



A photochemical C=C cleavage process: toward access to backbone *N*-formyl peptides

Haopei Wang and Zachary T. Ball*

Letter

Open Access

Address:
Department of Chemistry, Rice University, Houston TX, USA

Beilstein J. Org. Chem. **2021**, *17*, 2932–2938.
<https://doi.org/10.3762/bjoc.17.202>

Email:
Zachary T. Ball* - zach.ball@gmail.com

Received: 11 October 2021
Accepted: 30 November 2021
Published: 15 December 2021

* Corresponding author

This article is part of the thematic issue "Late-stage diversification of peptides".

Keywords:
formyl peptide; nitroaryl compound; nitroso compound; olefin cleavage; photocleavage

Associate Editor: N. Sewald

© 2021 Wang and Ball; licensee Beilstein-Institut.
License and terms: see end of document.

Abstract

Photo-responsive modifications and photo-uncaging concepts are useful for spatiotemporal control of peptides structure and function. While side chain photo-responsive modifications are relatively common, access to photo-responsive modifications of backbone N–H bonds is quite limited. This letter describes a new photocleavage pathway, affording *N*-formyl amides from vinylogous nitroaryl precursors under physiologically relevant conditions via a formal oxidative C=C cleavage. The *N*-formyl amide products have unique properties and reactivity, but are difficult or impossible to access by traditional synthetic approaches.

Findings

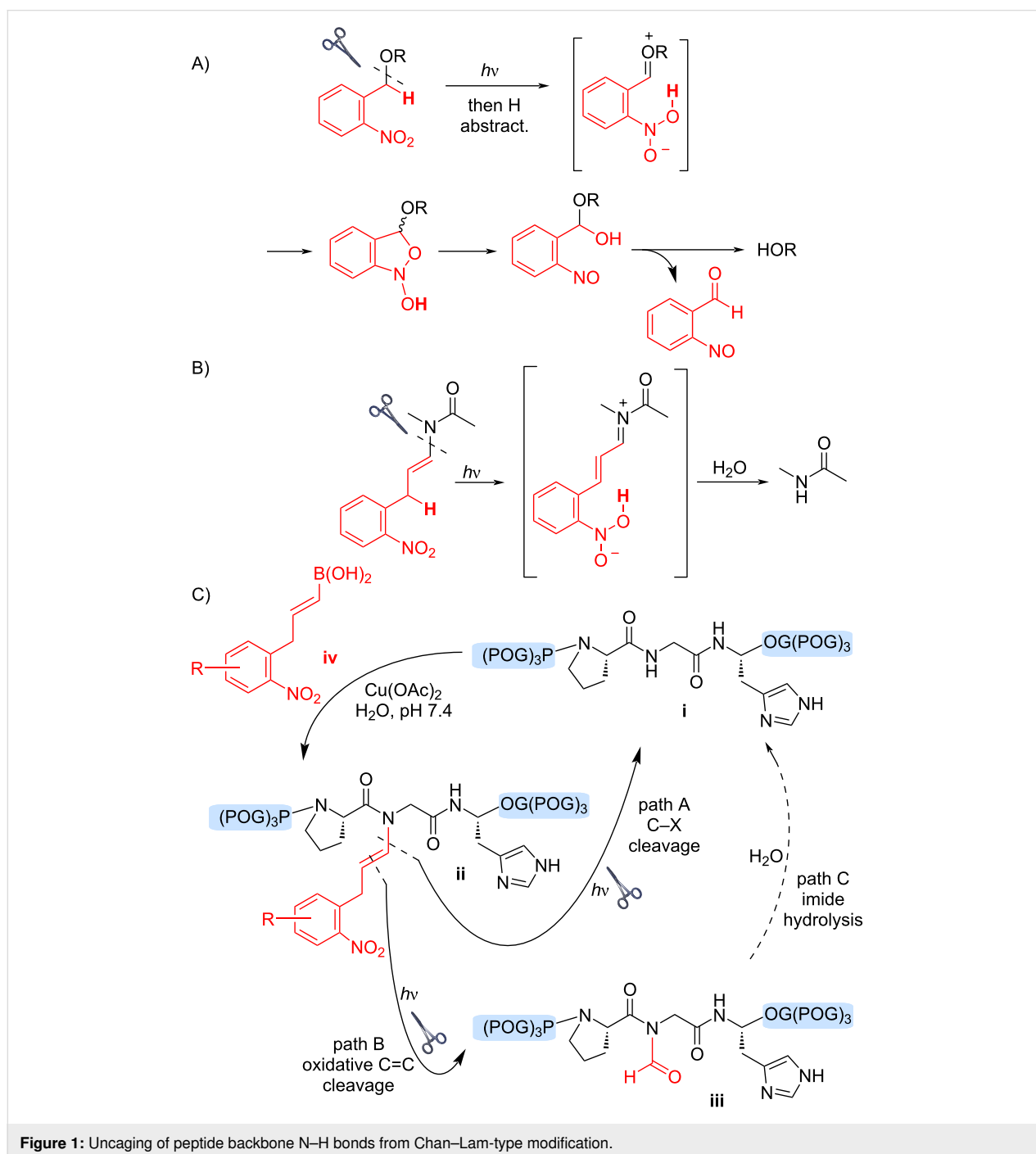
The photochemistry of nitroaromatic functional groups has a rich history that dates back decades [1-5]. Photochemical pathways allow access to diverse and interesting target structures [6-10], though photocleavage of C–X bonds for use as photoremovable protecting groups [11,12] has been the major thrust of the development of 2-nitroaryl compounds. Various 2-nitrobenzyl derivatives are used to photocage heteroatom functional groups, including alcohols, amines, carboxylic acids, and phosphates [11]. Typical photochemical pathways result in cleavage of a benzylic C–X bond following initial benzylic H-atom abstraction [11,13]. In contrast, photorelease systems based on

C–C or C=C bond photocleavage are quite rare [14,15]. We recently reported a vinylogous analogue of this photo-deprotection process, which allowed photocleavage of alkenyl sp² C–X bonds, rather than benzylic sp³ C–X cleavage [16,17]. We now report that further studies into this reaction demonstrate two mechanistically distinct photocleavage pathways, with selectivity dependent on pH. In addition to an anticipated alkenyl sp² C–X bond cleavage pathway, we identified a new photochemical reaction pathway, prevalent under neutral and acidic reaction conditions, which leads to formyl products from formal oxidative cleavage of a C=C bond.

Our interest in vinylogous analogues of 2-nitroaryl photoreactive groups stems from studies into alkenylboronic acid reagents for Chan–Lam-type modification of peptide backbone N–H bonds, directed by a proximal histidine residue (Figure 1C, step, **i** + **iv** → **ii**) [18–20]. Subsequent investigations validated the use of photoreactive boronic acids as an approach to reversible backbone N–H modification via photocleavage of an alkenyl C–N bond [16,17]. Traditional 2-nitroaryl groups allow cleavage of benzylic C–X bonds (e.g. C–O cleavage, Figure 1A)

through H-atom abstraction from a photoexcited intermediate, which produces an oxonium-type intermediate (in brackets). Hydrolysis of this intermediate then affords an alcohol product. Recently [16,17], we demonstrated that vinylogous analogues of this mechanism (Figure 1B) provide entry into similar photocleaving chemistries for amide release.

Figure 1C shows an example of this concept applied to a peptide substrate. Reaction of the peptide **i** with an alkenyl-



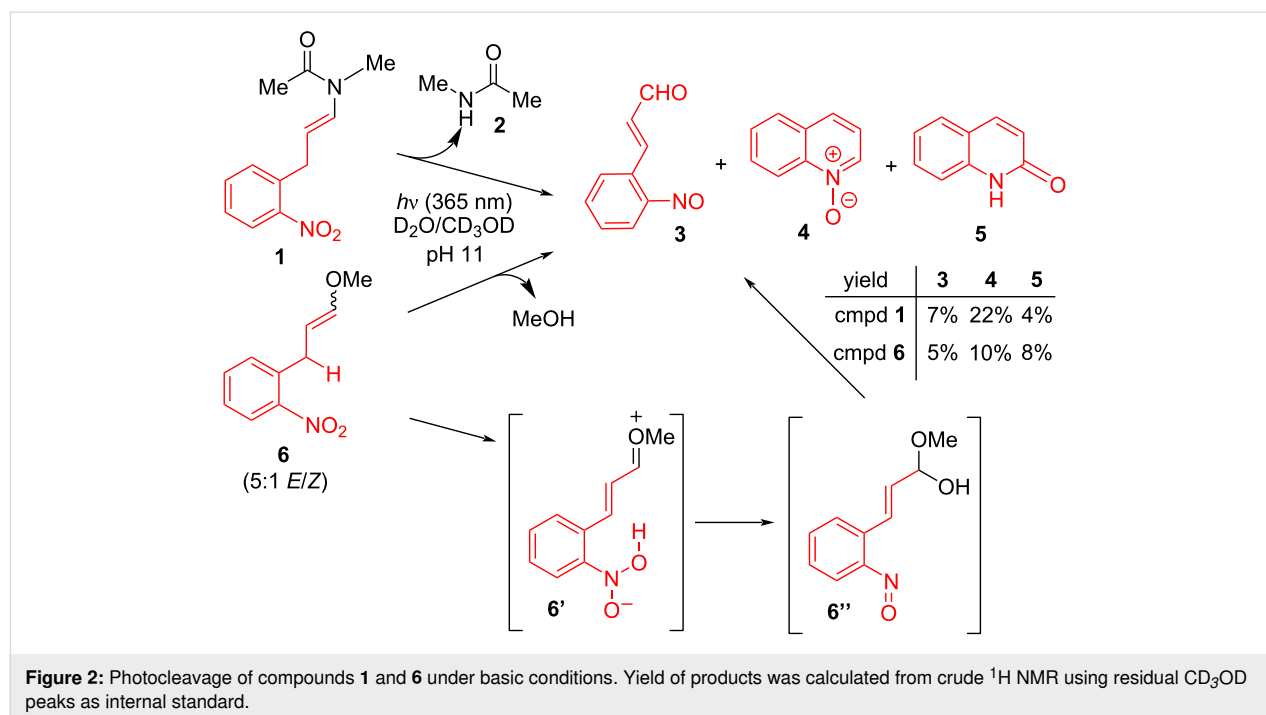
boronic acid reagent **iv** in the presence of a copper(II) salt in water provides access to the backbone N–H alkenylation product **ii**, directed by a neighboring histidine residue. Upon exposure to 365-nm light, photocleavage of the caging group (red) was observed, producing the free peptide **i**. Irradiation longer than 10 minutes were sometimes necessary for maximal yield of photo-deprotected product **i** [16]. Furthermore, byproducts or transiently stable intermediates were sometimes indicated by HPLC and/or NMR of these photocleavage reactions [16,17]. These observations prompted a more detailed study of the components present during photocleavage reactions of small-molecule models, leading to the identification of the *N*-formyl product **iii**, a possible intermediate on the path to product **i** via imide hydrolysis.

To better understand the mechanism of photocleavage and the appearance of the formyl product **iii**, we first identified the 2-nitroaryl-derived byproducts produced in this reaction. Model compound **1** was subjected to aqueous photocleavage in the presence of triethylamine, and the resulting reaction mixture was purified by reversed-phase HPLC (Figure 2). We isolated a nitroso product **3**, in addition to two other major identifiable components of the crude reaction: quinoline *N*-oxide (**4**) and quinolinone (**5**). The compounds **4** and **5** are C₉ compounds possibly derived from thermal or photochemical rearrangement of compound **3** or another intermediate. The yield of each product was calculated by NMR and verified by isolation (Figure 2). To test the generality of this process with other functional groups, we prepared and tested alkenyl ether **6** as a model of

C–O-bond cleavage. Photoirradiation of the ether **6** similarly provided a mixture of C₉-containing products **3**, **4**, and **5**. Under these reaction conditions, the C–X cleavage products (MeOH or **2**) were observed, but no formyl products were formed. The C₉ byproducts – the nitroso **3**, and related compounds **4** and **5** are all consistent with the classical C–X cleavage mechanism and with hydrolysis of the presumed oxonium intermediate **6'**, but are inconsistent with the production of formyl products.

In contrast, photoirradiation of the same alkenyl ether **6** under acidic conditions at pH 4.0 provided a mixture of methanol (53%) and methyl formate (38%, **7**) as determined by NMR, the latter product is the result of formal oxidative C=C cleavage (Figure 3a). Alkenyl amide **1** at pH 4.0 similarly gave mixtures of the C–N cleavage product **2** and C=C cleavage product **8**. We examined product selectivity in the irradiation of alkenyl amide **1** across a range of pH and found a significant correlation (Figure 3b–d). The formyl product **8** predominated at acidic and neutral pH. The amount of **8** decreased with increasing pH, and above pH 10 the C–X cleavage product **2** became the major product. Unfortunately, no products other than the formyl compound were isolated after photocleavage of compound **1** or **6** in acidic conditions. Instead, when irradiation of alkenyl amide **1** was conducted in acetone, crude NMR analysis indicated the appearance of product **8** as well as new peaks in the aromatic region.

Following acetylation of the reaction mixture, we were able to isolate small quantities of *O*-acetyl *N*-hydroxyindole (**9**,



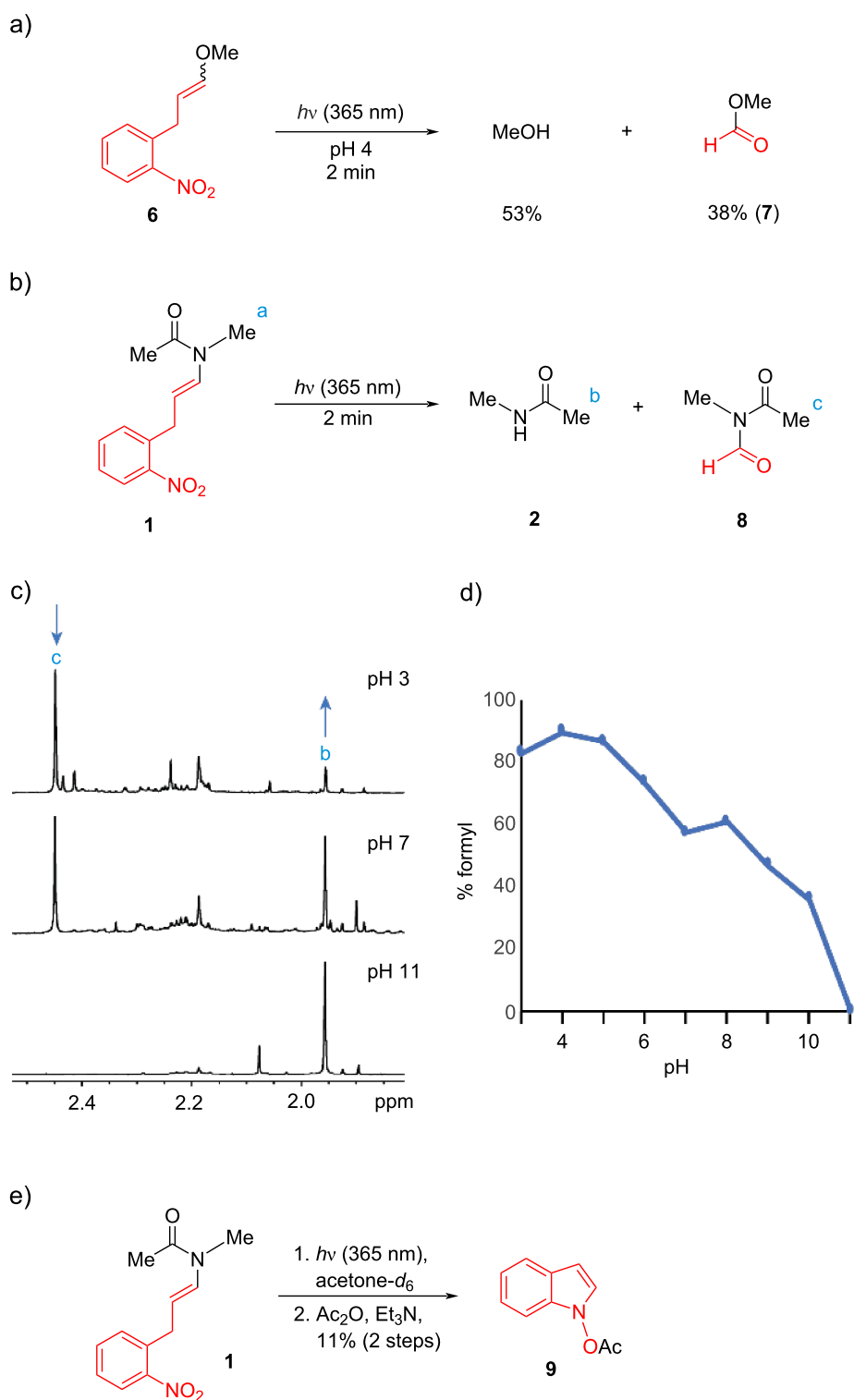


Figure 3: (a) Photocleavage of compound **6** under acidic conditions. Yields determined by ^1H NMR using residual CD_3OD as an internal standard. (b–d) Selectivity of photocleavage of alkenyl amide **1** as a function of pH. Product percentage of *N*-formyl **8** was assessed by crude NMR (c) and graphed (d). Formation of *N*-formyl-*N*-methyl acetamide **8** during photocleavage of compound **1**. Conditions: **1** (1.8 μmol) was dissolved in $\text{MeOD-}d_4$ (200 μL) and deuterated buffer (400 μL). The solution was irradiated at 365 nm for 2 min. (e) Photocleavage reaction of **1** in acetone.

Figure 3e), although the initial byproduct *N*-hydroxyindole itself proved too unstable to be isolated. It is noteworthy within this context that hydroxyindole is a C₈ compound, consistent with transfer of the C₁ formyl group to compound **8**. The formation of amide **2** at elevated pH could, in theory, derive from hydrolysis of the initially formed formyl product **8** (i.e. Figure 1C). However, the appearance of primarily C₉ byproducts in the formation of amide **2** at elevated pH precludes pathways involving the intermediacy of **8**. To provide additional support for this analysis, and to assess the stability of *N*-formyl amides formed in this reaction, we irradiated alkenyl amide **10**, which contains a 2-phenylethyl substituent that allowed easier isolation of *N*-formyl **11** (Figure 4). After irradiation, the product **11** was isolated in 28% yield, the modest yield reflecting the instability in water and on silica of this compound. The purified *N*-formyl **11** was then dissolved in buffer (pH 8), and its hydrolysis to amide **12** was assessed (Figure 4, inset). We observed clean first-order kinetics to give amide **12** with a half-life ($t_{1/2}$) of 6.4 h.

The observation of *N*-formyl products can be rationalized with a bifurcating mechanism (Figure 5). Following photoactivation, H-atom abstraction and nucleophilic addition of water would produce the key intermediate **B**. Such hemi-aminal compounds would be unstable under basic conditions, readily forming aldehyde products **3**. However, related hemi-aminal compounds are quite stable under non-basic conditions, and the motif is even

contained in some natural products, such as zampanolide [21] and spergualin [22]. We propose a competing electrocyclization pathway, affording the heterocycle **D**, a pathway which should not be base-catalyzed, and thus may be reasonably predominant under appropriate conditions. From heterocycle **D**, a C–C cleavage would produce the *N*-formyl product **8** and a re-aromatized C₈ heterocyclic byproduct **E**. Rearrangement to hydroxyindole (**F**) would then account for the isolation of the acetylated analogue **9**.

The photochemical pathway described here represents a formal oxidative olefin cleavage of vinylogous nitroaryl-modified amides and ethers. The pathway adds to the diversity of photochemical pathways known for 2-nitrophenyl systems, and the concept described here might be useful for the synthetic unmasking of relatively sensitive imido structures. For chemical biology applications, the results point to a far more diverse photochemistry than previously assumed for vinylogous photocleavage systems. Although formyl hydrolysis to the “expected” amide products can and does occur under physiological conditions, the rates of this hydrolysis are slow for the simple models in this study. Within more complex peptides or proteins, selectivity in photocleavage pathways may differ significantly, depending on local chemical environment. It is also worth noting that *N*-formyl products are themselves acylating reagents, and thus could find use in photochemical generation of selective acyl donors.

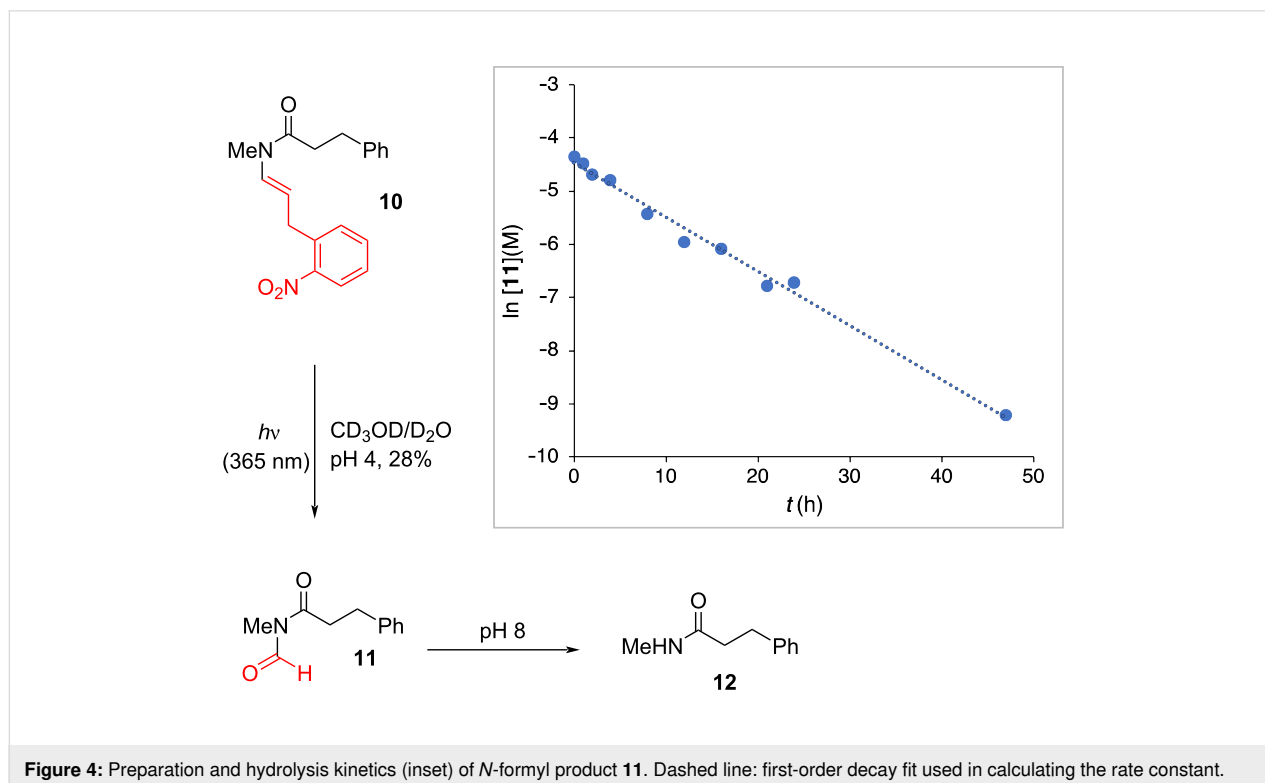
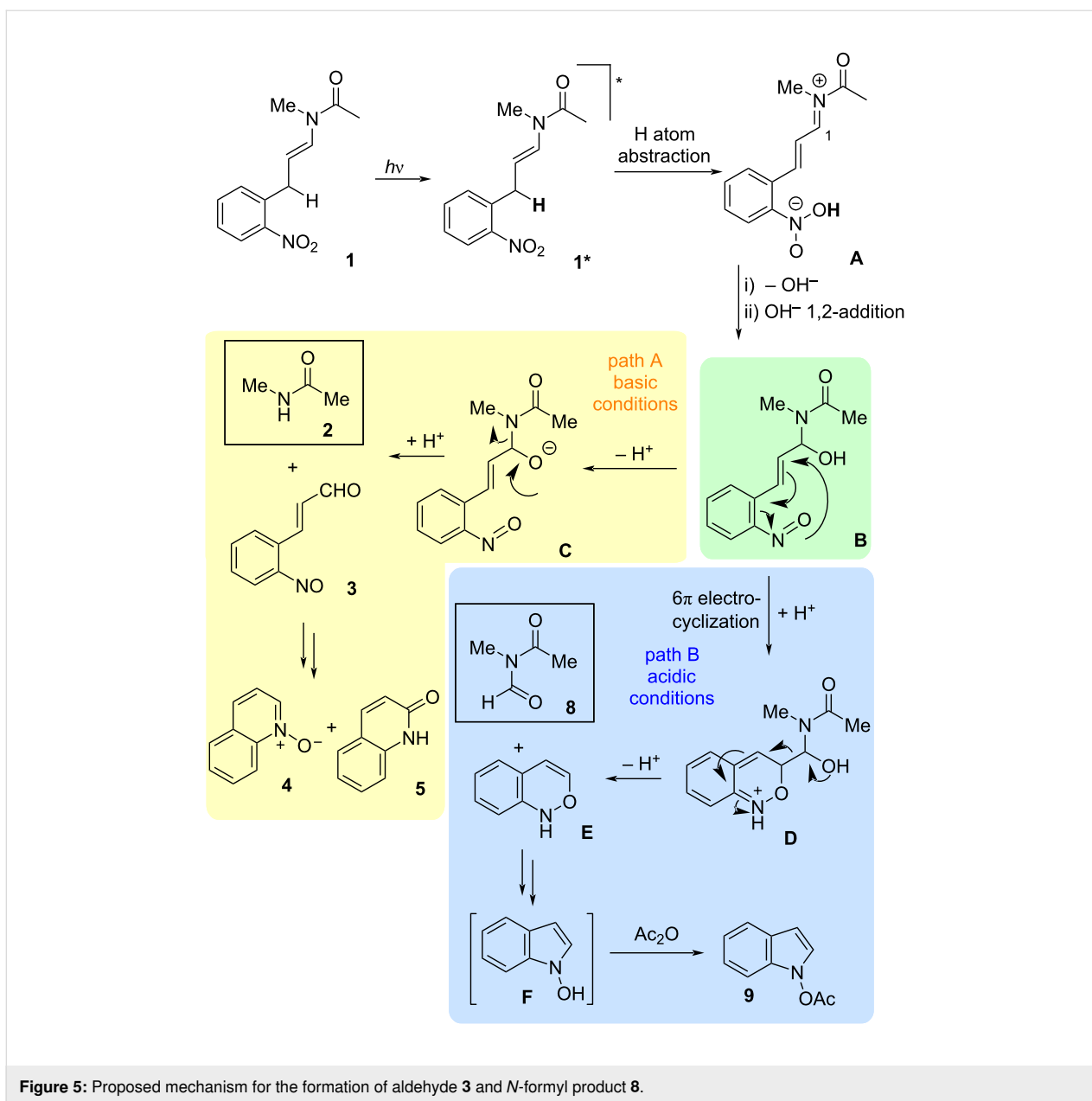


Figure 4: Preparation and hydrolysis kinetics (inset) of *N*-formyl product **11**. Dashed line: first-order decay fit used in calculating the rate constant.



Supporting Information

Supporting Information File 1

Experimental section and additional information.

[<https://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-17-202-S1.pdf>]

Funding

We acknowledge support from the Robert A. Welch Foundation Research Grant C-1680 (Z.T.B.), and the National Science Foundation under grant number CHE-1904865.

ORCID® iDs

Haopei Wang - <https://orcid.org/0000-0001-9756-8783>

Zachary T. Ball - <https://orcid.org/0000-0002-8681-0789>

References

- Jones, L. B.; Kudrna, J. C.; Foster, J. P. *Tetrahedron Lett.* **1969**, *10*, 3263–3265. doi:10.1016/s0040-4039(01)88404-5
- Bartrop, J. A.; Bunce, N. J. *J. Chem. Soc. C* **1968**, 1467–1474. doi:10.1039/j39680001467
- Finnegan, R. A.; Knutson, D. *J. Am. Chem. Soc.* **1968**, *90*, 1670–1671. doi:10.1021/ja01008a061
- Hamanoue, K.; Amano, M.; Kimoto, M.; Kajiwara, Y.; Nakayama, T.; Teranishi, H. *J. Am. Chem. Soc.* **1984**, *106*, 5993–5997. doi:10.1021/ja00332a041

5. Chapman, O. L.; Heckert, D. C.; Reasoner, J. W.; Thackaberry, S. P. *J. Am. Chem. Soc.* **1966**, *88*, 5550–5554. doi:10.1021/ja00975a037
6. Belligund, K.; Mathew, T.; Hunt, J. R.; Nirmalchandar, A.; Haiges, R.; Dawlaty, J.; Prakash, G. K. S. *J. Am. Chem. Soc.* **2019**, *141*, 15921–15931. doi:10.1021/jacs.9b07241
7. Nie, H.-J.; Guo, A.-D.; Lin, H.-X.; Chen, X.-H. *RSC Adv.* **2019**, *9*, 13249–13253. doi:10.1039/c9ra02466b
8. Zhu, J. S.; Kraemer, N.; Li, C. J.; Haddadin, M. J.; Kurth, M. J. *J. Org. Chem.* **2018**, *83*, 15493–15498. doi:10.1021/acs.joc.8b02356
9. Guo, A.-D.; Wei, D.; Nie, H.-J.; Hu, H.; Peng, C.; Li, S.-T.; Yan, K.-N.; Zhou, B.-S.; Feng, L.; Fang, C.; Tan, M.; Huang, R.; Chen, X.-H. *Nat. Commun.* **2020**, *11*, 5472. doi:10.1038/s41467-020-19274-y
10. Charlton, J. L.; Liao, C. C.; De Mayo, P. *J. Am. Chem. Soc.* **1971**, *93*, 2463–2471. doi:10.1021/ja00739a018
11. Klán, P.; Šolomek, T.; Bochet, C. G.; Blanc, A.; Givens, R.; Rubina, M.; Popik, V.; Kostikov, A.; Wirz, J. *Chem. Rev.* **2013**, *113*, 119–191. doi:10.1021/cr300177k
12. Pelliccioli, A. P.; Wirz, J. *Photochem. Photobiol. Sci.* **2002**, *1*, 441–458. doi:10.1039/b200777k
13. Gaplovsky, M.; Il'ichev, Y. V.; Kamzhilov, Y.; Kombarova, S. V.; Mac, M.; Schwörer, M. A.; Wirz, J. *Photochem. Photobiol. Sci.* **2005**, *4*, 33–42. doi:10.1039/b409927c
14. Ichinose, N.; Mizuno, K.; Otsuji, Y.; Caldwell, R. A.; Helms, A. M. *J. Org. Chem.* **1998**, *63*, 3176–3184. doi:10.1021/jo971111s
15. Sako, M.; Shimada, K.; Hirota, K.; Maki, Y. *Tetrahedron Lett.* **1986**, *27*, 3877–3880. doi:10.1016/s0040-4039(00)83904-0
16. Mangubat-Medina, A. E.; Martin, S. C.; Hanaya, K.; Ball, Z. T. *J. Am. Chem. Soc.* **2018**, *140*, 8401–8404. doi:10.1021/jacs.8b04893
17. Mangubat-Medina, A. E.; Trial, H. O.; Vargas, R. D.; Setegne, M. T.; Bader, T.; Distefano, M. D.; Ball, Z. T. *Org. Biomol. Chem.* **2020**, *18*, 5110–5114. doi:10.1039/d0ob00923g
18. Ohata, J.; Minus, M. B.; Abernathy, M. E.; Ball, Z. T. *J. Am. Chem. Soc.* **2016**, *138*, 7472–7475. doi:10.1021/jacs.6b03390
19. Ohata, J.; Zeng, Y.; Segatori, L.; Ball, Z. T. *Angew. Chem., Int. Ed.* **2018**, *57*, 4015–4019. doi:10.1002/anie.201800828
20. Hanaya, K.; Miller, M. K.; Ball, Z. T. *Org. Lett.* **2019**, *21*, 2445–2448. doi:10.1021/acs.orglett.9b00759
21. Troast, D. M.; Porco, J. A. *Org. Lett.* **2002**, *4*, 991–994. doi:10.1021/ol025558l
22. Kondo, S.; Iwasawa, H.; Ikeda, D.; Umeda, Y.; Ikeda, Y.; Iinuma, H.; Umezawa, H. *J. Antibiot.* **1981**, *34*, 1625–1627. doi:10.7164/antibiotics.34.1625

License and Terms

This is an open access article licensed under the terms of the Beilstein-Institut Open Access License Agreement (<https://www.beilstein-journals.org/bjoc/terms>), which is identical to the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0>). The reuse of material under this license requires that the author(s), source and license are credited. Third-party material in this article could be subject to other licenses (typically indicated in the credit line), and in this case, users are required to obtain permission from the license holder to reuse the material.

The definitive version of this article is the electronic one which can be found at:
<https://doi.org/10.3762/bjoc.17.202>