

Gut microbiota: an overlooked factor that plays a significant role in osteoporosis Journal of International Medical Research 2019, Vol. 47(9) 4095–4103 © The Author(s) 2019 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/0300060519860027 journals.sagepub.com/home/imr



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#### Abstract

Gut microbes are known as the body's second gene pool. Symbiotic intestinal bacteria play a major role in maintaining balance in humans. Bad eating habits, antibiotic abuse, diseases, and a poor living environment have a negative effect on intestinal flora. Abnormal intestinal microbes are prone to cause a variety of diseases, affecting life expectancy and long-term quality of life, especially in older people. Several recent studies have found a close association between intestinal microorganisms and osteoporosis. The potential mechanism of intestinal flora affecting bone formation or destruction by mediating nitric oxide, the immune and endocrine systems, and other factors is briefly described in this review. All of these factors may be responsible for the intestinal flora that causes osteoporosis. Studying the relationship between intestinal flora and bone health not only provides new ideas for studying the role of intestinal microorganism in osteoporosis, but also provides a new therapeutic direction for clinically refractory osteoporosis. Study of the relationship between intestinal microbiota and osteoporosis is important for maintaining bone health and minimizing osteoporosis.

#### **Keywords**

Intestinal flora, osteoporosis, nitric oxide, immunity, endocrine system, homocysteine

Date received: 21 March 2019; accepted: 6 June 2019

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# Introduction

There is an average of 100 trillion microorganisms in the human intestinal tract, including bacteria, archaea, fungi, and viruses. Among the many categories of microbes that have been identified, thickwalled bacteria, Bacteroides, microalgae, actinomycetes, and Proteus account for the majority of microbes. These microorganisms have far more genetic phenotypes than humans themselves, but their role in maintaining the health of the body has been largely overlooked.<sup>1</sup> Intestinal symbiotic bacteria play a major role in maintaining balance in humans by aiding food digestion and absorption, secreting microbial metabolites, and protecting mucosal barrier function. However, an imbalance in microorganisms releases proteins. peptides, and metabolites that act against organs, causing systemic multiple organ dysfunction.<sup>2</sup>

As the load-bearing organ of the body, bone has active metabolism and is of great significance in routine activities. Repeated falls, muscular atrophy, muscle-strength deficiency, limited activity, and weight loss can easily cause loss of bone mass, microstructure destruction, and other pathologiosteoporosis.<sup>3</sup> changes. such as cal A dynamic imbalance between osteoblast and osteoclast populations is the decisive factor for developing osteoporosis. Osteoporosis affects more than 200 million people worldwide, with more than one third of them suffering from osteoporotic fractures.<sup>4</sup> Therefore, osteoporosis places enormous economic and social pressure on society and individuals.<sup>5</sup> As the population of older people in society increases, so will the incidence of osteoporosis and the rates of disability and death due to osteoporosis. Early intervention during development of osteoporosis can reduce the burden on families and society and improve the long-term survival rate and quality of life of patients.

The symbiosis between gut microbiota and the host requires a delicate balance, which once disrupted, may increase the risk of osteoporosis. David et al. found that intestinal microorganisms affect bone through inflammation, nutrition, and calcium. The significance of inflammation was highlighted.<sup>6</sup> A review also proposed that the effect of intestinal flora on bone is mediated by effects on inflammation and immunity.<sup>7</sup> Under stress, the intestinal tract can cause changes in vasoactive substances,8 which has an adverse effect on bone formation. The autoimmunity of germ-bearing mice affected bone formation compared with sterile mice.<sup>9</sup> The intestinal tract is also involved in endocrine metabolism. which is associated with bone homeostasis. Studying the relationship between intestinal flora and bone health provides a new theoretical basis for delaying osteoporosis and provides a new therapeutic direction for clinically refractory osteoporosis. This article reviews the potential mechanism by which gut microbes mediate osteoporosis.

# Nitric oxide

Intestinal smooth muscle and intestinal capillary endothelial cells express inducible nitric oxide synthase (iNOS).<sup>10</sup> Nitric oxide synthase (NOS) is a rate-limiting enzyme for biosynthesis of nitric oxide (NO). Microbes can drive the binding of pathogenic bacteria bacterial or lipopolysaccharide-inducible transcription factor nuclear factor (NF- $\kappa$ B) to the *iNOS* promoter, thus upregulating iNOS transcription.<sup>11,12</sup> The number of osteoblasts are increased by iNOS under mechanical stress, but iNOS also promotes osteoclast increasing levels production by of RANKL (receptor activator of NF- $\kappa$ B).<sup>13</sup>

Intestinal microbes also promote the release of endothelial NOS (eNOS).<sup>14</sup> eNOS mRNA regulates production of osteoblasts and osteoclasts, as well as the

release of inflammatory mediators, such as tumor necrosis factor- $\alpha$ , interleukin (IL)-1, and IL-6. In NOS-deficient rats, trabecular bone mineralization and chondrocyte function are reduced, impeding lateral and lateral bone growth in 8- to 10-week-old mice. Low NO concentrations might affect bone growth. In a previous study, iNOS-deficient rats developed more severe osteoporosis than did eNOS-deficient rats, which indicated that intestinal microbes mainly maintained bone homeostasis by affecting iNOS activity.<sup>15</sup> However, after 12 to 18 weeks of age, the negative effect of NOS deficiency on bone homeostasis in mice was eliminated. This might be related to the negative effect of NOS deficiency on bone growth being counteracted by sex hormone-associated bone protection in mice.

High NO concentrations can impede osteoclast differentiation and activate the cGMP-dependent protein kinase G pathway, thus reducing acid secretion and adhesion.<sup>13</sup> However, at high concentrations, NO can compete for binding to RANKL, thus hindering the interaction of RANKL with NF- $\kappa$ B receptor agonists and impeding osteoclast activity.<sup>16</sup> These results indicate that the concentration of NO has opposite effects on bone. Moreover, studies have shown an association between NO and vitamin D. Vitamin D, as a direct transcriptional regulator of endothelial NOS, can positively regulate NO.17 However, epidemiology suggests otherwise. NO can also interact with vitamin D, affecting the number of osteoblasts, as well as their response under fluid-flow shear stress.<sup>18</sup> NO indirectly affects bone homeostasis by affecting vitamins. Changes in plasma NO concentrations can also affect changes in bone blood flow, which may affect bone homeostasis. NO plays a role in the endpoint of osteoporosis and this can be seen in various types of osteoporosis.

### Immune system

The immune system is a host defense system. Under normal conditions, intestinal microbes and the immune system maintain the health of the body. However, imbalance of gut flora due to various factors can also promote development of disease through the immune system. In sterile mice, transplantation of complex microorganisms can alleviate depletion of myeloid progenitor cells and obstruction of monocyte proliferation.<sup>19</sup> This finding suggests that intestinal flora are related to development of immune cells. The immune system is inextricably linked to development of osteoporosis. In mice, thick-walled bacteria, particularly Clostridium, promote accumulation of regulatory T cells (Tregs) in the lamina propria of the colon. Tregs inhibit osteoclast differentiation and hinder osteoclast formation. Decreased abundance of *Clostridium* strains cause a decrease in factor- $\beta$  and Foxp3<sup>+</sup> Treg levels and an increase in bone loss.<sup>20</sup> However, Treg cells secrete cytotoxic T-lymphocyte antigen 4, which binds to CD80/CD86 molecules on osteoclast precursors, and thus inhibits inflammatory responses.<sup>21</sup> Britton et al.<sup>22</sup> found that Lactobacillus reuteri can decrease the number of T lymphocytes and inhibit osteoclast formation. However, CD4<sup>+</sup>T cells interact with CD11c dendritic cells and osteoclasts.<sup>23</sup> develop into functional Bacterial colonization experiments in animals have also confirmed that CD4<sup>+</sup>T cells interact with CD11c dendritic cells to probably aggravate osteoporosis.9

An imbalance in intestinal microbes inhibits differentiation of type 1 and 2 T helper cells (Th1 and 2), as well as Tregs, thus inducing osteoclast differentiation and proliferation, and aggravating bone loss. An imbalance in intestinal flora also induces differentiation of Th17 cells, which belong to the CD4<sup>+</sup>T-cell osteoclast population. Th17 cells secrete IL-17a, tumor necrosis factors, IL-1, and IL-6, as well as low levels of interferon- $\gamma$ , which stimulate osteoblasts to release RANKL.<sup>24</sup> Kim et al.<sup>25</sup> found that mouse commensal segmented filamentous bacteria and human commensal bacteria can promote Th17 differentiation, providing new guidelines for intestinal microbiota-targeted therapy for osteoporosis.

B lymphocytes control the RANKL/ osteoprotegrin (OPG) (osteotrophin) ratio through the phosphoinositide 3-kinase/protein kinase B (Akt)/mammalian target of rapamycin (mTOR) signal transduction pathway, and consequently regulate the growth rate of bone cells. Intestinal flora affect the expression of mTOR transcription factors in the posterior midgut. Knocking out TSC1 expression (a negative regulator of mTOR) causes an increase in the RANKL/OPG ratio, thus accelerating osteoclast proliferation.<sup>26</sup> Gut microbiota can also affect B-cell development and OPG production by B cells.<sup>27</sup> The decoy receptor OPG can directly inhibit RANKL.<sup>28</sup> Furthermore, OPG can inhibit osteoclast differentiation and bone resorption by modulating autophagy-related genes and AMP-activated protein kinase/ mTOR/p70S6K signaling.<sup>29</sup> The immune system may be involved in the pathogenesis of osteoporosis mediated by intestinal microorganisms. Immune system abnormalities often result in secondary osteoporosis.

# Endocrine system

Endocrine hormones act on various organs of the body. These hormones are also involved in development of many diseases. Estrogen is directly related to the diversity of gut flora. Among gut microbes, those encoding  $\beta$ -glucuronidase are involved in the enterohepatic circulation of estrogen.<sup>1</sup> Estrogen accelerates osteoblast differentiation and survival through the NO/cGMPdependent phosphorylation pathway<sup>30</sup> and

the Fas/Fas-ligand system.<sup>31</sup> Moreover, estrogen increases the OPG/RANKL ratio that is dependent on the low-density lipoprotein receptor-related protein 5 pathway, reduces osteoclast differentiation, and maintains bone homeostasis.<sup>32</sup> Intestinal microbes affect the enterohepatic circulation of thyroid hormones by modulating glucuronidase and sulfate activities.<sup>33</sup> Among the thyroid hormones, calcitonin (secreted by thyroid C cells) can inhibit parathyroid hormone, thus reducing bone mobilization, promoting osteoclast apoptosis, and delaying the onset of osteoporosis. However, some studies have shown that calcitonin accelerates bone circulation and inhibits osteoblast proliferation, which is mediated by osteoclasts and bone cells after initiation of bone formation mechanisms.<sup>34</sup> Thyroid hormone membrane-bound receptors also increase eNOS activity,<sup>35</sup> indirectly stimulating osteoblast proliferation and survival through the cGMP-dependent protein kinase G pathway. Insulin-like growth factor-1 (IGF-1) is a growth factor that promotes the quality of life of chondrocytes and osteoblasts.<sup>36</sup> IGF-1 is regulated through the Akt-mTOR-dependent pathway and induces a sustained increase in eNOS levels.<sup>37</sup> Intestinal microbial imbalance indirectly affects osteoblast function and number by causing а decrease in IGF-1 levels.38

Glucagon-like peptide 1 (GLP-1), which is secreted by intestinal L cells, promotes osteogenic differentiation and inhibits mesenchymal stem cells, with conversion to fat.<sup>39</sup> GLP-2 is also secreted by L cells in the intestinal mucosa. GLP-2 has a strong intestinal affinity and can inhibit proliferation of osteoclasts through the transforming growth factor- $\beta$ -SMAD2/3– iNOS–NO–caspase3–B-cell lymphoma 2 signaling pathway.<sup>40</sup> GLP-2 can also upregulate butyric acid levels in the intestine, thus decreasing levels of stress byproducts and delaying apoptosis.<sup>41</sup> GLP-2 also maintains stability of various cells in the intestine. Testosterone and metabolites that protect bone homeostasis are also modulated by the gut microbiome.<sup>42</sup> Insulin-like factor 3 in testosterone affects the musculoskeletal system through the insulin-like factor 3/relaxin family peptide receptor 2 axis.<sup>43</sup> Osteoporosis caused by endocrine dysfunction is often secondary osteoporosis (Figure 1).

### Others

Homocysteine is a non-protein amino acid that is produced during metabolism of methionine. Folic acid is an important cofactor in the metabolism of homocysteine. Intestinal flora disorders lead to reduced absorption of folic acid in the jejunum, leading to hyperhomocysteinemia. Hyperhomocysteinemia not only causes degradation of the extracellular matrix and a decrease in bone blood flow, but also has a harmful effect on bone health.<sup>44</sup> This condition also affects osteoblast precursors, namely human mesenchymal stem cells, which attenuate osteoblast differentiation.<sup>45</sup> Moreover, hyperhomocysteinemia causes an increase in reactive oxygen species production and activation of matrix metalloproteinases, among which matrix metalloproteinase-9 and matrix metalloproteinase-13 play a role in cartilage osteogenesis.46 Additionally, hyperhomocysteinemia leads to activation of protein phospholipase 2A. This disrupts the FOXO1 and mitogen-activated protein kinase signaling cascades and alters the redox regulation mechanism of osteoblasts.

Bifidobacteria in the intestine can significantly enhance transcription of the lactase gene promoter and promote lactose absorption,<sup>47</sup> thus reducing hypocalcemia-induced bone mobilization. *Lactobacillus helveticus* fermentation further promotes the release of free calcium and accelerates calcium deposition.<sup>48</sup> Intestinal microbes are also involved in development of inflammatory bowel disease. This causes a decrease in calcium and vitamin D absorption in the intestine, and indirectly increases the risk of



Abbreviations: OB: osteoblast; OC:osteoclast; Wnt:Wnt singal molecule; MMP-8:matrix metalloproteinases 13; ROS: reactiveosygenspecies; HHCY:hyperhomocysteinemia; Vii D:vitamin D; NO:nitric oxide; High:high levels of NO; Hb-egf:heparin-binding epidermal growth factor; IGF-1:insulin like growth factor 1; GLP-1:glucagon-likepeptide1; Ca:calcium; IL-1:Interleukin 1; TNFc:Tumor necrosis factor cr; IL-6: Interleukin 6; Th17:type 17 T helper cells; Th1:helper T cell 1; Th2:helper T cell 2; Tregs:regulatory T cells; CTL44::ytotoxic T-lymphocyte antigen 4; B cells:B lymphocyte; OPG:osteoprotegenin; GLP-2: glucagon-likepeptide 2; PTH:parathyroid hormone;

**Figure I.** Association between intestinal microorganisms and osteoporosis. The figure shows differences in nitric oxide concentrations, endocrine hormones, immune system-related cells, and antigens in osteoblasts and osteoclasts. High homocysteine, calcium, and vitamin D levels are also involved in gut microbes and osteoblasts

osteoporosis. Intestinal pH values also change because of intestinal flora, thus affecting calcium absorption.<sup>49</sup> *L. reuteri* can inhibit Wnt10b.<sup>50</sup> Wnt, which is a highly conserved signaling molecule, effectively stimulates osteoblast differentiation, inhibits adipogenesis, and impedes bone degeneration.<sup>51</sup> Wnt shows osteoprotective effects by affecting the microenvironment necessary for differentiation of mesenchymal cells into bone precursor cells and preventing osteoblasts from transforming into chondrocytes.<sup>52</sup>

## Therapy

High-fiber probiotic supplements, shortchain fatty acid (SCFA) diets, and fecal transplants are the most studied therapies. They are anti-inflammatory and maintain balance of the gut microbiota. Each of these therapies has its own characteristics. Among them, SCFAs not only increase calcium absorption, but also have antiinflammatory effects and maintain intestimicrobial balance.<sup>6</sup> Additionally, nal SCFAs can reduce intestinal pH and inhibit formation of calcium chelates. SCFAs also increase calcium absorption by increasing the level of calcium binding, protein transcription, and upregulation of the vitamin D receptor.<sup>53</sup> These fatty acids also promote Treg development and stimulate the Treg immune response, which weaken immune system abnormalities caused by intestinal microbes.<sup>54</sup> However, SCFAs play a role in the pathogenesis of hepatic encephalopathy, and an excessive increase in SCFAs may aggravate hepatic coma in patients with liver disease. Even in patients without liver disease, octanoic acid (a SCFA) is associated with experimental hepatic encephalopathy.<sup>55</sup> Further research on the role of SCFAs in liver disease is required. In recent years, heparin-binding epidermal growth factor has been shown

to significantly increase NO and may provide a new treatment for osteoporosis.<sup>56</sup>

### **Conclusions and prospects**

Many studies have confirmed that there is a close relationship between gut microbiota and osteoporosis. Endocrine dysfunction plays an important role in osteoporosis. Immune system abnormalities and NO also affect osteoporosis. Various pathways work together and eventually lead to osteoporosis. The relationship of gut microbes with osteoporosis requires further research, especially the role of specific microorganisms in osteoporosis. This article is a preliminary discussion on the relationship between intestinal flora and osteoporosis. This review provides a theoretical basis for new prevention and treatment strategies for osteoporosis and suggests new ideas for immunomodulation and targeted treatment of osteoporosis.

### Author contributions

Meng-lei Hao and Guang-yao Wang were major contributors in writing and submitting the manuscript. Dong-lai Wang made substantial contributions to conception of the article. Xiao-qin Zuo, Chan-juan Qu, and Bo-chen Yao provided assistance and suggestions during the submission process. All authors read and approved the final manuscript.

### **Declaration of conflicting interest**

The authors declare that there is no conflict of interest.

### Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

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#### References

- Kwa M, Plottel CS, Blaser MJ, et al. The intestinal microbiome and estrogen receptor-positive female breast cancer. *J Natl Cancer Inst* 2016; 108. doi: 10.1093/jnci/ djw029. [Print 2016 Aug].
- 2. Khanna S and Tosh PK. A clinician's primer on the role of the microbiome in human health and disease. In: *Mayo clinic proceedings: 2014.* Elsevier, pp.107–114.
- Blain H, Rolland Y, Beauchet O, et al. Usefulness of bone density measurement in fallers. *Joint Bone Spine* 2014; 81: 403–408.
- 4. Neugebauer J, Heilig J, Hosseinibarkooie S, et al. Plastin 3 influences bone homeostasis through regulation of osteoclast activity. *Hum Mol Genet* 2018; 27: 4249–4262.
- Tatangelo G, Watts J, Lim K, et al. The cost of osteoporosis, osteopenia, and associated fractures in Australia in 2017. *J Bone Miner Res* 2019; 34: 616–625.
- 6. David Yatsonsky I, Pan K, Shendge VB, et al. Linkage of microbiota and osteoporosis: a mini literature review. *World J Orthop* 2019; 10: 123.
- Ohlsson C and Sjögren K. Effects of the gut microbiota on bone mass. *Trends Endocrinol Metab* 2015; 26: 69–74.
- Karl JP, Margolis LM, Madslien EH, et al. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress. *Am J Physiol Gastrointest Liver Physiol* 2017; 312: G559–G571.
- Sjögren K, Engdahl C, Henning P, et al. The gut microbiota regulates bone mass in mice. *J Bone Miner Res* 2012; 27: 1357–1367.
- Mahavadi S, Nalli AD, Kumar DP, et al. Cytokine-induced iNOS and ERK1/2 inhibit adenylyl cyclase type 5/6 activity and stimulate phosphodiesterase 4D5 activity in intestinal longitudinal smooth muscle. *Am J Physiol Cell Physiol* 2014; 307: C402.
- Kleinert H, Schwarz PM and Förstermann U. Regulation of the expression of inducible nitric oxide synthase. *Biol Chem* 2003; 384: 1343–1364.
- 12. Xie Q, Kashiwabara Y and Nathan C. Role of transcription factor NF-kappa B/Rel in

induction of nitric oxide synthase. J Biol Chem 1994; 269: 4705–4708.

- Kalyanaraman H, Schall N and Pilz RB. Nitric oxide and cyclic GMP functions in bone. *Nitric Oxide* 2018; 76: 62–70.
- Li X, Li X, Shang Q, et al. Fecal microbiota transplantation (FMT) could reverse the severity of experimental necrotizing enterocolitis (NEC) via oxidative stress modulation. *Free Radic Biol Med* 2017; 108: 32–43.
- Basso N and Heersche JN. Effects of hind limb unloading and reloading on nitric oxide synthase expression and apoptosis of osteocytes and chondrocytes. *Bone* 2006; 39: 807–814.
- Teixeira CC, Agoston H and Beier F. Nitric oxide, C-type natriuretic peptide and cGMP as regulators of endochondral ossification. *Dev Biol* 2008; 319: 171–178.
- Andrukhova O, Slavic S, Zeitz U, et al. Vitamin D is a regulator of endothelial nitric oxide synthase and arterial stiffness in mice. *Mol Endocrinol* 2014; 28: 53–64.
- Al-Daghri NM, Bukhari I, Yakout SM, et al. Associations of serum nitric oxide with vitamin D and other metabolic factors in apparently healthy adolescents. *Biomed Res Int* 2018; 2018: 1489132.
- Khosravi A, Yáñez A, Price JG, et al. Gut microbiota promote hematopoiesis to control bacterial infection. *Cell Host Microbe* 2014; 15: 374–381.
- 20. Luo C, Wang L, Sun C, et al. Estrogen enhances the functions of CD4+ CD25+ Foxp3+ regulatory T cells that suppress osteoclast differentiation and bone resorption in vitro. *Cell Mol Immunol* 2011; 8: 50.
- Wing K, Yamaguchi T and Sakaguchi S. Cell-autonomous and-non-autonomous roles of CTLA-4 in immune regulation. *Trends Immunol* 2011; 32: 428–433.
- Britton RA, Irwin R, Quach D, et al. Probiotic L. reuteri treatment prevents bone loss in a menopausal ovariectomized mouse model. *J Cell Physiol* 2014; 229: 1822–1830.
- Alnaeeli M, Penninger JM and Teng YT. Immune interactions with CD4+ T cells promote the development of functional osteoclasts from murine CD11c+ dendritic cells. *J Immunol* 2006; 177: 3314–3326.

- 24. Sato K, Suematsu A, Okamoto K, et al. Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction. *J Exp Med* 2006; 203: 2673–2682.
- Kim S, Kim H, Yim YS, et al. Maternal gut bacteria promote neurodevelopmental abnormalities in mouse offspring. *Nature* 2017; 549: 528.
- 26. Xu S, Zhang Y, Liu B, et al. Activation of mTORC1 in B lymphocytes promotes osteoclast formation via regulation of β-catenin and RANKL/OPG. *J Bone Miner Res* 2016; 31: 1320–1333.
- Wesemann DR. Microbes and B cell development. In: *Advances in immunology*. vol. 125. Elsevier, 2015, pp.155–178.
- Dougall WC. Molecular pathways: osteoclast-dependent and osteoclastindependent roles of the RANKL/RANK/ OPG pathway in tumorigenesis and metastasis. *Clin Cancer Res* 2012; 18: 326–335.
- Tong X, Gu J, Song R, et al. Osteoprotegerin inhibit osteoclast differentiation and bone resorption by enhancing autophagy via AMPK/mTOR/p70S6K signaling pathway in vitro. J Cell Biochem 2019; 120: 1630–1642.
- Marathe N, Rangaswami H, Zhuang S, et al. Pro-survival effects of 17β-estradiol on osteocytes are mediated by nitric oxide/ cGMP via differential actions of cGMPdependent protein kinases I and II. J Biol Chem 2012; 287: 978–988.
- Nakamura T, Imai Y, Matsumoto T, et al. Estrogen prevents bone loss via estrogen receptor α and induction of Fas ligand in osteoclasts. *Cell* 2007; 130: 811–823.
- 32. Zhang C, Peng J, Wu S, et al. Dioscin promotes osteoblastic proliferation and differentiation via Lrp5 and ER pathway in mouse and human osteoblast-like cell lines. *J Biomed Sci* 2014; 21: 30.
- Virili C and Centanni M. "With a little help from my friends"-The role of microbiota in thyroid hormone metabolism and enterohepatic recycling. *Mol Cell Endocrinol* 2017; 458: 39–43.
- Naot D, Musson DS and Cornish J. The activity of peptides of the calcitonin family in bone. *Physiol Rev* 2018; 99: 781–805.

- Hiroi Y, Kim HH, Ying H, et al. Rapid nongenomic actions of thyroid hormone. *Proc Natl Acad Sci U S A* 2006; 103: 14104–14109.
- 36. Wang Y, Bikle DD and Chang W. Autocrine and paracrine actions of IGF-I signaling in skeletal development. *Bone Res* 2013; 1: 249.
- 37. Cittadini A, Monti MG, Petrillo V, et al. Complementary therapeutic effects of dual delivery of insulin-like growth factor-1 and vascular endothelial growth factor by gelatin microspheres in experimental heart failure. *Eur J Heart Fail* 2011; 13: 1264–1274.
- Cani PD, Lecourt E, Dewulf EM, et al. Gut microbiota fermentation of prebiotics increases satietogenic and incretin gut peptide production with consequences for appetite sensation and glucose response after a meal–. Am J Clin Nutr 2009; 90: 1236–1243.
- Luo G, Liu H and Lu H. Glucagon-like peptide-1 (GLP-1) receptor agonists: potential to reduce fracture risk in diabetic patients? *Br J Clin Pharmacol* 2016; 81: 78–88.
- 40. Lu Y, Lu D and Hu Y. Glucagon-like peptide 2 decreases osteoclasts by stimulating apoptosis dependent on nitric oxide synthase. *Cell Prolif* 2018; 51: e12443.
- He C, Huang L, Lei P, et al. Sulforaphane normalizes intestinal flora and enhances gut barrier in mice with BBN-induced bladder cancer. *Mol Nutr Food Res* 2018; 62: 1800427.
- Markle JG, Frank DN, Mortin-Toth S, et al. Sex differences in the gut microbiome drive hormone-dependent regulation of autoimmunity. *Science* 2013; 339: 1084–1088.
- De Toni L, Agoulnik AI, Sandri M, et al. INSL3 in the muscolo-skeletal system. *Mol Cell Endocrinol* 2019; 487: 12–17.
- Schalinske KL and Smazal AL. Homocysteine imbalance: a pathological metabolic marker. *Adv Nutr* 2012; 3: 755–762.
- 45. Lanza D, Perna AF, Oliva A, et al. Impact of the uremic milieu on the osteogenic potential of mesenchymal stem cells. *PLoS One* 2015; 10: e0116468.
- Vacek TP, Kalani A, Voor MJ, et al. The role of homocysteine in bone remodeling. *Clin Chem Lab Med* 2013; 51: 579–590.

- Tishkoff SA, Reed FA, Ranciaro A, et al. Convergent adaptation of human lactase persistence in Africa and Europe. *Nat Genet* 2007; 39: 31.
- 48. Han K, Cao J, Wang J, et al. Effects of Lactobacillus helveticus fermentation on the Ca2+ release and antioxidative properties of sheep bone hydrolysate. *Korean J Food Sci Anim Resour* 2018; 38: 1144.
- Chen YC, Greenbaum J, Shen H, et al. Association between gut microbiota and bone health: potential mechanisms and prospective. *J Clin Endocrinol Metab* 2017; 102: 3635–3646.
- Zhang J, Motyl KJ, Irwin R, et al. Loss of bone and Wnt10b expression in male type 1 diabetic mice is blocked by the probiotic Lactobacillus reuteri. *Endocrinology* 2015; 156: 3169–3182.
- Iyer S, Ambrogini E, Bartell SM, et al. FOXOs attenuate bone formation by suppressing Wnt signaling. *J Clin Invest* 2013; 123: 3409–3419.

- 52. Hill TP, Später D, Taketo MM, et al. Canonical Wnt/β-catenin signaling prevents osteoblasts from differentiating into chondrocytes. *Dev Cell* 2005; 8: 727–738.
- Xu X, Jia X, Mo L, et al. Intestinal microbiota: a potential target for the treatment of postmenopausal osteoporosis. *Bone Res* 2017; 5: 17046.
- D'Amelio P and Sassi F. Gut microbiota, immune system, and bone. *Calcif Tissue Int* 2018; 102: 415–425.
- 55. Baraldi M, Zeneroli ML, Ventura E, et al. An increase in cerebral benzodiazepine receptors induced by a subacute administration of ammonia, mercaptans and shortchain fatty acids in rats. *Clin Sci (Lond)* 1987; 73: 669–671.
- Yu X, Radulescu A, Zorko N, et al. Heparinbinding EGF-like growth factor increases intestinal microvascular blood flow in necrotizing enterocolitis. *Gastroenterology* 2009; 137: 221–230.