

### Research Article

# Investigation of *BTLA* tagging variants with risk of esophagogastric junction adenocarcinoma

Weifeng Tang<sup>1,\*</sup>, Shuchen Chen<sup>2,\*</sup>, Mingqiang Kang<sup>2</sup>, Jun Liu<sup>3</sup> and Chao Liu<sup>1</sup>

<sup>1</sup>Department of Cardiothoracic Surgery, Affiliated People's Hospital of Jiangsu University, Zhenjiang, Jiangsu Province, China; <sup>2</sup>Department of Thoracic Surgery, Fujian Medical University Union Hospital, Fuzhou, Fujian Province, China; <sup>3</sup>Department of Medical Oncology, Fujian Medical University Cancer Hospital and Fujian Cancer Hospital, Fuzhou, Fujian Province, China

Correspondence: Weifeng Tang (twf001001@126.com) or Shuchen Chen (cscdoctor@163.com)



**Background:** Variants in *B- and T-lymphocyte attenuator* (*BTLA*) gene are likely to affect the function of BTLA protein.

**Methods:** In the present case–control study, we selected *BTLA* tagging single-nucleotide polymorphisms (SNPs) (rs16859629 T>C, rs1982809 G>A, rs2171513 G>A and rs3112270 A>G) and conducted a case–control study to identify the association of *BTLA* SNPs with risk of esophagogastric junction adenocarcinoma (EGJA). The present study involved 1236 new incident EGJA cases and 1540 cancer-free controls.

**Results:** The genotypes of *BTLA* SNPs were analyzed using a SNPscan Kit. No association was also found between the *BTLA* SNPs and the susceptibility of EGJA in overall comparsion. In subgroup analyses, the *BTLA* rs1982809 was found to be associated with an increased susceptibility of EGJA (AA versus GG:  $OR^{adjusted} = 2.09$ , 95% CI 1.08–4.07, P = 0.030; and AA versus GA/GG:  $OR^{adjusted} = 1.99$ , 95% CI 1.04–3.82, P = 0.039). In haplotype comparison, we identified that TAAG haplotype with the order of *BTLA* rs16859629, rs1982809, rs2171513 and rs3112270 SNPs might increase the susceptibility of EGJA (OR = 3.07, 95% CI = 1.41–6.71; P = 0.003).

**Conclusion:** To conclude, the present study suggests that BTLA T<sub>rs16859629</sub>A<sub>rs1982809</sub> A<sub>rs2171513</sub>G<sub>rs3112270</sub> haplotype may increase the susceptibility of EGJA. More studies should be conducted to evaluate whether BTLA polymorphisms may influence the susceptibility of cancer in the future.

## Introduction

The morbidity of esophagogastric junction adenocarcinoma (EGJA) is promoting rapidly, both in developing and developed contries [1–3]. EGJA comprises a vital portion esophageal and gastric cancer, with an increasing ratio. It is reported that EGJA is a common fatal tumor in China. EGJA is regarded as an entity with a specific clinical feature and molecular profile. The potential protective factor or a real cause of EGJA is unclear. Thus, an understanding of the potential risk factors influencing the development EGJA biology may be helpful to diagnosis and prognostic assessment for the supervision of EGJA patients.

During the activation of T lymphocytes, they can express some receptors for receiving various signals. B- and T-lymphocyte attenuator (BTLA), also named CD272, is a most recently identified and studied member of the immune globulin (Ig) superfamily [4–7]. BTLA is a glycoprotein and it contains two tyrosine-based inhibitory motifs [8]. During activation, BTLA is not expressed on T helper type 2 (Th2) cells, but Th1 cells. The expression of BTLA on T cells participates in negative regulation of T cell and then leads to an decreased T-lymphocytes proliferation [9]. Recently, many investigations have focused on the relationship of BTLA with inflammation, autoimmune disease and cancer. Shi et al. reported that BTLA-herpes virus entry mediator (HVEM) checkpoint axis might be implicated in the regulation of inflammation in liver [10]. A previous study indicated that the up-regulation of BTLA gene expression and

\*These authors have contributed equally to this

Received: 29 May 2019 Revised: 08 November 2019 Accepted: 18 November 2019

Accepted Manuscript online: 27 November 2019 Version of Record published: 13 December 2019



soluble BTLA (sBTLA) was validated in thymoma-associated myasthenia gravis [11]. A prognostic investigation showed that the levels of immune checkpoints sBTLA could be considered as a biomarker for unresectable pancreatic adenocarcinoma cases with a poor survival [12]. A functional study identified that IFN- $\gamma$  level in circulating T-lymphocytes could be promoted by inhibiting BTLA/HVEM pathway [13]. Additionally, Feng et al. [14] and Lan et al. [15] reported that the level of BTLA expression in gastric carcinoma (GC) might be a useful biomarker for the evalution of GC prognosis.

Single-nucleotide polymorphisms (SNPs) in *BTLA* gene are likely to affect the role of BTLA protein. Some studies have kept a watchful eye on the correlation of *BTLA* variants with the development of cancer [16–18]. Fu et al. reported that the frequencies of *BTLA* rs1844089 and rs2705535 SNPs may alter the risk of breast cancer [17]. In Polish population, it was found that *BTLA* rs1982809 G>A, a 3'-UTR SNP, might be a low-penetrating risk factor for the development of renal cell carcinoma [18]. In addition, another study indicated that *BTLA* rs1982809 G and rs2705511 C alleles were more frequent in patients with chronic lymphocytic leukemia compared with healthy controls [16]. In view of the vital role in cancer development and progress, we supposed that *BTLA* SNPs might be correlated with EGJA susceptibility. Here, *BTLA* tagging SNPs (rs16859629 T>C, rs1982809 G>A, rs2171513 G>A and rs3112270 A>G) were selected. The aim of the present study was to identify the association of *BTLA* tagging SNPs with risk of EGJA.

# Materials and methods Subjects

The present study involved 1236 new incident EGJA patients and 1540 cancer-free controls. Among these patients, 393 cases patients diagnosed with EGJA and treated at two affiliated hospitals of Fujian Medical University [Union Hospital (Fuzhou, China) and Fujian Cancer Hospital (Fuzhou, China)] from January 2014 to June 2018. In addition, 843 patients with EGJA were from Jiangsu University People's Hospital (Zhenjiang, China) from January 2008 to June 2018. Siewert type was used in our study [19]. Here, all EGJA cases included were Siewert type II (their centre within 1 cm proximal and 2-cm distal of the anatomical cardia). All included EGJA cases were diagnosed at the first time with histopathological test. For EGJA cases, the major included criteria were: (a) individuals who did not have a history of other cancers, (b) without any immunological diseases and (c) EGJA patients were not treated with any chemotherapy and/or radiotherapy before the enrolment. We recruited 1540 cancer-free subjects as controls matching to the EGJA patients by sex, year of birth ( $\pm 5$  year) and ethnicity (Eastern Chinese Han nationality). They were from the hospitals mentioned above for regular health examination. The major included criteria for controls were: (a) cancer-free individuals, (b) without any immunological diseases, (c) sex and age matching to EGJA cases and (d) Han nationality who living in Eastern China. Each patitcipant signed a consent form. The experimental protocol was authorized by the ethics committees of the Jiangsu University.

### **Selection of SNPs**

The tagging SNPs of *BTLA* [from 112458030 to 112504757 in chromosome 3 (extending 5 Kb, upstream and downstream, respectively)] were structured and collected from Chinese populations via Genome Variation Server data. The criteria of tagging SNPs selection were described in our previous studies [20,21].

### **DNA** extraction

Genomic DNA was extracted from the colletced blood samples with the Promega DNA Kit (Promega, Madison, U.S.A.), according to the explanatory memorandum. A 2-µl DNA was droped in NanoDrop ND-1000 spectrophotometer (Wilmington, U.S.A.) to evaluate concentration and purity of DNA sample.

### Genotyping

The genotypes of *BTLA* rs16859629, rs1982809, rs2171513 and rs3112270 SNPs were analyzed using a SNPscan Kit (Genesky Biotechnologies Inc., Shanghai, China) as described previously [22–24]. PCR process was conducted in a 20-µl mixture volume in 96-well plates. ABI 3730xl DNA Analyzer was used to identify the genotype. The data of the sequencing were read by GeneMapper 4.1 (AppliedBiosystems, U.S.A.). One hundred and eleven DNA specimens were randomly chosen for repeat genotyping by another person in a blind fashion, and the obtained variants were concordant.



Table 1 Distribution of selected demographic variables and risk factors in this case-control study

Variable	Overall cases	(n = 1236)	Overall contr	ols (n = 1540)	$P^1$
	n	%	n	%	
Age (years)	64.28 ( <u>+</u> 8.64)		64.17 ( <u>+</u> 10.32)		0.775
Age (years)					0.408
< 64	568	45.95	732	47.53	
≥64	668	54.05	808	52.47	
Sex					0.485
Male	885	71.60	1084	70.39	
Female	351	28.40	456	29.61	
Smoking status					0.087
Never	884	71.52	1146	72.73	
Ever	352	28.48	394	27.27	
Alcohol use					<0.001
Never	1,028	83.17	1359	88.25	
Ever	208	16.83	181	11.75	

<sup>&</sup>lt;sup>1</sup>Two-sided  $\chi^2$  test and Student's t test.

### Statistical method

For each locus in BTLA gene, an online  $\chi^2$  test was used to assess the Hardy–Weinberg equilibrium (HWE) [25]. The Student t test was performed to deal with continuous variables of demographic characteristics between two groups. And  $\chi^2$  test was harnessed to handle the categorical variables (e.g., age, sex, cigarette using and alcohol consumption) and variant distributions of BTLA SNPs between two groups. The haplotypes of BTLA gene were evaluated by SHESIS software [26]. Odds ratios (ORs) and 95% confidence intervals (CIs) were used to evaluate the strength of the correlation of BTLA SNPs with the risk of EGJA. Multiple logistic regression analysis was harnessed to check the distribution of BTLA rs16859629, rs1982809, rs2171513 and rs3112270 genotypes between two groups. Subgroup analyses between the BTLA variants and characteristic variables were also conducted. The adjusted P values, ORs and 95% CIs were calculated by adjustment for age, sex, cigarette using and drinking. A P < 0.05 (two-way tests) was defined as significance in all statistical tests. All statistical analyses described previously were performed in SAS 9.4 software (SAS Institute Inc., Cary, NC, U.S.A.). Using PS software (http://biostat.mc.vanderbilt.edu/twiki/bin/view/Main/PowerSampleSize), the power value ( $\alpha$  = 0.05) was calculated [27,28]. We also used the false-positive report probability (FPRP) to determine the significant findings [29].

# Results

### **Baseline characteristics**

Table 1 summarizes age, sex, cigarette using and alcohol consumption in two groups. EGJA patients had a mean age of  $64.28 \pm 8.64$  years. The age and sex ratio was not significant between two groups (P = 0.408 and P = 0.485, respectively). The success rate of BTLA rs16859629, rs1982809, rs2171513 and rs3112270 genotyping was of high quality (>97%) (Table 2). We pressented the data of minor allele frequency (MAF) in Table 2. In control group, the frequencies of genotype distribution met HWE (Table 2).

# Relationship of *BTLA* rs16859629, rs1982809, rs2171513 and rs3112270 SNPs with EGJA

The genotype distributions and frequencies of BTLA rs16859629, rs1982809, rs2171513 and rs3112270 genotypes are presented in Table 3. In a single SNP analysis, the BTLA rs2171513 G>A genotype frequencies were 62.83% (GG), 32.67% (GA) and 4.50% (AA) in EGJA patients and 63.70% (GG), 32.27% (GA) and 4.03% (AA) in the cancer-free controls. When the BTLA rs2171513 GG genotype was defined as the reference, the BTLA rs2171513 GA genotype was not correlated with the susceptibility for EGJA (GA versus GG: adjusted OR = 1.04, 95% CI: 0.88–1.22, P = 0.668); the BTLA rs2171513 AA genotype was not correlated with the susceptibility for EGJA (AA versus GG: adjusted OR = 1.23, 95% CI: 0.83–1.81, P = 0.302). In addition, the BTLA rs2171513 GA/AA genotypes did not conferred the risk to EGJA in the dominant model (GA/AA versus GG: adjusted OR = 1.06, 95% CI: 0.90–1.24, P = 0.497). In the recessive genetic compared model, when the BTLA rs2171513 GG/GA genotypes were defined as a reference,



**Table 2** Primary information for *BTLA* targging SNPs (rs2171513 G>A, rs3112270 A>G, rs1982809 G>A and rs16859629 T>C)

Genotyped polymorphisms	rs2171513 G>A	rs3112270 A>G	rs1982809 G>A	rs16859629 T>C
Chr	3	3	3	3
Position_38	112466080	112461797	112463893	112471533
Region	3'-UTR	Promoter	3'-UTR	intron_variant
MAF <sup>1</sup> in database (1000 g Chinese Han populatons)	0.188	0.269	0.216	0.067
MAF in our controls ( $n = 1540$ )	0.196	0.280	0.256	0.084
P value for HWE <sup>2</sup> test in our controls	0.625	0.114	0.796	0.898
% Genotyping value	98.34%	98.56%	98.52%	97.48%

<sup>&</sup>lt;sup>1</sup>MAF, minor allele frequency.

Table 3 Logistic regression analyses of associations between *BTLA* targging SNPs (rs2171513 G>A, rs3112270 A>G, rs1982809 G>A and rs16859629 T>C) and the risk of EGJA

Genotype	EGJA case (n = 1236)		Controls ( <i>n</i> = 1540)		Crude OR (95%CI)	P	Adjusted OR <sup>1</sup> (95% CI)	P
	n	%	n	%	(667,661,	•	(55 /5 5.)	
rs2171513 G>A	4							
GG	754	62.83	985	64.38	1.00		1.00	
GA	392	32.67	489	31.96	1.05(0.89-1.23)	0.580	1.04(0.88-1.22)	0.668
AA	54	4.50	56	3.66	1.26(0.86-1.85)	0.241	1.23(0.83-1.81)	0.302
GA+AA	446	37.17	545	35.62	1.07(0.91-1.25)	0.404	1.06(0.90-1.24)	0.497
GG+GA	1146	95.50	1,474	96.34	1.00		1.00	
AA	54	4.50	56	3.66	1.24(0.85-1.82)	0.269	1.21(0.83-1.78)	0.327
A allele	500	20.83	601	19.64				
rs3112270 A>0	3							
AA	639	52.99	782	51.11	1.00		1.00	
AG	472	39.14	641	41.90	0.90(0.77-1.06)	0.197	0.90(0.77-1.06)	0.192
GG	95	7.88	107	6.99	1.09(0.81-1.46)	0.582	1.10 (0.82–1.48)	0.538
AG+GG	567	47.02	748	48.89	0.93(0.80-1.08)	0.330	0.93(0.80-1.08)	0.333
AA+AG	1111	92.13	1423	93.01	1.00		1.00	
GG	95	7.88	107	6.99	1.14(0.85-1.52)	0.380	1.15(0.86-1.53)	0.343
G allele	662	27.45	855	27.94				
rs1982809 G>A	4							
GG	668	55.44	846	55.29	1.00		1.00	
GA	461	38.26	586	38.30	1.00 (0.85-1.17)	0.964	1.00(0.85-1.17)	0.984
AA	76	6.30	98	6.41	0.98(0.72-1.35)	0.911	1.00(0.85-1.37)	0.980
GA+AA	537	44.56	684	44.71	0.99(0.85-1.16)	0.941	1.00(0.86-1.16)	0.979
GG+GA	1129	93.70	1432	93.59	1.00		1.00	
AA	76	6.30	98	6.41	0.98(0.72-1.34)	0.917	1.00(0.73-1.36)	0.983
A allele	613	25.44	782	25.56				
rs16859629 T>	·C							
П	1028	85.74	1265	83.94	1.00		1.00	
TC	158	13.18	231	15.33	0.84(0.68-1.05)	0.122	0.84(0.67-1.04)	0.106
CC	13	1.08	11	0.73	1.45(0.65–3.26)	0.363	1.39(0.62-3.13)	0.426
CT+CC	171	14.26	242	16.06	0.87(0.70-1.08)	0.197	0.86(0.70-1.07)	0.166
TT+CT	1186	98.92	1496	99.27	1.00		1.00	
CC	13	1.08	11	0.73	1.49(0.67-3.34)	0.332	1.43(0.64-3.21)	0.389
C allele	184	7.67	253	8.39				

 $<sup>^1</sup>$ Adjusted for age, sex, smoking, status of Chronic hepatitis B virus infection and drinking. Bold values are statistically significant (P < 0.05).

<sup>&</sup>lt;sup>2</sup>HWE, Hardy–Weinberg equilibrium.



Table 4 Stratified analyses between BTLA rs1982809 G>A polymorphism and EGJA risk by sex, age, smoking status and alcohol consumption

Variable	BTLA rs1	BTLA rs1982809 (Case/Control)1			ted OR <sup>2</sup> (95% CI); P				
	GG	GA	AA	GG	GA versus GG	AA versus GG	GA/AA versus GG	AA versus (GG/GA)	
Sex									
Male	488/605	328/412	49/61	1.00	0.99(0.82–1.20); <i>P</i> : 0.925	1.01(0.68–1.49); <i>P</i> : 0.981	0.99(0.83–1.19); <i>P</i> : 0.937	1.01(0.68–1.49); <i>P</i> : 0.966	
Female	180/241	133/174	27/37	1.00	1.02(0.75–1.37); <i>P</i> : 0.916	0.99(0.58–1.69); <i>P</i> : 0.983	1.01(0.76–1.34); <i>P</i> : 0.932	0.99(0.59–1.66); <i>P</i> : 0.962	
Age									
<64	304/391	205/287	40/51	1.00	0.92(0.73–1.17); <i>P</i> : 0.502	1.00(0.64–1.56); <i>P</i> : 0.996	0.93(0.75–1.17); <i>P</i> : 0.553	1.04(0.67–1.60); <i>P</i> : 0.876	
≥64	364/455	256/299	36/47	1.00	1.07(0.86–1.33); <i>P</i> : 0.545	0.97(0.61–1.53); <i>P</i> : 0.896	1.06 (0.86–1.30); <i>P</i> : 0.609	0.94(0.60–1.48); <i>P</i> : 0.801	
Smoking status	3								
Never	487/606	325/424	50/81	1.00	0.95(0.79–1.15); <i>P</i> : 0.600	0.78(0.54–1.14); <i>P</i> : 0.199	0.93(0.77-1.11); <i>P</i> : 0.392	0.80(0.56–1.15); <i>P</i> : 0.229	
Ever	181/240	136/162	26/17	1.00	1.13(0.83–1.54); <i>P</i> : 0.449	2.09(1.08–4.07); P: 0.030	1.22(0.91–1.64); <i>P</i> : 0.193	1.99(1.04–3.82); P: 0.039	
Alcohol consumption									
Never	563/737	378/524	63/90	1.00	0.95(0.80–1.13); <i>P</i> : 0.543	0.92(0.66–1.30); <i>P</i> : 0.639	0.94(0.80–1.11); <i>P</i> : 0.493	0.94(0.68–1.32); <i>P</i> : 0.725	
Ever	105/109	83/62	13/8	1.00	1.40(0.91–2.17); <i>P</i> : 0.126	1.56(0.61–3.99); <i>P</i> : 0.350	1.42(0.94–2.16); <i>P</i> : 0.098	1.37(0.54-3.44); <i>P</i> : 0.504	

<sup>&</sup>lt;sup>1</sup>The genotyping was successful in 1205 (97.49%) EGJA cases, and 1530 (99.35%) controls for BTLA rs1982809.

the *BTLA* rs2171513 AA genotype was not correlated with susceptibility for EGJA (AA versus GG/GA: adjusted OR = 1.21, 95% CI: 0.83-1.78, P = 0.327) (Table 3). No association was also found between *BTLA* rs3112270 A>G, rs1982809 G>A and rs16859629 T>C SNPs and the susceptibility of EGJA (Table 3).

# Relationship of *BTLA* rs16859629, rs1982809, rs2171513 and rs3112270 SNPs with EGJA in subgroup analysis

Table 4 presents the variant frequencies of BTLA rs1982809 SNP in stratification analysis. When we conducted an adjustment for gender, age and alcohol consumption, we identified that the BTLA rs1982809 G>A was associated with an increased susceptibility of EGJA for ever smokers (AA versus GG: adjusted OR = 2.09, 95% CI 1.08–4.07, P = 0.030; and AA versus GA/GG: adjusted OR = 1.99, 95% CI 1.04–3.82, P = 0.039). We found that there was no significant association between BTLA rs1982809 G>A SNP and the risk of EGJA in other subgroups.

No association was found between the BTLA rs2171513 G>A, rs3112270 A>G and rs16859629 T>C SNPs and the susceptibility of EGJA in subgroup analyses (data was not shown).

### **SNP** haplotypes

Using haplotype constructing software mentioned above [26], we observed 12 BTLA gene haplotypes. We identified that TAAG haplotype with the order of BTLA rs16859629, rs1982809, rs2171513 and rs3112270 SNPs might increase the susceptibility of EGJA (OR = 3.07, 95% CI = 1.41-6.71; P = 0.003). However, other observed BTLA gene haplotypes did not alter the susceptibility of EGJA (Table 5).

# **Power calculation and FPRP determining**

Using PS software (http://biostat.mc.vanderbilt.edu/twiki/bin/view/Main/PowerSampleSize), the power value ( $\alpha=0.05$ ) was calculated [27,28]. For BTLA rs1982809 G>A SNP, the power value was 0.631 in AA versus GG genetic model and 0.589 in AA versus GG/GA genetic model among ever smokers. In haplotype comparison,  $T_{rs16859629}A_{rs1982809}A_{rs2171513}G_{rs3112270}$  haplotype could increase the susceptibility of EGJA (power value, 0.830).

<sup>&</sup>lt;sup>2</sup>Adjusted for age, sex, smoking status and alcohol consumption (besides stratified factors accordingly) in a logistic regression model.



Table 5 BTLA haplotypes frequency (%) and the association between BTLA haplotypes and risk of EGJA

Haplotypes	Case		Control		Crude OR (95%0	I) <i>P</i>	
	n	%	n	%			
TGGA	1159	48.64	1459	48.41	Reference		
TAGG	407	17.08	518	17.19	0.99(0.85-1.15)	0.887	
TGAA	283	11.88	350	11.61	1.02(0.85-1.21)	0.843	
CGGA	136	5.71	175	5.81	0.98(0.77-1.24)	0.856	
TGAG	120	5.04	154	5.11	0.98(0.76-1.26)	0.88	
TGGG	71	2.98	105	3.48	0.85(0.62-1.15)	0.309	
TAGA	70	2.94	87	2.89	1.01(0.73-1.40)	0.938	
TAAA	69	2.9	79	2.62	1.10(0.79-1.53)	0.575	
CAGG	34	1.43	57	1.89	0.75(0.49-1.16)	0.192	
TAAG	22	0.92	9	0.3	3.07(1.41-6.71)	0.003	
CAGA	7	0.29	19	0.63	0.46(0.19-1.11)	0.076	
Others	5	0.21	2	0.07	3.15(0.61-16.26)	0.149	

With the order of BTLA rs16859629 T>C, rs1982809 G>A rs2171513 G>A and rs3112270 A>G in gene position.

# **Discussion**

The incidence of EGJA is increasing in both the East and Western countries. It is reported that altered lifestyle and lower chronic Helicobacter pylori infection may result in an increasing incidence of EGJA [30,31]. The etiology of EGJA may be attribute to gene and environment factors. Recent evidence suggested that the variants of immune and inflammatory response related genes could alter the risk of cancer [21,32–35]. Considering an important role of *BTLA* gene in immune, we chose *BTLA* tagging SNPs (rs16859629, rs1982809, rs2171513 and rs3112270) and explored their effects on the development of EGJA. Here, we identified that *BTLA* TAAG haplotype with the order of rs16859629, rs1982809, rs2171513 and rs3112270 SNPs might be associated with the development of EGJA.

BTLA rs1982809 G>A SNP locates in 3'-UTR, which could participate in post-transcriptional control. Recently, studies have been conducted to identify a potential effect of BTLA rs1982809 locus on the development of malignancy. BTLA rs1982809 polymorphism, a 3'-UTR SNP, was found to be associated with the development of renal cell carcinoma in Polish populations [18]. Another case–control study also found that BTLA rs1982809 polymorphism were associated with chronic lymphocytic leukemia [16]. Subsequently, in the same study, the funcional investigation demonstrated that the presence of BTLA rs1982809 G allele was correlated with lower expression of BTLA mRNA in lymphocyte as compared with rs1982809 A allele [16]. In the present study, we first studied the relationship between BTLA rs1982809 locus and cancer risk in Asians. We found this SNP might not alter the overall EGJA risk. However, BTLA rs1982809 locus was identified as a risk factor to EGJA in smoking subgroup, which was similar to the previous reports [16,18]. The results suggested that the role of BTLA rs1982809 G>A polymorphism may be influenced by environmental factors. However, the subjects included in smoking subgroup were related small, these findings may be underpowered. In the future, more case–control studies should be conducted to evaluate whether BTLA rs1982809 G>A polymorphism might inhibit the function of B and T cells and influence the susceptibility of cancer.

In the present case–control study, the BTLA haplotypes were also constructed. We found BTLA Trs16859629 Ars1982809 Ars1982809 Ars1982809 haplotype might influence the risk of EGJA. However, this rare BTLA haplotypes only altered the susceptibility of a minor fraction of the EGJA patients. We first expolre the association of BTLA haplotypes with cancer risk in Asians. Our findings should be verified in the future studies.

It is necessary to acknowledge the limitations in the present case—control study. First, the present study was designed as hospital-based. Although the frequencies of genotype distribution in *BTLA* rs16859629, rs1982809, rs2171513 and rs3112270 SNPs met HWE and the MAFs of these selected SNPs in control group were close to the database for Chinese, the bias might have happened. Second, we only included four risk factors (gender, age, smoking and alcohol consumption). And other potential environment factors (e.g. body mass index, intake of vegetable and fruit, education level and economic income) were not considered. Thus, the potential interactions between gene and these environment factors could not addressed. Third, the participants included were related small in some subgroups, the observations may be insufficient evidence to identify a relationship with a definitive power. Fourth, in the present study, the biological functions of *BTLA* SNPs were not studied. Finally, only four *BTLA* tagging SNPs (rs16859629, rs1982809, rs2171513 and rs3112270) were selected, which could not fully assess the total hereditary susceptibility in *BTLA* gene.



To conclude, this investigation suggests that BTLA T<sub>rs16859629</sub>A<sub>rs1982809</sub>A<sub>rs2171513</sub>G<sub>rs3112270</sub> haplotype may increase the susceptibility of EGJA. More studies with multiple environment factors should be carried out to evaluate whether BTLA variants may influence the susceptibility of cancer in the future.

### **Acknowledgments**

We appreciate all subjects who participated in this study.

### **Author Contribution**

All authors contributed significantly to this study. Conceived and designed the experiments: W.T., S.C., Performed the experiments: C.L., J.L., W.T., Analyzed the data: M.K., Contributed reagents/materials/analysis tools: S.C., Wrote the manuscript: M.K., W.T.

### **Competing Interests**

The authors declare that there are no competing interests associated with the manuscript.

### **Funding**

This study was supported in part by 333 Talent Training Project of Organization Department in Jiangsu Province [grant number BRA2017147] and Young and Middle-aged Talent Training Project of Health Development Planning Commission in Fujian Province [grant number 2016-ZQN-25].

#### **Abbreviations**

FPRP, false-positive report probability; MAF, minor allele frequency; SNP, single-nucleotide polymorphism.

### References

- 1 Bollschweiler, E., Wolfgarten, E., Gutschow, C. et al. (2001) Demographic variations in the rising incidence of esophageal adenocarcinoma in white males. Cancer 92, 549–555, https://doi.org/10.1002/1097-0142(20010801)92:3%3c549::AID-CNCR1354%3e3.0.C0;2-L
- 2 Blaser, M.J. and Saito, D. (2002) Trends in reported adenocarcinomas of the oesophagus and gastric cardia in Japan. Eur. J. Gastroenterol. Hepatol. 14, 107–113, https://doi.org/10.1097/00042737-200202000-00003
- 3 Zhou, Y., Zhang, Z., Zhang, Z. et al. (2008) A rising trend of gastric cardia cancer in Gansu Province of China. Cancer Lett. 269, 18–25, https://doi.org/10.1016/j.canlet.2008.04.013
- 4 Pasero, C. and Olive, D. (2013) Interfering with coinhibitory molecules: BTLA/HVEM as new targets to enhance anti-tumor immunity. *Immunol. Lett.* **151**, 71–75, https://doi.org/10.1016/j.imlet.2013.01.008
- 5 Carreno, B.M. and Collins, M. (2003) BTLA: a new inhibitory receptor with a B7-like ligand. *Trends Immunol.* **24**, 524–527, https://doi.org/10.1016/j.it.2003.08.005
- 6 Croft, M. (2005) The evolving crosstalk between co-stimulatory and co-inhibitory receptors: HVEM-BTLA. Trends Immunol. 26, 292–294, https://doi.org/10.1016/j.it.2005.03.010
- 7 Zeng, C., Wu, T., Zhen, Y. et al. (2005) BTLA, a new inhibitory B7 family receptor with a TNFR family ligand. Cell. Mole. Immunol. 2, 427–432
- 8 Watanabe, N., Gavrieli, M., Sedy, J.R. et al. (2003) BTLA is a lymphocyte inhibitory receptor with similarities to CTLA-4 and PD-1. *Nat. Immunol.* **4**, 670–679, <a href="https://doi.org/10.1038/ni944">https://doi.org/10.1038/ni944</a>
- 9 Wang, W.D., Gao, Y.C., Lu, Y.B. et al. (2017) BTLA-expressing CD11c antigen presenting cells in patients with active tuberculosis exhibit low capacity to stimulate T cell proliferation. *Cell. Immunol.* **311**, 28–35, https://doi.org/10.1016/j.cellimm.2016.09.015
- 10 Shi, W., Shao, T., Li, J.Y. et al. (2019) BTLA-HVEM Checkpoint Axis Regulates Hepatic Homeostasis and Inflammation in a ConA-Induced Hepatitis Model in Zebrafish. *J. Immunol.* **203**, 2425–2442, https://doi.org/10.4049/jimmunol.1900458
- 11 Xi, J., Wang, L., Yan, C. et al. (2019) The Cancer Genome Atlas dataset-based analysis of aberrantly expressed genes by GeneAnalytics in thymoma associated myasthenia gravis: focusing on T cells. *J. Thoracic Disease* 11, 2315–2323, https://doi.org/10.21037/jtd.2019.06.01
- 12 Bian, B., Fanale, D., Dusetti, N. et al. (2019) Prognostic significance of circulating PD-1, PD-L1, pan-BTN3As, BTN3A1 and BTLA in patients with pancreatic adenocarcinoma. *Oncoimmunology* **8**, e1561120, https://doi.org/10.1080/2162402X.2018.1561120
- 13 Liu, J., Li, J., He, M. et al. (2018) Distinct Changes of BTLA and HVEM Expressions in Circulating CD4(+) and CD8(+) T Cells in Hepatocellular Carcinoma Patients. *J. Immunol. Res.* **2018**, 4561571, https://doi.org/10.1155/2018/4561571
- 14 Feng, X.Y., Wen, X.Z., Tan, X.J. et al. (2015) Ectopic expression of B and T lymphocyte attenuator in gastric cancer: a potential independent prognostic factor in patients with gastric cancer. *Mole. Med. Rep.* **11**, 658–664, https://doi.org/10.3892/mmr.2014.2699
- 15 Lan, X., Li, S., Gao, H. et al. (2017) Increased BTLA and HVEM in gastric cancer are associated with progression and poor prognosis. *OncoTargets Ther.* **10**, 919–926, https://doi.org/10.2147/OTT.S128825
- 16 Karabon, L., Partyka, A., Jasek, M. et al. (2016) Intragenic Variations in BTLA Gene Influence mRNA Expression of BTLA Gene in Chronic Lymphocytic Leukemia Patients and Confer Susceptibility to Chronic Lymphocytic Leukemia. *Arch. Immunol. Ther. Exp. (Warsz.)* **64**, 137–145, https://doi.org/10.1007/s00005-016-0430-x



- 17 Fu, Z., Li, D., Jiang, W. et al. (2010) Association of BTLA gene polymorphisms with the risk of malignant breast cancer in Chinese women of Heilongjiang Province. *Breast Cancer Res. Treat.* **120**, 195–202, https://doi.org/10.1007/s10549-009-0462-6
- 18 Partyka, A., Tupikowski, K., Kolodziej, A. et al. (2016) Association of 3' nearby gene BTLA polymorphisms with the risk of renal cell carcinoma in the Polish population. *Urol. Oncol.* **34**, 419 e13–9, https://doi.org/10.1016/j.urolonc.2016.04.010
- 19 Siewert, J.R. and Stein, H.J. (1998) Classification of adenocarcinoma of the oesophagogastric junction. Br. J. Surg. 85, 1457–1459, https://doi.org/10.1046/j.1365-2168.1998.00940.x
- 20 Tang, W., Zhang, S., Qiu, H. et al. (2014) Genetic variations in MTHFR and esophageal squamous cell carcinoma susceptibility in Chinese Han population. *Med. Oncol.* **31**, 915, https://doi.org/10.1007/s12032-014-0915-6
- 21 Zou, C., Qiu, H., Tang, W. et al. (2018) CTLA4 tagging polymorphisms and risk of colorectal cancer: a case-control study involving 2,306 subjects. Onco Targets Ther. 11, 4609–4619, https://doi.org/10.2147/OTT.S173421
- 22 Zheng, L., Yin, J., Wang, L. et al. (2013) Interleukin 1B rs16944 G>A polymorphism was associated with a decreased risk of esophageal cancer in a Chinese population. *Clin. Biochem.* **46**, 1469–1473
- 23 Yin, J., Wang, L., Shi, Y. et al. (2014) Interleukin 17A rs4711998 A>G polymorphism was associated with a decreased risk of esophageal cancer in a Chinese population. *Dis. Esophagus* 27, 87–92
- 24 Qiu, H., Lin, X., Tang, W. et al. (2017) Investigation of TCF7L2, LEP and LEPR polymorphisms with esophageal squamous cell carcinomas. *Oncotarget* 8, 109107–109119. https://doi.org/10.18632/oncotarget.22619
- 25 Tang, W., Chen, S., Liu, J. et al. (2019) Investigation of IGF1, IGF2BP2, and IGFBP3 variants with lymph node status and esophagogastric junction adenocarcinoma risk. *J. Cell. Biochem.* **120**, 5510–5518, https://doi.org/10.1002/jcb.27834
- 26 Shi, Y.Y. and He, L. (2005) SHEsis, a powerful software platform for analyses of linkage disequilibrium, haplotype construction, and genetic association at polymorphism loci. *Cell Res.* **15**, 97–98, https://doi.org/10.1038/sj.cr.7290272
- 27 Tang, W., Qiu, H., Ding, H. et al. (2013) Association between the STK15 F31I polymorphism and cancer susceptibility: a meta-analysis involving 43,626 subjects. *PLoS One* **8**, e82790, https://doi.org/10.1371/journal.pone.0082790
- 28 Tang, W., Wang, Y., Pan, H. et al. (2019) Association of miRNA-499 rs3746444 A>G variants with adenocarcinoma of esophagogastric junction (AEG) risk and lymph node status. *OncoTargets Ther.* **12**, 6245–6252
- 29 He, J., Wang, M.Y., Qiu, L.X. et al. (2013) Genetic variations of mTORC1 genes and risk of gastric cancer in an Eastern Chinese population. *Mol. Carcinog.* **52**, E70–E79, https://doi.org/10.1002/mc.22013
- 30 Hasegawa, S., Yoshikawa, T., Cho, H. et al. (2009) Is adenocarcinoma of the esophagogastric junction different between Japan and western countries? The incidence and clinicopathological features at a Japanese high-volume cancer center. *World J. Surg.* **33**, 95–103, https://doi.org/10.1007/s00268-008-9740-4
- 31 Chung, J.W., Lee, G.H., Choi, K.S. et al. (2009) Unchanging trend of esophagogastric junction adenocarcinoma in Korea: experience at a single institution based on Siewert's classification. *Dis. Esophagus* 22, 676–681, https://doi.org/10.1111/j.1442-2050.2009.00946.x
- 32 Tang, W., Wang, Y., Chen, S. et al. (2016) Investigation of Cytotoxic T-lymphocyte antigen 4 Polymorphisms in Gastric Cardia Adenocarcinoma. *Scand. J. Immunol.* **83**, 212–218, https://doi.org/10.1111/sji.12409
- 33 Chen, S., Wang, Y., Chen, Y. et al. (2017) Investigation of Cytotoxic T-lymphocyte antigen-4 polymorphisms in non-small cell lung cancer: a case-control study. *Oncotarget* **8**, 76634–76643
- 34 Tang, W., Chen, S., Chen, Y. et al. (2017) Programmed death-1 polymorphisms is associated with risk of esophagogastric junction adenocarcinoma in the Chinese Han population: A case-control study involving 2,740 subjects. *Oncotarget* 8, 39198–39208
- 35 Zhu, J., Liu, C., Teng, X. et al. (2016) Association of the interleukin-18 receptor 1 and interleukin-18 receptor accessory protein polymorphisms with the risk of esophageal cancer. *Biomed. Rep.* **4**, 227–235, https://doi.org/10.3892/br.2015.552