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Exploring the current molecular landscape and management of multiple myeloma patients with the t(11;14) translocation

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Multiple myeloma (MM) is a genetically complex disease. The key myelomainitiating genetic events are hyperdiploidy and translocations involving the immunoglobulin heavy chain (IgH) enhancer on chromosome 14, which leads to the activation of oncogenes (e.g., CCND1, CCND3, MAF, and MMSET). The t(11;14) translocation is the most common in MM (15%–20%) and results in cyclin D1 (CCND1) upregulation, which leads to kinase activation and tumor cell proliferation. Notably, t(11;14) occurs at a higher rate in patients with plasma cell leukemia (40%) and light chain amyloidosis (50%). Patients with myeloma who harbor the t(11;14) translocation have high levels of the antiapoptotic protein B-cell lymphoma 2 (BCL2). Multiple studies demonstrated that the presence of t(11;14) was predictive of BCL2 dependency, suggesting that BCL2 could be a target in this subtype of myeloma. Venetoclax, an oral

Abbreviations: ASCT, autologous stem cell transplantation; BAD, BCL2-associated death; BAK, BCL2 antagonist/killer; BAX, BCL2-associated X protein; BCL2, B-cell lymphoma 2; BCL2L1, B-cell lymphoma 2-like protein 1; BCLXL, B-cell lymphoma-extra large; BH3, BCL2 homology domain 3; BID, BCL2-interacting domain; BIK, BCL2-interacting killer; BIM, BCL2-interacting mediator; BM, bone marrow; BMF, B-cell maturation factor; CCND1, cyclin D1; chr., chromosome; CYLD, cylindromatosis tumor suppressor protein; HR, hazard ratio; i-FISH, interphase fluorescence *in-situ* hybridization; IgE, immunoglobulin E; IGH, immunoglobulin heavy chain; IgM, immunoglobulin M; IL-6, interleukin-6; IRF4, interferon regulatory factor 4; MCL1, myeloid cell leukemia 1; mg, milligrams; MM, multiple myeloma; ORR, overall response rate; OS, overall survival; PAX5, paired box protein 5; PCs, plasma cells; PFS, progression-free survival; PIK3IP1, PI3K-interacting protein 1; PUMA, p53-upregulated modulator of apoptosis; R/R, relapsed/refractory; TNT, time to next treatment; Ven-Vd, venetoclax–bortezomib-dexamethasone; VGPR, very good partial response; VRd, bortezomib-V, lenalidomide-R, dexamethasone-d.

BCL2 inhibitor, has shown remarkable activity in treating relapsed/refractory MM patients with t(11;14) and BCL2 overexpression, either as monotherapy or in combination with other anti-myeloma agents. In this review, we describe the molecular defects associated with the t(11;14), bring into question the standard cytogenetic risk of myeloma patients harboring t(11;14), summarize current efficacy and safety data of targeted venetoclax-based therapies, and discuss the future of individualized or precision medicine for this unique myeloma subgroup, which will guide optimal treatment.

KEYWORDS

Keywords: translocation t(11;14), multiple myeloma, precision medicine, BCL-2, venetoclax, genetic abnormalities, prognosis in myeloma

Introduction

Multiple myeloma (MM), a hematological malignancy of plasma cells (PCs), has a high level of molecular and clinical complexity (1, 2). Although MM remains an incurable disease, never before has existed such translational research progress and optimism about therapeutic options. Advances in genomic studies have increased our understanding of MM pathogenesis, demonstrating that it is not a single disease. Despite their morphological similarities, at least six different diseases, collectively known as MM, all derived from PCs, have been genetically defined (3, 4).

Early cytogenetic studies defined the most common MM abnormalities, including hyperdiploidy with chromosome (chr.) counts of 53–55 and the translocation t(11:14) (q13;q32), both described in the same paper (5). Subsequent research using interphase fluorescence *in-situ* hybridization (i-FISH) assays revealed that recurrent translocations involving the immunoglobulin heavy chain (IGH) enhancer of chr. 14 (known as 14q32 translocations) were as common in MM as hyperdiploidy (6). Both hyperdiploidy and IGH translocations are considered the main initiating events in the pathogenesis of myeloma (2, 4, 7–9).

The five most common 14q32 translocations (60% of all patients) are t(11;14), t(4;14), t(14;16), t(14;20), and t(6;14) (3, 10, 11). Translocations t(4;14), t(14;16), and t(14;20) are linked to poor prognosis and defined as high-risk cytogenetic prognostic markers, whereas patients with t(11;14), t(6;14), and/or hyperdiploidy are considered to have standard-risk disease (3, 10). The molecular classification into these subgroups has been widely used not only to predict survival but also to identify targeted therapies (9, 10, 12, 13).

MM is an incurable PC neoplasm with substantial inter- and intrapatient genetic heterogeneity, which cannot be fully captured by routine diagnostics, even within the same patient. Typically, precision medicine links molecular-genetic aberrations with matched targeted therapies. Nevertheless, the number of druggable targets in myeloma is extremely low. However, MM with t(11;14), which is susceptible to B-cell lymphoma 2 (BCL2) inhibition in the clinical setting, is the only subtype with a therapeutically exploitable vulnerability (14, 15). In addition, there is no doubt that MM with t(11;14) is a unique clinical and biological entity, corresponding most likely to B-cell myeloma (16, 17).

This review aims to characterize the molecular abnormalities associated with the t(11;14), define the cytogenetic risk of myeloma patients with t(11;14), and highlight current efficacy and safety data for venetoclax-based therapies. Furthermore, the potential of precision therapeutic intervention in this myeloma subgroup will be discussed.

Translocation t(11:14) in MM: update on molecular and clinical finding

The t(11:14) is historically the first translocation of the 14q32 chromosome, discovered using conventional karyotype (5). Better characterization was achieved by precisely mapping the translocation breakpoints on both chromosomes (18). Following that, several researchers investigated the clinical features and outcomes of t(11:14) patients (5, 19). Numerous studies established that t(11:14) is a unique myeloma subtype, initially considered as positive prognostic translocation (20–23). However, in the novel therapies era, large studies established that patients with t(11:14) may not exhibit superior clinical outcomes compared to patients without t(11:14) or other cytogenetically standard-risk individuals (15, 24–26).

However, importantly, nearly one-third of patients with t (11;14) harbor additional high-risk cytogenetic abnormalities (27). In an early study, among 212 newly diagnosed patients, the

most frequently coexisting aberrations with t(11;14) were del13q (37%), del(IGH) (33%), gain1q (20.7%), del16q (14.9%), del17p (7.7%), del1p (3.4%), and multiple gains1q (2.9%) (24). Thirty-five percent of these patients had at least two additional chromosomal abnormalities (24). The translocation t(11;14) has also been associated with cyclin D1 (CCND1) and IRF4 mutations (4), while an atypical form of t(11;14), characterized by unique FISH patterns, adding a particular biologic interest behind this lesion, has recently been reported (28).

The t(11;14) is the most frequent 14q32 translocation found in MM (15%–20%). It causes CCND1 upregulation, leading to kinase activation and tumor cell proliferation (16). A greater incidence (68%) of this translocation among relatively younger patients (<50 years) with the worse outcome has been reported (29). Furthermore, t(11;14) has been discovered in 10%–13% of individuals with monoclonal gammopathy of undetermined significance, implying that it is an early event in the disease pathogenesis (30, 31). This translocation can be found in other plasma cell-related neoplasms. It is significantly more common (~40%) in patients with plasma cell leukemia (32–34) and encountered nearly in half of those with light-chain amyloidosis (35).

African Americans or individuals with a high degree of African ancestry have an increased prevalence of the t(11;14) translocation (36, 37). This finding may partially explain the influence of race on prognosis and outcomes in MM, a complex subject with conflicting results under investigation (38, 39).

Patients with this specific translocation have a unique myeloma type and are less likely to be hyperdiploid (17). They show a two-fold increased incidence of light-chain or non-secretory myeloma, are more likely to present with bone disease, and harbor uncommon heavy-chain isotypes such as immunoglobulin M (IgM) or IgE (10, 39–42). Malignant PCs are less mature with scant cytoplasm, have a lymphoplasmacytic phenotype with CD20 expression in roughly half of the cases and, most critically, are prone to apoptosis through the anti-apoptotic protein BCL2, which is the target for venetoclax (39, 40, 43).

Because MM patients with t(11;14) overexpress BCL2, venetoclax, a highly selective oral BCL2 inhibitor, appears to be an effective therapeutic strategy (44–46). We focus on t(11:14) because it has emerged as the first predictor of response to BCL2 inhibition in MM, thereby establishing a very intriguing field of potential treatments.

The prognostic and predictive role of t(11;14) in MM

The presence of high-risk cytogenetic features identified originally as [del(17p), t(4;14), and t(14;16)] has been extensively documented as a negative prognostic factor in

MM. These patients typically have a worse outcome than standard-risk patients with t(11;14), t(6;14), and/or hyperdiploidy (42, 47, 48).

The risk classification of the t(11;14) has been a longrunning discussion, most likely in the wrong direction. The t (11;14) was associated with a relatively favorable outcome and classified as a standard-risk translocation when alkylators like melphalan or cyclophosphamide were the backbone of myeloma care (49). However, recent studies showed an inferior depth of response rates, progression-free survival (PFS), and overall survival (OS) for t(11:14) patients treated with novel agents compared to standard-risk myeloma (16, 50–52). These findings question the classification of t(11;14) patients in the standardrisk subgroup, claiming that they should be classified as intermediate risk and treated appropriately (24, 25).

One of the largest studies conducted by the Mayo Clinic first showed that MM patients with t(11;14) had better PFS, time to next treatment (TNT), and OS than the non-t(11;14)translocation group (high-risk abnormalities), but worse markers than the standard-risk group, which includes trisomies or normal cytogenetics. The outcome difference remained even after excluding patients with del(17p) from all subgroups. As a result, patients with t(11;14) were reclassified as "intermediate risk" (16).

Another retrospective, single-center study including 1,000 newly diagnosed MM patients analyzed the outcome of the largest cohort of t(11:14) patients ever evaluated. All patients were treated identically with a triplet regimen (VRd) consisting of a proteasome inhibitor (bortezomib, V), an immunomodulatory drug (lenalidomide, R), and dexamethasone (d) representing the standard of care in the era of novel therapies. In this study, the presence of t(11;14) was linked to lower response rates [very good partial response (VGPR) 50% *vs.* 76%; *p* < 0.001] and subsequently to shorter median PFS (51 *vs.* 75 months; *p* < 0.001) compared to patients with standard risk (51, 52).

According to several studies, autologous stem cell transplantation (ASCT) may abrogate the unfavorable outcome of patients harboring the t(11:14) with a superior survival for transplanted African Americans compared to Whites in the United States (53). However, the ASCT impact has not been adequately explored in these patients because they are often studied together with hyperdiploid patients as having standard-risk disease (54, 55).

Moreover, the presence of multiple cytogenetic abnormalities has a greater impact on the prognosis of MM than a single high-risk aberration (47, 56, 57). Individuals with multiple gains(1q), del(1p), del(IGH), and del(13q) have a significantly worse median OS (p < 0.05) than patients with a sole t(11;14). Worse outcome with weaker correlation has also been observed in patients with t(11;14) together with del(17p) (p = 0.07 > 0.05) (24). These results indicate that MM patients with t(11;14), who carry the extrachromosomal abnormalities

listed above, should be recognized as having a high-risk disease, rather than a standard-risk disease, and should be considered as at least intermediate-risk patients.

In a recent review, Gao et al. conclude that outcomes of t (11;14) MM are similar to standard-risk patients when they receive novel agent induction therapy consolidated by ASCT, regardless of coexisting with single gain(1q) or not (26). Significantly worse median OS has been observed for t(11;14) patients with multiple gains(1q), compared to single gain (1q) (24).

Translocation t(11;14) and BCL2 protein family codependency

BCL2 is a member of the regulator protein family controlling apoptosis by either initiating or ending cell death (44). BIM, BAD, PUMA, BID, BMF, HRK, NOXA, BAK, BAX, and BIK (BH3 domain-containing proteins) comprise the main proapoptotic BCL2 family proteins. Conversely, MCL1, BCLW, BCLXL, BFL1, and BCL2 are the main anti-apoptotic proteins. The balance of pro- and anti-apoptotic proteins affects cell fate, which varies between cells (46).

While chronic lymphocytic leukemia and follicular lymphoma are BCL2-dependent neoplasms, MM is typically MCL1 dependent. Specifically, most myeloma cell lines (80%) are dependent on the anti-apoptotic MCL1 protein or both MCL1 and BCLXL, being the most dominant pro-survival protein in MM (44, 58–61). Thus, MCL1 inhibitors exert antimyeloma properties (44, 46, 60).

In contrast, only 20% of myelomas favor signals through the BCL2 protein, which is a target of venetoclax (12, 44, 60–62). MM with t(11;14) has distinct biology with considerably increased BCL2 and decreased MCL1 expression (44, 60). Due to high BCL2 expression, patients with t(11;14) are dependent on anti-apoptotic BCL2 proteins (46, 58). The rationale behind the response to BCL2 inhibitors is that malignant cells exhibit anti-apoptotic properties to survive. The upregulation of BCL2 is the hallmark of resistance to apoptosis for myeloma cells. Thus, malignant PCs manage to overcome pro-apoptotic alterations, caused by hypoxia, growth factor withdrawal, or adhesion loss (12, 46).

Inflammatory cytokines such as IL-6, derived from bone marrow (BM), mediate BCL2, BCLXL, and MCL1 expression in MM, forming complex network interactions between the marrow stroma and myeloma cells (63–65). IL-6 is a paracrine factor delivered by the microenvironment, primarily by myeloid cells, promoting the growth and survival of malignant PCs (66, 67). These mechanisms could account for the complex signaling interactions between t(11;14) MM and the BM microenvironment.

Hence, there is ongoing research targeting survival pathways in MM, aiming to identify which patient subgroup will benefit more from venetoclax. Although the t(11;14) group has the highest response rates, this does not rule out the possibility of other MM subgroups responding. The response also depends on the ratio of pro- and anti-apoptotic protein expression in myeloma cells with t(11;14) (12, 46). Nevertheless, there is heterogeneity regarding the level of BCL2 and MCL1 expression (46, 61). Overall, sensitivity to venetoclax is closely associated with high BCL2 and low BCLXL or MCL1 expression levels (60, 61). Interestingly, venetoclax response does not always coincide with increased BCL2 expression (44, 46, 59, 68).

Translocation t(11;14) and precision intervention in MM

A better understanding of the t(11;14) prognostic consequences is required to improve present and future therapy options in MM. Establishing the t(11:14) translocation status is crucial for developing a personalized treatment myeloma strategy. Recently, a plethora of clinical trials using venetoclax, either alone or in combination with other drugs, has shown remarkable activity, particularly in the relapsed/ refractory (R/R) patients with t(11:14) and/or BCL2 overexpression (69–73). We present an update on clinical trials targeting this specific translocation with an emphasis on response rates and toxicity (Table 1).

The first evidence of the substantial clinical activity of venetoclax as a single agent was reported in the R/R myeloma setting in phase 1 clinical trials. Patients with very advanced MM were enrolled with a median of five prior therapies, the majority being refractory to both bortezomib and lenalidomide. Apart from two patients, all the rest of the responders (40%) were t (11;14) positive (74). Nevertheless, responses were also observed in non-t(11;14) patients who received venetoclax, although to a lesser extent (74).

Additionally, the efficacy of venetoclax was confirmed in R/R MM, when added to bortezomib and dexamethasone. This combination demonstrated 90% overall response rate (ORR) and >64% VGPR in individuals not refractory to bortezomib (69). Subsequently, in the phase III BELLINI trial (NCT02755597), venetoclax (800 mg/day) in combination with bortezomib and dexamethasone (Ven-Vd) demonstrated superior efficacy to placebo plus Vd in patients with t(11;14) R/R MM. After a median 4-year follow-up, the venetoclax arm achieved a PFS of 36.8 months compared to 9.3 months in the placebo arm [hazard ratio (HR), 0.12; 95% CI, 0.03-0.44; p = 0.0014]. The greatest benefit was observed for those who had high BCL2 expression or who were t(11;14) positive. In neither arm was the median OS reached (HR, 0.61; 95% CI, 0.16-2.32; p = 0.4654) (70, 77). In addition, while the trial's original analysis revealed a mortality increase among venetoclax-treated patients, the updated analysis did not report any further increase in early TABLE 1 Clinical trials involving venetoclax.

Reference	Trial phase	Targeted drugs/dose	Study MM population	Median lines of prior therapy/ refractoriness	Response rates	Grade 3-4 toxicity	Study status	Clinical trial identifier
Kumar S et al., 2017 (74)	Phase I Interventional Non- randomized	Venetoclax [300, 600, 900, 1,200 mg/day maximum tolerated dose (NR)], 1,200 mg/day (expansion)	R/R, 66 pts 30+ t(11;14)	5 prior lines Bortezomib (70%) Lenalidomide (77%) Pomalidomide (53%) ASCT (76%)	Total: ORR: 21% VGPR or better: 15% t(11;14): ORR: 40% VGPR or better: 27% DOR: 9.7 mts	Thrombopenia (26%) Neutropenia (21%) Anemia (14%) Lymphopenia (15%) Back Pain (8%) Pneumonia (8%)	Completed	NCT01794520
Kaufman JL et al., 2020 (71)	Phase I/II, non- randomized	Venetoclax (800 mg/day) Dexamethasone (40 mg/wk)	R/R t(11;14) 20 pts (phase I) 31 pts (phase II)	3 prior lines (phase I) 5 prior lines (phase II)	Phase I: ORR: 60% VGPR or better: 30% DOR: 12.4 mts Phase II: ORR: 48% VGPR or better: 36% DOR, OS: NR	Phase I: Thrombopenia (10%) Neutropenia (10%) Anemia (12%) Lymphopenia (20%) TLS: 10% Phase II: Similar 11 deaths 8 progression 2 adverse event	Completed	NCT01794520
Costa LJ et al., 2021 (72)	Phase II, non- randomized	Venetoclax Maximum tolerated dose (NR) 800 mg/day (expansion) Carfilzomib 70 mg/m ² (expansion) Dexamethasone	R/R, 49 pts	1–3 prior lines	t(11;14) median 2 prior lines: ORR: 92% VGPR or better: 85%	Novel safety concerns have not arisen	Ongoing	NCT02899052
Gasparetto C et al., 2021 (75)	Phase II, non- randomized	Venetoclax (400 mg/day) Pomalidomide (4 mg/day) Dexamethasone (40 mg/wk)	R/R, 8 pts		t(11;14) Positive or negative status	Toxicity	Terminated	NCT03567616
Mateos MV et al., 2020 (76)	Phase III, randomized, CANOVA	Venetoclax (400 mg/day) Dexamethasone (40 mg/wk) vs. Pomalidomide (4 mg/day) Dexamethasone (40 mg/wk)	244 pts (estimated)	≥2 prior lines Lenalidomide refractory (100%)			Ongoing	NCT03539744
Moreau P et al., 2017 (69)	Phase 1b, non- randomized	Venetoclax (100, 200, 300, 400, 500, 600, 800, 1,000, 1,200 mg/day) Bortezomib (1.3 mg/m ²) Dexamethasone (20 mg)	R/R, 66 pts t(11;14): 14%	3 prior lines Bortezomib (39%) Lenalidomide (53%) ASCT (59%)	Total: ORR: 67% VGPR or better: 42% DOR: 9.7 mts RR: 89% (1–3 prior lines), 50% (4–6), 11% (>6)	Thrombopenia (29%) Neutropenia (14%) Anemia (15%) Pneumonia (8%)	Completed	NCT01794507
Kumar S et al., 2020 (70) Kumar S et al., 2021 (77)	BELLINI, phase III, randomized, double blind Final cutoff data: March 2021 Median follow-up: 45.6 mts	Venetoclax (800 mg/day) Bortezomib (1.3 mg/m ²) Dexamethasone (20 mg) vs. Bortezomib (1.3 mg/m ²) Dexamethasone (20 mg)	R/R VEN: 194 pts t(11;14): 10% Placebo: 97 pts t(11;14): 15%	1–3 prior lines	Median OS: NR in both arms PFS median (total): VEN (23.4 mts) vs. placebo (11.4 mts) PFS median t (11;14): VEN (36.8 mts) vs. placebo (9.3 mts) ORR: VEN 82% vs. placebo 68%	Total deaths: VEN 78 (40%) vs. placebo 36 (37%) Serious infections: Ven: 35% placebo: 29% Fatal infections: VEN 9 pts vs. placebo 0 pts Thrombopenia Neutropenia Pneumonia Anemia Diarrhea	Suspended, due to safety	NCT02755597

(Continued)

Reference	Trial phase	Targeted drugs/dose	Study MM population	Median lines of prior therapy/ refractoriness	Response rates	Grade 3-4 toxicity	Study status	Clinical trial identifier
					VGPR or better: VEN 59% vs. placebo 36%			
Kaufman JL et al., 2020 (71)	Phases I and II, randomized Preliminary results (May 2020)	Venetoclax Various Doses Daratumumab 1800mg SC Or 16mg/kg IV Bortezomib (1.3mg/m ²) Dexamethasone 20 mg, cycles 1–3: 20 or 40 mg/wk, cycles 4–8: 20 mg/mth c. 9+	Part I (VEN-DARA- DEXA): t(11;14): 24 pts Part II (VEN-DARA- BORT- DEXA): 24 pts t(11;14): 6		ORR: >90%	Similar to previous VEN trials Infections: 9 pts Additional non-VEN arm (DARA-BORT- DEXA) to compare VEN toxicity	Ongoing	NCT03314181
Maples KT et al., 2020	Retrospective	Venetoclax	t(11;14) 68 pts		Total: ORR: 71% VGPR or better: 48.5% Median PFS: 14.1 or 23.2 mts for <3 prior lines		Completed	

TABLE 1 Continued

NR, not reached; DOR, duration of response; OS, overall survival; PFS, progression-free survival; ORR, overall response rate; RR, response rate; VGPR, very good partial response; TLS, tumor lysis syndrome; VEN, venetoclax; DARA, daratumumab; DEXA, dexamethasone; BORT, bortezomib; mts, months; pts, patients; MM, multiple myeloma.

deaths in the venetoclax arm. The BELLINI study was halted due to the abovementioned toxicity and the increased mortality rate in the venetoclax arm (40%, 78 deaths). Most deaths were due to infectious complications, especially during disease progression. Moreover, they were mainly confined to patients with low BCL2 expression of the non-t(11;14) subgroup (70, 77).

Venetoclax has been examined in combination with carfilzomib and dexamethasone (NCT02899052) in a nonrandomized, phase 2, open-label trial (72) as well as with daratumumab and dexamethasone with or without bortezomib (NCT03314181, a randomized phase 1 study) in cytogenetically unselected patients with R/R MM (73). The latter study demonstrated outstanding clinical responses with no new safety signals, confirming again that venetoclax is more effective in t(11;14) patients, with a high and deep ORR over 90% (73).

Based on the BELLINI trial results, the clinical development of venetoclax is currently limited to t(11;14) patients. For example, the CANOVA trial (NCT03539744) is a phase 3, randomized trial comparing venetoclax plus dexamethasone *vs.* pomalidomide plus dexamethasone in t(11:14) R/R MM patients (76). Nevertheless, a phase 2, non-randomized study of venetoclax in combination with pomalidomide and dexamethasone in R/R MM (NCT03567616), including patients with positive or negative t(11:14) status, was terminated due to limited enrollment and toxicity (75).

Preliminary data outside of clinical studies also confirmed that venetoclax-based combinations achieve remarkable responses in heavily pretreated R/R MM (78–80). While the BELLINI trial showed that a proper patient selection may reduce the risk of venetoclax-associated side effects, the natural history of myeloma relapsing on venetoclax is largely unknown. The first retrospective study to assess real-world outcomes from the time of venetoclax refractoriness demonstrated a median OS of 31.4 months. These findings support the use of venetoclax early in the treatment of the t (11;14) patient group, challenging the notion that venetoclax resistance results in a more aggressive disease phenotype (81).

These data significantly support BCL2-directed therapy, such as venetoclax in t(11;14) patients, comprising the first step toward a precise treatment strategy in myeloma, namely, "precision medicine" (9, 14, 61, 78, 82). We should underline, however, that venetoclax is still investigational and should only be used in the context of clinical trials. Although applying venetoclax outside of a clinical trial is sometimes helpful, we are still learning about the drug.

Currently, venetoclax is administered in the R/R setting to patients with MM harboring t(11;14). However, we assume that the drug could and should be a first-line treatment option. To the best of our knowledge, there are no ongoing clinical trials or research studies in this use. Patients with t(11;14) MM have an

unmet need for well-designed clinical trials evaluating the safety and efficacy of venetoclax as first-line therapy.

Response and resistance to venetoclax

Venetoclax is not beneficial for all patients with t(11;14). Certain patients may have a remarkable venetoclax response, while others do not. Although the biology underlying this heterogeneity is unknown, both *de-novo* and acquired resistance can occur (81). BCL2 expression has been correlated with a significantly higher response to venetoclax. Conversely, the expression of MCL1 and BCLXL in cell lines has been linked to a poor response (58, 60, 82). However, ongoing studies using BH3 profiling, a novel *ex-vivo* functional technique for determining a cancer cell's reliance on anti-apoptotic proteins, revealed that MM is a diverse disease in terms of anti-apoptotic protein dependency and cannot be considered exclusively BCL2, BCLXL, or MCL1 dependent (59, 60, 83).

Contrary to prior findings, Gupta et al. demonstrated that t (11;14) and CCND1 may not directly affect the likelihood of response to venetoclax in MM (68). They specifically identified other factors contributing to the venetoclax response, including increased expression of a signature panel of B-cell genes, exclusively observed in venetoclax-sensitive t(11;14) patients. Additionally, the authors developed a predictive flow cytometric score for venetoclax response including B markers (CD20, CD79A) used to detect venetoclax-sensitive myeloma without relying on t(11;14) (68).

Intriguingly, a patient subset lacking t(11;14) responds to venetoclax. This was observed in a subset of high-risk patients harboring the t(14;16) translocation, demonstrating a remarkable response to venetoclax, most likely due to their elevated CD2 expression signature (MS4A1 or CD20, CD79A, VPREB3, PIK3IP1), despite the absence of t(11;14) (68). They also reported that BIM binding to BCL2 is linked with venetoclax response, a finding supported by decreased sensitivity to venetoclax in BIM knockout mice.

Another study demonstrated that an immature plasma cell phenotype in myeloma patients with t(11;14) consisting of significantly low CD38 and CD138 expression and high levels of B-cell markers such as CD79A and PAX5, along with an increased BCL2/BCL2L1 ratio, was highly susceptible to venetoclax treatment and less sensitive to daratumumab-based therapies, supporting further the hypothesis that venetoclax sensitivity is predicted by a robust B-cell myeloma phenotype (84). Interestingly, CYLD inactivation and 1q gains were also reported to predict venetoclax response (85). In contrast, high neuregulin-2 expression has been linked to venetoclax resistance in non-t(11;14) MM (86). The shift in myeloma cell dependence from BCL2 to MCL1 or BCLXL is the hallmark of acquired resistance to venetoclax in t(11;14) MM, as most responders eventually relapse (87, 88). Another resistance mechanism has recently been identified as a *de-novo* D111A mutation in BCL2 caused by venetoclax treatment (88).

Concluding remarks

MM is a genetically complex disease associated with a number of recurrent translocations and mutations influencing prognosis, clinical presentation, and treatment response. Regardless of advances in the molecular disease landscape, this progress has not been translated into clinical benefit, primarily because the majority of drugs still target the neoplastic tumor burden rather than specific mutations, rearrangements, or underlying genomic defects of PCs (89). A remarkable exception seems to be venetoclax, the first targeted medication with clinical efficacy proven in a patient subset carrying a certain cytogenetic profile, like t(11;14). Defining MM patients who are most likely to benefit from venetoclax treatment is still a work in progress. Detection of features associated with venetoclax response is crucial. A recent study showed that a specific B-cell gene signature predicts better and stronger responses than the t(11;14) translocation (68).

Myeloma with t(11;14) has distinct biology with considerably increased expression of the anti-apoptotic protein BCL2. The translocation t(11;14) has been associated with a more severe disease phenotype in patients who carry additional chromosomal or molecular abnormalities and this should be further evaluated. A better understanding of the prognostic consequences of the t(11;14) translocation is necessary to guide existing and future therapy options (46, 68).

Despite the fact that our understanding from t(11:14) has been translated into clinical practice, the benefits are applicable to some patients. Most patients do not respond for long and their disease worsens. The combination of venetoclax or BCL2 inhibitors with other anti-myeloma drugs or novel intervention strategies has the potential to improve clinical outcomes.

Author contributions

D.D.M: collected and assembled the data, generated the main body and tables of the manuscript S.P. generated sections of the manuscript E.H: conceptualized and designed the document, guided the co-authors, generated sections of the manuscript, and was responsible for the final writing of the document. The entire manuscript was reviewed and approved by all co-authors.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Manier S, Salem KZ, Park J, Landau DA, Getz G, Ghobrial IM. Genomic complexity of multiple myeloma and its clinical implications. *Nat Rev Clin Oncol* (2017) 14(2):100–13. doi: 10.1038/nrclinonc.2016.122

2. Manier S, Salem K, Glavey SV, Roccaro AM, Ghobrial IM. Genomic aberrations in multiple myeloma. *Cancer Treat Res* (2016) 169:23-34. doi: 10.1007/978-3-319-40320-5_3

3. Fonseca R, Bergsagel PL, Drach J, Shaughnessy J, Gutierrez N, Stewart AK, et al. International myeloma working group molecular classification of multiple myeloma: spotlight review. *Leukemia* (2009) 23(12):2210–21. doi: 10.1038/leu.2009.174

4. Walker BA, Mavrommatis K, Wardell CP, Ashby TC, Bauer M, Davies FE, et al. Identification of novel mutational drivers reveals oncogene dependencies in multiple myeloma. *Blood* (2018) 132(6):587–97. doi: 10.1182/blood-2018-03-840132

5. Liang W, Hopper JE, Rowley JD. Karyotypic abnormalities and clinical aspects of patients with multiple myeloma and related paraproteinemic disorders. *Cancer* (1979) 44(2):630–44. doi: 10.1002/1097-0142(197908) 44:2<630::aid-cncr2820440233>3.0.co;2-g

6. Bergsagel PL, Kuehl WM. Molecular pathogenesis and a consequent classification of multiple myeloma. *J Clin Oncol* (2005) 23(26):6333-8. doi: 10.1200/jco.2005.05.021

7. Hultcrantz M, Yellapantula V, Rustad EH. Genomic profiling of multiple myeloma: New insights and modern technologies. *Best Pract Res Clin Haematol* (2020) 33(1):101153. doi: 10.1016/j.beha.2020.101153

8. Weaver CJ, Tariman JD. Multiple myeloma genomics: A systematic review. Semin Oncol Nurs (2017) 33(3):237-53. doi: 10.1016/j.soncn.2017.05.001

 Cardona-Benavides IJ, de Ramón C, Gutiérrez NC. Genetic abnormalities in multiple myeloma: Prognostic and therapeutic implications. *Cells* (2021) 10(2):336. doi: 10.3390/cells10020336

10. Abdallah N, Rajkumar SV, Greipp P, Kapoor P, Gertz MA, Dispenzieri A, et al. Cytogenetic abnormalities in multiple myeloma: association with disease characteristics and treatment response. *Blood Cancer J* (2020) 10(8):82. doi: 10.1038/s41408-020-00348-5

11. Robiou du Pont S, Cleynen A, Fontan C, Attal M, Munshi N, Corre J, et al. Genomics of multiple myeloma. *J Clin Oncol* (2017) 35(9):963–7. doi: 10.1200/jco.2016.70.6705

12. Perrot A, Corre J, Avet-Loiseau H. Risk stratification and targets in multiple myeloma: From genomics to the bedside. *Am Soc Clin Oncol Educ Book* (2018) 38:675–80. doi: 10.1200/edbk_200879

13. Rajkumar SV, Kumar S. Multiple myeloma current treatment algorithms. Blood Cancer J (2020) 10(9):94. doi: 10.1038/s41408-020-00359-2

14. Pan D, Richter J. Where we stand with precision therapeutics in myeloma: Prosperity, promises, and pipedreams. *Front Oncol* (2021) 11:819127. doi: 10.3389/ fonc.2021.819127

15. Paner A, Patel P, Dhakal B. The evolving role of translocation t(11;14) in the biology, prognosis, and management of multiple myeloma. *Blood Rev* (2020) 41:100643. doi: 10.1016/j.blre.2019.100643

16. Lakshman A, Alhaj Moustafa M, Rajkumar SV, Dispenzieri A, Gertz MA, Buadi FK, et al. Natural history of t(11;14) multiple myeloma. *Leukemia* (2018) 32 (1):131–8. doi: 10.1038/leu.2017.204

17. Fonseca R, Blood EA, Oken MM, Kyle RA, Dewald GW, Bailey RJ, et al. Myeloma and the t(11;14)(q13;q32); evidence for a biologically defined unique subset of patients. *Blood* (2002) 99(10):3735–41. doi: 10.1182/blood.v99.10.3735

18. Chesi M, Bergsagel PL, Brents LA, Smith CM, Gerhard DS, Kuehl WM. Dysregulation of cyclin D1 by translocation into an IgH gamma switch region in two multiple myeloma cell lines. *Blood* (1996) 88(2):674–81. doi: 10.1182/blood.V88.2.674.bloodjournal882674

19. Dewald GW, Kyle RA, Hicks GA, Greipp PR. The clinical significance of cytogenetic studies in 100 patients with multiple myeloma, plasma cell leukemia, or amyloidosis. *Blood* (1985) 66(2):380–90. doi: 10.1182/blood.V66.2.380.380

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20. Moreau P, Facon T, Leleu X, Morineau N, Huyghe P, Harousseau JL, et al. Recurrent 14q32 translocations determine the prognosis of multiple myeloma, especially in patients receiving intensive chemotherapy. *Blood* (2002) 100(5):1579– 83. doi: 10.1182/blood-2002-03-0749

21. Avet-Loiseau H, Attal M, Moreau P, Charbonnel C, Garban F, Hulin C, et al. Genetic abnormalities and survival in multiple myeloma: the experience of the intergroupe francophone du myélome. *Blood* (2007) 109(8):3489–95. doi: 10.1182/ blood-2006-08-040410

22. Dewald GW, Therneau T, Larson D, Lee YK, Fink S, Smoley S, et al. Relationship of patient survival and chromosome anomalies detected in metaphase and/or interphase cells at diagnosis of myeloma. *Blood* (2005) 106(10):3553–8. doi: 10.1182/blood-2005-05-1981

23. Gertz MA, Lacy MQ, Dispenzieri A, Greipp PR, Litzow MR, Henderson KJ, et al. Clinical implications of t(11;14)(q13;q32), t(4;14)(p16.3;q32), and -17p13 in myeloma patients treated with high-dose therapy. *Blood* (2005) 106(8):2837–40. doi: 10.1182/blood-2005-04-1411

24. Leiba M, Duek A, Amariglio N, Avigdor A, Benyamini N, Hardan I, et al. Translocation t(11;14) in newly diagnosed patients with multiple myeloma: Is it always favorable? *Genes Chromosomes Cancer* (2016) 55(9):710–8. doi: 10.1002/gcc.22372

25. Gran C, Uttervall K, Borg Bruchfeld J, Wallblom A, Alici E, Gahrton G, et al. Translocation (11;14) in newly diagnosed multiple myeloma, time to reclassify this standard risk chromosomal aberration? *Eur J Haematol* (2019) 103(6):588–96. doi: 10.1111/ejh.1325

26. Gao W, Du J, Liu J, Zhou H, Zhang Z, Jian Y, et al. What multiple myeloma with t(11;14) should be classified into in novel agent era: Standard or intermediate risk? *Front Oncol* (2020) 10:538126. doi: 10.3389/fonc.2020.538126

27. Atrash S, Flahavan EM, Xu T, Ma E, Karve S, Hong WJ, et al. Treatment patterns and outcomes according to cytogenetic risk stratification in patients with multiple myeloma: a real-world analysis. *Blood Cancer J* (2022) 12(3):46. doi: 10.1038/s41408-022-00638-0

28. Smadbeck JB, Sharma N, Abdallah N, Tang H, Binder M, Fonseca R, et al. Characterization of atypical t(11;14) CCND1/IGH translocations in multiple myeloma. *Blood* (2021) 138(Supplement 1):3771. doi: 10.1182/blood-2021-150569

29. Gonsalves WI, Parmar H. Ageism in the t(11;14) subtype of multiple myeloma. Acta Haematol (2021) 144(1):6-7. doi: 10.1159/000508234

30. Avet-Loiseau H, Facon T, Grosbois B, Magrangeas F, Rapp MJ, Harousseau JL, et al. Oncogenesis of multiple myeloma: 14q32 and 13q chromosomal abnormalities are not randomly distributed, but correlate with natural history, immunological features, and clinical presentation. *Blood* (2002) 99(6):2185–91. doi: 10.1182/blood.v99.6.2185

31. López-Corral L, Gutiérrez NC, Vidriales MB, Mateos MV, Rasillo A, García-Sanz R, et al. The progression from MGUS to smoldering myeloma and eventually to multiple myeloma involves a clonal expansion of genetically abnormal plasma cells. *Clin Cancer Res* (2011) 17(7):1692–700. doi: 10.1158/1078-0432.ccr-10-1066

32. Glavey SV, Flanagan L, Bleach R, Kelly C, Quinn J, Chonghaile TN, et al. Secondary plasma cell leukaemia treated with single agent venetoclax. Br J Haematol (2020) 190(4):e242-e5. doi: 10.1111/bjh.16858

33. Jelinek T, Mihalyova J, Kascak M, Duras J, Popkova T, Benkova K, et al. Single-agent venetoclax induces MRD-negative response in relapsed primary plasma cell leukemia with t(11;14). *Am J Hematol* (2019) 94(1):E35-e7. doi: 10.1002/ajh.25331

34. Gonsalves WI, Buadi FK, Kumar SK. Combination therapy incorporating bcl-2 inhibition with venetoclax for the treatment of refractory primary plasma cell leukemia with t (11;14). *Eur J Haematol* (2018) 100(2):215–7. doi: 10.1111/ejh.12986

35. Hayman SR, Bailey RJ, Jalal SM, Ahmann GJ, Dispenzieri A, Gertz MA, et al. Translocations involving the immunoglobulin heavy-chain locus are possible early genetic events in patients with primary systemic amyloidosis. *Blood* (2001) 98 (7):2266–8. doi: 10.1182/blood.v98.7.2266

36. Kazandjian D, Hill E, Hultcrantz M, Rustad EH, Yellapantula V, Akhlaghi T, et al. Molecular underpinnings of clinical disparity patterns in African American vs. Caucasian American multiple myeloma patients. *Blood Cancer J* (2019) 9(2):15. doi: 10.1038/s41408-019-0177-9

37. Baughn LB, Pearce K, Larson D, Polley MY, Elhaik E, Baird M, et al. Differences in genomic abnormalities among African individuals with monoclonal gammopathies using calculated ancestry. *Blood Cancer J* (2018) 8(10):96. doi: 10.1038/s41408-018-0132-1

38. Marinac CR, Ghobrial IM, Birmann BM, Soiffer J, Rebbeck TR. Dissecting racial disparities in multiple myeloma. *Blood Cancer J* (2020) 10(2):19. doi: 10.1038/s41408-020-0284-7

39. Chakraborty R, Lentzsch S. Prognostic impact of t(11;14) in multiple myeloma: Black and white or shades of gray? *Cancer* (2020) 127(1):31-4. doi: 10.1002/cncr.33206

40. Garand R, Avet-Loiseau H, Accard F, Moreau P, Harousseau JL, Bataille R. t (11;14) and t(4;14) translocations correlated with mature lymphoplasmacytoid and immature morphology, respectively, in multiple myeloma. *Leukemia* (2003) 17 (10):2032–5. doi: 10.1038/sj.leu.2403091

41. Avet-Loiseau H, Garand R, Lodé L, Harousseau JL, Bataille R. Translocation t(11;14)(q13;q32) is the hallmark of IgM, IgE, and nonsecretory multiple myeloma variants. *Blood* (2003) 101(4):1570–1. doi: 10.1182/blood-2002-08-2436

42. Greenberg AJ, Rajkumar SV, Therneau TM, Singh PP, Dispenzieri A, Kumar SK. Relationship between initial clinical presentation and the molecular cytogenetic classification of myeloma. *Leukemia* (2014) 28(2):398-403. doi: 10.1038/leu.2013.258

43. Hoyer JD, Hanson CA, Fonseca R, Greipp PR, Dewald GW, Kurtin PJ. The (11;14)(q13;q32) translocation in multiple myeloma. a morphologic and immunohistochemical study. *Am J Clin Pathol* (2000) 113(6):831-7. doi: 10.1309/4w8e-8f4k-bhup-ube7

44. Touzeau C, Maciag P, Amiot M, Moreau P. Targeting bcl-2 for the treatment of multiple myeloma. *Leukemia* (2018) 32(9):1899–907. doi: 10.1038/ s41375-018-0223-9

45. Kaufman JL, Gasparetto C, Schjesvold FH, Moreau P, Touzeau C, Facon T, et al. Targeting BCL-2 with venetoclax and dexamethasone in patients with relapsed/refractory t(11;14) multiple myeloma. *Am J Hematol* (2021) 96(4):418–27. doi: 10.1002/ajh.26083

46. Gupta VA, Ackley J, Kaufman JL, Boise LH. BCL2 family inhibitors in the biology and treatment of multiple myeloma. *Blood Lymphat Cancer* (2021) 11:11–24. doi: 10.2147/blctt.s245191

47. Sonneveld P, Avet-Loiseau H, Lonial S, Usmani S, Siegel D, Anderson KC, et al. Treatment of multiple myeloma with high-risk cytogenetics: a consensus of the international myeloma working group. *Blood* (2016) 127(24):2955–62. doi: 10.1182/blood-2016-01-631200

48. Nooka AK, Lonial S. New targets and new agents in high-risk multiple myeloma. *Am Soc Clin Oncol Educ Book* (2016) 35:e431-41. doi: 10.1200/edbk_159516

49. Fonseca R, Blood E, Rue M, Harrington D, Oken MM, Kyle RA, et al. Clinical and biologic implications of recurrent genomic aberrations in myeloma. *Blood* (2003) 101(11):4569–75. doi: 10.1182/blood-2002-10-3017

50. Kaufman GP, Gertz MA, Dispenzieri A, Lacy MQ, Buadi FK, Dingli D, et al. Impact of cytogenetic classification on outcomes following early high-dose therapy in multiple myeloma. *Leukemia* (2016) 30(3):633–9. doi: 10.1038/leu.2015.287

51. Joseph NS, Kaufman JL, Dhodapkar MV, Hofmeister CC, Almaula DK, Heffner LT, et al. Long-term follow-up results of lenalidomide, bortezomib, and dexamethasone induction therapy and risk-adapted maintenance approach in newly diagnosed multiple myeloma. *J Clin Oncol* (2020) 38(17):1928–37. doi: 10.1200/jco.19.02515

52. Kaufman JL, Joseph NS, Gupta VA, Gleason S, Hofmeister CC, Heffner LT, et al. Outcomes of myeloma patients with t(11;14) receiving lenalidomide, bortezomib, and dexamethasone (RVD) induction therapy. *Blood* (2018) 132 (Supplement 1):3282. doi: 10.1182/blood-2018-99-119051

53. Badar T, Hari P, Dávila O, Fraser R, Wirk B, Dhakal B, et al. African Americans with translocation t(11;14) have superior survival after autologous hematopoietic cell transplantation for multiple myeloma in comparison with whites in the united states. *Cancer* (2020) 127(1):82–92. doi: 10.1002/cncr.33208

54. Takamatsu H, Yamashita T, Kurahashi S, Saitoh T, Kondo T, Maeda T, et al. Clinical implications of t(11;14) in patients with multiple myeloma undergoing autologous stem cell transplantation. *Biol Blood Marrow Transplant* (2019) 25 (3):474–9. doi: 10.1016/j.bbmt.2018.11.003

55. Saini N, Ma J, Milton DR, Patel R, Varma A, Bashir Q, et al. Impact of autologous transplantation in patients with multiple myeloma with t(11;14): A propensity-score matched analysis. *Clin Cancer Res* (2019) 25(22):6781–7. doi: 10.1158/1078-0432.ccr-19-0706

56. Boyd KD, Ross FM, Chiecchio L, Dagrada GP, Konn ZJ, Tapper WJ, et al. A novel prognostic model in myeloma based on co-segregating adverse FISH lesions

and the ISS: analysis of patients treated in the MRC myeloma IX trial. *Leukemia* (2012) 26(2):349–55. doi: 10.1038/leu.2011.204

57. Jian Y, Chen X, Zhou H, Zhu W, Liu N, Geng C, et al. Prognostic impact of cytogenetic abnormalities in multiple myeloma: A retrospective analysis of 229 patients. *Medicine (Baltimore)* (2016) 95(19):e3521. doi: 10.1097/md.00000000003521

58. Touzeau C, Dousset C, Le Gouill S, Sampath D, Leverson JD, Souers AJ, et al. The bcl-2 specific BH3 mimetic ABT-199: a promising targeted therapy for t (11;14) multiple myeloma. *Leukemia* (2014) 28(1):210–2. doi: 10.1038/leu.2013.216

59. Touzeau C, Ryan J, Guerriero J, Moreau P, Chonghaile TN, Le Gouill S, et al. BH3 profiling identifies heterogeneous dependency on bcl-2 family members in multiple myeloma and predicts sensitivity to BH3 mimetics. *Leukemia* (2016) 30 (3):761–4. doi: 10.1038/leu.2015.184

60. Punnoose EA, Leverson JD, Peale F, Boghaert ER, Belmont LD, Tan N, et al. Expression profile of BCL-2, BCL-XL, and MCL-1 predicts pharmacological response to the BCL-2 selective antagonist venetoclax in multiple myeloma models. *Mol Cancer Ther* (2016) 15(5):1132–44. doi: 10.1158/1535-7163.mct-15-0730

61. Bonolo de Campos C, Meurice N, Petit JL, Polito AN, Zhu YX, Wang P, et al. "Direct to drug" screening as a precision medicine tool in multiple myeloma. *Blood Cancer J* (2020) 10(5):54. doi: 10.1038/s41408-020-0320-7

62. Vaxman I, Sidiqi MH, Gertz M. Venetoclax for the treatment of multiple myeloma. *Expert Rev Hematol* (2018) 11(12):915-20. doi: 10.1080/17474086.2018.1548931

63. Joshi S, Gupta N, Khan R, Kumar R, Sharma M, Kumar L, et al. Interrelationship between angiogenesis, inflammation and oxidative stress in Indian patients with multiple myeloma. *Clin Transl Oncol* (2016) 18(2):132–7. doi: 10.1007/s12094-015-1344-5

64. Moser-Katz T, Joseph NS, Dhodapkar MV, Lee KP, Boise LH. Game of bones: How myeloma manipulates its microenvironment. *Front Oncol* (2020) 10:625199. doi: 10.3389/fonc.2020.625199

65. Puthier D, Derenne S, Barillé S, Moreau P, Harousseau JL, Bataille R, et al. Mcl-1 and bcl-xL are co-regulated by IL-6 in human myeloma cells. *Br J Haematol* (1999) 107(2):392–5. doi: 10.1046/j.1365-2141.1999.01705.x

66. Musolino C, Allegra A, Innao V, Allegra AG, Pioggia G, Gangemi S. Inflammatory and anti-inflammatory equilibrium, proliferative and antiproliferative balance: The role of cytokines in multiple myeloma. *Mediators Inflamm* (2017) 2017:1852517. doi: 10.1155/2017/1852517

67. Matthes T, Manfroi B, Huard B. Revisiting IL-6 antagonism in multiple myeloma. Crit Rev Oncol Hematol (2016) 105:1-4. doi: 10.1016/j.critrevonc.2016.07.006

68. Gupta VA, Barwick BG, Matulis SM, Shirasaki R, Jaye DL, Keats JJ, et al. Venetoclax sensitivity in multiple myeloma is associated with b-cell gene expression. *Blood* (2021) 137(26):3604–15. doi: 10.1182/blood.2020007899

69. Moreau P, Chanan-Khan A, Roberts AW, Agarwal AB, Facon T, Kumar S, et al. Promising efficacy and acceptable safety of venetoclax plus bortezomib and dexamethasone in relapsed/refractory MM. *Blood* (2017) 130(22):2392–400. doi: 10.1182/blood-2017-06-788323

70. Kumar SK, Harrison SJ, Cavo M, de la Rubia J, Popat R, Gasparetto C, et al. Venetoclax or placebo in combination with bortezomib and dexamethasone in patients with relapsed or refractory multiple myeloma (BELLINI): a randomised, double-blind, multicentre, phase 3 trial. *Lancet Oncol* (2020) 21(12):1630–42. doi: 10.1016/s1470-2045(20)30525-8

71. Kaufman JL, Baz RC, Harrison SJ, Quach H, Ho SJ, Vangsted AJ, et al. Updated analysis of a phase I/II study of venetoclax in combination with daratumumab and dexamethasone, ± bortezomib, in patients with relapsed/ refractory multiple myeloma. *J Clin Oncol* (2020) 38(15_suppl):8511. doi: 10.1200/JCO.2020.38.15_suppl.8511

72. Costa LJ, Davies FE, Monohan GP, Kovacsovics T, Burwick N, Jakubowiak A, et al. Phase 2 study of venetoclax plus carfilzomib and dexamethasone in patients with relapsed/refractory multiple myeloma. *Blood Adv* (2021) 5(19):3748–59. doi: 10.1182/bloodadvances.2020004146

73. Bahlis NJ, Baz R, Harrison SJ, Quach H, Ho SJ, Vangsted AJ, et al. Phase I study of venetoclax plus daratumumab and dexamethasone, with or without bortezomib, in patients with relapsed or refractory multiple myeloma with and without t(11;14). *J Clin Oncol* (2021) 39(32):3602–12. doi: 10.1200/ ico.21.00443

74. Kumar S, Kaufman JL, Gasparetto C, Mikhael J, Vij R, Pegourie B, et al. Efficacy of venetoclax as targeted therapy for relapsed/refractory t(11;14) multiple myeloma. *Blood* (2017) 130(22):2401–9. doi: 10.1182/blood-2017-06-788786

75. Gasparetto C, Bowles KM, Abdallah AO, Morris L, Mander G, Coppola S, et al. A phase II study of venetoclax in combination with pomalidomide and dexamethasone in Relapsed/Refractory multiple myeloma. *Clin Lymphoma Myeloma Leuk* (2021) 21(11):775–84. doi: 10.1016/j.clml.2021.07.029

76. Mateos MV, Moreau P, Dimopoulos MA, Hong WJ, Cooper S, Yu Y, et al. A phase III, randomized, multicenter, open-label study of venetoclax or pomalidomide in combination with dexamethasone in ptients with t(11;14)-positive Relapsed/Refractory multiple myeloma. *J Clin Oncol* (2020) 38 (15_suppl):TPS8554. doi: 10.1200/JCO.2020.38.15_suppl.TPS8554

77. Kumar S, Harrison SJ, Cavo M, de la Rubia J, Popat R, Gasparetto C, et al. Final overall survival results from BELLINI, a phase 3 study of venetoclax or placebo in combination with bortezomib and dexamethasone in Relapsed/ Refractory multiple myeloma. *Blood* (2021) 138(Supplement 1):84. doi: 10.1182/ blood-2021-145757

78. Basali D, Chakraborty R, Rybicki L, Rosko N, Reed J, Karam M, et al. Realworld data on safety and efficacy of venetoclax-based regimens in relapsed/ refractory t(11;14) multiple myeloma. *Br J Haematol* (2020) 189(6):1136–40. doi: 10.1111/bjh.16454

79. Sidiqi MH, Al Saleh AS, Kumar SK, Leung N, Jevremovic D, Muchtar E, et al. Venetoclax for the treatment of multiple myeloma: Outcomes outside of clinical trials. *Am J Hematol* (2021) 96(9):1131–6. doi: 10.1002/ajh.26269

80. Abuelgasim KA, Alherz N, Alhejazi A, Damlaj M. Venetoclax in combination with carfilzomib and dexamethasone in relapsed/refractory multiple myeloma harboring t(11,14)(q13;q32): two case reports and a review of the literature. J Med Case Rep (2020) 14(1):54. doi: 10.1186/s13256-020-02376-y

81. Maples KT, Nooka AK, Gupta V, Joseph NS, Heffner LT, Hofmeister C, et al. Natural history of multiple myeloma patients refractory to venetoclax: A single center experience. *Am J Hematol* (2021) 96(3):E68–e71. doi: 10.1002/ajh.26064

82. Bodet L, Gomez-Bougie P, Touzeau C, Dousset C, Descamps G, Maïga S, et al. ABT-737 is highly effective against molecular subgroups of multiple myeloma. *Blood* (2011) 118(14):3901–10. doi: 10.1182/blood-2010-11-317438

83. Gomez-Bougie P, Maiga S, Tessoulin B, Bourcier J, Bonnet A, Rodriguez MS, et al. BH3-mimetic toolkit guides the respective use of BCL2 and MCL1 BH3mimetics in myeloma treatment. *Blood* (2018) 132(25):2656–69. doi: 10.1182/ blood-2018-03-836718

84. Kitadate A, Terao T, Narita K, Ikeda S, Takahashi Y, Tsushima T, et al. Multiple myeloma with t(11;14)-associated immature phenotype has lower CD38 expression and higher BCL2 dependence. *Cancer Sci* (2021) 112(9):3645–54. doi: 10.1111/cas.15073

85. Ziccheddu B, Da Vià MC, Lionetti M, Maeda A, Morlupi S, Dugo M, et al. Functional impact of genomic complexity on the transcriptome of multiple myeloma. *Clin Cancer Res* (2021) 27(23):6479–90. doi: 10.1158/1078-0432.ccr-20-4366

86. Dai Y, Li X, Hu F, Li J, Liang Y, Wang D, et al. Identification of a novel drug sensitivity biomarker neuregulin-2 for venetoclax in non-t(11;14) multiple myeloma. *Blood* (2021) 138(Supplement 1):459–61. doi: 10.1182/blood-2021-150370

87. Matulis SM, Gupta VA, Neri P, Bahlis NJ, Maciag P, Leverson JD, et al. Functional profiling of venetoclax sensitivity can predict clinical response in multiple myeloma. *Leukemia* (2019) 33(5):1291-6. doi: 10.1038/s41375-018-0374-8

88. Neri P, Maity R, Alberge JB, Sinha S, Donovan J, Kong M, et al. Mutations and copy number gains of the BCL2 family members mediate resistance to venetoclax in multiple myeloma (MM) patients. *Blood* (2019) 134 (Supplement_1):572. doi: 10.1182/blood-2019-127593

89. Ghobrial I. BELLINI: a renaissance for an era of precision therapy in multiple myeloma. *Lancet Oncol* (2020) 21(12):1547-9. doi: 10.1016/s1470-2045 (20)30587-8