

Opportunities in Primary and Enteric Hyperoxaluria at the Cross-Roads Between the Clinic and Laboratory



Barbara Cellini¹, Michelle A. Baum², Yaacov Frishberg³, Jaap W. Groothoff⁴, Peter C. Harris⁵, Sally A. Hulton⁶, Felix Knauf⁷, John Knight⁸, John C. Lieske⁷, W. Todd Lowther⁹, Shabbir Moochhala¹⁰, Lama Nazzal¹¹, Gregory E. Tasian¹², Jonathan M. Whittamore¹³ and David J. Sas¹⁴

¹Department of Medicine and Surgery, University of Perugia, Perugia, Italy; ²Department of Nephrology, Boston Children's Hospital, Boston, Massachusetts, USA; ³Division of Pediatric Nephrology, Shaare Zedek Medical Center, Jerusalem, Israel; ⁴Department of Pediatric Nephrology, Emma Children's Hospital, Amsterdam UMC, Amsterdam, the Netherlands; ⁵Division of Nephrology and Hypertension and Department of Biochemistry and Molecular Biology, Mayo Clinic, Rochester, Minnesota, USA; ⁶Department of Nephrology, Birmingham Women's and Children's Hospital NHS Foundation Trust, Birmingham, UK; ⁷Division of Nephrology and Hypertension, Mayo Clinic, Rochester, Minnesota, USA; ⁸Department of Urology, Heersink School of Medicine, University of Alabama at Birmingham, Birmingham, Alabama, USA; ⁹Center for Structural Biology, Department of Biochemistry, Wake Forest University School of Medicine, Winston-Salem, North Carolina, USA; ¹⁰UCL Department of Renal Medicine, Royal Free Hospital, London, UK; ¹¹Nephrology Division, NYU Langone Health and NYU Grossman School of Medicine, New York, New York, USA; ¹²Division of Pediatric Urology, Children's Hospital of Philadelphia, Pennsylvania, USA; ¹³Charles and Jane Pak Center for Mineral Metabolism and Clinical Research UT Southwestern Medical Center, Dallas, Texas, USA; and ¹⁴Division of Pediatric Nephrology and Hypertension, Mayo Clinic Children's Center, Rochester, Minnesota, USA

Hyperoxaluria is a condition in which there is a pathologic abundance of oxalate in the urine through either hepatic overproduction (primary hyperoxaluria [PH]) or excessive enteric absorption of dietary oxalate (enteric hyperoxaluria [EH]). Severity can vary with the most severe forms causing kidney failure and extrarenal manifestations. To address the current challenges and innovations in hyperoxaluria, the 14th International Hyperoxaluria Workshop convened in Perugia, Italy, bringing together international experts for focused presentation and discussion. The objective of the following report was to disseminate an overview of the proceedings and provide substrate for further thought. The format of this paper follows the format of the meeting, addressing, "PH type 1" (PH1) first, followed by "surgery, genetics, and ethics in PH", then "PH types 2 and 3," (PH2 and PH3) and, finally, "EH." Each session began with presentations of the current clinical challenges, followed by discussion of the latest advances in basic and translational research, and concluded with interactive discussions about prioritizing the future of research in the field to best serve the need of the patients.

Kidney Int Rep (2024) **9,** 3083–3096; https://doi.org/10.1016/j.ekir.2024.08.031 © 2024 International Society of Nephrology. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

yperoxaluria is a pathologic condition characterized by increased urinary excretion of oxalate, an end-metabolic product in humans, leading to the formation of the poorly-soluble calcium oxalate (CaOx) that precipitates as stones and/or nephrocalcinosis in the kidneys. The 2 main forms of hyperoxaluria, PH and EH, are characterized by a different origin of

Correspondence: Barbara Cellini, Department of Medicine and Surgery, University of Perugia, P.le Severi 1, 06132, Perugia, Italy. E-mail: barbara.cellini@unipg.it; or David J. Sas, Division of Pediatric Nephrology and Hypertension, Mayo Clinic Children's Center, 200 First St. SW Rochester, Minnesota 55905, USA. E-mail: sas.david@mayo.edu

Received 2 July 2024; accepted 26 August 2024; published online 1 September 2024

oxalate (Figure 1). In PH, increased endogenous production of oxalate occurs because of inherited mutations leading to the deficit of enzymes involved in the hepatic metabolism of glyoxylate, the direct precursor of oxalate through lactate dehydrogenase (LDH). The deficit of peroxisomal alanine:glyoxylate aminotransferase (AGT) causes PH1, 3-5 whereas the deficit of cytosolic/mitochondrial glyoxylate hydroxypyruvate reductase (GR/HPR) causes PH2. The pathogenesis of PH3 is still puzzling because it is genetically caused by pathogenic variants leading to a functional deficit of 4-hydroxy-2-oxoglutarate aldolase (HOGA), an enzyme that generates glyoxylate from hydroxyproline. Various hypotheses have been formulated to explain increased urinary oxalate excretion in PH3,

including the possible inhibitory effect of accumulating 4-hydroxy-2-oxoglutarate on GR/HPR, or the cleavage of 4-hydroxy-2-oxoglutarate by a still-unknown cytosolic aldolase. ^{9,10}

Because dietary oxalate accounts for a significant portion of urinary oxalate, excessive intake of oxalaterich foods can lead to hyperoxaluria A distinct form of oxalate hyperabsorption by the large bowel known as EH is a frequent complication of intestinal fat malabsorption following small intestinal resection or gastric bypass surgery or medical conditions such as inflammatory bowel disease or cystic fibrosis. 12,13 Patients with EH are at increased risk of nephrolithiasis and nephrocalcinosis, possibly leading to chronic kidney disease and end-stage kidney disease (ESKD). 14

Current therapies for all forms of hyperoxaluria aim at reducing CaOx supersaturation through high fluid intake and crystallization inhibitors, or, in the most severe forms, dialysis and kidney transplant. For patients with PH1, other treatment options are available. A subset of them is responsive to vitamin B6 (pyridoxine), which can reduce oxalate production. Traditionally, combined liver-kidney transplantation was needed to address kidney failure and replace the oxalate-producing liver. More recently, therapies based on RNA interference (RNAi) have been developed to reduce hepatic oxalate production in PH and,

though these biological drugs appear to be quite effective, they are not available for all patients and their long-term risks are not known. 18-21

The 14th International Hyperoxaluria Workshop took place on June 23 and 24, 2023 in Perugia, Italy, organized by the Oxalosis and Hyperoxaluria Foundation, with participation of an international group of the primary experts in the field, as well as representatives from patient advocacy groups. The conference focused on the most critical clinical and ethical challenges currently facing patient management in PH and EH, and the latest innovations offered by basic and translational research. This narrative review summarizes the highlights of the lectures through the eyes of the chairs of each session of the workshop. The detailed meeting program can be accessed at https://ohf.org/14th_international_workshop.

PH1 Clinical Update

PH1 is the most prevalent and severe type of PH. Over 200 pathogenic variants have been found in the AGT gene, most of them leading to an overall AGT-deficiency, but some only to AGT mistargeting to mitochondria where the enzyme does not detoxify glyoxylate. PH1 may cause recurrent kidney stones, nephrocalcinosis, and kidney failure in over 60% of patients. As kidney function declines, oxalate

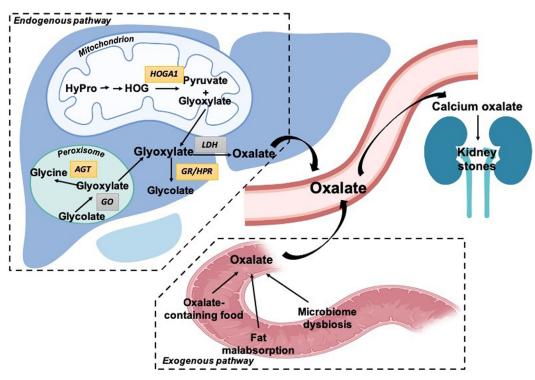


Figure 1. Overview of the pathways controlling oxalate homeostasis. AGT, alanine:glyoxylate aminotransferase; GO, glycolate oxidase; GR/HPR, glyoxylate hydroxypyruvate reductase; HOG, 4-hydroxy-2-oxoglutarate; HOGA1, HOG aldolase; HyPro, hydroxyproline; LDH, lactate dehydrogenase.

accumulates in the blood and is deposited in various tissues, leading to life-threatening condition referred to as systemic oxalosis. Unfortunately, even aggressive dialysis cannot keep up with oxalate generation, often resulting in ongoing systemic oxalosis. ²⁴ For non-pyridoxine responsive patients with kidney failure, combined liver-kidney transplantation was the only therapeutic option before RNAi therapies. ²⁵⁻²⁷

Two new drugs based on RNAi, lumasiran and nedosiran, have been shown to be safe and effective in clinical trials. In 2020, lumasiran was approved by both the US Food and Drug Administration and the European Medicines Agency as therapy for PH1.28 Nedosiran was approved by the US Food and Drug Administration for patients with PH1 aged >9 years with estimated glomerular filtration rate (GFR) >30 ml/ min per 1.73 m² in Fall 2023.²⁰ Both drugs aim to reduce endogenous oxalate production by gene silencing of enzymes involved in glyoxylate metabolism. Both are administrated subcutaneously at prescribed intervals and require lifelong treatment. Lumasiran blocks glycolate oxidase (GO), which promotes the conversion of glycolate into the oxalate precursor, glyoxylate. This effect results in an average reduction of about 65% in urine oxalate excretion from baseline. 26,28,29 The 12-month outcome studies have shown promising results with decreased kidney stone events and nephrocalcinosis, along with preservation of GFR over time, 30 with no significant side effects. Several case reports have described decrease in dialysis time or number of days, ability to stop dialysis in some patients, and ability to perform isolated renal transplant in patients due to significant reduction in plasma oxalate levels with lumasiran treatment.31-33

Nedosiran blocks LDHA, which converts glyoxylate to oxalate. Studies in patients with PH1 show similar effectiveness and safety as lumasiran. In the PHYOX2 randomized controlled study, nearly 65% had normal or nearly normal urinary oxalate levels ($<1.3 \times$ upper limit of normal) after 6 months of treatment with nedosiran. A case report demonstrated response of plasma oxalate in a patient with PH1 on dialysis, and successful kidney transplantation while receiving nedosiran.

The following unanswered questions with respect to RNAi therapy in PH1 were discussed:

 Should RNAi treatments be given to all patients with PH1 or only to those who do not fully respond to pyridoxine and have clinically severe disease? There are geographical differences on this point. In Europe, the level of recommendation differs based on severity of disease and pyridoxine-responsiveness,²⁵ whereas in the US, RNAi treatment is prescribed for any

- patient with PH1 who is not fully pyridoxine-sensitive. The scientific community agrees that in case of full pyridoxine responsiveness with normalization of urine oxalate, there is likely limited benefit of RNAi. There is debate on the indication in patients who do not fully respond to pyridoxine, but still have milder disease. The arguments for restrictive application are cost, unknown long-term effects, and unnecessary addition of a drug in fully pyridoxine-responsive patients or patients with minimal or no clinical symptoms. Arguments supporting a low threshold for RNAi prescription, also in patients with minimal or no clinical symptoms, include the unpredictable course of PH1 and the risk for sudden deterioration resulting in ESKD, as well as the prevention of lifelong treatment of ESKD such as dialysis and/or renal transplantation. In addition, long-term outcome data are warranted to fully appreciate the exact impact of RNAi on clinical outcome with respect to kidney function preservation and reduction of stone events.
- 2. Are RNAi treatments safe in pregnancy? Animal studies with high dose lumasiran showed no fetal abnormalities but no human pregnant patients with lumasiran maintenance therapy have been reported so far. Data on all pregnant patients with PH should be reported to determine outcomes. Indeed, some women may choose to discontinue therapy during pregnancy versus continue RNAi treatment; and these outcomes and safety information should also be documented.
- 3. Can RNAi therapy replace liver transplantation? The prospects for isolated kidney transplant in future patients on RNAi seem promising, but thus far only case reports are available. Most centers are performing isolated renal transplant in patients who have demonstrated significant reduction in urine or plasma oxalate levels on lumasiran. There is optimism that avoiding liver transplantation may be a viable option in those who have sufficient reduction in urine or plasma oxalate and long-term data are being collected.
- 4. What would be an acceptable plasma oxalate concentration in patients with PH1 on dialysis for isolated kidney transplantation? There is insufficient evidence to answer that question; however, there are some indications from case reports of kidney transplantation performed in patients treated with RNAi, as well as from data reported on plasma oxalate levels in non-PH patients on dialysis. Future data on kidney transplant in patients receiving RNAi treatment are required to answer emerging questions, including determination of the target plasma oxalate concentration at which this approach is safe.

- 5. Should RNAi therapy be individualized based on extent of enzyme deficit? A study with stable isotopes showed interpatient variability from 55% to over 90% in GO blockade from lumasiran. No data exist on increasing dosage or dosing frequency in patients to improve response to lumasiran. Potential individualized combination therapy (lumasiran and nedosiran, or other potential future treatments) could be considered for patients who do not respond sufficiently to a single agent. This requires further investigation.
- 6. Is there a need for native nephrectomy prior to renal transplant? For patients with long-term PH1, kidneys may be a source of deposited oxalate that can be released back into the bloodstream, causing ongoing oxalate burden to the newly transplanted kidney, though there are no published data on this. Until investigation is performed to determine if there is a benefit to performing native nephrectomy, practice will continue to vary center to center.

Basic Science Research Update

The interest of the scientific community in PH1 has increased in the past decade related to interpretation of genetic data for a correct diagnosis, the design of new therapeutic strategies, and the assessment of patients' responsiveness using appropriate endpoints. Indeed, the reduced costs and turn-around times for genetic sequencing have brought out a new aspect of the diagnosis of rare diseases, that is, the assessment of the pathogenicity of newly identified variants. This aspect is of fundamental importance in PH1, because most disease-causing variants are missense and can lead to the AGT-deficit through molecular mechanisms spanning from a merely functional or structural defect to a combination of the 2.3,22,39 In addition, as recently reported, the assessment of the minor or major haplotype in which a variant is inherited could remarkably affect disease manifestations and responsiveness to treatments. 25,40,41 Indeed, the AGT minor haplotype characterized by p.Prol1Leu and p.Ile340Met substitutions, can have broad effects on AGT fitness. Interestingly, the 2 variants act in different directions, where the introduction of Leull was destabilizing but Met340 stabilizing. Some variants can be pathogenic on both the major and minor haplotype, such as p.Ile56Asn, but the specific haplotype may further influence protein fitness and therefore be a major modifier of the clinical presentation or progression. 42 It follows that the AGT haplotype should be taken into consideration when assessing pathogenicity of likely penetrance of variants.

Improved understanding of PH1 pathogenesis has offered great opportunities for the design of new therapeutic approaches. Utilizing a small-molecule

approach, which may be of lower cost and more easily administered, progress has been made in reducing plasma oxalate by targeting LDH or GO, as well as by molecules leading to a combined inhibition of both proteins. 43-47 Some of these molecules have been already tested in clinical trials (www. clinicaltrials.gov, identifier NCT05367661). Moreover, efficient pharmacological chaperones to better rescue misfolded AGT and inhibitors of CaOx crystallization have been described. 48-50 Regarding biologicals, studies in AGT gene $1^{-/-}$ mice show targeting to hepatocytes, expression of AGT, reduction of oxalate levels, and prevention of nephrocalcinosis with specific doses.⁴⁸ Gene editing approaches are also on the horizon for PH1, aimed at reducing substrate load, by specific knock-out of LDH or GO. 51-53 A gene editing approach to correct the AGT gene pathogenic variant in PH1 patient-derived fibroblasts has been also proposed.⁵⁴ Patient fibroblasts were differentiated to hepatocytes by targeted reprograming, and using CRISPR-Cas9, the point defect within the gene was either corrected or a normal copy of the AGT gene was knocked into the endogenous locus. AGT expression was documented in the corrected cells, showing the proof of principle of this approach.

Another very active point of discussion in PH1 is the setup of suitable methods to monitor disease progression as well as treatment responsiveness. Until now, most of the studies relied on 24-hour urinary oxalate excretion. However, the latter does not represent an accurate measure for endogenous oxalate production in patients with PH and it is not applicable in those with anuria. Therefore, a gas chromatography tandem mass spectrometry analysis method following a stable isotope infusion protocol of ¹³C₂-oxalate and ¹³C₁-glycolate has been implemented. Using this method, the efficacy of lumasiran, which silences GO, was shown to vary between patients, likely reflecting different levels of drug effectiveness. ⁵⁷

Overall, the field of translational research in PH1 is living a particularly exciting moment, especially regarding approaches to test variant pathogenicity and modifying effects, monitoring the efficacy of existing treatments, and potential new treatments in the form of small molecules, transgenic expression, and pathogenic variant correction by gene therapy. Over the next few years these novel approaches are likely to be finding their way into the PH clinic.

Surgery, Genetics, and Ethics in PH Surgery

Endourological techniques for kidney stone removal continue to advance. Current laser and ultrasonic lithotripsy devices allow many stones to be fragmented and removed through ureteroscopic techniques.⁵⁸ Other larger stones may require percutaneous access for stone fragmentation and removal. Nevertheless, this is still much less invasive than open surgery which is now very rarely required. In addition, modern endourological techniques for the most part do not damage the kidneys; and chronic kidney disease strictly related to repeated surgery has become less of a concern. One major patient concern is, however, pain related to ureteral stents which are typically placed either before or after surgery or both to prevent ureteral obstruction secondary to stones or fragments of stones. To date, there are no consistently effective strategies to prevent pain and lower genitourinary tract symptoms when stents are in place.

One major surgery-related concern for patients with PH is the increased risk for acute oxalate nephropathy resulting from the procedure. To some extent, all patients can be susceptible to acute kidney injury secondary to oxalate during times of volume depletion or other factors that result in decreased renal perfusion. Thus, even during a routine kidney stone surgery, a patient with PH can develop acute kidney injury that is variably reversible if the surgery is complicated by hypotension, even when the hypotension is subtle. Patients with PH and diminished GFR are at even greater risk of acute kidney injury in the perioperative period.

Similar risk for acute kidney injury from acute oxalate nephropathy exists after kidney transplant, which can occur with either a liver or kidney transplant or combined liver-kidney transplant, as oxalate either already in the bloodstream or from tissue stores is excreted through the new allograft.⁵⁹ Thus, centers must be aware of this possibility and often aggressive dialysis before and after transplant is performed. It is very important that these procedures be performed in centers that are aware of these concerns and prepared to have access to appropriate dialysis and laboratory support. Patients who have had kidney failure for a significant period often have massive oxalate deposition in the native kidneys, prompting some centers to perform native nephrectomy at the time of transplant to remove this potential source of ongoing oxalate excretion. As discussed earlier, because there is only anecdotal evidence for this, controversy persists.

Genetics

Knowledge regarding the true prevalence of the 3 forms of PH is evolving as more genetic data are being generated worldwide, and as more novel variants have been confirmed in individual families and patients. Confirming pathogenicity remains potentially challenging, especially for missense changes and when

large informative families with strong phenotypic data are not available. Newer laboratory techniques to look at expressed protein in cultured cells and enzyme activity assays in vitro can help to demonstrate likely pathogenicity of a novel variant. Based on these evolving data, current estimates suggest that the frequency of biallelic cases worldwide could approximate 1:150,000 for PH1, 1:300,000 for PH2, and 1:135,000 for PH3, with estimated carrier frequencies of $\sim 1:200$, \sim 1:280, and \sim 1:185, respectively.²³ Thus, the prevalence of PH3 may exceed PH1 and PH2 remains the least common. Often patients present with risk factors for monogenic causes of kidney stone disease, including frequent stones at an early age, nephrocalcinosis, and kidney failure. However, reliably measuring biochemical data may be challenging where access to advanced laboratory testing is limited. Genetic testing with kidney stone or nephrocalcinosis candidate gene panels provide a method for rapid identification of monogenic causes, and current evidence indicates >40 monogenic causes of kidney stone disease, including PH1, PH2, and PH3.60-62 Due to overlapping clinical features, panel screening is recommended even when the diagnosis seems obvious. For example, 10% of those with a strong PH phenotype do not have biallelic variants in a PH gene. 60

Because PH is a biallelic disorder, worldwide prevalence varies widely among geographic and ethnic groupings. For example, whereas worldwide rates of consanguinity are estimated to be 10%, rates of consanguinity are highest in the Middle East, ranging from 35% to 80%, and less than 4% in Europe and the USA.63 A large national study in the UK showed that the risk of baby being born with a recessive disorder doubled for consanguineous couples.⁶⁴ Thus, consanguinity can be regarded as a modifiable risk factor for autosomal recessive diseases including PH. Additional challenges identifying cases of PH arise in countries that do not include PH in their disease-tracking databases. Finally, Deesker and colleagues highlight that underlying genetic causes are not the only factor in PH disease severity, showing that younger siblings of index cases often have milder courses despite having the same genetic variants for PH.65

Ethics

Access to genetic testing and reliable urinary and plasma oxalate measurement to make an accurate diagnosis is quite variable around the world. Similar disparities to access exist for state-of-the-art treatments such as intensive dialysis, organ transplant, and RNAi therapies. These disparities will only increase with the development of innovative and very expensive treatment strategies involving small molecules and gene

editing. Access to RNAi treatment is limited in many countries by the governing agencies due to its high price. Most low- or middle-income, and even some European countries, have no access at all. Beyond cost, limited access to RNAi treatments are also exacerbated by the requirement for ultra-low temperatures during shipping and storage. Strategies to overcome these health disparities remain an important challenge in the hyperoxaluria community. Patient organizations may be in a good position to shine a light on these issues and mobilize caregiver and industry partners.

PH2 and PH3 Clinical Update

PH2 is caused by a deficiency of the enzyme GR/ HPR^{66,67} encoded by GRHPR.⁶⁸ GR/HPR activity is highest in the liver but also present in leukocytes and kidneys. 69 Lack of GR/HPR leads to an accumulation of glyoxylate and hydroxypyruvate, both metabolized by LDH to oxalate and L-glycerate, respectively. A retrospective review of genetically confirmed PH2 cases in the OxalEurope registry was recently published and included 112 patients from 11 countries, the majority residing in the UK. 70 Clinical characteristics of PH2 are like those of PH1. The median age of first symptom and diagnosis duplicate those reported for PH1 (3.2/9 years vs. 3.9/8.1 years), with urolithiasis noted in the majority.²³ Nephrocalcinosis is less common than in PH1 but can occur in all age groups.⁷¹ Presentation of symptoms in infancy is as common as in PH1. In contrast, patients with PH2 are less likely to present in kidney failure at diagnosis, with no progression to ESKD recorded prior to the age of 15 years, unlike PH1. Renal survival and the overall long-term prognosis for patients with PH2 appears similar in outcome to the pyridoxine-sensitive PH1 genotypes. Genetic analysis revealed 18 novel mutations in the GRHPR gene. 70 Chronic Kidney Disease stage 2 or worse was recorded in 45 of 89 (50.6%), and 22 patients (24.7%) had reached stage 5. Median renal survival was 43.3 years and 15 transplants in 11 patients were recorded. Renal outcome did not correlate with genotype, biochemical parameters, or with the presence of nephrocalcinosis at presentation. Thus, PH2 is a disease with significant morbidity and accurate diagnosis by 24-hour urine analysis and careful follow-up is required.

The supportive management is as for PH1, but with no subset responsive to pyridoxine. High fluid intake and citrate are the mainstay. Although the risk for systemic oxalosis has traditionally been considered lower in patients with PH2, the increasing documentation of renal impairment places this group of patients in a category like PH1 where risks for systemic oxalosis increase as GFR falls.

The clinical practice recommendations for PH2 suggest that liver transplantation may be advocated in those with advanced disease (estimated GFR <30 ml/min/SA).²⁵ The 1-year and 5-year cumulative kidney allograft survival were 43% and 29%, respectively; and 3 patients underwent a repeat kidney transplant (at 5, 6, and 22 years). Successful liver-kidney transplantation in patients with PH2 has been reported in several cases.⁷²⁻⁷⁴

Nedosiran treatment reported from the PHYOX2 trial demonstrated no statistically relevant reduction in urine oxalate in the small number of patients with PH2 enrolled, ³⁴ though investigations into its potential effectiveness in patients with PH2 and PH3 are ongoing. Future therapeutic strategies may require a personalized approach, consideration of other medications (e.g., stiripentol as an LDH inhibitor ³²), and development of better biomarkers to inform disease progression.

PH3 was described in 2010 and appears to be the mildest form of PH in terms of risk for ESKD, though patients tend to present at a young age and can have abundant stone formation.⁷⁵ It is caused by pathogenic variants in HOGA1 encoding the mitochondrial enzyme HOGA. The exact mechanism through which this defect leads to increased oxalate production has not been clearly elucidated. It is likely that PH3 is more prevalent than PH2 but is newer and perhaps underdiagnosed due to the milder phenotype. Like PH2, because there is no effective targeted therapy, treatment consists of hyperhydration and crystal inhibition. In 1 report, 97% of patients with PH3 had intact GFR at age 40 years.⁷⁶ Few cases of PH3 resulting in ESKD exist and involve patients who required multiple surgical interventions, which could have contributed to their GFR decline. 23,77 For both PH2 and PH3, there is a significant unmet need for a safe and effective treatment.

Basic Science Research Update

The main research topics related to PH2 and PH3 dealt with the knowledge gaps in endogenous oxalate synthesis in humans and how it relates to the pathogenesis of the 2 diseases, particularly the role of hydroxyproline catabolism. Indeed, the liver is a major site of glyoxylate synthesis from hydroxyproline, glycolate, and glycine metabolism, ⁷⁸⁻⁸² and is therefore an important organ to target to reduce endogenous oxalate synthesis in all types of PH.

Common to both PH2 and PH3 is the central role of hydroxyproline catabolism to oxalate synthesis. Fargue and colleagues continuously infused i.v. carbon-13 hydroxyproline in adults with PH2 and PH3 and demonstrated that hydroxyproline metabolism

contributes 47% (PH2) and 33% (PH3) to urinary oxalate excretion.⁸¹ Bone turnover,⁸³ exercise,⁸⁴ dietary collagen intake, 79 and renal function 85 have all been shown to influence hydroxyproline levels in blood or urine. However, the impact of these factors on urinary oxalate excretion in those afflicted with PH2 or PH3 warrants further investigation. Nonetheless, inhibition of the hydroxyproline degradation pathway could have a significant impact on lowering urinary oxalate excretion in PH2 and PH3. In this regard, a key opportunity is offered by the possible use of hydroxyproline dehydrogenase inhibitors. Hydroxyproline dehydrogenase is the first enzyme in the hydroxyproline catabolic pathway and represents a promising target because humans⁸⁶ and mice⁸⁷ with hydroxyproline dehydrogenase deficiency is healthy. A recent report demonstrated that the inhibitor N-propargylglycine prevents 4-hydroxyproline catabolism in mouse liver and kidney. 88 However, this compound also interferes with proline metabolism, highlighting that agents targeting hydroxyproline catabolism must also be evaluated for potential side effects. Furthermore, hydroxyproline metabolism occurs in kidneys and thus blockage of hydroxyproline dehydrogenase activity at both sites may be needed for optimal reduction of urinary oxalate excretion.

Another strategy currently pursued to reduce oxalate synthesis in PH2 and PH3 is the inhibition of LDH. Approaches to reduce LDH function have focused on developing agents that only target this enzyme in the liver, because systemic LDH inhibition can result in serious side effects. ⁸⁹ Nedosiran, a siRNA that specifically knocks down expression of hepatic LDHA, is a US Food and Drug Administration-approved drug to reduce urinary oxalate excretion in PH1. However, it demonstrated a mixed efficacy in reducing urinary oxalate excretion in patients affected by PH2 and PH3. ^{34,90} This further underscores the need for a better understanding of endogenous oxalate synthesis in PH2 and PH3.

The role of non-hepatic tissues in endogenous oxalate synthesis is also an important topic of discussion within the scientific community. Indeed, the expanded tissue expression of GRHPR and HOGA1 suggests that the kidney may also contribute, although future work is needed to define its role as a premise to novel strategies for reducing urinary oxalate excretion in PH2/3.

Specific to PH3, challenges to our understanding include the involvement of HOGA in the catabolism of hydroxyproline, with this enzyme generating glyoxylate rather than detoxifying it.^{7,91,92} Moreover, the enzyme exhibits oxaloacetate decarboxylase activity suggesting it may have a wider role in metabolism,⁹³ whose impact on glyoxylate detoxification and/or

oxalate synthesis remains unknown. Hypotheses to explain the mechanism(s) by which oxalate synthesis is increased in PH3 may include the inhibition of GR/HPR by 4-hydroxy-2-oxoglutarate, breakdown of 4-hydroxy-2-oxoglutarate to glyoxylate by an alternative aldolase or aldolase(s), and an alteration in cellular redox balance resulting in increased glyoxylate oxidation to oxalate by LDH. However, the relative role of each of these potential mechanisms in oxalate synthesis in PH3 requires further investigation.

EH Clinical Update

EH is the abnormally high excretion of oxalate in urine due to fat malabsorption by the small intestine, which leads to excess oxalate absorption by the colon. Normal healthy adults typically excrete <40 mg (<0.5 mmol) of oxalate daily. Idiopathic CaOx stone-formers may have excretion rates up to 50 mg/d (0.5 mmol/d), whereas urine oxalate can range between 50 and 100 mg/d (0.5-1.0 mmol/d) for patients with EH. Conditions frequently associated with EH include bariatric surgery, specifically Roux-en-Y gastric bypass, short gut syndrome, Crohn's disease, cystic fibrosis with pancreatic insufficiency, and more rarely, celiac disease. 94 Notably, restrictive gastric bypass procedures (i.e., gastric banding), have not been associated with the development of hyperoxaluria or kidney stones. 95,96 If possible, the first line of intervention for EH should be correction of the underlying bowel disease. Adjunctive treatments can seek to reduce oxalate absorption through dietary manipulations to decrease the bioavailability of oxalate or limit calcium sequestration by fatty acids within the large intestine. These include a low-oxalate diet, but this is challenging to sustain and with mixed evidence of efficacy. 13 Restricting dietary fat consumption and maintaining sufficient calcium intake to maximize the opportunity for calcium and oxalate to bind within the gut thereby limiting absorption has long been recognized to help reduce urinary oxalate. 97 Similarly, some (but not all) older studies demonstrated that bile acid sequestrants such as cholestyramine could also decrease urinary oxalate excretion. 12 There is, however, a paucity of contemporary data for these approaches with patients with EH who have undergone modern bariatric procedures.98

Surgical treatment options for patients with EH with symptomatic kidney stones are much the same as those for idiopathic kidney stone disease and thus have similar challenges. Recent advances in endourology equipment, including single use ureteroscopes and high-power lasers offer technologic advantages and increased efficiency. ⁹⁹ In addition, the adoption of predictive models increasingly allows urologists to

select the appropriate intervention and power source to remove stones with greater effectiveness. ¹⁰⁰

Basic Science Research Update

Research efforts on oxalate handling by the gut have been focused along 2 main lines: oxalate degradation by commensal microbes and oxalate transport across the intestinal epithelium. Concerning the former, it is known that the gut microbiome has an essential role in human health, and its disruption has been implicated in the pathophysiology of a wide variety of diseases including kidney stones. 101-104 The presence of oxalate-degrading gut bacteria is associated with lower rates of urinary oxalate excretion which can help guard against hyperoxaluria and reduce the risk for CaOx stone formation. 105 Several of these bacteria have been identified although only a few are active in vivo, perhaps the best known of these is Oxalobacter formigenes. 106,107 Differences have been noted between the microbiome of stone formers compared to non-stone formers, including a lower prevalence of O. formigenes. 107,108 The rat Roux-en-Y gastric bypass model of EH could be robustly colonized with O. formigenes leading to normalized urinary oxalate excretion. Oxalate-degrading probiotics such as O. formigenes hold promise as a potential approach for treating EH. Although clinical trials with patients with PH and O. formigenes yielded conflicting results, 109-112 no such trials have been attempted for patients with EH. An alternative strategy to probiotic administration is orally available exogenous oxalate-degrading enzymes of microbial origin, such as reloxaliase (Allena Pharmaceuticals) or OX-1 (Oxidien Pharmaceuticals), whose safety and efficacy in heathy subjects 113,114 and patients with EH 115 has been investigated. It is also worth mentioning that though many studies have focused on the most abundant bacterial components of the microbiota, other less abundant microbes may play a role in oxalate handling. In this regard, commensal fungi may reveal a previously unrecognized role within the microbiota in oxalate degradation. 116

In addition to the gut microbiome, the relatively recent discovery that the urinary tract is not sterile, even in the absence of infection, has launched a new line of research into how this previously unrealized microbiome influences kidney stone formation and urine composition. Improvements in culture-based techniques, together with advances in "-omic" approaches, are allowing investigators to explore this "new" microbiome. However, given its low biomass, rigorous control of potential contamination during sample collection, processing, and analysis is paramount. So far, sequencing studies indicate that distinct microbial communities exist in the urinary tract of CaOx stone-formers compared with controls.

Over 3500 microbial genes were more highly expressed in non-stone formers and 19 % of these (including oxalate-degrading genes) bacteria such as Lactobacillus crispatus may therefore have important implications for hyperoxaluria and potential stone risk.

Oxalate transport in the intestine has also received considerable attention. Indeed, understanding how oxalate is absorbed across the intestinal epithelium is important for developing effective strategies to limit excess uptake by the colon in EH. The current model proposes that the absorption mechanism is entirely passive and paracellular along the length of the intestine. 120 There is, however, evidence from rodent models of a significant transcellular component to absorption by the large intestine involving the apical anion exchanger, Slc26a3. 121 Investigations of the mechanistic basis of EH with the Roux-en-Y gastric bypass rat model have emphasized how dietary fat increases luminal oxalate solubility and enhances epithelial permeability, which are predicted to drive paracellular absorption. 122 In this setting, any role for colonic Slc26a3 is likely to be insignificant. Studies on other Slc26a family members may provide further insights, including Slc26a1, recently characterized for its major role in sulfate homeostasis, but potentially able to contribute to oxalate influx. 123

The progress in our understanding of oxalate handling in the intestine toward the development of novel therapeutic strategies relies on the parallel refinement of animal models. In this regard, after gastric bypass, another prevalent cause for EH is Crohn's disease; however, suitable animal models resembling the human pathology of spontaneous ileal inflammation are few. A promising candidate is the SAMP1/YitFc (SAMP1) mouse model. 124 SAMP1 mice displayed clear signs of fat malabsorption and urinary oxalate excretion increased markedly with dietary fat content. Expression levels of tight junction proteins were altered in the intestine of SAMP1 mice suggesting enhanced gut permeability, as demonstrated by the Roux-en-Y gastric bypass rat model. 107 In addition, the SAMP1 microbiome was enriched with several oxalate degraders, indicating a possible adaptation to increased oxalate levels within the large intestine. Continued study of these potential contributors to the pathophysiology of EH will be crucial for the development of effective therapies.

CONCLUSION

The Oxalosis and Hyperoxaluria Foundation-sponsored 14th International Hyperoxaluria Workshop hosted in Perugia, Italy in 2023 brought together the world's experts to discuss the current challenges in managing patients with PH and EH; and visualized a path

forward for investigators to answer the most pressing questions to facilitate innovation in research and improved clinical care. The format allowed for open, honest, and respectful discussion between physicians, clinical investigators, and basic scientists to work toward this common goal.

As our coauthors and session chairs have outlined above, great work continues to be performed expanding on the strong foundation of dedicated predecessors. For PH1, we celebrate the victory of game-changing treatment with RNAi medications but acknowledge that many new questions have arisen that must be answered, all while newer treatment modalities evolve through various stages of development. For PH2 and PH3, patients continue to struggle without definitive treatment, highlighting the crucial need to improve our understanding of the underlying mechanisms of oxalate production in these 2 conditions. Similarly, EH warrants further study because elucidating the details of oxalate transport in the gut and the role of the microbiome may lead to targeted therapies to reduce the risk of hyperoxaluria in these patients. Finally, we acknowledged that all the hard work being performed in the field must include the voices of the patients and include an ethical perspective to maximize equity and inclusion.

It is crucial for conferences such as this to continue to facilitate progress in the management and research of rare diseases. Dedicated, focused time together with diverse experts and stakeholders sharing physical space and ideas results in efficient communication in real time providing opportunities for rich discussions that cannot be replicated in any other way. We are grateful for the opportunity and express great appreciation for the participation of all attendees.

DISCLOSURE

BC has received research funding from Novo Nordisk Pharmaceuticals. MAB has received research funding from Alnylam Pharmaceuticals and Novo Nordisk Pharmaceuticals as well as consulting fees from Alnylam Pharmaceuticals, Novo Nordisk, Cantero Therapeutics, and Chinook Therapeutics. YF has received consulting fees from Alnylam Pharmaceuticals and Arbor Biotechnology. JWG has received research funding and consulting fees from Alnylam Pharmaceuticals and Novo Nordisk Pharmaceuticals. SAH has received consulting fees from Alnylam Pharmaceuticals, Pharmaceuticals, and Arbor Biotechnology. FK has received research funding from Alnylam Pharmaceuticals and Novo Nordisk Pharmaceuticals as well as consulting fees Allena Pharmaceuticals, Oxthera Pharmaceuticals, Alnylam Pharmaceutical, Chinook Pharmaceuticals, Zai Pharmaceuticals, Advicenne Pharmaceuticals, Medice, and EcoR1. JK has received research funding from Novo Nordisk Pharmaceuticals and Arbor Biotechnology as consulting Nordisk fees from Novo Pharmaceuticals. GET is on the Scientific Advisory Boards for and has received research funding from Alnylam Pharmaceuticals and Novo Nordisk. DJS has received research funding from Alnylam Pharmaceuticals and Novo Nordisk Pharmaceuticals as well as consulting fees from Advicenne Pharmaceuticals. All the other authors declared no conflicting interests.

ACKNOWLEDGMENTS

We wish to acknowledge the Oxalosis Hyperoxaluria Foundation, especially Kim Hollander and Julie Bertarelli, for their relentless support for this meeting and all endeavors to improve the lives of patients and families with PH. We also thank the University of Perugia for their generous hospitality as host of this conference. Finally, we thank the physicians and scientists who participated in this conference and provided outstanding expertise, enthusiasm, and vision. Besides the session chairs who are the authors on this paper, the speakers were: Nicola Brunetti-Pierri, Michael Somers, Barbara Ruggiero, Mirco Dindo, Monica Diaz-Gavilan, Sander Garrelfs, Andrea Molinos Vincente, Virginia Nieto-Romero, Ana Clara Najenson, Kyle Wood, Timucin Taner, Gill Rumsby, Giorgia Mandrile, Lisa Deesker, Imran Khan Jalbani, Xiaojing Tang, Neveen Soliman, Reham Almardini, Govinda Regmi, Ana Prado, Efrat Ben-Shalom, Sonia Fargue, Kerry M. Loomes, Emmanuel Letavernier, Marta Leporati, Jennifer Adams, Megan Prochaska, Kevin Koo, Aaron Miller, and Luigina Romani.

Funding

The meeting upon which this work is based was funded by the Oxalosis and Hyperoxaluria Foundation, a 502(3)(c) non-profit organization. Other funding for individual authors: BC: Oxalosis and Hyperoxaluria Foundation, Italian Ministry of University and Research (PRIN2022 2022J7CKMJ); JK: Oxalosis and Hyperoxaluria Foundation, Innovative Science Accelerator Program (NIH/NIDDK, U24DK128851); and LN: 1R01DK129675.

REFERENCES

- Bhasin B, Urekli HM, Atta MG. Primary and secondary hyperoxaluria: understanding the enigma. World J Nephrol. 2015;4:235–244. https://doi.org/10.5527/wjn.v4.i2.235
- Ermer T, Nazzal L, Tio MC, Waikar S, Aronson PS, Knauf F. Oxalate homeostasis. Nat Rev Nephrol. 2023;19:123–138. https://doi.org/10.1038/s41581-022-00643-3

- Fargue S, Acquaviva Bourdain C. Primary hyperoxaluria type 1: pathophysiology and genetics. Clin Kidney J. 2022;15:i4–i8. https://doi.org/10.1093/ckj/sfab217
- Ben-Shalom E, Garrelfs SF, Groothoff JW. Primary hyperoxaluria: the pediatric nephrologist's point of view. Clin Kidney J. 2022;15:i23–i28. https://doi.org/10.1093/ckj/sfab231
- Moochhala SH, Worcester EM. Primary hyperoxaluria: the adult nephrologist's point of view. Clin Kidney J. 2022;15: i29-i32. https://doi.org/10.1093/ckj/sfac068
- Rumsby G, Hulton SA. Primary hyperoxaluria type 2. In: Adam MP, Feldman J, Mirzaa GM, et al., eds. *GeneReviews*. University of Washington; 1993-2024.
- Belostotsky R, Seboun E, Idelson GH, et al. Mutations in DHDPSL are responsible for primary hyperoxaluria type III. Am J Hum Genet. 2010;87:392–399. https://doi.org/10.1016/j.ajhg.2010.07.023
- Milliner DS, Harris PC, Sas DJ, et al. Primary hyperoxaluria type 3. In: Adam MP, Feldman J, Mirzaa GM, et al., eds. GeneReviews. University of Washington; 1993-2024.
- Riedel TJ, Knight J, Murray MS, Milliner DS, Holmes RP, Lowther WT. 4-hydroxy-2-oxoglutarate aldolase inactivity in primary hyperoxaluria type 3 and glyoxylate reductase inhibition. *Biochim Biophys Acta*. 2012;1822:1544–1552. https://doi.org/10.1016/j.bbadis.2012.06.014
- Belostotsky R, Pitt JJ, Frishberg Y. Primary hyperoxaluria type III-a model for studying perturbations in glyoxylate metabolism. *J Mol Med (Berl)*. 2012;90:1497–1504. https://doi.org/10.1007/s00109-012-0930-z
- Holmes RP, Goodman HO, Assimos DG. Contribution of dietary oxalate to urinary oxalate excretion. *Kidney Int.* 2001;59:270–276. https://doi.org/10.1046/j.1523-1755.2001.00488.x
- Asplin JR. The management of patients with enteric hyperoxaluria. *Urolithiasis*. 2016;44:33–43. https://doi.org/10.1007/ s00240-015-0846-5
- Witting C, Langman CB, Assimos D, et al. Pathophysiology and treatment of enteric hyperoxaluria. Clin J Am Soc Nephrol. 2021;16:487–495. https://doi.org/10.2215/CJN.08000520
- Nazzal L, Puri S, Goldfarb DS. Enteric hyperoxaluria: an important cause of end-stage kidney disease. Nephrol Dial Transplant. 2016;31:375–382. https://doi.org/10.1093/ndt/gfv005
- Burns Z, Knight J, Fargue S, Holmes R, Assimos D, Wood K. Future treatments for hyperoxaluria. *Curr Opin Urol.* 2020;30: 171–176. https://doi.org/10.1097/MOU.000000000000000000
- Dindo M, Conter C, Oppici E, Ceccarelli V, Marinucci L, Cellini B. Molecular basis of primary hyperoxaluria: clues to innovative treatments. *Urolithiasis*. 2019;47:67–78. https:// doi.org/10.1007/s00240-018-1089-z
- Bacchetta J, Lieske JC. Primary hyperoxaluria type 1: novel therapies at a glance. Clin Kidney J. 2022;15:i17–i22. https:// doi.org/10.1093/ckj/sfab245
- Shee K, Stoller ML. Perspectives in primary hyperoxaluria historical, current and future clinical interventions. Nat Rev Urol. 2022;19:137–146. https://doi.org/10.1038/s41585-021-00543-4
- Sawyer K, Leahy S, Wood KD. Progress with RNA interference for the treatment of primary hyperoxaluria. *BioDrugs*. 2022;36:437–441. https://doi.org/10.1007/s40259-022-00539-5

- Syed YY. Nedosiran: first approval. *Drugs*. 2023;83:1729–1733. https://doi.org/10.1007/s40265-023-01976-4
- Hoppe B, Martin-Higueras C. Improving treatment options for primary hyperoxaluria. *Drugs*. 2022;82:1077–1094. https://doi.org/10.1007/s40265-022-01735-x
- Oppici E, Montioli R, Cellini B. Liver peroxisomal alanine: glyoxylate aminotransferase and the effects of mutations associated with Primary hyperoxaluria type I: an overview. *Biochim Biophys Acta*. 2015;1854:1212–1219. https://doi.org/ 10.1016/j.bbapap.2014.12.029
- Hopp K, Cogal AG, Bergstralh EJ, et al. Phenotype-genotype correlations and estimated carrier frequencies of primary hyperoxaluria. *J Am Soc Nephrol.* 2015;26:2559–2570. https://doi.org/10.1681/ASN.2014070698
- Sas DJ, Enders FT, Gunderson TM, et al. Natural history of clinical, laboratory, and echocardiographic parameters of a primary hyperoxaluria cohort on long term hemodialysis. Front Med (Lausanne). 2021;8:592357. https://doi.org/10. 3389/fmed.2021.592357
- Groothoff JW, Metry E, Deesker L, et al. Clinical practice recommendations for primary hyperoxaluria: an expert consensus statement from ERKNet and OxalEurope. Nat Rev Nephrol. 2023;19:194–211. https://doi.org/10.1038/s41581-022-00661-1
- Metry EL, Garrelfs SF, Peters-Sengers H, et al. Long-term transplantation outcomes in patients with primary hyperoxaluria Type 1 included in the European hyperoxaluria consortium (OxalEurope) registry. Kidney Int Rep. 2022;7: 210–220. https://doi.org/10.1016/j.ekir.2021.11.006
- Michael M, Harvey E, Milliner DS, et al. Diagnosis and management of primary hyperoxalurias: best practices. Pediatr Nephrol. 2024;39:3143–3155. https://doi.org/10.1007/ s00467-024-06328-2
- Garrelfs SF, Frishberg Y, Hulton SA, et al. Lumasiran, an RNAi therapeutic for primary hyperoxaluria Type 1. N Engl J Med. 2021;384:1216–1226. https://doi.org/10.1056/NEJMo a2021712
- Sas DJ, Magen D, Hayes W, et al. Phase 3 trial of lumasiran for primary hyperoxaluria type 1: a new RNAi therapeutic in infants and young children. *Genet Med.* 2022;24:654–662. https://doi.org/10.1016/j.gim.2021.10.024
- Hulton SA, Groothoff JW, Frishberg Y, et al. Randomized clinical trial on the long-term efficacy and safety of lumasiran in patients with primary hyperoxaluria type 1. Kidney Int Rep. 2022;7:494–506. https://doi.org/10.1016/j.ekir.2021. 12.001
- Sellier-Leclerc AL, Metry E, Clave S, et al. Isolated kidney transplantation under lumasiran therapy in primary hyperoxaluria type 1: a report of five cases. Nephrol Dial Transplant. 2023;38:517–521. https://doi.org/10.1093/ndt/gfac295
- Lombardi Y, Isnard P, Chavarot N, et al. Stiripentol and lumasiran as a rescue therapy for oxalate nephropathy recurrence after kidney transplantation in an adult patient with primary hyperoxaluria type 1. Am J Kidney Dis. 2023;82:113–116. https://doi.org/10.1053/j.ajkd.2022.12.005
- Metry EL, Deesker LJ, Garrelfs SF, et al. Corrigendum to "Successful kidney-alone transplantation in a patient with PH1 on combination RNA-interference therapy.". Kidney International, 2023;104:203-204 Kidney Int. 2023;104:1038. https://doi.org/10.1016/j.kint.2023.08.002

- 34. Baum MA, Langman C, Cochat P, et al. PHYOX2: a pivotal randomized study of nedosiran in primary hyperoxaluria type 1 or 2. *Kidney Int.* 2023;103:207–217. https://doi.org/10.1016/j.kint.2022.07.025
- Hoppe B, Koch A, Cochat P, et al. Safety, pharmacodynamics, and exposure-response modeling results from a first-in-human phase 1 study of nedosiran (PHYOX1) in primary hyperoxaluria. *Kidney Int.* 2022;101:626–634. https://doi.org/10.1016/j.kint.2021.08.015
- Breeggemann MC, Gluck SL, Stoller ML, Lee MM. A case report of kidney-only transplantation in primary hyperoxaluria type 1: a novel approach with the use of nedosiran. Case Rep Nephrol Dial. 2023;13:63–69. https://doi.org/10. 1159/000531053
- Joher N, Moktefi A, Grimbert P, et al. Early post-transplant recurrence of oxalate nephropathy in a patient with primary hyperoxaluria type 1, despite pretransplant lumasiran therapy. *Kidney Int.* 2022;101:185–186. https://doi.org/10. 1016/j.kint.2021.10.022
- Garrelfs SF, Metry EL, van Harskamp D, et al. Glycolate oxidase inhibition by lumasiran varies between patients with primary hyperoxaluria type 1. Kidney Int. 2023;103:990–993. https://doi.org/10.1016/j.kint.2023.01.029
- Pey AL, Albert A, Salido E. Protein homeostasis defects of alanine-glyoxylate aminotransferase: new therapeutic strategies in primary hyperoxaluria type I. *BioMed Res Int.* 2013;2013:687658. https://doi.org/10.1155/2013/687658
- Dindo M, Mandrile G, Conter C, et al. The ILE56 mutation on different genetic backgrounds of alanine:glyoxylate aminotransferase: clinical features and biochemical characterization. *Mol Genet Metab*. 2020;131:171–180. https://doi.org/10. 1016/j.ymgme.2020.07.012
- Mandrile G, Beck B, Acquaviva C, et al. Genetic assessment in primary hyperoxaluria: why it matters. *Pediatr Nephrol.* 2023;38:625–634. https://doi.org/10.1007/s00467-022-05613-2
- Dindo M, Pascarelli S, Chiasserini D, et al. Structural dynamics shape the fitness window of alanine:glyoxylate aminotransferase. *Protein Sci.* 2022;31:e4303. https://doi.org/10.1002/pro.4303
- Alejo-Armijo A, Cuadrado C, Altarejos J, et al. Lactate dehydrogenase A inhibitors with a 2,8-dioxabicyclo [3.3.1] nonane scaffold: a contribution to molecular therapies for primary hyperoxalurias. *Bioorg Chem.* 2022;129:106127. https://doi.org/10.1016/j.bioorg.2022.106127
- Cabrera N, Cuesta SA, Mora JR, et al. Searching glycolate oxidase inhibitors based on QSAR, molecular docking, and molecular dynamic simulation approaches. *Sci Rep.* 2022;12:19969. https://doi.org/10.1038/s41598-022-24196-4
- Ding J, Gumpena R, Boily MO, et al. Dual glycolate oxidase/ lactate dehydrogenase A inhibitors for primary hyperoxaluria. ACS Med Chem Lett. 2021;12:1116–1123. https:// doi.org/10.1021/acsmedchemlett.1c00196
- Moya-Garzon MD, Gomez-Vidal JA, Alejo-Armijo A, et al. Small molecule-based enzyme inhibitors in the treatment of primary hyperoxalurias. *J Pers Med.* 2021;11:74. https://doi. org/10.3390/jpm11020074
- Salido S, Alejo-Armijo A, Parola AJ, et al. Chitosan derivatives as nanocarriers for hLDHA inhibitors delivery to hepatic cells: a selective strategy for targeting primary

- hyperoxaluria diseases. *Int J Pharm.* 2022;627:122224. https://doi.org/10.1016/j.ijpharm.2022.122224
- Martin-Higueras C, Luis-Lima S, Salido E. Glycolate oxidase is a safe and efficient target for substrate reduction therapy in a mouse model of primary hyperoxaluria type I. *Mol Ther*. 2016;24:719–725. https://doi.org/10.1038/mt.2015.224
- Grottelli S, Annunziato G, Pampalone G, et al. Identification of human alanine-glyoxylate aminotransferase ligands as pharmacological chaperones for variants associated with primary hyperoxaluria type 1. J Med Chem. 2022;65:9718– 9734. https://doi.org/10.1021/acs.jmedchem.2c00142
- Kletzmayr A, Mulay SR, Motrapu M, et al. Inhibitors of calcium oxalate crystallization for the treatment of oxalate nephropathies. Adv Sci (Weinh). 2020;7:1903337. https://doi.org/10.1002/advs.201903337
- Torella L, Klermund J, Bilbao-Arribas M, et al. Efficient and safe therapeutic use of paired Cas9-nickases for primary hyperoxaluria type 1. *EMBO Mol Med*. 2024;16:112–131. https://doi.org/10.1038/s44321-023-00008-8
- Chen Z, Zhang D, Zheng R, et al. In vivo base editing rescues primary hyperoxaluria type 1 in rats. *Kidney Int*. 2024;105: 496–507. https://doi.org/10.1016/j.kint.2023.11.029
- Zabaleta N, Barberia M, Martin-Higueras C, et al. CRISPR/ Cas9-mediated glycolate oxidase disruption is an efficacious and safe treatment for primary hyperoxaluria type I. Nat Commun. 2018;9:5454. https://doi.org/10.1038/s41467-018-07827-1
- Nieto-Romero V, Garcia-Torralba A, Molinos-Vicente A, et al. Restored glyoxylate metabolism after AGXT gene correction and direct reprogramming of primary hyperoxaluria type 1 fibroblasts. iScience. 2024;27:109530. https://doi.org/10. 1016/j.isci.2024.109530
- D'Costa MR, Kausz AT, Carroll KJ, et al. Subsequent urinary stone events are predicted by the magnitude of urinary oxalate excretion in enteric hyperoxaluria. Nephrol Dial Transplant. 2021;36:2208–2215. https://doi.org/10.1093/ndt/ gfaa281
- Sas DJ, Mara K, Mehta RA, et al. Natural history of urine and plasma oxalate in children with primary hyperoxaluria type 1. *Pediatr Nephrol.* 2024;39:141–148. https://doi.org/10.1007/ s00467-023-06074-x
- 57. van Harskamp D, Garrelfs SF, Oosterveld MJS, Groothoff JW, van Goudoever JB, Schierbeek H. Development and validation of a new gas chromatography-tandem mass spectrometry method for the measurement of enrichment of glyoxylate metabolism analytes in hyperoxaluria patients using a stable isotope procedure. *Anal Chem.* 2020;92:1826–1832. https://doi.org/10.1021/acs.anal-chem.9b03670
- Carrasco A Jr., Granberg CF, Gettman MT, Milliner DS, Krambeck AE. Surgical management of stone disease in patients with primary hyperoxaluria. *Urology*. 2015;85:522– 526. https://doi.org/10.1016/j.urology.2014.11.018
- Cornell LD, Amer H, Viehman JK, et al. Posttransplant recurrence of calcium oxalate crystals in patients with primary hyperoxaluria: incidence, risk factors, and effect on renal allograft function. Am J Transplant. 2022;22:85–95. https://doi.org/10.1111/ajt.16732
- Cogal AG, Arroyo J, Shah RJ, et al. Comprehensive genetic analysis reveals complexity of monogenic urinary stone

- disease. *Kidney Int Rep.* 2021;6:2862–2884. https://doi.org/10.1016/j.ekir.2021.08.033
- Gefen AM, Zaritsky JJ. Review of childhood genetic nephrolithiasis and nephrocalcinosis. Front Genet. 2024;15: 1381174. https://doi.org/10.3389/fgene.2024.1381174
- Halbritter J, Baum M, Hynes AM, et al. Fourteen monogenic genes account for 15% of nephrolithiasis/nephrocalcinosis. J Am Soc Nephrol. 2015;26:543–551. https://doi.org/10.1681/ ASN.2014040388
- Romeo G, Bittles AH. Consanguinity in the contemporary world. Hum Hered. 2014;77:6–9. https://doi.org/10.1159/ 000363352
- Sheridan E, Wright J, Small N, et al. Risk factors for congenital anomaly in a multiethnic birth cohort: an analysis of the Born in Bradford study. *Lancet*. 2013;382:1350–1359. https://doi.org/10.1016/S0140-6736(13)61132-0
- Deesker LJ, Garrelfs SF, Mandrile G, et al. Improved outcome of infantile oxalosis over time in Europe: data from the OxalEurope registry. *Kidney Int Rep.* 2022;7:1608–1618. https://doi.org/10.1016/j.ekir.2022.04.012
- Williams HE, Smith LH Jr. L-glyceric aciduria. A new genetic variant of primary hyperoxaluria. N Engl J Med. 1968;278: 233–238. https://doi.org/10.1056/NEJM196802012780502
- MISTRY J, DANPURE CJ, CHALMERS RA. Hepatic d-glycerate dehydrogenase and glyoxylate reductase deficiency in primary hyperoxaluria type 2. *Biochem Soc Trans*. 1988;16: 626–627. https://doi.org/10.1042/bst0160626
- Cramer SD, Ferree PM, Lin K, Milliner DS, Holmes RP. The gene encoding hydroxypyruvate reductase (GRHPR) is mutated in patients with primary hyperoxaluria type II. *Hum Mol Genet*. 1999;8:2063–2069. https://doi.org/10.1093/hmg/8. 11.2063
- Behnam JT, Williams EL, Brink S, Rumsby G, Danpure CJ. Reconstruction of human hepatocyte glyoxylate metabolic pathways in stably transformed Chinese-hamster ovary cells. *Biochem J.* 2006;394:409–416. https://doi.org/10.1042/ BJ20051397
- Garrelfs SF, Rumsby G, Peters-Sengers H, et al. Patients with primary hyperoxaluria type 2 have significant morbidity and require careful follow-up. *Kidney Int.* 2019;96:1389–1399. https://doi.org/10.1016/j.kint.2019.08.018
- Johnson SA, Rumsby G, Cregeen D, Hulton SA. Primary hyperoxaluria type 2 in children. *Pediatr Nephrol*. 2002;17: 597–601. https://doi.org/10.1007/s00467-002-0858-6
- Dhondup T, Lorenz EC, Milliner DS, Lieske JC. Combined liver-kidney transplantation for primary hyperoxaluria type
 a case report. Am J Transplant. 2018;18:253–257. https://doi.org/10.1111/ajt.14418
- Del Bello A, Cointault O, Delas A, Kamar N. Primary hyperoxaluria type 2 successfully treated with combined liverkidney transplantation after failure of isolated kidney transplantation. Am J Transplant. 2020;20:1752–1753. https://doi. org/10.1111/ajt.15829
- Genena KM, Sas DJ, Milliner DS, Lieske JC. Successful treatment of primary hyperoxaluria Type 2 with a combined liver and kidney transplant. Kidney Int Rep. 2023;8:1469– 1472. https://doi.org/10.1016/j.ekir.2023.03.013
- 75. Sas DJ, Harris PC, Milliner DS. Recent advances in the identification and management of inherited hyperoxalurias.

- *Urolithiasis.* 2019;47:79–89. https://doi.org/10.1007/s00240-018-1093-3
- Singh P, Viehman JK, Mehta RA, et al. Clinical characterization of primary hyperoxaluria type 3 in comparison to types 1 and 2: a retrospective cohort study. Nephrol Dial Transplant. 2022;37:869–875.
- Singh P, Granberg CF, Harris PC, et al. Primary hyper-oxaluria type 3 can also result in kidney failure: a case report. Am J Kidney Dis. 2021;79:125–128. https://doi.org/10.1053/j.ajkd.2021.05.016
- Hockaday TD, Clayton JE, Frederick EW, Smith LH. Primary hyperoxaluria. *Med (Baltim)*. 1964;43:315–345. https://doi. org/10.1097/00005792-196405000-00010
- Knight J, Jiang J, Assimos DG, Holmes RP. Hydroxyproline ingestion and urinary oxalate and glycolate excretion. *Kid-ney Int.* 2006;70:1929–1934. https://doi.org/10.1038/sj.ki. 5001906
- Knight J, Assimos DG, Callahan MF, Holmes RP. Metabolism of primed, constant infusions of [1,2-(1) (3)C(2)] glycine and [1-(1) (3)C(1)] phenylalanine to urinary oxalate. *Metabolism*. 2011;60:950–956. https://doi.org/10.1016/j.metabol.2010.09. 002
- Fargue S, Milliner DS, Knight J, Olson JB, Lowther WT, Holmes RP. Hydroxyproline metabolism and oxalate synthesis in primary hyperoxaluria. *J Am Soc Nephrol*. 2018;29: 1615–1623. https://doi.org/10.1681/ASN.2017040390
- Garrelfs SF, van Harskamp D, Peters-Sengers H, et al. Endogenous oxalate production in primary hyperoxaluria type 1 patients. J Am Soc Nephrol. 2021;32:3175–3186. https://doi.org/10.1681/ASN.2021060729
- Ziff M, Kibrick A, Dresner E, Gribetz HJ. Excretion of hydroxyproline in patients with rheumatic and non-rheumatic diseases. J Clin Invest. 1956;35:579–587. https://doi.org/10.1172/JCI103311
- Mavropalias G, Calapre L, Morici M, et al. Changes in plasma hydroxyproline and plasma cell-free DNA concentrations after higher- versus lower-intensity eccentric cycling. Eur J Appl Physiol. 2021;121:1087–1097. https://doi. org/10.1007/s00421-020-04593-1
- Hart W, Duursma SA, Visser WJ, Njio LK. The hydroxyproline content of plasma of patients with impaired renal function. Clin Nephrol. 1975;4:104–108.
- Staufner C, Haack TB, Feyh P, et al. Genetic cause and prevalence of hydroxyprolinemia. *J Inherit Metab Dis*. 2016;39:625–632. https://doi.org/10.1007/s10545-016-9940-2
- Buchalski B, Wood KD, Challa A, et al. The effects of the inactivation of hydroxyproline dehydrogenase on urinary oxalate and glycolate excretion in mouse models of primary hyperoxaluria. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866:165633. https://doi.org/10.1016/j.bbadis.2019. 165633
- Bons J, Tadeo A, Scott GK, et al. Therapeutic targeting of HYPDH/PRODH2 with N-propargylglycine offers a hyperoxaluria treatment opportunity. *Biochim Biophys Acta Mol Basis Dis.* 2024;1870:166848. https://doi.org/10.1016/j.bbadis.2023.166848
- Serrano-Lorenzo P, Rabasa M, Esteban J, et al. Clinical, biochemical, and molecular characterization of two families with novel mutations in the LDHA gene (GSD XI).

- Genes (Basel). 2022;13:1835. https://doi.org/10.3390/genes
- Goldfarb DS, Lieske JC, Groothoff J, et al. Nedosiran in primary hyperoxaluria subtype 3: results from a phase I, single-dose study (PHYOX4). *Urolithiasis*. 2023;51:80. https://doi.org/10.1007/s00240-023-01453-3
- Williams EL, Bockenhauer D, van't Hoff WG, et al. The enzyme 4-hydroxy-2-oxoglutarate aldolase is deficient in primary hyperoxaluria type 3. Nephrol Dial Transplant. 2012;27:3191–3195. https://doi.org/10.1093/ndt/gfs039
- Monico CG, Rossetti S, Belostotsky R, et al. Primary hyperoxaluria type III gene HOGA1 (formerly DHDPSL) as a possible risk factor for idiopathic calcium oxalate urolithiasis. Clin J Am Soc Nephrol. 2011;6:2289–2295. https://doi. org/10.2215/CJN.02760311
- Huang A, Burke J, Bunker RD, et al. Regulation of human 4-hydroxy-2-oxoglutarate aldolase by pyruvate and alpha-ketoglutarate: implications for primary hyperoxaluria type-3. *Biochem J.* 2019;476:3369–3383. https://doi.org/10.1042/BCJ20190548
- Lieske JC, Mehta RA, Milliner DS, Rule AD, Bergstralh EJ, Sarr MG. Kidney stones are common after bariatric surgery. Kidney Int. 2015;87:839–845. https://doi.org/10.1038/ki.2014. 352
- Penniston KL, Kaplon DM, Gould JC, Nakada SY. Gastric band placement for obesity is not associated with increased urinary risk of urolithiasis compared to bypass. J Urol. 2009;182:2340–2346. https://doi.org/10.1016/j.juro.2009.07. 041
- Semins MJ, Matlaga BR, Shore AD, et al. The effect of gastric banding on kidney stone disease. *Urology*. 2009;74: 746–749. https://doi.org/10.1016/j.urology.2009.04.093
- 97. Stauffer JQ. Hyperoxaluria and intestinal disease. The role of steatorrhea and dietary calcium in regulating intestinal oxalate absorption. *Am J Dig Dis.* 1977;22:921–928. https://doi.org/10.1007/BF01076170
- Campos GM, Khoraki J, Browning MG, Pessoa BM, Mazzini GS, Wolfe L. Changes in utilization of bariatric surgery in the United States from 1993 to 2016. Ann Surg. 2020;271:201–209. https://doi.org/10.1097/SLA.0000000000 003554
- Bahaee J, Plott J, Ghani KR. Single-use flexible ureteroscopes: how to choose and what is around the corner? Curr Opin Urol. 2021;31:87–94. https://doi.org/10.1097/MOU.0000000000000852
- Sudhir Pillai P, Hsieh SS, Vercnocke AJ, et al. In vivo prediction of kidney stone fragility using Radiomics-based regression models. *J Endourol*. 2023;37:443–452. https://doi.org/10.1089/end.2022.0483
- Mehta M, Goldfarb DS, Nazzal L. The role of the microbiome in kidney stone formation. *Int J Surg.* 2016;36:607–612. https://doi.org/10.1016/j.ijsu.2016.11.024
- Kelsey R. Stones: Individual gut microbiome in nephrolithiasis. *Nat Rev Urol.* 2018;15:202. https://doi.org/10.1038/nrurol.2018.16
- Miller AW, Penniston KL, Fitzpatrick K, Agudelo J, Tasian G, Lange D. Mechanisms of the intestinal and urinary microbiome in kidney stone disease. *Nat Rev Urol.* 2022;19:695– 707. https://doi.org/10.1038/s41585-022-00647-5

- 104. Galan-Llopis JA, Sanchez-Pellicer P, Navarro-Lopez V. Role of microbiome in kidney stone disease. *Curr Opin Urol.* 2023;33:84–89. https://doi.org/10.1097/MOU.000000000000000000151
- Liu M, Nazzal L. Enteric hyperoxaluria: role of microbiota and antibiotics. *Curr Opin Nephrol Hypertens*. 2019;28:352– 359. https://doi.org/10.1097/MNH.0000000000000518
- Hatch M, Cornelius J, Allison M, Sidhu H, Peck A, Freel RW.
 Oxalobacter sp. reduces urinary oxalate excretion by promoting enteric oxalate secretion. *Kidney Int.* 2006;69:691–698. https://doi.org/10.1038/sj.ki.5000162
- Canales BK, Hatch M. Oxalobacter formigenes colonization normalizes oxalate excretion in a gastric bypass model of hyperoxaluria. Surg Obes Relat Dis. 2017;13:1152–1157. https://doi.org/10.1016/j.soard.2017.03.014
- Stern JM, Moazami S, Qiu Y, et al. Evidence for a distinct gut microbiome in kidney stone formers compared to non-stone formers. *Urolithiasis*. 2016;44:399–407. https://doi.org/10. 1007/s00240-016-0882-9
- 109. Hoppe B, Pellikka PA, Dehmel B, Banos A, Lindner E, Herberg U. Effects of Oxalobacter formigenes in subjects with primary hyperoxaluria type 1 and end-stage renal disease: a Phase II study. Nephrol Dial Transplant. 2021;36: 1464–1473. https://doi.org/10.1093/ndt/gfaa135
- Milliner D, Hoppe B, Groothoff J. A randomised Phase II/III study to evaluate the efficacy and safety of orally administered Oxalobacter formigenes to treat primary hyperoxaluria. *Urolithiasis*. 2018;46:313–323. https://doi.org/10. 1007/s00240-017-0998-6
- 111. Hoppe B, Niaudet P, Salomon R, et al. A randomised Phase I/ Il trial to evaluate the efficacy and safety of orally administered Oxalobacter formigenes to treat primary hyperoxaluria. *Pediatr Nephrol.* 2017;32:781–790. https://doi.org/ 10.1007/s00467-016-3553-8
- Ariceta G, Collard L, Abroug S, et al. ePHex: a phase 3, double-blind, placebo-controlled, randomized study to evaluate long-term efficacy and safety of Oxalobacter formigenes in patients with primary hyperoxaluria. *Pediatr Nephrol.* 2023;38:403–415. https://doi.org/10.1007/s00467-022-05591-5
- Quintero E, Bird VY, Liu H, et al. A prospective, double-blind, randomized, placebo-controlled, crossover study using an orally administered oxalate decarboxylase (OxDC). Kidney360. 2020;1:1284–1290. https://doi.org/10.34067/KID. 0001522020
- Langman CB, Grujic D, Pease RM, et al. A double-blind, placebo controlled, randomized Phase 1 cross-over study with ALLN-177, an orally administered oxalate degrading enzyme. Am J Nephrol. 2016;44:150–158. https://doi.org/10. 1159/000448766
- Lingeman JE, Pareek G, Easter L, et al. ALLN-177, oral enzyme therapy for hyperoxaluria. *Int Urol Nephrol.* 2019;51: 601–608. https://doi.org/10.1007/s11255-019-02098-1
- Costantini C, Dindo M, Pariano M, et al. Commensal fungi and oxalate degradation: is there a link? *Microbiota and host*. 2024;2:e230020.
- Wolfe AJ, Toh E, Shibata N, et al. Evidence of uncultivated bacteria in the adult female bladder. *J Clin Microbiol*. 2012;50:1376–1383. https://doi.org/10.1128/JCM.05852-11

- Kachroo N, Lange D, Penniston KL, et al. Standardization of microbiome studies for urolithiasis: an international consensus agreement. Nat Rev Urol. 2021;18:303–311. https://doi.org/10.1038/s41585-021-00450-8
- 119. Kachroo N, Monga M, Miller AW. Comparative functional analysis of the urinary tract microbiome for individuals with or without calcium oxalate calculi. *Urolithiasis*. 2022;50:303– 317. https://doi.org/10.1007/s00240-022-01314-5
- Knauf F, Ko N, Jiang Z, et al. Net intestinal transport of oxalate reflects passive absorption and SLC26A6-mediated secretion. J Am Soc Nephrol. 2011;22:2247–2255. https://doi.org/10.1681/ASN.2011040433
- 121. Freel RW, Whittamore JM, Hatch M. Transcellular oxalate and CI- absorption in mouse intestine is mediated by the

- DRA anion exchanger Slc26a3, and DRA deletion decreases urinary oxalate. *Am J Physiol Gastrointest Liver Physiol.* 2013;305:G520–G527. https://doi.org/10.1152/ajpgi.00167. 2013
- Hatch M, Canales BK. The mechanistic basis of hyperoxaluria following gastric bypass in obese rats. *Urolithiasis*. 2016;44:221–230. https://doi.org/10.1007/s00240-015-0836-7
- Pfau A, Lopez-Cayuqueo KI, Scherer N, et al. SLC26A1 is a major determinant of sulfate homeostasis in humans. J Clin Invest. 2023;133. https://doi.org/10.1172/JCI161849
- 124. Pizarro TT, Pastorelli L, Bamias G, et al. SAMP1/YitFc mouse strain: a spontaneous model of Crohn's disease-like ileitis. *Inflamm Bowel Dis.* 2011;17:2566–2584. https://doi.org/10. 1002/ibd.21638