

Evidence of Clonal Expansion in the Genome of a Multidrug-Resistant *Mycobacterium tuberculosis* Clinical Isolate from Peru

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We report the genome sequence of *Mycobacterium tuberculosis* INS-MDR from Peru, a multidrug-resistant tuberculosis (MDR-TB) and Latin American-Mediterranean (LAM) lineage strain. Our analysis showed mutations related to drug resistance in the *rpoB* (D516V), *katG* (S315T), *kasA* (G269S), and *pnca* (Q10R) genes. Our evidence suggests that INS-MDR may be a clonal expansion related to the African strain KZN 1435.

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The emergence of multidrug-resistant (MDR) strains for tuberculosis (TB) is resulting in one of the major worldwide public health problems. In 2012, the incidence rate for TB in Peru was 95 cases/100,000 population, of which 3.9% were new cases of MDR-TB (1), and from these MDR-TB cases, 79% were from Lima (2). Currently, there is no genomic information on an MDR strain from Peru. Thus, obtaining this information is a key step in understanding the biology of the pathogen and improving treatment for TB (3). Here, we report genomic features of a multidrug-resistant strain of *Mycobacterium tuberculosis*, INS-MDR, from a patient with active TB from Peru.

The establishment of this strain's lineage was based on 24 mycobacterial interspersed repetitive unit-variable number of tandem repeat (MIRU-VNTR) loci (4) and on single-nucleotide polymorphisms (SNPs) based on phylogeny (5). The genome sequencing of INS-MDR was performed using an Illumina HiSeq 2000 sequencer with coverage of 1,331×. Resulting paired-end reads were assembled with BWA v 0.5.9-r16 (6), using the H37Rv genome (NC000962.3) as a reference. The genomic sequence was annotated with the RAST server (7), Prokaryotic Genome Annotation Pipeline (PGAAP), and Clusters of Orthologous Groups (COG) (8) databases. A comparative analysis was carried out between INS-MDR and KZN 1435, an MDR strain from South Africa (9), using SNPsFinder (10) to identify the differences in intergenic coding regions and gene ontology classifications (COG).

It has been determined that INS-MDR belongs to the LAM lineage. We obtained 58,157,302 paired-end reads that, after the assembly, resulted in 22 contigs comprising about 99.98% compared to the H37Rv genome. INS-MDR is 4,383,671 bp long, with an average GC content of 65.6%.

A total of 805 polymorphisms with respect to H37Rv were observed, with 703 of these located in the coding regions that were classified in COG categories as follows: secondary metabolite biosynthesis, transport, and catabolism ($n = 38$); lipid transport and metabolism ($n = 41$); replication, recombination, and repair ($n = 32$); energy production and conversion ($n = 35$); amino acid

transport and metabolism ($n = 30$); carbohydrate transport and metabolism ($n = 27$); cell motility ($n = 21$); cell wall/membrane/envelope biogenesis (M) ($n = 23$); coenzyme transport and metabolism ($n = 22$); signal transduction mechanisms ($n = 22$); inorganic ion transport and metabolism ($n = 22$); transcription ($n = 20$); translation, ribosomal structure, and biogenesis ($n = 16$); posttranslational modification and protein turnover ($n = 13$); nucleotide transport and metabolism ($n = 11$); defense mechanisms ($n = 5$); cell cycle control, cell division, and chromosome partitioning ($n = 9$); and RNA processing and modification ($n = 2$).

We identified a mutation on the *rpoB* gene (D516V) related to resistance to rifampin (11), mutations on the *katG* (S315T) and *kasA* genes (G269S) related to resistance to isoniazid (12, 13), and a mutation on the *pnca* gene (Q10R) related to pyrazinamide resistance (14). Also, we found mutations on the *gyrA* gene (E21Q, S95T, G247S, G668D) and a mutation on the *embB* gene (Y319S) that do not confer resistance (15–17).

Comparison of the KZN 1435 and INS-MDR strains demonstrated that, despite the differences in geographic origin and the high incidence of TB in South Africa (1), the strains have similar proportions of SNPs, showing high degrees of conservation of genome structure. These results suggest that the outbreak of drug-resistant tuberculosis in Lima may be a clonal expansion of the same strain; however, more genomic information is required.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [JAQ100000000](https://www.ncbi.nlm.nih.gov/nuccore/JAQ100000000). The version described in this paper is version [JAQ101000000](https://www.ncbi.nlm.nih.gov/nuccore/JAQ101000000).

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REFERENCES

1. World Health Organization. 2013. Global tuberculosis report 2013. World Health Organization, Geneva, Switzerland.
2. ESN-PCT. 2013. Informe de la Estrategia Sanitaria Nacional de Pre-

- vención y Control de la Tuberculosis sobre Situación y Control de TB-MDR y TB-XDR en Perú. <http://190.40.40.245/newtb/Archivos/RecursoInformacion/20140211114915.pdf>.
3. Abubakar I, Zignol M, Falzon D, Raviglione M, Ditiu L, Masham S, Adetifa I, Ford N, Cox H, Lawn SD, Marais BJ, McHugh TD, Mwaba P, Bates M, Lipman M, Zijenah L, Logan S, McNerney R, Zumla A, Sarda K, Nahid P, Hoelscher M, Pletschette M, Memish ZA, Kim P, Hafner R, Cole S, Migliori GB, Maeurer M, Schito M, Zumla A. 2013. Drug-resistant tuberculosis: time for visionary political leadership. *Lancet Infect. Dis.* 13: 529–539. [http://dx.doi.org/10.1016/S1473-3099\(13\)70030-6](http://dx.doi.org/10.1016/S1473-3099(13)70030-6).
 4. Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rüsç-Gerdes S, Willery E, Savine E, de Haas P, van Deutekom H, Roring S, Bifani P, Kurepina N, Kreiswirth B, Sola C, Rastogi N, Vatin V, Gutierrez MC, Fauville M, Niemann S, Skuce R, Kremer K, Loch C, van Soolingen D. 2006. Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*. *J. Clin. Microbiol.* 44:4498–4510. <http://dx.doi.org/10.1128/JCM.01392-06>.
 5. Filliol I, Motiwala AS, Cavatore M, Qi W, Hazbón MH, Bobadilla del Valle M, Fyfe J, García-García L, Rastogi N, Sola C, Zozio T, Guerrero MI, León CI, Crabtree J, Angiuoli S, Eisenach KD, Durmaz R, Joloba ML, Rendón A, Sifuentes-Osornio J, Ponce de León A, Cave MD, Fleischmann R, Whittam TS, Alland D. 2006. Global phylogeny of *Mycobacterium tuberculosis* based on single nucleotide polymorphism (SNP) analysis: insights into tuberculosis evolution, phylogenetic accuracy of other DNA fingerprinting systems, and recommendations for a minimal standard SNP set. *J. Bacteriol.* 188:759–772. <http://dx.doi.org/10.1128/JB.188.2.759-772.2006>.
 6. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25:1754–1760. <http://dx.doi.org/10.1093/bioinformatics/btp324>.
 7. Meyer F, Paarmann D, D'Souza M, Olson R, Glass EM, Kubal M, Paczian T, Rodriguez A, Stevens R, Wilke A, Wilkening J, Edwards RA. 2008. The metagenomics RAST server—a public resource for the automatic phylogenetic and functional analysis of metagenomes. *BMC Bioinformatics* 9:386. <http://dx.doi.org/10.1186/1471-2105-9-386>.
 8. Tatusov RL, Fedorova ND, Jackson JD, Jacobs AR, Kiryutin B, Koonin EV, Krylov DM, Mazumder R, Mekhedov SL, Nikolskaya AN, Rao BS, Smirnov S, Sverdlov AV, Vasudevan S, Wolf YI, Yin JJ, Natale DA. 2003. The COG database: an updated version includes eukaryotes. *BMC Bioinformatics* 4:41. <http://dx.doi.org/10.1186/1471-2105-4-41>.
 9. Ioerger TR, Koo S, No E-G, Chen X, Larsen MH, Jacobs WR, Jr, Pillay M, Sturm AW, Sacchetti JC. 2009. Genome analysis of multi- and extensively-drug-resistant tuberculosis from KwaZulu-Natal, South Africa. *PLoS One* 4:e7778. <http://dx.doi.org/10.1371/journal.pone.0007778>.
 10. Song J, Xu Y, White S, Miller KW, Wolinsky M. 2005. SNPsFinder—a web-based application for genome-wide discovery of single nucleotide polymorphisms in microbial genomes. *Bioinformatics* 21:2083–2084. <http://dx.doi.org/10.1093/bioinformatics/bti176>.
 11. Lipin MY, Stepanshina VN, Shemyakin IG, Shinnick TM. 2007. Association of specific mutations in katG, rpoB, rpsL and rrs genes with spoligotypes of multidrug-resistant *Mycobacterium tuberculosis* isolates in Russia. *Clin. Microbiol. Infect.* 13:620–626. <http://dx.doi.org/10.1111/j.1469-0691.2007.01711.x>.
 12. Campbell PJ, Morlock GP, Sikes RD, Dalton TL, Metchock B, Starks AM, Hooks DP, Cowan LS, Plikaytis BB, Posey JE. 2011. Molecular detection of mutations associated with first- and second-line drug resistance compared with conventional drug susceptibility testing of *Mycobacterium tuberculosis*. *Antimicrob. Agents Chemother.* 55:2032–2041. <http://dx.doi.org/10.1128/AAC.01550-10>.
 13. Lee AS, Lim IH, Tang LL, Telenti A, Wong SY. 1999. Contribution of kasA analysis to detection of isoniazid-resistant *Mycobacterium tuberculosis* in Singapore. *Antimicrob. Agents Chemother.* 43:2087–2089.
 14. Cheng SJ, Thibert L, Sanchez T, Heifets L, Zhang Y. 2000. pncA mutations as a major mechanism of pyrazinamide resistance in *Mycobacterium tuberculosis*: spread of a monoresistant strain in Quebec, Canada. *Antimicrob. Agents Chemother.* 44:528–532. <http://dx.doi.org/10.1128/AAC.44.3.528-532.2000>.
 15. Plinke C, Rüsç-Gerdes S, Niemann S. 2006. Significance of mutations in embB codon 306 for prediction of ethambutol resistance in clinical *Mycobacterium tuberculosis* isolates. *Antimicrob. Agents Chemother.* 50: 1900–1902. <http://dx.doi.org/10.1128/AAC.50.5.1900-1902.2006>.
 16. Lau RW, Ho PL, Kao RY, Yew WW, Lau TC, Cheng VC, Yuen KY, Tsui SK, Chen X, Yam WC. 2011. Molecular characterization of fluoroquinolone resistance in *Mycobacterium tuberculosis*: functional analysis of gyrA mutation at position 74. *Antimicrob. Agents Chemother.* 55:608–614. <http://dx.doi.org/10.1128/AAC.00920-10>.
 17. Malik S, Willby M, Sikes D, Tsodikov OV, Posey JE. 2012. New insights into fluoroquinolone resistance in *Mycobacterium tuberculosis*: functional genetic analysis of gyrA and gyrB mutations. *PLoS One* 7:e39754. <http://dx.doi.org/10.1371/journal.pone.0039754>.