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REVIEW ARTICLE

Advances in oral mesenchymal stem cell-derived extracellular vesicles in health and disease



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Received 19 November 2022; received in revised form 8 March 2023; accepted 15 March 2023 Available online 12 April 2023

KEYWORDS

Disease; Extracellular vesicles; Oral tissue MSC; Therapeutic application; Tissue regeneration **Abstract** Extracellular vesicles (EVs) are nano-size vesicles secreted naturally by all cells into the extracellular space and have been recognized as important cell—cell mediators in multicellular organisms. EVs contain nucleic acids, proteins, lipids, and other cellular components, regulating many basic biological processes and playing an important role in regenerative medicine and diseases. EVs can be traced to their cells of origin and exhibit a similar function. Moreover, EVs demonstrate low immunogenicity, good biocompatibility, and fewer side effects, compared to their parent cells. Mesenchymal stem cells (MSCs) are one of the most important resource cells for EVs, with a great capacity for self-renewal and multipotent differentiation, and play an essential role in stem cell therapy. The mechanism of MSC therapy was thought to be attributed to the differentiation of MSCs after targeted migration, as previously noted. However, emerging evidence shows the previously unknown role of MSCs (OMSC-EVs), show more advantages than those of all other MSCs in tissue repair and regeneration, due to their lower invasiveness and easier accessibility for sample collection. Here, we systematically review the biogenesis and biological characteristics of OMSC-EVs, as well as the role of OMSC-

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https://doi.org/10.1016/j.gendis.2023.03.015

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EVs in intercellular communication. Furthermore, we discuss the potential therapeutic roles of OMSC-EVs in oral and systemic diseases. We highlight the current challenges and future directions of OMSC-EVs to focus more attention on clinical translation. We aim to provide valuable insights for the explorative clinical application of OMSC-EVs.

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Introduction

Extracellular vesicles (EVs) released by cells of all organisms, such as eukarvotes, prokarvotes, or plants, exert different functions, primarily relying on the properties of the cell origin and the state of the cell at the time of purification.¹ One of the most studied EVs is derived from mesenchymal stem cells (MSCs), which possess characteristics that are homogeneous to those of their parental cells. MSCs are a heterogeneous population and exist in a variety of human tissues, 2^{-4} where they may express different tissue-specific molecular markers.⁵ Oral tissue MSCs (OMSCs) are a class of MSCs derived from oral tissues, including dental pulp stem cells (DPSCs),⁶ stem cells from exfoliated deciduous teeth (SHEDs),⁷ periodontal ligament stem cells (PDLSCs),⁸ dental follicle progenitor cells (DFPCs),⁹ stem cells from the apical papilla (SCAPs),¹⁰ and gingival mesenchymal stem cells (GMSCs).^{11,12} They are characterized by great self-renewal and multidirectional differentiation abilities, similar to those of all other tissuederived MSCs. Of note, OMSCs have remarkable advantages, such as being widely available, easy to obtain, and minimally invasive, and thus have become one of the most promising therapeutic options for tissue regeneration and repair.¹³ Previously, MSC therapy was believed to be achieved primarily through direct migration to the site after transplantation with the potential for increased proliferation and differentiation. However, in recent years, studies have established that MSC therapy is largely dependent on the paracrine factors of MSCs rather than the MSCs themselves.¹⁴ Among the many paracrine factors of MSCs, MSCderived EVs (MSC-EVs) have started to attract more attention, due to their drug delivery capacity, high specificity, low immunogenicity, good biocompatibility, high stability, and no cytotoxicity, compared to their parental cells.¹⁵ In this review, we summarize the biological origin, heterogeneity, and uptake mechanism of EVs, and focus on the role and potential application of OMSC-EVs in local and systemic diseases, in order to provide a valuable reference for the research on EVs, especially OMSC-EVs and their future potential clinical application.

The biogenesis, heterogeneity, and functionality of extracellular vesicles

EVs are membrane-containing vesicles released by all types of cells, which are divided into three main categories according to their biological origin and size (Table 1), including exosomes (EXOs), microvesicles (MVs), and apoptotic extracellular vesicles (ApoEVs).¹⁶ Other subpopulations such as supremeres and exomeres have also been recently referred to (Fig. 1).^{17,18}

EXOs, also called small extracellular vesicles (sEVs), are one group of EVs that originate from endosomes with diameters of around 30–150 nm.¹⁹ The biogenesis and development of EXOs have been extensively reviewed elsewhere^{20–22} and it mainly involves double invagination of the plasma membrane and the formation of multivesicular bodies (MVBs) containing abundant intraluminal vesicles (ILVs).²³ It is known that the cellular protein network plays a crucial role in the biogenesis and secretion of EXOs, including endosomal sorting complexes required for transport (ESCRTs), and tetraspanins such as CD63, CD81, and CD9.^{23–25} Functionally, EXOs carry many

| Characteristics | FXOs | MVs | ApoEVs | Reference |
|-----------------|---------------------------------------|---|-------------------------|-----------|
| | | 1414.5 | | Kererence |
| Source cells | Living cells | Living cells | Apoptotic cells | 46 |
| Size | ~50–150 nm | ~150–1000 nm | ~500–5000 nm | 19,47,48 |
| Mechanism | Double invagination | Plasma membrane | Microtubule spike, | 20,49 |
| of biogenesis | of the plasma membrane | budding outward directly | apoptopodia, and | |
| | | | beaded apoptopodia | |
| Cargo | Proteins, mRNA, and miRNA | Proteins, plasmid DNA, | Organelles, fragments, | 50-52 |
| | | mRNA, and miRNA | proteins, nucleic acids | |
| Markers | CD63, CD81, CD9 | Annexin A1, ADP-ribosylation Phosphatidylserine factor 6 exposure, activated | Phosphatidylserine | 28,53 |
| | | | exposure, activated | |
| | | | caspase-3 | |
| Centrifugal | 100,000-120,000 g | 1000-10,000 g | 500—16,000 g | 54,55 |
| Condition | , , , , , , , , , , , , , , , , , , , | , 3 | <i>,</i> 3 | |



Figure 1 The biogenesis and secretion of EVs. The biogenesis of EXOs involves double invagination of the plasma membrane and the formation of multivesicular bodies (MVBs). Briefly, the plasma membrane first invaginates to form early endosomes (EE), and then EEs undergo the second invagination of the membrane to mature into late endosomes (LE), also called MVB, which includes a large collection of ILVs; lastly, MVBs fuse with the plasma membrane and release ILVs into the extracellular space, which forms EXOs with diameters of \sim 30–150 nm; while MVs were produced by the plasma membrane directly budding outward, triggered by the change in cell membrane stiffness and curvature change. Unlike the ApoEV former, the formation is a dynamic process, which is regulated by the morphological steps of apoptotic cell disassembly and contains different forms such as microtubule spike, apoptopodia, and beaded apoptopodia. A variety of molecules, such as MLC, MLCK, and ESCRT, participate in the regulation of EV biogenesis and release. This graph was created with Biorender.com.

bioactive molecules and play an important role in intercellular communication, signal transduction, regulation of immune response, antigen presentation, and epigenetic reprogramming of recipient cells, and it depends largely on the physiological and pathological conditions of the tissues or cells of origin and the surface receptors of the recipient cells.²¹ MVs are small membrane sacs produced by exuding from the cell surface with a size of 150-1000 nm.²⁶ It is generally believed that the biogenesis of MVs is a multistep process, including the transport of cellular cargoes to the plasma membrane, the rearrangement of membrane lipids, and the mechanical contraction-mediated blebbing extrusion.^{27,28} MVs have been widely detected in various biological fluids such as peripheral blood, urine, and ascites, and play an important role in various diseases, including tumor invasion and metastasis, inflammation, and tissue regeneration by promoting horizontal transfer of bioactive molecules such as proteins, RNA, and microRNA.²⁹⁻³¹ ApoEVs are the largest type of EV produced by cells when they were undergoing apoptosis, typically having a size greater than 1 μ m. ApoEV formation is a dynamic process, which is regulated by the morphological steps of apoptotic cell disassembly, controlled by protein kinases such as Rhoassociated protein kinase 1 (ROCK1) and MLCK, the membrane channel PANX1.³² ApoEVs are "small, sealed sacs" that were previously considered garbage bags. Until recently, their ability to deliver useful materials, such as self-antigens, to neighboring or distant receptor cells has come to light. ApoEVs can not only promote efficient removal of cell debris by surrounding phagocytes, but also carry biomolecules, including protein, microRNA, and DNA, to regulate cell-to-cell communication.^{33,34}

EVs show great heterogeneity in cargo and functionality according to their sources of cells and tissues, and even the method of purification. It is generally believed that EVs obtained by differential centrifugation contain a heterogeneous mixture of all types of EVs and non-vesicular compartments and even some levels of soluble proteins.³⁵ Ultracentrifugation (UC) and size-exclusion chromatography (SEC) appeared to give a very high better purity of EVs.³⁶ Indeed, subtle molecular differences at the level of a single EV may lead to significant changes in the biological function of EVs.³⁷ It becomes increasingly critical to develop a novel technology to analyze EVs at a singlemolecule level to be able to identify the heterogeneity of the EV population. Recently, advances in novel fluorescence microscopy techniques make it possible to detect EVs at the single-molecule level.³⁸ More isolation methods have been reported, such as the combined enrichment method,³⁹ modern approaches based on size, charge, and affinity,⁴⁰ and apoplastic washing fluid.⁴¹ Additionally, nanoflow cytometry or advanced flow cytometry has been used to differentiate EVs by recognizing the different surface markers labeled with fluorophores.^{42,43} A single EV flow cytometry analysis (FCA)⁴⁴ is enabled by target-initiated engineering (TIE) of DNA nanostructures on individual EVs, which employs a conformation-switchable DNA probe to bind to the surface marker of the EV, triggering the engineering of a DNA nanostructure by a hybridization chain reaction (HCR).⁴⁵ HCR products not only enlarge the overall size of a single EV to be beyond 500 nm, but can also bind to multiple fluorophores to amplify the signal from the marker molecules located on the limited area of the EV surface, both enabling visualization of a single EV in a conventional flow cytometer and greatly simplifying the measurement of multiple markers on the same EV. However, it is still challenging to isolate the entire population of EVs using current methods as only certain known surface markers are available for EV detection.⁴⁴

The mechanisms of EVs in intercellular communication

Recent studies highlight that EVs released by cells, acting as paracrine factors, interact with the recipient cell by migrating to distant sites and reaching the target cells.⁵⁶ The topological structure of EVs is similar to that of cells, and functional communication between EVs and cells can involve different types of interactions, including fusion of the EV-plasma membrane, endocytosis uptake, and binding of EVs to the cell surface via the ligand-receptor without delivery of the contents (Fig. 2).⁵⁷

The direct fusion of the EV membrane with the lipid bilayer of the plasma membrane is one way to communicate with neighborhood cells, which is regulated by several proteins, such as soluble receptor for the N-ethyl-maleimide-sensitive factor attachment protein (SNARE), Rab and Sec1/Munc18-like proteins (SM),⁵⁷ although it maybe not be the main route for EVs entering cells under

physiological conditions.^{58,59} Endocytosis is considered one of the main pathways for the uptake of EVs, including clathrin and caveolin-mediated endocytosis, macrophagocytosis.60 pinocytosis. and Clathrin-mediated endocytosis is achieved by clathrin-coated vesicles that deform the membrane into bubbles and break off. In general, clathrin and caveola-mediated endocytosis play a role in EV uptake, but the precise mechanism of this pathway remains to be further investigated.⁶⁰ Macropinocytosis is another endocytosis uptake pathway, and the mechanism is similar to phagocytosis, but it does not require direct contact with internalizing substances, rather relies on the GTPase RAC1, actin, and cholesterol, and requires Na⁺/H⁺ exchange activity.^{61–63} These studies at least suggest that macropinocytosis may be a secondary pathway for the internalization of EV or an uptake mechanism for certain cell types. Phagocytosis is a receptor-ligand-mediated endocytosis event. When EVs bind to the cell membrane by the corresponding receptor and ligand, downstream signal transduction can be initiated, leading to aggregation of actin at the uptake site. Actin contraction causes the plasma membrane to protrude and form pseudopodia to envelop EVs or other extracellular molecules, and then the pseudopodia fuse to form bubbles. In the cytoplasm, the dynein at the neck of the bubbles contracts and breaks off the bubbles to form phagosomes. This process occurs primarily during the uptake of large EVs, such as ApoEVs, which are mainly carried out by specialized cells such as macrophages and dendritic cells.^{60,64} Phosphatidylserine (PS) exposure is the characteristic surface structure of



Figure 2 The mechanism of EV uptake. EVs can participate in cellular communication through molecule transport or signal transfer between cells. EVs can enter recipient cells by direct membrane fusion. Alternatively, EVs can also be internalized by macropinocytosis and clathrin and caveolin-mediated endocytosis. Otherwise, some larger EVs, such as apoEVs, can also be taken up by target cells through phagocytosis. This graph was created with Biorender.com.

ApoEVs, which differs from the other two types of EV, and PS can recruit phagocytes and promote the phagocytosis of ApoEVs.⁶⁵

The superiority of OMSC-EVs and their biological function in health

Compared to MSCs from other tissues, OMSCs have greater advantages of wide source, easy access, and being minimally invasive, together with their excellent proliferation and differentiation abilities,66,67 making them a strong regenerative and therapeutic potential for promising application prospects.^{68,69} Due to the origin of the neural crest, OMSCs have considerable potential for neurogenic differentiation, thereby OMSC-EVs have unique advantages in the prevention and treatment of neurotrophic and peripheral neurosystemic diseases.^{70–73} Furthermore, OMSC-EVs have stronger immunomodulatory abilities.^{74,75} DPSC-EXOs have been reported to inhibit differentiation of CD4⁺ T cells in Th17 to reduce the secretion of pro-inflammatory cytokines IL-17 and TNF- α , but promote differentiation in Tregs to increase the release of anti-inflammatory cytokines IL-10 and TGF- β , compared to BMSC-EXOs.⁷⁶ Even though different tissue-derived MSCs have similar stem cell characteristics, their relative differences in cytokine profiles can give them unique biological significance.⁷⁷ For example, OMSCs possess a unique cytokine profile and therefore have more significant advantages than BMSCs in proliferation, osteogenesis, and odontogenesis. 66,77,78 Especially in the aspect of tooth and periodontal tissue regeneration, OMSCs have significant advantages over other MSCs, such as PDLSCs in periodontal regeneration⁷⁹ and DPSCs in dental pulp regeneration.⁸⁰ Therefore, we focused on OMSC-EVs in this review and discussed their biological functions in health and disease.

Emerging evidence supports that MSC-induced paracrine signaling is important to maintain tissue homeostasis.81,82 EVs, as a component of the MSC secretome, participate in the transfer of proteins and genetic material, and the regulation of various physiological processes.⁸² It is known that EVs secreted by SCAP, which are derived from the apical papilla of incompletely developed teeth, play an important role in root formation and development.83,84 OMSC-EVs may have a positive effect on odontogenic differentiation and dentin-pulp formation. PIWI-interacting RNAs (piRNAs) are a new class of small ncRNAs, which form RNA-induced silencing complexes by binding to the Pelement-induced wimpy testis protein family (PIWI), one of the argonautes, contributing to maintaining the functional integrity of stem cells.⁸⁵ Compared to the piRNA expression profiles of SCAP-EXOs with BMSC-EXOs, the higher expression of piRNAs in SCAP-EXOs activated the MAPK signaling pathway, resulting in odonto-/osteogenic differentiation.⁸ It has been suggested that dentin-pulp formation involves complex epithelial-mesenchymal interactions between Hertwig epithelial root sheath cells (HERS) and dental cells (DPC), and EXOs secreted by HERS (ELVs-H1) containing the Wnt3a protein, can up-regulate the expression of β -catenin and activate Wnt/β -catenin signaling pathway in DPC to induce angiogenesis, odontoblast differentiation, and neural differentiation of dental MSCs.⁸⁷ The beneficial effect of OMSC-EVs in inducing odontogenic differentiation of stem cells suggests that OMSC-EVs may play an important role in the formation of pulp and dentin, and also provides a valuable theory for pulp and dentin regeneration.88,89 DPSC-EXOs promote odontogenic differentiation through transporting miR-27a-5p to down-regulate the latent TGF- β -binding protein 1 (LTBP1), one of the inhibitory molecules of TGF β 1 signaling, to activate TGF β 1/Smad signaling pathway.⁸⁸ DPSC-EXOs also induce odontogenic differentiation through the p38/MAPK pathway, which increases the expression of key regulators such as DMP1 and DPP.⁸⁹ In addition, the balance between bone formation of osteoblasts and bone resorption of osteoclasts is crucial for the development of oral and facial bones. Studies exploring the communication between OMSC and osteoclasts found that miR-3-4p from OMSC-EXOs is an important downstream factor of GATA4 that directly targets BMP-3 and NFATc1 in activated T cells to regulate the functions of OMSCs and osteoclasts,⁹⁰ which plays a key role in the development of oral facial bone.

The role of OMSC-EVs in the regeneration

Effective tissue repair and regeneration after injury are crucial for maintaining the physiological functions of tissues. Here, we focus on the role and current application of OMSC-EVs in various tissue regeneration (Fig. 3).

The role of OMSC-EVs in periodontal bone remodeling

OMSC-EVs have prominent advantages in periodontal bone regeneration.^{83,91} Periodontal bone defects generally include inflammatory bone loss caused by periodontitis and mechanical bone loss caused by trauma or surgery. MVs obtained from DFPCs have been reported to activate the phospholipase C (PLC)/protein kinase C (PKC)/mitogenactivated protein kinase (MAPK) pathway in DFPCs to promote alveolar bone regeneration.⁹² Healthy PDLSC-EXOs can inhibit the overactivation of classical Wnt signal in an inflammatory environment to maintain the osteogenic capacity of endogenous stem cells and promote periodontal bone regeneration.93 SHED-EXOs have been shown to activate the BMP/Smad and Wnt/ β -catenin signaling pathways by transporting Wnt3a and BMP2 mRNA and proteins to recipient cells, effectively promoting osteogenic differentiation of PDLSCs in a dose-dependent manner,⁹⁴ and also promote periodontal bone regeneration by promoting neovascularization and new bone formation through the AMPK signaling pathway.⁹⁵ In addition, SHED-EXOs also promote bone formation and inhibit lipogenesis, which up-regulated the expression of Runx2 and p-Smad5 and decreased the expression of $Ppar\gamma$.⁹⁶ EV derived from inflammatory DPSCs (iDPSC-EVs) transport the limb development membrane protein 1 (LMBR1)-targeting miR-758-5p to promote PDLSCs osteogenic and odontogenic differentiation via BMP signaling.⁹⁷ The increased EXOs produced by TNF-a-pretreated GMSCs up-regulated the expression of CD73, which induced anti-inflammatory M2 macrophages and inhibit inflammation and osteoclast activity to prevent periodontal bone loss.⁹⁸ EVs are also often combined with materials for



Figure 3 Role of oral MSC-EVs in disease and tissue regeneration. EVs have a potential therapeutic role in the regeneration of tissue injuries such as bone, nerve, and skin soft tissue injuries, and in the inflammatory repair of immune-inflammatory diseases such as osteoarthritis, systemic lupus erythematosus, and periodontitis. Furthermore, EVs are associated with tumor cell proliferation, migration, and invasion, so they may have potential applications for tumor diagnosis and targeted therapy. This graph was created with Biorender.com.

bone tissue repair after injury. EVs derived from GMSCs and DPSCs combined with materials can improve the expression of osteogenic markers *RUNX2*, vascular endothelial growth factor (*VEGFA*), and osteopontin (*OPN*) and promote the repair of bone defects in rats.^{99,100}

The role of OMSC-EVs in tooth regeneration

SCAP-EXOs have been suggested to be a potential therapeutic approach for the regeneration of dentine-pulp complex and bone regeneration. SCAP generated during early tooth root development and SCAP-EXOs were found to promote the regeneration of dentine-pulp complex by inducing specific odontogenesis and osteogenesis by activating the mitogen-activated protein kinase (MAPK) signaling pathway.^{86,101} Furthermore, SCAP-EXOs can promote Rho-GTPase Cdc42-mediated angiogenesis, providing essential blood supply and nutrients to support tissue regeneration.¹⁰² In bioengineered tooth regeneration, EXOs derived from human DPSC (hDPSC) aggregates promoted the tooth regeneration process by up-regulating the odontogenic and angiogenic capacity of hDPSCs, leading to functional tooth regeneration, supporting ongoing root development in humans.¹⁰³ The regenerative effects of DPSC-EV on jawbone defects suggested that JB-MSC could efficiently uptake DPSC-EVs, which in turn significantly promoted the expression of osteogenic genes, such as runtrelated transcription factor 2 (RUNX2), alkaline phosphatase (ALP), and osteocalcin (OCN), and the osteogenic differen-tiation capacity of JB-MSCs.¹⁰⁴ Similar beneficial effects for bone regeneration were observed in rat calvarial bone defects.¹⁰⁰ ELVs-H1 can also be combined with biomaterials such as collagen to form a sustained release system, which is beneficial to prolong the biological activity of ELV and highly biocompatible ELV-H1 collagen provides a microenvironment suitable for odontoblast differentiation.⁸¹

The role of OMSC-EVs in soft tissue regeneration

The skin, as the first major defence barrier in the body, is prone to damage due to acute or chronic injuries after trauma, burns, or diabetes. Currently, studies on skin regeneration focus on accelerating wound healing after injury, and MSC-EVs have been considered to have good potential in repairing skin injury.¹⁰⁵ SCAP-EXOs promote the formation of Cdc42-dependent filaments in endothelial cells to promote cell migration by transporting Cdc42 to recipient cells, which then accelerate angiogenesis and wound healing of mouse palatal gingival soft tissue.¹⁰² HDPSC-EVs were also shown to have the ability to promote angiogenesis without being influenced by the periodontal inflammatory environment, providing a possible source of EVs that can be obtained from extracted teeth due to periodontitis.¹⁰⁶ Compared to skin wounds, gingival wounds heal faster and GMSCs have been found to rely on the Fas/ Fas-associated phosphatase-1 (Fap-1)/caveolin-1 (Cav-1) mechanism to generate EV with high expression of IL-1RA, leading to the promotion of the healing of gingival wounds in mice. Local application of GMSC-EVs with high expression of IL-1RA can also promote wound healing of the skin, further demonstrating the beneficial effect of GMSC-EVs on the repair of soft tissue injury.¹⁰⁷ Furthermore, chronic, delayed, or even non-healing wounds often occur in patients with systemic diseases such as diabetes due to high glycemic load, impaired angiogenesis, and unbalanced cytokine profiles, which can lead to the risk of gangrene, amputation, and even death. Fortunately, the combination treatment of GMSC-EXO with hydrogel shows the potential to address the problem, which has been identified to effectively promote skin wound healing in diabetic rats by promoting collagen reepithelialization, deposition, and remodeling.¹⁰⁸ These findings highlight the role of GMSC-EXOs in wound healing and provide a novel non-invasive method with practical value for skin repair using OMSC-EVs.

The role of OMSC-EVs in neural tissue regeneration

OMSCs appear to have a more prominent potential for neural regeneration due to their origin in the neural crest and the expression of some neural progenitor cells and mature cell markers. It is known to promote the repair and regeneration of nerve injuries, but the specific underlying mechanism has not been fully elucidated. Recent studies have shown that paracrine factors of OMSC, especially EVs, play an important role in nerve regeneration and have great therapeutic potential.¹⁰⁹ In a mouse model of sciatic nerve injury, GMSC-derived EVs (GMSC-EVs) up-regulate the protein level of C-Jun. Notch1, glial fibrillary acid protein (GFAP), and the sex-determined region Y-box 2 (SOX2), which are the characteristic genes for dedifferentiation, and promote the proliferation and migration of Schwann cells to promote regeneration and repair of injured nerves.¹¹⁰ Similarly, GMSC-EVs were also found to promote nerve regeneration and functional recovery by increasing the proliferation of rat dorsal root ganglion cells.¹¹¹ MSCs stimulated by inflammatory cytokines like TNF- α tend to secrete a large number of soluble mediators to exert their immunosuppressive potential. For instance, EXOs derived from TNF-a-stimulated GMSCs enhanced the protective effects of RGC and microglia and reduced neuroinflammation by inhibiting microglial activation through MEG3/miR-21a-5p axis.¹¹² SHED-EXOs can also reduce neuroinflammation by altering microglia polarization.¹¹³ Furthermore. SHED-EXOs grown on three-dimensional laminin-coated alginate microcarriers have been found to become a potential treatment for Parkinson's disease, which can inhibit the apoptosis of dopaminergic neurons to protect nerves.¹

The role of OMSC-EVs in immune disease

MSCs exert an immunomodulatory effect by interacting with the innate and adaptive immune systems. EVs, as one of the important paracrine factors of MSCs, play a crucial role in a variety of immune and inflammatory diseases. OMSC-EVs also exhibit good immunomodulatory properties, which play a role in immune and inflammation-related diseases. Compared to BMSCs and DPSCs-derived EXO immunomodulatory capacity, DPSC-EXOs show stronger inhibition of CD4⁺ T cell differentiation in Th17 cells to reduce the release of pro-inflammatory cytokines, while promoting CD4⁺ T cell differentiation in Tregs to increase the release of anti-inflammatory factors, thus exerting immunomodulatory function.⁷⁶ GMSC-EXOs can reduce the release of inflammatory cytokines, inhibit lipid accumulation, and promote the polarization of pro-inflammatory macrophages into anti-inflammatory phenotypes in a highlipid microenvironment.¹¹⁵ Therefore, OMSC-EVs have promise in the treatment of various inflammatory diseases.

MSC-derived secretomes also show great potential to treat systemic immune-inflammatory diseases as a novel alternative treatment. It has been shown that hPDLSCderived EXOs/MVs (hPDLSCs-EMVs) obtained from relapseretreat-multiple sclerosis patients or healthy donors have been shown to improve experimental autoimmune encephalomyelitis through anti-inflammatory and immunosuppressive effects and reverse disease progression by restoring tissue integrity through remyelination in the spinal cord.¹¹⁶ Studies have shown that intravenous injection of DPSC-EXOs can reduce cerebral edema, cerebral infarction, and nerve injury induced by cerebral ischemia/ reperfusion (I/R) in mice, which inhibits inflammatory responses through box 1 protein of the high mobility group (HMGB1)/toll-like receptor 4 (TLR4)/myeloid differentiation protein 88 (MyD88)/nuclear factor kappa B (NF- κ B) pathway to improve neuroinflammation after cerebral I/R injury.¹¹⁷ Sonoda S et al found that intravenous injection of SHED-EV could improve the phenotype similar to systemic lupus ervthematosus of MRL/lpr mice, which is achieved by improving the hematopoietic niche and immunomodulatory function of BMSCs to promote telomerase activity.¹¹⁸ EVs generated by miR-140-5p-overexpressed DPSCs can exert anti-apoptotic effects, which induce a significant reduction in IL-1_β-induced chondrocyte apoptosis to improve knee osteoarthritis in rat osteoarthritis. These results suggest that the combination of genetic engineering of DPSCs with transgenic DPSC-derived EVs can be a novel strategy for the treatment of osteoarthritis.¹¹⁹ Furthermore, DPSC-EXOs can inhibit the production of reactive oxygen species (ROS) and reduce the polarization of M1 macrophages through the ROS-MAPK-NF κ B-P65 signaling pathway, becoming a potential treatment for spinal cord injury.¹

In addition to systemic diseases, OMSC-EVs also have great therapeutic potential for oral inflammation-related diseases. In the rat pulp exposure model, DPSC-EVs, enriched with miR125a-3p, can promote the polarization of M2 macrophages through TLR and NF κ B signals to inhibit the pulpitis response. Furthermore, those EVs were found to promote macrophage release of BMP2 and activate the BMP pathway to facilitate the dentin formation of DPSCs.¹²¹ Some studies have proven that EVs derived from bacterial lipopolysaccharide (LPS) treated with DFPC also inhibit inflammation to promote periodontitis healing because they have an antioxidant effect and can inhibit intracellular ROS, reducing the ratio of the NF-B receptor activator of NFkB ligand (RANKL)/osteoprotegerin (OPG) of PDLSCs by inhibiting the ROS/c-Jun N-terminal kinase (JNK) signal under inflammatory conditions, and promote the polarization of M2-type macrophages through ROS/extracellular signal-regulated kinase (ERK) signaling.¹²² However, EVs produced by LPS-treated PDLSCs have been reported to encourage inflammatory progression by inducing M1-type macrophage polarization.⁷⁷ Unlike LPS-treated PDLSCs, healthy PDLSC-EVs can decrease NF-kB activity through the TLR4 signaling pathway to inhibit inflammation, suggesting that PDLSC-EVs may be used to treat chronic periodontitis.¹²³ PDLSC-EXOs also alleviate the inflammatory microenvironment through the Th17/Treg/miR-155-5p/SIRT1 regulatory network and enhance the ability of PDLSCs to promote angiogenesis by regulating EXO-mediated VEGFA transfer.¹²⁴ These results indicate that the biological composition and function of OMSC-EVs are closely related to the parental cells from which they are derived, which can be not only a potential therapeutic tool but also a potential therapeutic target for periodontitis to some extent. In a rat model of periodontal disease, GMSC-EVs also significantly improve the regeneration of damaged periodontal tissue by reducing pro-inflammatory cytokines secreted by monocytes/macrophages and T cells, which inhibit T cell activation and induce Treg formation.¹²⁵ In conclusion, OMSC-EVs can inhibit inflammation and promote disease recovery in a variety of ways, which is a potential treatment for diseases related to immune inflammation.

The role of OMSC-EVs in tumor progression and their therapeutic potential

EVs are nanoscale messengers and play a role in intercellular communication not only in normal tissues but also in tumor tissue, coordinating all kinds of autocrine and paracrine functions to change the tumor microenvironment, thus facilitating tumor growth and progression.^{126,127} Previous studies have demonstrated MSCs-EVs and their roles in the regulation of the tumor microenvironment and the potential application prospect in the treatment of tumors.¹²⁸ Cancer stem cells-derived EXOs (CSC-EXOs) also contain stem cell-specific proteins, regulatory miRNA, and survival factors that contribute to maintaining tumor heterogeneity and altering tumor progression.¹²⁹ Compared to MSCs-EVs that are derived from other tissues, OMSCs have unique neural crest origin and show many unique stem-like characteristics, 130 indicating that they have potential application value for tumor progression and treatment, although few studies on OMSC-EVs in tumors are available up to now.

Recent studies have shown that EVs derived from oral leukoplakia MSCs (LK-MSCs) and oral cancer MSCs (CA-MSCs) can transport miR-8485 to promote tumor cell proliferation, migration, and invasion instead of the normal oral mucosa (N-MSCs). However, inhibiting the release of EVs using GW4869, a commonly used exosome secretion inhibitor, attenuates the pro-tumoral effect of LK-MSC and EVs derived from CA-MSCs.¹³¹ Taken together, LK-MSC-derived EXO can be involved in the recurrence and malignant transformation of oral leukoplakia, and miR-8485 may be a potential marker of disease progression and a potential target for clinical intervention. Interestingly, due to their inherent ability to deliver biomolecules, EVs can be used as unique vectors for the intracellular transport of various therapeutic materials, such as miRNA and proteins, to cancer tissues.¹³² Compared to traditional liposome agents, EVs exhibit relatively satisfactory tissue tolerance and greater specificity for cancer targeting, and cancer cells can ingest and internalize more EVs than normal cells.¹³³ OMSC-EVs have good drug-loading capability and tumortargeting properties. For example, EXOs, which are derived from hDPSCs cultivated with gemcitabine (GCB), significantly inhibited the cell growth of pancreatic carcinoma cell lines in vitro.¹³⁴ OMSC-EVs also have the additional advantages of being less invasive, easy to access, and rapid to expand, which are favored in the treatment of tumors.¹³⁵ DPSCs are considered valuable sources for EV production due to their great potential for multidirectional differentiation and reproductive activity. MiR-34a is highly methylated in several cancers, leading to tumor cell apoptosis. Some researchers established DPSCs engineered with miR-34a and then obtained DPSC-EXOs that are abundant in miR-34a to evaluate the role of engineered EXOs in the treatment and prognosis of breast cancer. The results showed that DPSC-EXOs engineered with miR-34a enriched with miR-34a induced apoptosis of breast cancer cells in a dose-dependent manner, thus reducing the migration and invasion of cancer cells,¹³⁶ indicating the role of OMSC-EV in tumor development and migration, thus may provide a potential application value for tumor diagnosis and treatment.

Conclusions and perspectives

To date, EVs have become emerging mediators in cell-cell communication, having a variety of physiological functions, such as participating in intercellular information transport and regulating the physiological and pathological processes of distal cells in health and disease. Although increasing investigations have been conducted in the field of EV biology, the biological characteristics, molecular composition, function, targeting, and uptake mechanisms of EVs are still not completely elucidated to some extent. EVs have been emphasized to play a role in the transfer of biological molecules such as proteins, lipids, and nucleic acids, and are expected to be used as a new cell-free therapy in various diseases, such as immune diseases, bone and tooth regeneration, neurological diseases, wound healing, etc. Among all MSC-EVs, OMSC-EVs show great application prospects in many local and systemic diseases. Although there is not enough evidence to fully evaluate and compare specific molecular differences, biological functions, and therapeutic applications between OMSC-EVs and other different types of MSC-EVs, many studies revealed that OMSC-EVs had stronger immunomodulatory, antiapoptotic, and pro-regeneration characteristics compared to other MSC-EVs. However, it should be noted that a more accurate comparison between these studies in MSC-EVs is needed, as the methodology for the purification, characterization, and evaluation of the function of EVs varies greatly from study to study. Despite relatively few studies on OMSC-derived ApoEVs, the role of OMSC-derived EXOs and other EVs in local or systemic diseases has been widely studied, suggesting the potential role of OMSC-EVs in the development and treatment of diseases. There is no doubt that the research and application of OMSC-EVs still have limitations and unsolved problems that are similar to other studies of MSC-EVs at present, such as the lack of the gold standard for the separation and quantification of OMSC-EVs, and the standardized methods for the storage, transportation, and mass production of OMSC-EVs. Furthermore, the nomenclature of OMSC-EVs and the molecular and functional variation among EVs secreted from tissue-specific origin cells under normal and disease statutes are still unresolved questions. This is now being addressed by constantly updated guidelines on extraocular vesicle research, and the minimal requirements for studies of extracellular vesicles (MISEV).¹³⁷ Furthermore, the exploration of appropriate EV dose application methods, the proper treatment of their parental cells, and efficient EV purification methods are also needed to achieve the desired effect. Using further investigation of these questions, we hope to gain deeper insight into the biology of OMSC-EVs shortly to accelerate the transformation of OMSC-EVs into clinical applications in a rational way.

Conflict of interests

The authors declare that they have no conflict of interests.

Funding

This study was supported by the National Key R&D Program of China (No. 2022YFC2504200), the National Natural Science Foundation of China (No. 82270960), and the Science & Technology Development Talent Project of Jilin Financial Department, Jilin, China (No. JCSZ2021893-35) to AZ.

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