

Genome Sequences of Rhinovirus B Isolates from Wisconsin Pediatric Respiratory Studies

Stephen B. Liggett,^a Yury A. Bochkov,^b Tressa Pappas,^b Robert F. Lemanske, Jr,^b James E. Gern,^b Naomi Sengamalay,^c Xuechu Zhao,^c Qi Su,^c Claire M. Fraser,^c Ann C. Palmenberg^d

Department of Medicine, University of South Florida Morsani College of Medicine, Tampa, Florida, USA^a; Department of Pediatrics, University of Wisconsin School of Medicine and Public Health, Madison, Wisconsin, USA^b; Institute for Genome Sciences, University of Maryland School of Medicine, Baltimore, Maryland, USA^c; Institute for Molecular Virology, University of Wisconsin-Madison, Madison, Wisconsin, USA^d

Nearly full-length RNA genome sequences for 39 rhinovirus B isolates (RV-B), representing 13 different genotypes, were resolved as part of ongoing studies at the University of Wisconsin that attempt to link rhinovirus (RV) diversity and respiratory disease in infants.

Received 24 February 2014 Accepted 6 March 2014 Published 27 March 2014

Citation Liggett SB, Bochkov YA, Pappas T, Lemanske RF, Jr, Gern JE, Sengamalay N, Zhao X, Su Q, Fraser CM, Palmenberg AC. 2014. Genome sequences of rhinovirus B isolates from Wisconsin pediatric respiratory studies. *Genome Announc.* 2(2):e00202-14. doi:10.1128/genomeA.00202-14.

Copyright © 2014 Liggett et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Ann C. Palmenberg, acpalmen@wisc.edu.

The University of Wisconsin hospitals and clinics in Madison, WI, support several ongoing studies examining why some children have mild illnesses while others have severe colds or even asthma attacks when they catch a rhinovirus-induced cold. The Childhood Origins of Asthma (COAST), RhinoGen, and T Regulatory Cells and Childhood Asthma (T-Reg) studies screen cohort infant nasal secretions from well/sick visits, with the goal of correlating respiratory viruses with the severity of illness. Of samples collected between 1999 and 2010, hundreds were identified as solitary infections by RNA rhinoviruses (RV), organisms in the *Enterovirus* genus of the *Picornaviridae* family. The screens used multiplex PCR assays (1), rhinovirus PCR (2), or both. The initial partial sequencing assigned species (A, B, or C) and preliminary genotype identifications, as has been reported (3). Multiple isolates were then reexamined using massively parallel sequencing techniques. The methods (4) were applied directly to patient samples without viral propagation in cells, a procedure that automated the sequence derivations and eliminated variant selection bias that might have been introduced by amplification. The single-pass methodology gave, on average, 93% full-genome coverage to a depth of 8 to 10 reads for 179 study-specific isolates. The sample titers that contributed to successful datasets ranged from 6×10^2 to 1.6×10^8 virions (viral RNA [vRNA] copy number per 350 μ l of nasal sample, determined by quantitative real-time PCR [qRT-PCR]), with a median value of $\sim 6 \times 10^4$ virions.

In historic RV taxonomy, panels of clinical isolates, as archived by the American Type Culture Collection, were indexed into an initial 99 RV-A and RV-B types after an assessment of antigenic cross-reactivity in rabbits (5). Current classification schemes assign RV strains to one of 3 species (A, B, or C) if they share >70% amino acid identity in the P1, 2C, and 3CD regions with other known members. Within species, the isolates are subdivided into numeric genotypes that respect the historic serotype-naming system but now rely almost entirely on observed pairwise nucleotide identity (>87 to 88%) in the VP1 or VP4/VP2 coding sequences (6). The delineations and poten-

tial new genotypes are periodically reviewed by the *Picornaviridae* Study Group of the International Committee on Taxonomy of Viruses (ICTV). The preferred nomenclature (6) designates the species letter (A, B, or C) and type number (e.g., A16). The isolate designations are unique to each accession number.

Within this context, the Wisconsin study resolved nearly full-genomes for 39 RV-B isolates representing 13 different genotypes. The data add depth to observations for the new genotypes B101 and B102 (7) and define novel genotypes, B103 and B104. As a consequence, a previous B52 field isolate (accession no. FJ445188) was also reclassified as B104 (3). Relative to prototype RV-B genomes, which average $\sim 7,209$ bases (b) (8), most of the 39 new assemblies were missing the difficult-to-sequence 5' and/or 3' termini (average, $\Delta 404$ b) and occasionally, short internal fragments (<100 b) for which the contigs were not explicitly linked. Nevertheless, every derived sequence (average, 6,753 b; median, 6,806 b) was unambiguously aligned with the index RV compilation (8) for accurate type identification (6).

Nucleotide sequence accession numbers. Each contiguous RV-B data set has been deposited in GenBank using the listed accession numbers. Each unit described here is the first genome version of the sequence of that isolate: B04, [JN798573](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN798573); B06, [JN562723](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN562723), [JN815243](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN815243), [JQ747745](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ747745), [JQ747748](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ747748), and [JX193795](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JX193795); B42, [JF781498](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781498), [JF781507](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781507), and [JN562724](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN562724); B48, [JN990698](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN990698); B69, [JN562721](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN562721) and [JQ245970](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ245970); B70, [JN990706](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN990706), [JQ245974](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ245974), and [JX074054](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JX074054); B72, [JN562726](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN562726), [JN614997](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN614997), [JN798562](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN798562), and [JQ245969](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ245969); B83, [JN990701](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN990701); B84, [JF781499](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781499), [JF781502](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781502), [JN541271](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN541271), [JN614991](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN614991), [JN798588](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN798588), [JQ837723](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ837723), and [JX074048](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JX074048); B101, [JF781500](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781500), [JF781501](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781501), and [JX074052](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JX074052); B102, [JX074053](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JX074053); B103, [JN614996](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN614996), [JN798572](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN798572), [JN815239](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JN815239), [JQ245972](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ245972), [JQ837717](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ837717), [JQ837721](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ837721), and [JQ994497](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JQ994497); B104, [JF781506](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/JF781506).

ACKNOWLEDGMENTS

This work was supported by National Institutes of Health Public Health Service grants P01-HL70831 (COAST), U19-AI070503 (Rhino-

Gen), R01-HL080072 (T-Reg), U19-AI104317 (to A.C.P.), and HL091490 (to S.B.L.). This project was also funded in part from the National Institute of Allergy and Infectious Diseases, National Institutes of Health, Department of Health and Human Services, under contract HHSN272200900009C.

REFERENCES

1. Lee WM, Grindle K, Pappas T, Marshall DJ, Moser MJ, Beaty EL, Shult PA, Prudent JR, Gern JE. 2007. High-throughput, sensitive, and accurate multiplex PCR-microsphere flow cytometry system for large-scale comprehensive detection of respiratory viruses. *J. Clin. Microbiol.* 45:2626–2634. <http://dx.doi.org/10.1128/JCM.02501-06>.
2. Lee WM, Kiesner C, Pappas T, Lee I, Grindle K, Jartti T, Jakiela B, Lemanske RF, Shult PA, Gern JE. 2007. A diverse group of previously unrecognized human rhinoviruses are common causes of respiratory illnesses in infants. *PLoS One* 2:e966. <http://dx.doi.org/10.1371/journal.pone.0000966>.
3. Lee WM, Lemanske RF, Evans MD, Vang F, Pappas T, Gangnon R, Jackson DJ, Gern JE. 2012. Human rhinovirus species and season of infection determine illness severity. *Am. J. Respir. Crit. Care Med.* 186: 886–891. <http://dx.doi.org/10.1164/rccm.201202-0330OC>.
4. Liggett SB, Bochkov YA, Pappas T, Lemanske RF, Jr, Gern JE, Sengamaly N, Zhao X, Su Q, Fraser CM, Palmenberg AC. 2014. Genome sequences of rhinovirus A isolates from Wisconsin pediatric respiratory studies. *Genome Announc.* 2(2):e00200-14. <http://dx.doi.org/10.1128/genomeA.00200-14>.
5. Hamparian VV, Colonno RJ, Cooney MK, Dick EC, Gwaltney JM, Hughes JH, Jordan WS, Kapikian AZ, Mogabgab WJ, Monto A, Phillips CA, Rueckert RR, Scheible JH, Stott EJ, Tyrrell DAJ. 1987. A collaborative report: rhinoviruses—extension of the numbering system from 89 to 100. *Virology* 159:191–192. [http://dx.doi.org/10.1016/0042-6822\(87\)90367-9](http://dx.doi.org/10.1016/0042-6822(87)90367-9).
6. McIntyre CL, Knowles NJ, Simmonds P. 2013. Proposals for the classification of human rhinovirus species A, B and C into genotypically assigned types. *J. Gen. Virol.* 94:1791–1806. <http://dx.doi.org/10.1099/vir.0.053686-0>.
7. McIntyre CL, Savolainen-Kopra C, Hovi T, Simmonds P. 2013. Recombination in the evolution of human rhinovirus genomes. *Arch. Virol.* 158: 1497–1515. <http://dx.doi.org/10.1007/s00705-013-1634-6>.
8. Palmenberg AC, Spiro D, Kuzmickas R, Wang S, Djikeng A, Rathe JA, Fraser-Liggett CM, Liggett SB. 2009. Sequencing and analyses of all known human rhinovirus genomes reveal structure and evolution. *Science* 324: 55–59. <http://dx.doi.org/10.1126/science.1165557>.