



# **Circular RNAs as New Regulators in Gastric Cancer: Diagnosis and Cancer Therapy**

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Gastric cancer (GC) is one of the most commonly diagnosed cancers that causes high mortality in the world. Although the surgery tools and chemotherapies have significantly improved the overall survival of patients with GC, the early diagnosis of GC remains insufficient and many patients diagnosed with advanced stages of GC are not able to benefit from curative therapy. Circular RNAs (circRNAs), novel members of the non-coding cancer genome, are being explored with regards to various cancer types including GC. CircRNAs could work as miRNA sponges to regulate cell proliferation, cell migration, and cell cycle in GC. In addition, it was found that abnormal expression of circRNAs was associated with pathological characteristics in GC tissues, which could help to act as potential markers of early diagnosis or predictors of prognosis. Although various functional circRNAs have been discovered and characterized, the studies of circRNAs in GC are still at early stages compared with other RNAs. In order to provide a whole view to better understand the circRNAs in the occurrence and development of GC, we review the current knowledge on circRNAs in relation to their expression and regulation in GC as well as their potential to be diagnosis markers, and their role in drug resistance will be mentioned. It is helpful to address their possibility from basic research into practical application.

Keywords: gastric cancer, circRNA, chemotherapy, cell growth, cell cycle

# BACKGROUND

### **Gastric Cancer**

Gastric cancer (GC) is one of the most commonly diagnosed cancers. The mortality of GC accounts for 8.2% of all cancer death, being the third leading cause of cancer-related death in the world. Eastern Asia is well known to be a high incidence area of GC, the incident rates are markedly elevated especially in Mongolia, Japan, and the South Korea (1). Anatomically, GC could be classified into true gastric (non-cardia) and gastro-oesophageal-junction cancers (cardia). Through genomic and statistical analysis GC could be classified into four subgroups: Epstein-Barr virus positive tumors (9%), microsatellite unstable tumors (22%), genomically stable tumors (20%), and chromosomally unstable tumors (50%), which might be used in the determination of prognosis and treatment (2). The important causes of GC include Helicobacter pylori infection and Epstein-Barr virus (EBV) infection, and 10% of the cases are aggregated in families (3).

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Most patients with early stage GC cannot be diagnosed because it begins as asymptomatic, and, when they are diagnosed with symptoms of anorexia, dyspepsia, weight loss and abnormal pain, often it is at an advanced stage, with an overall survival (OS) rate of less than 30% (4). Despite the improvement of surgical and medical managements, it is still a big challenge to completely eliminate tumor cells via surgery, radio therapies and chemotherapies. In this scenario, it is essential to find new potential diagnosis biomarkers and therapeutic targets to improve the diagnosis and treatment of GC. In recent years, the functions of circRNAs in GC have been significantly evaluated since their discovery 40 years ago. In this review, the biogenetic process and biological functions of circRNAs, and their close relationship with GC are summarized, and the clinical significance of circRNAs with regulatory potency in the diagnosis and treatment of GC are emphasized.

#### CircRNAs

Non-coding RNAs (ncRNA) represent 60% of the total RNA by genome wide analysis. According to their functions, ncRNA can be classified into house-keeping ncRNAs and regulatory ncRNAs. House-keeping ncRNAs include, ribosomal RNA (rRNA), transfer RNAs (tRNA), small nuclear RNA (snRNA)and small nucleolar RNA (snoRNA), which participate in mRNA and rRNA processing. Regulatory ncRNAs are composed of microRNAs (miRNAs), siRNAs, piwi-interacting RNAs (piRNAs) and others (<200 bp) and long ncRNAs (lncRNAs) (>200 bp), which show their ability to control gene expression as epigenetic regulators. Among lncRNAs, circular RNAs (circRNAs) featured in high abundance, structural stability, and tissue- and developmentalspecific expression, which have recently been regarded as an emerging key player in regulating various physiological processes and pathogenic disease progress (5). Recent studies indicated that circRNAs are crucial in several types of diseases, which include neurological disorders (6), atherosclerosis (7, 8), osteoarthritis (9), diabetes (10) and cancer (11). Based on the increased evidences of the significance of circRNAs in human diseases, circRNAs may act as a diagnosis biomarker and a new therapeutic target in the near future.

Circular RNAs are defined as a distinct subclass of endogenous lncRNAs which form a locked continuous loop covalently with the 5' and 3' ends joined together, transcribed through RNA polymerase II as most lncRNAs are. Through canonical and non-canonical splicing, circRNAs can be categorized into three types: exonic circRNA (ecRNAs), originated from pre-mRNAs consisting of single or multiple exons; circular intronic RNAs (ciRNAs), derived from linear RNAs with one or more introns; and those composed of both exons and introns are named exonintron circRNAs (ElciRNAs) (Figure 1). CircRNAs are highly valued because of their distinctive roles in gene expression regulation and biological process (12). Regulatory roles in gene by circRNAs can be classified as follows: (1) CircRNAs can bind to the promoter of target genes as well as RNA polymerase II to regulate gene transcription; (2) CircRNAs can have different gene splicing patterns by sequestering RBPs that regulate mRNA processing; (3) CircRNAs can compete endogenous miRNA-binding sites by sponging miRNAs in target genes; and (4) CircRNAs can be translated into proteins. Although the functions and molecular mechanism of circRNAs remains largely unexplored, recent evidences suggested that circRNAs are aberrant expressed in several cancer types with tissue-and disease-specificity, making them a potential diagnosis biomarker in cancer (13). For instance, the expression of circRNAs can change in many cancers including, prostate cancer (14), bladder cancer (15), breast cancer (16) and glioma (17). A large number of researches also indicated that circRNAs evaluation in cancer patients may be helpful for prognosis prediction of certain cancers. For instance, compared with low circ-FBXW7 expression in glioblastoma patients, higher circ-FBXW7 expression exhibited higher overall survival (OS) (18). The downregulation of circHIPK3 in bladder cancer was closely related to increased invasion as well as lymph node metastasis (19). Hereinafter we emphasize the expression profiles of circRNAs and circRNAs-associated therapeutic potentials in GC.

# **CircRNAs and Gastric Cancer**

As significant attention has been received in recent years regarding the role of circRNAs in GC diagnosis and treatment, many papers report high-throughput sequencing or microarray results that describes the differential circRNAs expression profiles in GC patients and healthy donors (20, 21). These identified circRNAs are down-regulated or up-regulated during carcinogenesis, and therefore serve as tumor suppressors or oncogenes in GC, which make possible that circRNAs become diagnosis biomarkers and prognosis predictors. We searched recent published articles in key journals, from which the expressions of circRNAs and their clinical significances associated with human GC are summarized below (**Table 1** and **Figure 1**).

#### CircRNA and Gene Expression Profiles in GC

Several articles published in 2017 show us a preliminary expression profile of circRNAs in GC. Huang's group screened with circRNA chip using five pairs of matched GC and non-GC tissues, and it was revealed that 191 circRNAs were increased in GC tissues, whereas 522 were decreased, among which hsa\_circ\_0076304, hsa\_circ\_0076305, and hsa\_circ\_0035431 were mostly changed (43). Guo's group detected a total of 5396 circRNAs between GC tissues and adjacent nontumorous tissues, revealed 107 increased and 201 decreased circRNAs. Among which hsa circ 0035445, hsa circ 0003789, hsa\_circ\_0007099, hsa\_circ\_0001897, and hsa\_circ\_0007707 had the highest magnitude of change. Moreover, most differentially expressed circRNAs were located in chr1, chr2, chr3, chr9, and chr17 (44). Dai's group screened circRNA and mRNA profiles in the GC tissues and adjacent tissues and found 1,285 significant differently expressed circRNAs, with 594 decreased and 691 increased, and 69 among them might play regulatory roles in their target gene mRNAs expression. 5,460 differentially expressed genes were found in 29,112 genes with 2,390 genes increased and 3,070 genes decreased, such as MYH9, MALAT1 may involve in the occurrence and development of GC (45). A Brazil group verified 66 annotated circRNAs in Gastric tissue without GC, 620 in paired tumor-adjacent gastric tissue and 220 in GC samples, among them, hsa\_circ\_0001136, hsa\_circ\_0000284,



hsa\_circ\_0000211, and hsa\_circ\_0004771 were differentially expressed which may become novel biomarkers of GC (46). Guo's group screened the profiles in circRNAs expression between the plasma of GC patients and healthy donors and the circRNA levels were confirmed by both RT-qPCR and RT-droplet digital PCR. They found 343 differentially expressed circRNAs with hsa\_circ\_0088300 and hsa\_circ\_0075825 increased, hsa\_circ\_0001017 and hsa\_circ\_0061276 decreased in patients. The plasma hsa\_circ\_0001017 and hsa\_circ\_0061276 in patients after operation recovered to normal level indicating that these two circRNAs can be new ideal biomarkers for GC (47). In 2019, Zhang's group identified 196 increased and 311 decreased genes in GC tissues. In the meantime, they identified two decreased circRNAs hsa circ 0000332 and hsa circ 0021087 in GC tissues. It was also found that hsa\_circ\_0021087 may be more crucial in the GC network than hsa\_circ\_0000332 through prediction of target genes for circRNAs and miRNAs and construction of differentially expressed circRNA-miRNA-mRNA networks (48).

#### CircRNA Serves as MiRNA Sponges in GC

MicroRNAs are key players in the pathogenesis of almost all human cancer types (49, 50). Previously we mentioned that circRNAs can regulate gene expression by serving as miRNA sponges. CircRNAs have a miRNA response element that can compete for miRNA-binding sites in target genes. Although circRNAs regarding their potential molecular mechanisms in GC are largely unexplored, the function as regulators of miRNA activity suggested their involvement in GC. Recent papers drew some inspiration on the role of circRNAs acting as miRNA sponges in regulating GC cell proliferation, invasion and prognosis. CircPVT1 was increased in GC and promoted cell growth by sponging the miR-125 family (22). CircLARP4 was decreased and inhibited proliferation and invasion in GC cells by sponging miR-424 and upregulating LATS1 gene, whose expression is correlated with the pathological stage as well as unfavorable prognosis. Thus circLARP4 could act as a novel tumor suppressive factor in GC (25). CircRNA\_100269 was found decreased in GC cells and inhibited the cell proliferation. Furthermore, miR-630 was verified a directly interacting with circRNA\_100269 and could suppress its function in GC cells (26). CircYAP1 was decreased the overexpression of which inhibited cell proliferation as well as invasion in GC tissues by sponging miR-367-5p to inhibit p27 Kip1 expression and GC progression (27). Meltzer's group identified the overexpression of miR-21 in GC samples based on miR sequence data from 446 GC and 45 normal samples within The Cancer Genome Atlas (TCGA). Based on miR-21 they designed a synthetic circRNA containing five repeated miR-21 binding sites and revealed the role of the synthetic circRNA in suppressing GC growth by sponging miR-21 (51). Has\_circ\_0001461 was verified as decreased in GC tissues whose overexpression inhibited GC cell proliferation, migration and invasion by sponging miR-548g and upregulating RUNX1 expression in GC cells (31). CircRBMS3 was increased in GC tissues, and its expression was closely related to advanced TNM stage and lymph-node metastasis. CircRBMS3 could promote gastric

TABLE 1	Expression	of CircRNAs	associated v	with human	gastric cancers.
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CircRNAs	Gene symbol	Deregulation	References	Gene affected	Function affected
circPVT1	PVT1	Increased	(22)	miR-125	Cell growth
circLMTK2	LMTK2	Increased	(23)		Cell growth and prognosis
hsa_circ_0000096	HIAT1	Decreased	(24)	Cyclin D1, CDK6, MMP-2, MMP-9	Cell growth and migration
circLARP4	LARP4	Decreased	(25)	miR-424, LATS1	Cell growth and invasion
circRNA_100269	LPHN2	Decreased	(26)	miR-630	Cell growth
circYAP1	YAP1	Decreased	(27)	miR-367-5p	Cell growth and invasion
hsa_circ_0130810	KIAA1244	Decreased	(20)		Cell metastasis
hsa_circ_0000993	ATL2	Decreased	(28)	miR-214-5p	Cell growth and metastasis
circAGO2	AGO2	Increased	(29)		Cell growth, invasion, and metastasis
circ-DONSON	DONSON	Increased	(30)	SOX4	Cell growth and invasion
has_circ_0001461	FAT1	Decreased	(31)	miR-548g, RUNX1	Cell growth and metastasis
hsa_circ_0006156	FNDC3B	Increased	(32)	E-cadherin, CD44	Cell migration and invasion
hsa_circ_0000199	AKT3	Increased	(33)	miR-198, PIK3R1	Cell growth and apoptosis
hsa_circ_0064644	RBMS3	Increased	(34)	miR-153, SNAI1	Cell growth and invasion
circPDSS1	PDSS1	Increased	(35)	miR-186-5p, NEK2	Cell cycle and apoptosis
hsa_circ_0000234	CUL2	Decreased	(36)	mir-142-3p, VAMP3	Cell growth and metastasis
circPSMC3	PSMC3	Decreased	(37)	miR-296-5p	Cell growth and migration
circNF1	NF1	Increased	(38)	miR-16	Cell growth
hsa_circ_0010522	RAP1GAP	Increased	(39)	miR-133	White adipose tissue browning, cancer-associated cachexia
has_circ_0032627	DLST	Increased	(40)	miR-502-3p, NRAS/MEK1/ERK1/2	Cell viability, invasion, and metastasis
hsa_circ_0092303	CACTIN	Increased	(41)	miR-331-3p, TGFBR1	Cell growth and metastasis
circNRIP1	NRIP1	Increased	(42)	miR-149-5p	Cell growth and metastasis

cancer cells proliferation and invasion by sponging miR-153 thereby upregulating SNAI1 expression (34). CircPDSS1 was increased in GC tissues and promoted GC cell cycle and inhibited apoptosis by sponging miR-186-5p and upregulating NEK2 expression (35). Hsa\_circ\_0000234 was decreased in GC tissues and regulated VAMP3 expression by sponging miR-142-3p in the development of GC (36). CircPSMC3 was decreased in GC tissues which was correlated with higher TNM stage and shorter overall survival. CircPSMC3 overexpression suppressed the tumorigenesis of GC by competitively binding endogenous miR-296-5p to regulate the PTEN expression (37). CircNF1 was increased in GC tissues and promotes cell proliferation by targeting miR-16, thereby decreasing the expression of its downstream target MAP7 and AKT3 (38). Ba's group identified an exosomal circRNA named ciRS-133, which was delivered into preadipocytes thereby promoting the white adipose tissue (WAT) browning in GC patients by suppressing miR-133 and activating PRDM16 (39). CircDLST was elevated in GC tissues and promotes the tumorigenesis and metastasis by sponging miR-502-5p thereby activating the miR-502-5p mediated NRAS/MEK1/ERK1/2 signaling (40). CircCACTIN expression was highly increased in GC tissues which promoted GC cells migration, invasion and EMT by sponging miRNA-331-3p and regulating mRNA expression of TGFBR1 (41). CircNRIP1 promotes tumorigenesis in GC by sponging miR-149-5p to downregulate AKT1. Moreover, circNRIP1 promotes tumor metastasis by transmitting exosomal communication among GC cells (42).

We realized that the functional study of circRNAs in GC is limited in their role as miRNA sponges in GC cell proliferation,

invasion and prognosis. However, this may be the very first step to understanding the circRNAs functions in cancers. Some of the circRNAs which have RBPs, binding sites, or enzymes may function as protein sponges or protein scaffolds while their exact roles in cancer have not been explored (52, 53).

#### CircRNA Serves as Diagnostic Biomarkers in GC

Circular RNAs feature in abundance, structurally conserved and stable, which exhibits high tissue-specific expression (54). The high structural stability makes circRNAs resistive to ribonucleases and enables them to be detected in human blood, saliva, and gastric fluid (44, 55, 56). These characteristics increase the potential of circRNAs to become diagnostic biomarkers in GC. Additionally, circRNAs significantly correlated with distal metastasis, tumor node metastasis (TNM) stage, gender, and age in GC, which make them suitable as a cancer biomarkers (57). In recent years, plenty of studies have verified circRNAs regarding their clinical significance and the roles in GC. Their clinical relevance with GC diagnosis was summarized below.

Hsa\_circ\_0000190 exhibits a decreased level in GC tissues and GC patients' plasma samples, the expressions of which are correlated with tumor diameter, lymphatic metastasis, distal metastasis, and TNM stage. The AUC in tissues and plasma was better than CEA and CA19-9 as GC biomarkers which were reached to 0.75 and 0.60, respectively (58). Hsa\_circ\_0000745 was decreased in GC tissues and plasma samples from patients with GC. Hsa\_circ\_0000745 expression in GC tissues and plasma correlated with tumor differentiation and TNM, respectively. The AUC of hsa\_circ\_0000745 in plasma

was 0.683 and the combination of plasma hsa\_circ\_0000745 level and CEA level increased the AUC to 0.775, indicating hsa\_circ\_0000745 could be a good GC diagnosis biomarker (59). Hsa circ 0001017 and hsa circ 0061276 are decreased in GC tissues whose levels were significantly correlated with distal metastasis. Combinative use of above two circRNAs increases the AUG to 0.966 with 95.5% sensitivity and 95.7% specificity, suggesting their suitability as potential biomarkers in GC diagnosis (47). CircPVRL3 was decreased in GC tissues with 90.3% sensitivity and 56.4% specificity. The potential diagnostic value of the circPVLR3 was analyzed with AUC of 0.7626. Furthermore, the AUC of circPVRL3 expression varied in different TNM stages, creating the possibility for circPVLR3 to become a prognostic biomarker (60). The circSMARCA5 level was decreased in GC tissues and cell lines. CircSMARCA5 upregulation inhibited GC cells proliferation, migration and invasion, inversely, circSMARCA5 knockdown promoted GC progression. The AUC of circSMARCA5 was 0.806, indicating that the circRNA could function as a potential diagnosis biomarker (61). Hsa\_circ\_0000467 was significantly increased in GC tissue and in GC cell lines as well as in the plasma samples from GC patients. The AUC of hsa\_circ\_0000467 to discriminate GC patients and healthy individuals was 0.790, suggesting that hsa\_circ\_0000467 might be a potential diagnosis biomarker in GC. Moreover, hsa circ 0000467 might act as a prognosis biomarker due to the close association between hsa circ 0000467 and TNM stage (62). CircPSMC3 was decreased in GC tissues, GC patients' plasmas and GC cell lines. Down-expression of circPSMC3 was negatively associated with TNM stage and lymphatic metastasis with the AUC up to 0.9326 along with the 85.85% sensitivity and 95.24% specificity, indicating that circPSMC3 is a potential biomarker with a high diagnosis value (37). Circ-DCAF6 was found increased in human GC tissues compared to adjacent non-tumor samples. The high circ-DCAF6 expression was evidently correlated with depth of invasion, lymph node invasion and TNM stages but was not associated with gender, age, tumor size and differentiation grade. At a prognosis level, the positive expression of circ-DCAF6 is highly correlated

with GC patient overall survival. Therefore, circ-DCAF6 may act as a useful biomarker for GC (63).

# CircRNA IN CANCER THERAPY RESISTANCE

As illustrated above, gene expression profiles screening implicated the possible role of circRNAs in GC progression, and the deregulation of circRNAs that influenced the GC cell proliferation, migration and invasion signaling, furthermore, correlated with the occurrence and development of GC mean circRNAs could become a potential diagnosis and prognosis biomarker for GC. In this manner, circRNAs may be used as a therapeutic target in GC treatment in the near future. Although we did not find any clinical reports regarding circRNAs treatment at the time of writing, accumulated evidences have demonstrated significant clinical relevance of circRNAs in cancer therapy. Here we will summarize the current status of circRNAs in cancer therapy resistance (**Table 2**).

First circRNA expression profiles based on microarray analysis using chemoradiation-resistant colorectal cancer (CRC) cells identified 71 differentially expressed circRNAs and the most modulated circRNAs were involved in signaling pathways associated with CRC development, which provided a useful database for further understanding chemoradiation resistance and the role of circRNAs in chemoradiation-resistant CRC development (64). CircPVT1, as mentioned above as a tumor promoter in GC, was found positively expressed in osteosarcoma (OS)tissues, serums as well as chemo-resistant cell lines. CircPVT1 knockdon weakens the resistance to doxorubicin and cisplatin of OS cells by decreasing ABCB1 gene expression (65). Differentially expressed circRNAs were demonstrated in Gemcitabine resistant pancreatic ductal adenocarcinoma (PDAC) cell lines and the two most increased circRNAs in patients with Gemcitabine resistance compared to the nonresistant patients were identified. Two circRNAs knockdown keep the sensitivity of PANC-1-GR cells to Gemcitabine

CircRNAs	Cancer Deregulation Gene affected		Associated cell process	References					
circPVT1	Osteosarcoma	Increased	ABCB1	Resistance to doxorubicin and cisplatin	in (65)				
circ_0001258	Osteosarcoma	Decreased	miR-744-3p/GSTM2		(75)				
circ_0000285	Bladder cancer	Decreased		Prognosis and cisplatin resistance	(69)				
circ_0025202	Breast cancer	Decreased	miR-182-5p/FOXO3a	Sensitivity to tamoxifen	(71)				
circRNA-007874	Breast cancer	Increased	TRAF4/Eg5	Reverse monastrol resistance	(76)				
circ_0004015	NSCLC	Increased	miR-1183/PDPK1	Promotes TKI drug resistance	(77)				
circ_0004870	CRPC	Decreased	RBM39	Resistance to enzalutamide	(70)				
circ_100053	Chronic myeloid leukemia	Increased		Resistance to imatinib	(78)				
circBA9.3	Chronic myeloid leukemia	Increased	c-ABL1/BCR-ABL1	Resistance against TKI therapy	(72)				
circPAN3	Acute myeloid leukemia	Increased	miR-153-5p/miR-183-5p-XIAP	Resistance to doxorubicin	(73)				
hg19_circ_0005033	LSCC	Increased	miR-4521	Resistance to chemotherapy	(79)				
hsa_circ_0000199	Gastric cancer	Increased	miR-198/PIK3R1	Resistance to CDDP	(33)				
hsa_circ_0081143	Gastric cancer	Increased	miR-646/CDK6	Resistance to CDDP resistance	(74)				

CRPC, castration-resistant prostate cancer; NSCLC, non-small cell lung cancer; LSCC, laryngeal squamous cell carcinoma.

TABLE 2 | CircBNA in cancer therapy resistance



treatment, however, overexpression of them increased the resistance (66). Screening of differentially expressed circRNA profiles in taxol-resistant and AZD9291-resistant non-small cell lung cancer (NSCLC) cells provides a database for further understanding the drug resistance in NSCLC (67, 68). Hsa\_circ\_0000285 was significantly reduced in bladder cancer tissues and serum and had lower expression level in cisplatin-resistant bladder cancer patients. And the association of hsa\_circ\_0000285 with tumor size, differentiation, lymph node metastasis, distant metastasis and TNM stage was analyzed (69). Hsa\_circ\_0004870 was identified of its significance in enzalutamide resistance development in castration-resistant prostate cancer (CRPC) through association with RBM39 gene (70). Hsa\_circ\_0025202 was significantly decreased in

tamoxifen-resistant breast cancer and regulated the miR-182-5p/FOXO3a axis to achieve tamoxifen sensitivity and tumor progression (71). CircBA9.3 promoted the proliferation and inhibited apoptosis of CML cells, and was validated as increased in patients with TKI resistance, which was positively correlated with BCR-ABL1 expression (72). CircPAN3 regulated doxorubicin (ADM) resistance in acute myeloid leukemia by regulating the miR-153-5p/miR-183-5p-XIAP signaling (73). At the time of writing, two reports had demonstrated the role of circRNAs in GC resistance and the molecular mechanisms were explored. Hsa\_circ\_0000199 was increased in CDDP-resistant GC tissues and cells and was proved regulating the expression of PIK3R1, activating the PI3K/AKT signaling pathway and targeting miR-198 to facilitate the resistance of CDDP in GC cells (33). Hsa\_circ\_0081143 was increased in GC tissues and inhibition of hsa\_circ\_0081143 induced the GC cells sensitivity to CDDP *in vitro* and through increasing miR-646 expression to downregulate CDK6 expression *in vivo* (74). These results indicated the possibility of circRNAs to serve as potential biomarkers and therapeutic targets for various cancer types.

# **FUTURE PERSPECTIVES**

Even though the studies focusing on the roles of circRNAs in cancers are just beginning, researchers obviously have good prospects regarding the circRNAs functioning as potential therapeutic targets for cancer treatment. As in GC, a growing number of circRNAs were characterized pathologically associated with GC and the mechanisms in regulating GC cell proliferation, invasion and prognosis were studied (**Figure 2**). The circRNAs are structurally stable, differentially expressed with tissue-and disease-specificity in GC making them a potential diagnosis biomarker. However, the study regarding their biogenesis, cellular location, function, and the regulation mechanism for most of the circRNAs remains insufficient.

More attention was paid to the relationship of endogenous circRNAsin GC occurrence and development in recent studies. The functional efficacy of synthetic circRNA was first reported in decreasing GC cell proliferation, suppressing the miR-21 regulatory activity in posttranscriptional levels and increasing miR-21 downstream proteins expression. In future studies, the role of the synthetic circRNA in inhibiting tumor growth and the toxicity in an animal model should be evaluated. Furthermore, the synthetic circRNA can be easily synthesized and designed thereby applied to target other cancer types or diseases and can be tested in clinical practices (51). Exosomes have been reported to be involved in almost every process of cancer (80, 81), and cell derived exosomes were used as cancer drug delivery systems that obviously enhanced drug cytotoxicity in cancer treatment (82, 83). Recent study reported that circulating exosomal miRNAs in adipose tissue was able to regulate gene expression in distant tissues, implying exosomes are important mediators in signal transduction (84). The cancer-secreted circRNAs were first demonstrated in GC that the exosomal ciRS-133 can be transported into preadipocytes and promoted white adipose tissue browning. In addition, exosome-delivered ciRS-133 aggravated tumor cachexia, of which the underlying mechanism required further study (39). The studies with new points of view not only provided a new horizon toward the possible roles of circRNAs in GC progression but also offer new perspectives for using circRNAs as a novel tool in GC treatment.

It is beyond doubt that the research of circRNAs is still in its beginning, continued work should be carried until the practical application of circRNAs for cancer treatment is better understood. Most screening studies of circRNAs mentioned above were performed using RNA seq on a limited number of GC tissues and healthy controls. Although most differently expressed circRNAs in certain cancer types can be validated using more than one prediction algorithm and RT-qPCR, the expression level of these circRNAs in other cancer types has never been evaluated. Therefore, it hampers the application of singular circRNAs as specific biomarker for a certain cancer type. So far, the studies of mechanisms of certain circRNA in cancer progression were emphasized on its regulatory function as a miRNA sponge. This is somewhat biased because we know very little about what, in practice, causes the deregulation of circRNAs in cancer at the time of writing. There is speculation that the high dysregulation may be partially due to the gene changes genetically and/or epigenetically involved in their biogenesis (85), as of yet, this is still a question that remains to be answered. Moreover, the involved miRNA might target hundreds of genes thus the regulatory network from circRNA to miRNA and mRNA must be complicated and might influence multiple cancer related signaling pathway. At the time of writing, we know almost nothing about circRNAs-targeted therapy even if a few reports have highlighted the circRNAs as master-regulators of drug resistance in several cancer types (86). Continued work should be carried out to reveal the biogenesis, regulatory functions and molecular mechanisms both pathologically and physiologically.

# CONCLUSION

In conclusion, circRNA research is at the beginning of its route to clinical practices. A growing number of new circRNAs have been identified that were deregulated in GC tissues, plasma and cell lines, and the deregulation was closely correlated with GC cell proliferation, apoptosis, invasion and metastasis by mediating endogenous miRNA/target genes signaling pathway. The stability and its tissue- and disease-specificity of circRNAs make them novel biomarkers and potential therapeutic targets in GC diagnosis and treatment. Moreover, circRNAs associated with drug resistance were validated in GC and other cancer types although the research is still in its infancy. In the future, the physiological aspects of circRNAs regarding the biogenesis, cellular location and functions need to be determined and their regulatory networks in GC and other diseases remain to be extensively and systemically investigated.

# **AUTHOR CONTRIBUTIONS**

WW and QY designed the concept and draft the manuscript. TZ and JY analyzed the data in the references and revised the language. All authors contributed to the article and approved the submitted version.

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# REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2018) 68:394–424. doi: 10.3322/caac.21492
- Van Cutsem E, Sagaert X, Topal B, Haustermans K, Prenen H. Gastric cancer. Lancet. (2016) 388:2654–64.
- Digklia A, Wagner AD. Advanced gastric cancer: current treatment landscape and future perspectives. World J Gastroenterol. (2016) 22:2403–14. doi: 10. 3748/wjg.v22.i8.2403
- Allemani C, Weir HK, Carreira H, Harewood R, Spika D, Wang X-S, et al. Global surveillance of cancer survival 1995-2009: analysis of individual data for 25 676 887 patients from 279 population-based registries in 67 countries (CONCORD-2). *Lancet*. (2015) 385:977–1010. doi: 10.1016/s0140-6736(14) 62038-9
- Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature*. (2013) 495:333–8. doi: 10.1038/nature11928
- van Rossum D, Verheijen BM, Pasterkamp RJ. Circular RNAs: novel regulators of neuronal development. *Front Mol Neurosci*. (2016) 9:74. doi: 10.3389/fnmol. 2016.00074
- Burd CE, Jeck WR, Liu Y, Sanoff HK, Wang Z, Sharpless NE. Expression of linear and novel circular forms of an INK4/ARF-associated non-coding RNA correlates with atherosclerosis risk. *PLoS Genet.* (2010) 6:e1001233. doi: 10.1371/journal.pgen.1001233
- Holdt LM, Stahringer A, Sass K, Pichler G, Kulak NA, Wilfert W, et al. Circular non-coding RNA ANRIL modulates ribosomal RNA maturation and atherosclerosis in humans. *Nat Commun.* (2016) 7:12429.
- Zhou Z-B, Du D, Huang G-X, Chen A, Zhu L. Circular RNA Atp9b, a competing endogenous RNA, regulates the progression of osteoarthritis by targeting miR-138-5p. *Gene.* (2018) 646:203–9. doi: 10.1016/j.gene.2017.12. 064
- Zhang S-J, Chen X, Li C-P, Li X-M, Liu C, Liu B-H, et al. Identification and characterization of circular RNAs as a new class of putative biomarkers in diabetes retinopathy. *Invest Ophthalmol Vis Sci.* (2017) 58:6500–9. doi: 10.1167/iovs.17-22698
- Yang Z, Xie L, Han L, Qu X, Yang Y, Zhang Y, et al. Circular RNAs: regulators of cancer-related signaling pathways and potential diagnostic biomarkers for human cancers. *Theranostics*. (2017) 7:3106–17. doi: 10.7150/thno. 19016
- 12. Zhang H-D, Jiang L-H, Sun D-W, Hou J-C, Ji Z-L. CircRNA: a novel type of biomarker for cancer. *Breast Cancer*. (2018) 25:1–7.
- 13. Zhang M, Xin Y. Circular RNAs: a new frontier for cancer diagnosis and therapy. J Hematol Oncol. (2018) 11:21.
- Kong Z, Wan X, Zhang Y, Zhang P, Zhang Y, Zhang X, et al. Androgenresponsive circular RNA circSMARCA5 is up -regulated and promotes cell proliferation in prostate cancer. *Biochem Biophys Res Commun.* (2017) 493:1217–23. doi: 10.1016/j.bbrc.2017.07.162
- Zhong Z, Huang M, Lv M, He Y, Duan C, Zhang L, et al. Circular RNA MYLK as a competing endogenous RNA promotes bladder cancer progression through modulating VEGFA/VEGFR2 signaling pathway. *Cancer Lett.* (2017) 403:305–17. doi: 10.1016/j.canlet.2017.06.027
- Liang H-F, Zhang X-Z, Liu B-G, Jia G-T, Li W-L. Circular RNA circ-ABCB10 promotes breast cancer proliferation and progression through sponging miR-1271. Am J Cancer Res. (2017) 7:1566–76.
- Song X, Zhang N, Han P, Moon B-S, Lai RK, Wang K, et al. Circular RNA profile in gliomas revealed by identification tool UROBORUS. *Nucleic Acids Res.* (2016) 44:e87. doi: 10.1093/nar/gkw075
- Yang Y, Gao X, Zhang M, Yan S, Sun C, Xiao F, et al. Novel role of FBXW7 circular RNA in repressing glioma tumorigenesis. J Natl Cancer Inst. (2018) 110:304–15. doi: 10.1093/jnci/djx166
- Li Y, Zheng F, Xiao X, Xie F, Tao D, Huang C, et al. CircHIPK3 sponges miR-558 to suppress heparanase expression in bladder cancer cells. *EMBO Rep.* (2017) 18:1646–59. doi: 10.15252/embr.201643581
- Tang W, Fu K, Sun H, Rong D, Wang H, Cao H. CircRNA microarray profiling identifies a novel circulating biomarker for detection of gastric cancer. *Mol Cancer.* (2018) 17:137.

- Liang M, Liu Z, Lin H, Shi B, Li M, Chen T, et al. High-throughput sequencing reveals circular RNA hsa\_circ\_0000592 as a novel player in the carcinogenesis of gastric carcinoma. *Biosci Rep.* (2019) 39:BSR20181900.
- 22. Chen J. Circular RNA profile identifies circPVT1 as a proliferative factor and prognostic marker in gastric cancer. *Ann Oncol.* (2016) 27:ix68–85.
- 23. Chen J, Wang Y. Comprehensive circular RNA profile and bioinformatics analyses reveal that circLMTK2 is upregulated and as a proliferative factor and prognostic marker in gastric cancer. *Ann Oncol.* (2017) 28:7.
- Li P, Chen H, Chen S, Mo X, Li T, Xiao B, et al. Circular RNA 0000096 affects cell growth and migration in gastric cancer. *Br J Cancer*. (2017) 116:626–33. doi: 10.1038/bjc.2016.451
- Zhang J, Liu H, Hou L, Wang G, Zhang R, Huang Y, et al. Circular RNA\_LARP4 inhibits cell proliferation and invasion of gastric cancer by sponging miR-424-5p and regulating LATS1 expression. *Mol Cancer*. (2017) 16:151.
- Zhang Y, Liu H, Li W, Yu J, Li J, Shen Z, et al. CircRNA\_100269 is decreased in gastric cancer and suppresses tumor cell growth by targeting miR-630. *Aging*. (2017) 9:1585–94. doi: 10.18632/aging.101254
- Liu H, Liu Y, Bian Z, Zhang J, Zhang R, Chen X, et al. Circular RNA YAP1 inhibits the proliferation and invasion of gastric cancer cells by regulating the miR-367-5p/p27 (Kip1) axis. *Mol Cancer*. (2018) 17:151.
- Zhong S, Wang J, Hou J, Zhang Q, Xu H, Hu J, et al. Circular RNA hsa\_circ\_0000993 inhibits metastasis of gastric cancer cells. *Epigenomics*. (2018) 10:1301–13. doi: 10.2217/epi-2017-0173
- Chen Y, Yang F, Fang E, Xiao W, Mei H, Li H, et al. Circular RNA circAGO2 drives cancer progression through facilitating HuR-repressed functions of AGO2-miRNA complexes. *Cell Death Differ*. (2019) 26:1346–64. doi: 10.1038/ s41418-018-0220-6
- Ding L, Zhao Y, Dang S, Wang Y, Li X, Yu X, et al. Circular RNA circ-DONSON facilitates gastric cancer growth and invasion via NURF complex dependent activation o transcription factor SOX4. *Mol Cancer*. (2019) 18:45.
- Fang J, Hong H, Xue X, Zhu X, Jiang L, Qin M, et al. A novel circular RNA, circFAT1(e2), inhibits gastric cancer progression by targeting miR-548g in the cytoplasm and interacting with YBX1 in the nucleus. *Cancer Lett.* (2019) 442:222–32. doi: 10.1016/j.canlet.2018.10.040
- Hong Y, Qin H, Li Y, Zhang Y, Zhuang X, Liu L, et al. FNDC3B circular RNA promotes the migration and invasion of gastric cancer cells via the regulation of E-cadherin and CD44 expression. *J Cell Physiol.* (2019) 234:19895–910. doi: 10.1002/jcp.28588
- Huang X, Li Z, Zhang Q, Wang W, Li B, Wang L, et al. Circular RNA AKT3 upregulates PIK3R1 to enhance cisplatin resistance in gastric cancer via miR-198 suppression. *Mol Cancer*. (2019) 18:71.
- Li G, Xue M, Yang F, Jin Y, Fan Y, Li W. CircRBMS3 promotes gastric cancer tumorigenesis by regulating miR-153-SNAI1 axis. *J Cell Physiol.* (2019) 234:3020–8. doi: 10.1002/jcp.27122
- Ouyang Y, Li Y, Huang Y, Li X, Zhu Y, Long Y, et al. CircRNA circPDSS1 promotes the gastric cancer progression by sponging miR-186-5p and modulating NEK2. J Cell Physiol. (2019) 234:10458–69. doi: 10.1002/jcp.27714
- Peng L, Zhang G. Hsa\_Circ\_0000234 functions as a competitive endogenous RNA to regulate VAMP3 expression by sponging MIR-142-3P in gastric cancer. *Gastroenterology*. (2019) 156:S1376–1376.
- Rong D, Lu C, Zhang B, Fu K, Zhao S, Tang W, et al. CircPSMC3 suppresses the proliferation and metastasis of gastric cancer by acting as a competitive endogenous RNA through sponging miR-296-5p. *Mol Cancer*. (2019) 18:25.
- Wang Z, Ma K, Cheng Y, Meltzer SJ. Novel circular RNA NF1 acts as a sponge, promoting gastric cancer by absorbing MIR-16. *Gastroenterology*. (2019) 156:S1175–1175.
- Zhang H, Zhu L, Bai M, Liu Y, Zhan Y, Deng T, et al. Exosomal circRNA derived from gastric tumor promotes white adipose browning by targeting the miR-133/PRDM16 pathway. *Int J Cancer.* (2019) 144:2501–15. doi: 10.1002/ ijc.31977
- Zhang J, Hou L, Liang R, Chen X, Zhang R, Chen W, et al. CircDLST promotes the tumorigenesis and metastasis of gastric cancer by sponging miR-502-5p and activating the NRAS/MEK1/ERK1/2 signaling. *Mol Cancer*. (2019) 18:80.
- Zhang L, Song X, Chen X, Wang Q, Zheng X, Wu C, et al. Circular RNA CircCACTIN promotes gastric cancer progression by sponging MiR-331-3p and regulating TGFBR1 expression. *Int J Biol Sci.* (2019) 15:1091–103. doi: 10.7150/ijbs.31533

- Zhang X, Wang S, Wang H, Cao J, Huang X, Chen Z, et al. Circular RNA circNRIP1 acts as a microRNA-149-5p sponge to promote gastric cancer progression via the AKT1/mTOR pathway. *Mol Cancer.* (2019) 18:20.
- Dang Y, Ouyang X, Zhang F, Wang K, Lin Y, Sun B, et al. Circular RNAs expression profiles in human gastric cancer. *Sci Rep.* (2017) 7:9060.
- Shao Y, Li J, Lu R, Li T, Yang Y, Xiao B, et al. Global circular RNA expression profile of human gastric cancer and its clinical significance. *Cancer Med.* (2017) 6:1173–80. doi: 10.1002/cam4.1055
- Sui W, Shi Z, Xue W, Ou M, Zhu Y, Chen J, et al. Circular RNA and gene expression profiles in gastric cancer based on microarray chip technology. *Oncol Rep.* (2017) 37:1804–14. doi: 10.3892/or.2017.5415
- 46. Vidal AF, Ribeiro-dos-Santos AM, Vinasco-Sandoval T, Magalhaes L, Pinto P, Anaissi AKM, et al. The comprehensive expression analysis of circular RNAs in gastric cancer and its association with field cancerization. *Sci Rep.* (2017) 7:14551.
- Li T, Shao Y, Fu L, Xie Y, Zhu L, Sun W, et al. Plasma circular RNA profiling of patients with gastric cancer and their droplet digital RT-PCR detection. *J Mol Med.* (2018) 96:85–96. doi: 10.1007/s00109-017-1600-y
- Hao S, Lv J, Yang Q, Wang A, Li Z, Guo Y, et al. Identification of key genes and circular RNAs in human gastric cancer. *Med Sci Monit*. (2019) 25:2488–504. doi: 10.12659/msm.915382
- Kohlhapp FJ, Mitra AK, Lengyel E, Peter ME. MicroRNAs as mediators and communicators between cancer cells and the tumor microenvironment. *Oncogene*. (2015) 34:5857–68. doi: 10.1038/onc.2015.89
- Rupaimoole R, Calin GA, Lopez-Berestein G, Sood AK. miRNA deregulation in cancer cells and the tumor microenvironment. *Cancer Discov.* (2016) 6:235–46. doi: 10.1158/2159-8290.cd-15-0893
- Liu X, Abraham JM, Cheng Y, Wang Z, Wang Z, Zhang G, et al. Synthetic circular RNA functions as a MIR-21 sponge to suppress gastric carcinoma cell proliferation. *Mol Ther Nucleic Acids*. (2018) 13:312–21. doi: 10.1016/j.omtn. 2018.09.010
- Abdelmohsen K, Panda AC, Munk R, Grammatikakis I, Dudekula DB, De S, et al. Identification of HuR target circular RNAs uncovers suppression of PABPN1 translation by CircPABPN1. *RNA Biol.* (2017) 14:361–9. doi: 10.1080/15476286.2017.1279788
- Du WW, Fang L, Yang W, Wu N, Awan FM, Yang Z, et al. Induction of tumor apoptosis through a circular RNA enhancing Foxo3 activity. *Cell Death Differ*. (2017) 24:357–70. doi: 10.1038/cdd.2016.133
- Salzman J, Chen RE, Olsen MN, Wang PL, Brown PO. Cell-type specific features of circular RNA expression. *PLoS Genet.* (2013) 9:e1003777. doi: 10. 1371/journal.pgen.1003777
- Bahn JH, Zhang Q, Li F, Chan T-M, Lin X, Kim Y, et al. The landscape of microRNA, Piwi-interacting RNA, and circular RNA in human saliva. *Clin Chem.* (2015) 61:221–30. doi: 10.1373/clinchem.2014. 230433
- Memczak S, Papavasileiou P, Peters O, Rajewsky N. Identification and characterization of circular RNAs as a new class of putative biomarkers in human blood. *PLoS One.* (2015) 10:e0141214. doi: 10.1371/journal.pone. 0141214
- Li P, Chen S, Chen H, Mo X, Li T, Shao Y, et al. Using circular RNA as a novel type of biomarker in the screening of gastric cancer. *Clin Chim Acta*. (2015) 444:132–6. doi: 10.1016/j.cca.2015.02.018
- Chen S, Li T, Zhao Q, Xiao B, Guo J. Using circular RNA hsa\_circ\_0000190 as a new biomarker in the diagnosis of gastric cancer. *Clin Chim Acta*. (2017) 466:167–71. doi: 10.1016/j.cca.2017.01.025
- Huang M, He Y-R, Liang L-C, Huang Q, Zhu Z-Q. Circular RNA hsa\_circ\_0000745 may serve as a diagnostic marker for gastric cancer. *World J Gastroenterol.* (2017) 23:6330–8. doi: 10.3748/wjg.v23. i34.6330
- Sun H-D, Xu Z-P, Sun Z-Q, Zhu B, Wang Q, Zhou J, et al. Down-regulation of circPVRL3 promotes the proliferation and migration of gastric cancer cells. *Sci Rep.* (2018) 8:10111.
- Cai J, Chen Z, Zuo X. circSMARCA5 functions as a diagnostic and prognostic biomarker for gastric cancer. *Dis Mark*. (2019) 2019:2473652.
- Lu J, Zhang P-Y, Xie J-W, Wang J-B, Lin J-X, Chen Q-Y, et al. Hsa\_circ\_0000467 promotes cancer progression and serves as a diagnostic and prognostic biomarker for gastric cancer. J Clin Lab Anal. (2019) 33:e22726. doi: 10.1002/jcla.22726

- Wu L, Liu D, Yang Y. Enhanced expression of circular RNA circ-DCAF6 predicts adverse prognosis and promotes cell progression via sponging miR-1231 and miR-1256 in gastric cancer. *Exp Mol Pathol.* (2019) 110:104273. doi: 10.1016/j.yexmp.2019.104273
- 64. Xiong W, Ai YQ, Li YF, Ye Q, Chen ZT, Qin JY, et al. Microarray analysis of circular RNA expression profile associated with 5-Fluorouracil-based chemoradiation resistance in colorectal cancer cells. *Biomed Res Int.* (2017) 2017:8421614.
- Kun-Peng Z, Xiao-Long M, Chun-Lin Z. Overexpressed circPVT1, a potential new circular RNA biomarker, contributes to doxorubicin and cisplatin resistance of osteosarcoma cells by regulating ABCB1. *Int J Biol Sci.* (2018) 14:321–30. doi: 10.7150/ijbs.24360
- Shao F, Huang M, Meng F, Huang Q. Circular RNA signature predicts gemcitabine resistance of pancreatic ductal adenocarcinoma. *Front Pharmacol.* (2018) 9:584. doi: 10.3389/fphar.2018.00584
- Xu N, Chen S, Liu Y, Li W, Liu Z, Bian X, et al. Profiles and bioinformatics analysis of differentially expressed circrnas in taxol-resistant non-small cell lung cancer cells. *Cell Physiol Biochem*. (2018) 48:2046–60. doi: 10.1159/ 000492543
- Chen T. Comprehensive analysis of circular RNA profiling in AZD9291resistant non-small cell lung cancer cell lines. *Mol Cancer*. (2019) 10:930–41. doi: 10.1111/1759-7714.13032
- Chi BJ, Zhao DM, Liu L, Yin XZ, Wang FF, Bi S, et al. Downregulation of hsa\_circ\_0000285 serves as a prognostic biomarker for bladder cancer and is involved in cisplatin resistance. *Neoplasma*. (2019) 66:197–202. doi: 10.4149/ neo\_2018\_180318n185
- Greene J, Baird AM, Casey O, Brady L, Blackshields G. Circular RNAs are differentially expressed in prostate cancer and are potentially associated with resistance to enzalutamide. *Sci Rep.* (2019) 9:10739.
- Sang Y, Chen B, Song X, Li Y, Liang Y, Han D, et al. circRNA\_0025202 regulates Tamoxifen sensitivity and tumor progression via regulating the miR-182-5p/FOXO3a axis in breast cancer. *Mol Ther.* (2019) 27:1638–52. doi: 10.1016/j.ymthe.2019.05.011
- Pan Y, Lou J, Wang H, An N, Chen H, Zhang Q, et al. CircBA9.3 supports the survival of leukaemic cells by up-regulating c-ABL1 or BCR-ABL1 protein levels. *Blood Cells Mol Dis.* (2018) 73:38–44. doi: 10.1016/j.bcmd.2018.09.002
- Shang J, Chen WM, Wang ZH, Wei TN, Chen ZZ, Wu WB. CircPAN3 mediates drug resistance in acute myeloid leukemia through the miR-153-5p/miR-183-5p-XIAP axis. *Exp Hematol.* (2019) 70:e3.
- Xue M, Li G, Fang X, Wang L, Jin Y, Zhou Q. hsa\_circ\_0081143 promotes cisplatin resistance in gastric cancer by targeting miR-646/CDK6 pathway. *Cancer Cell Int.* (2019) 19:25.
- Zhu KP, Zhang CL, Ma XL, Hu JP, Cai T, Zhang L. Analyzing the interactions of mRNAs and ncRNAs to predict competing endogenous RNA networks in osteosarcoma chemo-resistance. *Mol Ther.* (2019) 27:518–30. doi: 10.1016/j. ymthe.2019.01.001
- Liu Y, Dong Y, Zhao L, Su L, Luo J. Circular RNAMTO1 suppresses breast cancer cell viability and reverses monastrol resistance through regulating the TRAF4/Eg5 axis. *Int J Oncol.* (2018) 53:1752–62.
- 77. Zhou Y, Zheng X, Xu B, Chen L, Wang Q, Deng H, et al. Circular RNA hsa\_circ\_0004015 regulates the proliferation, invasion, and TKI drug resistance of non-small cell lung cancer by miR-1183/PDPK1 signaling pathway. *Biochem Biophys Res Commun.* (2019) 508:527–35. doi: 10.1016/j. bbrc.2018.11.157
- 78. Gu Y, Huang J, Luo Q, Yang Y, Ping L, Jian-Jun C, et al. High circ\_100053 predicts a poor outcome for chronic myeloid leukemia and is involved in imatinib resistance. *Oncol Res.* (2019):doi: 10.3727/ 096504018X15412701483326 [Epub ahead of print],
- Wu Y, Zhang Y, Niu M, Shi Y, Liu H, Yang D, et al. Whole-transcriptome analysis of CD133+CD144+ cancer stem cells derived from human laryngeal squamous cell carcinoma cells. *Cell Physiol Biochem*. (2018) 47:1696–710.
- Yin Y, Cai X, Chen X, Liang H, Zhang Y, Li J, et al. Tumor-secreted miR-214 induces regulatory T cells: a major link between immune evasion and tumor growth. *Cell Res.* (2014) 24:1164–80. doi: 10.1038/cr.2014.121
- Zhang H, Bai M, Deng T, Liu R, Wang X, Qu Y, et al. Cell-derived microvesicles mediate the delivery of miR-29a/c to suppress angiogenesis in gastric carcinoma. *Cancer Lett.* (2016) 375:331–9. doi: 10.1016/j.canlet.2016. 03.026

- Kooijmans SAA, Vader P, van Dommelen SM, van Solinge WW, Schiffelers RM. Exosome mimetics: a novel class of drug delivery systems. *Int J Nanomed*. (2012) 7:1525–41. doi: 10.2147/ijn.s29661
- Saari H, Lazaro-Ibanez E, Viitala T, Vuorimaa-Laukkanen E, Siljander P, Yliperttula M. Microvesicle- and exosome-mediated drug delivery enhances the cytotoxicity of Paclitaxel in autologous prostate cancer cells. J Control Release. (2015) 220:727–37. doi: 10.1016/j.jconrel.2015.09.031
- Thomou T, Mori MA, Dreyfuss JM, Konishi M, Sakaguchi M, Wolfrum C, et al. Adipose-derived circulating miRNAs regulate gene expression in other tissues. *Nature*. (2017) 542:450–5. doi: 10.1038/nature 21365
- Kristensen LS, Hansen TB, Veno MT, Kjems J. Circular RNAs in cancer: opportunities and challenges in the field. *Oncogene*. (2018) 37:555–65. doi: 10.1038/onc.2017.361
- Wang KW, Dong M. Role of circular RNAs in gastric cancer: recent advances and prospects. World J Gastrointest Oncol. (2019) 11:459–69. doi: 10.4251/ wjgo.v11.i6.459

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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