

DATA REPORT

Open Access

Novel *SLC20A2* variant in a Japanese patient with idiopathic basal ganglia calcification-1 (IBGC1) associated with dopa-responsive parkinsonism

Yaeko Ichikawa¹, Masaki Tanaka^{2,3}, Eriko Kurita¹, Masanori Nakajima¹, Masaki Tanaka¹, Chizuko Oishi¹, Jun Goto⁴, Shoji Tsuji^{1,2,3,5} and Atsuro Chiba¹

Abstract

Idiopathic basal ganglia calcification-1 (IBGC1) is an autosomal dominant disorder characterized by calcification in the basal ganglia, which can manifest a range of neuropsychiatric symptoms, including parkinsonism. We herein describe a 64-year-old Japanese IBGC1 patient with bilateral basal ganglia calcification carrying a novel *SLC20A2* variant (p.Val322Glufs*92). The patient also presented with dopa-responsive parkinsonism with decreased dopamine transporter (DAT) density in the bilateral striatum and decreased cardiac ¹²³I-meta-iodobenzylguanidine uptake.

Idiopathic basal ganglia calcification (IBGC), also known as Fahr disease or primary familial brain calcification (PFBC), is a disorder characterized by bilateral calcifications in the basal ganglia and other brain regions. Clinical manifestations of IBGC range from asymptomatic to neuropsychiatric symptoms, including dystonia, parkinsonism, ataxia, and cognitive impairment¹. Typically, the inheritance mode of familial IBGC is an autosomal dominant one and to date, four dominant causal genes of familial IBGC have been identified, including *SLC20A2* (IBGC1, MIM: #213600), *PDGFRB* (IBGC4, MIM: #615007), *PDGFB* (IBGC5, MIM: #615483), and *XPR1* (IBGC6, MIM: #616413)^{2–5}. Recently, *MYORG* was reported as an autosomal recessive causal gene for IBGC (IBGC7, MIM: #618317)^{6,7}. Variants in *SLC20A2*, encoding the type III sodium-dependent phosphate transporter 2 (PiT-2), are a major cause of IBGC^{8,9}. Herein, we report an IBGC1 patient with a novel variant in *SLC20A2* associated with dopa-responsive parkinsonism.

The patient was a 63-year-old Japanese woman who presented to our hospital with a one-month history of lumbago

and unsteady gait. Neurological examination revealed gait disturbance with stooped posture and short steps, but rigidity, tremor, weakness, and cerebellar symptoms were not observed. Computed tomography (CT) images of her brain revealed marked calcifications in the bilateral basal ganglia, thalami, and dentate nuclei (Fig. 1a). Laboratory tests showed that serum calcium, phosphate, and intact parathyroid hormone levels were all within the normal ranges. There was no family history of IBGC or parkinsonism. After written informed consent was obtained, we analyzed all the coding regions of the IBGC causative genes, *SLC20A2*, *PDGFRB*, and *PDGFB*, by Sanger sequencing as previously reported¹⁰. We diagnosed her as IBGC1 based on the identification of a novel heterozygous frameshift variant, p.Val322Glufs*92 (NM_006749.4:c.965_966delTG, exon 8), in *SLC20A2* (Fig. 1b). The variant was absent in the following genome databases: dbSNP 151 (<https://www.ncbi.nlm.nih.gov/projects/SNP/>), Integrative Japanese Genome Variation Database (<http://ijgvd.megabank.tohoku.ac.jp/>), Exome Aggregation Consortium database version 0.3.1 (<http://exac.broadinstitute.org/>), and Human Gene Mutation Database (HGMD® Professional 2019.1).

Ten months after her first visit, she was hospitalized because of difficulties in standing up without assistance at the age of 64. She showed severe bradykinesia, postural

Correspondence: Yaeko Ichikawa (yaeko-ty@umin.ac.jp)

¹Department of Neurology, Kyorin University School of Medicine, Tokyo, Japan

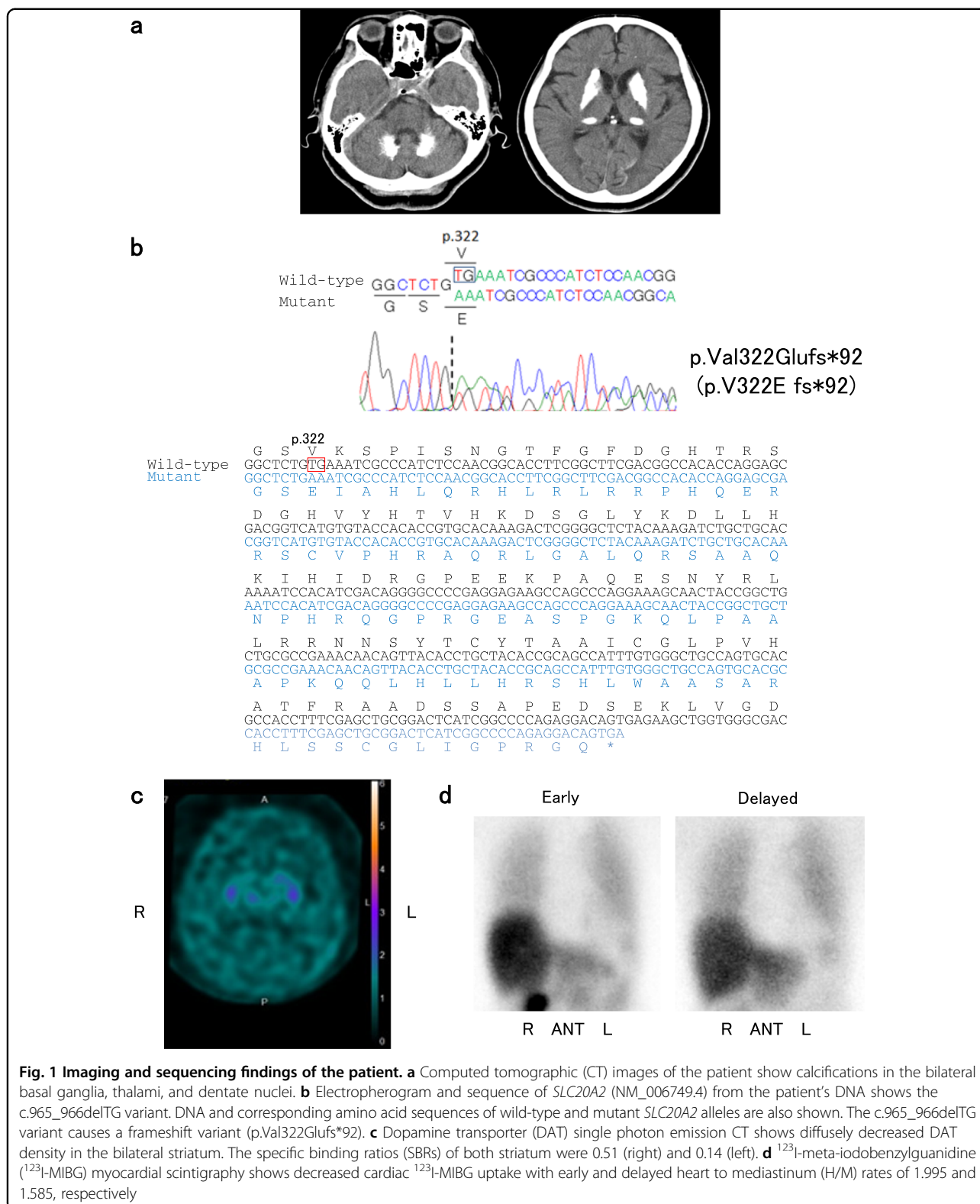
²Department of Neurology, The University of Tokyo Hospital, Tokyo, Japan

Full list of author information is available at the end of the article.

© The Author(s) 2019



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.



instability, and mild symmetric rigidity without tremor. Her Unified Parkinson Disease Rating Scale part III (UPDRS-III) score was 43 of 108 on the ninth hospital day. Her Mini-

Mental State Examination score was 24 of 30, and her Hasegawa dementia scale revised was 22 of 30. Dopamine transporter (DAT) single photon emission CT using

Table 1 Variants of *SLC20A2* and clinical features of genetically confirmed IBGC1 Japanese patients with parkinsonism

Case	1	2	3	4	5	6	7	8	9
Variant	c.212G>A R71H Exon 2	c.269G>T G90V Exon 2	G90V	c.516+1G>A	V144Gfs*85	IVS 4	c.965_966delTG V322Efs*92 Exon 8	c.1909A>C S637R Exon 11	
Patient	Proband	Proband	Son	Mother	Proband	Son	Proband	Brother	Proband
Age/sex	73/F	79/M	52/M	89/F	62/M	27/M	64/F	NA/M	62/M
Age at onset (years)	71	74	50	NA	60		63	NA	62
Onset symptom	Clumsiness of hands and unsteady gait	Dementia	Depression	NA	Slowness and gait disturbance	Asymptomatic	Unsteady gait	NA	Difficulty in driving a car
Parkinsonism	(+)	(+)	None	(+)	(+)	None	(+)	(+)	(+)
Levodopa responsiveness	(+)	NA		NA	(+)		(+)	NA	NA
Cognitive impairment	(+)	(+)	None	NA	None	None	Mild	NA	(+)
MMSE	16/30	13/30	30/30	NA	30/30	NA	24/30	NA	NA
HDS-R	NA	NA	NA	NA	NA	NA	22/30	NA	14/30
FAB	NA	3/18	NA	NA	NA	NA	Not examined	NA	NA
DAT SPECT	NA	Decreased	Normal	NA	Decreased	NA	Decreased	NA	NA
MIBG scintigraphy	NA	Decreased	Normal	NA	Decreased	NA	Decreased	NA	NA
Early H/M	NA	1.62	3.24	NA	1.43	NA	1.995	NA	NA
Delayed H/M	NA	NA	NA	NA	NA	NA	1.585	NA	NA
Autopsy	(+)	NA	NA	NA	NA	NA	(-)	NA	(+)
Lewy bodies	(+)								(+)
Reference	Yamada et al. ¹⁰	Koyama et al. ¹⁴		Koyama et al. ¹⁴			This report	Kimura et al. ¹³	

F female, M male, NA not applicable, MMSE Mini-Mental State Examination, HDS-R Hasegawa dementia scale revised, FAB frontal assessment battery, DAT SPECT dopamine transporter single photon emission CT, MIBG scintigraphy ¹²³I-meta-iodobenzylguanidine myocardial scintigraphy

¹²³I-ioflupane showed diffusely decreased DAT density in the bilateral striatum (Fig. 1c). The specific binding ratios (SBRs) of both striatum were 0.51 (right) and 0.14 (left). Her ¹²³I-meta-iodobenzylguanidine (¹²³I-MIBG) myocardial scintigraphy revealed reduced cardiac ¹²³I-MIBG uptake with early and delayed heart to mediastinum (H/M) rates of 1.995 and 1.585, respectively (Fig. 1d). Levodopa therapy (200 mg/day) was started on the 14th hospital day and was effective against bradykinesia and postural instability. She was able to walk without assistance in her room. On the 122nd hospital day, she received 600 mg/day of levodopa, and her UPDRS-III score markedly improved from 43 to 11.

The variants associated with IBGC are located widely in *SLC20A2* among the patients with IBGC, and the correlation of genotype and phenotype remains unclear.^{1,9,11}

Parkinsonism is one of the common clinical symptoms of IBGC. Tadic et al. showed that 13% of patients with *SLC20A2* or *PDGFRB* variants presented with

parkinsonism¹. Another review reported motor improvement with dopatherapy in five patients with genetically confirmed IBGC¹². Genetically confirmed Japanese IBGC1 patients presenting with parkinsonism have also been reported (Table 1)^{10,13,14}. Among the five variants summarized in Table 1, two variants (c.516+1G>A and c.965_966delTG) are frameshift variants, presumably resulting in loss of function of *SLC20A2*. In addition, a decreased level of *SLC20A2* protein was described in the case with the missense variant (c.1909A>C, S637R), raising the possibility of unstable mutant protein¹³. Although the functional investigations were not reported for the two missense variants (R71H and G90V), loss-of-function variants are considered for the three variants shown in Table 1. Consistent with previous reports, the majority of variants associated with IBGC are loss-of-function variants^{8,9}, and the present study also suggests that loss-of-function mechanisms are likely involved in at least of the

three variants. The present case demonstrated decreased DAT density in the bilateral striatum and decreased cardiac ^{123}I -MIBG uptake (Fig. 1c, d). The decreased DAT density in the bilateral striatum suggested presynaptic dopaminergic dysfunction, which was reported in patients with IBGC^{14–17}. Saito et al. also showed that postsynaptic dopaminergic dysfunction in the bilateral striatum matched calcified regions¹⁶. These findings suggested that basal ganglia calcification might result in dopaminergic dysfunction in IBGC patients. The three cases with reduced DAT density in the striatum (cases 2, 5, and 7. Table 1) also presented with decreased cardiac ^{123}I -MIBG uptake, which was indistinguishable from that observed in patients with Lewy body diseases, including idiopathic Parkinson disease (PD)¹⁸. Since PD is a relatively common disease in Japan (prevalence of ~150 per 100,000 persons in Japan)¹⁹, the coincidental presence of idiopathic PD and IBGC remains a possibility concerning dopa-responsive parkinsonism of patients with IBGC1. However, it is important to pay attention to patients with IBGC who show dopa-responsive parkinsonism to provide appropriate treatment. To clarify the etiologies of dopa-responsive parkinsonism occasionally observed in patients with IBGC, further functional analyses including DAT SPECT and ^{123}I -MIBG myocardial scintigraphy will be required in a larger number of patients with genetically confirmed IBGC.

HGV Database

The relevant data from this Data Report are hosted at the Human Genome Variation Database at <https://doi.org/10.6084/m9.figshare.hgv.2603>

Acknowledgements

This work was supported by Grant-in-Aid (No. H26-Jitsuyoka [Nanbyo]-Ippan-080) from the Ministry of Health, Labour and Welfare, Japan (S.T.).

Author details

¹Department of Neurology, Kyorin University School of Medicine, Tokyo, Japan. ²Department of Neurology, The University of Tokyo Hospital, Tokyo, Japan. ³Institute of Medical Genomics, International University of Health and Welfare, Chiba, Japan. ⁴Department of Neurology, International University of Health and Welfare Mita Hospital, Tokyo, Japan. ⁵Department of Molecular Neurology, Graduate School of Medicine, The University of Tokyo, Tokyo, Japan

Conflict of interest

The authors declare that they have no conflict of interest.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 1 June 2019 Revised: 18 July 2019 Accepted: 25 July 2019.
Published online: 4 September 2019

References

- Tadic, V. et al. Primary familial brain calcification with known gene mutations: a systematic review and challenges of phenotypic characterization. *JAMA Neurol.* **72**, 460–467 (2015).
- Wang, C. et al. Mutations in SLC20A2 link familial idiopathic basal ganglia calcification with phosphate homeostasis. *Nat. Genet.* **44**, 254–256 (2012).
- Keller, A. et al. Mutations in the gene encoding PDGF-B cause brain calcifications in humans and mice. *Nat. Genet.* **45**, 1077–1082 (2013).
- Nicolas, G. et al. Mutation of the PDGFRB gene as a cause of idiopathic basal ganglia calcification. *Neurology* **80**, 181–187 (2013).
- Legati, A. et al. Mutations in XPR1 cause primary familial brain calcification associated with altered phosphate export. *Nat. Genet.* **47**, 579–581 (2015).
- Yao, X. P. et al. Biallelic mutations in MYORG cause autosomal recessive primary familial brain calcification. *Neuron* **98**, 1116–1123 (2018). e5.
- Arkadir, D. et al. MYORG is associated with recessive primary familial brain calcification. *Ann. Clin. Transl. Neurol.* **6**, 106–113 (2019).
- Hsu, S. C. et al. Mutations in SLC20A2 are a major cause of familial idiopathic basal ganglia calcification. *Neurogenetics* **14**, 11–22 (2013).
- Lemos, R. R. et al. Update and mutational analysis of SLC20A2: a major cause of primary familial brain calcification. *Hum. Mutat.* **36**, 489–495 (2015).
- Yamada, M. et al. Evaluation of SLC20A2 mutations that cause idiopathic basal ganglia calcification in Japan. *Neurology* **82**, 705–712 (2014).
- Ding, Y. & Dong, H. Q. A novel SLC20A2 mutation associated with familial idiopathic basal ganglia calcification and analysis of the genotype-phenotype association in Chinese patients. *Chin. Med J. (Engl.)* **131**, 799–803 (2018).
- Nicolas, G. et al. Phenotypic spectrum of probable and genetically-confirmed idiopathic basal ganglia calcification. *Brain* **136**, 3395–3407 (2013).
- Kimura, T. et al. Familial idiopathic basal ganglia calcification: Histopathologic features of an autopsied patient with an SLC20A2 mutation. *Neuropathology* **36**, 365–371 (2016).
- Koyama, S. et al. Clinical and radiological diversity in genetically confirmed primary familial brain calcification. *Sci. Rep.* **7**, 12046 (2017).
- Paschali, A. et al. Dopamine transporter SPECT/CT and perfusion brain SPECT imaging in idiopathic basal ganglia calcinosis. *Clin. Nucl. Med.* **34**, 421–423 (2009).
- Saito, T. et al. Neuroradiologic evidence of pre-synaptic and post-synaptic nigrostriatal dopaminergic dysfunction in idiopathic Basal Ganglia calcification: a case report. *J. Neuroimaging* **20**, 189–191 (2010).
- Paghera, B., Caobelli, F. & Giubbini, R. ^{123}I -ioflupane SPECT in Fahr disease. *J. Neuroimaging* **23**, 157–158 (2013).
- Orimo, S., Suzuki, M., Inaba, A. & Mizusawa, H. ^{123}I -MIBG myocardial scintigraphy for differentiating Parkinson's disease from other neurodegenerative parkinsonism: a systematic review and meta-analysis. *Park. Relat. Disord.* **18**, 494–500 (2012).
- Yamawaki, M., Kusumi, M., Kowa, H. & Nakashima, K. Changes in prevalence and incidence of Parkinson's disease in Japan during a quarter of a century. *Neuroepidemiology* **32**, 263–269 (2009).