



Article Bifunctional 3-Hydroxy-4-Pyridinones as Potential Selective Iron(III) Chelators: Solution Studies and Comparison with Other Metals of Biological and Environmental Relevance

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Abstract:** The binding ability of five bifunctional 3-hydroxy-4-pyridinones towards Cu^{2+} and Fe^{3+} was studied by means of potentiometric and UV–Vis spectrophotometric measurements carried out at I = 0.15 mol L⁻¹ in NaCl_(aq), T = 298.15 K and 310.15 K. The data treatments allowed us to determine speciation schemes featured by metal-ligand species with different stoichiometry and stability, owing to the various functional groups present in the 3-hydroxy-4-pyridinones structures, which could potentially participate in the metal complexation, and in the Cu²⁺ and Fe³⁺ behaviour in aqueous solution. Furthermore, the sequestering ability and metal chelating affinity of the ligands were investigated by the determination of $pL_{0.5}$ and pM parameters at different pH conditions. Finally, a comparison between the Cu²⁺ and Fe³⁺/3-hydroxy-4-pyridinones data herein presented with those already reported in the literature on the interaction of Zn²⁺ and Al³⁺ with the same ligands showed that, from the thermodynamic point of view, the 3-hydroxy-4-pyridinones are particularly selective towards Fe³⁺ and could therefore be considered promising iron-chelating agents, also avoiding the possibility of competition, and eventually the depletion, of essential metal cations of biological and environmental relevance, such as Cu²⁺ and Zn²⁺.

Keywords: iron-chelation; chelating agents; 3-hydroxy-4-pyridinone; Fe-complexation; Cu-interaction; sequestering ability

1. Introduction

Copper (Cu) and iron (Fe) are essential metals for plants, animals and humans, ensuring their normal biochemical and physiological functions [1,2]. In healthy situations, living organisms are provided with homeostatic mechanisms and buffers to keep normal metal concentration levels and to avoid anomalous phenomena, as metal decompartmentalization, release and mobilization [3].

In plants, Cu plays important roles in photosynthetic and respiratory electron transport processes, occurring in chloroplasts and mitochondria. It participates in the oxidative stress protection, acts as cofactor of many enzymes and plays key functions in the cell wall metabolism, namely for Fe-mobilization, oxidative phosphorylation and the biogenesis of the molybdenum cofactor [4]. In the human body, copper favours the normal development of the brain and nervous system and maintains a fair level of white blood cells. Cu is also necessary to keep the muscle tone and functions; it is involved in the formation of red blood cells and in the processes of absorption and transport of iron (Fe³⁺) in the body. Furthermore, the generation of cellular energy in the form of ATP into the mitochondria depends on the participation of a copper-containing enzyme.

As regards iron, in living organisms it plays essential functions in metabolic processes, like photosynthesis, respiration and DNA synthesis [5]. In plants, mainly present in ferric (Fe^{3+}) form [6], it participates in the chlorophyll production [7], being also necessary for nitrogen fixation processes and for plants growth [8]. It is detected in iron-containing hemeproteins, like cytochromes present in electron transfer systems within the mitochondria and chloroplasts, and also in non-heme proteins, as ferredoxin. Iron can become a toxic element for soils and plants when it is accumulated at significant concentration levels. In these matrices, Fenton reaction (iron redox cycle) can produce reactive oxygen species (ROS) like -OH radicals [3], which are able to damage DNA, lipids and proteins so that effects like soils bronzing and leaves stippling may occur. As an example, the reason for colour alterations in leaves could be the plants' synthesis of enzymes aimed to control the free radicals effects, like in the case of basil, tomato, impatiens and phlox plants [8,9]. In addition, Fe³⁺ could also compete with Cu²⁺ and Zn²⁺ in its uptake and transport within plant cells [10]. In humans and other mammals, iron is a fundamental constituent of myoglobin and haemoglobin, proteins able to transport oxygen along the body. It is also important for the normal functions of hormones and cells, also being the cofactor of many enzymes, like in cytochromes B5, C and P450 [8]. Similarly to what was observed in plants and soils, in humans and mammals iron excess can also be very toxic. In fact, a Fenton reaction produces hydroxyl radicals which can react with nucleic acids, proteins, sugars and lipids, leading to pathological situations such as DNA and RNA damage, proteins and sugars oxidation and lipid peroxidation, respectively. Unfortunately, since physiological mechanisms for iron elimination do not exist, acute and chronic effects due to its overload can be often observed [3]. Examples of harmful effects provoked by iron accumulation are the deterioration of the gastric and intestinal mucosae, Bantu siderosis, cardiovascular diseases, neurodegenerative disorders and carcinogenic risks [8]. Iron overload could also cause haemoglobinopathy diseases such as transfusional hemosiderosis, owing to the metal parenteral administration for the treatment of β -thalassemia major, or hemochromatosis, a genetic disorder correlated with iron over-absorption [3].

In this light, many research efforts have been devoted to the development of new strategies aimed to remove metals from different matrices without side effects. All the green and sustainable approaches developed for metals extraction from environmental matrices are called "*chelation technologies*" [11], whereas the treatment of human diseases related to the metal intoxication is called "*chelation therapy*". This last approach is based on the administration of chelating agents to patients suffering from metals overload, inducing their sequestration and systemic excretion [12,13]. As regards iron chelation, potential chelating agents should fulfil some criteria, such as: absence of toxicity of the chelators and the corresponding iron-complexes, economic availability, drug-likeness properties, selectivity towards the metal cation of interest without involving the depletion of essential components, good intestinal absorption, oral activity, affinity towards biological membranes, good bioavailability to the target cells, higher metal chelating capacity and specificity with respect to the commercial molecules [14].

The bidentate chelators 3-hydroxy-4-pyridinones (3,4-HPs) match these criteria and therefore this family of compounds have been considered as promising drug candidates. They are characterized by an aromatoid *N*-heterocyclic ring, containing an exocyclic pair of electron donor atoms (*O*-*O*), featured by a ketone and a hydroxyl substituent groups in the *ortho* position, which confers them a high affinity towards divalent and trivalent metal cations [15]. In fact, a 3,4-HP derivative, namely the 1,2-dimethyl-3-hydroxy-4-pyridinone, (Deferiprone, *DFP*) is approved as an orally active chelating drug for the treatment of iron overload patients. Since its disclosure and later approval [16,17], many 3,4-HP derivatives, have been developed with the aim of overcoming some *DFP* drawbacks and efficacy improving [3]. Following this strategy, we have recently explored a small family of compounds, namely bifunctional bidentate 3,4-HP ligands, with the aim of improving their lipophilic–hydrophilic balance, bioavailability and affinity towards biological membranes,

as well as their chelating efficacy towards Fe^{3+} with respect to the commercially available chelating agents [13,18].

Herein, pursuing our previous strategy, we present the results of a potentiometric and UV–Vis (Ultraviolet–Visible) spectrophotometric investigation on the interaction of five bifunctional 3-hydroxy-4-pyridinones (Figure 1) with Cu^{2+} and Fe^{3+} , metal cations with a *borderline* and a *hard* character, respectively, that are carried out at I = 0.15 mol L⁻¹ in NaCl_(aq), T = 298.15 K and 310.15 K. Furthermore, the obtained thermodynamic data are compared with those already reported in the literature on the binding ability and chelating affinity of the five ligands towards Zn²⁺ [19] and Al³⁺ [13], also featured by *borderline* and *hard* behaviour, respectively, at the same experimental conditions. The aim of this work was to evaluate whether the 3-hydroxy-4-pyridinones under study could be exploited as selective chelating agents for the treatment of Fe³⁺ overload in humans or, alternatively, in environmental matrices. Another relevant issue worth investigating was to ascertain whether, from a thermodynamic point of view, along with an effective Fe³⁺-sequestration, a significant competition, and possibly depletion, of divalent metals of biological and environmental relevance such as Cu²⁺ and Zn²⁺ may occur, despite the different charge density, acid–base behaviour [20–22] and ionic radius [23].



Figure 1. Structures of the 3-hydroxy-4-pyridinone ligands, with the protonable groups enclosed in dotted rectangles.

Along the text, in the tables and figures, the five 3-hydroxy-4-pyridinones under study will be indicated with the abbreviations:

 $H_2(L1) = 4-(3-hydroxy-2-methyl-4-oxopyridin-1(4H)-yl)butanoic acid;$

 $H_2(L2) = (S)-2-amino-4-((2-(3-hydroxy-2-methyl-4-oxopyridin-1(4H)-yl)ethyl)amino)-4-oxobutanoic acid;$

 $H_2(L3) = (S)-2-amino-4-((3-(3-hydroxy-2-methyl-4-oxopyridin-1(4H)-yl)propyl)amino)-4-oxobutanoic acid;$

 $H_2(L4) = (S)$ -2-amino-5-(3-hydroxy-2-methyl-4-oxopyridin-1(4*H*)-yl)pentanoic acid; H(L5) = 1-(3-aminopropyl)-3-hydroxy-2-methylpyridin-4(1*H*)-one.

2. Results and Discussion

2.1. Equilibria for the Formation of Metal-Ligand Species

The formation or stability constants of the metal-ligand species are expressed considering the following stepwise (1) and overall (2) equilibria:

$$pM^{n+} + H_rL_q^{-(zq-r)} = M_pH_rL_q^{(pn+r-qz)} K_{pqr}$$
 (1)

$$pM^{n+} + qL^{z-} + rH^{+} = M_pH_rL_q^{(pn+r-qz)} \qquad \beta_{pqr}$$
 (2)

The equilibrium constants, concentrations and ionic strengths are expressed in the molar (c, mol L⁻¹) concentration scale.

2.2. Synthesis of the Ligands

The five 3-hydroxy-4-pyridinones (Figure 1) have been synthesized and characterized in the neutral form (H_rL^0), following procedures already reported in the literature [13].

2.3. Acid–Base Properties of Ligands and the Metal Cations

The 3-hydroxy-4-pyridinones under study are featured by different protonable groups highlighted in Figure 1 with dotted rectangles. The ligands' structure consists of a hydroxyl group as substituent on the *N*-heterocyclic ring, a -NH₂ and/or -COOH on the alkyl chain and a pyridinone nitrogen atom (proton provided by an excess of inorganic acid) [13], each of them with different acidity. The 3-hydroxy-4-pyridinones protonation constants have been already reported in the literature at I = 0.15 mol L⁻¹ in NaCl_(aq), T = 298.15 and 310.15 K (Table S1) [13].

The hydrolytic constants of Cu^{2+} and Fe^{3+} have already been published [20–22]. In the case of Fe^{3+} , the solubility product related to the formation of $Fe(OH)_3^{0}{}_{(s)}$ sparingly soluble species has been also considered [21].

2.4. Metal-Ligand Studies

The elaboration of potentiometric and UV–Vis spectrophotometric data on the binding ability of the ligands towards Cu^{2+} and Fe^{3+} allowed us to determine various speciation schemes, based on the different acid–base properties of the 3,4-HPs in $NaCl_{(aq)}$, the hydrolytic behaviour and the charge density of the metal cations. The best possible speciation models were selected on the basis of criteria such as the simplicity and probability of the model, the species formation percentages in the whole investigated pH, the statistical parameters (like the standard deviation on equilibrium constants and on the fitting values), the corresponding ratios between single variances compared with those from the accepted model. The high number of experiments carried out and experimental points collected allowed for the consideration of differences in variance between the accepted model and others to be significant.

In the case of the Cu²⁺ and Fe³⁺/(3,4-HPs) interactions investigated with both of the mentioned analytical techniques, an average of the potentiometric and UV–Vis stability constants was calculated with the aim of describing the systems in a more complete way, considering a wide range of metal and ligand concentrations used, namely $c \sim 10^{-3}$ mol L⁻¹ and $\sim 10^{-4}$ – 10^{-5} mol L⁻¹, for potentiometric and UV–Vis spectrophotometric measurements, respectively.

2.4.1. Cu²⁺/(3,4-HPs) Systems

For each investigated $Cu^{2+}/(3,4-HPs)$ system, the treatment of potentiometric and UV– Vis spectrophotometric data recorded at $I = 0.15 \text{ mol } L^{-1}$ in NaCl_(aq), T = 298.15 K and pH ranges 2.0-10.0 and 2.0-11.0, respectively, allowed us to obtain speciation models featured by complex species with 1:1 stoichiometry ($CuL^{(2-z)}$) and different protonation degrees $(CuH_2L^{(4-z)}, CuHL^{(3-z)})$. The experimental formation constants obtained by each analytical techniques are in accordance with each other, and they are reported in Table 1. As can be observed in Table 1, at the mentioned experimental conditions a trend of the complexes' stability can be observed, based on the common species $CuL^{(2-z)}$: $Cu(L3)^{0}_{(aq)} > Cu(L4)^{0}_{(aq)}$ $> Cu(L2)^{0}_{(aq)} > Cu(L5)^{+} > Cu(L1)^{0}_{(aq)}$. This trend could be explained considering that the stability of the $Cu^{2+}/(3,4-HPs)$ species may be favoured by the concomitant presence of the extra-functional groups in the 3,4-HP ligand molecules, namely -COOH, -NH₂ and -CHNH₂COOH bearing groups in the alkyl chain bound to the *N*-heterocyclic ring, which, in some cases have also inserted an amide moiety $(H_2(L2), H_2(L3))$ (Figure 1). Generally, the complexes with higher stability are those where the 3,4-HP ligands are extra-functionalized with α -amino-carboxylic groups (H₂(L2), H₂(L3), H₂(L4)), probably due to their inherent chelating capacity [24]. The different length of the alkyl moiety is also another factor influencing the stability of the species; in fact, from the comparison between the data obtained for $Cu(L2)^{0}_{(aq)}$ and $Cu(L3)^{0}_{(aq)}$ species, which only differ in the ligand structures

by an additional -CH₂ group present in the H₂(*L*3) alkyl chain (Figure 1), a decrease of the formation constants with alkyl moiety length decreasing can be observed (Table 1). Furthermore, from the comparison among the Cu(*L*1)⁰_(aq) and Cu(*L*5)⁺ stability constants it can be observed that the ligand featured by the only amino group (H(*L*5)) in the alkyl chain forms Cu²⁺ complexes with higher stability than the carboxylic-3-hydroxy-4-pyridinone (H₂(*L*1)), a trend which is in accordance with data reported in the literature [24–26] on the interactions of alkylamines and carboxylic acids towards Cu²⁺, also following the Pearson's principle of "*hard* and *soft* acids and bases" theory (HSAB) for ligand-metal preferences [27–29].

Table 1. Experimental stability constants ¹ of	Cu ²⁺ /3-hydroxy-4-pyridinone species obtained by
different analytical techniques at $I = 0.15$ mol L	$^{-1}$ in NaCl _(aq) , $T = 298.15$ K and $p = 0.1$ MPa.

Species	Potentiometry	logβ _{pqr} (logK _{pqr}) UV–Vis Spectrophotometry	Average Stability Constants ²
$CuH(L1)^+$ $Cu(L1)^0_{(aq)}$	$\begin{array}{c} 14.23 \pm 0.04 \ ^3 \ (4.28) \\ 9.76 \pm 0.06 \end{array}$	- -	- -
$CuH_2(L2)^{2+}$ $CuH(L2)^+$ $Cu(L2)^0_{(aq)}$	$\begin{array}{c} 24.98 \pm 0.02 \ ^3 \ (5.46) \\ 21.75 \pm 0.03 \ (11.02) \\ 15.10 \pm 0.10 \end{array}$	$\begin{array}{c} 24.93 \pm 0.05 \ ^3 \ (5.41) \\ 21.00 \pm 0.04 \ (10.27) \\ 14.48 \pm 0.105 \end{array}$	$\begin{array}{c} 24.95 \pm 0.07\ ^{4} \ (5.43) \\ 21.37 \pm 0.30 \ (10.64) \\ 14.79 \pm 0.26 \end{array}$
$CuH_2(L3)^{2+}$ $Cu(L3)H^+$ $Cu(L3)^0_{(aq)}$	$\begin{array}{c} 26.93 \pm 0.02 \ ^3 \ (6.26) \\ 22.84 \pm 0.03 \ (11.85) \\ 15.95 \pm 0.03 \end{array}$	- - -	- - -
$CuH_2(L4)^{2+}$ $CuH(L4)^+$ $Cu(L4)^0_{(aq)}$	$\begin{array}{c} 25.65 \pm 0.02 \ ^3 \ (5.21) \\ 21.63 \pm 0.03 \ (10.53) \\ 15.62 \pm 0.07 \end{array}$	$\begin{array}{c} 25.66 \pm 0.04 \ ^3 \ (5.22) \\ 21.78 \pm 0.02 \ (10.68) \\ 15.62 \pm 0.05 \end{array}$	$\begin{array}{c} 25.65 \pm 0.01\ ^{4} \ (5.21) \\ 21.70 \pm 0.04 \ (10.60) \\ 15.62 \pm 0.06 \end{array}$
CuH(<i>L5</i>) ²⁺ Cu(<i>L5</i>) ⁺	$20.00 \pm 0.03 \ {}^3 \ (8.92) \\ 12.62 \pm 0.06$	$\begin{array}{c} 20.241 \pm 0.006 \ ^3 \ (9.161) \\ 12.67 \pm 0.04 \end{array}$	$\begin{array}{c} 20.12 \pm 0.09 \ ^4 \ (9.04) \\ 12.64 \pm 0.04 \end{array}$

¹ logβ_{pqr} and logK_{pqr} refer to Equations (2) and (1), respectively; ² data obtained by an average of potentiometric and UV–Vis spectrophotometric data; ³ ±Std. Dev.; ⁴ errors on weighed data. Standard uncertainties: u(T) = 0.1 K; u(I) = 0.01 mol L⁻¹.

A further comparison between the speciation of the different Cu²⁺/3-hydroxy-4pyridinone systems may be performed based on the distribution diagrams reported in Figure 2, for H₂(*L*3), and Figure S1, for the other ligands. In the case of H₂(*L*1) (Figure S1a), the diagram shows that the formation of the CuH(*L*1)⁺ and Cu(*L*1)⁰_(aq) species reaches the 68% and 99% maximum percentages at pH ~ 3.9 and pH ~ 6.6, respectively. As regards the distribution of Cu²⁺/H₂(*L*2) (Figure 2), H₂(*L*3) (Figure S1b) and H₂(*L*4) (Figure S1c) species, the metal-ligands complexation occurs up to pH ~ 3.2–3.3 with the formation of CuH₂*L*²⁺ species exceeding the 52% formation. The CuH*L*⁺ complex achieves more than the 86% formation at pH ~ 5.0–5.1 for H₂(*L*2) and H₂(*L*4), pH ~ 5.5 for H₂(*L*3). The 1:1 stoichiometry complex starts to form at pH ~ 4.0, 4.5 and 3.6 and reaches more than the 99% formation at pH ~ 8.8, 9.1 and 8.2, for H₂(*L*2), H₂(*L*3) and H₂(*L*4), respectively. In the case of the Cu²⁺/H(*L*5) system (Figure S1d), the formation of the Cu(*L*5)H²⁺ and Cu(*L*5)⁺ species reaches their maximum percentages at pH ~ 5.4 and pH ~ 9.6, respectively.

Concerning the UV–Vis spectrophotometric behaviour, a representative example of the spectra recorded for the Cu²⁺/H₂(*L*2) system is shown in Figure 3a. An absorption band with $\lambda_{max} = 278$ nm can be observed at pH ~ 2.0–2.5. Its intensity increases with the pH, up to pH ~ 3.5, and then it starts to decrease, up to pH ~ 9.0–9.7, undergoing a bathochromic shift. Above pH ~ 10.0, the band intensity raises again with a subsequent red shift, up to pH ~ 11.0. The deconvolution of the UV–Vis data allowed us to calculate the molar absorptivity (ε/L (mol⁻¹ cm⁻¹)) values for each metal-ligand species. As an example, a graphical representation of the ε determined for the Cu²⁺/H₂(*L*2) system is reported in Figure 3b. At *I* = 0.15 mol L⁻¹ in NaCl_(aq), *T* = 298.15 K, the molar absorptivities are: $\varepsilon_{max}(CuH_2(L2)^{2+}) = 7514$ at $\lambda_{max} = 280$ nm, $\varepsilon_{max}(CuH(L2)^+) = 7813$ at $\lambda_{max} = 302$ nm, $\varepsilon_{max}(Cu(L2)_{(aq)}) = 5222$ at $\lambda_{max} = 304$ nm.



Figure 2. Distribution diagram of the Cu²⁺/H₂(*L*2) system at *T* = 298.15 K, *I* = 0.15 mol L⁻¹ in NaCl_(aq), $c_{Cu}^{2+} = 5.0 \cdot 10^{-4}$ mol L⁻¹ and $c_{ligand} = 1.5 \cdot 10^{-3}$ mol L⁻¹.



Figure 3. (a) UV–Vis absorption profile of the Cu²⁺/H₂(*L*2) system at different pH values and (b) calculated molar absorptivity of CuH₂(*L*2)²⁺, CuH(*L*2)⁺, Cu(*L*2)⁰_(aq) species at *T* = 298.15 K, $I = 0.15 \text{ mol } L^{-1}$ in NaCl_(aq), $c_{Cu}^{2+} = 2.0 \cdot 10^{-5} \text{ mol } L^{-1}$ and $c_{\text{ligand}} = 8.5 \cdot 10^{-5} \text{ mol } L^{-1}$.

2.4.2. Fe³⁺/(3,4-HPs) Systems

The investigation on the binding ability of the H₂(*L*1), H₂(*L*2), H₂(*L*4) and H(*L*5) ligands towards Fe³⁺ was carried out by potentiometric titrations at *I* = 0.15 mol L⁻¹ in NaCl_(aq) and *T* = 298.15 K. In the case of H₂(*L*2) and H(*L*5), UV–Vis experiments were performed also at the same ionic strength and *T* = 310.15 K. The data treatment allowed for the determination of FeHL^(4-z), FeL^(3-z) and FeL₂^(3-2z) species in the pH range 2.0–5.0, due to the formation of a red colour precipitate, attributable to the sparingly soluble Fe(OH)₃⁰_(s) species [21]. This limitation was overcome by spectrophotometric titrations performed at more diluted conditions, thus allowing to explore the measurements in a wider pH range (2.0–9.1).

The stability constants determined at the different experimental conditions are reported in Table 2. The values obtained by the two analytical techniques are in quite good agreement. Similarly to the $Cu^{2+}/(3,4-HPs)$ studies, in this case the data average was also calculated.

Table 2. Experimental stability constants ¹ of Fe³⁺/(3,4-HP) species obtained by different analytical techniques at I = 0.15 mol L⁻¹ in NaCl_(aq), different temperatures and p = 0.1 MPa.

		$\log \beta_{pqr}$ (lo		
Species	Potentiometry	<i>T</i> = 298.15 K UV–Vis Spectrophotometry	Average Stability Constants ²	UV-Vis Spectrophotometry
FeH($L1$) ²⁺ Fe($L1$) ⁺ Fe($L1$) ₂ ⁻	$\begin{array}{c} 17.37 \pm 0.10 \ {}^3(7.42) \\ 13.23 \pm 0.19 \\ 22.52 \pm 0.20 \ (9.29) \end{array}$		- - -	- - -
FeH($L2$) ²⁺ Fe($L2$) ⁺ Fe($L2$) ₂ ⁻	$\begin{array}{c} 26.16 \pm 0.03 \ ^3(15.43) \\ 22.06 \pm 0.06 \\ 38.01 \pm 0.04 \ (15.95) \end{array}$	$\begin{array}{c} 25.91 \pm 0.04^3 (15.18) \\ 21.78 \pm 0.03 \\ 38.01 {}^5 (16.23) \end{array}$	$\begin{array}{c} 26.03 \pm 0.10 \ {}^{4}(15.30) \\ 21.97 \pm 0.11 \\ 38.01 \pm 0.11(16.04) \end{array}$	$\begin{array}{c} 26.23 \pm 0.01 \ {}^3(15.24) \\ 22.86 \pm 0.04 \\ 39.79 \pm 0.08 \ (16.83) \end{array}$
$FeH(L4)^{2+}$ $Fe(L4)^{+}$ $Fe(L4)_2^{-}$	$\begin{array}{c} 26.31 \pm 0.02\ {}^3(15.21) \\ 22.48 \pm 0.03 \\ 39.08 {\pm} 0.03\ (16.60) \end{array}$	- - -	- - -	- - -
FeH(<i>L5</i>) ³⁺ Fe(<i>L5</i>) ²⁺	$25.05 \pm 0.04 \ {}^3(13.97) \\ 20.93 \pm 0.03$	$\begin{array}{c} 24.85 \pm 0.04 \ {}^{3} (13.77) \\ 20.30 \pm 0.03 \end{array}$	24.95±0.15 ⁴ (13.87) 20.61±0.27	$\begin{array}{c} 25.35 \pm 0.04 \ {}^3(14.75) \\ 21.16 \pm 0.08 \end{array}$

¹ log β_{pqr} and log K_{pqr} refer to Equations (2) and (1), respectively; ² values obtained by an average of potentiometric and UV–Vis spectrophotometric data; ³ ±Std. Dev.; ⁴ errors on weighed data; ⁵ value kept constant from potentiometric data. Standard uncertainties: u(*T*) = 0.1 K; u(*I*) = 0.01 mol L⁻¹.

Based on the common $\operatorname{Fe} L^{(3-z)}$ species, the following trend can be observed: $\operatorname{Fe}(L4)^+ > \operatorname{Fe}(L2)^+ > \operatorname{Fe}(L5)^{2+} > \operatorname{Fe}(L1)^+$ at T = 298.15 K. The stability of the $\operatorname{Fe}^{3+}/(3,4-\operatorname{HP})$ species is favoured by the concomitant presence of the extra-functional groups on the 3-hydroxy-4-pyridinone derivatives, in particular the α -amino-carboxylic groups in alkyl chain ((H₂(L2) and H₂(L4), Figure 1) [24]. Furthermore, from the comparison among the $\operatorname{Fe}(L1)^+$ and $\operatorname{Fe}(L5)^+$ formation constant values, it can be observed that the ligand featured by the only amino group (H(L5)) in the alkyl chain forms Fe^{3+} complexes with higher stability than the carboxylic-3-hydroxy-4-pyridinone (H₂(L1)), in accordance with literature data [24] reported on the interaction of alkylamines and carboxylic acids towards Fe^{3+} .

In the case of the H₂(*L*2) and H(*L*5) ligands, the formation constants were also determined at $I = 0.15 \text{ mol } \text{L}^{-1}$ in NaCl_(aq) and T = 310.15 K, as reported in Table 2: the obtained values increase with temperature.

A further deepening on the speciation of the different $\text{Fe}^{3+}/(3,4\text{-HP})$ systems may be performed considering the distribution diagrams drawn from potentiometric data at $I = 0.15 \text{ mol } \text{L}^{-1}$ in $\text{NaCl}_{(\text{aq})}$ and T = 298.15 K, as reported in Figure 4 for $\text{H}_2(L2)$ and in Figure S2 for $\text{H}_2(L1)$, $\text{H}_2(L4)$ and H(L5).



Figure 4. Distribution diagram of the Fe³⁺/H₂(*L*2) system at *T* = 298.15 K, *I* = 0.15 mol L⁻¹ in NaCl_(aq), $c_{\text{Fe}}^{3+} = 5.0 \cdot 10^{-4} \text{ mol L}^{-1}$ and $c_{\text{ligand}} = 1.1 \cdot 10^{-3} \text{ mol L}^{-1}$.

In the case of H₂(*L*1) (Figure S2a), the diagram shows the FeH(*L*1)²⁺ and Fe(*L*1)⁺ species reaching the maximum percentages of 93% and 19% at pH ~ 2.8 and 3.5, respectively. Introducing the solubility product of Fe(OH)⁰_{3(s)} [21] in the speciation model in the HySS programme [30], used to calculate the formation percentages and to represent the distribution diagrams, the formation of the sparingly soluble species should occur at pH ~ 3.5, hindering the possible formation of the Fe(*L*1)₂⁻ species which should only start to form at the mentioned pH value. However, since the precipitation was experimentally observed at pH ~ 5.0, the formation of the Fe(*L*1)₂⁻ species in the pH range 3.5–5.0 could be considered as a probable complex, and therefore it was reported in Table 2. Regarding the species distribution of Fe³⁺/H₂(*L*2) (Figure 4), Fe³⁺/H₂(*L*4) (Figure S2b) and Fe³⁺/H(*L*5) (Figure S2c) species, the FeHL^(4-z) species achieves 99% formation at pH ~ 3.8, 3.7, 3.2 for H₂(*L*2), H₂(*L*4) and H(*L*5), respectively. As regards the FeL₂^(3-2z) complex, it starts to form from pH ~ 3.2 with ligands as H₂(*L*2) and H₂(*L*4), from pH ~ 2.6 with H(*L*5).

Figures 5, 6, S3 and S4 show the UV–Vis behaviour of $Fe^{3+}/H_2(L2)$ and $Fe^{3+}/H(L5)$ systems at different component concentration and temperatures.



Figure 5. UV–Vis absorption profiles of the Fe³⁺/H₂(*L*2) system at *I* = 0.15 mol L⁻¹ in NaCl_(aq), *T* = 298.15 K and at different pH values. (a) $c_{\text{Fe3+}} = 2.5 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$, $c_{\text{ligand}} = 1.8 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$; (b) $c_{\text{Fe3+}} = 2.5 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$, $c_{\text{ligand}} = 5.8 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$.



Figure 6. UV–Vis absorption profiles of the Fe³⁺/H(*L*5) system at I = 0.15 mol L⁻¹ in NaCl_(aq), different temperatures and pH values. (a) T = 298.15 K, $c_{Fe3+} = 2.0 \cdot 10^{-4}$ mol L⁻¹, $c_{ligand} = 6.4 \cdot 10^{-4}$ mol L⁻¹; (b) T = 310.15 K, $c_{Fe3+} = 2.5 \cdot 10^{-4}$ mol L⁻¹, $c_{ligand} = 5.7 \cdot 10^{-4}$ mol L⁻¹.

For the Fe³⁺/H₂(*L*2) system (Figure 5 and Figure S3), a band with $\lambda_{max} = 568$ nm is observed at pH ~ 2.0, followed by an intensity decrease at pH ~ 3.7. A first band hypsochromic shift ($\lambda_{max} = 510$ nm) and an absorbance increase occurs at pH ~ 4.5. Then, a second blue shift and a band is observed with $\lambda_{max} = 460$ nm from pH ~ 6.1–7.1, depending on the experimental conditions, up to the formation of precipitate, which hindered further investigations.

In the case of the Fe³⁺/H(*L*5) system, at metal/ligand stoichiometric conditions (Figure S4), the mentioned band with $\lambda_{max} = 568$ nm at pH ~ 2.0, as well as its two hypsochromic shifted bands ($\lambda_{max} = 510$ nm, 460 nm) at pH ~ 4.9 and 5.9–6.0, respectively, are observed. For $c_{Fe3+}/c_{ligand} = 1/2$ and $c_{Fe3+}/c_{ligand} = 1/3$ (Figure 6), the first recorded band is featured by $\lambda_{max} = 536$ nm at pH ~ 2.0, with a blue shift occurring at pH ~ 4.9 with a band at $\lambda_{max} = 515$ nm, whilst the last blue shift corresponds to a band at $\lambda_{max} = 460$ nm, similarly to the previous described spectra.

The deconvolution of the UV–Vis spectrophotometric data allowed us to calculate the molar absorptivity ($\varepsilon/L \pmod{1} \operatorname{cm}^{-1} \operatorname{cm}^{-1}$)) values for each metal-ligand species. Graphical representations of the ε determined for the Fe³⁺/H₂(*L*2) and Fe³⁺/H(*L*5) systems are reported in Figure S5 and 7, respectively, at *I* = 0.15 mol L⁻¹ in NaCl_(aq) and different temperatures. As a representative example, the calculated molar absorptivities for the Fe³⁺/H(*L*5) species are: $\varepsilon_{\text{max}}(\text{FeH}(L5)_{3+}) = 1440$ at $\lambda_{\text{max}} = 571$ nm, ε_{max} (Fe(*L*5)²⁺) = 2160 at $\lambda_{\text{max}} = 520$ nm, $\varepsilon_{\text{max}}(\text{Fe}(L5)^{2+}) = 2736$ at $\lambda_{\text{max}} = 492$ nm at *T* = 298.15 K; $\varepsilon_{\text{max}}(\text{FeH}(L5)_{3+}) = 1293$ at $\lambda_{\text{max}} = 578$ nm, $\varepsilon_{\text{max}}(\text{Fe}(L5)^{-2+}) = 2124$ at $\lambda_{\text{max}} = 518$ nm, $\varepsilon_{\text{max}}(\text{Fe}(L5)^{2+}) = 3006$ at $\lambda_{\text{max}} = 460$ nm at *T* = 310.15 K.



Figure 7. Graphical representation of molar absorptivity of Fe³⁺/H(*L*5) species at I = 0.15 mol L⁻¹ in NaCl_(aq), T = 298.15 K (solid line) and 310.15 K (dot line).

2.5. Literature Data Comparison

From the best of our knowledge, no studies have been reported on the Cu²⁺/(3,4-HP) systems. Two papers have been published by Santos et al. [31,32] on the binding ability of the same H₂(*L*1) and H₂(*L*4) ligands (Figure 1) towards Fe³⁺ at *I* = 0.10 mol L⁻¹ in KNO_{3(aq)} and *T* = 298.15 K. The authors determined a speciation scheme featured by FeH_rL_q^(3+r-qz) (q, r = 1–3) species with different stoichiometry, including FeHL²⁺. This complex was also reported in the current work for the same two ligands (Table 2), and so a comparison between the experimental and literature data can be made. The stability constants determined by Santos et al. are log*K*₁₁₁ = 9.58 for H₂(*L*1) [32] and log*K*₁₁₁ = 15.16 for H₂(*L*4) [31] (Table S2). The value obtained for H₂(*L*4) is in good accordance with the log*K*₁₁₁ = 15.21 (Table 2) presented in this paper at *I* = 0.15 mol L⁻¹ in NaCl_(aq) and *T* = 298.15 K. However, the value previously reported for H₂(*L*1) is slightly higher than the value determined herein (log*K*₁₁₁ = 7.42, Table 2).

Some other comparisons could also be made considering metal-ligand investigations on compounds with similar structures and functional groups (Figure S6) with respect to the 3,4-HP ligands under study. Nevertheless, some little differences in the ligand struc-

tures, discrepancies between the experimental conditions and, in particular, the different approaches sometimes used by the authors for the data treatment (determination of ligands' acid–base properties, apparent neglect or very few information reported on the metals' hydrolytic behaviour), make it difficult to establish a direct comparison among the stability constants. However, an attempt of comparison could be performed, considering the log K_{110} values reported in this paper for the M $L^{(n-z)}$ species and the data published in the literature for complexes with the same stoichiometry.

Concerning the $\text{Cu}^{2+}/\text{ligand systems}$, the stability constants reported in Table 1, for $\text{Cu}(L1)^{0}_{(\text{aq})}$, $\text{Cu}(L2)^{0}_{(\text{aq})}$, $\text{Cu}(L3)^{0}_{(\text{aq})}$, $\text{Cu}(L4)^{0}_{(\text{aq})}$ and $\text{Cu}(L5)^{+}$ species at I = 0.15 mol L⁻¹ in $\text{NaCl}_{(\text{aq})}$ and T = 298.15 K, can be compared with those published for the $\text{Cu}L^{(2-2)}$ complexes determined with Deferiprone (*DFP*) [33,34] and *AcNPrHP* ([35] (Figure S6, Table S2) at I = 0.10 mol L⁻¹ in $\text{KCl}_{(\text{aq})}$ and the same temperature. As regards the $\text{Cu}(L1)^{0}_{(\text{aq})}$ species, its stability is three orders of magnitude lower than the mentioned literature data. In the case of the other four 3-hydroxy-4-pyridinones, the copper-experimental values were found to be about two ($\text{Cu}(L5)^{+}$) and five ($\text{Cu}(L2)^{0}_{(\text{aq})}$, $\text{Cu}(L3)^{0}_{(\text{aq})}$, $\text{Cu}(L4)^{0}_{(\text{aq})}$) logarithmic units higher (Table 1), respectively, with respect to the literature ones (Table S2) [33–35]. The formation constant published for the Cu^{2+}/L -Aspartic acid (*Asp*) 1:1 stoichiometry species (Table S2) [22] can be compared with the corresponding H₂(L2) and H₂(L3) derivatives. In this case, the experimental values (Table 1) are six orders of magnitude higher with respect to the literature data [22].

As regards the $Fe^{3+}/(3,4-HPs)$ systems, the stability of the $Fe(L1)^+$, $Fe(L2)^+$, $Fe(L4)^+$ and $Fe(L5)^{2+}$ species (Table 2), at $I = 0.15 \text{ mol } L^{-1}$ in $NaCl_{(aq)}$ and T = 298.15 K and 310.15 K, can be compared with the literature data (Table S2) reported for the $Fe(DFP)^{2+}$ complex at $I = 0.10 \text{ mol } L^{-1}$ in KCl_(aq), at the same temperatures [33,34,36]. The Fe(L1)⁺ experimental value is about one-two logarithmic units lower with respect to the mentioned literature data. The $Fe(DFP)^{2+}$ stability constants [33,34,36] are instead about six–seven orders of magnitude lower than the $Fe(L2)^+$, $Fe(L4)^+$ and $Fe(L5)^{2+}$ stability constants reported in Table 2. Similar observations can be made for the formation constants published for the $Fe(Asp)^{2+}$ species, at $I = 1.00 \text{ mol } L^{-1}$ in a Na⁺ background electrolyte, T = 293.15 K [22], as well as for Fe(Orn)⁺ (L-Ornithine) at $I = 0.10 \text{ mol } L^{-1}$ in NaClO_{4(aq)}, at the same temperature [37], which can be compared with their $H_2(L2)$ and $H_2(L4)$ derivatives, respectively. In particular, in the case of $Fe(L2)^+$ species (Table 2), the metal-ligand stability is about eleven logarithmic units higher with respect to the $Fe(Asp)^{2+}$ one [22]. The $Fe(L4)^+$ experimental value (Table 2) is almost fourteen orders of magnitude higher than the $Fe(Orn)^+$ literature constant [37]. The data reported in the literature (Table S2) for other 3,4 HP analogues $(H_2Si, i = 1-3)$, for the Fe $(S1)^{2+}$ complex at $I = 0.10 \text{ mol } L^{-1}$ in KNO_{3(aq)} [38] and for the $Fe(S2)^+$ and $Fe(S3)^+$ ones at the same ionic strength and temperature but in a MOPS (3-(Nmorpholino)propanesulphonic acid) buffer at pH = 7.4 [39], present a stability six-height orders of magnitude lower than those observed for all the $Cu^{2+}/(3,4-HPs)$ 1:1 stoichiometry species, with the exception of the $Fe(L1)^+$ experimental value, which was between five and seven logarithmic units higher with respect to the literature data [38,39].

Overall, the generally much higher values found for the stability of the 1:1 metal complex with the ligands bearing a terminal α -amino-carboxylic group (H₂(*L*2), H₂(*L*3), H₂(*L*4)) may be mainly attributed to the probable co-adjuvation of the main hydroxypyridinone (*O*,*O*) metal coordination by the (*N*,*O*) glycine type coordination, and also the inserted amide bond, which can further interfere in the length and rigidity of the linker between both main groups.

2.6. Sequestering Ability

The evaluation of the sequestering ability of the 3-hydroxy-4-pyridinones towards Cu^{2+} and Fe^{3+} can be performed by calculating the $pL_{0.5}$ empirical parameter which represents the total ligand concentration required for the 50% sequestration of a metal cation if present in trace amount in solution. The $pL_{0.5}$ can be described using a sigmoidal

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type Boltzmann equation, with asymptotes equal to 1 for $pL \rightarrow -\infty$ and 0 for $pL \rightarrow +\infty$ (Equation (3)):

$$x_M = \frac{1}{1 + 10^{(pL - pL_{0.5})}} \tag{3}$$

where x_M is the mole fraction of metal cation complexed by the ligand, $pL = -\log c_L$ and $pL_{0.5} = -\log c_L$, if $x_M = 0.5$. The evaluation of the sequestering ability is very important for detoxification, remediation of polluted systems and water treatment processes, requiring the use of a chelating agent with the aim of trying to optimize the working conditions. A more detailed description of the $pL_{0.5}$ determination, its importance and other possible applications is reported in the literature [40].

The study of the sequestering ability of the ligands towards Cu^{2+} and Fe^{3+} was performed at I = 0.15 mol L⁻¹ in NaCl_(aq), T = 298.15 K and different pHs. In the case of Fe³⁺/H₂(*L*2) and Fe³⁺/H(*L*5) systems, the pL_{0.5} was also determined at the same ionic strength and T = 310.15 K.

From the analysis of the data reported in Table 3 and Figure S7a for the $Cu^{2+}/L2$ system, it can be concluded that the sequestering ability increases with pH, probably due to the gradual ligand deprotonation, which favours the metal-ligand electrostatic interaction. At pH = 7.4 (physiological value), I = 0.15 mol L⁻¹ in NaCl_(aq) and T = 298.15 K, the pL_{0.5} trend is: $H_2(L2) \ge H_2(L3) > H_2(L4) > H(L5) > H_2(L1)$ (Table 3, Figure S7b). As already observed for the stability constants, the sequestering ability is also influenced by the presence in the ligands structure of the -CO₂H, -NH₂, -CHNH₂CO₂H [24] and, possibly, the amidic moiety in the alkyl chain. In addition, the pL_{0.5} value obtained for the amino-3-hydroxy-4-pyridinone (H(L5)) is slightly higher than the one calculated for H₂(L1) (terminal -CO₂H group), highlighting a better Cu²⁺ sequestration by the ligand featured by the terminal -NH₂ group (H(L5)) with respect to the carboxylic one [24–26].

Table 3. $pL_{0.5}$ ¹ values of Cu²⁺/ligands systems at different pHs, $I = 0.15 \text{ mol } L^{-1}$ in NaCl_(aq) and T = 298.15 K.

Ligand	рН	pL _{0.5}
H ₂ (L1)	7.4	7.09
$\overline{H_2}(L2)$	2.5	2.74
	3.0	3.70
	4.0	5.48
	5.0	6.99
	6.0	8.18
	7.4	10.29
	8.1	10.54
	9.0	11.98
$H_2(L3)$	2.5	3.10
	3.0	4.05
	4.0	5.70
	5.0	7.18
	6.0	8.41
	7.4	10.30
	8.1	11.43
	9.0	12.20
	10.0	12.14
H ₂ (L4)	7.4	9.90
H(L5)	7.4	7.25

¹ values calculated by Equation (3).

As regards the iron-containing systems, the formation of precipitate at pH ~ 5.0 during the potentiometric measurements allowed us to evaluate the sequestering ability of the ligands in a quite narrow pH range. As can be observed in Figure 8, the pL_{0.5} trend at pH = 4.0 is: H₂(L2) (8.12) > H₂(L4) (7.94) > H(L5) (6.77) > H₂(L1) (5.14), confirming that, analogously to what was observed for the stability constants, the sequestration is mainly favoured by the presence in the ligand structures of the amide-amino-carboxylic, aminocarboxylic or amino moieties [24] in the alkyl chain bound to the *N*-heterocyclic ring.



Figure 8. Sequestration diagram of the ligands towards Fe^{3+} at pH = 4.0, *I* = 0.15 mol L⁻¹ in NaCl_(aq) and *T* = 298.15 K.

Furthermore, since in the case of $H_2(L2)$ and H(L5) UV–Vis experiments were carried out in the pH range 2.0–9.1, for these ligands the p $L_{0.5}$ values were also calculated at different pHs, T = 298.15 K and 310.15 K (Table S3), considering the spectrophotometric data (Table 2).

The sequestering ability of $H_2(L2)$ and H(L5) towards Fe³⁺ was found to increase with pH, possibly owing to the gradual ligand deprotonation with pH increasing. The p $L_{0.5}$ values also increase with temperature, in accordance with the stability constants trend (Table 2).

2.7. Analysis of the pM Values

The study of the metal-chelating affinity of a ligand or the comparison between different ligands' behaviour towards one or more metal cations can be performed by means of the pM parameter, with pM = $-\log [M]_{\text{free}}$ (with M = Cu or Fe) for $c_{\text{Mn+}} = 1.0 \cdot 10^{-6} \text{ mol } \text{L}^{-1}$ and $c_{\text{ligand}} = 1.0 \cdot 10^{-5} \text{ mol } \text{L}^{-1}$ [41].

The pM values of all the Cu²⁺ and Fe³⁺/(3,4-HPs) systems investigated in this paper were calculated at pH = 7.4 (physiological value). Furthermore, an attempt to compare the obtained data with those determined for ligands with similar molecular structures and functional groups (Figure S5) was carried out using literature stability constants [22,31–39,42], taking into account the already mentioned experimental and methodological differences used for the data treatment.

The analysis of the pCu values reported in Table 4 and in Figure 9a showed that, at physiological pH, the copper-chelating affinity by the ligands is favoured by the concomitant presence of -COOH, -NH₂ and amino-carboxylic groups in the 3,4-HP molecules. They follow the trend: $H_2(L3) > H_2(L2) > H_2(L4) > H_2(L1) > H(L5)$. At the selected pH value, an inversion of pCu tendency can be observed for $H_2(L1)$ (terminal -COOH) and H(L5) (terminal -NH₂) with respect to the already mentioned stability constants and sequestration trend. This aspect could be explained considering that at pH = 7.4, the amino group present in H(L5) is still protonated while the carboxylic one in $H_2(L1)$ is already deprotonated, thus favouring the $Cu^{2+}/H_2(L1)$ electrostatic interaction. At higher pH values, with the deprotonation of -NH₃⁺ to NH₂ in H(L5), the metal affinity increases with respect to $H_2(L1)$, and the pCu trend becomes analogous to that observed for the stability constants and $pL_{0.5}$ values. A comparison between the pCu data (Table 4, Figure 9b) determined for the

 $Cu^{2+}/H_2(L2)$ system and those calculated for other ligands (Figure S5), such as *DFP* [33,34], *L*-Aspartic acid (*Asp*) [22] and *AcNPrHP* [35], showed that the H₂(*L*2) copper-chelating affinity at physiological pH and micromolar concentration conditions is higher with respect to the other compounds, following the trend: H₂(*L*2) > *AcNPrHP* > *DFP* > *Asp*, with $\Delta pCu = pCu_{H2(L2)}-pCu_{literature} = 0.60, 0.74, 3.16$, respectively. This trend highlights that the ligands featured by the pyridinone ring display a higher copper affinity with respect to the others, although taking into account some little differences, probably due to the experimental conditions reported in Table 4.

Table 4. pM values calculated for different Cu^{2+} and $Fe^{3+}/(3,4-HPs)$ systems based on 3-hydroxy-4-pyridinone ligands and similar structures' ligands at pH = 7.4 from stability constants reported in the literature.

M ⁿ⁺	Ligand	pМ	Ref.	M ⁿ⁺	Ligand	pМ	Ref.
Cu ²⁺	H ₂ (L1)	8.60	This work	Fe ³⁺	$H_2(L4)$	21.90 ³	[31]
Cu ²⁺	$H_2(L2)$	11.00	This work	Fe ³⁺	Deferiprone	20.70 ¹	[34]
Cu ²⁺	$H_2(L3)$	11.10	This work	Fe ³⁺	$H_2(\hat{S}1)$	20.59 ¹	[38]
Cu ²⁺	$H_2(L4)$	10.90	This work	Fe ³⁺	$H_2(S2)$	20.19 ⁶	[39]
Cu ²⁺	$H_2(L5)$	8.20	This work	Fe ³⁺	$H_2(S3)$	19.97 ⁶	[39]
Cu ²⁺	Deferiprone	10.69 ¹	[34]	Fe ³⁺	L-Aspartic acid	15.63 ⁴	[22]
Cu ²⁺	L-Aspartic acid	7.84 ²	[22]	Fe ³⁺	<i>L-</i> Ornithine	15.64 ⁵	[37]
Cu ²⁺	AcNPrHP	$10.40^{\ 1}$	[35]	A1 ³⁺	$H_2(L2)$	14.20 ²	[13]
Fe ³⁺	$H_2(L2)$	24.38	This work	A1 ³⁺	$H_2(L5)$	13.17 ²	[13]
Fe ³⁺	$H_2(L5)$	23.98	This work	Zn ²⁺	$H_2(L2)$	6.03 ²	[19]
Fe ³⁺	$H_2(L1)$	19.90 ³	[32]	Zn ²⁺	$H_2(L5)$	8.28 ²	[19]

 $\overline{I} = 0.10 \text{ mol } L^{-1} \text{ in } \text{KCl}_{(aq)}, T = 298.15 \text{ K}; {}^{2}I = 0.15 \text{ mol } L^{-1} \text{ in } \text{NaCl}_{(aq)}, T = 298.15 \text{ K}; {}^{3}I = 0.10 \text{ mol } L^{-1} \text{ in } \text{KNO}_{3(aq)}, T = 298.15 \text{ K}; {}^{4}I = 1.00 \text{ mol } L^{-1} \text{ in } \text{Na}^{+} \text{ ionic medium}, T = 293.15 \text{ K}; {}^{5}I = 0.10 \text{ mol } L^{-1} \text{ in } \text{KNO}_{3(aq)}, T = 293.15 \text{ K}; {}^{6}I = 0.10 \text{ mol } L^{-1} \text{ in } \text{MOPS}$ (3-(*N*-morpholino)propanesulphonic acid) buffer at pH = 7.4, T = 298.15 \text{ K}.



Figure 9. Graphical representation of pCu values calculated vs. pH: (a) the (3,4-HPs) under study and (b) H₂(*L*2) and ligands with similar structures. Experimental conditions: $c_{Cu}^{2+} = 1.0 \cdot 10^{-6}$ mol L⁻¹ and $c_{ligand} = 1.0 \cdot 10^{-5}$ mol L⁻¹.

The Fe³⁺ chelating efficiency was evaluated at pH = 7.4 only for *L*2 and *L*5 ligands, since their interaction with the metal cation was also investigated by UV–Vis spectrophotometry, an analytical technique not used for the Fe³⁺/H₂(*L*1) and Fe³⁺/H₂(*L*4) systems. In fact, the UV–Vis studies were performed at lower component concentrations ($c \sim 10^{-4}$ mol L⁻¹) than those used for the potentiometric ones ($c \sim 10^{-3}$ mol L⁻¹), allowing us to investigate a wider pH range (2.0–9.1) without being stopped at pH ~ 5.0, as occurred for potentiometric titrations, owing to the formation of a precipitate possibly attributable to the formation of the Fe(OH)_{3(s)} species [21].

Analysis of the pFe values, reported in Table 4 for the systems studied herein, showed that at physiological pH the iron-chelating affinity is favoured by the concomitant pres-

ence of extra-functional groups in the 3,4-HP ligand molecules, namely the amide-aminocarboxylic moiety (H₂(*L*2)), with respect to the simple terminal group -NH₂ (H(*L*5)). The pFe data (Table 4) determined for Fe³⁺/H₂(*L*2) and Fe³⁺/H(*L*5) systems were also compared with the values reported in the literature for ligands such as H₂(*L*1) [32], H₂(*L*4) [31], *DFP* [33,34,36], *Asp* [22], *Orn* [37], H₂(*S*1) [38], H₂(*S*2) [39] and H₂(*S*3) [39]. H₂(*L*2) and H(*L*5) iron-chelating affinities at pH = 7.4 and micromolar conditions were found to be higher with respect to the literature compounds and follow the trend: H₂(*L*2) > H(*L*5) > H₂(*L*4)_{Santos} > *DFP* > H₂(*S*1) > H₂(*S*2) > H₂(*S*3) > H₂(*L*1)_{Santos} > *Orn* ~ *Asp*, with Δ pFe = pFe_{H2(*L*2)}pFe_{H(*L*5) or literature = 0.40, 2.48, 3.71, 3.79, 4.28, 4.41, 4.48, 8.74, 8.75, respectively. The mentioned trend highlights that the ligands featured by the hydroxo-oxo functionality from the pyridinone ring and amino, amino-carboxylic or amide-amino-carboxylic moieties present a higher metal affinity with respect to the others, even, obviously, taking also into account some little differences, probably due to the experimental conditions reported in Table 4.}

2.8. Comparison between $M^{n+}/(3,4-HPs)$ Systems

The data presented in the current paper for Cu^{2+} and $Fe^{3+}/(3-hydroxy-4-pyridinones)$ systems (Tables 1 and 2) were compared with those already reported in the literature on the interaction of the five ligands with Zn^{2+} [19] and Al^{3+} [13] (Table S4) at I = 0.15 mol L⁻¹ in $NaCl_{(aq)}$ and T = 298.15 K. The speciation models determined for the different systems display a common species, namely the ML^(n-z), which can be used as reference to evaluate and compare the binding ability of the ligands towards the metal cations. The data analysis showed that the log K_{110} trend is: Fe³⁺ > Al³⁺ > Cu²⁺ > Zn²⁺, meaning that the 3,4-HPs are featured by a much higher tendency to form very stable complex species with Fe³⁺, followed by Al³⁺, with respect to the M²⁺. In addition, considering the stability constants reported in the literature for the Zn^{2+} [19] and $Al^{3+}/(H_2(L2), H(L5))$ systems [13], the pZn and pAl values were also calculated at physiological pH (Table 4) and compared with the analogous results presented in this paper for Cu²⁺ and Fe³⁺. Analysing the data obtained for the different metal cations and the graphs in Figure 10 and Figure S8, we can conclude that, similarly to the $\log K_{110}$ behaviour, the pM values follow the trend for the metal ions: $Fe^{3+} > Al^{3+} > Cu^{2+} > Zn^{2+}$. Thus, both of the 3-hydroxy-4-pyridinones present a higher chelating affinity towards Fe³⁺, and in a lesser extent also to Al³⁺, with respect to divalent metal cations.



Figure 10. Calculated pM values vs. pH for the different $M^{n+}/H_2(L2)$ systems at T = 298.15 K, I = 0.15 mol L⁻¹ in NaCl_(aq), $c_M^{n+} = 1.0 \cdot 10^{-6}$ mol L⁻¹ and $c_{\text{ligand}} = 1.0 \cdot 10^{-5}$ mol L⁻¹.

These trends could probably be justified taking into account the already mentioned *"hard-soft* acids and bases" theory (HSAB) [27–29], according to which a *hard* acid–*hard*

base or a *soft* acid–*soft*-base interactions are kinetically and thermodynamically favoured if compared with *hard–soft* ones. On this basis, the affinity between *hard* metal cations (acids: Fe^{3+} , Al^{3+}) and *hard-base* functional groups (bases: -OH, -COOH) is higher with respect to those with *borderline* acids like Cu²⁺ and Zn²⁺.

In light of these considerations, it can be claimed that, from a thermodynamic point of view, most of the bifunctional 3,4-HP ligands studied herein are particularly selective towards Fe^{3+} and could be considered promising iron-chelating agents, also avoiding the possibility of a significant competition, and eventually a depletion, of divalent metals with biological and environmental relevance, such as Cu^{2+} and Zn^{2+} .

3. Materials and Methods

3.1. Chemicals

Riedel–deHäen concentrated ampoules were used to prepare sodium hydroxide and hydrochloride solutions standardized against potassium hydrogen phthalate and sodium carbonate, respectively. NaOH solutions were preserved from atmospheric carbon dioxide by means of soda lime traps. CuCl₂·2H₂O and FeCl₃·6H₂O salts purchased by Fluka were weighed to prepare the metal solutions without further purification and standardized against EDTA standard solutions [43]; their purity was always \geq 98%. The synthesis of the functionalized 3-hydroxy-4-pyridinones was already reported in the literature [13]. The ligand solutions were prepared by weighing the products in the neutral form (H_r(*L*)⁰) without any further purification. Their purity was checked by means of alkalimetric measurements and, for all the ligands, it was found to be \geq 99.5%. The ionic medium aqueous solutions were prepared by weighing the pure Fluka NaCl salt, previously dried in an oven at *T* = 383.15 K for two hours. The reagents used to carry out the studies were of the best available purity. The preparation of the solutions was performed using analytical grade water (R = 18 M Ω cm⁻¹) and grade A glassware.

3.2. Analytical Instrumentation and Procedures

3.2.1. Potentiometric Equipment and Procedure

The interactions of the five 3-hydroxy-4-pyridinones towards Cu²⁺ and Fe³⁺ were experimentally investigated using a Metrohm 809 Titrando and a potentiometer with a combined Thermo-Orion glass electrode (Ross type 8102) connected to an automatic burette. This apparatus was coupled to a personal computer, and automatic titrations were performed by means of the MetrohmTiAMO 1.2 software, useful for the control of titrant delivery, data acquisition and e.m.f. stability. The estimated accuracy, for e.m.f. and titrant volume readings, was ± 0.15 mV and ± 0.003 mL, respectively. The measurements were carried out in 25 mL thermostatted cells under magnetic stirring, and purified presaturated nitrogen was bubbled into the solutions for at least 5 min to exclude the presence of oxygen and carbon dioxide inside. For all the experiments, titrations of hydrochloric acid with standard NaOH solutions were carried out at the same temperature, ionic strength and ionic medium conditions with respect to those used for the systems under study, for refining the value of the electrode potential (E^0), the acidic junction potential ($E_j = j_a[H^+]$) and the ionic product of water (K_w). The pH scale employed was the free scale and pH \equiv $-\log[H^+]$, with $[H^+]$ that is the free concentration of the proton. From sixty to one hundred data points were collected during each titration, depending on the possible formation of sparingly soluble species.

The potentiometric titrations were carried out at $I = 0.15 \text{ mol } L^{-1}$ in NaCl_(aq), T = 298.15 Kand different concentrations of ligands ($c_{\text{ligand}} = 5.0 \cdot 10^{-4} - 1.5 \cdot 10^{-3} \text{ mol } L^{-1}$) and metal cations ($c_{\text{Mn+}} = 4.3 \cdot 10^{-4} - 1.0 \cdot 10^{-3} \text{ mol } L^{-1}$). The pH ranges investigated were 2.0–10.0 and 2.0–5.0 for Cu²⁺ and Fe³⁺/(3,4-HPs) measurements, respectively, due to the formation of sparingly soluble species.

3.2.2. UV-Vis Spectrophotometric Apparatus and Procedure

The UV–Vis spectrophotometric titrations were carried out using a Varian Cary 50 spectrophotometer presenting an optic fibre probe with a fixed 1-cm path length. This instrument was connected to a computer, and the recording of absorbance (A) signal vs. wavelength (λ / nm) was carried out by means of the Varian Cary WinUV software. At the same time, a Thermo-Orion combined glass electrode (Ross type 8102), linked to a potentiometer, was employed to collect potentiometric data. The NaOH titrant solution was delivered in a 25-mL titration cell using an automatic burette (Metrohm 665 model). The homogeneity of the solutions during the measurements was ensured using a magnetic stirrer. Nitrogen was bubbled in the solutions for at least 5 min before starting the experiments, also in this case, for excluding the presence of O_{2(g)} and CO_{2(g)} inside.

The binding ability of H₂(*L*2), H₂(*L*4) and H(*L*5) towards Cu²⁺ and Fe³⁺ was also studied by means of UV–Vis spectrophotometric titrations of solutions containing different concentrations of ligands ($c_{\text{ligand}} = 1.0 \cdot 10^{-5} - 2.1 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$) and metal cations ($c_{\text{Mn+}} = 5.0 \cdot 10^{-6} - 1.5 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$). The experiments were performed at *I* = 0.15 mol L⁻¹ in NaCl_(aq), *T* = 298.15 K, a wavelength of $200 \le \lambda$ / nm ≤ 800 , pH ranges 2.0–11.0 and 2.0–9.1 for Cu²⁺ and Fe³⁺/(3,4-HPs) investigations, respectively. In the case of the Fe³⁺/H₂(*L*2) and Fe³⁺/H(*L*5) systems, the measurements were also carried out at *I* = 0.15 mol L⁻¹ in NaCl_(aq) and *T* = 310.15 K.

3.3. Computer Programmes

Appropriate computer programmes were employed for the treatment of experimental data from different analytical techniques. The non-linear least squares ESAB2M computer program [44] was used for the determination of the acid–base titrations parameters (E^0 , pK_w , j_a) and the reagents' analytical concentration. The elaboration of potentiometric data was carried out by means of the BSTAC computer program [45], while the UV–Vis spectrophotometric ones were processed using the HypSpec 2008 [46]. The calculation of the $M^{n+}/3$ -hydroxy-4-pyridinone species formation percentages and the representation of distribution diagrams was performed using the HySS program [30].

4. Conclusions

The binding ability of five bifunctional 3-hydroxy-4-pyridinones towards Cu²⁺ and Fe³⁺ was studied by means of potentiometric and UV-Vis spectrophotometric measurements carried out at I = 0.15 mol L⁻¹ in NaCl_(aq) and T = 298.15 K. The data treatments allowed us to determine the speciation schemes featured by metal-ligand species with different stoichiometry and stability, due to the various functional groups present in the 3-hydroxy-4-pyridinones structures, which could potentially participate in the metal complexation and in the Cu²⁺ and Fe³⁺ behaviour in an aqueous solution. The stability of metal-ligand species follows the trends: $Cu(L3)^{0}_{(aq)} > Cu(L4)^{0}_{(aq)} > Cu(L2)^{0}_{(aq)} > Cu(L5)^{+} > Cu(L1)^{0}_{(aq)}$ and: $Fe(L4)^{+} > Fe(L2)^{+}_{(aq)} > Cu(L3)^{0}_{(aq)} > Cu(L4)^{0}_{(aq)} > Cu(L4$ > Fe(L5)²⁺ > Fe(L1)⁺, respectively. They were favoured by the simultaneous presence of amino or amino-carboxylic bearing groups in the 3,4-HP ligands, and showed some dependence on the length and structure of the chains between the pyridinone ring and the extra-functional groups. The investigation of the sequestering ability and metal-chelating efficiency was carried out by the calculation of the $pL_{0.5}$ and pM parameters at different pHs and physiological value (pH = 7.4), respectively. Similarly to the complexation behaviour, the sequestration and Cu^{2+} and Fe^{3+} affinity by the ligands under study is affected by the presence in the whole 3-hydroxy-4-pyridinone molecules of terminal amino-carboxylic groups and amidic moiety in the alkyl chain or, at least, of the one single terminal group, as -NH₂ group (H(L5)), with respect to the carboxylic group (H₂(L1)). In addition, the data presented in this paper for Cu²⁺ and Fe³⁺/3-hydroxy-4-pyridinone systems were compared with those reported in the literature, for the interaction of the ligands with Al³⁺ and Zn^{2+} at I = 0.15 mol L⁻¹ in NaCl_(aq) and T = 298.15 K. The log K_{110} and pM trend show a clear dependence on the metal ion ($Fe^{3+} > Al^{3+} > Cu^{2+} > Zn^{2+}$), meaning that the

3-hydroxy-4-pyridinones display a higher stability and chelating affinity towards Fe^{3+} and (in a lesser degree) also Al^{3+} , with respect to divalent metal cations. In light of these considerations, it can be claimed that, from a thermodynamic point of view, the ligands are particularly selective towards Fe^{3+} and could be considered promising iron-chelating agents, also avoiding the possibility of a significant competition, and eventually a depletion, of divalent metals with biological and environmental relevance, such as Zn^{2+} and Cu^{2+} .

Supplementary Materials: The following are available online, Table S1. Overall and stepwise protonation constants of the 3-hydroxy-4-pyridinones under study reported in the literature at $I = 0.15 \text{ mol } L^{-1}$ in NaCl_(aq) and different temperatures; Table S2. Literature stability constants of Cu²⁺ and Fe³⁺/ligand species reported at different temperatures, ionic strengths and ionic media in molar concentration scale; Table S3. $pL_{0.5}$ values of Fe³⁺/H₂(L2) and H(L5) systems at different pHs and temperature, from UV–Vis data at I = 0.15 mol L⁻¹ in NaCl_(ac); Table S4. Literature stability constants of $ZnL^{(2-z)}$ and $AlL^{(3-z)}$ species at I = 0.15 mol L^{-1} in $NaCl_{(aq)}$, T = 298.15 K; Figure S1. Distribution diagram of $Cu^{2+}/(3,4-HPs)$ systems at T = 298.15 K, I = 0.15 mol L^{-1} in NaCl_(aq), $c_{\text{Cu}}^{2+} = 5.0 \cdot 10^{-4} \text{ mol } \text{L}^{-1} \text{ and } c_{\text{ligand}} = 1.5 \cdot 10^{-3} \text{ mol } \text{L}^{-1}. \text{ Ligands} = \text{H}_2(L1) \text{ (a), } \text{H}_2(L3) \text{ (b); } \text{H}_2(L4) \text{ (c),}$ H(L5) (d); Figure S2. Distribution diagram of Fe³⁺ / (3,4-HPs) systems at T = 298.15 K, I = 0.15 mol L⁻¹ in NaCl_(aq), $c_{\text{Fe}}^{3+} = 5.0 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$ and $c_{\text{ligand}} = 1.1 \cdot 10^{-3} \text{ mol } \text{L}^{-1}$. Ligands = H₂(L1) (a), H₂(L4) (b), H(L5) (c); Figure S3. UV–Vis absorption profile $Fe^{3+}/H_2(L2)$ system at $I = 0.15 \text{ mol } L^{-1}$ in NaCl_(aq), T = 310.15 K and at different pH values. (a) $c_{\text{Fe3+}} = 2.3 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$, $c_{\text{ligand}} = 2.2 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$; (b) $c_{\text{Fe3+}} = 2.0 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$, $c_{\text{ligand}} = 4.0 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$; Figure S4. UV–Vis absorption profile Fe³⁺/H(L5) system at I = 0.15 mol L⁻¹ in NaCl_(aq), different temperatures and pH values. (a) T = 298.15 K, $c_{\text{Fe3+}} = 2.4 \cdot 10^{-4} \text{ mol } \text{L}^{-1}, c_{\text{ligand}} = 2.6 \cdot 10^{-4} \text{ mol } \text{L}^{-1};$ (b) $T = 310.15 \text{ K}, c_{\text{Fe3+}} = 2.5 \cdot 10^{-4} \text{ mol } \text{L}^{-1},$ $c_{\text{ligand}} = 2.4 \cdot 10^{-4} \text{ mol } \text{L}^{-1}$; Figure S5. Graphical representation of molar absorptivity of Fe³⁺/H₂(L2) species at T = 298.15 K (a) and 310.15 K (b), I = 0.15 mol L⁻¹ in NaCl_(aq); Figure S6. Molecular structures of compounds with similar structures and functional groups with respect to 3-hydroxy-4-pyridinones; Figure S7. Sequestration diagrams of: (a) $Cu^{2+}/H_2(L2)$ species at I = 0.15 mol L⁻¹ in $NaCl_{(aq)}$ and T = 298.15 K and different pHs, $pL_{0.5}$ values: 2.74 (pH = 2.5), 3.70 (pH = 3.0), 5.48 (pH = 4.0), 6.99 (pH = 5.0), 8.18 (pH = 6.0), 10.29 (pH = 7.4), 10.54 (pH = 8.1), 11.98 (pH = 9.0); (b) $Cu^{2+}/(3,4-HPs)$ systems at the same ionic strengths and temperature, pH = 7.4. pL_{0.5} values: 7.09 (H₂(L1)), 10.29 (H₂(L2)), 10.30 (H₂(L3)), 9.90 (H₂(L4)), 7.25 (H(L5)); Figure S8. Calculated pM values vs. pH for the different $M^{n+}/L5$ systems at T = 298.15 K, I = 0.15 mol L^{-1} in NaCl_(aq), $c_{\rm M}^{n+} = 1.0 \cdot 10^{-6} \text{ mol } L^{-1} \text{ and } c_{\rm ligand} = 1.0 \cdot 10^{-5} \text{ mol } L^{-1}.$

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