# **Variation in Active Site Amino Residues of H1N1 Swine Flu Neuraminidase**

G. Nageswara Rao, P. Srinivasarao, A. Apparao, and T.K. Rama Krishna Rao

Aditya Institute of Technology & Management {gnraoaitam,peri.srinivasarao}@yahoo.com, {apparaoallam,ramakrishnatk}@gmail.com

**Abstract.** In this paper, we report the variations of amino acid residues between H5N1 and H1N1 swine flu neuraminidase sequences at protein level. Random search in NCBI Flu database resulted in Canadian viral gene and analysis using blast technique revealed sites that are variant among sequences for which 3 dimensional structures were known. PDB summary database and multiple alignments were employed for validation of the results. Based on the mutations observed within active site region, homology derived model was constructed using swiss-pdb viewer. The residue variation observed was with respect to Tyr347 in H5N1 versus Asn344 in H1N1 neuraminidase sequence, which resulted in geometrical modification of ligand binding domain.

#### **1 Introduction**

Swine influenza was first proposed to be a disease related to human influenza during the 1918 flu pandemic. The H1N1 form of swine flu is one of the descendants of the strain that caused the 1918 flu pandemic [Jeffery K. Taubenberger, David M. Morens. 1918 Influenza: The mother of all pandemics. Rev Biomed 2006; 17:69-79]. The human influenza a virus continues to thrive among populations and continues to be a major cause of morbidity and mortality [Frost WH. Statistics of influenza morbidity. Public Health Rep. 1920; 35:584–97]. The virus showed various mutations [Glaser L, Stevens J, Zamarin D, Wilson IA, Garcia-Sastre A, Tumpey TM, et al. A single amino acid substitution in the 1918 influenza virus hemagglutinin changes the receptor binding specificity. J Virol. 2005; 79:11533–6] since it first originated thereby making the existing vaccines ineffective on a regular basis [Elodie Ghedin, Naomi A. Sengamalay, Martin Shumway et. al. Large-scale sequencing of human influenza reveals the dynamic nature of viral genome evolution. Nature 2005; 437, 1162-1166].

Influenza, commonly referred to as the [flu,](#page-8-0) is an infectious disease caused by RNA viruses of the family Orthomyxviridae (the influenza viruses), that affects birds and mammals. The most common symptoms of the disease are chills, fever, sore throat, muscle pains, severe headache, coughing, weakness and general discomfort. Typically, influenza is transmitted through the air by coughs or sneezes, creating aerosols containing the virus. Influenza can also be transmitted by bird droppings, saliva, nasal secretions, faeces and blood. An avian strain named H5N1 raised the concern of a

Aswatha Kumar M. et al. (Eds.): Proceedings of ICAdC, AISC 174, pp. 575–583. springerlink.com © Springer India 2013 new influenza pandemic, after it emerged in Asia in the 1990s. In April 2009 a novel flu strain evolved that combined genes from human, pig, and bird flu, referred as 'swine flu' [Yasushi Itoh, Kyoko Shinya, Maki Kiso et. al. In vitro and in vivo characterization of new swine-origin H1N1 influenza viruses. Nature 2009; 460, 1021-1025].

### **2 Materials and Methods**

The viral gene sequences were accessed and extracted from NCBI (National Centre for Biotechnology Information) Flu database [www.ncbi.nlm.nih.gov]. From the H1N1 sequences deposited in NCBI, the Canadian origin neuraminidase gene (Figure 1) was selected randomly to perform sequence comparisons.

The fasta format of the sequence selected for analysis is given below.

>gi|255734960|gb|ACU31180.1| neuraminidase [Influenza A virus (A/Canada-NS/RV1554/2009(H1N1))]

MNPNQKIITIGSVCMTIGMANLILQIGNIISIWISHSIQLGNQNQIETCNQSVIT YENNTWVNQTYVNISNTNFAAGQSVVSVKLAGNSSLCPVSGWAIYSKDNSV RIGSKGDVFVIREPFISCSPLECRTFFLTQGALLNDKHSNGTIKDRSPYRTLMSC PIGEVPSPYNSRFESVAWSASACHDGINWLTIGISGPDNGAVAVLKYNGIITDT IKSWRNNILRTQESECACVNGSCFTVMTDGPSNGQASYKIFRIEKGKIVKSVE MNAPNYHYEECSCYPDSSEITCVCRDNWHGSNRPWVSFNQNLEYQIGYICSG IFGDNPRPNDKTGSCGPVSSNGANGVKGFSFKYGNGVWIGRTKSISSRNGFE MIWDPNGWTGTDNNFSIKQDIVGINEWSGYSGSFVQHPELTGLDCIRPCFWV ELIRGRPKENTIWTSGSSISFCGVNSDTVGWSWPDGAELPFTIDK

ClustalW program [www.ebi.ac.uk/clustalw] was utilized to perform multiple sequence alignments. The template 3D structures were downloaded from Protein Data Bank [www.recsb.org/pdb]. PDB summary database [www.ebi.ac.uk/pdbsum] was employed to study active site residue region. Viral neuraminidase structure was built using Swiss-PdbViewer. Initially the sequence (H1N1 neuraminidase) to be modelled is loaded from Swiss model menu and then the option move raw sequence into the structure followed by move structure into raw sequence is performed. Then the reference sequence (3CL2) is loaded from open pdb file option of the file menu and performed iterative magic fit of the fit menu by which the target sequence and the template structure fits into each other.

GenBank: GO465699.1

## Influenza A virus (A/Canada-NS/RV1554/2009(H1N1)) segment 6 neuraminidase (NA) gene, complete cds



**Fig. 1.** H1N1 Neuraminidase gene selected for analysis

#### **3 Results and Discussion**

Initially BLAST analysis was employed to evaluate the percent identities, similarities and number of gaps. Apart from this, based on PAM and BLOSUM matrices, considering Score and E-value, alignments are chosen for structure predictions.

The neuraminidase belongs to sialidase superfamily and the data from NCBI suggests that Sialidases or neuraminidases function to bind and hydrolyze terminal sialic acid residues from various glycoconjugates as well as playing roles in pathogenesis, bacterial nutrition and cellular interactions. They have a six-bladed, beta-propeller fold with the non-viral sialidases containing 2-5 Asp-box motifs (most commonly Ser/Thr-X-Asp-[X]-Gly-X-Thr- Trp/Phe). This Conserved Domain also includes eubacterial, eukaryotic, and viral sialidases.

BLAST analysis was carried out with default parameters and the scores, top alignments are given in Figures 2 and 3.



**Fig. 2.** BLAST analysis graphical representation

From the above top two and below alignments, although 3CL2, a H5N1 neuraminidase was considered for further work because 3CL2 was bound with oseltamivir. Therefore, structural and sequential differences between 3CL2 and H1N1 sequences were performed (Figure 4).

<b>Accession</b>	<b>Description</b>	<b>Max score</b>	<b>Total score</b>	<b>Query coverage</b>	<b>E</b> value	Links
3NSS A	Chain A, The 2009 Pandemic H1n1 Neuraminidase N1 Lacks The 150-	797	797	82%	0.0	S
2HTY A	Chain A, N1 Neuraminidase >pdb 2HTY B Chain B, N1 Neuraminidase	750	750	82%	0.0	S
3CL2 A	Chain A, N1 Neuraminidase N294s + Oseltamivir >pdb 3CL2 B Chain E	744	744	82%	0.0	S
3CKZ A	Chain A, N1 Neuraminidase H274y + Zanamivir >pdb 3CL0   A Chain A,	743	743	82%	0.0	S
3CYE A	Chain A, Cyrstal Structure Of The Native 1918 H1n1 Neuraminidase F	737	737	82%	0.0	S
3BEQ A	Chain A, Neuraminidase Of ABREVIG MISSION11918 H1N1 STRAIN >p	733	733	82%	0.0	S
2HTV A	Chain A, N4 Neuraminidase >pdb 2HTV B Chain B, N4 Neuraminidase	558	558	82%	0.0	S
2HT5 A	Chain A, N8 Neuraminidase >pdb 2HT7 A Chain A, N8 Neuraminidase	468	468	81%	6e-164	S
309J A	Chain A, Influenza Na In Complex With Compound 5 >pdb 309K A Ch	466	466	80%	4e-163	S
3SAL A	Chain A, Crystal Structure Of Influenza A Virus Neuraminidase N5 >p	453	453	82%	4e-158	S
1NMB N	Chain N, The Structure Of A Complex Between The Nc10 Antibody Ar	382	382	99%	3e-129	S
1NNA A	Chain A, Three-Dimensional Structure Of Influenza A N9 Neuraminida	363	363	80%	7e-123	sy.
1XOE A	Chain A, N9 Tern Influenza Neuraminidase Complexed With (2r,4r,5r)-	363	363	80%	8e-123	s y
<b>INCA N</b>	Chain N, Refined Crystal Structure Of The Influenza Virus N9 Neuram	363	363	80%	9e-123	S
1A14 N	Chain N, Complex Between Nc10 Anti-Influenza Virus Neuraminidase :	363	363	80%	$1e-122$	s y
5NN9 A	Chain A, Refined Atomic Structures Of N9 Subtype Influenza Virus Ne	362	362	80%	2e-122	S
<b>1NCB N</b>	Chain N, Crystal Structures Of Two Mutant Neuraminidase-Antibody	362	362	80%	2e-122	S
1L7H A	Chain A, Crystal Structure Of R292k Mutant Influenza Virus Neuramin	361	361	80%	3e-122	sy.
3NN9 A	Chain A, Refined Atomic Structures Of N9 Subtype Influenza Virus Ne	361	361	80%	4e-122	S
1INY A	Chain A, A Sialic Acid Derived Phosphonate Analog Inhibits Different	361	361	80%	$5e-122$	S
6NN9 A	Chain A, Refined Atomic Structures Of N9 Subtype Influenza Virus Ne	361	361	80%	$5e-122$	S
<b>INCC N</b>	Chain N, Crystal Structures Of Two Mutant Neuraminidase-Antibody	360	360	80%	7e-122	S
4NN9 A	Chain A, Refined Atomic Structures Of N9 Subtype Influenza Virus Ne	360	360	80%	8e-122	S
1NMA N	Chain N, N9 Neuraminidase Complexes With Antibodies Nc41 And Nc1	360	360	80%	9e-122	S
<b>INCD N</b>	Chain N, Refined Crystal Structure Of The Influenza Virus N9 Neuram	360	360	80%	$1e-121$	S
1L7G A	Chain A, Crystal Structure Of E119g Mutant Influenza Virus Neuramir	360	360	80%	2e-121	S
2B8H A	Chain A, ANWSWHALEMAINE184 (H1N9) REASSORTANT INFLUENZA V	360	360	80%	2e-121	S
1V0Z A	Chain A, Structure Of Neuraminidase From English Duck Subtype N6	355	355	79%	1e-119	S

Sequences producing significant alignments:

#### **Fig. 3.** BLAST analysis result

However, a careful observation of active site lining residues resulted in residue mutation in human H1N1 sequence. In other words, the residue variation was observed with respect to Tyr347 in H5N1 versus Asn344 in H1N1 neuraminidase sequence. Owing to the active site residue mutation, the protein model was built using SPDBV software.

An active site residue mutation was identified on comparison with H5N1 avian flu Neuraminidase enzyme. Hence, the 3D structure of H1N1 neuraminidase was built which can aid in detecting more potent binding inhibitor using computer-aided drug binding and screening studies (Figures 4-7).

```
> Dpdb|3CL2|A S Chain A, N1 Neuraminidase N294s + Oseltamivir
pdb|3CL2|B S Chain B, N1 Neuraminidase N294s + Oseltamivir
pdb|3CL2|C S Chain C, N1 Neuraminidase N294s + Oseltamivir
pdb13CL21D S Chain D. N1 Neuraminidase N294s + Oseltamivir
pdb|3CL2|E S Chain E, N1 Neuraminidase N294s + Oseltamivir
pdb|3CL2|F S Chain F, N1 Neuraminidase N294s + Oseltamivir
pdb|3CL2|G S Chain G, N1 Neuraminidase N294s + Oseltamivir
pdb|3CL2|H S Chain H. N1 Neuraminidase N294s + Oseltamivir
Lenath=385Score = 744 bits (1920), Expect = 0.0, Method: Compositional matrix adjust.
Identities = 351/385 (91%), Positives = 375/385 (97%), Gaps = 0/385 (0%)
Query 83 VKLAGNSSLCPVSGWAIYSKDNSVRIGSKGDVFVIREPFISCSPLECRTFFLTQGALLND 142
           VKLAGNSSLCP++GWA+YSKDNS+RIGSKGDVFVIREPFISCS LECRTFFLTOGALLND
Sbjct 1 VKLAGNSSLCPINGWAVYSKDNSIRIGSKGDVFVIREPFISCSHLECRTFFLTQGALLND 60
Ouery 143 KHSNGTIKDRSPYRTLMSCPIGEVPSPYNSRFESVAWSASACHDGINWLTIGISGPDNGA 202
           KHSNGT+KDRSP+RTLMSCP+GE PSPYNSRFESVAWSASACHDG +WLTIGISGPDNGA
Sbjct 61 KHSNGTVKDRSPHRTLMSCPVGEAPSPYNSRFESVAWSASACHDGTSWLTIGISGPDNGA 120
Query 203 VAVLKYNGIITDTIKSWRNNILRTQESECACVNGSCFTVMTDGPSNGQASYKIFRIEKGK 262
           VAVLKYNGIITDTIKSWRNNILRTQESECACVNGSCFTVMTDGPSNGQASYKIF++EKGK
Sbict 121 VAVLKYNGIITDTIKSWRNNILRTOESECACVNGSCFTVMTDGPSNGOASYKIFKMEKGK 180
Ouerv 263 IVKSVEMNAPNYHYEECSCYPDSSEITCVCRDNWHGSNRPWVSFNONLEYOIGYICSGIF 322
           +VKSVE++APNYHYEECSCYP++ EITCVCRD+WHGSNRPWVSFNONLEYOIGYICSG+F
Sbjct 181 VVKSVELDAPNYHYEECSCYPNAGEITCVCRDSWHGSNRPWVSFNONLEYQIGYICSGVF 240
Ouerv 323 GDNPRPNDKTGSCGPVSSNGANGVKGFSFKYGNGVWIGRTKSISSRNGFEMIWDPNGWTG 382
           GDNPRPND TGSCGPVSSNGA GVKGFSFKYGNGVWIGRTKS +SR+GFEMIWDPNGWT
Sbjct 241 GDNPRPNDGTGSCGPVSSNGAYGVKGFSFKYGNGVWIGRTKSTNSRSGFEMIWDPNGWTE 300
Query 383 TDNNFSIKQDIVGINEWSGYSGSFVQHPELTGLDCIRPCFWVELIRGRPKENTIWTSGSS 442
           TD++FS+KQDIV I +WSGYSGSFVQHPELTGLDCIRPCFWVELIRGRPKE+TIWTSGSS
Sbjct 301 TDSSFSVKQDIVAITDWSGYSGSFVQHPELTGLDCIRPCFWVELIRGRPKESTIWTSGSS 360
Query 443 ISFCGVNSDTVGWSWPDGAELPFTI 467
           ISFCGVNSDTVGWSWPDGAELPFTI
Sbjct 361 ISFCGVNSDTVGWSWPDGAELPFTI 385
```
**Fig. 4.** Variations of amino acid residues between H5N1 and H1N1 neuraminidase sequences



**Fig. 5.** Active site region of 3CL2 bound to oseltamivir



**Fig. 6.** Comparison of docked images of superimposed H5N1 and H1N1 acitve site regions showing Tyr347 of H5N1 replaced by Asn344 in H1N1



**Fig. 7.** Modelled protein with interacting H-bonds

### **Conclusion**

Literature reports suggest the importance of computational tools in finding few features that are relevant and important in understanding the structure and function of various mutational events in genes or proteins. One such study reported in this paper suggested the fact that with few computational efforts, variations in amino acid residue regions within the protein sequence can be known and accurate homology models can be built within short period of time. In this work, an active site residue mutation was identified in H1N1 neuraminidase upon comparison with H5N1 avian flu Neuraminidase enzyme. Hence, the 3D structure of H1N1 neuraminidase was built which can aid in detecting more potent binding inhibitor using computer-aided drug binding and screening studies.

### **References**

- 1. Olsen, C.W., Brown, I.H., Easterday, B.C., Reeth, K.V.: Diseases of swine By Straw, B.E., Taylor, D.J., Swine Influenza. ch. 28 , pp. 469–470
- 2. http://www.who.int
- 3. AlKhawaja, S.: Consultant, Infectious Disease Physician. Ministry of Health Kingdom of Bahrain Medical Bulletin 31(2), 1–4 (2009)
- 4. Brown, D.: System set up after SARS epidemic was slow to alert global authorities (2009), http://www.washingtonpost.com/wpdyn/content/article/2009/04/29/AR2009042904911.html
- 5. Chiu, S.S., Lau, Y.L., Chan, K.H., Wong, W.H.S., Peiris, J.S.M.: Influenza-related hospitalizations among children in HongKong. N. Engl. J. Med. 347, 2097–2103 (2002)
- <span id="page-8-0"></span>6. Monto, A.S.: The role of antivirals in the control of influenza. Vaccine 21, 1796–1800 (2003)
- 7. Knox, C., Law, V., Jewison, T., Liu, P., Ly, S., Frolkis, A., Pon, A., Banco, K., Mak, C., Neveu, V., Djoumbou, Y., Eis-ner, R., Guo, A.C., Wishart, D.S.: DrugBank 3.0: a comprehensive resource for 'omics' research on drugs. Nucleic Acids Res. 39(Database issue), D1035–D1041 (2011)
- 8. Berman, H.M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T.N., Weissig, H., Shindyalov, I.N., Bourne, P.E.: The Protein Data Bank. Nucleic Acids Res. 28, 235–242 (2000)
- 9. Jones, G., Willett, P., Glen, R.C.: Molecular recognition of receptor sites using a genetic algorithm with a description of desolvation. J. Mol. Biol. 245, 43–53 (1995)
- 10. Schneidman-Duhovny, D., Inbar, Y., Nussinov, R., Wolfson, H.J.: PatchDock and Symm-Dock: servers for rigid and symmetric docking. Nucl. Acids. Res. 33, W363–W367 (2005)
- 11. Chang, D.T.-H., Oyang, Y.-J., Lin, J.-H.: MEDock: a web server for efficient prediction of ligand binding sites based on a novel optimization algorithm. Nucleic Acids Research 33(suppl. 2), W233–W238
- 12. Schames, J.R., Henchman, R.H., Siegel, J.S., Sotriffer, C.A., Ni, H., McCammon, J.A.: Discovery of a novel binding trench in HIV integrase. J. Med. Chem. 47(8), 1879–1881 (2004)
- 13. Kitchen, D.B., Decornez, H., Furr, J.R., Bajorath, J.: Nat. Rev. Drug. Discov. 3, 935–949 (2004)
- 14. Wang, R., Lu, Y., Wang, S.: J. Med. Chem. 46, 2287–2303 (2003)
- 15. Charifson, P.S., Corkery, J.J., Murcko, M.A., Walters, W.P.: J. Med. Chem. 42, 5100– 5109 (1999)