



# Article Radiomic Features Prognosticate Treatment Response in CAR-T Cell Therapy

Yoganand Balagurunathan <sup>1,2,\*</sup>, Jung W. Choi <sup>2</sup>, Zachary Thompson <sup>3</sup>, Michael Jain <sup>4</sup> and Frederick L. Locke <sup>4,\*</sup>

- <sup>1</sup> Department of Machine Learning, H Lee Moffitt Cancer Center, Tampa, FL 33612, USA
- <sup>2</sup> Department of Diagnostic & Interventional Radiology, H Lee Moffitt Cancer Center, Tampa, FL 33612, USA; jung.choi@moffitt.org
- <sup>3</sup> Department of Biostatistics & Bioinformatics, H Lee Moffitt Cancer Center, Tampa, FL 33612, USA; zachary.thompson@moffitt.org
- <sup>4</sup> Department of Blood and Marrow Transplant, H Lee Moffitt Cancer Center, Tampa, FL 33612, USA; michael.jain@moffitt.org
- \* Correspondence: yoganand.balagurunathan@moffitt.org (Y.B.); frederick.locke@moffitt.org (F.L.L.)

**Simple Summary:** We propose using quantitative imaging metrics (radiomics) derived from the standard of care patient imaging (positron emission tomography/computed tomography, PET/CT) to prognosticate response to advanced axicabtagene ciloleucel (axi-cel) cellular therapy in patients treated for refractory/relapsed Diffused large B-cell lymphoma (DLBCL). We find radiomic (shape, size) characteristics on the imaging scans (PET/CT) of extra nodal lesions are prognostic of survival outcome. These metrics have a high translational ability and can be used for patient selection that can benefit and spare others from these advanced treatments.

Abstract: Background: Diffuse large B-cell lymphomas (DLBCLs) are the most common, aggressive disease form that accounts for 30% of all lymphoma cases. Identifying patients who will respond to these advanced cell-based therapies is an unaddressed challenge. Methods: We propose to develop a radiomics- (quantitative image metric) based signature on the patients' imaging scans (positron emission tomography/computed tomography, PET/CT) and use these metrics to prognosticate response to axi-cel (axicabtagene ciloleucel), autologous CD19 chimeric antigen receptor (CAR) T-cell (CAR-T) therapy. We curated a cohort of 155 patients with relapsed/refractory (R/R) DLBCL who were treated with axi-cel. Using their baseline image scan (PET/CT), the largest lesions related to nodal/extranodal disease were identified and characterized using imaging metrics (radiomics). We used principal component (PC) analysis to reduce the dimensionality of these features across the functional categories (size, shape, and texture). We evaluated the prognostic ability of radiomic-based PC to treatment response (1-year), measured by overall survival (OS) and progression-free survival (PFS). Results: We found that radiomic PC was prognostic of overall survival (Shape-PC, q < 0.013/0.0108, Size-PC, q < 0.003/0.0088), in CT/PET, respectively. In comparison, the metabolic tumor volume (MTV) was prognostic (q < 0.0002/0.0007). The radiomic PCs across the functional categories showed moderate to weak correlation with MTV, Spearman's  $\rho$  of 0.44/0.35/0.27, and 0.45/0.36/0.55 for Size/Shape/Texture-PC1 obtained on PET and CT, respectively. Conclusions: We found radiomic PC based on size and shape metrics that are able to prognosticate treatment response to CAR-T therapy.

**Keywords:** image biomarkers in lymphoma; radiomics in CAR-T; response to CAR-T; axi-cel biomarkers



Academic Editor: Alexandre Escargueil

Received: 15 April 2025 Revised: 22 May 2025 Accepted: 27 May 2025 Published: 30 May 2025

Citation: Balagurunathan, Y.; Choi, J.W.; Thompson, Z.; Jain, M.; Locke, F.L. Radiomic Features Prognosticate Treatment Response in CAR-T Cell Therapy. *Cancers* **2025**, *17*, 1832. https://doi.org/10.3390/ cancers17111832

Copyright: © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/ licenses/by/4.0/).

## 1. Introduction

Lymphomas, heterogeneously diverse hematological malignancies that arises from B cells, T cells, or natural killer (NK) cells, are broadly classified into Hodgkin lymphoma (HL) and non-Hodgkin lymphoma (NHL). Each disease presents unique clinical and genetic features requiring tailored treatments [1,2]. Diffuse Large B-Cell Lymphomas (DLBCLs) are the most common form of non-Hodgkin lymphoma, accounting for about 30% of the cases, and are aggressive disease subtypes with a 5-year survival of 65% across all grades [3,4]. It is reported that about 30 to 40% of cases may present with advanced relapsed/refractory (R/R) stages that currently rely on salvage treatment that includes intensive chemotherapy and a stem cell transplantation [5]. Axicabtagene ciloleucel (axi-cel), chimeric antigen receptor (CAR) T-cell therapy using a CD19 target, has demonstrated superior levels of durable response in R/R DLBCL, currently recommended as a second-line treatment [6–8]. There have been efforts to develop clinical biomarkers, such as the International Prognostic Index (IPI), to prognosticate survival across risk groups in DLBCL based on retrospective data [9]. This index uses pretreatment clinical variables such as serum Lactate Dehydrogenase (LDH) and the Eastern Cooperative Oncology Group (ECOG) status, clinical stage, and extranodal site, which have been shown to be useful at diagnosis of the disease [10]. These markers do not provide a clear benefit in predicting disease progression or relapse beyond disease assessment at baseline [11]. Current clinical consensus criteria for disease staging and response include the Cheson and Lugano classification [12,13], but they are unable to prognosticate treatment response. Metabolic tumor volume (MTV) measure gross tumor burden, and these metrics have been recently shown to be prognostic of treatment response in CAR-T [14–16]. However, these metrics are time-consuming to compute and do not provide any insights into the disease progression.

There is an urgent clinical need to develop a biomarker that can be used to identify patients who would benefit from cellular therapy. Improvements in imaging modalities have led to better staging and detection of DLBCL on <sup>18</sup>F fluorodeoxyglucose (<sup>18</sup>F-FDG) positron emission tomography (PET), along with high-resolution computed tomography (CT) imaging that has allowed for more precise evaluation of disease condition, tumor biology, and its microenvironment [17].

Quantitative imaging metrics (radiomics) have shown enormous promise in reflecting the pathophysiology of the tumor and describing the textural heterogeneity and shape characteristics, and have been shown to prognosticate the disease progression across oncological diseases [18–20]. These metrics have been recently shown to be useful in predicting outcomes in DLBCL [21,22]. In this study, we propose to use radiomic features on patient imaging (PET, CT) of DLBCL and test their ability to prognosticate treatment response for CAR-T therapy, complementing our predictive models presented in our previous work [22]. Our study overview is illustrated in Figure 1.

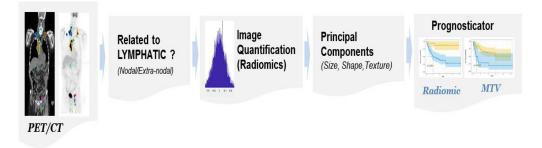


Figure 1. The study process flow shows the use of patient-level imaging metrics (radiomics, MTV).

## 2. Materials and Methods

#### 2.1. Patient Data

The study retrospectively curated 155 patient records with R/R diffuse large B-cell lymphoma (DLBCL) who had received CAR-T therapy (axi-cel). We obtained 100 patients from the H Lee Moffitt Cancer Center, and 55 anonymized patient records were obtained from the clinical trial consortium (Zuma-1, sponsor Kite Pharma). Patients with non-measurable lesions or with or without baseline PET imaging were excluded from the study. Bridging therapy was defined as any lymphoma-specific therapy given before CAR T-cell infusion, prior to the start of fludarabine cyclophosphamide chemotherapy for lymphodepletion, previously presented [14,22].

Our study was approved by the Institutional Review Board (IRB) at the University of South Florida /Moffitt Cancer Center. Patients with baseline imaging (<sup>18</sup>F FDG-PET/CT) prior to CAR T-cell therapy were included. Most patients received bridging therapy as a standard of care before treatment, defined as any lymphoma-specific therapy. Table 1 shows the patient cohort characteristics. Imaging data corresponding to whole-body CT and FDG-PET (attenuated corrected) image voxel values were converted to standardized uptake value (SUV) reference before our analysis.

		Subcohorts (Lesion Level)		
Characteristics	All Patients * (N = 155)	Extra-Nodal ( $n = 94$ )	Lymphatic ( <i>n</i> = 124)	
Age (mean, median, std.dev)	60.1	59.4	61	
rige (mean, meanin, statuev)	(63, 12.2)	(63.5, 12.8)	(63, 10.9)	
Sex (male/female/unavailable)	61/39/55	36/22	53/29	
IDU (maan madian atd day)	400.5	448.3	408.6	
LDH (mean, median, std.dev)	(266, 348.25)	(275.5, 406.79)	(267.5, 353.4)	
ECOG				
0–1	83	48	66	
2–3	17	10	16	
	/death			
One Very Program	No	24 (41.4%)	38(46.3%)	
One Year Progression	Yes	34 (58.6%)	44 (53.7%)	
	Unavailable 55			
Stage				
I/II	22	10	14	
III/IV	78	48	68	
Bridge Therapy				
Yes	Yes: 50	Yes: 28	Yes: 41	
No	No: 50	No: 30	No: 41	
Unavailable	55			
Axi-cel therapy				
Trial (cancer center)	100	58	82	
Consortium (Zuma-1)	55	36	42	

 Table 1. Patient demographics for the data used for radiomic analysis.

\* Some clinical variables were unavailable for consortium patients. Lactate Dehydrogenase (LDH), Eastern Cooperative Oncology Group (ECOG), axicabtagene ciloleucel (axi-cel).

#### 2.2. Metabolic Tumor Volume

We used custom tools implemented on MIM PACS (version 6.8.4, MIM Software<sup>®</sup>, Cleveland, OH) to semi-automatically identify lesions with SUV uptake over a reference

liver region ( $\geq$ 2 cm diameter), which was manually located, following the PERSIST criteria [23]. A clinical expert (radiologist J.W.C.) evaluated these abnormal regions and removed falsely detected regions. Most common false detections were due to physiological processes (brain/bladder/etc.), and a few others were removed due to inflammation. We followed the consensus criteria for lesion identification on the PET image scans [23,24], and considered voxels over the 41% of SUVmax to define the lesion boundary on these images. We then summed metabolically active regions across the body to obtain Metabolic Tumor Volume (MTV), reported in multiple scales, milliliters (mL), and cubic centimeters (cc). The metrics have been previously presented in DLBCL patients [14].

#### 2.3. Radiological Review

Lesions were semi-automatically identified on the PET scan and reviewed by our research/clinical radiologist (J.W.C). Additional details about the lesion's anatomical location and association with the lymphatics (nodal vs. extranodal) and volume/size were recorded. There were 1058 lesions reviewed across the patient cohort, of which 342 were related to nodal (lymphatics), and 616 were related to extranodal (non-lymphatic). The patient sub-cohort was organized by grouping the largest lesion in the nodal (n = 124) and extranodal (n = 94), independently. The nodal cohort had most lesions in the abdomen, pelvis, and neck, which are three major organ sites. The extranodal group had the most lesions in the lungs, bone, and liver, which were the major organ sites across the patients in the cohort.

#### 2.4. Radiomics

The CT and FDG-PET imaging were resampled to a common reference resolution of  $1 \times 1 \times 1 \text{ mm}^3$  using bilinear interpolation. The PET images were standardized to SUV units using the activity concentration to the dosage of <sup>18</sup>F-FDG injected volume and patient body weight. The abnormal region was converged on the PET images and was translated to CT. We extracted 306 radiomic features for the identified lesion in each modality: CT and PET (SUV) images, totaling 612 features for a lesion. We categorized imaging radiomic features into three broad functional categories: Size (n = 38), Shape (n = 9), and Texture (n = 259); details on the feature descriptors are deferred to Supplemental Tables S1–S3, which were previously reported [22]. The radiomic feature descriptor definition followed the recommendations of the Image Biomarker Standardization Initiative (IBSI) consensus criteria [25–27]. Features with a minimal change (coefficient of variance  $\leq 3\%$ ) across the patient samples were removed due to the invariant nature of the metric.

#### 2.5. Statistical Analysis

The relationship between the radiomics feature-based principal components (PC) and the metabolic tumor volume (MTV) was assessed using Spearman's correlation coefficients (see Table 2). We used Kaplan–Meier (KM) survival analysis to estimate the survival function of the identified patient groups, separated using the imaging metrics (radiomics). We used the midpoint (Median) on the PC metric to divide patients into groups and evaluate the treatment response measured by survival time to event (OS/PFS). The process was independently tested across the feature categories (size, shape, texture). We quantified the difference in the survival functions using log-rank statistical hypothesis testing to estimate the significance [28]. We computed the proportional hazard ratio (HR) for these cohorts using the Cox-regression model [29] and reported concordance (C-index), to continuous event time measured by survival time (OS/PFS) [30]. A *p*-value of less than 0.05 was considered statistically significant in our analysis. We corrected for multiple testing by computing adjusted *p*-value (q-value or false discovery rate) [31].

Radiomic Metric (Principal Component, PC)	Spearman Correlation (ρ)	<i>p</i> -Value
Lym	phatics (CT Images)	
Size PC1	0.368	0.0000268
Shape PC1	0.3002	0.0007
Texture PC1	-0.345	0.00008
Extra	-Nodal (CT Images)	
Size PC1	-0.4519	0.000004
Shape PC1	0.3698	0.00024
Texture PC1	0.5535	0.00
Lymp	phatics (PET Images)	
Size PC1	0.3698	0.000023
Shape PC1	0.2717	0.0023
Texture PC1	0.175	0.0518
Extra-	Nodal (PET Images)	
Size PC1	0.4496	0.0535
Shape PC1	0.3590	0.00038
Texture PC1	-0.275	0.0073

**Table 2.** The relationship between metabolic tumor volume (MTV) and radiomics features PC (principal components) across feature categories was assessed using Spearman's correlation coefficient.

#### 3. Results

The study assessed patients' baseline PET and CT scans (n = 155), and we formed sub-cohorts based on the anatomical location of the largest lesion related to the lymphatic or nodal (n = 124) and non-lymphatic or extranodal (n = 94). The most frequently involved nodal sites (lymphatic) were the abdominal (46.77%), pelvic (17.74%), and mediastinal (8.87%). The most frequently involved extranodal sites were lung (36.17%), musculoskeletal (21.28%), and pelvic (8.51%). The radiomic features were categorized into functional categories: Size (n = 38), Shape (n = 9), and Texture (n = 259), and the lesions were characterized independently across the imaging modalities (CT and PET).

We obtained principal components (PC) on features in each of the feature categories. We found the size-based PCs in extranodal CT and PET (-SUV) images showed the highest correlation with metabolic tumor volume (MTV),  $\rho = -0.451$  and 0.449, respectively. Texture and Shape-based PCs on lymphatic lesions in PET (SUV) scans had the lowest correlation with MTV, a  $\rho = 0.175$  and 0.271, respectively; see Table 2. We report the top five individual features in each of the functional categories, selected based on those with the highest loading factors for the respective principal components. In the shape category, the shape-PCs are related to compactness and sphericity-based features (loading of -0.43). We found that size-based PCs showed higher loading related to volume fraction (at 10% and 90%), which are extracted in PET images. The texture-based PCs showed higher loading factors related to co-occurrence type features in PET images (see Table 3).

		Loading Factors			Median Value	
	Features	PC1	PC2	PC3	MTV (Median): 169.94 mL	
	Shape-Related Feat	ures				
1	Compactness_1	-0.431	-0.122	0.024	_	
2	Compactness_2	-0.414	-0.074	0.025	- Shape-PC1 (Median):	
3	Spherical_disproportion	0.405	0.224	0.017	-0.1909	
4	Sphericity	-0.434	-0.141	0.020	_	
5	Asphericity	0.405	0.224	0.017	_	
	Size-Related Featu	res				
1	SUV (volume_at_intensity_fraction_10)	-0.031	-0.123	-0.338	_	
2	SUV (volume_at_intensity_fraction_90)	-0.147	-0.032	-0.149	– Size-PC1 (Median):	
3	SUV (intensity_at_volume_fraction_10)	-0.097	-0.101	-0.297	-0.2013	
4	SUV (intensity_at_volume_fraction_90)	0.023	-0.129	-0.331	_	
5	SUV (volume_at_intensity_fraction_difference)	0.011	-0.121	-0.316	_	
	Texture-Related Fea	tures				
1	SUV (avg_coocurrence_joint_max)	0.030	0.001	-0.078	_	
2	SUV (avg_coocurrence_joint_average)	-0.012	0.054	0.003	- Texture-PC1 (Median):	
3	SUV (avg_coocurrence_joint_variance)	0.050	-0.104	0.020	-2.9435	
4	SUV (avg_coocurrence_joint_entropy)	-0.064	0.055	0.044	_	
5	SUV (avg_coocurrence_difference_average)	0.074	-0.129	0.060	_	

**Table 3.** Top loading factors (absolute value) for principal components (PC1 to PC3) on individual radiomic features estimated in the entire cohort.

We tested the ability of radiomics-based principal components to predict treatment outcome, measured by the overall survival, OS, and progression-free survival, PFS (censored at 1 year). We then evaluated shape, size, and texture-based radiomic PCs derived from PET and CT image data in the extranodal cohort that were significant (p < 0.05) for OS and PFS.

We found that the shape- and size-based radiomic PC metrics on extranodal cohort were significant (p < 0.05) for OS and PFS, in PET images (see Table 4). We illustrate patients' image scans with high and low shape metrics (see Figure 2). We used the Cox-regression model to assess patients' proportional hazard (HR) to treatment outcome, measured by survival (OS/PFS). We first evaluated the clinical metric, MTV, to outcome, and found an HR of 3.885 and 3.64 for nodal (lymphatic) and extra-nodal (non-lymphatic) cohorts. The c-index is 0.67/0.65 for nodal (lymphatic)/extra-nodal (non-lymphatic) cohorts (see Table 5). We found that size and shape PCs on extra nodal CT and PET radiomic PCs were significant and showed higher risk for adverse outcomes (c-index 0.59 to 0.6 for shape and 0.61 to 0.6 for size, in OS/PFS, respectively). Texture-based radiomic PCs on extranodal CT show significance (c-index 0.61/0.6 for OS) (see Table 5).

**Table 4.** Prognostic value of radiomics feature-based metrics (principal components, PCs), measured by overall survival (OS) and progression-free survival (PFS), 1 Year after axi-cel therapy. The lesions were stratified into sub-cohorts (nodal and extra-nodal) across modalities: (A) CT and (B) PET (SUV) imaging. The MTV (metabolic tumor volume) was computed at the patient level. Adjusted *p*-value (false discovery rate, q-value) was also reported; \* *p* < 0.1.

		A. Metrics on C	Computed Tomography (CT	) Imaging		
		Survival Statistics (Log-rank <i>p</i> -Value, q-Value)				
	Variable	OverAll Survival (OS)		Progression Free Survival (PFS)		
		Nodal ( <i>n</i> = 126)	Extra Nodal ( $n = 94$ )	Nodal ( <i>n</i> = 126)	Extra Nodal ( $n = 94$ )	
1	MTV (total body)	<0.0001 * (0.0003)	<0.0001 * (0.0002)	0.00024 * (0.0096)	0.00017 * (0.00096)	
2	Shape-PC1	0.75 (0.75)	0.008 * (0.013)	0.67 (0.72)	0.017 * (0.0272)	
3	Size-PC	0.18 (0.24)	0.0012 * (0.003)	0.22 (0.293)	0.00046 * (0.0012)	
4	Texture-PC	0.58 (0.663)	0.0037 * (0.007)	0.72 (0.72)	0.0022 * (0.0044)	

B. Metrics on Positron Emission Tomography (PET) Imaging

		Survival Statistics (Log-rank <i>p</i> -Value, adjusted <i>p</i> -Value)				
	Variable	Over All Survival (OS)		Progression free Survival (PFS)		
		Nodal ( <i>n</i> = 126)	Extra Nodal ( $n = 94$ )	Nodal ( <i>n</i> = 126)	Extra Nodal ( $n = 94$ )	
1	MTV (total body)	<0.0001 * (0.0003)	<0.0001 * (0.00069)	0.00024 * (0.00096)	0.00017 * (0.00096)	
2	Shape-PC1	0.072 (0.1152)	0.0054 * (0.0108)	0.052 (0.0832)	0.0074 * (0.0148)	
3	Size-PC	0.16 (0.2133)	0.0033 * (0.0088)	0.21 (0.2400)	0.00061 * (0.0016)	
4	Texture-PC	0.37 (0.4229)	0.73 (0.730)	0.089 (0.11897)	0.43 (0.430)	

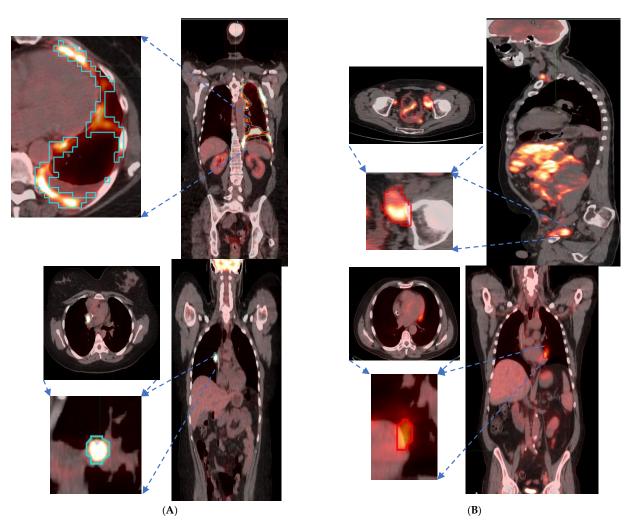
**Table 5.** The proportional hazard ratio using the Cox regression model was used to assess the role of MTV and Radiomic features in treatment outcome (Overall survival, OS) and Progression-free survival, PFS). Evaluated across the sub-cohorts divided based on nodal status (nodal and extranodal) and modalities: (A) CT and (B) PET (SUV) related to OS, (C) CT and (D) PET (SUV) metrics related to PFS. Adjusted *p*-value (false discovery rate, q-value) was also reported.

	A. Metrics on CT Imaging: OS					
		Nodal ( <i>n</i> = 126)		Extra Nodal ( $n = 94$ )		
	Variable	HR ( <i>p-</i> Value, q-Value)	C-Index (CI)	HR ( <i>p-</i> Value, q-Value)	C-Index (CI)	
1	MTV (total body)	3.885 (0.00001), 0.0001	0.67 [0.616,0.721]	3.644 (0.000093), 0.0004	0.65 [0.575, 0.721]	
2	Shape-PC1	1.183 (0.5424), 0.5762	0.52 [0.426, 0.615]	2.383 (0.0062), 0.0123	0.6 [0.516, 0.69]	
3	Size-PC	1.524 (0.1293), 0.172	0.56 [0.464, 0.647]	2.546 (0.0021), 0.0056	0.62 [0.54, 0.695]	
4	Texture-PC	0.858 (0.5762), 0.5762	0.52 [0.427, 0.623]	2.227 (0.0103), 0.0164	0.59 [0.495, 0.682]	

		B. Me	trics on PET Imaging	: OS		
		<b>Nodal</b> ( <i>n</i> = 2	126)	Extra Nodal ( $n = 94$ )		
	Variable	HR ( <i>p-</i> Value, q-value)	C-Index (CI)	HR ( <i>p-</i> Value, q-Value)	C-Index (CI)	
1	MTV (total body)	3.885 (0.000014), 0.0001	0.67 [0.603, 0.733]	3.644 (0.000093), 0.0004	0.65 (0.57, 0.725)	
2	Shape-PC1	1.488 (0.1518), 0.2023	0.55 [0.438, 0.653]	2.139 (0.0148), 0.0296	0.59 (0.504, 0.683)	
3	Size-PC	1.542 (0.1188), 0.1901	0.56 [0.466, 0.65]	2.477 (0.0044), 0.0117	0.61 (0.516, 0.696)	
4	Texture-PC	1.175 (0.5568), 0.6364	0.52 [0.426, 0.608]	0.899 (0.7275), 0.7275	0.51 (0.424, 0.602)	
		C. Me	trics on CT Imaging:	PFS		
	<b>X7 + 1 1</b>	Nodal ( <i>n</i> = 126)		Extra Nodal ( $n = 94$ )		
	Variable	HR ( <i>p</i> -Value, q-Value)	C-Index (CI)	HR ( <i>p</i> -Value, q-Value)	C-Index (CI)	
1	MTV (total body)	2.486 (0.0004), 0.0015	0.62 [0.553, 0.68]	2.814 (0.0003), 0.0015	0.62 [0.565, 0.685]	
2	Shape-PC1	1.111 (0.6683), 0.7241	0.51 [0.434, 0.586]	1.921 (0.0192), 0.0309	0.58 [0.494, 0.662]	
3	Size-PC	1.355 (0.2194), 0.2926	0.54 [0.439, 0.639]	2.417 (0.0014), 0.0037	0.61 [0.519, 0.699]	
4	Texture-PC	0.917 (0.7241), 0.724	0.52 [0.429, 0.611]	2.323 (0.003), 0.006	0.6 [0.51, 0.682]	
		D. Me	trics on PET Imaging:	: PFS		
	X7 1. 1 .	Nodal ( $n = 1$	Nodal ( <i>n</i> = 126)		n = 94)	
	Variable	HR ( <i>p</i> -Value, q-Value)	C-Index (CI)	HR ( <i>p</i> -Value, q-Value)	C-Index (CI)	
1	MTV (total body)	2.486 (0.0003), 0.0015	0.62 [0.563, 0.67]	2.814 (0.0003), 0.0015	0.62 [0.56, 0.689]	
2	Shape-PC1	1.615 (0.0545), 0.0872	0.56 [0.463, 0.659]	2.078 (0.0087), 0.0175	0.6 [0.529, 0.665]	
3	Size-PC	1.36 (0.2135), 0.2440	0.54 [0.447, 0.633]	2.598 (0.0009), 0.0025	0.6 [0.533, 0.674]	
4	Texture-PC	1.521 (0.0910), 0.1214	0.56 [0.477, 0.638]	0.806 (0.4284), 0.4284	0.54 [0.424, 0.647]	

#### Table 5. Cont.

We compared the radiomic feature-based principal component (PC1) computed in each of the feature categories (size, shape, texture) across patients separated based on their clinical disease condition at follow-up time (disease progression and non-progression). We found a significant difference in the radiomics PC metric across all feature categories (see Figure 3a). We then repeated the comparison in patients with lower tumor volume (MTV < 147.5) and higher tumor volume (MTV  $\geq$  147.5). The MTV cut-point of 147.5 mL was established in previously published findings [14], to prognosticate patient response. We found that shape PC and size PC metrics show significant differences between the progressors and non-progressors in the lower tumor volume group (see Figure 3b). In the larger tumor volume groups, we do not see significant differences in clinical progression (See Figure 3c). We used the radiomic PCs to prognosticate treatment outcomes (OS/PFS), using the Kaplan–Meier survival analysis across, and statistical significance was assessed using a log-rank test (See Figure 4).



**Figure 2.** Patient scans showed representative slices in the fused image (CT/PET) with a pointing arrow towards lesions with significant uptake. Representative patients (Lung and Abdominal) image slice for (**A**) High shape metric and (**B**) Low shape metric.

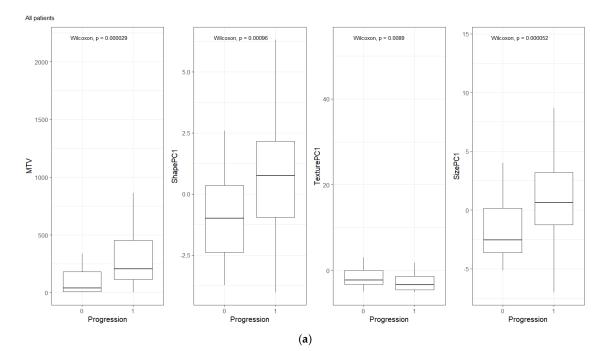
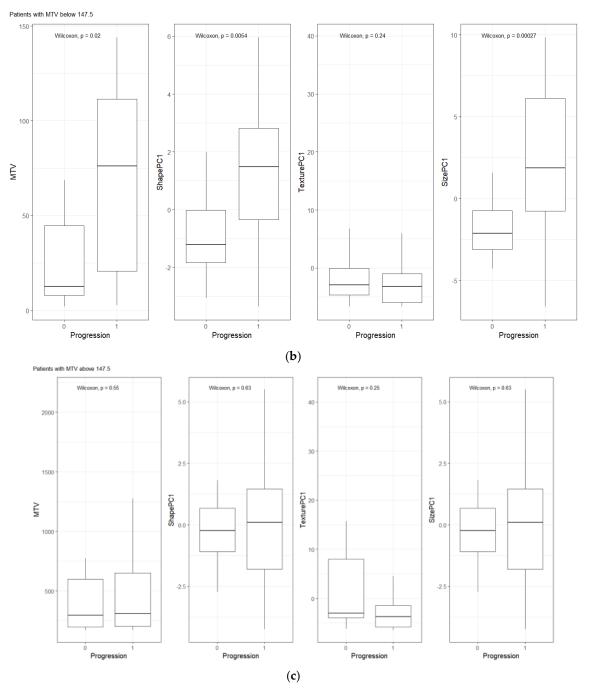


Figure 3. Cont.



**Figure 3.** Distribution of patients based on clinical outcome (progression at 1-year) for MTV, Shape PC and Texture PC for (**a**) all patients, (**b**) patients with lower total tumor burden, and (**c**) patients with higher tumor burden. The cut-point value was based on prior work (*Blood Adv.*, 4(14) 2020) [14].

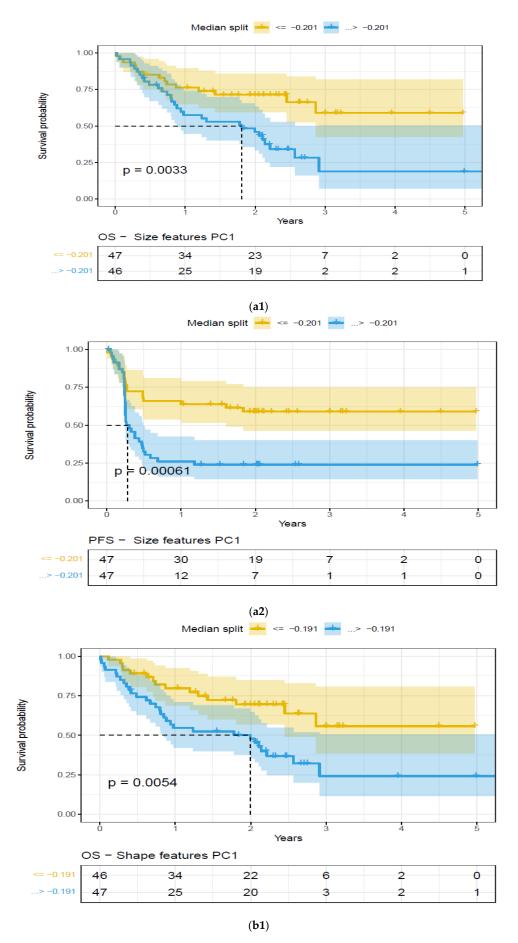


Figure 4. Cont.

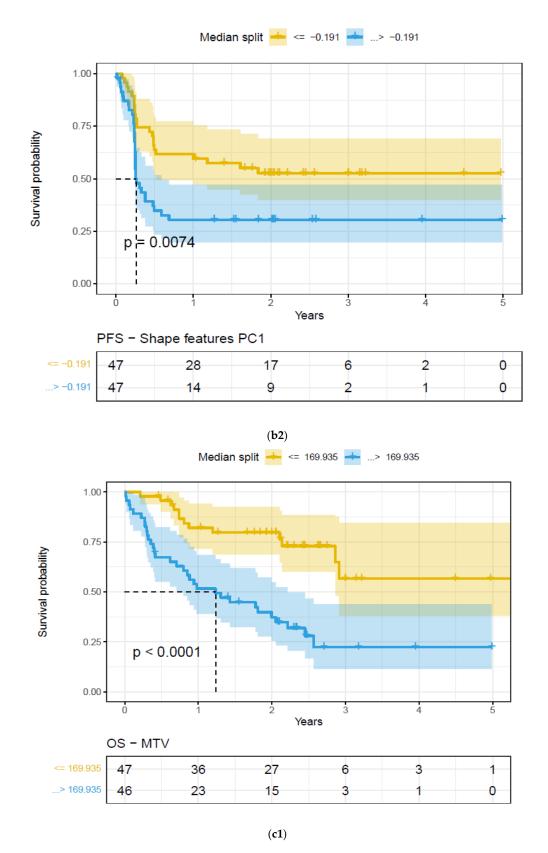
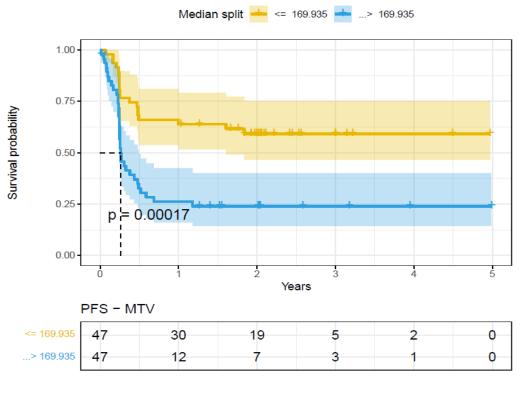


Figure 4. Cont.



(c2)

**Figure 4.** Kaplan–Meier (KM) plots of patient cohort separated using median split based on features extracted on the largest extra-nodal lesions in a patient's PET/SUV scans, for feature categories based on Principal Component (PC1)—Overall survival. Feature categories PC1, KM plots, (**a**) size-based principal components (PC1)—OS (**a1**) and PFS (**a2**), (**b**) shape-based principal components (PC1)—OS (**b1**) and PFS (**b2**), (**c**) metabolic tumor volume—OS (**c1**) and PFS (**c2**), (See details in Table 4).

## 4. Discussion

This study used retrospective patient data to develop a quantitative imaging metric (radiomics) signature derived from their baseline PET/CT scans that could be used as a biomarker to prognosticate response (1-year) in patients undergoing axi-cel therapy. We systematically assessed the lesions based on their role in the lymphatic system (nodal, extra-nodal) and categorized them based on the functional description (size, shape, texture). We showed that radiomic-based features (PC on shape and size) derived from the extra-nodal lesions have a higher risk of adverse outcomes and are a prognostic indicator of survival.

The heterogeneous nature of lymphoma disease makes it challenging to assess and prognosticate treatment response [32,33]. Current clinical consensus assessment criteria [12,13], do not fully capture the disease condition, nor can they predict disease prognosis, especially in axi-cel therapy, leaving a need for better biomarkers. Characterizing subtle physiological changes seen on radiological imaging with quantitative metrics (radiomics) that describe them has been shown to be related to disease conditions and their response outcomes [18–20]. This study is unique in many aspects; firstly, we use lesion-based metrics across many organ sites and form predictors to evaluate the ability to prognosticate survival outcome. Secondly, we subdivided the lesions at the patient level based on nodal (or lymphatic) and extra-nodal (or non-lymphatic) sites. Our study showed that the principal components of radiomic features have the ability to prognosticate patient treatment outcomes to axi-cel treatment. Few other teams have shown similar outcomes without distinguishing the disease sites [21,34].

Beyond tumor metabolic activity, the shape of the lesions observed on imaging scans (PET, CT) can potentially play a diagnostic role in lymphoma [25,35–38]. In current clinical practice, lesion tracer uptake (PET activity) is assessed on a point scale with qualitative reference to the normal metabolic uptake (liver or mediastinum) [12,39]. Grayscale characteristics in PET/CT image scans can be quantified with metrics describing the lesions' size, shape, or texture. There has been abundant literature that has shown the usefulness of radiomic features in predicting the pathological cancer status, oncogenic mutations, or clinical response [18,40–43]. There have been many studies that have attempted to assess the response of axi-cel therapy in DLBCL [7,44]; most useful quantitative biomarkers have been limited to gross measures such as the MTV. These metrics are known to describe the extent of the disease and have been reported to be prognostic of treatment response [14,45-47]. In our prior work [22], we tested the ability of the radiomic metrics to predict the treatment outcome of axi-cel therapy (AUC 0.68, compared to TMV 0.72), while others have shown the utility of radiomic features (on PET images) related to maximum intensity, skewness, major axis, and gray-level emphasis, which were predictive of the treatment outcome (AUC of 0.73, compared to TMV of 0.66) and prognostic of OS/PFS for 3-month follow up [21]. In comparison, our study identifies the largest lesion in nodal/extra nodal and characterizes them using descriptors in three major feature categories (size, shape, texture), and tests their ability to prognosticate treatment outcome (OS/PFS). The study shows that shape-based descriptors in the extra-nodal cohort on PET images show a significant survival difference (OS/PFS) after axi-cel treatment (1-year). Our findings have been validated by several groups, and patients with extra-nodal lesions are reported to have worse outcomes [48,49]. In a multivariate analysis, the number of sites of extra nodal disease with initial high LDH at lymphodepletion was reported to have an inferior survival outcome [49]. In lymphoma, single variable-based models with reproducible thresholds are often used to prognosticate patient outcomes [50,51]. Our methodology uses PCs on the radiomic features in functional categories, where the size category has 38 features, the shape category has nine features, and the texture category has 259 features, see Supplementary Materials Table S1. An ensemble combination (like principal component, PC) of these features with similar function, in these categories, provides a better basis that could be used as a biomarker to prognosticate treatment response. In our study, we found non-size-based PCs (Shape-based PC1) on extranodal lesions to be prognostic (p < 0.05) of OS/PFS on PET, and non-size-based PCs (Shape and Texture based PC1) on CT images were also found to be prognostic (p < 0.05) of OS/PFS (see Table 4, Figure 4) and higher risk for survival outcome (see Table 5). We find these radiomic PCs (Shape, Texture and Size) had a low level of correlation with the metabolic tumor volume metric (MTV), computed at the patient level (highest Spearman's,  $\rho$  of 0.45, 0.37, 0.55, respectively). In solid tumors, it has been reported that most proliferative lesions are non-circular, and irregular compared to benign lesions [37,52,53]. A recent publication [54] reported that a high metabolic heterogeneity measured as a cumulative SUV-histogram on PET images has a poor prognosis for patients with DLBCL.

There are few studies that have tracked the toxicities related to advanced immunotherapy [55], which can certainly be an investigative area. Recent advancements in genomics biomarkers in other blood-based malignancies have shown promise in prognosticating disease progression [56,57]. Advancements in artificial intelligence methods will certainly play a role in identifying finer patterns in patients that could be used to improve patient care in these advanced treatments [58–60], there are several data requirements that need to be considered prior to clinical utility [61,62].

Our study demonstrates the usefulness of lesion-based radiomic PCs to prognosticate treatment outcomes. This study has identified shape and size characteristics (shape PC and size PC, texture in CT) on imaging scans that can serve as surrogates to treatment

response. The study contributions can be summarized as follows: (a) identifying the role of shape/size-based features, (b) association with extra nodal lesions.

These metrics, independently or collectively, could help select patients who would benefit from advanced axi-cel therapy and improve care delivery effectively. Our study, despite showing novel findings, would require secondary validation for clinical translation.

### 5. Limitations

Our study is limited by retrospective data collection, and our findings would need an independent validation cohort prior to clinical translation. Our study did not collect or relate to cell therapy-related toxicities like cytokine release syndrome (CRS), immune effector cell-associated neurotoxicity (ICANS), prolonged cytopenia, tumor lysis syndrome, and others as previously reported [55].

## 6. Conclusions

The study investigated the prognostic role of radiological image metrics (radiomics) on PET/CT images and related them to responses to immunotherapy. The study identified radiomic signatures based on shape and size features to be prognostic of axi-cel therapy. These image-based metrics derived at the lesion level will potentially allow greater clinical translation in recruiting patients who will benefit from these treatments.

**Supplementary Materials:** The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/cancers17111832/s1, Table S1: Feature categories (3D Image features\*; Table S2: Quantitative Image features\* used to describe the lesion of interest, computed independently in each image modality (CT/PET); Table S3: Description of Texture Features.

**Author Contributions:** J.W.C., F.L.L. and Y.B.: Hypothesis, methods, direction; F.L.L. and M.J.: Clinical data curation, clinical care; J.W.C.: lesion identification and marking; Y.B. and F.L.L.: results inference, implementation of methods; Z.T.: statistical analysis; Y.B., F.L.L. and J.W.C.: manuscript writing; All: manuscript editing and approval. All authors have read and agreed to the published version of the manuscript.

**Funding:** Y.B. and F.L.L.: Research grant from Kite pharma<sup>®</sup>, which partly supported their research time.

**Institutional Review Board Statement:** The study was approved by the University of South Florida/Moffitt Cancer Center's IRB. Patients waived consent to use clinical data, including imaging, for retrospective research without identifiable patient information.

**Informed Consent Statement:** The study used de-identified patient records, including imaging data. Patients waived informed consent for research use.

**Data Availability Statement:** The de-identified patient records for those treated at Moffitt Cancer Center will be shared after the institutional data transfer agreement.

Acknowledgments: The authors would like to acknowledge the Moffitt Cancer Center's CCSG Support award (NCI's P30-CA076292) and the services of the Bioinformatics and Biostatistics Shared Resources core, clinical staff, and unwavering trust of our patients, who consented to the use of their data for the research study. This work partially supported the research time for the study authors (Y.B. and F.L.L.) with funding from Kite Pharma<sup>®</sup>, a Gilead Company. The authors would like to acknowledge the data curation, radiology reads/processing, and scholarly discussion of the following individuals: Hong Lu, Jin Qi, Erin Dean, Salvatore Corallo, Zhouping Wei, and Abdalah Mahmoud, all of whom have contributed directly or indirectly to the presented work. We would like to recognize the unwavering motivational support from (late) Gillies, former chair of Cancer Physiology at H Lee Moffitt Cancer Center, an NCI-designated Comprehensive Cancer Center. F.L.L.: This work was also supported by the National Cancer Institute (P30CA076292, PI Cleveland; and R01CA244328-01,

MPI Jim, Locke, Small), and generous donations from the Hyer family, Thiel family, and Hano family. F.L.L. is supported in part by the Leukemia and Lymphoma Society as a Clinical Scholar.

**Conflicts of Interest:** Y.B. and F.L.L. received partial coverage through a research grant from Kite Pharma<sup>®</sup>, to support research time for the work. There are no conflicts to report that are related to this work. F.L.L.: Scientific Advisory Role/Consulting Fees: A2, Adaptive Biotechnologies, Adaptimmune, Allogene, Amgen, Astra-Zeneca, Bluebird Bio, BMS, Calibr, Caribou, EcoR1, Gerson Lehrman Group (GLG), Iovance, Kite Pharma, Janssen, Legend Biotech, Miltenyi, Novartis, Sana, Pfizer, Poseida. Data Safety Monitoring Board: Data and Safety Monitoring Board for the NCI Safety Oversight CAR T-cell Therapies Committee. Research Contracts or Grants to my Institution for Service: 2SeventyBio (Institutional), Allogene (Institutional), BMS (Institutional), Incyte (Institutional), Kite Pharma (Institutional), Leukemia and Lymphoma Society Scholar in Clinical Research (PI: Locke), Mark Foundation, National Cancer Institute, Patents, Royalties, Other Intellectual Property: Several patents held by the institution in my name (unlicensed) in the field of cellular immunotherapy. Education or Editorial Activity: Aptitude Health, ASH, ASTCT, Clinical Care Options Oncology, Society for Immunotherapy of Cancer. J.M.: Consultancy/Advisory for Kite/Gilead, Novartis, and J&J. Research Funding from Kite/Gilead, Incyte, Lilly.

# Abbreviations

The following abbreviations are used in this manuscript:

PC	Principal components
СТ	Computed tomography
PET	Positron emission tomography
CAR-T	Chimeric antigen receptor T-cell therapy
Axi-cel	Axicabtagene ciloleucel
LDH	Lactate Dehydrogenase
ECOG	Eastern Cooperative Oncology Group
MTV	Metabolic tumor volume

# References

- Kulkarni, N.M.; Pinho, D.F.; Narayanan, S.; Kambadakone, A.R.; Abramson, J.S.; Sahani, D.V. Imaging for Oncologic Response Assessment in Lymphoma. *Am. J. Roentgenol.* 2016, 208, 18–31. [CrossRef] [PubMed]
- Paquin, A.R.; Oyogoa, E.; McMurry, H.S.; Kartika, T.; West, M.; Shatzel, J.J. The diagnosis and management of suspected lymphoma in general practice. *Eur. J. Haematol.* 2023, 110, 3–13. [CrossRef] [PubMed]
- A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. The Non-Hodgkin's Lymphoma Classification Project. *Blood* 1997, 89, 3909–3918.
- 4. Metzger, M.L.; Mauz-Körholz, C. Epidemiology, outcome, targeted agents and immunotherapy in adolescent and young adult non-Hodgkin and Hodgkin lymphoma. *Br. J. Haematol.* **2019**, *185*, 1142–1157. [CrossRef]
- 5. Susanibar-Adaniya, S.; Barta, S.K. 2021 Update on Diffuse large B cell lymphoma: A review of current data and potential applications on risk stratification and management. *Am. J. Hematol.* **2021**, *96*, 617–629. [CrossRef]
- Locke, F.L.; Neelapu, S.S.; Bartlett, N.L.; Siddiqi, T.; Chavez, J.C.; Hosing, C.M.; Ghobadi, A.; Budde, L.E.; Bot, A.; Rossi, J.M.; et al. Phase 1 Results of ZUMA-1: A Multicenter Study of KTE-C19 Anti-CD19 CAR T Cell Therapy in Refractory Aggressive Lymphoma. *Mol. Ther.* 2017, 25, 285–295. [CrossRef]
- Neelapu, S.S.; Locke, F.L.; Bartlett, N.L.; Lekakis, L.J.; Miklos, D.B.; Jacobson, C.A.; Braunschweig, I.; Oluwole, O.O.; Siddiqi, T.; Lin, Y.; et al. Axicabtagene Ciloleucel CAR T-Cell Therapy in Refractory Large B-Cell Lymphoma. N. Engl. J. Med. 2017, 377, 2531–2544. [CrossRef]
- Brudno, J.N.; Kochenderfer, J.N. Recent advances in CAR T-cell toxicity: Mechanisms, manifestations and management. *Blood Rev.* 2019, 34, 45–55. [CrossRef]
- 9. Maurer, M.J. The International Prognostic Index in aggressive B-cell lymphoma. Haematologica 2023, 108, 2874–2879. [CrossRef]
- 10. International Non-Hodgkin's Lymphoma Prognostic Factors Project. A predictive model for aggressive non-Hodgkin's lymphoma. *N. Engl. J. Med.* **1993**, *329*, 987–994. [CrossRef]
- Sehn, L.H.; Berry, B.; Chhanabhai, M.; Fitzgerald, C.; Gill, K.; Hoskins, P.; Klasa, R.; Savage, K.J.; Shenkier, T.; Sutherland, J.; et al. The revised International Prognostic Index (R-IPI) is a better predictor of outcome than the standard IPI for patients with diffuse large B-cell lymphoma treated with R-CHOP. *Blood* 2007, *109*, 1857–1861. [CrossRef] [PubMed]

- 12. Cheson, B.D.; Fisher, R.I.; Barrington, S.F.; Cavalli, F.; Schwartz, L.H.; Zucca, E.; Lister, T.A. Recommendations for initial evaluation, staging, and response assessment of Hodgkin and non-Hodgkin lymphoma: The Lugano classification. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* **2014**, *32*, 3059–3068. [CrossRef] [PubMed]
- Johnson, S.A.; Kumar, A.; Matasar, M.J.; Schöder, H.; Rademaker, J. Imaging for Staging and Response Assessment in Lymphoma. *Radiology* 2015, 276, 323–338. [CrossRef]
- 14. Dean, E.A.; Mhaskar, R.S.; Lu, H.; Mousa, M.S.; Krivenko, G.S.; Lazaryan, A.; Bachmeier, C.A.; Chavez, J.C.; Nishihori, T.; Davila, M.L.; et al. High metabolic tumor volume is associated with decreased efficacy of axicabtagene ciloleucel in large B-cell lymphoma. *Blood Adv.* **2020**, *4*, 3268–3276. [CrossRef]
- 15. Vercellino, L.; Cottereau, A.S.; Casasnovas, O.; Tilly, H.; Feugier, P.; Chartier, L.; Fruchart, C.; Roulin, L.; Oberic, L.; Pica, G.M.; et al. High total metabolic tumor volume at baseline predicts survival independent of response to therapy. *Blood* **2020**, *135*, 1396–1405. [CrossRef] [PubMed]
- Voorhees, T.J.; Zhao, B.; Oldan, J.; Hucks, G.; Khandani, A.; Dittus, C.; Smith, J.; Morrison, J.K.; Cheng, C.J.; Ivanova, A.; et al. Pretherapy metabolic tumor volume is associated with response to CD30 CAR T cells in Hodgkin lymphoma. *Blood Adv.* 2022, 6, 1255–1263. [CrossRef]
- 17. Caimi, P.F.; Hill, B.T.; Hsi, E.D.; Smith, M.R. Clinical approach to diffuse large B cell lymphoma. *Blood Rev.* **2016**, *30*, 477–491. [CrossRef]
- Aerts, H.J.W.L.; Velazquez, E.R.; Leijenaar, R.T.H.; Parmar, C.; Grossmann, P.; Carvalho, S.; Bussink, J.; Monshouwer, R.; Haibe-Kains, B.; Rietveld, D.; et al. Decoding tumour phenotype by noninvasive imaging using a quantitative radiomics approach. *Nat. Commun.* 2014, *5*, 4006. [CrossRef]
- 19. Grossmann, P.; Stringfield, O.; El-Hachem, N.; Bui, M.M.; Rios Velazquez, E.; Parmar, C.; Leijenaar, R.T.; Haibe-Kains, B.; Lambin, P.; Gillies, R.J.; et al. Defining the biological basis of radiomic phenotypes in lung cancer. *Elife* **2017**, *6*, e23421. [CrossRef]
- 20. Kumar, V.; Gu, Y.; Basu, S.; Berglund, A.; Eschrich, S.A.; Schabath, M.B.; Forster, K.; Aerts, H.J.; Dekker, A.; Fenstermacher, D.; et al. Radiomics: The process and the challenges. *Magn. Reson. Imaging* **2012**, *30*, 1234–1248. [CrossRef]
- Ligero, M.; Simó, M.; Carpio, C.; Iacoboni, G.; Balaguer-Montero, M.; Navarro, V.; Sánchez-Salinas, M.A.; Bobillo, S.; Marín-Niebla, A.; Iraola-Truchuelo, J.; et al. PET-based radiomics signature can predict durable responses to CAR T-cell therapy in patients with large B-cell lymphoma. *eJHaem* 2023, 4, 1081–1088. [CrossRef]
- Balagurunathan, Y.; Wei, Z.; Qi, J.; Thompson, Z.; Dean, E.; Lu, H.; Vardhanabhuti, S.; Corallo, S.; Choi, J.W.; Kim, J.J.; et al. Radiomic Features on PET/CT Imaging of Large B cell Lymphoma Lesions Predicts CAR T-cell Therapy Efficacy. *Front. Oncol. Sect. Hematol. Malig.* 2024, 14, 1485039. [CrossRef]
- O, J.H.; Lodge, M.A.; Wahl, R.L. Practical PERCIST: A Simplified Guide to PET Response Criteria in Solid Tumors 1.0. *Radiology* 2016, 280, 576–584. [CrossRef] [PubMed]
- 24. Moghbel, M.C.; Kostakoglu, L.; Zukotynski, K.; Chen, D.L.; Nadel, H.; Niederkohr, R.; Mittra, E. Response Assessment Criteria and Their Applications in Lymphoma: Part 1. *J. Nucl. Med.* **2016**, *57*, 928–935. [CrossRef]
- Balagurunathan, Y.; Gu, Y.; Wang, H.; Kumar, V.; Grove, O.; Hawkins, S.; Kim, J.; Goldgof, D.B.; Hall, L.O.; Gatenby, R.A.; et al. Reproducibility and Prognosis of Quantitative Features Extracted from CT Images. *Transl. Oncol.* 2014, 7, 72–87. [CrossRef] [PubMed]
- 26. Balagurunathan, Y.; Kumar, V.; Gu, Y.; Kim, J.; Wang, H.; Liu, Y.; Goldgof, D.B.; Hall, L.O.; Korn, R.; Zhao, B.; et al. Test-retest reproducibility analysis of lung CT image features. *J. Digit. Imaging* **2014**, *27*, 805–823. [CrossRef] [PubMed]
- 27. Zwanenburg, A.; Vallières, M.; Abdalah, M.A.; Aerts, H.; Andrearczyk, V.; Apte, A.; Ashrafinia, S.; Bakas, S.; Beukinga, R.J.; Boellaard, R.; et al. The Image Biomarker Standardization Initiative: Standardized Quantitative Radiomics for High-Throughput Image-based Phenotyping. *Radiology* 2020, 295, 328–338. [CrossRef]
- 28. Bland, J.M.; Altman, D.G. The logrank test. BMJ Clin. Res. Ed. 2004, 328, 1073. [CrossRef]
- 29. Cox, D.R. Regression models and life-tables. J. R. Stat. Soc. Ser. B Methodol. 1972, 34, 187–220. [CrossRef]
- 30. Uno, H.; Cai, T.; Pencina, M.J.; D'Agostino, R.B.; Wei, L.J. On the C-statistics for evaluating overall adequacy of risk prediction procedures with censored survival data. *Stat. Med.* **2011**, *30*, 1105–1117. [CrossRef]
- 31. Storey, J.D.; Tibshirani, R. Statistical significance for genomewide studies. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 9440–9445. [CrossRef] [PubMed]
- 32. Campo, E.; Swerdlow, S.H.; Harris, N.L.; Pileri, S.; Stein, H.; Jaffe, E.S. The 2008 WHO classification of lymphoid neoplasms and beyond: Evolving concepts and practical applications. *Blood* **2011**, *117*, 5019–5032. [CrossRef]
- 33. Arber, D.A.; Orazi, A.; Hasserjian, R.; Thiele, J.; Borowitz, M.J.; Le Beau, M.M.; Bloomfield, C.D.; Cazzola, M.; Vardiman, J.W. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood* 2016, 127, 2391–2405. [CrossRef] [PubMed]
- Leithner, D.; Flynn, J.R.; Devlin, S.M.; Mauguen, A.; Fei, T.; Zeng, S.; Zheng, J.; Imber, B.S.; Hubbeling, H.; Mayerhoefer, M.E.; et al. Conventional and novel [(18)F]FDG PET/CT features as predictors of CAR-T cell therapy outcome in large B-cell lymphoma. *J. Hematol. Oncol.* 2024, 17, 21. [CrossRef]

- Feeney, J.; Horwitz, S.; Gönen, M.; Schöder, H. Characterization of T-cell lymphomas by FDG PET/CT. *AJR. Am. J. Roentgenol.* 2010, 195, 333–340. [CrossRef]
- Wang, S.; Chen, A.; Yang, L.; Cai, L.; Xie, Y.; Fujimoto, J.; Gazdar, A.; Xiao, G. Comprehensive analysis of lung cancer pathology images to discover tumor shape and boundary features that predict survival outcome. *Sci. Rep.* 2018, *8*, 10393. [CrossRef]
- 37. Grove, O.; Berglund, A.E.; Schabath, M.B.; Aerts, H.J.; Dekker, A.; Wang, H.; Velazquez, E.R.; Lambin, P.; Gu, Y.; Balagurunathan, Y.; et al. Quantitative computed tomographic descriptors associate tumor shape complexity and intratumor heterogeneity with prognosis in lung adenocarcinoma. *PLoS ONE* **2015**, *10*, e0118261. [CrossRef] [PubMed]
- 38. Palumbo, B.; Bianconi, F.; Palumbo, I.; Fravolini, M.L.; Minestrini, M.; Nuvoli, S.; Stazza, M.L.; Rondini, M.; Spanu, A. Value of Shape and Texture Features from 18F-FDG PET/CT to Discriminate between Benign and Malignant Solitary Pulmonary Nodules: An Experimental Evaluation. *Diagnostics* 2020, 10, 696. [CrossRef]
- 39. Cheson, B.D.; Pfistner, B.; Juweid, M.E.; Gascoyne, R.D.; Specht, L.; Horning, S.J.; Coiffier, B.; Fisher, R.I.; Hagenbeek, A.; Zucca, E.; et al. Revised response criteria for malignant lymphoma. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* 2007, *25*, 579–586. [CrossRef]
- 40. Ather, S.; Kadir, T.; Gleeson, F. Artificial intelligence and radiomics in pulmonary nodule management: Current status and future applications. *Clin. Radiol.* **2020**, *75*, 13–19. [CrossRef]
- 41. Balagurunathan, Y.; Schabath, M.B.; Wang, H.; Liu, Y.; Gillies, R.J. Quantitative Imaging features Improve Discrimination of Malignancy in Pulmonary nodules. *Sci. Rep.* **2019**, *9*, 8528. [CrossRef]
- 42. Napel, S.; Giger, M. Special Section Guest Editorial:Radiomics and Imaging Genomics: Quantitative Imaging for Precision Medicine. *J. Med. Imaging* 2015, 2, 041001. [CrossRef] [PubMed]
- Zhou, M.; Scott, J.; Chaudhury, B.; Hall, L.; Goldgof, D.; Yeom, K.W.; Iv, M.; Ou, Y.; Kalpathy-Cramer, J.; Napel, S.; et al. Radiomics in Brain Tumor: Image Assessment, Quantitative Feature Descriptors, and Machine-Learning Approaches. *AJNR Am. J. Neuroradiol.* 2018, *39*, 208–216. [CrossRef]
- 44. Levstek, L.; Janžič, L.; Ihan, A.; Kopitar, A.N. Biomarkers for prediction of CAR T therapy outcomes: Current and future perspectives. *Front. Immunol.* **2024**, *15*, 1378944. [CrossRef] [PubMed]
- 45. Cottereau, A.S.; Meignan, M.; Nioche, C.; Capobianco, N.; Clerc, J.; Chartier, L.; Vercellino, L.; Casasnovas, O.; Thieblemont, C.; Buvat, I. Risk stratification in diffuse large B-cell lymphoma using lesion dissemination and metabolic tumor burden calculated from baseline PET/CT(†). *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* **2021**, *32*, 404–411. [CrossRef]
- Hwang, S.H.; Jung, M.; Jeong, Y.H.; Jo, K.; Kim, S.; Wang, J.; Cho, A. Prognostic value of metabolic tumor volume and total lesion glycolysis on preoperative (18)F-FDG PET/CT in patients with localized primary gastrointestinal stromal tumors. *Cancer Metab.* 2021, *9*, 8. [CrossRef]
- Ilyas, H.; Mikhaeel, N.G.; Dunn, J.T.; Rahman, F.; Møller, H.; Smith, D.; Barrington, S.F. Defining the optimal method for measuring baseline metabolic tumour volume in diffuse large B cell lymphoma. *Eur. J. Nucl. Med. Mol. Imaging* 2018, 45, 1142–1154. [CrossRef]
- Bobillo, S.; Joffe, E.; Lavery, J.A.; Sermer, D.; Ghione, P.; Noy, A.; Caron, P.C.; Hamilton, A.; Hamlin, P.A.; Horwitz, S.M.; et al. Clinical characteristics and outcomes of extranodal stage I diffuse large B-cell lymphoma in the rituximab era. *Blood* 2021, 137, 39–48. [CrossRef]
- 49. Beyar Katz, O.; Perry, C.; Grisariu-Greenzaid, S.; Yehudai-Ofir, D.; Luttwak, E.; Avni, B.; Zuckerman, T.; Sdayoor, I.; Stepensky, P.; Ringelstein-Harlev, S.; et al. Response rates of extra-nodal diffuse large B cell lymphoma to anti-CD19-CAR T cells: A real word retrospective multicenter study. *Eur. J. Haematol.* **2023**, *111*, 63–71. [CrossRef]
- 50. Raskin, W.; Harle, I.; Hopman, W.M.; Booth, C.M. Prognosis, Treatment Benefit and Goals of Care: What do Oncologists Discuss with Patients who have Incurable Cancer? *Clin. Oncol. R. Coll. Radiol.* **2016**, *28*, 209–214. [CrossRef] [PubMed]
- 51. Ethier, J.L.; Paramsothy, T.; You, J.J.; Fowler, R.; Gandhi, S. Perceived Barriers to Goals of Care Discussions With Patients With Advanced Cancer and Their Families in the Ambulatory Setting: A Multicenter Survey of Oncologists. *J. Palliat. Care* **2018**, *33*, 125–142. [CrossRef] [PubMed]
- Jalaguier-Coudray, A.; Thomassin-Piana, J. Solid masses: What are the underlying histopathological lesions? *Diagn. Interv. Imaging* 2014, 95, 153–168. [CrossRef] [PubMed]
- Zhang, J.; Li, Y.; Zhao, Y.; Qiao, J. CT and MRI of superficial solid tumors. *Quant. Imaging Med. Surg.* 2018, 8, 232–251. [CrossRef] [PubMed]
- 54. Senjo, H.; Hirata, K.; Izumiyama, K.; Minauchi, K.; Tsukamoto, E.; Itoh, K.; Kanaya, M.; Mori, A.; Ota, S.; Hashimoto, D.; et al. High metabolic heterogeneity on baseline 18FDG-PET/CT scan as a poor prognostic factor for newly diagnosed diffuse large B-cell lymphoma. *Blood Adv.* 2020, *4*, 2286–2296. [CrossRef]
- 55. Stella, F.; Chiappella, A.; Casadei, B.; Bramanti, S.; Ljevar, S.; Chiusolo, P.; Di Rocco, A.; Tisi, M.C.; Carrabba, M.G.; Cutini, I.; et al. A Multicenter Real-life Prospective Study of Axicabtagene Ciloleucel versus Tisagenlecleucel Toxicity and Outcomes in Large B-cell Lymphomas. *Blood Cancer Discov.* 2024, *5*, 318–330. [CrossRef]
- 56. Zhang, X.; Yang, X.; Wang, C.; Huang, L.; Zhang, Y.; Wei, J. High Expression of Plasma IL-1β Levels and Transition of Regulatory T-Cell Subsets Correlate with Disease Progression in Myelodysplastic Syndrome. *Blood* **2022**, *140*, 9761–9762. [CrossRef]

- 57. Peng, J.; Yang, X.; Li, C.; Zhang, Y.; Ma, L.; Wei, J. High Serum ST2 Level Predicts Progression and Poor Prognosis of Denovomyelodysplastic Syndrome. *Blood* 2023, 142, 6455. [CrossRef]
- 58. Gong, Y.; Fei, P.; Zhang, Y.; Xu, Y.; Wei, J. From Multi-Omics to Visualization and Beyond: Bridging Micro and Macro Insights in CAR-T Cell Therapy. *Adv. Sci.* 2025, e2501095. [CrossRef]
- 59. Luciani, F.; Safavi, A.; Guruprasad, P.; Chen, L.; Ruella, M. Advancing CAR T-cell Therapies with Artificial Intelligence: Opportunities and Challenges. *Blood Cancer Discov.* **2025**, *6*, 159–162. [CrossRef]
- Shahzadi, M.; Rafique, H.; Waheed, A.; Naz, H.; Waheed, A.; Zokirova, F.R.; Khan, H. Artificial intelligence for chimeric antigen receptor-based therapies: A comprehensive review of current applications and future perspectives. *Ther. Adv. Vaccines Immunother.* 2024, 12. [CrossRef]
- 61. Balagurunathan, Y.; Mitchell, R.; El Naqa, I. Requirements and reliability of AI in the medical context. *Phys. Medica* 2021, *83*, 72–78. [CrossRef] [PubMed]
- 62. Afroogh, S.; Akbari, A.; Malone, E.; Kargar, M.; Alambeigi, H. Trust in AI: Progress, challenges, and future directions. *Humanit. Social. Sci. Commun.* **2024**, *11*, 1568. [CrossRef]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.