



The Role of Obesity-Induced Perivascular Adipose Tissue (PVAT) Dysfunction in Vascular Homeostasis

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Abstract: Perivascular adipose tissue (PVAT) is an additional special type of adipose tissue surrounding blood vessels. Under physiological conditions, PVAT plays a significant role in regulation of vascular tone, intravascular thermoregulation, and vascular smooth muscle cell (VSMC) proliferation. PVAT is responsible for releasing adipocytes-derived relaxing factors (ADRF) and perivascularderived relaxing factors (PDRF), which have anticontractile properties. Obesity induces increased oxidative stress, an inflammatory state, and hypoxia, which contribute to PVAT dysfunction. The exact mechanism of vascular dysfunction in obesity is still not well clarified; however, there are some pathways such as renin–angiotensin–aldosterone system (RAAS) disorders and PVAT-derived factor dysregulation, which are involved in hypertension and endothelial dysfunction development. Physical activity has a beneficial effect on PVAT function among obese patients by reducing the oxidative stress and inflammatory state. Diet, which is the second most beneficial non-invasive strategy in obesity treatment, may have a positive impact on PVAT-derived factors and may restore the balance in their concentration.

Keywords: obesity; perivascular adipose tissue; exercise; endothelial dysfunction

1. Introduction

Today, an increasing prevalence of obesity is observed in many countries, since a third of the worldwide population is described as obese or overweight [1]. National survey data from 2000 to 2018 in the USA reported that obesity prevalence increased to over 42% among adults, and the prevalence of severe obesity ($BMI \ge 40 \text{ kg/m}^2$) doubled to 9.2% over the study period [2]. Weight problems and obesity are increasing at a rapid rate in most of the EU Member States, with estimates of 52.7% of the EU's population being overweight in 2019 [3]. According to the Global Burden of Disease study, 4.7 million people died prematurely in 2017 as a result of obesity [4]. Obesity is a risk factor for developing many disorders such as diabetes mellitus, hypertension, cardiovascular events, obstructive sleep apnea syndrome, certain cancers, and musculoskeletal diseases [5]. Obesity also has a negative impact on quality of life and increases the costs of healthcare [6,7].

Adipose tissue is known as an endocrine organ. By producing adipokines, it regulates various metabolism pathways and processes such as insulin sensitivity, energy metabolism, blood flow, and even inflammatory stage [8,9]. Adipose tissue is divided into two main subtypes: white (WAT) and brown (BAT), according to their characteristic and different properties. WAT is responsible for storage of the excess of energy as fatty acids, while BAT mostly specializes in thermogenesis [10]. There is also a third type of adipocyte, termed the "beige" adipocyte. It is a brown adipocyte that arises within white adipose depots and also has thermogenic capacity [11].



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Perivascular adipose tissue (PVAT) is an additional special type of adipose tissue surrounding blood vessels. PVAT is located around the aorta, coronary arteries, small and resistance vessels, and vasculature of the musculoskeletal system [12–14] On the contrary, PVAT is absent among cerebral vessels [15]. It consists of stem cells, adipocytes, mast cells, and nerves [16,17]. PVAT lies outside the adventitia, with no laminar structures or any organized barrier separating them from each other. Although PVAT characteristics resemble both brown and white adipose tissues, recent evidence suggests that PVAT develops from its own distinct precursors, implying a closer link between PVAT and the vascular system [18]. PVAT at different anatomical locations presents different phenotypes. PVAT demonstrates WAT, BAT, and mixed phenotypes, depending on their anatomical placement [19]. In the abdominal PVAT, white adipocytes are more abundant, whereas thoracic PVAT contains more brown adipocytes. These regional differences in PVAT could explain the higher susceptibility of the abdominal aorta to atherosclerosis compared to the thoracic aorta [20–22]. Moreover, gender influences differences in PVAT. After menopause in women, there is an increase in perivascular and pericardial adipose tissue, and additionally, the volume of aortic PVAT positively correlates with the reduction in estradiol [23,24]. Additionally, in obesity experimental models, the PVAT mass and adipocyte size are increased [25]. PVAT, similar to every other adipose tissue, secretes cytokines, hormones, growth factors, and adipokines. It plays a beneficial role as long as adipokine levels with opposing properties remain in equilibrium. In obesity, PVAT becomes dysfunctional and exerts detrimental effects on vascular homeostasis [26].

2. The Influence of PVAT-Derived Factors on Vascular Function

Under physiological conditions, PVAT plays a significant role in the regulation of vascular tone, intravascular thermoregulation, and vascular smooth muscle cell (VSMC) proliferation (Figure 1) [27–29]. PVAT exhibits an anticontractile effect as a response to several factors such as endothelin-1, phenylephrine, angiotensin II, and serotonin [30–32]. PVAT anticontractile factors are divided into adipocytes-derived relaxing factors (ADRF) and perivascular-derived relaxing factors (PDRF) [30–33]. On the other hand, PVAT induces vasoconstriction by releasing angiotensin II [34] and the superoxide anion [35]. These factors affect vascular tone via endocrine and paracrine mechanisms. Moreover, VSMCs play a significant role in maintaining the balance between vasoconstriction and vasodilator signals. However, PVAT, as a special adipose tissue, is not only a mechanical support for the vasculature but plays a vital role in the homeostasis of the vascular system, sharing a status no less important than that of the endothelium [36].

2.1. Adiponectin

Adiponectin is profusely produced and released by PVAT under physiological conditions [37]. It is secreted in different polymeric forms, which differ from each other by molecular weight: multimeric, hexameric, trimeric, and globular [38]. Adiponectin is known as a vasodilator, which acts through different mechanisms affecting endothelial cells and VSMCs directly. First of all, adiponectin activates in both mentioned locations 5' adenosine monophosphate-activated protein kinase (AMPK), which is responsible for phosphorylation of endothelial nitric oxide synthase (eNOS) [39,40]. Moreover, adiponectin increases the production of 3,4-tetrahydrobiopterin (BH4), which is an important cofactor of eNOS [41]. As a consequence, the synthesis of the well-known vasodilator nitric oxide (NO) is increased. In addition, AMPK induces the opening of large-conductance calcium-activated potassium channels (BK channels) in VSMCs, which also contributes to vasodilation [42]. AMPK is activated mainly by globular and trimeric forms of adiponectin. Additional effects of multimeric adiponectin are the downregulation of glucose blood levels by the stimulation of glucose uptake by muscles, the improvement of insulin sensitivity, or a reduction in hepatic glucose production [43]. Moreover, it also regulates fatty acid metabolism by increasing the high-density lipoprotein (HDL) concentration and decreasing the triglycerides [44].



Figure 1. Vasodilatory and hyperpolarizing effect of perivascular adipose tissue–derived factors (PVAT-derived factors) on vascular smooth muscle cells (VSMCs) and hyperpolarizing effect of PVAT-derived factors on endothelial cells. Adiponectin causes vasodilation by affecting adiponectin receptors (AR) in endothelial cells, which contributes to the activation of locations 5' adenosine monophosphate-activated protein kinase (AMPK), which is responsible for the activation of endothelial NO synthase (eNOS). Enhanced NO concentration induces activation of cyclic guanosine monophosphate (cGMP), which is responsible for opening large-conductance calcium-activated potassium channels (BKCa). eNOS is present in both endothelial cells and adipocytes. Leptin activates leptin receptors (LR), which are responsible for activation of not only AMPK, but also endothelium-derived hyperpolarizing factor (EDHF), which activates BKCa. Moreover, AMPK independently activates BKCa and induces a hyperpolarization effect. Hydrogen sulfide (H2S) induces activation of BKCa in VSMCs and endothelial cells. Moreover, it induces a decrease in intracellular pH by the activates eNOS and increases the NO concentration. Hydrogen peroxide (H₂O₂) stimulates the soluble guanylyl cyclase (sGC-1), which induces vasodilation through the NO/GC-1/cGMP pathway.

2.2. Leptin

Leptin is another adipokine highly produced by adipose tissue. It regulates appetite by centrally affecting the hypothalamus and activating a sympathetic effect by affecting the arcuate nucleus in the hypothalamus [45]. Leptin synthesis is directly proportional to adipocyte size [46]. Leptin-induced vasodilation acts via endothelium-dependent and -independent mechanisms, which additionally depends on the type of vessel. In large arteries, such as the aorta, leptin increases endothelium-dependent vasodilation, in a similar way to adiponectin, by AMPK activation, which is responsible for eNOS phosphorylation [47]. In small arteries, such as the mesenteric artery, leptin-induced increased synthesis of NO and endothelium-derived hyperpolarizing factor (EDHF) contribute to endotheliumdependent vasodilation [48]. VSMCs are also known as a target for leptin, which impairs the contraction effect of angiotensin II by reducing the Ca²⁺ release from cellular reserves and inducing VSMC proliferation [47]. In addition, leptin in a high concentration can induce vasoconstriction by increasing the endothelin-1 release from endothelium [49]. Leptin is known as an immune-modulatory factor, which induces the production of proinflammatory cytokines such as IL-6, TNF α , or IL-12 and the differentiation of monocytes into macrophages [50].

2.3. H_2O_2

PVAT-derived hydrogen peroxide (H_2O_2) is known as both a vasoconstrictor and vasodilator factor by different mechanisms, which depend on the concentration of H_2O_2 , vessel type and vessel contractile status [51]. In healthy individuals, the concentration of H_2O_2 is nontoxic. First of all, H_2O_2 has membrane-permeable properties and freely diffuses to smooth muscle cells, where it stimulates the soluble guanylyl cyclase (sGC-1), which plays the role of receptor for NO in smooth muscles and induces vasodilation by the NO/sGC-1/cGMP pathway [52]. On the other hand, a high concentration of H_2O_2 stimulates cyclooxygenase and increases the level of Ca²⁺ [51].

2.4. H_2S

Hydrogen sulfide (H₂S) is a gaseous factor, which is produced by PVAT, endothelial cells, and VSMCs, controlling the vascular tone. H₂S-induced vasodilation is caused by activation of BK channels in VSMCs, which leads towards cell membrane hyperpolarization, inactivation of voltage-dependent l-type Ca²⁺ channels, and a decrease in intracellular Ca²⁺ concentration [53]. In addition, H₂S leads to a dose-dependent decrease in intracellular pH, which causes the vasodilation. It is suggested that the Cl⁻/HCO₃⁻ ionic exchanger is engaged in this process [54]. The shortage of H₂S is important in the development of various cardiovascular diseases, such as hypertension, atherosclerosis, and heart failure [55].

2.5. Angiotensin 1-7 and Angiotensin II

Components of the renin–angiotensin–aldosterone system (RAAS) are present in the aortic and mesenteric PVAT, except renin [56]. The effect of factors on vascular tone is different. Angiotensin 1–7 induce vasodilation by endothelium-dependent mechanisms. After the activation of the Mas receptors, located in the endothelium, the synthesis of NO is increased, which leads to vasodilation by the activation of BK channels [57]. On the contrary, angiotensin II, which is also produced by PVAT, induces vasoconstriction. There are regional differences in angiotensin II synthesis by PVAT, while it is greater in mesenteric adipose tissue than in the periaortic adipose tissue [35]. Moreover, angiotensin II in increased concentration activates immune cells, which can produce cytokines and proinflammatory mediators [58].

2.6. NO

Nitric oxide (NO) is a well-known endogenous gas with vasodilative properties, which is produced in almost all human cells. There are three isoforms of NOS: neuronal NOS (nNOS), inducible NOS (iNOS), and endothelial NOS (eNOS) [59]. Each of them is characterized by different attributes. nNOS is present in cells of the central and peripheral nervous system, where produced NO acts as a neurotransmitter and plays a role in the central regulation of blood pressure [60]. iNOS is activated by inflammatory cytokines and plays a role in inflammation; additionally, iNOS is Ca²⁺ independent, unlike other isoforms [59]. eNOS, which is located in endothelial cells, regulates blood pressure locally and has an antiatherosclerotic effect [60,61]. PVAT is responsible for increased production of NO by a direct mechanism, while eNOS isoform is also present in PVAT, where NO is directly produced and released affecting vasculature [62]. In addition, NO produced in PVAT positively regulates adiponectin release by PVAT [37]. On the other hand, PVAT-derived factors, mentioned in previous sections, increase NO production, which is responsible for activations of BK channels and stimulating cGMP synthesis endothelium and smooth muscle cells.

2.7. COX-Derived Factors

PVAT is also known as a source of a group of adipose-derived factors, which are produced by cyclooxygenase (COX), such as thromboxane A2 (TXA2), prostaglandin D2, prostaglandin E2, prostaglandin, F2a, prostaglandin H2, and prostaglandin I (prostacyclin) [63,64]. Both COX-1 and COX-2 isoforms are present in adipocytes of PVAT [64]. COX-derived factors have a different effect on vascular tone. Under normal conditions, serotonin stimulates the production of TXA2, which is characterized by pro-contractile and pro-inflammatory properties [65]. Prostacyclin is known as a vasodilator, which additionally reacts with receptors on the vasculature and plays a significant role in pro-tection against endothelium dysfunction and atherosclerosis [27,66,67]. Moreover, the concentration of prostacyclin declines with age progression [27].

2.8. Noradrenaline

PVAT is known as a source of noradrenaline (NA), which is the main neurotransmitter in the sympathetic nervous system (SNS). NA induces vasoconstriction of vessels by activation of adrenoreceptors localized on VSMCs [68]. NA is released by the endings of sympathetic nerves, which are spread among adipose tissue. However, it is reported that NA is additionally synthesized by PVAT independently of SNS activation, while surgical denervation of PVAT decreased NA concentration insignificantly [69,70]. Moreover, Ayala-Lopez et al. also indicated that NA is stored in PVAT, while tyramine stimulation induced an increased release of catecholamines from PVAT. In addition, NA stimulates the synthesis of H_2O_2 by PVAT, which has an anti-contractile effect [71].

3. The Role of Inflammation, Oxidative Stress, and Hypoxia in Obesity

Obesity is characterized by an excessive level of triglycerides and lipids, which are stored in adipocytes. It leads to their hyperplasia and hypertrophy, where hyperplasia is a well-tolerated complication. In contrast, it is suggested that the capacity of lipid storage and subsequent growth in adipocyte size is limited, and exceeding this threshold induces serious molecular changes and induces cellular dysfunction and death of adipocytes [72]. Moreover, enlarged adipocytes induce elevation of IL-6, IL-8, and leptin and decrease the level of adiponectin, which leads to consequent accumulation of inflammatory factors in PVAT [73,74]. Cytokines, fatty acids, and cell-free DNA, which are excreted after adipocytes apoptosis, induce migration of macrophages to the adipose tissue. Adipose tissue macrophages are divided into two subgroups, which differ from each other by type of secreted cytokines and cell markers: M1 with an inflammatory profile and M2 with an immunosuppressive feature [75]. The M1 subclass secretes cytokines such as TNF- α , IL-6, and IL-1 β and plays a significant role in inducing an inflammatory state in adipose tissue, which is important in the development of vascular disorders. Obesity is accompanied by a chronic low-grade inflammatory state, which is confirmed by an elevated level of inflammatory markers, especially C-reactive protein and IL-6, which are significantly higher among obese nonmorbid patients and positively correlates with BMI [76,77].

The excess of carbohydrates, fatty acids, and hyper nutrition induce oxidative stress activation by various pathways such as glycoxidation, oxidative phosphorylation in mitochondria, and NADPH oxidase (NOX) activation with consequent reactive oxygen species (ROS) production [78,79]. The increase in NOX activity leads to excessive production of the superoxide anion (O_2^-), which can react with DNA, lipids, and proteins leading to their destruction [80]. Moreover, O_2^- leads to the alteration of NO activity and consequent endothelial dysfunction and cardiovascular events among obese populations [81]. In addition, an elevated level of ROS induces VSMC proliferation and remodeling, which contribute to hypertension development and increased risk of cardiovascular events [82,83].

Moreover, hypertrophy of adipocytes does not proceed hand in hand with angiogenesis, and the demand of tissues for oxygen is greater than the supply. As a result, hypoxia and consequent necrosis and inflammation occur [84]. Hypoxia-inducible factor (HIF-1 α), which is increased in adipose tissue among obese individuals, plays the role of mediator in hypoxia. HIF-1 α induces the elevation of IL-6 and TNF- α activity and reduces adiponectin concentration [85].

4. Dysregulation of Vascular Function Induced by PVAT Dysfunction in Obesity

PVAT dysfunction, which is a result of an increased inflammatory state among obese patients, induces the dysregulation of vascular function by increased peripheral resistance and vascular tone [86]. In animal models, with diet-induced obesity, loss of the PVAT anticontractile effect was correlated with increased blood pressure [87,88]. The exact mechanism of vascular dysfunction in obesity is still not well clarified; however, there are some pathways such as renin–angiotensin–aldosterone system (RAAS) disorders and PVAT-derived factors' dysregulation, which are involved in the process. Pathological modifications in the synthesis and secretion of ADR and PDRF are responsible for the loss of their anticontractile effect (Figure 2). Endothelial dysfunction plays a crucial role in the pathophysiological process of microvascular and macrovascular complications of obesity [89,90].



Figure 2. The influence of obesity on PVAT-derived factors' synthesis resulting in vascular dysfunction. (PVAT: perivascular adipose tissue, RAAS: renin–angiotensin–aldosterone system, NO: nitric oxide, H₂S: hydrogen sulfide, H₂O₂: hydrogen peroxide, TNF– α : tumor necrosis factor α , and NA: noradrenaline).

4.1. The RAAS in Obesity

The renin–angiotensin–aldosterone system (RAAS) plays a significant role in the regulation of blood pressure. Components of RAAS are produced by the adrenal gland, liver, and adipose tissue. The hypertrophy of adipocytes among obese patients contributes to increased production of angiotensinogen, Ang-II, and aldosterone by PVAT and subcutaneous adipose tissue [91,92]. The concentration of factors mentioned above positively correlates with BMI among obese nonhypertensive subjects [93]. Moreover, the animal model study with diet-induced obese rats revealed the increased production of angiotensinogen by adipose tissue, while the liver expression remained unchanged [94], which shows the significant role of adipose tissue in RAAS factor production. Each component of RAAS plays an important role in the development of dysfunction of microcirculation, hypertension, and arterial stiffness. Increased concentrations of Ang-II and aldosterone induce microvascular constriction via different mechanisms. First of all, chronic activation of angiotensin-II type 1 receptors (AT1R), which suppresses eNOS activity, induces a decrease in NO concentration and bioavailability [95]. Moreover, Ang-II stimulates the synthesis and release of endothelium-dependent vasoconstrictors such as endothelin-1 by increased expression of preproendothelin-1, which is consequently converted to endothelin-1 and COX-1-derived prostanoids by increased expression of COX in human endothelial cells [96,97]. In addition, each component of RAAS, especially aldosterone, by the activation of mineralocorticoid

receptors, induces reabsorption of sodium in distal nephron and consequently increases blood pressure [98]. Ang-II and aldosterone also induce arterial stiffening via enhanced fibrosis, proliferation of VSMCs, and increased collagen deposits [99].

4.2. PVAT-Derived Factors Dysregulation with Effect on Vascular Tone

Among obese patients, the regulatory effect of PVAT on vascular tone is attenuated because of the dysregulation in PVAT-derived factors release. Increased inflammation in adipose tissue, especially IL-6 and TNF α , decreases adiponectin secretion, while the expression of mRNA remains unchanged. Almabrouk et al. reported that concentration of adiponectin was 70% decreased among mice, which were fed a high-fat diet for 12 weeks [100]. Aghamohammadzadeh et al. presented similar findings among humans, while the adiponectin level was significantly lower among obese patients compared to healthy individuals; however, the main limitation of this study was the small study group [101]. Peroxisome proliferator-activated receptor gamma (PPAR- γ) belongs to liganddependent nuclear receptors, which play a significant role in adipocyte differentiation and metabolism [102]. Obesity induces downregulation of PPAR- γ , which decreases the expression of adiponectin [103]. Decreased synthesis and secretion of adiponectin is responsible for the decrease in anticontractile effect.

Leptin synthesis is proportional to adipose size; thus, in obesity, the production of leptin is increased in PVAT and visceral adipose tissue [104]. An increased level of leptin is related to selective insensitivity in appetite and weight regulation and a biphasic effect on vascular function. Even though leptin has an anticontractile effect by inducing NO synthesis, long-lasting exposure of endothelium to leptin induces the inverse result by decreasing the bioavailability of NO [105]. Initially, in diet-induced obesity in animal models, the impairment in leptin-induced NO synthesis and release was compensated by enhanced EDHF-mediated vasodilation. After three months of a high-fat diet, both NO and EDHF-mediated vasodilation effects were reduced and led to an increase in blood pressure [44,48]. In addition, an increased level of leptin decreased the level of L-arginine, resulting in eNOS uncoupling and overproduction of O_2^- , which reacted with NO forming the cytotoxic ONOO-inducing endothelial dysfunction [105]. Increased leptin levels predispose the development of atherosclerosis, by inducing the production of IL-6, IL-12, and TNF α , which plays a significant role in atherogenesis [106]. Moreover, leptin induces the proliferation of vascular and endothelial cells, which also play an important role in atherosclerosis development [107]. In addition, increased blood pressure induced by an enhanced level of leptin is a result of the SNS stimulation. In experimental studies, the external infusion of leptin increased the concentration of norepinephrine and epinephrine dose dependently [107]. Harlan et al. suggested that leptin induces SNS stimulation by activation of the arcuate nucleus in hypothalamus [108].

Geng et al. showed that H_2S bioavailability and synthesis in adipose tissue was decreased in mice on a highfat diet [109]. It is probably a result of the decreased activation of cystathionine γ -lyase (CSE), which is responsible for H_2S production. Moreover, obesity is related to an increased concentration of ROS, which induces H_2S degradation [110]. A decreased concentration of H_2S results in increased systolic and diastolic blood pressure and contributes to the progression of atherosclerosis [111]. H_2O_2 is another vasorelaxant factor, which has the opposite abilities to O_2^- , which induces vasoconstriction. The final effect on vessels depends on the balance between them and the activation of superoxide dismutase (SOD), which is an essential antioxidant enzyme in inactivation of the mentioned factors [25]. The oxidative stress, present among obese individuals, reduces SOD activation, which results in the loss of anticontractile effect and consequent vascular dysfunction by increased concentration of O_2^- and H_2O_2 [40]. It turns out that H_2O_2 is changed into a hydroxyl radical, which induces cell damage.

Decreased NO release is present among obese patients compared to controls [56]. First of all, disturbances in secretion of the PVAT-derived factors mentioned above induces changes in NO synthesis and releasing. Elevated ROS concentration among obese individ-

uals reacts with the NO forming cytotoxic product, which interferes with endothelial cells. Among diet-induced obese mice, reduced expression of eNOS in mesenteric PVAT [87] and impaired eNOS function in thoracic aorta PVAT [49] was observed.

Obesity is connected with increased COX-1 and consequent COX-2 activation, affecting increased production of contractive (TXA2), which contributes to an increased vascular smooth muscles tone [64]. Moreover, due to pro-inflammatory TXA2, the increased concentration of inflammatory markers, ROS, and consequent endothelial dysfunction is present [112,113]. HF diet attenuates prostacyclin secretion, which induces endothelial dysfunction [27]. Moreover, obese subjects present impairment in the prostacyclin ability to increase adenosine 3',5'-cyclic monophosphate (cAMP) and consequent dysregulation in platelets function [114].

5. The Influence of Exercise among Obese on PVAT

Exercise is one of the beneficial nonpharmacological interventions, which is essential in obesity treatment. Physical activity increases the muscles' demands for oxygen and nutrition, resulting in increased blood flow and the vasodilation of muscle vessels. Exercise induces body weight loss, improvement in endothelial function, and reduction in blood pressure [115]. The influence of physical activity on PVAT is multiple. First of all, exercise has anti-inflammatory properties and reduces oxidative stress [116]. It was also shown that exercise prevents or attenuates infiltration of immune cells into PVAT improving vascular function [117]. It contributes to decreasing adipocyte size and consequent reduction in inflammatory markers and pro-inflammatory factors secretion such as TNF α , IL-6, or leptin [118,119]. Physical activity contributes to enhanced adiponectin synthesis by PVAT, inducing an improvement in endothelium-dependent vasodilation and vascular function [120]. Moreover, exercise training is responsible for a reduction in ROS concentration and increased NO bioavailability [121]. In obese rats, physical activity increases the eNOS expression and phosphorylation, which is essential in enzyme activation [120,122]. SNS modulates vascular tone via a double mechanism. Noradrenaline, which is the main SNS mediator, induces the expression of β 3-adrenoreceptors, which are responsible for adiponectin release, while organic cation transporter 3 (OCT3) separates the excess of NA [38]. Saxton et al. reported that obesity contributes to the downregulation of mentioned receptors among mice [123]. However, exercise performed on obese mice turned out to increase the expression of β 3-adrenoreceptors and returned control levels of OCT3. Moreover, stimulation of β 3-adrenoreceptors plays an important role in eNOS activation in PVAT [124]. Additionally, it was postulated that skeletal muscle activity regulates PVAT function through myokines such as FGF21, meteorin-like, irisin, IL-15, and IL-6, acting in a paracrine fashion to antagonize dysfunction of PVAT (i.e., inflammation and dysregulated secretion of adipokines) [117]. DeVallance et al. showed that 8-week aerobic training in rats with metabolic syndrome prevented the increase in oxidant load and inflammation, while enhancing •NO and proteasome function in PVAT, which favorably influenced the function of aortic endothelium [125]. Wang et al. reported that among obese mice, the dominant population of macrophages in the PVAT are M1 subtype, which has proinflammatory properties [126]. Physical activity has an influence on the population of macrophages in PVAT, while 8-week exercise induces reduction in M1 cells with rising of M2 cells among exercised mice, which was not observed among the obese nonexercised group of mice [126]. Moreover, mentioned study confirms the thesis presented by the remaining reports that physical activity has anti-inflammatory properties, while 8-week exercise induces a significant reduction in IL-6 and TNF- α concentration with a consequent significant increase in adiponectin and IL-10 level. Uncoupling protein 1 (UCP1), especially highly expressed in BAT, is responsible for heat production by distracting the proton gradient in mitochondria [127]. Exercise intervention induces upregulation of expression of UCP1 and increases the number of multilocular brown adipocytes in PVAT among obese, exercised mice [126]. Similar findings were reported by Liao et al., who reported

an increase in UCP1 expression in the mesenteric artery PVAT after exercise intervention among rats [128].

6. The Potential Influence of Diet on PVAT-Derived Factors among Obese Patients

Besides exercise, an adequate diet is another beneficial intervention in weight loss and obesity treatment. Nowadays, there are a wide variety of diet strategies, which differ from one another in terms of the percentage content of macronutrients, such as carbohydrates, proteins, and fats. However, a reduction in daily calorie intake is a universal rule and a recommended strategy in weight loss [129]. Some data that present the influence of dietary intervention directly on PVAT are available. Nevertheless, the association between diets and PVAT are not clearly understood. Reports mainly refer to animal models, in which high-carbohydrate (HC) diets induce obesity and the consequent loss of the anticontractile effect of PVAT by an imbalance in PVAT-derived factor secretion [100]. In contrast, Costa et al. have shown that consuming an HC diet for 4 weeks enhanced the release of vasodilatory factors from PVAT, suggesting that this could be a compensatory adaptive characteristic in order to preserve the vascular function during the initial stages of obesity [130]. Additionally, it is suggested that imbalanced diets can cause PVAT inflammation and dysfunction as well as impaired vascular function. The recent published study has showed that a high-fat (HF) and a high-sucrose (HS) diet affected PVAT at different sites. Sasoh et al. have presented characteristic differences in the effects of HF and HS diets on PVAT and aortae [131]. A HF diet induced an increased number of large-sized lipid droplets and increased cluster of differentiation (CD) 68+ macrophage- and monocyte chemotactic protein (MCP)-1-positive areas in the abdominal aortic PVAT (aPVAT). Furthermore, a HF diet caused a decreased collagen fiber-positive area and increased CD68+ macrophage- and MCP-1-positive areas in the abdominal aorta. In contrast, a HS diet induced an increased number of large-sized lipid droplets, increased CD68+ macrophageand MCP-1-positive areas, and decreased UCP-1 positive area in the thoracic aortic PVAT (tPVAT). Moreover, a HS diet caused a decreased collagen fiber-positive area and increased CD68+ macrophage- and MCP-1-positive areas in the thoracic aorta. However, there were some factors that did not follow the trend to this variation. For example, angiotensinogen levels were increased in both tPVAT and aPVAT of the HF group. The authors concluded that the potential mechanisms underlying these effects may be related to the different adipocyte species that comprise tPVAT and aPVAT [131]. Victorio et al. reported that the effect of HF and HS diets on PVAT differs depending on sex [132]. The anti-contractile effect of PVAT was measured by comparing the phenylephrine-induced contraction in mesenteric arteries after 3 and 5 months of HF or HF+HS diet among male and female mice. The results showed that anticontractile function was impaired after 3 months of both obesogenic diets among females, while among males, the anti-contractile effect remained comparable during the experiment. Moreover, the assessment of PVAT-derived endothelial function after acetylcholine administration likewise demonstrated differences between sexes, while obesogenic diet among females induces endothelial dysfunction after 3 months and only after 5 months among males.

However, there are many reports that relate the positive impact of different diets on inflammatory state, oxidative stress, NO, adiponectin, or leptin concentration. Thus, one could conclude that similar changes could be observed in PVAT; however, further studies should be conducted.

The Mediterranean diet (MD) is the most popular diet and is commonly known as a healthy, balanced diet with proven efficiency in reducing the cardiovascular risk among high-risk patients and reducing overall mortality [133,134]. A typical MD contains 55–60% carbohydrates, mainly complex ones, 25–30% polyunsaturated and monounsaturated fats, and 15–20% proteins, and meals are generally based on fish, nuts, olive oil, and plant-based foods [135]. Luisi et al. reported that the implementation of an MD for 3 months among overweight/obese patients, with high-quality extra virgin olive oil, induced weight loss and the significant elevation of adiponectin levels [136]. Interestingly, among normal

weight controls, the MD has no impact on weight, and the increase in adiponectin concentration was not as considerable as that found among overweight/obese patients. It can be concluded that weight loss and the consequent reduction in adipose tissue contribute to a size reduction in adipocytes and an improvement in adiponectin synthesis and release. Among both groups, the concentration of IL-6 significantly decreased after dietary intervention, which provides proof of the anti-inflammatory properties of MD [136]. Moreover, it is suggested that the higher the amount of fiber in one's diet, the greater the adiponectin concentration in one's blood [137].

Recently, the ketogenic diet has become very popular due to its therapeutic properties in relation to different diseases. It has been widely used in drug-resistant epilepsy with good outcomes and is increasingly being used in metabolic disorders such as obesity or diabetes mellitus [138,139]. The ketogenic diet is characterized by low carbohydrates and high fat, inducing changes in the metabolism of energy substrates, with a switch from glucose to fatty acids [138]. A very low-calorie ketogenic diet (VLCKD) is a special type of caloric reduction diet characterized by a very low or extremely low daily food energy intake, circa 800 kcal per day [140]. It provides 30–50 g of carbohydrates, about 30-40 g of fats, and 0.8-1.5 g/kg of ideal body weight (IBW) of proteins [141]. Monda et al. reported that obese patients who consumed a VLCKD diet for 8 weeks presented with a significant body mass reduction, a decreased concentration of inflammatory markers such as IL-6, TNF- α , and CRP, and a significant elevation in the level of adiponectin in their blood [140]. This relatively short period of intervention induced a significant multifactorial improvement; however, the main limitation of the aforementioned study is the small sample size. In other reports, the ketogenic diet has also been proven to have anti-inflammatory properties [142,143].

A low-calorie diet is a balanced diet with a 20–30% reduction in one's daily calorie intake, differing from the ketogenic diet by its macronutrient content, and consisting of 45–55% carbohydrates, 25–35% fat, and 15–25% proteins, with an additional 30 g of fiber [144]. The beneficial effects of a calorie reduction among humans include weight loss, a reduction in superoxide and inflammatory factor production, and the upregulation of the activity of eNOS [145,146]. Vink et al. found that 12 weeks of a low-calorie diet among obese patients resulted in a significant reduction in leptin and the elevation of the adiponectin concentration [147]. Another report among overweight patients confirmed a reduction in body weight and leptin levels after 6 months of a low-calorie diet [148]. It is reported that the supplementation of melatonin in animal models, which is known for its anti-inflammatory and anti-oxidative properties, could be used in the prevention of obesity [149,150]. Szewczyk-Golec et al. reported that 30 days of daily oral administration of 10 g of melatonin, accompanied by a low-calorie diet, induced a statistically significant reduction in body mass weight and led to the elevation of the adiponectin concentration among obese patients, which was not observed among the control group, who received a low-calorie diet alone [151].

Intermittent fasting (IF) is the type of diet that is based on taking intermittent breaks from eating. There are two types of IF: alternative day fasting and time-restricted fasting. The first type includes 24 h of fasting followed by a 24-h period of eating with mixing fast days with nonrestricted days during a week, while time-restricted fasting consists of different variations, f.e., 16 h of fasting with 8 h of eating [152]. IF is reported to have a positive impact on adipose tissue function especially among obese individuals. Liu et al. reported that eight weeks of IF among high-fat diet (HFD)-fed mice reduced adipocytes hypertrophy and concentration of inflammatory markers by bodyweight reduction and improvement in insulin sensitivity [153]. Moreover, IF is more effective in reducing inflammatory markers concentration than a calorie-restriction diet among animal models [154]. Interestingly, the comparison between IF and a calorie-restricted diet among humans showed an intermittent increase in M1 markers of inflammation, which was a result of lipolysis and consequent increase in non-esterified fatty acids serum concentration after IF [155]. On the other hand, IF in mice was reported to induce activation of macrophages

with M2 subtype polarization [156]. The M2 macrophages, which are known in the literature for their anti-inflammatory properties, induce the production of IL-10, phagocyte apoptotic cells, and promote wound healing [157,158]. Moreover, fasting-mediated activation of AMPK is reported among animal models [159] and seems to play a significant role in mitochondrial homeostasis [160]. In addition, AMPK, opposite to mTOR, turned out to activate autophagy, which seems to play a significant role in maintaining autophagic homeostasis [161]. Permanent overnutrition induces suppression of autophagy and the accumulation of impaired cellular components, such as mitochondria, in metabolic tissues, which contributes to metabolic dysfunction and consequent development of metabolic disorders such as obesity [162].

7. Conclusions

PVAT, as a special adipose tissue, is not only a mechanical support for the vascular system but plays a vital role in the homeostasis of the vascular system, sharing a status no less important than that of endothelium. Obesity plays an important role in the development of vascular dysfunction, dysregulation of vascular tone, and endothelial dysfunction. The pathogenesis of obesity contains hypoxia, increased oxidative stress, and enhanced inflammatory factors. All these components induce PVAT dysfunction, dysregulation in the synthesis of PVAT-derived factors, decreased bioavailability of NO, an increased inflammatory state in PVAT, and increased activation of RAAS. Exercise training, commonly known as an essential nonpharmacological intervention in obesity treatment, contributes to an improvement in PVAT activity, which could have a positive effect on vascular tone. Diet, which is the second most beneficial non-invasive strategy in obesity treatment, may have a positive impact on PVAT-derived factors and may restore the balance in their concentration. Independent of the type of diet, a decrease in body mass weight, which is connected with a reduction in adipose tissue, may restore the balance of the synthesis and release of adipokines. However, further studies should be conducted in order to demonstrate the exact influence of diet on PVAT among humans. Moreover, additional studies are also needed to help researchers better understand the pathophysiology of PVAT and evaluate whether targeting PVAT function could be used as a novel approach for the treatment of cardiovascular diseases.

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References

- Global Burden of Disease Study 2015 (GBD 2015) Obesity and Overweight Prevalence 1980–2015 | GHDx. Available online: http://ghdx.healthdata.org/record/ihme-data/gbd-2015-obesity-and-overweight-prevalence-1980-2015 (accessed on 3 August 2021).
- 2. Hales, C.M. Prevalence of Obesity and Severe Obesity Among Adults: United States, 2017–2018. NCHS Data Brief 2020, 360, 8.
- 3. Overweight and Obesity-BMI Statistics. Available online: https://ec.europa.eu/eurostat/statistics-explained/index.php?title= Overweight_and_obesity_-_BMI_statistics (accessed on 6 September 2021).
- Dai, H.; Alsalhe, T.A.; Chalghaf, N.; Riccò, M.; Bragazzi, N.L.; Wu, J. The Global Burden of Disease Attributable to High Body Mass Index in 195 Countries and Territories, 1990–2017: An Analysis of the Global Burden of Disease Study. *PLoS Med.* 2020, 17, e1003198. [CrossRef] [PubMed]

- Singh, G.M.; Danaei, G.; Farzadfar, F.; Stevens, G.A.; Woodward, M.; Wormser, D.; Kaptoge, S.; Whitlock, G.; Qiao, Q.; Lewington, S.; et al. The Age-Specific Quantitative Effects of Metabolic Risk Factors on Cardiovascular Diseases and Diabetes: A Pooled Analysis. *PLoS ONE* 2013, 8, e65174. [CrossRef] [PubMed]
- 6. Kolotkin, R.L.; Andersen, J.R. A Systematic Review of Reviews: Exploring the Relationship between Obesity, Weight Loss and Health-related Quality of Life. *Clin. Obes.* 2017, *7*, 273–289. [CrossRef]
- 7. Tremmel, M.; Gerdtham, U.-G.; Nilsson, P.M.; Saha, S. Economic Burden of Obesity: A Systematic Literature Review. *Int. J. Environ. Res. Public Health* **2017**, *14*, 435. [CrossRef] [PubMed]
- Fernández-Alfonso, M.S.; Gil-Ortega, M.; García-Prieto, C.F.; Aranguez, I.; Ruiz-Gayo, M.; Somoza, B. Mechanisms of Perivascular Adipose Tissue Dysfunction in Obesity. *Int. J. Endocrinol.* 2013, 2013, 402053. [CrossRef]
- Petersen, M.C.; Shulman, G.I. Mechanisms of Insulin Action and Insulin Resistance. *Physiol. Rev.* 2018, 98, 2133–2223. [CrossRef] [PubMed]
- Saely, C.H.; Geiger, K.; Drexel, H. Brown versus White Adipose Tissue: A Mini-Review. Gerontology 2012, 58, 15–23. [CrossRef] [PubMed]
- 11. Kiefer, F.W.; Cohen, P.; Plutzky, J. Fifty Shades of Brown: Perivascular Fat, Thermogenesis, and Atherosclerosis. *Circulation* **2012**, *126*, 1012–1015. [CrossRef]
- Police, S.B.; Thatcher, S.E.; Charnigo, R.; Daugherty, A.; Cassis, L.A. Obesity Promotes Inflammation in Periaortic Adipose Tissue and Angiotensin II-Induced Abdominal Aortic Aneurysm Formation. *Arter. Thromb. Vasc. Biol.* 2009, 29, 1458–1464. [CrossRef]
- 13. Verlohren, S.; Dubrovska, G.; Tsang, S.-Y.; Essin, K.; Luft, F.C.; Huang, Y.; Gollasch, M. Visceral Periadventitial Adipose Tissue Regulates Arterial Tone of Mesenteric Arteries. *Hypertension* **2004**, *44*, 271–276. [CrossRef]
- 14. Yudkin, J.S.; Eringa, E.; Stehouwer, C.D.A. "Vasocrine" Signalling from Perivascular Fat: A Mechanism Linking Insulin Resistance to Vascular Disease. *Lancet* 2005, *365*, 1817–1820. [CrossRef]
- 15. Gao, Y.-J. Dual Modulation of Vascular Function by Perivascular Adipose Tissue and Its Potential Correlation with Adiposity/Lipoatrophy-Related Vascular Dysfunction. *Curr. Pharm. Des.* **2007**, *13*, 2185–2192. [CrossRef]
- 16. Takaoka, M.; Suzuki, H.; Shioda, S.; Sekikawa, K.; Saito, Y.; Nagai, R.; Sata, M. Endovascular Injury Induces Rapid Phenotypic Changes in Perivascular Adipose Tissue. *Arter. Thromb. Vasc. Biol.* **2010**, *30*, 1576–1582. [CrossRef]
- 17. Hu, Y.; Zhang, Z.; Torsney, E.; Afzal, A.R.; Davison, F.; Metzler, B.; Xu, Q. Abundant Progenitor Cells in the Adventitia Contribute to Atherosclerosis of Vein Grafts in ApoE-Deficient Mice. *J. Clin. Investig.* **2004**, *113*, 1258–1265. [CrossRef] [PubMed]
- Qi, X.-Y.; Qu, S.-L.; Xiong, W.-H.; Rom, O.; Chang, L.; Jiang, Z.-S. Perivascular Adipose Tissue (PVAT) in Atherosclerosis: A Double-Edged Sword. *Cardiovasc. Diabetol.* 2018, 17, 134. [CrossRef]
- 19. Gil-Ortega, M.; Somoza, B.; Huang, Y.; Gollasch, M.; Fernández-Alfonso, M.S. Regional Differences in Perivascular Adipose Tissue Impacting Vascular Homeostasis. *Trends Endocrinol. Metab.* **2015**, *26*, 367–375. [CrossRef] [PubMed]
- Lasar, D.; Julius, A.; Fromme, T.; Klingenspor, M. Browning Attenuates Murine White Adipose Tissue Expansion during Postnatal Development. *Biochim. Biophys Acta* 2013, 1831, 960–968. [CrossRef] [PubMed]
- Lu, D.; Wang, W.; Xia, L.; Xia, P.; Yan, Y. Gene Expression Profiling Reveals Heterogeneity of Perivascular Adipose Tissues Surrounding Coronary and Internal Thoracic Arteries. *Acta Biochim. Biophys. Sin.* 2017, 49, 1075–1082. [CrossRef]
- Britton, K.A.; Pedley, A.; Massaro, J.M.; Corsini, E.M.; Murabito, J.M.; Hoffmann, U.; Fox, C.S. Prevalence, Distribution, and Risk Factor Correlates of High Thoracic Periaortic Fat in the Framingham Heart Study. J. Am. Heart Assoc. 2012, 1, e004200. [CrossRef]
- El Khoudary, S.R.; Shields, K.J.; Janssen, I.; Hanley, C.; Budoff, M.J.; Barinas-Mitchell, E.; Everson-Rose, S.A.; Powell, L.H.; Matthews, K.A. Cardiovascular Fat, Menopause, and Sex Hormones in Women: The SWAN Cardiovascular Fat Ancillary Study. J. Clin. Endocrinol. Metab. 2015, 100, 3304–3312. [CrossRef]
- 24. Victorio, J.A.; da Costa, R.M.; Tostes, R.C.; Davel, A.P. Modulation of Vascular Function by Perivascular Adipose Tissue: Sex Differences. *Curr. Pharm. Des.* **2020**, *26*, 3768–3777. [CrossRef]
- Marchesi, C.; Ebrahimian, T.; Angulo, O.; Paradis, P.; Schiffrin, E.L. Endothelial Nitric Oxide Synthase Uncoupling and Perivascular Adipose Oxidative Stress and Inflammation Contribute to Vascular Dysfunction in a Rodent Model of Metabolic Syndrome. *Hypertension* 2009, 54, 1384–1392. [CrossRef]
- 26. Sowka, A.; Dobrzyn, P. Role of Perivascular Adipose Tissue-Derived Adiponectin in Vascular Homeostasis. *Cells* **2021**, *10*, 1485. [CrossRef] [PubMed]
- Chang, L.; Villacorta, L.; Li, R.; Hamblin, M.; Xu, W.; Dou, C.; Zhang, J.; Wu, J.; Zeng, R.; Chen, Y.E. Loss of Perivascular Adipose Tissue on Peroxisome Proliferator-Activated Receptor-γ Deletion in Smooth Muscle Cells Impairs Intravascular Thermoregulation and Enhances Atherosclerosis. *Circulation* 2012, *126*, 1067–1078. [CrossRef] [PubMed]
- 28. Ozen, G.; Daci, A.; Norel, X.; Topal, G. Human perivascular adipose tissue dysfunction as a cause of vascular disease: Focus on vascular tone and wall remodeling. *Eur. J. Pharm.* 2015, 766, 16–24. [CrossRef] [PubMed]
- 29. Miao, C.-Y.; Li, Z.-Y. The Role of Perivascular Adipose Tissue in Vascular Smooth Muscle Cell Growth. *Br. J. Pharm.* **2012**, *165*, 643–658. [CrossRef] [PubMed]
- 30. Gollasch, M. Vasodilator Signals from Perivascular Adipose Tissue. Br. J. Pharmacol. 2012, 165, 633. [CrossRef]
- 31. Szasz, T.; Webb, R.C. Perivascular Adipose Tissue: More than Just Structural Support. *Clin. Sci.* 2012, 122, 1–12. [CrossRef]
- Gálvez-Prieto, B.; Somoza, B.; Gil-Ortega, M.; García-Prieto, C.F.; de las Heras, A.I.; González, M.C.; Arribas, S.; Aranguez, I.; Bolbrinker, J.; Kreutz, R.; et al. Anticontractile Effect of Perivascular Adipose Tissue and Leptin Are Reduced in Hypertension. *Front. Pharm.* 2012, 3, 103. [CrossRef]

- Lee, Y.-C.; Chang, H.-H.; Chiang, C.-L.; Liu, C.-H.; Yeh, J.-I.; Chen, M.-F.; Chen, P.-Y.; Kuo, J.-S.; Lee, T.J.F. Role of Perivascular Adipose Tissue-Derived Methyl Palmitate in Vascular Tone Regulation and Pathogenesis of Hypertension. *Circulation* 2011, 124, 1160–1171. [CrossRef]
- Gálvez-Prieto, B.; Bolbrinker, J.; Stucchi, P.; de Las Heras, A.I.; Merino, B.; Arribas, S.; Ruiz-Gayo, M.; Huber, M.; Wehland, M.; Kreutz, R.; et al. Comparative Expression Analysis of the Renin-Angiotensin System Components between White and Brown Perivascular Adipose Tissue. J. Endocrinol. 2008, 197, 55–64. [CrossRef] [PubMed]
- 35. Gao, Y.; Takemori, K.; Su, L.; An, W.; Lu, C.; Sharma, A.; Lee, R. Perivascular Adipose Tissue Promotes Vasoconstriction: The Role of Superoxide Anion. *Cardiovasc. Res.* **2006**, *71*, 363–373. [CrossRef]
- Cheng, C.K.; Bakar, H.A.; Gollasch, M.; Huang, Y. Perivascular Adipose Tissue: The Sixth Man of the Cardiovascular System. *Cardiovasc. Drugs* 2018, 32, 481–502. [CrossRef]
- Weston, A.H.; Egner, I.; Dong, Y.; Porter, E.L.; Heagerty, A.M.; Edwards, G. Stimulated Release of a Hyperpolarizing Factor (ADHF) from Mesenteric Artery Perivascular Adipose Tissue: Involvement of Myocyte BKCa Channels and Adiponectin. *Br. J. Pharmacol.* 2013, *169*, 1500–1509. [CrossRef]
- Saxton, S.N.; Ryding, K.E.; Aldous, R.G.; Withers, S.B.; Ohanian, J.; Heagerty, A.M. Role of Sympathetic Nerves and Adipocyte Catecholamine Uptake in the Vasorelaxant Function of Perivascular Adipose Tissue. *Arterioscler. Thromb. Vasc. Biol.* 2018, 38, 880–891. [CrossRef] [PubMed]
- 39. Chen, H.; Montagnani, M.; Funahashi, T.; Shimomura, I.; Quon, M.J. Adiponectin Stimulates Production of Nitric Oxide in Vascular Endothelial Cells. J. Biol. Chem. 2003, 278, 45021–45026. [CrossRef]
- 40. Steinberg, G.R.; Kemp, B.E. AMPK in Health and Disease. Physiol. Rev. 2009, 89, 1025–1078. [CrossRef] [PubMed]
- 41. Margaritis, M.; Antonopoulos, A.S.; Digby, J.; Lee, R.; Reilly, S.; Coutinho, P.; Shirodaria, C.; Sayeed, R.; Petrou, M.; De Silva, R.; et al. Interactions between Vascular Wall and Perivascular Adipose Tissue Reveal Novel Roles for Adiponectin in the Regulation of Endothelial Nitric Oxide Synthase Function in Human Vessels. *Circulation* **2013**, *127*, 2209–2221. [CrossRef]
- 42. Föller, M.; Jaumann, M.; Dettling, J.; Saxena, A.; Pakladok, T.; Munoz, C.; Ruth, P.; Sopjani, M.; Seebohm, G.; Rüttiger, L.; et al. AMP-Activated Protein Kinase in BK-Channel Regulation and Protection against Hearing Loss Following Acoustic Overstimulation. *FASEB J.* **2012**, *26*, 4243–4253. [CrossRef]
- 43. Yanai, H.; Yoshida, H. Beneficial Effects of Adiponectin on Glucose and Lipid Metabolism and Atherosclerotic Progression: Mechanisms and Perspectives. *Int. J. Mol. Sci* 2019, *20*, 1190. [CrossRef]
- 44. Christou, G.A.; Kiortsis, D.N. Adiponectin and Lipoprotein Metabolism. Obes. Rev. 2013, 14, 939–949. [CrossRef]
- 45. Rahmouni, K.; Correia, M.L.G.; Haynes, W.G.; Mark, A.L. Obesity-Associated Hypertension. *Hypertension* **2005**, *45*, 9–14. [CrossRef] [PubMed]
- Fain, J.N.; Madan, A.K.; Hiler, M.L.; Cheema, P.; Bahouth, S.W. Comparison of the Release of Adipokines by Adipose Tissue, Adipose Tissue Matrix, and Adipocytes from Visceral and Subcutaneous Abdominal Adipose Tissues of Obese Humans. *Endocrinology* 2004, 145, 2273–2282. [CrossRef]
- 47. Bełtowski, J. Leptin and the Regulation of Endothelial Function in Physiological and Pathological Conditions. *Clin. Exp. Pharmacol. Physiol.* **2012**, *39*, 168–178. [CrossRef] [PubMed]
- Jamroz-Wiśniewska, A.; Gertler, A.; Solomon, G.; Wood, M.E.; Whiteman, M.; Bełtowski, J. Leptin-Induced Endothelium-Dependent Vasorelaxation of Peripheral Arteries in Lean and Obese Rats: Role of Nitric Oxide and Hydrogen Sulfide. *PLoS ONE* 2014, 9, e86744. [CrossRef] [PubMed]
- 49. Quehenberger, P.; Exner, M.; Sunder-Plassmann, R.; Ruzicka, K.; Bieglmayer, C.; Endler, G.; Muellner, C.; Speiser, W.; Wagner, O. Leptin Induces Endothelin-1 in Endothelial Cells in Vitro. *Circ. Res.* **2002**, *90*, 711–718. [CrossRef]
- Tilg, H.; Moschen, A.R. Adipocytokines: Mediators Linking Adipose Tissue, Inflammation and Immunity. *Nat. Rev. Immunol.* 2006, *6*, 772–783. [CrossRef]
- 51. Ardanaz, N.; Pagano, P.J. Hydrogen Peroxide as a Paracrine Vascular Mediator: Regulation and Signaling Leading to Dysfunction. *Exp. Biol. Med.* **2006**, 231, 237–251. [CrossRef]
- 52. Montfort, W.R.; Wales, J.A.; Weichsel, A. Structure and Activation of Soluble Guanylyl Cyclase, the Nitric Oxide Sensor. *Antioxid. Redox Signal.* **2017**, *26*, 107–121. [CrossRef]
- 53. Wang, R. Shared Signaling Pathways among Gasotransmitters. *Proc. Natl. Acad. Sci. USA* 2012, 109, 8801–8802. [CrossRef] [PubMed]
- 54. Lee, S.W.; Cheng, Y.; Moore, P.K.; Bian, J.-S. Hydrogen Sulphide Regulates Intracellular PH in Vascular Smooth Muscle Cells. *Biochem. Biophys. Res. Commun* 2007, 358, 1142–1147. [CrossRef]
- Zhong, X.; Wang, Y.; Wu, J.; Sun, A.; Yang, F.; Zheng, D.; Li, T.; Dong, S.; Zhao, Y.; Yang, G.; et al. Calcium Sensing Receptor Regulating Smooth Muscle Cells Proliferation through Initiating Cystathionine-Gamma-Lyase/Hydrogen Sulfide Pathway in Diabetic Rat. *Cell Physiol. Biochem.* 2015, 35, 1582–1598. [CrossRef]
- 56. Xia, N.; Li, H. The Role of Perivascular Adipose Tissue in Obesity-induced Vascular Dysfunction. *Br. J. Pharm.* **2017**, 174, 3425–3442. [CrossRef] [PubMed]
- 57. Lee, R.M.K.W.; Lu, C.; Su, L.-Y.; Gao, Y.-J. Endothelium-Dependent Relaxation Factor Released by Perivascular Adipose Tissue. *J. Hypertens* **2009**, *27*, 782–790. [CrossRef] [PubMed]
- 58. Kusters, P.J.H.; Lutgens, E.; Seijkens, T.T.P. Exploring Immune Checkpoints as Potential Therapeutic Targets in Atherosclerosis. *Cardiovasc. Res.* **2018**, *114*, 368–377. [CrossRef] [PubMed]

- 59. Bredt, D.S.; Snyder, S.H. Isolation of Nitric Oxide Synthetase, a Calmodulin-Requiring Enzyme. *Proc. Natl. Acad. Sci. USA* **1990**, 87, 682–685. [CrossRef] [PubMed]
- 60. Förstermann, U.; Closs, E.I.; Pollock, J.S.; Nakane, M.; Schwarz, P.; Gath, I.; Kleinert, H. Nitric Oxide Synthase Isozymes. Characterization, Purification, Molecular Cloning, and Functions. *Hypertension* **1994**, *23 Pt* 2, 1121–1131. [CrossRef]
- 61. Hong, F.; Liang, X.; Liu, W.; Lv, S.; He, S.; Kuang, H.; Yang, S. Roles of ENOS in Atherosclerosis Treatment. *Inflamm. Res.* 2019, 68, 429–441. [CrossRef]
- 62. Xia, N.; Horke, S.; Habermeier, A.; Closs, E.I.; Reifenberg, G.; Gericke, A.; Mikhed, Y.; Münzel, T.; Daiber, A.; Förstermann, U.; et al. Uncoupling of Endothelial Nitric Oxide Synthase in Perivascular Adipose Tissue of Diet-Induced Obese Mice. *Arterioscler. Thromb. Vasc. Biol.* **2016**, *36*, 78–85. [CrossRef]
- 63. Félétou, M.; Vanhoutte, P.M. Endothelial Dysfunction: A Multifaceted Disorder (The Wiggers Award Lecture). *Am. J. Physiol. Heart Circ. Physiol.* **2006**, 291, H985–H1002. [CrossRef] [PubMed]
- 64. Meyer, M.R.; Fredette, N.C.; Barton, M.; Prossnitz, E.R. Regulation of Vascular Smooth Muscle Tone by Adipose-Derived Contracting Factor. *PLoS ONE* **2013**, *8*, e79245. [CrossRef] [PubMed]
- 65. Ahmad, A.A.; Randall, M.D.; Roberts, R.E. Sex Differences in the Role of Phospholipase A2-dependent Arachidonic Acid Pathway in the Perivascular Adipose Tissue Function in Pigs. J. Physiol. 2017, 595, 6623–6634. [CrossRef]
- 66. Egan, K.M.; Lawson, J.A.; Fries, S.; Koller, B.; Rader, D.J.; Smyth, E.M.; Fitzgerald, G.A. COX-2-Derived Prostacyclin Confers Atheroprotection on Female Mice. *Science* **2004**, *306*, 1954–1957. [CrossRef]
- 67. Mendizábal, Y.; Llorens, S.; Nava, E. Vasoactive Effects of Prostaglandins from the Perivascular Fat of Mesenteric Resistance Arteries in WKY and SHROB Rats. *Life Sci.* 2013, 93, 1023–1032. [CrossRef]
- Ayala-Lopez, N.; Watts, S.W. New Actions of an Old Friend: Perivascular Adipose Tissue's Adrenergic Mechanisms. *Br. J. Pharm.* 2017, 174, 3454–3465. [CrossRef] [PubMed]
- 69. Ayala-Lopez, N.; Martini, M.; Jackson, W.F.; Darios, E.; Burnett, R.; Seitz, B.; Fink, G.D.; Watts, S.W. Perivascular Adipose Tissue Contains Functional Catecholamines. *Pharm. Res. Perspect.* **2014**, *2*, e00041. [CrossRef]
- Vargovic, P.; Ukropec, J.; Laukova, M.; Cleary, S.; Manz, B.; Pacak, K.; Kvetnansky, R. Adipocytes as a New Source of Catecholamine Production. *FEBS Lett.* 2011, 585, 2279–2284. [CrossRef] [PubMed]
- Costa, R.M.; Filgueira, F.P.; Tostes, R.C.; Carvalho, M.H.C.; Akamine, E.H.; Lobato, N.S. H2O2 Generated from Mitochondrial Electron Transport Chain in Thoracic Perivascular Adipose Tissue Is Crucial for Modulation of Vascular Smooth Muscle Contraction. *Vasc. Pharm.* 2016, *84*, 28–37. [CrossRef] [PubMed]
- 72. Cotillard, A.; Poitou, C.; Torcivia, A.; Bouillot, J.-L.; Dietrich, A.; Klöting, N.; Grégoire, C.; Lolmede, K.; Blüher, M.; Clément, K. Adipocyte Size Threshold Matters: Link with Risk of Type 2 Diabetes and Improved Insulin Resistance After Gastric Bypass. J. Clin. Endocrinol. Metab. 2014, 99, E1466–E1470. [CrossRef]
- 73. Chandalia, M.; Lin, P.; Seenivasan, T.; Livingston, E.H.; Snell, P.G.; Grundy, S.M.; Abate, N. Insulin Resistance and Body Fat Distribution in South Asian Men Compared to Caucasian Men. *PLoS ONE* **2007**, *2*, e812. [CrossRef] [PubMed]
- 74. Skurk, T.; Alberti-Huber, C.; Herder, C.; Hauner, H. Relationship between Adipocyte Size and Adipokine Expression and Secretion. *J. Clin. Endocrinol. Metab.* 2007, *92*, 1023–1033. [CrossRef] [PubMed]
- 75. Martinez, F.O.; Gordon, S. The M1 and M2 Paradigm of Macrophage Activation: Time for Reassessment. *F1000Prime Rep.* **2014**, *6*, 13. [CrossRef]
- Sudhakar, M.; Silambanan, S.; Chandran, A.S.; Prabhakaran, A.A.; Ramakrishnan, R. C-Reactive Protein (CRP) and Leptin Receptor in Obesity: Binding of Monomeric CRP to Leptin Receptor. *Front. Immunol.* 2018, *9*, 1167. [CrossRef]
- 77. Gnacińska, M.; Małgorzewicz, S.; Guzek, M.; Lysiak-Szydłowska, W.; Sworczak, K. Adipose Tissue Activity in Relation to Overweight or Obesity. *Endokrynol. Pol.* **2010**, *61*, 160–168.
- Savini, I.; Catani, M.V.; Evangelista, D.; Gasperi, V.; Avigliano, L. Obesity-Associated Oxidative Stress: Strategies Finalized to Improve Redox State. *Int. J. Mol. Sci.* 2013, 14, 10497–10538. [CrossRef]
- Codoñer-Franch, P.; Tavárez-Alonso, S.; Murria-Estal, R.; Tortajada-Girbés, M.; Simó-Jordá, R.; Alonso-Iglesias, E. Elevated Advanced Oxidation Protein Products (AOPPs) Indicate Metabolic Risk in Severely Obese Children. *Nutr. Metab. Cardiovasc. Dis.* 2012, 22, 237–243. [CrossRef]
- 80. Jiang, F.; Lim, H.K.; Morris, M.J.; Prior, L.; Velkoska, E.; Wu, X.; Dusting, G.J. Systemic Upregulation of NADPH Oxidase in Diet-Induced Obesity in Rats. *Redox Rep.* 2011, *16*, 223–229. [CrossRef]
- 81. Beckman, J.S.; Koppenol, W.H. Nitric Oxide, Superoxide, and Peroxynitrite: The Good, the Bad, and the Ugly. *Am. J. Physiol.-Cell Physiol.* **1996**, 271, C1424–C1437. [CrossRef] [PubMed]
- 82. Rani, V.; Deep, G.; Singh, R.K.; Palle, K.; Yadav, U.C.S. Oxidative Stress and Metabolic Disorders: Pathogenesis and Therapeutic Strategies. *Life Sci.* **2016**, *148*, 183–193. [CrossRef]
- 83. Mulvany, M.J. Small Artery Remodelling in Hypertension. Basic Clin. Pharm. Toxicol. 2012, 110, 49-55. [CrossRef] [PubMed]
- 84. Goossens, G.H.; Bizzarri, A.; Venteclef, N.; Essers, Y.; Cleutjens, J.P.; Konings, E.; Jocken, J.W.E.; Cajlakovic, M.; Ribitsch, V.; Clément, K.; et al. Increased Adipose Tissue Oxygen Tension in Obese Compared with Lean Men Is Accompanied by Insulin Resistance, Impaired Adipose Tissue Capillarization, and Inflammation. *Circulation* 2011, 124, 67–76. [CrossRef]
- Cancello, R.; Henegar, C.; Viguerie, N.; Taleb, S.; Poitou, C.; Rouault, C.; Coupaye, M.; Pelloux, V.; Hugol, D.; Bouillot, J.-L.; et al. Reduction of Macrophage Infiltration and Chemoattractant Gene Expression Changes in White Adipose Tissue of Morbidly Obese Subjects after Surgery-Induced Weight Loss. *Diabetes* 2005, 54, 2277–2286. [CrossRef] [PubMed]

- 86. Karaca, Ü.; Schram, M.T.; Houben, A.J.H.M.; Muris, D.M.J.; Stehouwer, C.D.A. Microvascular Dysfunction as a Link between Obesity, Insulin Resistance and Hypertension. *Diabetes Res. Clin. Pract.* **2014**, *103*, 382–387. [CrossRef]
- 87. Bussey, C.E.; Withers, S.B.; Aldous, R.G.; Edwards, G.; Heagerty, A.M. Obesity-Related Perivascular Adipose Tissue Damage Is Reversed by Sustained Weight Loss in the Rat. *Arterioscler. Thromb. Vasc. Biol.* **2016**, *36*, 1377–1385. [CrossRef] [PubMed]
- Withers, S.B.; Forman, R.; Meza-Perez, S.; Sorobetea, D.; Sitnik, K.; Hopwood, T.; Lawrence, C.B.; Agace, W.W.; Else, K.J.; Heagerty, A.M.; et al. Eosinophils Are Key Regulators of Perivascular Adipose Tissue and Vascular Functionality. *Sci. Rep.* 2017, 7, 44571. [CrossRef]
- 89. Koliaki, C.; Liatis, S.; Kokkinos, A. Obesity and Cardiovascular Disease: Revisiting an Old Relationship. *Metabolism* **2019**, *92*, 98–107. [CrossRef]
- 90. Stanek, A.; Fazeli, B.; Bartuś, S.; Sutkowska, E. The Role of Endothelium in Physiological and Pathological States: New Data. *BioMed Res. Int.* 2018, 2018, e1098039. [CrossRef]
- 91. Harte, A.; McTernan, P.; Chetty, R.; Coppack, S.; Katz, J.; Smith, S.; Kumar, S. Insulin-Mediated Upregulation of the Renin Angiotensin System in Human Subcutaneous Adipocytes Is Reduced by Rosiglitazone. *Circulation* 2005, *111*, 1954–1961. [CrossRef]
- Briones, A.M.; Nguyen Dinh Cat, A.; Callera, G.E.; Yogi, A.; Burger, D.; He, Y.; Corrêa, J.W.; Gagnon, A.M.; Gomez-Sanchez, C.E.; Gomez-Sanchez, E.P.; et al. Adipocytes Produce Aldosterone Through Calcineurin-Dependent Signaling Pathways. *Hypertension* 2012, 59, 1069–1078. [CrossRef] [PubMed]
- Rossi, G.P.; Belfiore, A.; Bernini, G.; Fabris, B.; Caridi, G.; Ferri, C.; Giacchetti, G.; Letizia, C.; Maccario, M.; Mannelli, M.; et al. Body Mass Index Predicts Plasma Aldosterone Concentrations in Overweight-Obese Primary Hypertensive Patients. *J. Clin. Endocrinol. Metab.* 2008, 93, 2566–2571. [CrossRef]
- Boustany, C.M.; Bharadwaj, K.; Daugherty, A.; Brown, D.R.; Randall, D.C.; Cassis, L.A. Activation of the Systemic and Adipose Renin-Angiotensin System in Rats with Diet-Induced Obesity and Hypertension. *Am. J. Physiol.-Regul. Integr. Comp. Physiol.* 2004, 287, R943–R949. [CrossRef] [PubMed]
- 95. Muniyappa, R.; Yavuz, S. Metabolic Actions of Angiotensin II and Insulin: A Microvascular Endothelial Balancing Act. *Mol. Cell Endocrinol.* **2013**, *378*, 59–69. [CrossRef] [PubMed]
- Kobayashi, T.; Nogami, T.; Taguchi, K.; Matsumoto, T.; Kamata, K. Diabetic State, High Plasma Insulin and Angiotensin II Combine to Augment Endothelin-1-Induced Vasoconstriction via ET A Receptors and ERK. *Br. J. Pharmacol.* 2008, 155, 974–983. [CrossRef] [PubMed]
- 97. Kane, M.O.; Etienne-Selloum, N.; Madeira, S.V.F.; Sarr, M.; Walter, A.; Dal-Ros, S.; Schott, C.; Chataigneau, T.; Schini-Kerth, V.B. Endothelium-Derived Contracting Factors Mediate the Ang II-Induced Endothelial Dysfunction in the Rat Aorta: Preventive Effect of Red Wine Polyphenols. *Pflug. Arch.-Eur. J. Physiol.* **2010**, 459, 671–679. [CrossRef]
- Schütten, M.T.J.; Houben, A.J.H.M.; de Leeuw, P.W.; Stehouwer, C.D.A. The Link Between Adipose Tissue Renin-Angiotensin-Aldosterone System Signaling and Obesity-Associated Hypertension. *Physiology* 2017, 32, 197–209. [CrossRef] [PubMed]
- Kushibiki, M.; Yamada, M.; Oikawa, K.; Tomita, H.; Osanai, T.; Okumura, K. Aldosterone Causes Vasoconstriction in Coronary Arterioles of Rats via Angiotensin II Type-1 Receptor: Influence of Hypertension. *Eur. J. Pharmacol.* 2007, 572, 182–188. [CrossRef] [PubMed]
- 100. Almabrouk, T.A.M.; White, A.D.; Ugusman, A.B.; Skiba, D.S.; Katwan, O.J.; Alganga, H.; Guzik, T.J.; Touyz, R.M.; Salt, I.P.; Kennedy, S. High Fat Diet Attenuates the Anticontractile Activity of Aortic PVAT via a Mechanism Involving AMPK and Reduced Adiponectin Secretion. *Front. Physiol.* 2018, *9*, 51. [CrossRef]
- Aghamohammadzadeh, R.; Unwin, R.D.; Greenstein, A.S.; Heagerty, A.M. Effects of Obesity on Perivascular Adipose Tissue Vasorelaxant Function: Nitric Oxide, Inflammation and Elevated Systemic Blood Pressure. J. Vasc. Res. 2016, 52, 299. [CrossRef]
- 102. Tontonoz, P.; Spiegelman, B.M. Fat and Beyond: The Diverse Biology of PPARy. Annu. Rev. Biochem. 2008, 77, 289-312. [CrossRef]
- 103. Chatterjee, T.K.; Stoll, L.L.; Denning, G.M.; Harrelson, A.; Blomkalns, A.L.; Idelman, G.; Rothenberg, F.G.; Neltner, B.; Romig-Martin, S.A.; Dickson, E.W.; et al. Proinflammatory Phenotype of Perivascular Adipocytes: Influence of High-Fat Feeding. *Circ. Res.* 2009, *104*, 541–549. [CrossRef] [PubMed]
- 104. Schroeter, M.R.; Eschholz, N.; Herzberg, S.; Jerchel, I.; Leifheit-Nestler, M.; Czepluch, F.S.; Chalikias, G.; Konstantinides, S.; Schäfer, K. Leptin-Dependent and Leptin-Independent Paracrine Effects of Perivascular Adipose Tissue on Neointima Formation. *Arter. Thromb. Vasc. Biol.* 2013, 33, 980–987. [CrossRef] [PubMed]
- 105. Korda, M.; Kubant, R.; Patton, S.; Malinski, T. Leptin-Induced Endothelial Dysfunction in Obesity. *Am. J. Physiol. Heart Circ. Physiol.* **2008**, 295, H1514–H1521. [CrossRef]
- 106. Martin, S.S.; Qasim, A.; Reilly, M.P. Leptin Resistance: A Possible Interface of Inflammation and Metabolism in Obesity-Related Cardiovascular Disease. J. Am. Coll. Cardiol. 2008, 52, 1201–1210. [CrossRef]
- 107. Hou, N.; Luo, J.-D. Leptin and Cardiovascular Diseases. Clin. Exp. Pharmacol. Physiol. 2011, 38, 905–913. [CrossRef]
- Harlan, S.M.; Morgan, D.A.; Agassandian, K.; Guo, D.-F.; Cassell, M.D.; Sigmund, C.D.; Mark, A.L.; Rahmouni, K. Ablation of the Leptin Receptor in the Hypothalamic Arcuate Nucleus Abrogates Leptin-Induced Sympathetic Activation. *Circ. Res.* 2011, 108, 808–812. [CrossRef] [PubMed]
- Geng, B.; Cai, B.; Liao, F.; Zheng, Y.; Zeng, Q.; Fan, X.; Gong, Y.; Yang, J.; Cui, Q.H.; Tang, C.; et al. Increase or Decrease Hydrogen Sulfide Exert Opposite Lipolysis, but Reduce Global Insulin Resistance in High Fatty Diet Induced Obese Mice. *PLoS ONE* 2013, *8*, e73892. [CrossRef]

- 110. Stein, A.; Bailey, S.M. Redox Biology of Hydrogen Sulfide: Implications for Physiology, Pathophysiology, and Pharmacology. *Redox Biol.* **2013**, *1*, 32–39. [CrossRef] [PubMed]
- 111. Whiteman, M.; Gooding, K.M.; Whatmore, J.L.; Ball, C.I.; Mawson, D.; Skinner, K.; Tooke, J.E.; Shore, A.C. Adiposity Is a Major Determinant of Plasma Levels of the Novel Vasodilator Hydrogen Sulphide. *Diabetologia* 2010, 53, 1722–1726. [CrossRef]
- 112. Zhang, M.; Song, P.; Xu, J.; Zou, M.-H. Activation of NAD(P)H Oxidases by Thromboxane A2 Receptor Uncouples Endothelial Nitric Oxide Synthase. *Arter. Thromb. Vasc. Biol.* **2011**, *31*, 125–132. [CrossRef]
- Drouin, A.; Farhat, N.; Bolduc, V.; Thorin-Trescases, N.; Gillis, M.-A.; Villeneuve, L.; Nguyen, A.; Thorin, E. Up-Regulation of Thromboxane A2 Impairs Cerebrovascular ENOS Function in Aging Atherosclerotic Mice. *Pflug. Arch.* 2011, 462, 371–383. [CrossRef] [PubMed]
- 114. Trovati, M.; Mularoni, E.M.; Burzacca, S.; Ponziani, M.C.; Massucco, P.; Mattiello, L.; Piretto, V.; Cavalot, F.; Anfossi, G. Impaired Insulin-Induced Platelet Antiaggregating Effect in Obesity and in Obese NIDDM Patients. *Diabetes* 1995, 44, 1318–1322. [CrossRef] [PubMed]
- 115. McDowell, C.P.; Campbell, M.J.; Herring, M.P. Sex-Related Differences in Mood Responses to Acute Aerobic Exercise. *Med. Sci.* Sports Exerc. 2016, 48, 1798–1802. [CrossRef] [PubMed]
- Bai, Y.; Sigala, W.; Adams, G.R.; Vaziri, N.D. Effect of Exercise on Cardiac Tissue Oxidative and Inflammatory Mediators in Chronic Kidney Disease. Am. J. Nephrol. 2009, 29, 213–221. [CrossRef]
- 117. Boa, B.C.S.; Yudkin, J.S.; van Hinsbergh, V.W.M.; Bouskela, E.; Eringa, E.C. Exercise Effects on Perivascular Adipose Tissue: Endocrine and Paracrine Determinants of Vascular Function. *Br. J. Pharm.* **2017**, *174*, 3466–3481. [CrossRef]
- 118. Fritzen, A.M.; Lundsgaard, A.-M.; Jordy, A.B.; Poulsen, S.K.; Stender, S.; Pilegaard, H.; Astrup, A.; Larsen, T.M.; Wojtaszewski, J.F.P.; Richter, E.A.; et al. New Nordic Diet-Induced Weight Loss Is Accompanied by Changes in Metabolism and AMPK Signaling in Adipose Tissue. J. Clin. Endocrinol. Metab. 2015, 100, 3509–3519. [CrossRef]
- Ruffino, J.S.; Davies, N.A.; Morris, K.; Ludgate, M.; Zhang, L.; Webb, R.; Thomas, A.W. Moderate-Intensity Exercise Alters Markers of Alternative Activation in Circulating Monocytes in Females: A Putative Role for PPARγ. *Eur. J. Appl. Physiol.* 2016, 116, 1671–1682. [CrossRef] [PubMed]
- 120. Meziat, C.; Boulghobra, D.; Strock, E.; Battault, S.; Bornard, I.; Walther, G.; Reboul, C. Exercise Training Restores ENOS Activation in the Perivascular Adipose Tissue of Obese Rats: Impact on Vascular Function. *Nitric. Oxide* **2019**, *86*, 63–67. [CrossRef]
- 121. Cao, S.; Li, B.; Yi, X.; Chang, B.; Zhu, B.; Lian, Z.; Zhang, Z.; Zhao, G.; Liu, H.; Zhang, H. Effects of Exercise on AMPK Signaling and Downstream Components to PI3K in Rat with Type 2 Diabetes. *PLoS ONE* **2012**, *7*, e51709. [CrossRef]
- 122. Sousa, A.S.; Sponton, A.C.S.; Trifone, C.B.; Delbin, M.A. Aerobic Exercise Training Prevents Perivascular Adipose Tissue-Induced Endothelial Dysfunction in Thoracic Aorta of Obese Mice. *Front. Physiol.* **2019**, *10*, 1009. [CrossRef]
- 123. Saxton, S.N.; Toms, L.K.; Aldous, R.G.; Withers, S.B.; Ohanian, J.; Heagerty, A.M. Restoring Perivascular Adipose Tissue Function in Obesity Using Exercise. *Cardiovasc. Drugs* **2021**. [CrossRef]
- 124. Farah, C.; Kleindienst, A.; Bolea, G.; Meyer, G.; Gayrard, S.; Geny, B.; Obert, P.; Cazorla, O.; Tanguy, S.; Reboul, C. Exercise-Induced Cardioprotection: A Role for ENOS Uncoupling and NO Metabolites. *Basic Res. Cardiol.* **2013**, *108*. [CrossRef]
- 125. DeVallance, E.; Branyan, K.W.; Lemaster, K.C.; Anderson, R.; Marshall, K.L.; Olfert, I.M.; Smith, D.M.; Kelley, E.E.; Bryner, R.W.; Frisbee, J.C.; et al. Exercise Training Prevents the Perivascular Adipose Tissue-Induced Aortic Dysfunction with Metabolic Syndrome. *Redox Biol.* 2019, 26, 101285. [CrossRef]
- 126. Wang, J.; Polaki, V.; Chen, S.; Bihl, J.C. Exercise Improves Endothelial Function Associated with Alleviated Inflammation and Oxidative Stress of Perivascular Adipose Tissue in Type 2 Diabetic Mice. Oxid. Med. Cell Longev. 2020, 2020, 8830537. [CrossRef] [PubMed]
- Cannon, B.; Nedergaard, J. Brown Adipose Tissue: Function and Physiological Significance. *Physiol. Rev.* 2004, 84, 277–359. [CrossRef] [PubMed]
- Liao, J.; Yin, H.; Huang, J.; Hu, M. Dysfunction of Perivascular Adipose Tissue in Mesenteric Artery Is Restored by Aerobic Exercise in High-Fat Diet Induced Obesity. *Clin. Exp. Pharmacol. Physiol.* 2021, 48, 697–703. [CrossRef] [PubMed]
- 129. Julia, C.; Péneau, S.; Andreeva, V.A.; Méjean, C.; Fezeu, L.; Galan, P.; Hercberg, S. Weight-Loss Strategies Used by the General Population: How Are They Perceived? *PLoS ONE* **2014**, *9*, e97834. [CrossRef] [PubMed]
- 130. dos Reis Costa, D.E.F.; Silveira, A.L.M.; Campos, G.P.; Nóbrega, N.R.C.; de Araújo, N.F.; de Figueiredo Borges, L.; dos Santos Aggum Capettini, L.; Ferreira, A.V.M.; Bonaventura, D. High-Carbohydrate Diet Enhanced the Anticontractile Effect of Perivascular Adipose Tissue Through Activation of Renin-Angiotensin System. *Front. Physiol.* 2021, *11*, 628101. [CrossRef]
- 131. Sasoh, T.; Kugo, H.; Kondo, Y.; Miyamoto, K.; Minami, M.; Higashihara, M.; Kawamoto, H.; Takeshita, F.; Moriyama, T.; Zaima, N. Different effects of high-fat and high-sucrose diets on the physiology of perivascular adipose tissues of the thoracic and abdominal aorta. *Adipocyte* 2021, 10, 412–423. [CrossRef]
- 132. Victorio, J.A.; Guizoni, D.M.; Freitas, I.N.; Araujo, T.R.; Davel, A.P. Effects of High-Fat and High-Fat/High-Sucrose Diet-Induced Obesity on PVAT Modulation of Vascular Function in Male and Female Mice. *Front. Pharm.* **2021**, *12*, 720224. [CrossRef] [PubMed]
- 133. Estruch, R.; Ros, E.; Salas-Salvadó, J.; Covas, M.-I.; Corella, D.; Arós, F.; Gómez-Gracia, E.; Ruiz-Gutiérrez, V.; Fiol, M.; Lapetra, J.; et al. Primary Prevention of Cardiovascular Disease with a Mediterranean Diet Supplemented with Extra-Virgin Olive Oil or Nuts. *N. Engl. J. Med.* **2018**, *378*, e34. [CrossRef] [PubMed]
- 134. Sofi, F.; Macchi, C.; Abbate, R.; Gensini, G.F.; Casini, A. Mediterranean Diet and Health Status: An Updated Meta-Analysis and a Proposal for a Literature-Based Adherence Score. *Public Health Nutr.* **2014**, *17*, 2769–2782. [CrossRef]

- 135. Luisi, M.L.E.; Biffi, B.; Gheri, C.F.; Sarli, E.; Rafanelli, E.; Graziano, E.; Vidali, S.; Fattirolli, F.; Gensini, G.F.; Macchi, C. Efficacy of a Nutritional Education Program to Improve Diet in Patients Attending a Cardiac Rehabilitation Program: Outcomes of a One-Year Follow-Up. Intern. Emerg. Med. 2015, 10, 671–676. [CrossRef]
- 136. Luisi, M.L.E.; Lucarini, L.; Biffi, B.; Rafanelli, E.; Pietramellara, G.; Durante, M.; Vidali, S.; Provensi, G.; Madiai, S.; Gheri, C.F.; et al. Effect of Mediterranean Diet Enriched in High Quality Extra Virgin Olive Oil on Oxidative Stress, Inflammation and Gut Microbiota in Obese and Normal Weight Adult Subjects. *Front. Pharm.* 2019, 10, 1366. [CrossRef]
- 137. Mantzoros, C.S.; Williams, C.J.; Manson, J.E.; Meigs, J.B.; Hu, F.B. Adherence to the Mediterranean Dietary Pattern Is Positively Associated with Plasma Adiponectin Concentrations in Diabetic Women. Am. J. Clin. Nutr. 2006, 84, 328–335. [CrossRef] [PubMed]
- 138. Ułamek-Kozioł, M.; Czuczwar, S.J.; Januszewski, S.; Pluta, R. Ketogenic Diet and Epilepsy. Nutrients 2019, 11, 2510. [CrossRef]
- 139. Muscogiuri, G.; Barrea, L.; Laudisio, D.; Pugliese, G.; Salzano, C.; Savastano, S.; Colao, A. The Management of Very Low-Calorie Ketogenic Diet in Obesity Outpatient Clinic: A Practical Guide. *J. Transl. Med.* **2019**, *17*. [CrossRef]
- Monda, V.; Polito, R.; Lovino, A.; Finaldi, A.; Valenzano, A.; Nigro, E.; Corso, G.; Sessa, F.; Asmundo, A.; Di Nunno, N.; et al. Short-Term Physiological Effects of a Very Low-Calorie Ketogenic Diet: Effects on Adiponectin Levels and Inflammatory States. *Int. J. Mol. Sci.* 2020, *21*, 3228. [CrossRef] [PubMed]
- 141. Kirkpatrick, C.F.; Bolick, J.P.; Kris-Etherton, P.M.; Sikand, G.; Aspry, K.E.; Soffer, D.E.; Willard, K.-E.; Maki, K.C. Review of Current Evidence and Clinical Recommendations on the Effects of Low-Carbohydrate and Very-Low-Carbohydrate (Including Ketogenic) Diets for the Management of Body Weight and Other Cardiometabolic Risk Factors: A Scientific Statement from the National Lipid Association Nutrition and Lifestyle Task Force. J. Clin. Lipidol. 2019, 13, 689–711.e1. [CrossRef]
- Puchalska, P.; Crawford, P.A. Multi-Dimensional Roles of Ketone Bodies in Fuel Metabolism, Signaling, and Therapeutics. *Cell Metab.* 2017, 25, 262–284. [CrossRef]
- 143. Bhanpuri, N.H.; Hallberg, S.J.; Williams, P.T.; McKenzie, A.L.; Ballard, K.D.; Campbell, W.W.; McCarter, J.P.; Phinney, S.D.; Volek, J.S. Cardiovascular Disease Risk Factor Responses to a Type 2 Diabetes Care Model Including Nutritional Ketosis Induced by Sustained Carbohydrate Restriction at 1 Year: An Open Label, Non-Randomized, Controlled Study. *Cardiovasc. Diabetol.* 2018, 17, 56. [CrossRef]
- 144. Dąbek, A.; Wojtala, M.; Pirola, L.; Balcerczyk, A. Modulation of Cellular Biochemistry, Epigenetics and Metabolomics by Ketone Bodies. Implications of the Ketogenic Diet in the Physiology of the Organism and Pathological States. *Nutrients* 2020, 12, 788. [CrossRef]
- 145. Mattagajasingh, I.; Kim, C.-S.; Naqvi, A.; Yamamori, T.; Hoffman, T.A.; Jung, S.-B.; DeRicco, J.; Kasuno, K.; Irani, K. SIRT1 Promotes Endothelium-Dependent Vascular Relaxation by Activating Endothelial Nitric Oxide Synthase. *Proc. Natl. Acad. Sci. USA* 2007, *104*, 14855–14860. [CrossRef]
- 146. Ungvari, Z.; Parrado-Fernandez, C.; Csiszar, A.; de Cabo, R. Mechanisms Underlying Caloric Restriction and Lifespan Regulation: Implications for Vascular Aging. *Circ. Res.* **2008**, *102*, 519–528. [CrossRef]
- 147. Vink, R.G.; Roumans, N.J.; Mariman, E.C.; van Baak, M.A. Dietary Weight Loss-induced Changes in RBP4, FFA, and ACE Predict Weight Regain in People with Overweight and Obesity. *Physiol. Rep.* **2017**, *5*, e13450. [CrossRef]
- 148. Lambert, E.A.; Sari, C.I.; Eikelis, N.; Phillips, S.E.; Grima, M.; Straznicky, N.E.; Dixon, J.B.; Esler, M.; Schlaich, M.P.; Head, G.A.; et al. Effects of Moxonidine and Low-Calorie Diet: Cardiometabolic Benefits from Combination of Both Therapies. *Obesity* 2017, 25, 1894–1902. [CrossRef] [PubMed]
- Ríos-Lugo, M.J.; Jiménez-Ortega, V.; Cano-Barquilla, P.; Mateos, P.F.; Spinedi, E.J.; Cardinali, D.P.; Esquifino, A.I. Melatonin Counteracts Changes in Hypothalamic Gene Expression of Signals Regulating Feeding Behavior in High-Fat Fed Rats. *Horm. Mol. Biol. Clin. Investig.* 2015, 21. [CrossRef] [PubMed]
- Prunet-Marcassus, B.; Desbazeille, M.; Bros, A.; Louche, K.; Delagrange, P.; Renard, P.; Casteilla, L.; Pénicaud, L. Melatonin Reduces Body Weight Gain in Sprague Dawley Rats with Diet-Induced Obesity. *Endocrinology* 2003, 144, 5347–5352. [CrossRef] [PubMed]
- 151. Szewczyk-Golec, K.; Rajewski, P.; Gackowski, M.; Mila-Kierzenkowska, C.; Wesołowski, R.; Sutkowy, P.; Pawłowska, M.; Woźniak, A. Melatonin Supplementation Lowers Oxidative Stress and Regulates Adipokines in Obese Patients on a Calorie-Restricted Diet. Oxid. Med. Cell Longev. 2017, 2017, 8494107. [CrossRef] [PubMed]
- 152. Dong, T.A.; Sandesara, P.B.; Dhindsa, D.S.; Mehta, A.; Arneson, L.C.; Dollar, A.L.; Taub, P.R.; Ssperling, L.S. Intermittent Fasting: A Heart Healthy Dietary Pattern? *Am. J. Med.* **2020**, *133*, 901–907. [CrossRef]
- 153. Liu, B.; Page, A.J.; Hatzinikolas, G.; Chen, M.; Wittert, G.A.; Heilbronn, L.K. Intermittent Fasting Improves Glucose Tolerance and Promotes Adipose Tissue Remodeling in Male Mice Fed a High-Fat Diet. *Endocrinology* **2019**, *160*, 169–180. [CrossRef]
- Wang, X.; Yang, Q.; Liao, Q.; Li, M.; Zhang, P.; Santos, H.O.; Kord-Varkaneh, H.; Abshirini, M. Effects of Intermittent Fasting Diets on Plasma Concentrations of Inflammatory Biomarkers: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Nutrition* 2020, 79–80, 110974. [CrossRef]
- 155. Liu, B.; Hutchison, A.T.; Thompson, C.H.; Lange, K.; Heilbronn, L.K. Markers of Adipose Tissue Inflammation Are Transiently Elevated during Intermittent Fasting in Women Who Are Overweight or Obese. *Obes. Res. Clin. Pract.* 2019, 13, 408–415. [CrossRef] [PubMed]

- 156. Kim, K.-H.; Kim, Y.H.; Son, J.E.; Lee, J.H.; Kim, S.; Choe, M.S.; Moon, J.H.; Zhong, J.; Fu, K.; Lenglin, F.; et al. Intermittent Fasting Promotes Adipose Thermogenesis and Metabolic Homeostasis via VEGF-Mediated Alternative Activation of Macrophage. *Cell Res.* 2017, 27, 1309–1326. [CrossRef]
- 157. Fuentes, L.; Roszer, T.; Ricote, M. Inflammatory Mediators and Insulin Resistance in Obesity: Role of Nuclear Receptor Signaling in Macrophages. *Mediat. Inflamm.* 2010, 2010, 219583. [CrossRef] [PubMed]
- 158. Sica, A.; Mantovani, A. Macrophage Plasticity and Polarization: In Vivo Veritas. J. Clin. Investig. 2012, 122, 787–795. [CrossRef]
- 159. Kajita, K.; Mune, T.; Ikeda, T.; Matsumoto, M.; Uno, Y.; Sugiyama, C.; Matsubara, K.; Morita, H.; Takemura, M.; Seishima, M.; et al. Effect of Fasting on PPARγ and AMPK Activity in Adipocytes. *Diabetes Res. Clin. Pract.* **2008**, *81*, 144–149. [CrossRef]
- Weir, H.J.; Yao, P.; Huynh, F.K.; Escoubas, C.C.; Goncalves, R.L.; Burkewitz, K.; Laboy, R.; Hirschey, M.D.; Mair, W.B. Dietary Restriction and AMPK Increase Lifespan via Mitochondrial Network and Peroxisome Remodeling. *Cell Metab.* 2017, 26, 884–896.e5.
 [CrossRef]
- 161. Clemente-Postigo, M.; Tinahones, A.; El Bekay, R.; Malagón, M.M.; Tinahones, F.J. The Role of Autophagy in White Adipose Tissue Function: Implications for Metabolic Health. *Metabolites* **2020**, *10*, 179. [CrossRef]
- Rocchi, A.; He, C. Emerging Roles of Autophagy in Metabolism and Metabolic Disorders. *Front. Biol.* 2015, 10, 154–164. [CrossRef]
 [PubMed]