ORIGINAL RESEARCH

Clinical significance and prospective molecular mechanism of MALAT1 in pancreatic cancer exploration: a comprehensive study based on the GeneChip, GEO, Oncomine, and TCGA databases

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Purpose: Long noncoding RNAs (lncRNAs) are known to function as regulators in the development and occurrence of various tumors. MALAT1 is a highly conserved lncRNA and has vital functions in diverse tumors, including pancreatic cancer (PC). However, the underlying molecular regulatory mechanism involved in the occurrence and development of PC remains largely unknown. Thus, it is important to explore MALAT1 in PC and elucidate its function, which might offer a new perspective for clinical diagnosis and therapy.

Methods: First, we used the Gene Expression Omnibus, Oncomine, and The Cancer Genome Atlas databases to determine the clinical diagnostic and prognostic values of MALAT1. We next used our own GeneChip and The Cancer Genome Atlas database to collect the possible target genes of MALAT1 and further utilized a bioinformatics analysis to explore the underlying significant pathways that might be crucial in PC. Finally, we identified several key target genes of MALAT1 and hope to offer references for future research.

Results: We found that the expression of MALAT1 was significantly elevated in patients with PC. A receiver operating characteristics curve analysis showed a moderate diagnostic value (area under the curve =0.75, sensitivity =0.66, specificity =0.72). A total of 224 important overlapping genes were collected, and six hub genes (*CCND1*, *MAPK8*, *VEGFA*, *FOS*, *CDH1*, and *HSP90AA1*) were identified, of which *CCND1*, *MAPK8*, and *VEGFA*, are important genes in PC. Several pathways, including the mTOR signaling pathway, pathways in cancer, and the MAPK signaling pathway, were suggested to be the vital MALAT1 pathways in PC.

Conclusion: MALAT1 is suggested to be a promising diagnostic biomarker in PC. Six hub genes (*CCND1*, *MAPK8*, *VEGFA*, *FOS*, *CDH1*, and *HSP90AA1*), and specifically *CCND1*, *MAPK8*, and *VEGFA*, might be key MALAT1 target genes in PC. Due to their possible clinical significance in PC, several pathways, such as the mTOR signaling pathway, pathways in cancer, and the MAPK signaling pathway, are worthy of further study.

Keywords: MALAT1, pancreatic cancer, bioinformatics, target gene

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Introduction

Pancreatic cancer (PC) is the seventh most common cause of cancer mortality and causes 330,000 deaths per year worldwide, accounting for approximately 4.0% of all cases of cancer.[1] Currently, while the incidence of PC tends to linearly increase, the mortality of PC is high, and the prognosis is dismal. It is estimated that patients who present with early disease and have a negative resection margin only have a 5-year survival rate of 24%.[2] There is an even higher mortality for unresectable patients.

Currently, PC lacks a sensitive diagnosis marker to monitor the disease at an early stage and to predict the prognosis. Therefore, PC is in desperate need of a new index.

Long noncoding RNAs (lncRNAs) do not encode proteins and were once regarded as transcriptional noise or cloning artifacts.[3,4] According to recent research, lncRNAs are thought to be regulators of cancer development, which make them a promising target for cancer treatment.[5,6] MALAT1, with a length of over 8,000 nucleotides, is a highly conserved lncRNA that is derived from chromosome 11q13. It controls cancer cell proliferation, differentiation, apoptosis, and invasion by regulating the synthesis of proteins and the expression of genes. MALAT1 is well known as the first lncRNA, indicating the poor prognosis of non-small-cell lung cancer.[7] Currently, emerging research has shown that MALAT1 plays a crucial role in ovarian cancer,[8] esophageal cancer,[9] prostate cancer[10] breast cancer,[11] thyroid cancer,[12] nasopharyngeal carcinoma,[13] and other types of malignant diseases. Previously, studies have found that elevated MALAT1 levels are involved in PC proliferation and metastasis through the stimulation of autophagy.[14] Nevertheless, the potential molecular mechanism is still unclear and needs additional research. Over the last few years, there have been numerous novel techniques related to cancer mechanism and therapy exploration emerging. For instance, molecular docking and molecular dynamics simulation, which helps identify the cancer-associated single nucleotide polymorphisms and their possible molecular mechanism.[15–17] And other analysis including protein–ligand interaction analysis, principal component analysis, shape analysis of binding pocket, and kinase inhibitor screening are also used to explore the therapeutic molecules for cancers.[18,19] Researchers are also trying to figure out a methodology to optimize a synergistic and clinically achievable combination of multiple agents for cancer clinical therapy.[20] Despite their particular perspectives in uncovering the cancer mechanism and therapy, the complicated and demanding operation of the experimental techniques themselves might limit their further development. As a result, we hope to use computational methods to explore the cancer mechanism by virtue of online databases. Thus, in this paper, we performed a comprehensive study to further explore the clinical significance and underlying molecular mechanism of MALAT1 in PC on a computational level.

To explore the relationship between MALAT1 and PC, we collected gene expression data and clinical data from patients using The Cancer Genome Atlas (TCGA) and Gene Expression Omnibus (GEO). The relationship between MALAT1 and PC was further analyzed based on the TCGA and GEO data.

For the purpose of exploring the molecular mechanism, we analyzed the overlapping genes in the differentially expressed genes after MALAT1 knockout and the coexpressed genes from TCGA. We performed a bioinformatics functional analysis for the overlapping genes and used Gene Ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEGG), and Protein–Protein Interaction (PPI) databases.

Taken together, we hope to understand the clinical value of MALAT1 in PC and the molecular mechanism of MALAT1. We hope that the data shown here may offer new perspectives in future research and clinical application.

Materials and methods
Selection of microarrays in GEO datasets and profiles
For the purpose of understanding the diagnostic value of MALAT1 in PC, we systematically retrieved the GEO microarrays performed before December 2016 for a meta-analysis. The retrieval strategies were as follows: Pancrea* AND (adenocarcinoma OR carcinoma OR cancer OR neoplasm OR tumor OR tumour OR neoplas* OR malignan*). The inclusion criteria were as follows: 1) the enrolled subjects included patients with PC and normal control samples, 2) MALAT1 was expressed, detected, and available for extraction in the enrolled subjects, and 3) there were more than three subjects.

Validation of MALAT1 expression in PC based on the Oncomine database
The Oncomine database (http://www.oncomine.org) incorporates 264 independent datasets that include 35 cancer types and supports various methods of analysis, including molecular concepts analysis, interactome analysis, and meta-analysis.[21] Thus, we adopted the Oncomine database to further validate the expression of MALAT1 in PC. The differential expression analysis was directly performed using Oncomine online analysis tools.

Identification of the clinical prognostic value of MALAT1 in PC
TCGA (http://cancergenome.nih.gov) data portal is one of the largest available public resources offering genomic, transcriptomic, methylomic, and copy number variation datasets for more than 20 cancer types.[22,23] We evaluated the TCGA survival of patients with PC in two online analysis websites, cBioPortal for Cancer Genomics[24,25] (http://www.cbioportal.org/) and OncoLnc[26] (http://www.oncolnc.org/), which link to the TCGA database.

Identification of key differentially expressed genes associated with MALAT1 in PC

For the purpose of investigating the genes influenced by MALAT1, we performed GeneChip analysis after MALAT1 was knocked down. The human PC Miapaca-2 cell line was purchased from the Institute of Biochemistry and Cell Biology of the Chinese Academy of Sciences (Wuhan, People's Republic of China), and was maintained routinely in Dulbecco's Modified Eagle's Medium (Thermo Fisher Scientific, Waltham, MA, USA). The four siRNAs for MALAT1 were designed on the internet (http://design.RNAi.jp/) and synthesized by Shanghai Jima company. Lentivirus MALAT1-shRNA was synthesized using lentivirus, packaged, and screened. The knockdown rate of MALAT1-stable cells was 76% in Miapaca-2 cell line with the best siRNA sequence (F:GGCAGCTTTAACAGATAACA; R:CCGTCCGACAAGGGTCATTCA).The three groups of control vector cells and MALAT1-shRNA stable cells were tested with GeneChip prime view human kit (Affymetrix., 901838, Santa Clara, CA, USA). Using MALAT1 knockdowns, we first acquired the differentially expressed genes. After performing GO analysis, we obtained the enriched GO data and the corresponding genes. Furthermore, we collected MALAT1 and mRNA gene coexpression data from the TCGA database. We adopted 0.15 as the cut-off value for screening the significant coexpression genes. Aiming at determining the most likely key differentially expressed MALAT1-associated genes in PC, we further correlated the enriched GO items with correlative genes and TCGA coexpression genes. The overlapping genes we obtained were used for further analysis.

Bioinformatics analysis of the overlapping genes

For the overlapping genes, The Database for Annotation, Visualization, and Integrated Discovery version 6.7 (https://david-d.ncifcrf.gov/), an online bioinformatics functional enrichment resource for a list of genes analysis,[27,28] was used to perform GO enrichment and KEGG pathway analysis. The analysis was achieved via uploading the overlapping genes though the Functional Annotation porch. We chose those items whose P-values were below 0.05 for further study. Further, we used BiNGO, a plugin of Cytoscape 3.40,[29] to visualize the GO enrichment pathway, which includes biological process (BP), cellular component (CC), and molecular function (MF). Meanwhile, we also aimed to identify the most crucial target genes of MALAT1 from the overlapping genes. The STRING database version 10 (http://string-db.org/) is aimed at providing a critical assessment and integration of PPI through direct (physical) and indirect (functional) associations network construction.[30] Thus, STRING database was further used to perform a protein–protein interaction network analysis. The PPI network construction was achieved through StringApp, a plugin of Cytoscape 3.40, which links to STRING database. Interaction score (0.7) was adopted as a high confidence cut-off value to determine the PPI. Hub genes, which are the key genes in PC, were obtained from the network.

Statistical analysis

Stata 12.0 (Stata Corp., College Station, TX, USA) was used to perform a GEO microarray meta-analysis in this study. Receiver operating characteristic (ROC) curve analyses were utilized to evaluate the diagnostic value of MALAT1 in PC and normal tissue. Kaplan–Meier curves and the log-rank test were applied to analyze the effect of MALAT1 on the survival outcome of patients with PC. Only $P<0.05$ was considered statistically significant.

Results

Characteristics of the included datasets

A total of 27 eligible datasets published from 2008 to 2016 in the GEO database were used in this study. The characteristics of the selected datasets are shown in Table 1. In total, 764 PC samples and 469 healthy controls were used.

Clinical diagnostic value of MALAT1 as a biomarker in PC based on the GEO datasets

After pooling the data from the 27 eligible datasets, we found that there was significant heterogeneity among the datasets (I^2=61.1%, P=0.000, Figure 1). As a result, we performed a sensitivity analysis to screen the possible datasets that might cause the heterogeneity. As can be seen in Figure 2, two datasets, namely, GSE15471 and GSE62165, may contribute to significant heterogeneity. Thereby, we removed the two datasets and recalculated the pooled standard mean difference. As shown in Figure 3, the heterogeneity appears to have disappeared (I^2=44.9%, P=0.009). The pooled standard mean difference was 0.23 (95% confidence interval [CI]: 0.10–0.37, P=0.001), which indicates a significant difference in the effect of MALAT1 between patients with PC and healthy control patients. The expression of MALAT1 was significantly upregulated. Publication bias among the 25 datasets was assessed through Begg's funnel plot. As shown in Figure 4,

Table 1 Characteristics of the GEO datasets included in the meta-analysis

Series	Country	Year	Platform	Sample source	PC patients	Normal tissues
GSE14245	USA	2008	GPL570	Saliva	12	12
GSE11838	USA	2008	GPL6977	Tissue	28	79
GSE15471	Romania	2009	GPL570	Tissue	39	39
GSE16515	USA	2009	GPL570	Tissue	36	16
GSE22780	USA	2011	GPL570	Tissue	8	8
GSE32676	USA	2011	GPL570	Tissue	25	7
GSE18670	Belgium	2012	GPL570	Tissue/peripheral blood	6	6
GSE15932	People's Republic of China	2012	GPL570	Peripheral blood	16	8
GSE28735	USA	2012	GPL6244	Tissue	45	45
GSE23397	Germany	2013	GPL5188	Tissue	15	6
GSE41368	Italy	2013	GPL6244	Tissue	6	6
GSE43795	South Korea	2013	GPL10558	Tissue	26	5
GSE49641	Spain	2013	GPL6244	Peripheral blood	18	18
GSE43288	UK	2013	GPL96/GPL97	Tissue	4	3
GSE27890	USA	2014	GPL570	Tissue	4	4
GSE56560	UK	2014	GPL5175	Tissue	28	7
GSE58561	Norway	2014	GPL14550	Tissue	3	2
GSE55643	UK	2014	GPL6480	Tissue	45	8
GSE60979	Norway	2015	GPL14550	Tissue	49	12
GSE71008	USA	2015	GPL9052	Plasma	6	50
GSE74629	Spain	2015	GPL10558	Peripheral blood	36	14
GSE71989	USA	2015	GPL570	Tissue	14	8
GSE62165	Belgium	2016	GPL13667	Tissue	118	13
GSE62452	USA	2016	GPL6244	Tissue	69	61
GSE86436	People's Republic of China	2016	GPL13825	Tissue	6	6
GSE77858	USA	2016	GPL7264	Tissue	77	3
GSE91035	USA	2016	GPL22763	Tissue	25	23

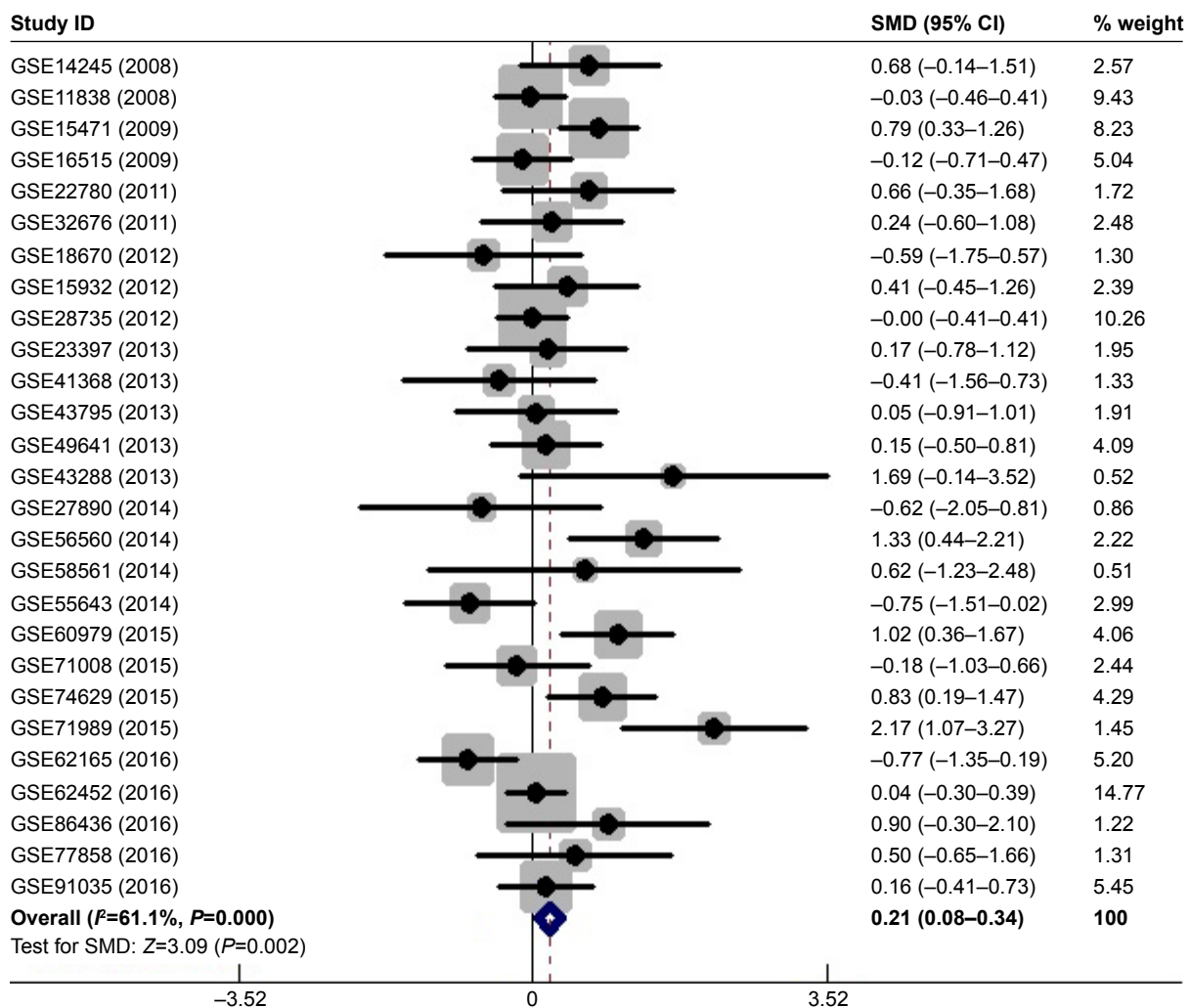**Abbreviations:** GEO, Gene Expression Omnibus; PC, pancreatic cancer.

the funnel plot was basically considered symmetrical. Taking $P>0.05$ as cut-off value, we found that $P=0.414$ was obtained by Begg's test, which indicated that no significant publication bias existed. Furthermore, a summary ROC curve revealed that the area under the curve was 0.75 (95% CI: 0.71–0.78), and the overall sensitivity and specificity was 0.66 (95% CI: 0.57–0.74) and 0.72 (95% CI: 0.61–0.81) (Figures 5 and 6A). The pooled diagnostic odds ratio was also calculated as 4.41 (95% CI: 2.71–7.17), which indicates that upregulated MALAT1 results in a higher risk of PC (Figure 6B).

MALAT1 expression difference validation based on the Oncomine database

Five PC datasets in the Oncomine database were adopted for the validation of MALAT1 expression (Grutzmann's dataset [http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-950]; Badea's datasets [http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE15471]; Iacobuzio-Donahue's dataset [http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE3654; http://genome-

www.stanford.edu/pancreatic1/index.shtml]; Pie's dataset [http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE16515]; Ishikawa's dataset [http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE1542]). In Grutzmann's and Badea's datasets, the expression of MALAT1 was significantly elevated in PC, which was in accordance with the results of the meta-analysis (Figure 7A and B). The P-values for these datasets were 0.025 and <0.01, respectively. However, the remaining three datasets did not show a statistically significant difference in MALAT1 expression (Figure 7C–E).

Prognostic value of MALAT1 based on the TCGA database

For the analysis using the cBioPortal for Cancer Genomics, the survival data for a total of 178 PC patients was analyzed. We found 10 of 178 patients had alterations in the expression of MALAT1 (Supplementary material 1). Thus, we evaluated the survival of the cases with MALAT1 alteration compared to those without MALAT1 alteration. The log-rank test showed that the P-value was 0.0380 (Figure 8A).

Study ID		SMD (95% CI)	% weight
GSE14245 (2008)		0.68 (−0.14–1.51)	2.57
GSE11838 (2008)		−0.03 (−0.46–0.41)	9.43
GSE15471 (2009)		0.79 (0.33–1.26)	8.23
GSE16515 (2009)		−0.12 (−0.71–0.47)	5.04
GSE22780 (2011)		0.66 (−0.35–1.68)	1.72
GSE32676 (2011)		0.24 (−0.60–1.08)	2.48
GSE18670 (2012)		−0.59 (−1.75–0.57)	1.30
GSE15932 (2012)		0.41 (−0.45–1.26)	2.39
GSE28735 (2012)		−0.00 (−0.41–0.41)	10.26
GSE23397 (2013)		0.17 (−0.78–1.12)	1.95
GSE41368 (2013)		−0.41 (−1.56–0.73)	1.33
GSE43795 (2013)		0.05 (−0.91–1.01)	1.91
GSE49641 (2013)		0.15 (−0.50–0.81)	4.09
GSE43288 (2013)		1.69 (−0.14–3.52)	0.52
GSE27890 (2014)		−0.62 (−2.05–0.81)	0.86
GSE56560 (2014)		1.33 (0.44–2.21)	2.22
GSE58561 (2014)		0.62 (−1.23–2.48)	0.51
GSE55643 (2014)		−0.75 (−1.51–0.02)	2.99
GSE60979 (2015)		1.02 (0.36–1.67)	4.06
GSE71008 (2015)		−0.18 (−1.03–0.66)	2.44
GSE74629 (2015)		0.83 (0.19–1.47)	4.29
GSE71989 (2015)		2.17 (1.07–3.27)	1.45
GSE62165 (2016)		−0.77 (−1.35–0.19)	5.20
GSE62452 (2016)		0.04 (−0.30–0.39)	14.77
GSE86436 (2016)		0.90 (−0.30–2.10)	1.22
GSE77858 (2016)		0.50 (−0.65–1.66)	1.31
GSE91035 (2016)		0.16 (−0.41–0.73)	5.45
Overall (I^2=61.1%, *P*=0.000)		**0.21 (0.08–0.34)**	**100**
Test for SMD: *Z*=3.09 (*P*=0.002)			

Figure 1 Forest plot of 27 GEO datasets evaluating the SMD of MALAT1 in the PC group and the healthy control group (a fixed-effect model).
Abbreviations: CI, confidence interval; GEO, Gene Expression Omnibus; PC, pancreatic cancer; SMD, standard mean difference.

As reported on the Oncolnc website, we analyzed the survival of patients with high MALAT1 expression and compared it to that of patients with low MALAT1 expression in a total of 174 patients with PC. It showed a trend that high MALAT1 expression indicates better survival, although no statistical significance was observed (*P*=0.304, Figure 8B).

Collection of the overlapping genes

After knocking down MALAT1, we performed a GO enrichment analysis and collected 656 enriched GO gene symbols. Furthermore, using 0.15 as *P*-value cut-off, we obtained 5,887 coexpressed genes from the TCGA database. We then intersected the two part genes and gathered 224 overlapping genes (Supplementary material 2), which might become the crucial targets genes of MALAT1 in PC.

Bioinformatics analysis of the overlapping genes

As displayed in Table 2, the overlapping genes contain three enriched GO pathway categories: BP, CC, and MF. In the BP category, the enriched items were mainly intracellular transport, regulation of phosphorus metabolic process, regulation of phosphate process, etc. In the CC category, membrane-enclosed lumen, nuclear lumen, and nuclear chromosome, among others, were the main components of the enrichment. In regards to the MF category, protein dimerization activity, nucleoside binding, and purine nucleoside binding ranked as the top three enriched items. All of the enriched GO pathways, including the BP, CC, and MF categories, were further visualized through Cytoscape 3.40 and the results are displayed in Supplementary materials 3 and 4 and Figure 9, respectively. In the KEGG pathway

Meta-analysis estimates, given named study is omitted
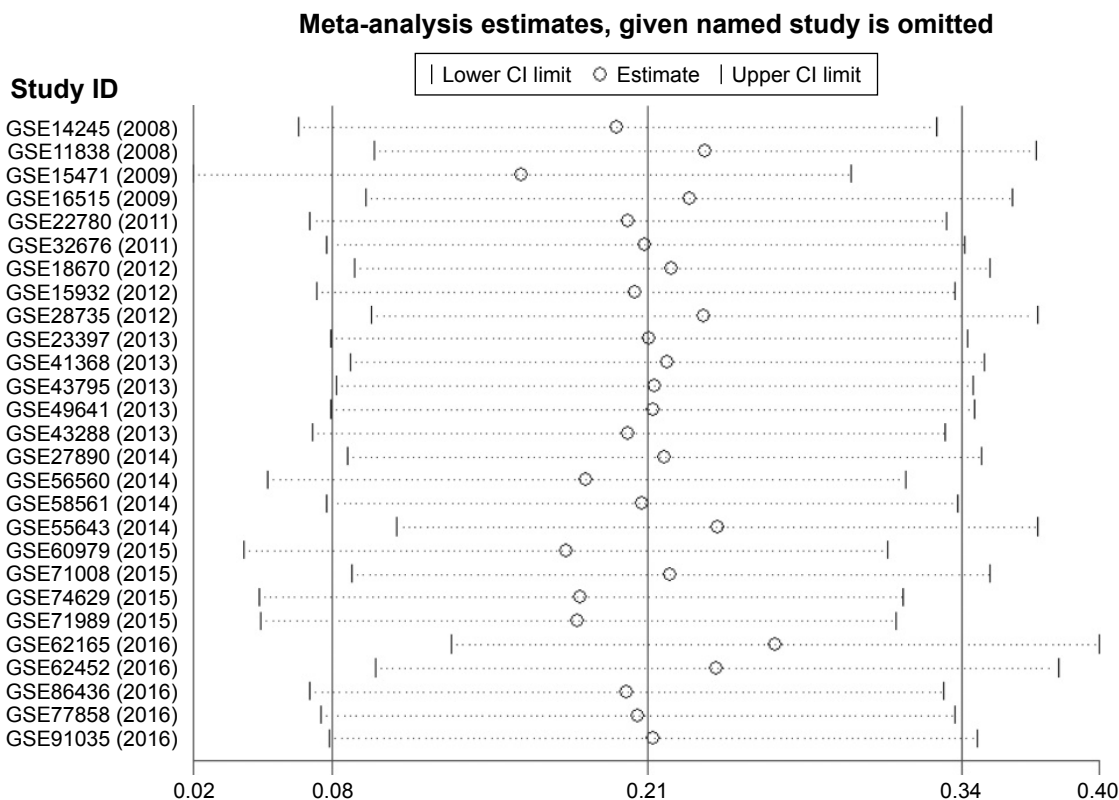
Figure 2 Sensitivity analysis to evaluate the heterogeneity among 27 GEO datasets.
Note: The results were computed by omitting each dataset in turn.
Abbreviations: CI, confidence interval; GEO, Gene Expression Omnibus.

analysis, we identified the mTOR signaling pathway, pathways in cancer, and the MAPK signaling pathway as the top three enriched pathways (Table 2). Interestingly, we also found that the PC pathway was one of the significantly enriched pathways. Five target genes (*VEGFC*, *CCND1*, *PIK3CB*, *VEGFA*, and *MAPK8*) were involved in the PC pathway. A PPI was constructed (Supplementary material 5) and six hub genes (*CCND1*, *MAPK8*, *VEGFA*, *FOS*, *CDH1*, and *HSP90AA1*) were identified (Figure 10).
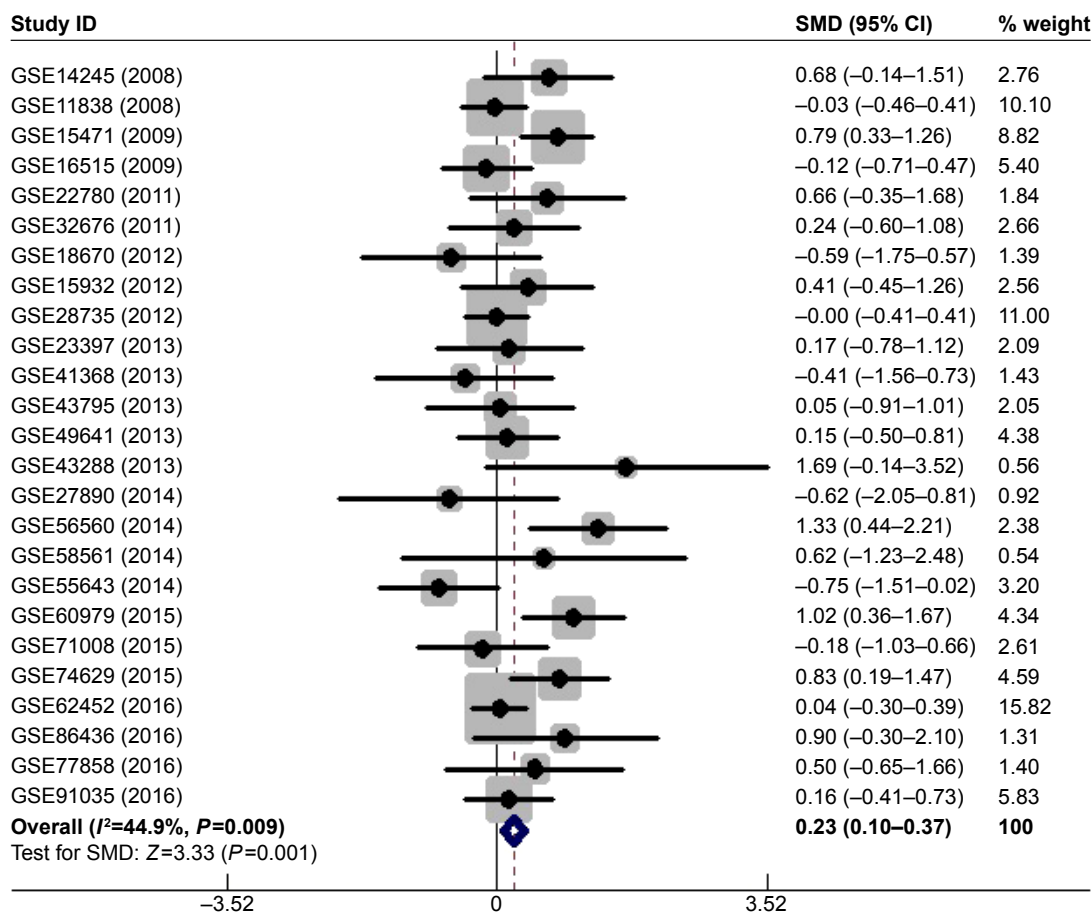
Discussion

In this paper, we first discovered that the expression of MALAT1 was significantly upregulated in PC patients by using a meta-analysis of 27 published GEO microarrays, which included a total of 764 patients with PC and 469 healthy controls. Further validation via the Oncomine database was consistent with the GEO microarray meta-analysis. Using the TCGA database, we acquired the survival data of the patients and investigated the influence of the alteration of MALAT1 expression on the patients' prognosis. Furthermore, using our own GeneChip and TCGA database, we collected and intersected the enriched GO genes after knocking down MALAT1 and the coexpressed genes in

TCGA. Further bioinformatics analysis of the overlapping genes, which aims to explore the underlying molecular regulatory mechanism, was performed.

During the past few years, alteration of MALAT1 has been found to be involved in malignancies and other diseases. Shuai et al[31] performed a meta-analysis that revealed that MALAT1 served as a promising biomarker for the prognosis of numerous malignancies, including digestive system cancers, urinary system cancers, and respiratory system cancers. In our study, we observed upregulated MALAT1, which is consistent with other previous reports, such as those by Li et al,[14] Pang et al,[32] and Liu et al.[33] Elevated MALAT1 has also been reported in gastric cancer,[34] hepatocellular carcinoma,[35] gallbladder cancer,[36] esophageal cancer,[37] and clear cell renal cell carcinoma.[38] Considering all of the above information, MALAT1 appears to play a distinct role in different tumors, which provides a brand new perspective for future cancer therapy.

In the present study, we discovered that MALAT1 showed a moderate performance in diagnosing PC after meta-analysis of a large number of PC, and corresponding control, samples. The area under the curve for the summary ROC curve was 0.75, with a sensitivity of 0.66 and a specificity of 0.72. Thus far in the published literature, there is

Study ID		SMD (95% CI)	% weight
GSE14245 (2008)		0.68 (−0.14–1.51)	2.76
GSE11838 (2008)		−0.03 (−0.46–0.41)	10.10
GSE15471 (2009)		0.79 (0.33–1.26)	8.82
GSE16515 (2009)		−0.12 (−0.71–0.47)	5.40
GSE22780 (2011)		0.66 (−0.35–1.68)	1.84
GSE32676 (2011)		0.24 (−0.60–1.08)	2.66
GSE18670 (2012)		−0.59 (−1.75–0.57)	1.39
GSE15932 (2012)		0.41 (−0.45–1.26)	2.56
GSE28735 (2012)		−0.00 (−0.41–0.41)	11.00
GSE23397 (2013)		0.17 (−0.78–1.12)	2.09
GSE41368 (2013)		−0.41 (−1.56–0.73)	1.43
GSE43795 (2013)		0.05 (−0.91–1.01)	2.05
GSE49641 (2013)		0.15 (−0.50–0.81)	4.38
GSE43288 (2013)		1.69 (−0.14–3.52)	0.56
GSE27890 (2014)		−0.62 (−2.05–0.81)	0.92
GSE56560 (2014)		1.33 (0.44–2.21)	2.38
GSE58561 (2014)		0.62 (−1.23–2.48)	0.54
GSE55643 (2014)		−0.75 (−1.51–0.02)	3.20
GSE60979 (2015)		1.02 (0.36–1.67)	4.34
GSE71008 (2015)		−0.18 (−1.03–0.66)	2.61
GSE74629 (2015)		0.83 (0.19–1.47)	4.59
GSE62452 (2016)		0.04 (−0.30–0.39)	15.82
GSE86436 (2016)		0.90 (−0.30–2.10)	1.31
GSE77858 (2016)		0.50 (−0.65–1.66)	1.40
GSE91035 (2016)		0.16 (−0.41–0.73)	5.83
Overall (I^2=44.9%, *P*=0.009)		**0.23 (0.10–0.37)**	**100**
Test for SMD: *Z*=3.33 (*P*=0.001)			

−3.52   0   3.52

Figure 3 Forest plot of 25 GEO datasets evaluating the SMD of MALAT1 in the PC group and the healthy control group after omitting two datasets (GSE15471 and GSE62165) (a fixed-effect model).
Abbreviations: CI, confidence interval; GEO, Gene Expression Omnibus; PC, pancreatic cancer; SMD, standard mean difference.

no report evaluating the diagnostic value of MALAT1 in PC through comprehensive meta-analysis. As a consequence, the information in this study will provide a valuable reference for clinical PC diagnosis. To date, several studies have

Begg's funnel plot with pseudo 95% CI
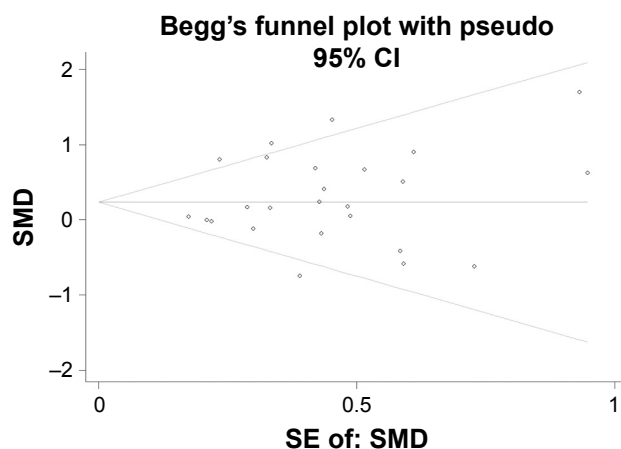
Figure 4 Begg's funnel plot evaluating the publication bias among the 25 datasets.
Abbreviations: CI, confidence interval; SE, standard error; SMD, standard mean difference.

reported that higher MALAT1 expression suggests poorer survival.[31,32] Regarding our results, we found that patients with MALAT1 alteration had better survival than patients without MALAT1 alteration (*P*=0.0380). We also found that the high expression of MALAT1 suggested better survival in the Kaplan–Meier curves. However, the *P*-values did not show statistical significance. Therefore, the exact prognostic value of MALAT1 in PC still remains unknown.

Since lncRNAs exert their regulatory function by specifically binding with target genes, we also aimed to determine possible targets of MALAT1 and to further explore the possible molecular pathways. Through a GO enrichment analysis, some top-ranked GO items, such as intracellular transport, membrane-enclosed lumen, and protein dimerization activity, were found to be possible crucial events in PC development. Regarding the KEGG pathway analysis, several significant pathways, such as the mTOR signaling pathway, pathways in cancer, and the MAPK signaling pathway, were discovered. Importantly, the PC pathway was also one of the enriched KEGG pathways. Five genes, *VEGFC*,

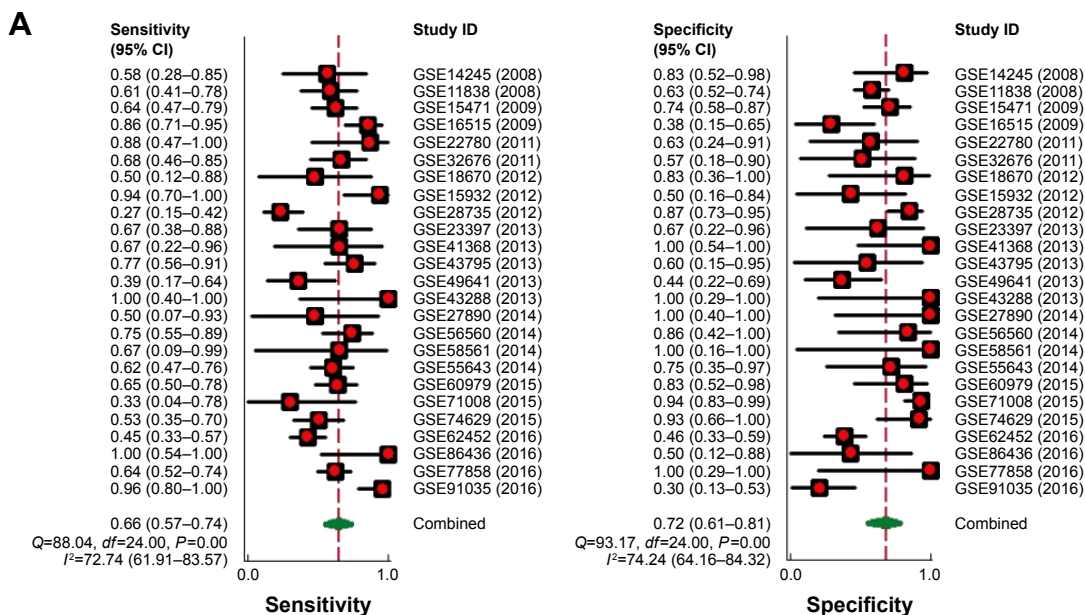## SROC with prediction and confidence contours
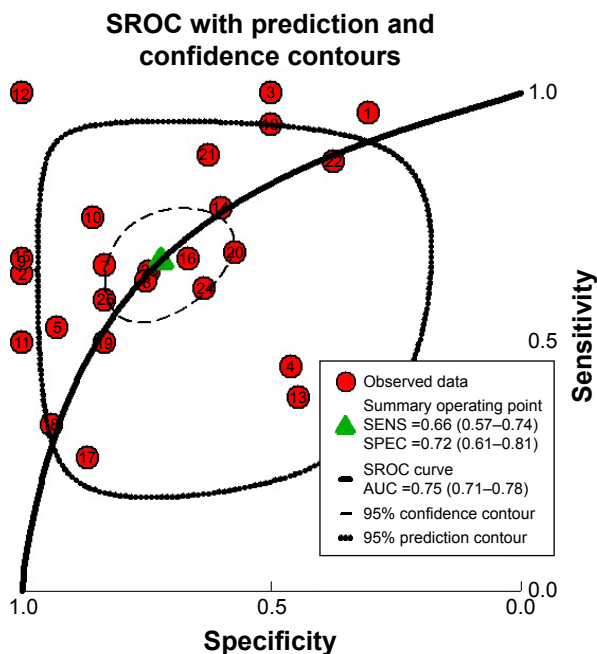
Figure 5 Summary of the ROC curve of 25 datasets evaluating the diagnostic value of MALAT1 in PC.
Abbreviations: AUC, area under the curve; CI, confidence interval; PC, pancreatic cancer; ROC, receiver operating characteristic; SENS, sensitivity; SPEC, specificity; SROC, summary receiver operating characteristic.

CCND1, *PIK3CB*, *VEGFA*, and *MAPK8*, are target genes of MALAT1, which might also be key target genes in PC. Interestingly, according to previous studies, we found that the mTOR signaling pathway has been reported to be involved in PC.[39,40] Additionally, the MAPK signaling pathway also has been reported to play a role in PC.[41] Therefore, to some extent, the pathways identified here might provide more

evidence for elucidating the potential molecular mechanism in PC. Finally, the PPI network suggests that six hub genes (*CCND1*, *MAPK8*, *VEGFA*, *FOS*, *CDH1*, and *HSP90AA1*) might serve as pivotal target genes of MALAT1 in PC. We discovered that *CCND1*, *MAPK8*, and *VEGFA* also participate in the PC pathway, as mentioned. As a consequence, we concentrate on the top three genes (*CCND1*, *MAPK8*, and *VEGFA*), which may be important genes, for detailed discussion.

CCND1, also known as BCL1, encodes a highly conserved cyclin protein, which exerts regulatory functions on CDK kinases. Mutation, amplification, and overexpression of CCND1 alter cell cycle progression, which might contribute to tumorigenesis. As a result, CCND1 alteration was frequently discovered in various tumors. The CCND1 genotype has been reported to be related to the risk for various cancers, such as colorectal cancer,[42] acute lymphoblastic leukemia,[43] glioma,[44] esophageal squamous cell carcinoma,[45] gastric cancer,[46] and nasopharyngeal carcinoma.[47,48] Moreover, CCND1 was also reported to play different roles in cancer survival, such as in lung adenocarcinoma,[49,50] prostate cancer,[51] meningioma[52] gastric adenocarcinoma,[53] and colorectal cancer.[54] In regards to PC, Deharvengt et al[55] reported that CCND1 is a target of a B lentivirus-delivered shRNA, which exerts suppressive effects on the growth, invasiveness, tumorigenicity, and proangiogenic potential of PC. Another study found that CCND1 was a strong prognostic biomarker for PC survival.[56] Taken together, it is obvious that CCND1 is involved in various cancers, including PC. However, whether
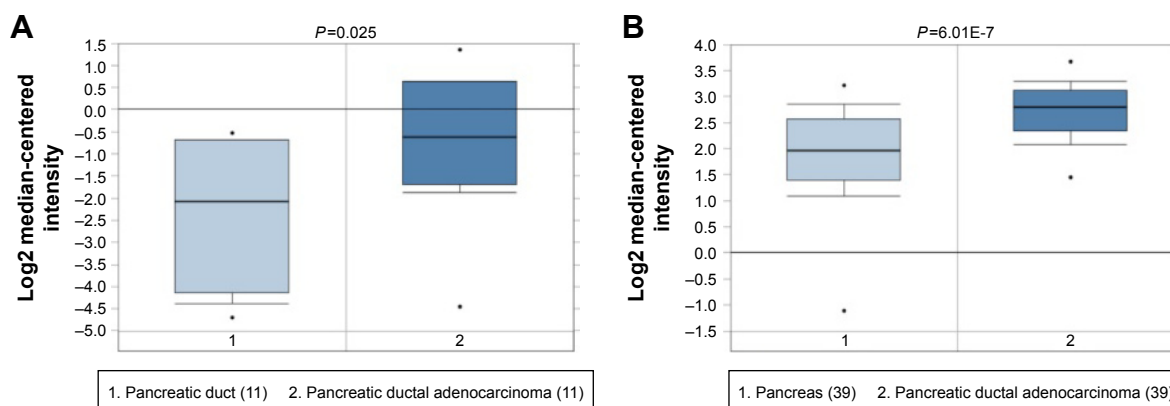

Figure 6 (*Continued*)

B

Study ID	Diagnostic OR (95% CI)
GSE14245	7.00 (1.04–46.95)
GSE11838	2.66 (1.10–6.46)
GSE15471	5.18 (1.96–13.69)
GSE16515	3.72 (0.93–14.85)
GSE22780	11.67 (0.92–147.56)
GSE32676	2.83 (0.51–15.77)
GSE18670	5.00 (0.34–72.77)
GSE15932	15.00 (1.29–174.39)
GSE28735	2.36 (0.80–6.99)
GSE23397	4.00 (0.54–29.80)
GSE41368	23.40 (0.89–612.99)
GSE43795	5.00 (0.67–37.26)
GSE49641	0.51 (0.13–1.92)
GSE43288	63.00 (0.98–4,042.12)
GSE27890	9.00 (0.30–271.66)
GSE56560	18.00 (1.84–176.56)
GSE58561	8.33 (0.22–320.39)
GSE55643	4.94 (0.89–27.32)
GSE60979	9.41 (1.85–47.95)
GSE71008	7.83 (1.00–61.46)
GSE74629	14.53 (1.72–123.07)
GSE62452	0.69 (0.35–1.38)
GSE86436	13.00 (0.51–330.48)
GSE77858	12.16 (0.61–243.91)
GSE91035	10.50 (1.18–93.70)

Random effects model
Pooled diagnostic odds ratio =4.41 (2.71 to 7.17)
Cochran-Q =44.40; df=24 (P=0.0068)
Inconsistency I^2=45.9%
τ^2=0.5836

0.01   1   100
Diagnostic odds ratio

Figure 6 Forest plot of 25 datasets evaluating the diagnostic value of MALAT1 in PC.
Notes: (**A**) Pooled sensitivity and specificity of 25 datasets. (**B**) Pooled diagnostic OR of 25 datasets.
Abbreviations: OR, odds ratio; PC, pancreatic cancer.

it is related to MALAT1 has not been reported to date, and further studies are required.

MAPK8, also named JNK or JNK1, belongs to the MAPK kinase family that acts as an integration point in numerous cellular processes, such as proliferation, differentiation, transcription regulation, and development. Increasing amounts of evidence have proven that MAPK8 has a crucial role in a wide variety of cancers. Chang et al[57] discovered that JNK1 activation was capable of predicting poor prognosis in hepatocellular carcinoma. Moreover, Okada et al[58] reported that JNK can inhibit temozolomide, thus acting as a rational therapeutic biomarker in glioblastoma. JNK activity
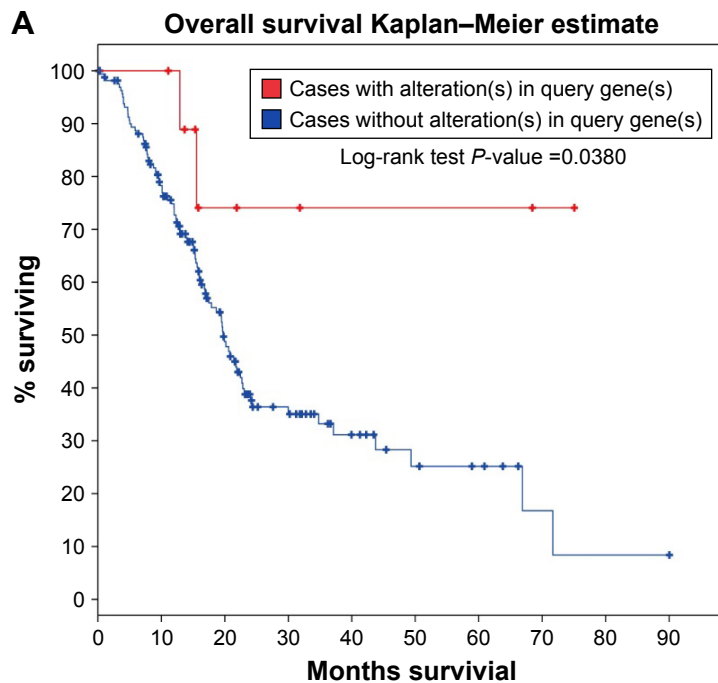
A

P=0.025

Log2 median-centered intensity

1. Pancreatic duct (11)   2. Pancreatic ductal adenocarcinoma (11)

B

P=6.01E-7

Log2 median-centered intensity

1. Pancreas (39)   2. Pancreatic ductal adenocarcinoma (39)

Figure 7 (*Continued*)

C *P*=0.114

1. Pancreas (5)   2. Pancreatic adenocarcinoma (12)

D *P*=0.159

1. Pancreas (16)   2. Pancreatic carcinoma (36)

E *P*=0.476

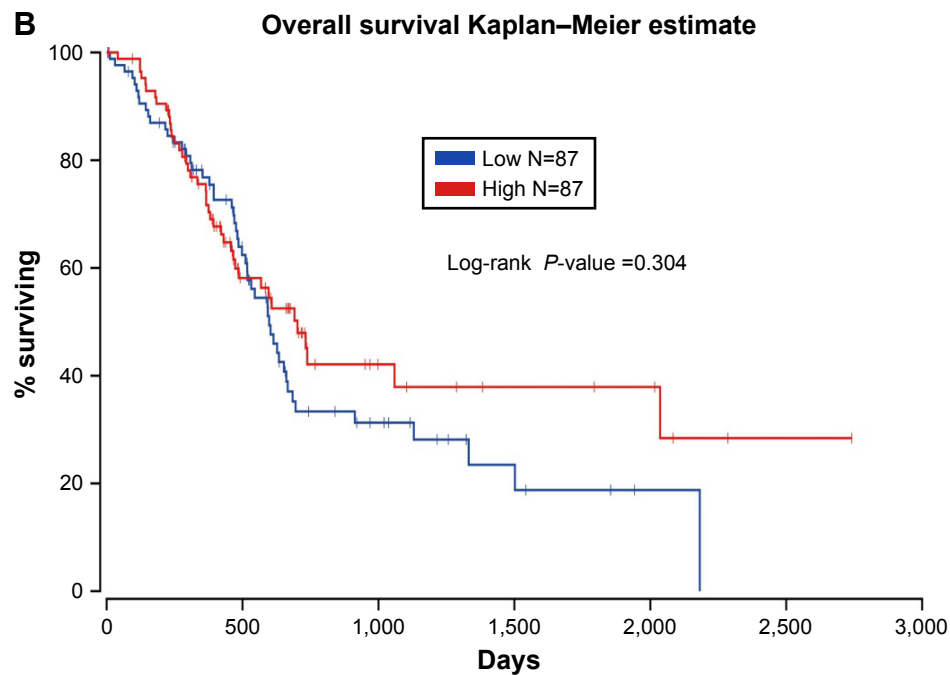1. Pancreatic duct (25)   2. Pancreatic ductal adenocarcinoma (24)
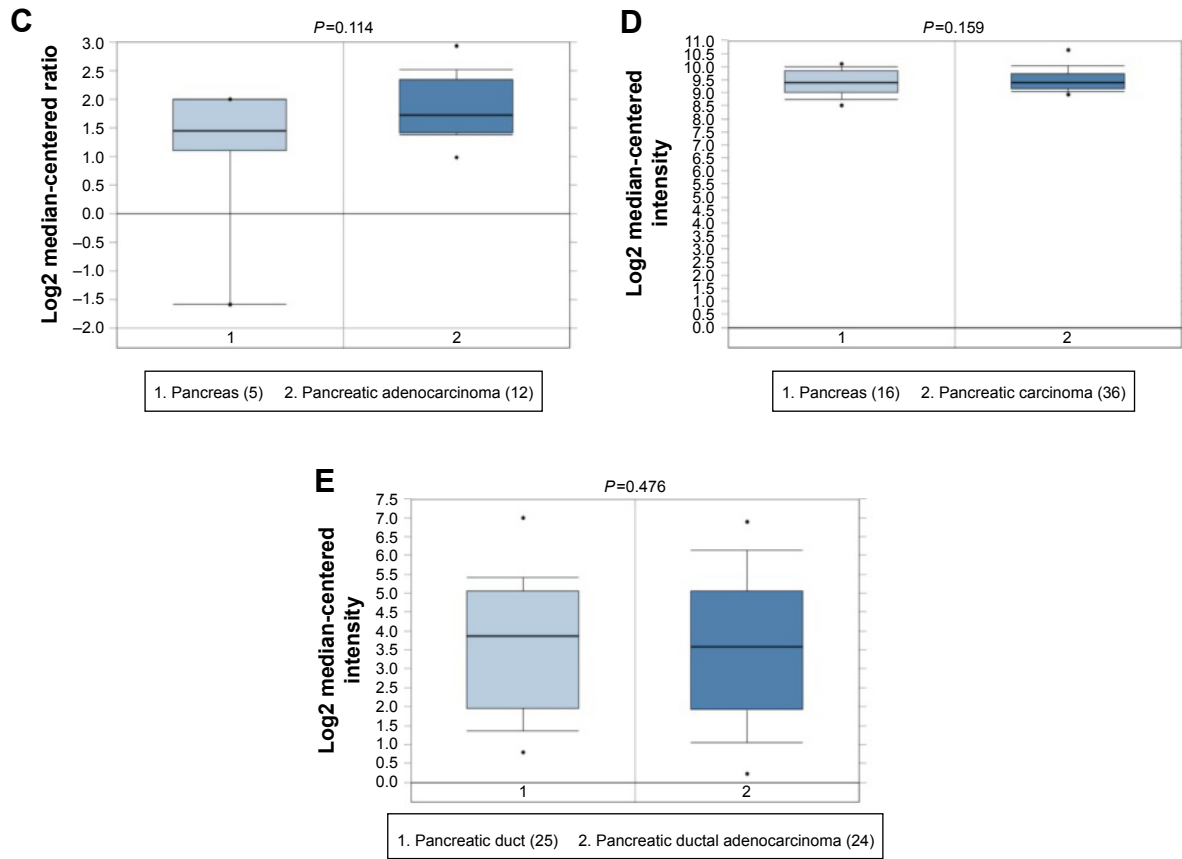
Figure 7 Box plot validating the expression of MALAT1 in the PC group and the healthy control group based on the Oncomine database.
Notes: (**A**) The expression of MALAT1 in Grutzmann's dataset (http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-950). (**B**) The expression of MALAT1 in Badea's datasets (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE15471). (**C**) The expression of MALAT1 in Iacobuzio-Donahue's dataset (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE3654; http://genome-www.stanford.edu/pancreatic1/index.shtml). (**D**) The expression of MALAT1 in Pie's dataset (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE16515). (**E**) The expression of MALAT1 in Ishikawa's dataset (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE1542).
Abbreviation: PC, pancreatic cancer.

A   **Overall survival Kaplan–Meier estimate**

☐ Cases with alteration(s) in query gene(s)
☐ Cases without alteration(s) in query gene(s)
Log-rank test *P*-value =0.0380

Figure 8 (*Continued*)

B

Overall survival Kaplan–Meier estimate

Figure 8 Survival analysis evaluating the prognostic value of MALAT1 in PC.
Notes: (**A**) Patients with PC were divided into two groups according to MALAT1 alteration. (**B**) Patients with PC were divided into two groups according to MALAT1 expression.
Abbreviation: PC, pancreatic cancer.

**Table 2** Significant GO and KEGG items of the overlapping genes

Category	Term	Count	P-value	Benjamini	FDR
GOTERM_BP_FAT	GO:0046907~intracellular transport	26	2.06E-05	0.032809	0.034527
GOTERM_BP_FAT	GO:0051174~regulation of phosphorus metabolic process	21	4.84E-05	0.038433	0.081107
GOTERM_BP_FAT	GO:0019220~regulation of phosphate metabolic process	21	4.84E-05	0.038433	0.081107
GOTERM_BP_FAT	GO:0042325~regulation of phosphorylation	20	8.74E-05	0.046107	0.146486
GOTERM_BP_FAT	GO:0042127~regulation of cell proliferation	26	0.000361	0.136005	0.603502
GOTERM_BP_FAT	GO:0045859~regulation of protein kinase activity	15	0.000791	0.226216	1.318661
GOTERM_BP_FAT	GO:0006915~apoptosis	21	0.00083	0.200893	1.383248
GOTERM_BP_FAT	GO:0012501~programmed cell death	21	0.000996	0.205897	1.656699
GOTERM_BP_FAT	GO:0043549~regulation of kinase activity	15	0.001101	0.199873	1.829703
GOTERM_BP_FAT	GO:0006468~protein amino acid phosphorylation	22	0.001215	0.196549	2.01821
GOTERM_CC_FAT	GO:0031974~membrane-enclosed lumen	43	0.000252	0.073845	0.337322
GOTERM_CC_FAT	GO:0031981~nuclear lumen	36	0.000284	0.042315	0.380158
GOTERM_CC_FAT	GO:0000228~nuclear chromosome	10	0.000299	0.029887	0.400137
GOTERM_CC_FAT	GO:0043233~organelle lumen	41	0.000671	0.049713	0.894362
GOTERM_CC_FAT	GO:0043232~intracellular nonmembrane-bounded organelle	53	0.000793	0.047075	1.056321
GOTERM_CC_FAT	GO:0043228~nonmembrane-bounded organelle	53	0.000793	0.047075	1.056321
GOTERM_CC_FAT	GO:0005793~ER–Golgi intermediate compartment	5	0.00204	0.098288	2.697165
GOTERM_CC_FAT	GO:0000267~cell fraction	27	0.002078	0.086396	2.747502
GOTERM_CC_FAT	GO:0005624~membrane fraction	22	0.002295	0.083602	3.029516
GOTERM_CC_FAT	GO:0005875~microtubule-associated complex	7	0.002312	0.075206	3.05179
GOTERM_MF_FAT	GO:0046983~protein dimerization activity	21	0.000154	0.061492	0.215993
GOTERM_MF_FAT	GO:0001882~nucleoside binding	43	0.000155	0.031399	0.217151
GOTERM_MF_FAT	GO:0001883~purine nucleoside binding	42	0.000274	0.036878	0.383329
GOTERM_MF_FAT	GO:0000166~nucleotide binding	53	0.000428	0.043003	0.597241
GOTERM_MF_FAT	GO:0032553~ribonucleotide binding	45	0.000667	0.053362	0.929846
GOTERM_MF_FAT	GO:0032555~purine ribonucleotide binding	45	0.000667	0.053362	0.929846
GOTERM_MF_FAT	GO:0017076~purine nucleotide binding	46	0.000898	0.059685	1.25016

(Continued)

Table 2 (*Continued*)

Category	Term	Count	*P*-value	Benjamini	FDR
GOTERM_MF_FAT	GO:0019003~GDP binding	4	0.001858	0.103456	2.570935
GOTERM_MF_FAT	GO:0032559~adenyl ribonucleotide binding	37	0.002135	0.104001	2.948878
GOTERM_MF_FAT	GO:0030554~adenyl nucleotide binding	38	0.00282	0.120983	3.87696
KEGG_PATHWAY	hsa04150:mTOR signaling pathway	6	0.001995	0.195612	2.227286
KEGG_PATHWAY	hsa05200:Pathways in cancer	13	0.011181	0.458169	11.91115
KEGG_PATHWAY	hsa04010:MAPK signaling pathway	11	0.017441	0.472326	18.00035
KEGG_PATHWAY	hsa04110:Cell cycle	7	0.021605	0.44854	21.83569
KEGG_PATHWAY	hsa05218:Melanoma	5	0.035037	0.540452	33.12075
KEGG_PATHWAY	hsa05219:Bladder cancer	4	0.036427	0.490394	34.19945
KEGG_PATHWAY	hsa05212:PC	5	0.036617	0.440598	34.34575

Abbreviations: FDR, false discovery rate; GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; PC, pancreatic cancer.
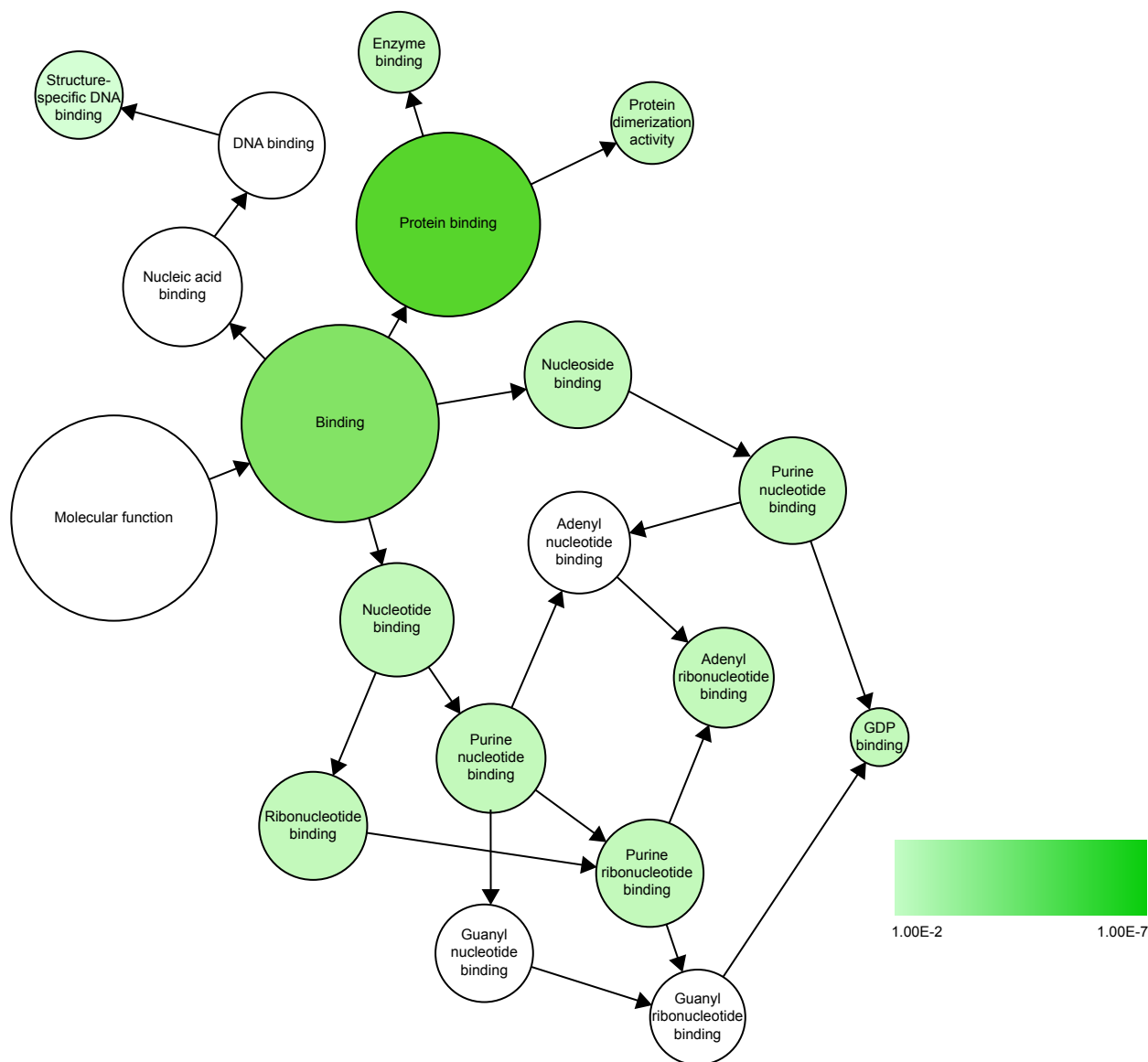
Figure 9 Significant MF network constructed by Cytoscape 3.40.
Notes: Each node represents a GO item. Colored nodes indicate significance (*P*<0.01). Node size represents the involved MF items.
Abbreviations: GO, Gene Ontology; MF, molecular function.
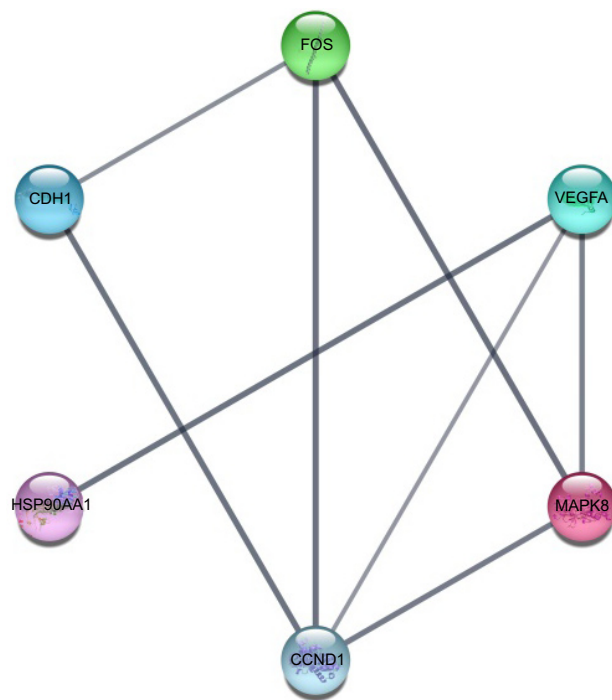
Figure 10 Hub genes obtained from the PPI network.
Note: A degree ≥10 was chosen as the cut-off point.
Abbreviation: PPI, protein–protein interaction.

has also been reported in endometrial cancer,[59] melanoma,[60] colon cancer,[61] etc. In regard to PC, Sahu et al[62] reported that JNK was involved in benzyl isothiocyanate (BITC)-mediated G(2)/M arrest to mediate apoptosis in human PC cells. Another study also found that JNK1 is activated by Irofulven treatment, which induces apoptosis in human PC cells.[63] Therefore, based on the abovementioned information, MAPK8 could possibly serve as a target gene of MALAT1 in PC, but additional studies are needed.

VEGFA, also known as VPF, VEGF, and MVCD1, is a member of the PDGF/VEGF growth factor family. Encoding a heparin-binding protein, it induces the proliferation and migration of vascular endothelial cells, which is fundamental for physiological and pathological angiogenesis. Upregulated VEGFA is common in many tumors and is related to tumor stage and progression. Liu et al[64] reported that VEGFA polymorphisms might play a potential role in the development and clinical outcome of hepatocellular carcinoma. VEGFA has also been found to be inhibited by miR-1, which exerts inhibitory functions in osteosarcoma cells, as well as promotes proliferation, migration, and invasion.[65] Furthermore, Zeng et al[66] found that downregulation of VEGFA can inhibit proliferation, migration, and invasion and can promote apoptosis in renal clear cell carcinoma. VEGFA in tumors has also been studied in

breast cancer,[67,68] laryngeal carcinoma,[69] colorectal cancer,[70,71] bladder cancer,[72] and lung cancer.[73] In PC, Liu et al[74] found that VEGFA was involved in the Twist/miR-497/VEGFA axis, which is significantly correlated to metastasis and angiogenesis in PC. VEGFA secretion has also been reported to be regulated by myoferlin and affects tumor-associated angiogenesis in PC.[75] In summary, as VEGFA has been shown to be important in cancers, it is suggested that VEGFA is a target of MALAT1 in PC. However, more studies are needed.

There are limitations in this study. Here, we have attempted to explore the relationship between MALAT1 and PC clinical parameters based on the TCGA database. However, no statistical significance was observed. Therefore, it still remains to be seen if there is a correlation between MALAT1 and PC. In addition, in this study, the exploration occurred at a bioinformatics level. Further experiments are needed to validate these assumptions.

In sum, we discovered that upregulated MALAT1 acts as a biomarker for the diagnosis of PC. The alteration or upregulation of MALAT1 expression tends to predict better prognostic outcome in PC. Several pathways, such as the mTOR signaling pathway, pathways in cancer, and the MAPK signaling pathway, might be targeted by MALAT1 and participate in the PC process. Six hub genes (*CCND1*, *MAPK8*, *VEGFA*, *FOS*, *CDH1*, and *HSP90AA1*), and specifically *CCND1*, *MAPK8* and *VEGFA*, were identified as key target genes of MALAT1 in PC. Here, we provided clinical reference values and underlying molecular mechanisms for MALAT1 in PC. These findings still need to be validated in future studies.

Conclusion

MALAT1 seems to act as a promising diagnostic biomarker in PC. Six hub genes (*CCND1*, *MAPK8*, *VEGFA*, *FOS*, *CDH1*, and *HSP90AA1*), specifically *CCND1*, *MAPK8*, and *VEGFA*, might be key correlative genes of MALAT1 in PC. Several pathways related to MALAT1, such as the mTOR signaling pathway, pathways in cancer, and the MAPK signaling pathway may play a crucial role in PC. The information we have obtained might shed light on clinical application and future research. Nevertheless, more experimental studies are needed to further validate these findings.

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Disclosure

The authors report no conflicts of interest in this work.

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