cell-derived granulocyte-macrophage colony-stimulating factor can induce excessive Gr-1⁺CD11b⁺ immature myeloid cells and CD8⁺ T cells to assist in TME remodeling [33]. Similarly, melanoma cell secretion of CCL21 and IL-25 cause myeloid-derived suppressor cells (MDSCs) to accumulate in the TME, creating an inflammatory milieu that supports early tumor growth [37].

Primary tumor-derived extracellular vesicles (EVs) are crucial for TME reconstruction in CRC. For example, IFN-induced cholesterol 25-hydroxylase (CH25H) and type I interferon (IFN) receptor expression can be downregulated by CRC-secreted EVs in the TME, which encourages CRC lung spread [38]. The neurogenic switch, activated by tumor-derived EVs with a high abundance of IL-1 β , CCL2, PGE₂, and other chemotactic factors, attracts adjacent peripheral fibers and neural progenitor cells that can guide cancer cell dissemination [39]. Through HMGB1/toll-like receptor (TLR) 4/NF-B signaling, EVs isolated from cancer cell cultures can activate neutrophils in a way that promotes tumor growth [40]. In colorectal cancer circPACRGL serves as a sponge for miR-142-3p/miR-506-3p, facilitating transforming growth factor (TGF) β 1 expression, thus promoting colorectal cancer (CRC) cell proliferation, migration, and invasion—as well as differentiation of N1 neutrophils into N2 neutrophils via the miR-142-3p/miR-506-3p-TGF- β 1 axis [41]. TME remodeling is also closely linked to the level of cholesterol in cancer cells. Abundance of cholesterol in the TME is strongly associated with tumor recurrence and metastasis. Oxysterols in the TME promote reciprocal changes in the LXR and sterol regulatory element-binding protein 2 (SREBP2) pathways, depleting T cells of cholesterol, resulting in abnormal metabolic and signaling pathways that induce T cell exhaustion and dysfunction [5]. In CRC, by encouraging the production and secretion of cholesterol, the unfolded protein response component X-box binding protein 1 (XBP1) activates myeloid-derived suppressor cells and compromises immunity [42].

2.2. Formation of an immunosuppressive microenvironment

Tumor-associated myeloid cells that have been recruited promote the development of an immunosuppressive microenvironment [43]. and reorganized host stromal cells [42]. Myeloid-derived suppressor cells can alter the expression of genes linked to lipid/cholesterol metabolism, endosomal sorting pathways, and cell cytoskeleton remodeling by activating NF-B/STAT1 and inhibiting STAT6. This promotes the colonization and growth of tumor cells as well as tumor metastasis. Regulatory T (Treg) cells are essential for immunological tolerance in melanoma and also stimulate immunosuppression in the TME [44]. Blocking SREBP or PD-1 signaling causes dysregulated PI3K activation in intratumoral Treg cells. Through a process dependent on SREBP activity, Treg cells in CRC display elevated production of programmed cell death protein 1, which further promotes protein geranylgeranylation fueled by mevalonate metabolism [45]. Yan et al. [10]reported an uneven distribution of cholesterol in intratumoral immune cells. They showed that CD8⁺ T cells exhibited higher AKT-mTORC1-SREBP2 signaling, resulting in more active cholesterol uptake and more rapid proliferation of the cell's genetic material. Notably however, a low level of cholesterol may result in reduced survival and proliferation, as well as the initiation of autophagy [5,46]. Several investigations have revealed a unique, as yet uncharacterized poor-outcome immunomodulatory milieu in triplenegative breast tumors. This microenvironment includes stromal restriction of CD8⁺ T cells, stromal expression of PD-L1, and enrichment for markers of cholesterol synthesis [47]. Thus, Treg cells may be key cellular components involved in TME remodeling.

An immunosuppressive TME and tumor immune escape can be caused by tumor-associated neutrophils (TANs) [48]. Activation of the catenin pathway within CRC cells has been shown to predominately exclude immune cell activation and produce a TME without neutrophil inflammation in in vivo tests, leading to the formation of an immunosuppressive microenvironment [49]. Wang and Johnson et al. [50,51] reported that TANs can alter the ECM by generating neutrophil elastase (NE), matrix metallopeptidase (MMP) 8 and MMP9, and vascular endothelial growth factor (VEGF), which promotes metastasis.

The most prevalent immune cells in the TME are tumor-associated macrophages. A number of cytokines such as C-C motif chemokine ligand 2 (CCL2), tumor necrosis factor, VEGF, C-X-C motif chemokine ligand 12 (CXCL12), and TGF [52] contribute to early recruitment of macrophages to sites of tumor formation [53]. In addition to soluble factors, a recent study found that tumor-derived exosomes (TDEs) metabolically rewire macrophages by interacting with TLR2, activating MyD88, initiating NF-B signaling, increasing cholesterol activity, producing more lactate, and polarizing them into an immunosuppressive phenotype [54].

2.3. Dysregulation of host lipid metabolism

Lipid metabolites make up the physiological host mesenchymal environment, including cholesterol, sphingolipid, phospholipid, and fatty acids, which are required for TME homeostasis. Tumor hypoxia initiates the metabolic reprogramming of fatty acid oxidation and increases free fatty acid (FFA) intake, which promotes the growth of CRC cells, thus supporting tumor invasion, antiangiogenic

drug resistance, and TME remodeling [55]. Inhibiting fatty acid binding protein 5 (FABP5) can cause mtDNA release and subsequent cGAS-STING-dependent type I IFN signaling, which increases production of the regulatory cytokine IL-10 and encourages Treg cell suppressive activity [56].

In CRC, elevated cholesterol can activate the endoplasmic reticulum stress sensor XBP1 and regulate PD-1 and 2B4 transcription, thereby suppressing CD8⁺ T cell antitumor activity and transforming the TME into an immunosuppressive phenotype [5]. Faulty cholesterol efflux in epithelial progenitor cells controlled their transcriptional landscape, promoting growth and producing a pro-tolerogenic TME [57]. Similarly, in hematopoietic malignancies, increased cholesterol needs enable leukemic cells to proliferate at a rapid rate and create a TME suitable for tumor growth [58].

3. Accumulation and function of cholesterol in TME remodeling

3.1. Accumulation of cholesterol in the TME

Cholesterol accumulation is the initial stage of TME remodeling in CRC. Different elements derived from primary lesions and immunosuppressive cells can activate and upregulate the cholesterol production pathway. Because the messenger RNA levels of important genes encoding cholesterol synthesis proteins and transport routes are upregulated in colon carcinoma, while those of the cholesterol efflux pathway are downregulated, cholesterol builds up in cancer cells [59]. Ma et al. [5] reported that reduced generation of cytotoxic cytokines and diminished antitumor activity were the results of CD36-mediated absorption of fatty acids by tumor-infiltrating CD8⁺ T cells in the TME. Lipid peroxidation and ferroptosis were also brought on by this process, which finally produced a microenvironment that was tumor-tolerant [5,44,60]. Moreover, high levels of cholesterol can promote the accumulation of MDSCs in host organs through the IRE1 α -XBP1 pathway [42,61]. Goossens et al. [62]found that membrane-cholesterol efflux and the depletion of lipid rafts from macrophages are encouraged by ovarian cancer cells. By enhancing cholesterol efflux and IL-4-mediated reprogramming, which involves suppressing IFN-induced gene expression, the tumor-growth-promoting effects of tumor-associated macrophages can be countered. The actions of TAMs that promote tumor growth are reversed and tumor progression is decreased by genetic deletion of ABC transporters, which facilitate cholesterol efflux. Lipid rafts, which are membrane microdomains rich in cholesterol, may prevent the destruction of the matrix caused by invadopodia in order to direct the distant colonization of cancer cells. This slows the propagation of tumors [62,63].

In addition to immune cells, transcription factors can also activate the cholesterol synthesis pathway in primary lesions [64]. Gu et al. [65] reported that in glioblastoma (GBM) tumor cores, as opposed to invasive tumor margins, cholesterol biosynthesis enzymes were expressed at higher quantities. SREBP2 enhanced CRC stem cell proliferation, self-renewal, and tumor growth, especially in starving circumstances.It's interesting to note that a different research team studying GBM came to the conclusion that the IDH mutation affects the proliferation and invasion of cancer cells by acting as a unique post-transcriptional regulator of cholesterol uptake mediated by the miR~19a/LDLR axis [66,67]. Thus, transcription factors are crucial to TME remodeling.

TDFs and tumor-derived EVs contribute to the cholesterol synthesis pathway. For example, The ATF3 transcription factor CH25H can be activated and cholesterol 25hydroxylase can be increased by stimuli emanating from the TME. CH25H upregulation is linked to CRC progression and tumor growth in antigen-presenting cells isolated from human colorectum tumors [12,36,68]. EVs with an abundance of cholesterol can also assist in remodeling the TME. Keeping CD8 alpha-negative dendritic cells from being activated by toll-like receptors and preventing a protective Th1 type response promotes the growth of an immunosuppressive environment [69].

3.2. ECM remodeling

The ECM is a complex network of macromolecules secreted from cells, including glycoproteins, enzymes, and collagen [70]. It gives cancer cells and tissues structural and biochemical support [71]. The primary building block of the tumor matrix and an important player in ECM remodeling is cholesterol. Lipid biosynthesis and adipogenesis are regulated by the SREBP family of transcription factors, which also controls the expression of many enzymes needed for cholesterol production [65,72]. A recent study has demonstrated that in CRC, the energy sensor AMPK upregulates SREBP1 activation, promoting the production of cholesterol, evidently triggering ECM rigidity and affecting physiological and pathophysiological procedures such as tissue fibrosis and mesenchymal stem cell differentiation [73].

Bone morphogenetic protein and activin membrane bound inhibitor, a TGF pseudoreceptor, is downregulated in response to TGFinduced activation through stimulation of TLR4 [74,75]. In primary malignancies such as those of the colon [76], lung [77], and breast [78], ECM remodeling is conducive to epithelial-mesenchymal transition (EMT) or migration of tumor cells. A fluid-phase absorption mechanism mediated by the cholesterol-dependent pathway enables internalization and degradation of extracellular protein by cells expressing oncogenic RAS, which accelerates the cell cycle and/or suppresses cell death regulated by ECM, leading to uncontrolled cancer expansion [79,80]. ECM protein 1 (ECM1) expression was positively correlated with hypercholesterolemia of triple-negative breast cancer cells, enhancing ECM stiffness in the TME [81]. It is interesting that various studies have emphasized how MMPs affect control of the ECM and cholesterol homeostasis [82]. For example, by degrading cholesterol, MMP2 and MMP9 can facilitate the invasion of CRC cells into the basement membrane, leading to tumor spread and diffusion [83].

3.3. Immunosuppression and angiogenesis

When it comes to the establishment of an immunosuppressive TME in CRC, cholesterol is essential. Depending on the circumstance,

cholesterol can help tumor cells avoid immune surveillance by interacting with different kinds of immune cells [84]. For example, in a mouse CRC model, cholesterol caused CD8⁺ T cell "exhaustion" [5,10]. Furthermore, by inhibiting TCR phosphorylation and restricting TCR allosteric transitions in human CD4⁺ Jurkat T cells, cholesterol and its metabolites can adversely affect TCR signaling [85], thus suppressing T cell infiltration into the TME. MHC class I chain-related protein A expression on human CRC cell lines can be reduced by cholesterol in NK cells, shielding them from being targeted by NK cells, thus providing an immunosuppressive microenvironment for melanoma cells [86]. Furthermore, mTORC1-induced cholesterol accumulation through SREBP activation encourages myeloid cell differentiation into MDSC [87].

Cholesterol encourages angiogenesis [88], which facilitates colorectal tumor growth. Local cholesterol is important for promoting angiogenesis in the TME. Researchers reported that apoA-I binding protein affects VEGFR2 dimerization and accelerates cholesterol efflux from endothelial cells to high-density lipoprotein (HDL), thereby promoting VEGF-induced angiogenesis [89]. Older macrophages have a polarized phenotype that is abnormally alternately activated and promotes pathologic vascular development when intracellular cholesterol is increased, indicating that it could be an effective therapeutic target [90]. Activated cholesterol synthesis induced by tumor-derived C-X-C motif chemokines is also involved in angiogenesis and TME reconstruction [91].

3.4. Metabolism reprogramming

Cholesterol homeostatic imbalance is responsible for TME remodeling. Enzymes [92], derivatives [93], and transcription factors [72,94] involved in cholesterol metabolic pathways all regulate metabolic TME [95] reprogramming in CRC. Squalene monooxygenase, or squalene epoxidase (SQLE), is an enzyme that controls the rate at which cholesterol is produced [96,97]. Several studies have reported that the N-terminal 100 amino acids of SQLE can be ubiquitinated by MARCHF6, causing proteasomal degradation. This enables tumor cells to proliferate through the production of migratory cancer stem cells, senescence bypass, anoikis resistance, and EMT [98,99]. Additionally, cholesterol accumulation that reduces SQLE promotes p53 breakdown and catenin activation, which ultimately speeds up TME remodeling and tumor progression in colorectal cancer (CRC) by inhibiting GSK3, which in turn leads p53 to dissociate from it and upregulate MDM2 [99]. Similar findings were made by another team, which stated that SQLE encourages the conversion of squalene to 2,30xidosqualene in the cholesterol synthesis pathway and that its absence results in an accumulation of the upstream metabolite squalene. This process modifies the cellular lipid profile and prevents ferroptotic cell death in cancer cells, giving them an advantage during oxidative stress and in tumor xenografts [100].

As well as enzymes, related derivatives are involved in TME remodeling. In breast cancer, ER and the liver X receptor (LXR) ligand 27-hydroxycholesterol, a major metabolite of cholesterol, promotes ER-dependent proliferation and LXR-dependent metastasis [8]. Recent studies suggest that 27-hydroxycholesterol may play a pro-metastatic role by inducing immune reprogramming [101]. Baek and his colleagues showed that this oxysterol promotes an immunologically suppressive milieu by increasing the number of polymorphonuclear-neutrophils and $\gamma\delta$ T cells at distal metastatic sites. Myeloid immune cell activity is required for the potent



Fig. 1. Function of cholesterol in tumor microenvironment remodeling. Cholesterol derived from tumor cells or ingested externally can help tumor cells escape immune surveillance. In primary lesions cholesterol interacts with other cells such as T cells and NK cells, remodeling the tumor microenvironment (TME), increasing the survival of cancer cells. Cholesterol reshapes the TME through extracellular matrix remodeling, immunosuppression, angiogenesis, and metabolism reprogramming. ECM, extracellular matrix; EMT, epithelial-mesenchymal transition.

anti-metastatic effects of 27-hydroxycholesterol [7].

In recent years growing numbers of researchers have focused on transcription factors and related receptors involved in the cholesterol metabolic pathway. For example, via the downregulation of LXR and HIVEP2, YTHDF2 promotes carcinogenesis in GBM cells and prevents LXR-dependent cholesterol homeostasis in GBM cells by activating the EGFR/SRC/ERK pathway [102]. Another team investigating CRC reported that with promotion of cholesterol manufacture in the tumor core and absorption in the margin, the transcription factor SREBP2 exhibited context-specific regulation of cholesterol biology based on its availability in the microenvironment [65]. In triple-negative breast cancer and CRC, nuclear receptor ROR γ increases the tumor cholesterol synthesis rate and total tumor cholesterol content, while disrupting host cholesterol homeostasis. Cai et al. [103] demonstrated that ROR γ interacts with SREBP2 and promotes chromatin acetylation at the locations of the genes responsible for cholesterol production, which advances tumors. p53, a well-known tumor suppressor gene, exerts anti-tumor effects by inhibiting cholesterol synthesis. Moon et al. [104] reported that p53 inhibits the mevalonate pathway and prevents SREBP-2 activation by transcriptionally activating the ABCA1 cholesterol transporter gene.

Recent research indicates that cholesterol influences EMT/mesenchymal transition, which in turn increases TME remodeling (MMT). In CRC cancer stem cells (CSCs), ZMYND8, a master transcriptional regulator of 27-HC metabolism, promotes EMT, oncogenic transformation, and tumor initiation by activating liver X receptor while decreasing 27-HC catabolism. This results in an accumulation of 27-HC in CSCs, and increases cholesterol biosynthesis and oxidation while decreasing 27-HC efflux [105]. Moreover, statins and NAD (P) dependent steroid dehydrogenase-like gene loss activate SREBP1, which promotes TGF- β 1 expression and facilitates epithelial-mesenchymal transition [106]. Cholesterol has significant functions in TME remodeling, promotion of tumor incidence, and growth of the main lesion. It is necessary to conduct more research on the functions of cholesterol in ECM, metabolic reprogramming, immunosuppression, and angiogenesis in TME reshaping (Fig. 1).

4. Cholesterol-related CRC biomarkers and therapeutic strategies

4.1. Finding cholesterol-related biomarkers to predict metastasis

Studies investigating the influences of particular cholesterol subtypes on tumor progression have become more prevalent as metabolomics techniques have advanced [107]. Modern studies are no longer limited to the conventional cholesterol classifications "HDL" and "LDL." Various malignancies, organs, and even different regions of the same organ have different cholesterol characteristics. Cholesterol and associated indicators can be used as therapeutic targets or biomarkers.

A range of cholesterol-related indicators in the initial tumor are reportedly strongly associated with tumor growth and metastasis by metabolomics and other multi-omics studies. According to Mihajlovic et al. [108], colorectal cancer incidence is affected by the activity of PON1 and lactonase (LACT), two of cholesterol's primary antioxidant components. This in turn causes a drop in HDL cholesterol concentration. In CRC, increased cholesterol inhibits expression of the scavenger receptor class B type 1 (SR-B1) in mitochondria, which prevents the mitochondria from inducing apoptosis [109]. In her2-positive heterogeneous gastric cancer, levels of CAV1 tumoral protein, a significant protein of cholesterol-rich membrane domains, were reportedly negatively correlated with TDM1 tumor uptake, causing limited trastuzumab benefit and poor patient survival [110]. Similarly recombinant HDL, a drug delivery nanoparticle that is currently being used to deliver small-molecule drugs, siRNAs, therapeutic proteins, and vaccine antigens, can be created from HDL via a processing step. Recent years have seen a significant increase in studies investigating the use of recombinant HDL and SR-B1 for the treatment of CRC [111].

In addition to HDL, recent studies have also revealed that some derivatives and receptors that maintain cholesterol homeostasis may help to improve CRC prevention and treatment [7]. For example, Noguchi et al. [112] discovered that inducing an adaptive response by activating the liver X receptor signaling system, 24S-OHC reportedly protects neuronal cells from malignant growth at sublethal dosages. Furthermore, the primary protein of HDL, apolipoprotein A-I, is a versatile protein that regulates inflammation and the immune system as well as cholesterol traffic. It might be a valuable biomarker that aids earlier CRC diagnosis, follow-up, and prognostic stratification in cancer patients as well as improved estimation of cancer risk [113]. Zeng et al. [114]found that the sigma-2 receptor TMEM97 has already been created and validated as a PET imaging biomarker of tumor proliferative status, and as a predictor of the effectiveness of cancer therapy. Sterol depletion and SREBP expression levels, among other cholesterol-regulating cues, control the expression of TMEM97. Understanding the several types of cholesterol-related indicators found in CRC and their properties and effects on TME remodeling will be crucial for developing new therapeutic approaches for the prevention of CRC.

TDFs and EVs with high cholesterol abundances have drawn a lot of interest. Potential biomarkers for primary and metastatic CRC include TDFs and EVs with elevated cholesterol concentrations in the TME or metastatic locations. For example, in CRC TEV reduced expression of the IFN-inducible cholesterol 25-hydroxylase and type I interferon (IFN) receptor (CH25H). A worse prognosis was associated with low CH25H levels in leukocytes from CRC patients [115]. In CRC, early-onset mammalian immune surveillance mechanisms can become "blind" to a growing cancer and lose their capacity to recognize and launch measures to eradicate a tumor due to the formation of cholesterol-enriched pmEV [34].

4.2. Targeting cholesterol for CRC therapy

Cholesterol is a prospective therapeutic target for CRC therapies due to its protumor activities. Cholesterol tumors are challenging to target due to the absence of distinct tumor markers. The following list describes the alternatives for current anti-cholesterol therapy in main tumor locations and metastases.

The mechanical tension in the membranes of cancer cells is changed. A novel approach to controlling cell membrane tension offers a possible course of action for cancer treatment. One study describes how cholesterol oxidase (COD) increases cell membrane tension in vitro by depleting cholesterol and how Hf-TBP/COD, a COD-functionalized nanoscale metal-organic framework, is designed to deplete cholesterol and regulate tumor mechanogenesis in vivo. It has been discovered that COD reduces cholesterol and modifies the mechanical characteristics of lipid bilayers, which inhibits cell motility, proliferation, and oxidative stress tolerance. Hf-TBP/COD increases mechanical tension of plasma membranes and osmotic fragility of cancer cells, which induces influx of zinc ions, inhibits cell migration, increases rupturing propensity for effective caspase-1 mediated pyroptosis, and decreases tolerance to oxidative stress. In a subcutaneous colon cancer model, Hf-TBP/COD increases anti-tumor immune response and tumor growth inhibition from 79.8 % to 95 % when compared to Hf-TBP alone [115,116].

Cholesterol primarily aids CRC cell survival and proliferation, and the development of medication resistance in primary tumors. In CRC patients, overexpression of transcriptional coactivator with PDZ-binding motif (TAZ) is closely associated with poor survival. However, it was demonstrated that cholesterol synthesis restricted TAZ expression in CRC by inhibiting 3-hydroxy-3-methylglutaryl-coenzyme A reductase, farnesyl pyrophosphate synthase, farnesyl-diphosphate farnesyltransferase 1, or sterol regulatory element-binding protein 2. The mevalonate-cholesterol-TAZ-TEAD2-Anln/Kif23 pathway has been identified as a novel tumor-specific target for CRC treatment in the past investigation. A combination of statins and immune checkpoint inhibitors greatly reduced tumor development, despite statins' moderate therapeutic effects on this pathway's blockage [116]. Additionally, discoveries made by Yan et al. [10]have intriguing therapeutic ramifications. It may be possible to develop methods that successfully stop cancer cells from synthesizing cholesterol while maintaining the availability of cholesterol in lymphocytes. Conversely newly created LXR-inverse agonists, which have previously been demonstrated to improve CD8⁺ T cell activity in preclinical models, may offer therapeutic promise in the future [117]. Given these encouraging findings, a more thorough analysis of regulating cholesterol metabolism to enhance chimeric antigen receptor T-Cell immunotherapy is unquestionably required.

In metastases, high cholesterol levels are more likely to promote immunosuppressive cells, stimulate angiogenesis, and create favorable ECM conditions and immunosuppressive environments for the colonization and proliferation of CRC cells. There may be a "tipping point" whereby metastases starts with cholesterol buildup. In a subcutaneous inoculation mouse model, Uchida et al. [118] used polyplex nanomicelles with a cholesterol moiety to treat metastatic CRC via the delivery of mRNA encoding an anti-angiogenic protein (sFlt-1). PEG-PAsp(TEP)-Chol nanomicelles, as opposed to those without Chol, effectively synthesized protein from that mRNA in tumor tissue, leading to a detectable decrease in tumor growth [119]. Further research is needed to identify more precise molecular targets that affect cholesterol signals and effectors in CRC. Moreover, traditional regimens' combinatorial methods with anti-cholesterol treatments need to be investigated.

5. Conclusions and perspective

The rate at which CRC develops is mostly determined by how invasive the tumor cells are, as well as by a number of other factors. The intricate molecular mechanisms enabling TME reconfiguration have recently been the subject of increased study. Developments have improved CRC diagnosis techniques and our understanding of the mechanisms underlying tumor spread, laying the groundwork for viable treatment plans. In order for tumor cells to attach, live, and multiply at primary and metastatic sites, as well as to drive tumor spread, the TME must interact with host stromal cells and cholesterol. With an emphasis on TDFs and EVs, more study has been done in recent years on cholesterol in primary lesions and at metastatic sites. Notably however, a number of questions about the origin, purpose, dynamics, and importance of the TME remain unanswered. For the prevention and treatment of metastatic CRC, a deeper comprehension of the mechanisms underpinning cholesterol driving TME remodeling, and the discovery of its effects on CRC metastasis will be extremely beneficial.

CRediT authorship contribution statement

Ke Wang: Writing – original draft. Yuanyuan Zhang: Writing – review & editing, Writing – original draft. Chengshuai Si: Supervision. Yuepeng Cao: Supervision, Funding acquisition. Peng Shao: Validation. Pei Zhang: Validation, Supervision. Nannan Wang: Visualization, Validation. Guoqing Su: Writing – original draft. Jinghang Qian: Writing – original draft. Liu Yang: Validation, Supervision.

Ethics approval and consent to participate

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Abbreviations

CRC	colorectal cancer
TME	tumor microenvironment
TDFs	tumor-derived factors
ECM	extracellular matrix
GM-CSF	granulocyte-macrophage colony-stimulating factor
CCL	chemotactic ligand
PGE2	prostaglandin E2
Pdcd1	programmed cell death protein 1
SREBP2	Sterol Regulatory Element-Binding Protein 2
FABP5	fatty acid-binding protein 5
XBP1	X-box binding protein-1
HCC	hepatocellular carcinoma
PDA	Pancreatic Ductal Adenocarcinoma
EMT	epithelial-mesenchymal transition
CH25H	cholesterol 25-hydroxylase

References

- [1] R.L. Siegel, K.D. Miller, N.S. Wagle, A. Jemal, Cancer statistics, 2023, CA Cancer J Clin 73 (2023) 17–48, https://doi.org/10.3322/caac.21763.
- [2] H. Sung, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal, F. Bray, Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, CA Cancer J Clin 71 (2021) 209–249, https://doi.org/10.3322/caac.21660.
- [3] O. Meurette, P. Mehlen, Notch signaling in the tumor microenvironment, Cancer Cell 34 (2018) 536–548, https://doi.org/10.1016/j.ccell.2018.07.009.
 [4] J.M. Pitt, A. Marabelle, A. Eggermont, J.C. Soria, G. Kroemer, L. Zitvogel, Targeting the tumor microenvironment: removing obstruction to anticancer immune
- responses and immunotherapy, Ann. Oncol. 27 (2016) 1482–1492, https://doi.org/10.1093/annonc/mdw168. [5] X. Ma, E. Bi, Y. Lu, P. Su, C. Huang, L. Liu, Q. Wang, M. Yang, M.F. Kalady, J. Qian, et al., Cholesterol induces CD8(+) T cell exhaustion in the tumor
- microenvironment, Cell Metab 30 (2019) 143-156.e145, https://doi.org/10.1016/j.cmet.2019.04.002.
- [6] S.F. Nielsen, B.G. Nordestgaard, S.E. Bojesen, Statin use and reduced cancer-related mortality, N. Engl. J. Med. 367 (2012) 1792–1802, https://doi.org/ 10.1056/NEJMoa1201735.
- [7] A.E. Baek, Y.A. Yu, S. He, S.E. Wardell, C.Y. Chang, S. Kwon, R.V. Pillai, H.B. McDowell, J.W. Thompson, L.G. Dubois, et al., The cholesterol metabolite 27 hydroxycholesterol facilitates breast cancer metastasis through its actions on immune cells, Nat. Commun. 8 (2017) 864, https://doi.org/10.1038/s41467-017-00910-z.
- [8] E.R. Nelson, S.E. Wardell, J.S. Jasper, S. Park, S. Suchindran, M.K. Howe, N.J. Carver, R.V. Pillai, P.M. Sullivan, V. Sondhi, et al., 27-Hydroxycholesterol links hypercholesterolemia and breast cancer pathophysiology, Science 342 (2013) 1094–1098, https://doi.org/10.1126/science.1241908.
- [9] L. Xiong, H.S. Liu, C. Zhou, X. Yang, L. Huang, H.Q. Jie, Z.W. Zeng, X.B. Zheng, W.X. Li, Z.Z. Liu, et al., A novel protein encoded by circINSIG1 reprograms cholesterol metabolism by promoting the ubiquitin-dependent degradation of INSIG1 in colorectal cancer, Mol. Cancer 22 (2023) 72, https://doi.org/ 10.1186/s12943-023-01773-3.
- [10] C. Yan, L. Zheng, S. Jiang, H. Yang, J. Guo, L.Y. Jiang, T. Li, H. Zhang, Y. Bai, Y. Lou, et al., Exhaustion-associated cholesterol deficiency dampens the cytotoxic arm of antitumor immunity, Cancer Cell 41 (2023) 1276–1293.e1211, https://doi.org/10.1016/j.ccell.2023.04.016.
- [11] O.F. Kuzu, M.A. Noory, G.P. Robertson, The role of cholesterol in cancer, Cancer Res. 76 (2016) 2063–2070, https://doi.org/10.1158/0008-5472.Can-15-2613.

- [12] V. Janelle, C. Rulleau, S. Del Testa, C. Carli, J.S. Delisle, T-cell immunotherapies targeting histocompatibility and tumor antigens in hematological malignancies, Front. Immunol. 11 (2020) 276, https://doi.org/10.3389/fimmu.2020.00276.
- [13] K.C. Corn, M.A. Windham, M. Rafat, Lipids in the tumor microenvironment: from cancer progression to treatment, Prog. Lipid Res. 80 (2020) 101055, https:// doi.org/10.1016/i.plipres.2020.101055.
- [14] D. Howie, A. Ten Bokum, A.S. Necula, S.P. Cobbold, H. Waldmann, The role of lipid metabolism in T lymphocyte differentiation and survival, Front. Immunol. 8 (2017) 1949, https://doi.org/10.3389/fimmu.2017.01949.
- [15] Y. Wang, Y. Wang, Y. Ren, Q. Zhang, P. Yi, C. Cheng, Metabolic modulation of immune checkpoints and novel therapeutic strategies in cancer, Semin. Cancer Biol. 86 (2022) 542–565, https://doi.org/10.1016/j.semcancer.2022.02.010.
- [16] B. Huang, B.L. Song, C. Xu, Cholesterol metabolism in cancer: mechanisms and therapeutic opportunities, Nat. Metab. 2 (2020) 132–141, https://doi.org/ 10.1038/s42255-020-0174-0.
- [17] J.M. Pitt, A. Marabelle, A. Eggermont, J.C. Soria, G. Kroemer, L. Zitvogel, Targeting the tumor microenvironment: removing obstruction to anticancer immune responses and immunotherapy, Ann. Oncol. 27 (2016) 1482–1492, https://doi.org/10.1093/annonc/mdw168.
- [18] D. Hanahan, L.M. Coussens, Accessories to the crime: functions of cells recruited to the tumor microenvironment, Cancer Cell 21 (2012) 309–322, https://doi.org/10.1016/j.ccr.2012.02.022.
- [19] S.A. Bergfeld, Y.A. DeClerck, Bone marrow-derived mesenchymal stem cells and the tumor microenvironment, Cancer Metastasis Rev. 29 (2010) 249–261, https://doi.org/10.1007/s10555-010-9222-7.
- [20] T.F. Gajewski, H. Schreiber, Y.X. Fu, Innate and adaptive immune cells in the tumor microenvironment, Nat. Immunol. 14 (2013) 1014–1022, https://doi.org/ 10.1038/ni.2703.
- [21] G.L. Beatty, E.G. Chiorean, M.P. Fishman, B. Saboury, U.R. Teitelbaum, W. Sun, R.D. Huhn, W. Song, D. Li, L.L. Sharp, et al., CD40 agonists alter tumor stroma and show efficacy against pancreatic carcinoma in mice and humans, Science 331 (2011) 1612–1616, https://doi.org/10.1126/science.1198443.
- [22] I. Comerford, Y. Harata-Lee, M.D. Bunting, C. Gregor, E.E. Kara, S.R. McColl, A myriad of functions and complex regulation of the CCR7/CCL19/CCL21 chemokine axis in the adaptive immune system, Cytokine Growth Factor Rev. 24 (2013) 269–283, https://doi.org/10.1016/j.cytogfr.2013.03.001.
- [23] S.K. Gautam, S.K. Batra, M. Jain, Molecular and metabolic regulation of immunosuppression in metastatic pancreatic ductal adenocarcinoma, Mol. Cancer 22 (2023) 118. https://doi.org/10.1186/s12943-023-01813-v.
- [24] S. Gomez, T. Tabernacki, J. Kobyra, P. Roberts, K.B. Chiappinelli, Combining epigenetic and immune therapy to overcome cancer resistance, Semin. Cancer Biol. 65 (2020) 99–113, https://doi.org/10.1016/j.semcancer.2019.12.019.
- [25] F. Long, Z. Lin, L. Li, M. Ma, Z. Lu, L. Jing, X. Li, C. Lin, Comprehensive landscape and future perspectives of circular RNAs in colorectal cancer, Mol. Cancer 20 (2021) 26, https://doi.org/10.1186/s12943-021-01318-6.
- [26] C. Cheng, F. Geng, X. Cheng, D. Guo, Lipid metabolism reprogramming and its potential targets in cancer, Cancer Commun. 38 (2018) 27, https://doi.org/ 10.1186/s40880-018-0301-4.
- [27] M. Reina-Campos, M. Heeg, K. Kennewick, I.T. Mathews, G. Galletti, V. Luna, Q. Nguyen, H. Huang, J.J. Milner, K.H. Hu, et al., Metabolic programs of T cell tissue residency empower tumour immunity, Nature (2023), https://doi.org/10.1038/s41586-023-06483-w.
- [28] Y. Zhao, J. Du, X. Shen, Targeting myeloid-derived suppressor cells in tumor immunotherapy: current, future and beyond, Front. Immunol. 14 (2023) 1157537, https://doi.org/10.3389/fimmu.2023.1157537.
- [29] B.K. Nirala, T.D. Patel, L. Kurenbekova, R. Shuck, A. Dasgupta, N. Rainusso, C. Coarfa, J.T. Yustein, MYC regulates CSF1 expression via microRNA 17/20a to modulate tumor-associated macrophages in osteosarcoma, JCI Insight 8 (2023), https://doi.org/10.1172/jci.insight.164947.
- [30] D.H. Hwang, J.A. Kim, J.Y. Lee, Mechanisms for the activation of Toll-like receptor 2/4 by saturated fatty acids and inhibition by docosahexaenoic acid, Eur. J. Pharmacol. 785 (2016) 24–35, https://doi.org/10.1016/j.ejphar.2016.04.024.
- [31] S. Kobayashi, T. Wannakul, K. Sekino, Y. Takahashi, Y. Kagawa, H. Miyazaki, B.A. Umaru, S. Yang, Y. Yamamoto, Y. Owada, Fatty acid-binding protein 5 limits the generation of Foxp3(+) regulatory T cells through regulating plasmacytoid dendritic cell function in the tumor microenvironment, Int. J. Cancer 150 (2022) 152–163, https://doi.org/10.1002/ijc.33777.
- [32] Y. Xu, Q. Liu, M. Zhong, Z. Wang, Z. Chen, Y. Zhang, H. Xing, Z. Tian, K. Tang, X. Liao, et al., 2B4 costimulatory domain enhancing cytotoxic ability of anti-CD5 chimeric antigen receptor engineered natural killer cells against T cell malignancies, J. Hematol. Oncol. 12 (2019) 49, https://doi.org/10.1186/s13045-019-0732-7.
- [33] L.J. Bayne, G.L. Beatty, N. Jhala, C.E. Clark, A.D. Rhim, B.Z. Stanger, R.H. Vonderheide, Tumor-derived granulocyte-macrophage colony-stimulating factor regulates myeloid inflammation and T cell immunity in pancreatic cancer, Cancer Cell 21 (2012) 822–835, https://doi.org/10.1016/j.ccr.2012.04.025.
- [34] L. Mashouri, H. Yousefi, A.R. Aref, A.M. Ahadi, F. Molaei, S.K. Alahari, Exosomes: composition, biogenesis, and mechanisms in cancer metastasis and drug resistance, Mol. Cancer 18 (2019) 75, https://doi.org/10.1186/s12943-019-0991-5.
- [35] D.M. Kuang, Y. Wu, N. Chen, J. Cheng, S.M. Zhuang, L. Zheng, Tumor-derived hyaluronan induces formation of immunosuppressive macrophages through transient early activation of monocytes, Blood 110 (2007) 587–595, https://doi.org/10.1182/blood-2007-01-068031.
- [36] Z. Lu, N. McBrearty, J. Chen, V.S. Tomar, H. Zhang, G. De Rosa, A. Tan, A.M. Weljie, D.P. Beiting, Z. Miao, et al., ATF3 and CH25H regulate effector trogocytosis and anti-tumor activities of endogenous and immunotherapeutic cytotoxic T lymphocytes, Cell Metab 34 (2022) 1342–1358.e1347, https://doi. org/10.1016/j.cmet.2022.08.007.
- [37] J.D. Shields, I.C. Kourtis, A.A. Tomei, J.M. Roberts, M.A. Swartz, Induction of lymphoidlike stroma and immune escape by tumors that express the chemokine CCL21, Science 328 (2010) 749–752, https://doi.org/10.1126/science.1185837.
- [38] E.V. Dang, J.G. McDonald, D.W. Russell, J.G. Cyster, Oxysterol restraint of cholesterol synthesis prevents AIM2 inflammasome activation, Cell 171 (2017) 1057–1071.e1011, https://doi.org/10.1016/j.cell.2017.09.029.
- [39] R.D. Cervantes-Villagrana, D. Albores-García, A.R. Cervantes-Villagrana, S.J. García-Acevez, Tumor-induced neurogenesis and immune evasion as targets of innovative anti-cancer therapies, Signal Transduct Target Ther 5 (2020) 99, https://doi.org/10.1038/s41392-020-0205-z.
- [40] X. Zhang, H. Shi, X. Yuan, P. Jiang, H. Qian, W. Xu, Tumor-derived exosomes induce N2 polarization of neutrophils to promote gastric cancer cell migration, Mol. Cancer 17 (2018) 146, https://doi.org/10.1186/s12943-018-0898-6.
- [41] A. Shang, C. Gu, W. Wang, X. Wang, J. Sun, B. Zeng, C. Chen, W. Chang, Y. Ping, P. Ji, et al., Exosomal circPACRGL promotes progression of colorectal cancer via the miR-142-3p/miR-506-3p- TGF-β1 axis, Mol. Cancer 19 (2020) 117, https://doi.org/10.1186/s12943-020-01235-0.
- [42] Z. Yang, Y. Huo, S. Zhou, J. Guo, X. Ma, T. Li, C. Fan, L. Wang, Cancer cell-intrinsic XBP1 drives immunosuppressive reprogramming of intratumoral myeloid cells by promoting cholesterol production, Cell Metab 34 (2022) 2018–2035.e2018, https://doi.org/10.1016/j.cmet.2022.10.010.
- [43] H.M. Chen, W. van der Touw, Y.S. Wang, K. Kang, S. Mai, J. Zhang, D. Alsina-Beauchamp, J.A. Duty, S.K. Mungamuri, B. Zhang, et al., Blocking immunoinhibitory receptor LILRB2 reprograms tumor-associated myeloid cells and promotes antitumor immunity, J. Clin. Invest. 128 (2018) 5647–5662, https://doi.org/10.1172/jci97570.
- [44] N.M. Chapman, M.R. Boothby, H. Chi, Metabolic coordination of T cell quiescence and activation, Nat. Rev. Immunol. 20 (2020) 55–70, https://doi.org/ 10.1038/s41577-019-0203-y.
- [45] S.A. Lim, J. Wei, T.M. Nguyen, H. Shi, W. Su, G. Palacios, Y. Dhungana, N.M. Chapman, L. Long, J. Saravia, et al., Lipid signalling enforces functional specialization of T(reg) cells in tumours, Nature 591 (2021) 306–311, https://doi.org/10.1038/s41586-021-03235-6.
- [46] Y. Kidani, H. Elsaesser, M.B. Hock, L. Vergnes, K.J. Williams, J.P. Argus, B.N. Marbois, E. Komisopoulou, E.B. Wilson, T.F. Osborne, et al., Sterol regulatory element-binding proteins are essential for the metabolic programming of effector T cells and adaptive immunity, Nat. Immunol. 14 (2013) 489–499, https:// doi.org/10.1038/ni.2570.
- [47] T. Gruosso, M. Gigoux, V.S.K. Manem, N. Bertos, D. Zuo, I. Perlitch, S.M.I. Saleh, H. Zhao, M. Souleimanova, R.M. Johnson, et al., Spatially distinct tumor immune microenvironments stratify triple-negative breast cancers, J. Clin. Invest. 129 (2019) 1785–1800, https://doi.org/10.1172/jci96313.
- [48] L. Yang, Y. Zhang, Tumor-associated macrophages: from basic research to clinical application, J. Hematol. Oncol. 10 (2017) 58, https://doi.org/10.1186/ s13045-017-0430-2.

- [49] S.G. Pai, B.A. Carneiro, J.M. Mota, R. Costa, C.A. Leite, R. Barroso-Sousa, J.B. Kaplan, Y.K. Chae, F.J. Giles, Wnt/beta-catenin pathway: modulating anticancer immune response, J. Hematol. Oncol. 10 (2017) 101, https://doi.org/10.1186/s13045-017-0471-6.
- [50] Y. Wang, K.C.C. Johnson, M.E. Gatti-Mays, Z. Li, Emerging strategies in targeting tumor-resident myeloid cells for cancer immunotherapy, J. Hematol. Oncol. 15 (2022) 118, https://doi.org/10.1186/s13045-022-01335-y.
- [51] C.A. Dumitru, S. Lang, S. Brandau, Modulation of neutrophil granulocytes in the tumor microenvironment: mechanisms and consequences for tumor progression, Semin. Cancer Biol. 23 (2013) 141–148, https://doi.org/10.1016/j.semcancer.2013.02.005.
- [52] M.G. Cecchini, M.G. Dominguez, S. Mocci, A. Wetterwald, R. Felix, H. Fleisch, O. Chisholm, W. Hofstetter, J.W. Pollard, E.R. Stanley, Role of colony stimulating factor-1 in the establishment and regulation of tissue macrophages during postnatal development of the mouse, Development 120 (1994) 1357–1372, https://doi.org/10.1242/dev.120.6.1357.
- [53] T. Kitamura, B.Z. Qian, D. Soong, L. Cassetta, R. Noy, G. Sugano, Y. Kato, J. Li, J.W. Pollard, CCL2-induced chemokine cascade promotes breast cancer metastasis by enhancing retention of metastasis-associated macrophages, J. Exp. Med. 212 (2015) 1043–1059, https://doi.org/10.1084/jem.20141836.
- [54] S.M. Morrissey, F. Zhang, C. Ding, D.E. Montoya-Durango, X. Hu, C. Yang, Z. Wang, F. Yuan, M. Fox, H.G. Zhang, et al., Tumor-derived exosomes drive immunosuppressive macrophages in a pre-metastatic niche through glycolytic dominant metabolic reprogramming, Cell Metab 33 (2021) 2040–2058.e2010, https://doi.org/10.1016/j.cmet.2021.09.002.
- [55] H. Iwamoto, M. Abe, Y. Yang, D. Cui, T. Seki, M. Nakamura, K. Hosaka, S. Lim, J. Wu, X. He, et al., Cancer lipid metabolism confers antiangiogenic drug resistance, Cell Metab 28 (2018) 104–117.e105, https://doi.org/10.1016/j.cmet.2018.05.005.
- [56] C.S. Field, F. Baixauli, R.L. Kyle, D.J. Puleston, A.M. Cameron, D.E. Sanin, K.L. Hippen, M. Loschi, G. Thangavelu, M. Corrado, et al., Mitochondrial integrity regulated by lipid metabolism is a cell-intrinsic checkpoint for Treg suppressive function, Cell Metab 31 (2020) 422–437.e425, https://doi.org/10.1016/j. cmet.2019.11.021.
- [57] E. Guilbaud, T. Barouillet, M. Ilie, C. Borowczyk, S. Ivanov, V. Sarrazy, N. Vaillant, M. Ayrault, A. Castiglione, G. Rignol, et al., Cholesterol efflux pathways hinder KRAS-driven lung tumor progenitor cell expansion, Cell Stem Cell 30 (2023) 800–817.e809, https://doi.org/10.1016/j.stem.2023.05.005.
- [58] A. Brendolan, V. Russo, Targeting cholesterol homeostasis in hematopoietic malignancies, Blood 139 (2022) 165–176, https://doi.org/10.1182/ blood.2021012788.
- [59] W. Yang, Y. Bai, Y. Xiong, J. Zhang, S. Chen, X. Zheng, X. Meng, L. Li, J. Wang, C. Xu, et al., Potentiating the antitumour response of CD8(+) T cells by modulating cholesterol metabolism, Nature 531 (2016) 651–655, https://doi.org/10.1038/nature17412.
- [60] X. Ma, L. Xiao, L. Liu, L. Ye, P. Su, E. Bi, Q. Wang, M. Yang, J. Qian, Q. Yi, CD36-mediated ferroptosis dampens intratumoral CD8(+) T cell effector function and impairs their antitumor ability, Cell Metab 33 (2021) 1001–1012.e1005, https://doi.org/10.1016/j.cmet.2021.02.015.
- [61] K. Lee, W. Tirasophon, X. Shen, M. Michalak, R. Prywes, T. Okada, H. Yoshida, K. Mori, R.J. Kaufman, IRE1-mediated unconventional mRNA splicing and S2Pmediated ATF6 cleavage merge to regulate XBP1 in signaling the unfolded protein response, Genes Dev. 16 (2002) 452–466, https://doi.org/10.1101/ gad.964702.
- [62] P. Goossens, J. Rodriguez-Vita, A. Etzerodt, M. Masse, O. Rastoin, V. Gouirand, T. Ulas, O. Papantonopoulou, M. Van Eck, N. Auphan-Anezin, et al., Membrane cholesterol efflux drives tumor-associated macrophage reprogramming and tumor progression, Cell Metab 29 (2019) 1376–1389.e1374, https://doi.org/ 10.1016/j.cmet.2019.02.016.
- [63] A. Sica, A. Bleve, M.C. Garassino, Membrane cholesterol regulates macrophage plasticity in cancer, Cell Metab 29 (2019) 1238–1240, https://doi.org/ 10.1016/j.cmet.2019.05.011.
- [64] D. Guo, F. Reinitz, M. Youssef, C. Hong, D. Nathanson, D. Akhavan, D. Kuga, A.N. Amzajerdi, H. Soto, S. Zhu, et al., An LXR agonist promotes glioblastoma cell death through inhibition of an EGFR/AKT/SREBP-1/LDLR-dependent pathway, Cancer Discov. 1 (2011) 442–456, https://doi.org/10.1158/2159-8290.Cd-11-0102.
- [65] D. Gu, F. Zhou, H. You, J. Gao, T. Kang, D. Dixit, Q. Wu, K. Yang, S. Ci, D. Shan, et al., SREBP2 maintains glioblastoma stem cells through keeping the balance between cholesterol biosynthesis and uptake, Neuro Oncol. (2023), https://doi.org/10.1093/neuonc/noad060.
- [66] T. Wang, Y. Zhou, Y. Fan, H. Duan, X. Guo, J. Chang, Y. Jiang, C. Li, Z. Fu, Y. Gao, et al., PERK-mediated cholesterol excretion from IDH mutant glioma determines anti-tumoral polarization of microglia, Adv. Sci. 10 (2023) e2205949, https://doi.org/10.1002/advs.202205949.
- [67] G.R. Villa, J.J. Hulce, C. Zanca, J. Bi, S. Ikegami, G.L. Cahill, Y. Gu, K.M. Lum, K. Masui, H. Yang, et al., An LXR-cholesterol Axis creates a metabolic Codependency for brain cancers, Cancer Cell 30 (2016) 683–693, https://doi.org/10.1016/j.ccell.2016.09.008.
- [68] Z. Lu, J. Chen, P. Yu, M.J. Atherton, J. Gui, V.S. Tomar, J.D. Middleton, N.T. Sullivan, S. Singhal, S.S. George, et al., Tumor factors stimulate lysosomal degradation of tumor antigens and undermine their cross-presentation in lung cancer, Nat. Commun. 13 (2022) 6623, https://doi.org/10.1038/s41467-022-34428-w.
- [69] A.T. Shamshiev, F. Ampenberger, B. Ernst, L. Rohrer, B.J. Marsland, M. Kopf, Dyslipidemia inhibits Toll-like receptor-induced activation of CD8alpha-negative dendritic cells and protective Th1 type immunity, J. Exp. Med. 204 (2007) 441–452, https://doi.org/10.1084/jem.20061737.
- [70] J. Huang, L. Zhang, D. Wan, L. Zhou, S. Zheng, S. Lin, Y. Qiao, Extracellular matrix and its therapeutic potential for cancer treatment, Signal Transduct Target Ther 6 (2021) 153, https://doi.org/10.1038/s41392-021-00544-0.
- [71] V. Mohan, A. Das, I. Sagi, Emerging roles of ECM remodeling processes in cancer, Semin. Cancer Biol. 62 (2020) 192–200, https://doi.org/10.1016/j. semcancer.2019.09.004.
- [72] H. Shimano, R. Sato, SREBP-regulated lipid metabolism: convergent physiology divergent pathophysiology, Nat. Rev. Endocrinol. 13 (2017) 710–730, https://doi.org/10.1038/nrendo.2017.91.
- [73] R. Bertolio, F. Napoletano, M. Mano, S. Maurer-Stroh, M. Fantuz, A. Zannini, S. Bicciato, G. Sorrentino, G. Del Sal, Sterol regulatory element binding protein 1 couples mechanical cues and lipid metabolism, Nat. Commun. 10 (2019) 1326, https://doi.org/10.1038/s41467-019-09152-7.
- [74] T. Tsuchida, S.L. Friedman, Mechanisms of hepatic stellate cell activation, Nat. Rev. Gastroenterol. Hepatol. 14 (2017) 397–411, https://doi.org/10.1038/ nrgastro.2017.38.
- [75] T. Teratani, K. Tomita, T. Suzuki, T. Oshikawa, H. Yokoyama, K. Shimamura, S. Tominaga, S. Hiroi, R. Irie, Y. Okada, et al., A high-cholesterol diet exacerbates liver fibrosis in mice via accumulation of free cholesterol in hepatic stellate cells, Gastroenterology 142 (2012) 152–164.e110, https://doi.org/10.1053/j. gastro.2011.09.049.
- [76] X. Li, J. Pan, T. Liu, W. Yin, Q. Miao, Z. Zhao, Y. Gao, W. Zheng, H. Li, R. Deng, et al., Novel TCF21(high) pericyte subpopulation promotes colorectal cancer metastasis by remodelling perivascular matrix, Gut 72 (2023) 710–721, https://doi.org/10.1136/gutjnl-2022-327913.
- [77] G. Burgstaller, B. Oehrle, M. Gerckens, E.S. White, H.B. Schiller, O. Eickelberg, The instructive extracellular matrix of the lung: basic composition and alterations in chronic lung disease, Eur. Respir. J. 50 (2017), https://doi.org/10.1183/13993003.01805-2016.
- [78] C. Li, S. Qiu, X. Liu, F. Guo, J. Zhai, Z. Li, L. Deng, L. Ge, H. Qian, L. Yang, et al., Extracellular matrix-derived mechanical force governs breast cancer cell stemness and quiescence transition through integrin-DDR signaling, Signal Transduct Target Ther 8 (2023) 247, https://doi.org/10.1038/s41392-023-01453-0.
- [79] C. Ramirez, A.D. Hauser, E.A. Vucic, D. Bar-Sagi, Plasma membrane V-ATPase controls oncogenic RAS-induced macropinocytosis, Nature 576 (2019) 477–481, https://doi.org/10.1038/s41586-019-1831-x.
- [80] H. Yamaguchi, Y. Takeo, S. Yoshida, Z. Kouchi, Y. Nakamura, K. Fukami, Lipid rafts and caveolin-1 are required for invadopodia formation and extracellular matrix degradation by human breast cancer cells, Cancer Res. 69 (2009) 8594–8602, https://doi.org/10.1158/0008-5472.Can-09-2305.
- [81] J. Wang, Y. Chen, Z. Luo, Q. Huang, Y. Zhang, H. Ning, S. Liu, J. Wang, X. Han, Citri Reticulatae Pericarpium-Reynoutria japonica Houtt. herb pair suppresses breast cancer liver metastasis by targeting ECM1-mediated cholesterol biosynthesis pathway, Phytomedicine 116 (2023) 154896, https://doi.org/10.1016/j. phymed.2023.154896.
- [82] L.G.N. de Almeida, H. Thode, Y. Eslambolchi, S. Chopra, D. Young, S. Gill, L. Devel, A. Dufour, Matrix metalloproteinases: from molecular mechanisms to physiology, pathophysiology, and pharmacology, Pharmacol. Rev. 74 (2022) 712–768, https://doi.org/10.1124/pharmrev.121.000349.

- [83] Z. Yuan, Y. Li, S. Zhang, X. Wang, H. Dou, X. Yu, Z. Zhang, S. Yang, M. Xiao, Extracellular matrix remodeling in tumor progression and immune escape: from mechanisms to treatments, Mol. Cancer 22 (2023) 48, https://doi.org/10.1186/s12943-023-01744-8.
- [84] R.J. King, P.K. Singh, K. Mehla, The cholesterol pathway: impact on immunity and cancer, Trends Immunol. 43 (2022) 78–92, https://doi.org/10.1016/j. it.2021.11.007.
- [85] M. Swamy, K. Beck-Garcia, E. Beck-Garcia, F.A. Hartl, A. Morath, O.S. Yousefi, E.P. Dopfer, E. Molnár, A.K. Schulze, R. Blanco, et al., A cholesterol-based allostery model of T cell receptor phosphorylation, Immunity 44 (2016) 1091–1101, https://doi.org/10.1016/j.immuni.2016.04.011.
- [86] C. Pich, I. Teiti, P. Rochaix, B. Mariamé, B. Couderc, G. Favre, A.F. Tilkin-Mariamé, Statins reduce melanoma development and metastasis through MICA overexpression, Front. Immunol. 4 (2013) 62, https://doi.org/10.3389/fimmu.2013.00062.
- [87] L. Strauss, M.A.A. Mahmoud, J.D. Weaver, N.M. Tijaro-Ovalle, A. Christofides, Q. Wang, R. Pal, M. Yuan, J. Asara, N. Patsoukis, et al., Targeted deletion of PD-1 in myeloid cells induces antitumor immunity, Sci Immunol 5 (2020), https://doi.org/10.1126/sciimmunol.aay1863.
- [88] O.E. Rahma, F.S. Hodi, The intersection between tumor angiogenesis and immune suppression, Clin. Cancer Res. 25 (2019) 5449–5457, https://doi.org/ 10.1158/1078-0432.Ccr-18-1543.
- [89] L. Fang, S.H. Choi, J.S. Baek, C. Liu, F. Almazan, F. Ulrich, P. Wiesner, A. Taleb, E. Deer, J. Pattison, et al., Control of angiogenesis by AIBP-mediated cholesterol efflux, Nature 498 (2013) 118–122, https://doi.org/10.1038/nature12166.
- [90] A. Sene, A.A. Khan, D. Cox, R.E. Nakamura, A. Santeford, B.M. Kim, R. Sidhu, M.D. Onken, J.W. Harbour, S. Hagbi-Levi, et al., Impaired cholesterol efflux in senescent macrophages promotes age-related macular degeneration, Cell Metab 17 (2013) 549–561, https://doi.org/10.1016/j.cmet.2013.03.009.
- [91] B. Han, F. Alonso-Valenteen, Z. Wang, N. Deng, T.Y. Lee, B. Gao, Y. Zhang, Y. Xu, X. Zhang, S. Billet, et al., A chemokine regulatory loop induces cholesterol synthesis in lung-colonizing triple-negative breast cancer cells to fuel metastatic growth, Mol. Ther. 30 (2022) 672–687, https://doi.org/10.1016/j. ymthe.2021.07.003.
- [92] J. Yarmolinsky, C.J. Bull, E.E. Vincent, J. Robinson, A. Walther, G.D. Smith, S.J. Lewis, C.L. Relton, R.M. Martin, Association between genetically proxied inhibition of HMG-CoA reductase and epithelial ovarian cancer, JAMA 323 (2020) 646–655, https://doi.org/10.1001/jama.2020.0150.
- [93] M. Soncini, G. Corna, M. Moresco, N. Coltella, U. Restuccia, D. Maggioni, L. Raccosta, C.Y. Lin, F. Invernizzi, R. Crocchiolo, et al., 24-Hydroxycholesterol participates in pancreatic neuroendocrine tumor development, Proc Natl Acad Sci U S A 113 (2016) E6219–e6227, https://doi.org/10.1073/ pnas.1613332113.
- [94] P. Romani, I. Brian, G. Santinon, A. Pocaterra, M. Audano, S. Pedretti, S. Mathieu, M. Forcato, S. Bicciato, J.B. Manneville, et al., Extracellular matrix mechanical cues regulate lipid metabolism through Lipin-1 and SREBP, Nat. Cell Biol. 21 (2019) 338–347, https://doi.org/10.1038/s41556-018-0270-5.
- [95] M.T. Snaebjornsson, S. Janaki-Raman, A. Schulze, Greasing the wheels of the cancer machine: the role of lipid metabolism in cancer, Cell Metab 31 (2020) 62–76, https://doi.org/10.1016/j.cmet.2019.11.010.
- [96] S. Gill, J. Stevenson, I. Kristiana, A.J. Brown, Cholesterol-dependent degradation of squalene monooxygenase, a control point in cholesterol synthesis beyond HMG-CoA reductase, Cell Metab 13 (2011) 260–273, https://doi.org/10.1016/j.cmet.2011.01.015.
- [97] N.K. Chua, H.W. Coates, A.J. Brown, Squalene monooxygenase: a journey to the heart of cholesterol synthesis, Prog. Lipid Res. 79 (2020) 101033, https://doi. org/10.1016/j.plipres.2020.101033.
- [98] A.J. Brown, N.K. Chua, N. Yan, The shape of human squalene epoxidase expands the arsenal against cancer, Nat. Commun. 10 (2019) 888, https://doi.org/ 10.1038/s41467-019-08866-y.
- [99] S.Y. Jun, A.J. Brown, N.K. Chua, J.Y. Yoon, J.J. Lee, J.O. Yang, I. Jang, S.J. Jeon, T.I. Choi, C.H. Kim, et al., Reduction of squalene epoxidase by cholesterol accumulation accelerates colorectal cancer progression and metastasis, Gastroenterology 160 (2021) 1194–1207.e1128, https://doi.org/10.1053/j. gastro.2020.09.009.
- [100] J. Garcia-Bermudez, L. Baudrier, E.C. Bayraktar, Y. Shen, K. La, R. Guarecuco, B. Yucel, D. Fiore, B. Tavora, E. Freinkman, et al., Squalene accumulation in cholesterol auxotrophic lymphomas prevents oxidative cell death, Nature 567 (2019) 118–122, https://doi.org/10.1038/s41586-019-0945-5.
- [101] L. Ma, L. Wang, A.T. Nelson, C. Han, S. He, M.A. Henn, K. Menon, J.J. Chen, A.E. Baek, A. Vardanyan, et al., 27-Hydroxycholesterol acts on myeloid immune cells to induce T cell dysfunction, promoting breast cancer progression, Cancer Lett. 493 (2020) 266–283, https://doi.org/10.1016/j.canlet.2020.08.020.
- [102] R. Fang, X. Chen, S. Zhang, H. Shi, Y. Ye, H. Shi, Z. Zou, P. Li, Q. Guo, L. Ma, et al., EGFR/SRC/ERK-stabilized YTHDF2 promotes cholesterol dysregulation and invasive growth of glioblastoma, Nat. Commun. 12 (2021) 177, https://doi.org/10.1038/s41467-020-20379-7.
- [103] D. Cai, J. Wang, B. Gao, J. Li, F. Wu, J.X. Zou, J. Xu, Y. Jiang, H. Zou, Z. Huang, et al., RORγ is a targetable master regulator of cholesterol biosynthesis in a cancer subtype, Nat. Commun. 10 (2019) 4621, https://doi.org/10.1038/s41467-019-12529-3.
- [104] S.H. Moon, C.H. Huang, S.L. Houlihan, K. Regunath, W.A. Freed-Pastor, J.P.t. Morris, D.F. Tschaharganeh, E.R. Kastenhuber, A.M. Barsotti, R. Culp-Hill, et al., p53 represses the mevalonate pathway to mediate tumor suppression, Cell 176 (2019) 564–580.e519, https://doi.org/10.1016/j.cell.2018.11.011.
- [105] M. Luo, L. Bao, Y. Chen, Y. Xue, Y. Wang, B. Zhang, C. Wang, C.D. Corley, J.G. McDonald, A. Kumar, et al., ZMYND8 is a master regulator of 27-hydroxycholesterol that promotes tumorigenicity of breast cancer stem cells, Sci. Adv. 8 (2022) eabn5295, https://doi.org/10.1126/sciadv.abn5295.
- [106] L. Gabitova-Cornell, A. Surumbayeva, S. Peri, J. Franco-Barraza, D. Restifo, N. Weitz, C. Ogier, A.R. Goldman, T.R. Hartman, R. Francescone, et al., Cholesterol pathway inhibition induces TGF-β signaling to promote basal differentiation in pancreatic cancer, Cancer Cell 38 (2020) 567–583.e511, https://doi.org/ 10.1016/j.ccell.2020.08.015.
- [107] M. Tan, S. Yang, X. Xu, High-density lipoprotein cholesterol and carcinogenesis, Trends Endocrinol Metab 34 (2023) 303–313, https://doi.org/10.1016/j. tem.2023.02.009.
- [108] M. Mihajlovic, T. Gojkovic, S. Vladimirov, M. Miljkovic, A. Stefanovic, J. Vekic, D. Zeljkovic, B. Trifunovic, J. Kotur-Stevuljevic, V. Spasojevic-Kalimanovska, et al., Changes in lecithin: cholesterol acyltransferase, cholesteryl ester transfer protein and paraoxonase-1 activities in patients with colorectal cancer, Clin. Biochem. 63 (2019) 32–38, https://doi.org/10.1016/j.clinbiochem.2018.11.010.
- [109] T.J. Zhao, N. Zhu, Y.N. Shi, Y.X. Wang, C.J. Zhang, C.F. Deng, D.F. Liao, L. Qin, Targeting HDL in tumor microenvironment: new hope for cancer therapy, J. Cell. Physiol. 236 (2021) 7853–7873, https://doi.org/10.1002/jcp.30412.
- [110] P.M.R. Pereira, K. Mandleywala, S. Monette, M. Lumish, K.M. Tully, S.S. Panikar, M. Cornejo, A. Mauguen, A. Ragupathi, N.C. Keltee, et al., Caveolin-1 temporal modulation enhances antibody drug efficacy in heterogeneous gastric cancer, Nat. Commun. 13 (2022) 2526, https://doi.org/10.1038/s41467-022-30142-9.
- [111] J. Kim, B. Thompson, S. Han, Y. Lotan, J.G. McDonald, J. Ye, Uptake of HDL-cholesterol contributes to lipid accumulation in clear cell renal cell carcinoma, Biochim. Biophys. Acta Mol. Cell Biol. Lipids 1864 (2019) 158525, https://doi.org/10.1016/j.bbalip.2019.158525.
- [112] N. Noguchi, Y. Urano, W. Takabe, Y. Saito, New aspects of 24(S)-hydroxycholesterol in modulating neuronal cell death, Free Radic. Biol. Med. 87 (2015) 366–372, https://doi.org/10.1016/j.freeradbiomed.2015.06.036.
- [113] K. Georgila, D. Vyrla, E. Drakos, Apolipoprotein A-I (ApoA-I), immunity, inflammation and cancer, Cancers 11 (2019), https://doi.org/10.3390/ cancers11081097.
- [114] C. Zeng, A. Riad, R.H. Mach, The biological function of sigma-2 receptor/TMEM97 and its utility in PET imaging studies in cancer, Cancers 12 (2020), https:// doi.org/10.3390/cancers12071877.
- [115] A. Ortiz, J. Gui, F. Zahedi, P. Yu, C. Cho, S. Bhattacharya, C.J. Carbone, Q. Yu, K.V. Katlinski, Y.V. Katlinskaya, et al., An interferon-driven oxysterol-based defense against tumor-derived extracellular vesicles, Cancer Cell 35 (2019) 33–45.e36, https://doi.org/10.1016/j.ccell.2018.12.001.
- [116] Y. Saito, D. Yin, N. Kubota, X. Wang, A. Filliol, H. Remotti, A. Nair, L. Fazlollahi, Y. Hoshida, I. Tabas, et al., A therapeutically targetable TAZ-TEAD2 pathway drives the growth of hepatocellular carcinoma via ANLN and KIF23, Gastroenterology 164 (2023) 1279–1292, https://doi.org/10.1053/j.gastro.2023.02.043.

- [117] A.B. Chaves-Filho, A. Schulze, Cholesterol atlas of tumor microenvironment provides route to improved CAR-T therapy, Cancer Cell 41 (2023) 1204–1206, https://doi.org/10.1016/j.ccell.2023.05.013.
- [118] S. Uchida, H. Kinoh, T. Ishii, A. Matsui, T.A. Tockary, K.M. Takeda, H. Uchida, K. Osada, K. Itaka, K. Kataoka, Systemic delivery of messenger RNA for the treatment of pancreatic cancer using polyplex nanomicelles with a cholesterol moiety, Biomaterials 82 (2016) 221–228, https://doi.org/10.1016/j. biomaterials.2015.12.031.
- [119] C. Hong, J. Liang, J. Xia, Y. Zhu, Y. Guo, A. Wang, C. Lu, H. Ren, C. Chen, S. Li, et al., One stone four birds: a novel liposomal delivery system multifunctionalized with ginsenoside Rh2 for tumor targeting therapy, Nano-Micro Lett. 12 (2020) 129, https://doi.org/10.1007/s40820-020-00472-8.