Cortical differences across psychiatric disorders and associated common and rare genetic variants

Kuldeep Kumar PhD¹, Zhijie Liao PhD¹, Jakub Kopal PhD², Clara Moreau PhD¹, Christopher R. K. Ching PhD³, Claudia Modenato PhD⁴, Will Snyder⁵, Sayeh Kazem MSc¹, Charles-Olivier Martin PhD¹, Anne-Marie Bélanger MSc¹, Valérie K. Fontaine MSc¹, Khadije Jizi MSc¹, Rune Boen PhD³, Guillaume Huguet PhD¹, Zohra Saci PhD¹, Leila Kushan MSc³, Ana I. Silva PhD^{8,22}, 16p11.2 European Consortium, Simons Searchlight Consortium, Marianne B.M. van den Bree PhD^{8,9,10}, David E.J. Linden MD^{8,10,11}, Michael J. Owen MD PhD^{8,9}, Jeremy Hall MD PhD^{8,9,10}, Sarah Lippé PhD¹, Guillaume Dumas PhD¹, Bogdan Draganski MD^{4,12,13,14}, Laura Almasy PhD^{15,16,17}, Sophia I. Thomopoulos MSc³, Neda Jahanshad PhD³, Ida E. Sønderby PhD^{2,18,19}, Ole A. Andreassen MD PhD^{2,19}, David C. Glahn PhD^{20,21}, Armin Raznahan MD PhD⁵, Carrie E. Bearden PhD³, Tomas Paus MD PhD^{1,23}, Paul M. Thompson PhD³, Sébastien Jacquemont MD¹.

¹Centre de recherche CHU Sainte-Justine and University of Montreal, Canada

² Centre for Precision Psychiatry, Division of Mental Health and Addiction, Institute of Clinical Medicine, University of Oslo, Oslo, Norway

³ Imaging Genetics Center, Mark and Mary Stevens Neuroimaging and Informatics Institute, Keck School of Medicine, University of Southern California, Marina del Rey, California, USA

⁴ LREN - Department of Clinical Neurosciences, Centre Hospitalier Universitaire Vaudois and University of Lausanne, Switzerland

⁵ Section on Developmental Neurogenomics, Human Genetics Branch, NIMH, NIH, Bethesda, MD, USA

⁶ Department of Psychiatry, University of Cambridge, Cambridge, UK

⁷ Semel Institute for Neuroscience and Human Behavior, Departments of Psychiatry and Biobehavioral Sciences and Psychology, UCLA, Los Angeles, USA

- ⁸ Centre for Neuropsychiatric Genetics and Genomics, Cardiff University, Cardiff, United Kingdom
- ⁹ Division of Psychological Medicine and Clinical Neurosciences, School of Medicine, Cardiff University, Cardiff, United Kingdom
- Neuroscience and Mental Health Research Institute, Cardiff University, Cardiff, United Kingdom
- ¹¹ Mental Health and Neuroscience Research Institute, Maastricht University, Netherlands
- ¹² Neurology Department, Max-Planck-Institute for Human Cognitive and Brain Sciences, Leipzig, Germany
- ¹³ Department of Neurology, Inselspital, University of Bern, Bern, Switzerland
- ¹⁴University Institute for Diagnostic and Interventional Neuroradiology, Inselspital, University of Bern, Bern, Switzerland
- ¹⁵ Department of Biomedical and Health Informatics, Children's Hospital of Philadelphia, PA, USA
- ¹⁶ Lifespan Brain Institute, Children's Hospital of Philadelphia and Penn Medicine, PA, USA
- ¹⁷ Department of Genetics, University of Pennsylvania, PA, USA
- ¹⁸ Department of Medical Genetics, Oslo University Hospital, Oslo, Norway
- ¹⁹ KG Jebsen Centre for Neurodevelopmental Disorders, University of Oslo, Oslo, Norway.
- ²⁰ Harvard Medical School, Department of Psychiatry, 25 Shattuck St, Boston, MA, USA
- ²¹ Boston Children's Hospital, Tommy Fuss Center for Neuropsychiatric Disease Research, 300 Longwood Avenue, Boston, MA, USA
- ²² Center for Magnetic Resonance Research, Department of Radiology, University of Minnesota, MN, USA
- ²³ Departments of Psychiatry and Neuroscience, University of Montreal, Montreal, Quebec, Canada

Abstract

Genetic studies have identified common and rare variants increasing the risk for neurodevelopmental and psychiatric disorders (NPDs). These risk variants have also been shown to influence the structure of the cerebral cortex. However, it is unknown whether cortical differences associated with genetic variants are linked to the risk they confer for NPDs. To answer this question, we analyzed cortical thickness (CT) and surface area (SA) for common and rare variants associated with NPDs, in ~33000 individuals from the general population and clinical cohorts, as well as ENIGMA summary statistics for 8 NPDs. Rare and common genetic variants increasing risk for NPDs were preferentially associated with total SA, while NPDs were preferentially associated with mean CT. Larger effects on mean CT, but not total SA, were observed in NPD medicated subgroups. At the regional level, genetic variants were preferentially associated with effects in sensorimotor areas, while NPDs showed higher effects in association areas. We show that schizophrenia- and bipolar-disorderassociated SNPs show positive and negative effect sizes on SA suggesting that their aggregated effects cancel out in additive polygenic models. Overall, CT and SA differences associated with NPDs do not relate to those observed across individual genetic variants and may be linked with critical non-genetic factors, such as medication and the lived experience of the disorder.

Introduction

Neurodevelopmental and psychiatric disorders (NPDs) represent a significant burden on global health, characterized by complex etiologies rooted in brain development and function¹⁻⁴. NPDs are highly heritable, with a broad spectrum of common and rare genetic variants implicated in their pathogenesis^{2,5-13}. The increasing availability of large-scale magnetic resonance imaging (MRI) and genetics datasets has afforded the opportunity to discover genetic variants, both common and rare, influencing the structure of the human cerebral cortex^{14–20}. Studies have shown that cortical morphology measures, such as cortical thickness (CT) and surface area (SA), are highly heritable 14,17,18, are driven by distinct neurobiological processes^{21–23}, and are influenced by largely separate genetic factors^{14,17,18,24,25}. Consequently, considerable research has explored the genetic overlap between variants associated with NPDs and those influencing cortical morphology 14,17,18,26-29, revealing small genetic correlations between NPDs and SA, and even weaker overlaps for CT. In parallel to genetic investigations, large scale case-control neuroimaging studies have consistently identified cortical differences associated with a range of psychiatric disorders ^{30,31}. Cross-disorder analyses have further revealed shared regional patterns of cortical variation^{32–37}, often aligning along the well-established sensorimotor-association cortical gradient^{18,38-40}. Cortical differences have also been reported for NPD-associated rare genetic variants, including copy number variations (CNVs)^{16,37,41–45}. Neuroimaging of these NPD-associated CNVs has demonstrated, on average, much larger effects on CT and SA^{41–43,46}, than those observed in NPDs^{15,42,43}. However, a critical question remains unanswered: whether the cortical differences associated with both common and rare genetic

variants are related to the risk they confer for psychiatric disorders. The lack of large-scale cohorts integrating genetics and neuroimaging data across healthy controls and individuals with psychiatric disorders has hindered direct investigation of this crucial relationship. Therefore, to address this knowledge gap, we designed an analytical approach to systematically compare the neuroimaging signatures of common and rare genetic variants that increase the risk for psychiatric disorders to the neuroimaging signatures associated with NPDs.

Here we aggregated multiple datasets as well as published summary statistics from ENIGMA³¹ consortium to compare clinical diagnoses-related (8 NPDs) and gene-related (common and rare variants) case-control group differences on global and regional CT and SA. Specifically, we aimed to compare effect sizes on CT and SA for three primary categories: i) 8 NPDs³¹ (attention deficit hyperactivity disorder (ADHD)⁴⁷; autism spectrum disorder (ASD)^{48,49}; bipolar disorder (BD)⁵⁰; clinical high-risk for psychosis (CHR-PS)⁵¹; conduct disorder (CD)⁵²; major depressive disorder (MDD)⁵³; obsessive-compulsive disorder (OCD)⁵⁴; and schizophrenia (SCZ)⁵⁵), considering medicated and unmedicated subgroups where available; ii) common variants⁵⁶⁻⁵⁹ associated with NPDs; and iii) 18 different CNV and aneuploidy rare variants associated with NPDs. Overall, we provide the first systematic evaluation of cortical differences associated with NPDs, their genetic variants (both common and rare), and place these findings in the context of twin and SNP heritability estimates¹⁴. These comprehensive analyses reveal a notable pattern: while psychiatric disorders preferentially affect mean CT, and association cortical regions, genetic variants impact total-SA and sensorimotor cortical regions.

Results

Effects of psychiatric disorders and associated genetic variants on

global cortical metrics

We sought to quantify effect sizes of neurodevelopmental and psychiatric disorders (NPDs)

and associated genetic variants on total cortical surface area (total SA) and mean cortical

thickness (mean CT). Effect sizes (Cohen's d) on total SA were an order of magnitude larger

for rare genetic variants compared with NPDs (11-fold, Wilcoxon rank-sum test, FDR

q<0.05, Figure 2B). This difference in effect sizes was less pronounced for mean CT (3-fold,

Wilcoxon rank-sum test, FDR q<0.05, Figure 2A). Rare genetic variants were associated

with larger effect sizes on total SA compared to mean CT while NPD-diagnosis showed

preferential association with mean CT (paired t-test and ratio of Cohen's d, FDR q<0.05,

Figure 2C-D). Of note, some early onset (childhood/adolescence subgroups) conditions

showed preferential effects on total SA (conduct disorder, and MDD-young, Figure 2C), but

we did not observe a preferential effect on surface when stratifying for pediatric/young NPDs

(Supplement Figure 1). A sensitivity analysis showed similar mean CT and total SA effect

sizes for rare variants across deletions, duplications, and sex chromosome aneuploidies

(Supplement Figure 2).

We then asked if preferential effects on total SA also applied to common variants associated

with NPDs (ADHD⁵⁷, BD⁵⁶, MDD⁵⁸, and SCZ⁵⁹). Previous work has shown that genetic

contribution to total SA was weakly correlated with NPDs. Genetic correlation between CT

and NPDs has not been detected¹⁴. We therefore sought to identify, beyond genetic

correlation, the potential genetic overlap between NPDs and cortical structure by computing a

non-parametric enrichment. NPD-associated SNPs were mildly enriched (above-median

ranking) in SA-associated SNPs (non-parametric p-value<0.0001; Figure 2E). In contrast,

none of the NPD-associated SNPs showed any enrichment for association with CT. Similarly,

the SA-genome-wide associated SNPs, but not CT, were also enriched in NPD associations

(above-median ranking, Supplement Figure 3).

We performed sensitivity analyses to test if differences observed above between genetic

variants and psychiatric diagnoses could be in part influenced by medication (also a proxy for

severity⁶⁰) and the age of participants (a proxy for severity or neurodevelopmental processes

in earlier onset participants, as well as duration of illness in adults). Effect sizes on mean CT

were 3.4-fold larger in medicated compared to unmedicated NPD subgroups (FDR q<0.05,

Figure 3A), while total SA remained the same in both subgroups (Figure 3B). As a result,

medicated sub-groups exhibited larger effects on mean CT compared to total SA (FDR

q<0.05), while unmedicated sub-groups had similar effect sizes for both metrics **Figure 3C**).

The preferential effect on mean CT observed for NPDs could not be explained by stratifying

participants according to age (pediatric, young, and adult participants, Supplement Figure

1).

Opposing regional patterns of cortical differences between NPDs and

genetic variants

We investigated if the preferential effects on total SA for genetic variants and mean CT for

psychiatric diagnoses were uniformly distributed across the cerebral cortex or localized to

specific cortical regions. To contextualize the regional profiles, we tested their similarities

with the well-established cortical gradient (**Methods**), which ranks regions from primary sensory-motor cortices, to higher order association cortices^{38,39}. We first examined how this gradient relates to both twin and SNP heritability estimates for regional CT and SA¹⁴. Heritabilities were higher in sensorimotor regions compared to association regions (negative correlations with the cortical gradient, r=-0.47 to -0.75, p-spin<0.05, **Figure 4B, Supplement Figure 4**).

We then investigated the regional cortical effect size maps of all NPDs examined above, 257 genome-wide NPD-associated SNPs¹⁴, and 11 CNVs (Supplement Table 1). Significant associations with regional SA or CT were observed for 18 NPDs, 20 NPD-associated common variants (SNPs), and all 11 CNVs (Figure 4A). Cohen's d maps without any FDR-significant ROI association were excluded from the regional analyses below (non-significant maps included medications subgroups, Figure 4A, Supplement Figure 5). We computed 4 set of consensus cortical maps, including 3 disregarding the directionality of effects, using following approaches: i) mean absolute effect size; ii) percentage of significance; iii) variance in effect size values; and iv) latent dimensions of cortical differences using principal component analysis across effect size profiles (Supplement Figure 6-7). The rare and common variant maps showed positive correlations (r=0.29 to 0.31, p-spin < 0.05, **Supplement Figure 8**), and did not show correlation with the NPD maps (Supplement Figure 8). NPD-associated rare and common genetic variants showed higher effect sizes in sensorimotor regions across all consensus map methods compared to association regions (Figure 4B, 5C, correlations with the cortical gradient: r=-0.43, p-spin=9.5e-3, for CT; r=-0.3, p-spin=2e-2, for SA). NPDs maps showed the opposite correlations with the cortical gradient highlighting larger effects in association regions

(**Figure 4B, 5C**, CT: r=0.28, p-spin=9.3e-2; SA: r=0.32, p-spin=3.2e-2). Overall,

NPD-associated common and rare variant effect sizes as well as twin and SNP heritability

estimates for cortical measures were higher in sensorimotor regions, while the opposite was

observed for NPDs (Figure 4-5).

Polygenic and familial liability to NPDs

We asked if the results showing preferential associations with total SA for individual rare and

common genetic variants computed above may be reconciled with the difficulty to detect

associations between SA and NPDs, as well as polygenic or familial liability to NPDs. For

global metrics, polygenic risk scores (PRS), PRS-BD and PRS-SCZ, were not associated with

either metric in 31,000 UK Biobank participants of European ancestry (Supplement Figure

9). To investigate the lack of PRS signal, we used regression estimates derived from cortical

structure GWAS. Half of the BD and SCZ associated SNPs had a negative beta estimate for

total SA (BD: 51.05%, and SCZ: 48.26%, Supplement Figure 9), while the other half

showed positive beta estimate, likely resulting in cancellation of the effects of additive

psychiatric PRS models on cortical structure. Using summary statistics⁶¹ in first degree

relatives, a proxy for multifactorial polygenic liability, we observed similar effects on total

SA for the family member and those with a diagnosis of SCZ or BD (Supplement Figure

10).

To understand why genetic variants show preferential effects on sensorimotor regions for

surface area, while NPDs demonstrate little or none, we examined the loadings of genetic

variants on the first latent dimension. Half of the rare variants loaded positively, while the

other loaded negatively on the SA latent dimension (i.e., increased and decreased surface in

sensorimotor regions, **Figure 5D**). The same was true for common variants (**Figure 5D**). As a result, we were unable to detect associations between PRS for BD and SCZ and surface area of sensorimotor regions (**Supplement Figure 9**).

Discussion

Individual genetic variants and NPD diagnoses showed, on average, opposing effects on global cortical differences, where rare and common genetic variants associated with NPDs preferentially affected total SA while the diagnosis of NPDs was associated with mean CT. The effect of NPDs on mean CT was larger in medication sub-groups, which are proxies for disorder severity. Beyond global effects, psychiatric diagnoses preferentially affected association cortical regions (involved in higher-order functions such as language, decision-making, and social cognition) while rare and common genetic variants impacted sensorimotor cortical regions (involved in basic sensory and motor functions). We also show that the polygenic architecture of NPDs (BD and SCZ) together with the positive and negative effects sizes of genetic variants on cortical structure may leads to the cancellation of most of the effects of NPD polygenic scores on cortical structure.

The preferential effects on total SA, compared to mean CT, of NPD-associated rare and common variants is concordant with the higher heritability estimates for total SA compared to mean CT¹⁴. The observed preferential effect of NPDs on mean CT may be attributable to critical non-genetic factors including medication, environmental factors, and plasticity in the context of a psychiatric disorder. While directly modeling these effects remains challenging,

our findings suggest a potential role for medication⁶², proxy for disorder severity. This also aligns with longitudinal findings from a randomized clinical trial in MDD showing medication-related changes in mean CT but not total SA⁶². Studies of familial risk, also support this notion, where individuals with either SZ or BD exhibited larger effect sizes on mean CT compared to their unaffected first degree relatives, while effects on total SA were not significant in both groups⁶¹. Furthermore, comparing 22q11.2 deletion carriers with and without Psychosis⁴⁵, showed differences in mean CT but not total SA. These differential effects may be consistent with the distinct developmental trajectories of cortical SA and CT^{24,25}. While the majority of cortical expansion occurs before the first two years^{24,63} and is relatively stable thereaftere^{23,64}, CT undergoes protracted maturation involving changes in dendritic arborization and myelination that continue into adulthood^{24,38,63-65}. As a result, further investigation of childhood onset conditions should reveal larger association with SA. Rare and common genetic variants preferentially impact sensorimotor cortical regions, which is concordant with higher twin and SNP heritability estimates for SA and CT in those same regions¹⁴. In contrast, NPDs tend to affect association regions (subserving executive, socioemotional, and mentalizing functions), which is in line with previous studies^{33–35}. Theses preferential effects in association areas (focal reduction in CT) have also been reported in 22q11.2 deletion carriers with a diagnosis of psychosis compared to those without⁴⁵. These differential regional effects in individual genetic variants and those with psychiatric disorders may be consistent with studies showing that the spatiotemporal patterns of CT and SA maturation follow a sensorimotor-to-association gradient, where sensorimotor cortical regions mature before the association regions^{24,38,63}.

Studies reported higher twin and SNP heritability estimates for SA compared to CT, and this study suggests the preferential effects of NPD-associated common and rare genetic variants on SA. However, detecting effects on SA in psychiatric conditions has been challenging³¹, raising a paradox given the high heritability estimates of psychiatric disorders^{1,5}. Notably, some early onset (childhood/adolescence subgroups) conditions showed preferential effects on total-SA (Conduct Disorder, and MDD-young) but detecting effects on SA remains challenging for other highly heritable childhood disorders such as ASD and ADHD. Future studies of pediatric NPDs are required to shed light on these questions.

Challenges to identify effects on SA may be a consequence of polygenic architecture of NPDs. Our investigation of individual SZ- and BD-associated common variants and their aggregated effects on total and regional SA (i.e., in the sensorimotor regions) using polygenic risk scores (PRS) suggests that many of the SA effects of individual SNPs are cancelled out in a PGS model. The effects of individual variants on total and regional SA (in the sensorimotor regions) may instead point to a hidden underlying mechanism indirectly related to SA. As an example, the level of transcriptomic differences observed in autistic brains (compared to controls) follows the cortical gradient, with the larger levels of differentially expressed genes occurring in the sensorimotor compared to association regions⁶⁶.

There are several limitations of our study. We are only investigating 8 NPDs and selected recurrent CNVs, this is largely due to the limited availability of the large CNV MRI cohorts, as well as robust associations of these CNVs with other conditions with published summary statistics^{43,67}. While our analysis showed similar observations across sex chromosome aneuploidies and CNVs, we have to note that aneuploidies have specific effects during puberty, and which will require careful delineation^{64,68}. Finally, the ENIGMA medication

analyses were not always able to account for confounding factors like illness severity and duration, movement, treatment dose and duration, and other co-occurring conditions that might be influencing the reported medication effects. While we see some consistent medication-related signals across sub-groups (and many ENIGMA studies are the largest of their kind), other factors could still be at play. Additionally, in uncontrolled studies, patients with the most severe symptoms often need the highest doses and experience the greatest brain changes, so medication is confounded with disease severity and duration of illness⁶². Nevertheless, our finding of differential effects on CT compared to SA aligns with randomized clinical trial evidence of medication-related brain structural changes⁶², suggesting that medication status can influence case-control brain profiles across disorders. Further research is warranted to determine the extent to which these findings generalizes to other neuroimaging measures and modalities⁶².

In summary, our study contributes to the ongoing effort to understand the genetic basis of the development of the cerebral cortex and its deviation in psychiatric disorders, by bringing together findings from neuroimaging, psychiatry, and genetics. We reported the distinct neuroimaging profiles observed in NPDs, compared to those associated with individual genetic variants. Our study suggests that the neuroimaging differences in NPDs are likely a manifestation of the aggregated effects of their polygenic architecture, influenced by critical non-genetic factors like medication and the lived experience of the disorder, highlighting the need for integrated models.

Methods

Data summary: In this study, we aggregated multiple datasets as well as published summary

statistics from ENIGMA Consortium³¹ to compare case control effect sizes (Cohen's d)

across 8 psychiatric disorders and associated common and rare variants on global and

regional CT and SA.

Rare genetic variant participants: Recurrent deletions and duplications were included in the

analysis based on previous analysis of T1-weighted MRI data^{41,44,67}, and where at least 18

carriers of the same CNV were available 41,67. Clinically ascertained groups: CNV carriers

were recruited after either being referred for genetic testing due to the diagnosis of a

neurodevelopmental disorder or as the relative (e.g., parent) of a CNV carrier. Controls were

defined as individuals who did not carry any NPD-associated CNVs.

Unselected population group: CNV carriers were identified in the UK Biobank. Controls

were defined as individuals who did not carry any of the recurrent CNVs selected from this

study. Demographic details and coordinates of each of the 11 CNVs are provided in

Supplement Material. Signed consents were obtained by investigators from each cohort for

all participants and/or their legal representatives prior to the investigation. This study, using

13

an aggregate dataset, obtained ethics approval from the CHU Sainte-Justine Hospital.

Rare genetic variant MRI image acquisition and preprocessing: The data sample included 3D

T1-weighted (T1w) volumetric brain images at 0.8-1 mm isotropic resolution across all sites.

MRI parameters for each cohort are detailed in the Supplemental Material. *Quality control*:

Visual quality inspection was performed by the two raters (CM, KK) using the ENIGMA

standardized quality control protocol (https://github.com/ENIGMA-git).

NPD participants, MRI image acquisition and preprocessing: In this study, we aggregated

published summary statistics for 8 NPDs from the following published ENIGMA studies:

ADHD⁴⁹; ASD⁴⁹; BD ⁵⁰; clinical high-risk for psychosis (CHR-PS)⁵¹; conduct disorder

(CD)⁵²; MDD⁵³; OCD⁴⁹; and SCZ⁵⁵. All these studies followed the ENIGMA standardized

quality control protocol (https://github.com/ENIGMA-git), and processing. An extensive

description of MRI image acquisition, methods for pre-processing the data are available in

the respective published study.

Cortical thickness and surface area measures: FreeSurfer 5.3.0 was used to extract cortical

thickness and surface area for 68 Desikan ROIs ⁶⁹, as well as total surface area (total SA), and

mean cortical thickness (mean CT). See Supplementary Methods for details.

Statistical analysis: Linear regression models (R version 3.6.3) were used to compute

CNV-control differences (Cohen's d) for each CNV using age, sex, and site, as covariates.

For regional surface area, we additionally used total Surface Area, as a covariate. This

approach was used for CNVs. The FDR procedure 70 was applied for multiple comparison

correction. The significance was set at FDR-corrected q < 0.05. See Supplementary Methods

for additional details.

Effect sizes (NPDs and CNVs): Cohen's d were computed based on case-control linear

regression. Cohen's d values for neurodevelopmental and psychiatric disorders were

extracted from published ENIGMA studies ^{30,31,71}: ADHD ⁴⁹; ASD ⁴⁹; BD ⁵⁰; clinical high-risk

for psychosis (CHR-PS)⁵¹; conduct disorder (CD)⁵²; MDD⁵³; OCD⁵⁴; and SCZ⁵⁵. All effect

sizes were computed after regressing for age, sex, and site. For regional surface area analysis,

adjustment for brain size was added either using total SA or ICV as a covariate, except for

MDD ⁵³, for which such results were unavailable. In addition, we used previously published

meta-analysis effect sizes for total SA and mean CT for 9 genetic mutations ⁴³ including 6

aneuploidies^{37,72,73} (Turner Syndrome (TS); Down Syndrome; XXX; XXY; XYY; and XXYY)

and 3 CNVs (7q11.23⁷⁴, 16p11.2 distal deletion and duplication ^{42,75}).

Effect size comparison metrics: For comparisons across metrics, the following effect size

metrics were used: absolute Cohen's d for global measures, and average absolute Cohen's d

of the top decile across regional cortical thickness and surface area ⁶⁷. As the proportion of

significant regions varied across CNVs and NPDs (due to differences in effect and sample

sizes), we chose to focus on the top decile Cohen's d for all CNVs and NPDs to avoid biases

and to provide effect sizes comparable across CNVs and NPDs. Statistical testing of spatially

correlated Cohen's d profiles was performed using spin permutation testing 76,77 . See details in

Supplementary Methods. All cortical projections were generated using the ggseg R package

15

Common genetic risk associated with NPDs: To assess common genetic risk associated with

NPDs, we investigated the GWAS summary statistics and genome-wide significant loci from

ENIGMA and PGC. Specifically, we leveraged the following published statistics: cortical

thickness and surface area¹⁴; ADHD ⁵⁷; BD⁵⁶; MDD ⁵⁸; and SCZ ⁵⁹. We used the effect sizes

from ENIGMA cortical summary statistics¹⁴ for all the NPD-associated genome-wide

significant loci, both global and regional MRI metrics.

Ranking and significance of NPD-associated genome-wide significant loci: To investigate

whether the common genetic risk factors associated with NPDs are also implicated in the

genetic architecture of the human cortex, we conducted the following analysis. We ranked

independent single nucleotide polymorphisms (SNPs) from GWAS of NPDs (ADHD, BD,

MDD, and SCZ) according to their association with cortical GWAS (cortical thickness and

surface area). Subsequently, we calculated the median rank of NPD SNPs within the ranked

cortical GWAS. To assess the significance of this observation, we performed 10,000 null

permutations by randomly sampling an equal number of SNPs and recomputing the median

rank. This yielded a permutation-based p-value for each NPD. Finally, we applied a

significance threshold (FDR q < 0.05) based on the permutation testing. This was also

assessed for the cortical thickness and surface area SNPs, by ranking them based on NPD

GWASes. SNPs from SCZ and cortical thickness GWAS were excluded from further analyses

16

due to the lack of significant enrichment.

Consensus maps of regional cortical thickness and surface area differences: We computed

consensus maps (per ROI across CNVs/NPDs) using: i) mean absolute effect size; ii) %

significance; iii) variance in effect size values; and iv) latent dimensions of cortical

differences across CNVs and across NPDs: Principal component analysis identified latent

dimensions of cortical thickness and surface area differences across all CNVs and across all

NPDs separately. We used the FactoMineR 79 package in R and ran PCA on Cohen's d values.

Correlation with normative maps of cortical gradients: We used the previously published

normative maps of cortical organization hierarchies ³⁹, specifically, the spatial transcriptomic

map (PC1 gene expression) ^{38,40}. To compute a spatial correlation with the normative map, we

mapped these maps to 68 cortical regions of Desikan parcellation using the neuromaps

python package ³⁹. Finally, a spin permutation method ^{76,77} was used to assess the significance

of the spatial correlation.

Polygenic Risk Scores (PRS): We used the standard PRS scores for bipolar disorder and

schizophrenia, provided by the UK Biobank (data fields 26214 and 26275)⁸⁰. We only kept

individuals of European Ancestry and removed any recurrent CNV carriers.

Data and materials availability

UK Biobank data was downloaded under the application 40980 and may be accessed via their

standard data access procedure (see http://www.ukbiobank.ac.uk/register-apply). UK Biobank

CNVs were called using the pipeline developed in the Jacquemont Lab, as described at

https://github.com/MartineauJeanLouis/MIND-GENESPARALLELCNV. The final CNV

calls are available for download from the UK Biobank returned datasets (Return ID: 3104,

https://biobank.ndph.ox.ac.uk/ukb/dset.cgi?id=3104). The 22q11.2 UCLA raw data are

currently available by request from the project PI. Raw neuroimaging data for rare variants

are available through request and data access agreement from the PIs of the projects (Brain

Canada: S.J. CHUSJ Montreal; 22q11.2: C.E.B. UCLA, Cardiff: D.E.J.L., M.J.O., M.V.B.,

J.H, Cardiff University; SCA: A.R. NIMH). References to the processing pipeline and R

package versions used for analysis are listed in the methods.

Code availability

The code for generating all the figures and supplement figures, along with processed

summary measures is available in the following GitHub repository:

https://github.com/kkumar-iitkgp/ct sa across disorders and variants.git

Acknowledgments

Funding

This research was supported by Calcul Quebec (http://www.calculquebec.ca) and Compute

Canada (http://www.computecanada.ca), the Brain Canada Multi-Investigator initiative, NIH

U01 grant for CAMP (1U01MH119690-01), the Canadian Institutes of Health Research,

CIHR 400528, The Institute of Data Valorization (IVADO) through the Canada First

Research Excellence Fund, Healthy Brains for Healthy Lives through the Canada First

Research Excellence Fund. Dr Jacquemont is a recipient of a Canada Research Chair in neurodevelopmental disorders and a chair from the Jeanne et Jean Louis Levesque Foundation. The Cardiff CNV cohort was supported by the Wellcome Trust Strategic Award "DEFINE" and the National Centre for Mental Health with funds from Health and Care Research Wales (code 100202/Z/12/Z). The CHUV cohort was supported by the SNF (Maillard Anne, Project, PMPDP3 171331). Data from the UCLA cohort provided by Dr. Bearden (participants with 22q11.2 deletions or duplications and controls) was supported through grants from the NIH (U54EB020403), NIMH (R01MH085953, R01MH100900, R03MH105808), and the Simons Foundation (SFARI Explorer Award). Claudia Modenato was supported by the doc.mobility grant provided by the Swiss National Science Foundation (SNSF). Kuldeep Kumar was supported by The Institute of Data Valorization (IVADO) Postdoctoral Fellowship program, through the Canada First Research Excellence Fund. CRKC and PMT are supported in part by NIMH grants R01MH116147, R01MH123163, and R01MH121246, and by the Milken Institute and the Baszucki Brain Research Fund. Dr. Sønderby is supported by the Research Council of Norway (#223273), South-Eastern Norway Regional Health Authority (#2020060), European Union's Horizon2020 Research and Innovation Programme (CoMorMent project; Grant #847776) and Kristian Gerhard Jebsen Stiftelsen (SKGJ-MED-021). BD is supported by the Swiss National Science Foundation (NCCR Synapsy, project grant numbers 32003B 135679, 32003B 159780, 324730 192755, and CRSK-3 190185), the Roger De Spoelberch and the Leenaards Foundations. G.D. is supported by the Institute for Data Valorization, Montreal (IVADO; CF00137433), the Fonds de recherche du Québec (FRQ; 285289), the Natural Sciences and Engineering Research Council of Canada (NSERC; DGECR-2023-00089), and the Azrieli Global Scholars

Fellowship from the Canadian Institute for Advanced Research (CIFAR) in the Brain, Mind,

& Consciousness program. We thank all of the families participating at the Simons

Searchlight sites, as well as the Simons Searchlight Consortium. We appreciate obtaining

access to imaging and phenotypic data on SFARI Base. Approved researchers can obtain the

Simons Searchlight population dataset described in this study by applying at

https://base.sfari.org. We are grateful to all families who participated in the 16p11.2 European

Consortium.

Disclosures

MvdB reports grants from Takeda Pharmaceuticals, outside the submitted work. P.M.T. and

CRKC received a research grant from Biogen, Inc., for work unrelated to this manuscript. All

other authors reported no biomedical financial interests or potential conflicts of interest.

Author contributions

K.K., Z.L., C.Mod., C.C, C.E.B., P.M.T., T.P., and S.J. designed the study, analyzed imaging

data, and drafted the manuscript.

<u>Analyses:</u> K.K. and C.Mod. performed all the analyses of neuroimaging data. W.S. and A.R.

performed analyses of neuroimaging data from sex chromosome aneuploidies.

Data collection: C.Mod., A.M., B.R-H., A.P., S.R., and S.M-B. recruited and scanned

participants in the 16p11.2 European Consortium. S.L., C.O.M., E.D., F. T-D., V.C., A.R.C.,

F.D. recruited and scanned participants in the Brain Canada cohort. L.K., C.E.B. collected

and provided the data for the UCLA cohort. D.E.J.L., M.J.O., M.B.M. V.d.B., J.H., and

A.I.S., provided the data for the Cardiff cohort. W.S. and A.R. provided the data for sex chromosome aneuploidies.

All authors provided feedback on the manuscript.

Article Information

Each cohort and corresponding study received approval from their local institutional review board and this study was approved by the Institutional review board of the CHU Ste Justine research center. The Simons Searchlight Consortium principal investigator is Wendy K. Chung. Contributors to the Simons Searchlight Consortium include the following: Hanalore Alupay, BS, Benjamin Aaronson, BS, Sean Ackerman, MD, Katy Ankenman, MSW, Ayesha Anwar, BA, Constance Atwell, PhD, Alexandra Bowe, BA, Arthur L. Beaudet, MD, Marta Benedetti, PhD, Jessica Berg, MS, Jeffrey Berman, PhD, Leandra N. Berry, PhD, Audrey L. Bibb, MS, Lisa Blaskey, PhD, Jonathan Brennan, PhD, Christie M. Brewton, BS, Randy Buckner, PhD, Polina Bukshpun, BA, Jordan Burko, BA, Phil Cali, EdS, Bettina Cerban, BA, Yishin Chang, MS, Maxwell Cheong, BE, MS, Vivian Chow, BA, Zili Chu, PhD, Darina Chudnovskaya, BS, Lauren Cornew, PhD, Corby Dale, PhD, John Dell, BS, Allison G. Dempsey, PhD, Trent Deschamps, BS, Rachel Earl, BA, James Edgar, PhD, Jenna Elgin, BS, Jennifer Endre Olson, PsyD, Yolanda L Evans, MA, Anne Findlay, MA, Gerald D Fischbach, MD, Charlie Fisk, BS, Brieana Fregeau, BA, Bill Gaetz, PhD, Leah Gaetz, MSW, BSW, BA, Silvia Garza, BA, Jennifer Gerdts, PhD, Orit Glenn, MD, Sarah E Gobuty, MS, CGC, Rachel Golembski, BS, Marion Greenup, MPH, MEd, Kory Heiken, BA, Katherine Hines, BA, Leighton Hinkley, PhD, Frank I. Jackson, BS, Julian Jenkins III, PhD, Rita J. Jeremy, PhD, Kelly Johnson, PhD, Stephen M. Kanne, PhD, Sudha Kessler, MD, Sarah Y. Khan, BA, Matthew Ku, BS, Emily Kuschner, PhD, Anna L. Laakman, MEd, Peter Lam, BS, Morgan

W. Lasala, BA, Hana Lee, MPH, Kevin LaGuerre, MS, Susan Levy, MD, Alyss Lian Cavanagh, MA, Ashlie V. Llorens, BS, Katherine Loftus Campe, MEd, Tracy L. Luks, PhD, Elysa J. Marco, MD, Stephen Martin, BS, Alastair J. Martin, PhD, Gabriela Marzano, HS, Christina Masson, BFA, Kathleen E. McGovern, BS, Rebecca McNally Keehn, PhD, David T. Miller, MD, PhD, Fiona K. Miller, PhD, Timothy J. Moss, MD, PhD, Rebecca Murray, BA, Srikantan S. Nagarajan, PhD, Kerri P. Nowell, MA, Julia Owen, PhD, Andrea M. Paal, MS, Alan Packer, PhD, Patricia Z. Page, MS, Brianna M. Paul, PhD, Alana Peters, BS, Danica Peterson, MPH, Annapurna Poduri, PhD, Nicholas J. Pojman, BS, Ken Porche, MS, Monica B. Proud, MD, Saba Qasmieh, BA, Melissa B. Ramocki, MD, PhD, Beau Reilly, PhD, Timothy P. L. Roberts, PhD, Dennis Shaw, MD, Tuhin Sinha, PhD, Bethanny Smith-Packard, MS, CGC, Anne Snow Gallagher, PhD, Vivek Swarnakar, PhD, Tony Thieu, BA, MS, Christina Triantafallou, PhD, Roger Vaughan, PhD, Mari Wakahiro, MSW, Arianne Wallace, PhD, Tracey Ward, BS, Julia Wenegrat, MA, and Anne Wolken, BS. European 16p11.2 Consortium principal investigator Sébastien Jacquemont. Members of the European 16p11.2 Consortium include the following: Addor Marie-Claude, Service de génétique médicale, Centre Hospitalier Universitaire Vaudois, Lausanne University, Switzerland; Andrieux Joris, Institut de Génétique Médicale, CHRU de Lille, Hopital Jeanne de Flandre, France; Arveiler Benoît, Service de génétique médicale, CHU de Bordeaux- GH Pellegrin, France; Baujat Geneviève, Service de Génétique Médicale, CHU Paris - Hôpital Necker-Enfants Malades, France; Sloan-Béna Frédérique, Service de médecine génétique, Hôpitaux Universitaires de Genève - HUG, Switzerland; Belfiore Marco, Service de génétique médicale, Centre Hospitalier Universitaire Vaudois, Lausanne University, Switzerland; Bonneau Dominique, Service de génétique médicale, CHU d'Angers, France;

Bouquillon Sonia, Institut de Génétique Médicale, Hopital Jeanne de Flandre, Lille, France; Boute Odile, Hôpital Jeanne de Flandre, CHRU de Lille, Lille, France; Brusco Alfredo, Genetica Medica, Dipartimento di Scienze Mediche, Università di Torino, Italy; Busa Tiffany, Département de génétique médicale, CHU de Marseille, Hôpital de la Timone, France; Caberg Jean- Hubert, Centre de génétique humaine, CHU de Liège, Belgique; Campion Dominique, Service de psychiatrie, Centre hospitalier de Rouvray, Sotteville lès Rouen, France; Colombert Vanessa, Service de génétique médicale, Centre Hospitalier Bretagne Atlantique CH Chubert- Vannes, France; Cordier Marie-Pierre, Service de génétique clinique, CHU de Lyon, Hospices Civils de Lyon, France; David Albert, Service de Génétique Médicale, CHU de Nantes, Hôtel Dieu, France; Debray François-Guillaume, Service de Génétique Humaine, CHU Sart Tilman - Liège, Belgique; Delrue Marie-Ange, Service de génétique médicale, CHU de Bordeaux, Hôpital Pellegrin, France; Doco-Fenzy Martine, Service de Génétique et Biologie de la Reproduction, CHU de Reims, Hôpital Maison Blanche, France; Dunkhase- Heinl Ulrike, Department of Pediatrics, Aabenraa Hospital, Sonderjylland, Denmark; Edery Patrick, Service de génétique clinique, CHU de Lyon, Hospices Civils de Lyon, France; Fagerberg Christina, Department of Clinical Genetics, Odense University hospital, Denmark; Faivre Laurence, Centre de génétique, Hôpital d'Enfants, CHU Dijon Bourgogne - Hôpital François Mitterrand, France; Forzano Francesca, Ambulatorio di Genetica Medica, Ospedali Galliera di Genova, Italy and Clinical Genetics Department, 7th Floor Borough Wing, Guy's Hospital, Guy's & St Thomas' NHS Foundation Trust, Great Maze Pond, London SE1 9RT, UK; Genevieve David, Département de Génétique Médicale, Maladies Rares et Médecine Personnalisée, service de génétique clinique, Université Montpellier, Unité Inserm U1183, CHU Montpellier, Montpellier,

France; Gérard Marion, Service de Génétique, CHU de Caen, Hôpital Clémenceau, France; Giachino Daniela, Genetica Medica, Dipartimento di Scienze Cliniche e Biologiche, Università di Torino, Italy; Guichet Agnès, Service de génétique, CHU d'Angers, France; Guillin Olivier, Service de psychiatrie, Centre hospitalier du Rouvray, Sotteville lès Rouen, France; Héron Delphine, Service de Génétique clinique, CHU Paris-GH La Pitié Salpêtrière-Charles Foix - Hôpital Pitié Salpêtrière, France; Isidor Bertrand, Service de Génétique Médicale, CHU de Nantes, Hôtel Dieu, France; Jacquette Aurélia, Service de Génétique clinique, CHU Paris-GH La Pitié Salpêtrière-Charles Foix - Hôpital Pitié-Salpêtrière, France; Jaillard Sylvie, Service de Génétique Moléculaire et Génomique – Pôle biologie, CHU de Rennes, Hôpital Pontchaillou, France; Journel Hubert, Service de génétique médicale, Centre Hospitalier Bretagne Atlantique CH Chubert- Vannes, France; Keren Boris, Centre de Génétique Moléculaire et Chromosomique, CHU Paris-GH La Pitié Salpêtrière-Charles Foix - Hôpital Pitié-Salpêtrière, France; Lacombe Didier, Service de génétique médicale, CHU de Bordeaux-GH Pellegrin, France; Lebon Sébastien, Pediatric Neurology Unit, Department of Pediatrics, Lausanne University Hospital, Lausanne, Switzerland; Le Caignec Cédric, Service de Génétique Médicale - Institut de Biologie, CHU de Nantes, France; Lemaître Marie-Pierre, Service de Neuropédiatrie, Centre Hospitalier Régional Universitaire de Lille, France; Lespinasse James, Service génétique médicale et oncogénétique, Hotel Dieu, Chambéry, France; Mathieu-Dramart Michèle, Service de Génétique Clinique, CHU Amiens Picardie, France; Mercier Sandra, Service de Génétique Médicale, CHU de Nantes, Hôtel Dieu, France; Mignot Cyril, Service de Génétique clinique, CHU Paris-GH La Pitié Salpêtrière-Charles Foix - Hôpital Pitié-Salpêtrière, France; Missirian Chantal, Département de génétique médicale, CHU de Marseille, Hôpital de la

Timone, France; Petit Florence, Service de génétique clinique Guy Fontaine, Hôpital Jeanne de Flandre, CHRU de Lille, France; Pilekær Sørensen Kristina, Department of Clinical Genetics, Odense University Hospital, Denmark; Pinson Lucile, Département de Génétique Médicale, Maladies Rares et Médecine Personnalisée, service de génétique clinique, Université Montpellier, Unité Inserm U1183, CHU Montpellier, Montpellier, France; Plessis Ghislaine, Service de Génétique, CHU de Caen, Hôpital Clémenceau, France; Prieur Fabienne, Service de génétique clinique, CHU de Saint-Etienne - Hôpital Nord, France; Raymond Alexandre, Center for Integrative Genomics, Lausanne University, Switzerland; Rooryck-Thambo Caroline, Laboratoire de génétique moléculaire, CHU de Bordeaux-GH Pellegrin, France; Rossi Massimiliano, Service de génétique clinique, CHU de Lyon, Hospices Civils de Lyon, France; Sanlaville Damien, Laboratoire de Cytogénétique Constitutionnelle, CHU de Lyon, Hospices Civils de Lyon, France; Schlott Kristiansen Britta, Department of Clinical Genetics, Odense University Hospital, Denmark; Schluth-Bolard Caroline, Laboratoire de Cytogénétique Constitutionnelle, CHU de Lyon, Hospices Civils de Lyon, France; Till Marianne, Service de génétique clinique, CHU de Lyon, Hospices Civils de Lyon, France; Van Haelst Mieke, Department of Genetics, University Medical Center Utrecht, Holland; Van Maldergem Lionel, Centre de Génétique humaine, CHRU de Besançon - Hôpital Saint-Jacques, France.

Figures

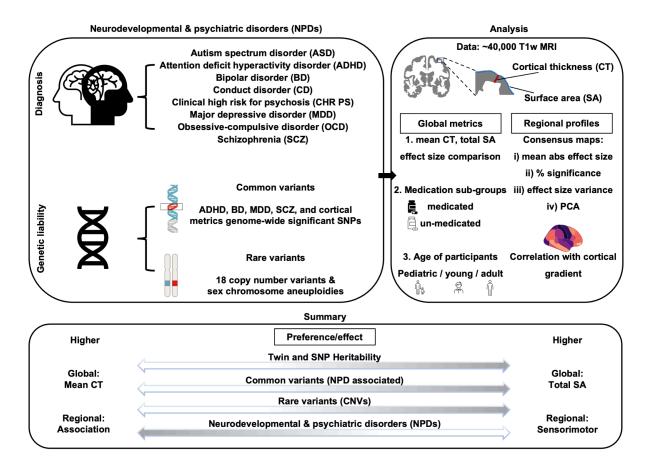


Figure 1. Graphical abstract.

Study overview. We aimed to compare effect sizes on CT and SA for three primary categories: i) 8 NPDs³¹ (attention deficit hyperactivity disorder (ADHD)⁴⁷; autism spectrum disorder (ASD)^{48,49}; bipolar disorder (BD)⁵⁰; clinical high-risk for psychosis (CHR-PS)⁵¹; conduct disorder (CD)⁵²; major depressive disorder (MDD)⁵³; obsessive-compulsive disorder (OCD)⁵⁴; and schizophrenia (SCZ)⁵⁵), considering medicated and unmedicated subgroups where available; ii) common variants^{56–59} associated with NPDs; and iii) 18 different CNV and aneuploidy rare variants associated with NPDs. Towards this, we aggregated multiple datasets as well as published summary statistics from ENIGMA³¹ consortium to compare case control effect sizes (Cohen's d) across 8 psychiatric disorders and associated common and

rare variants on global and regional CT and SA. Global and regional effect sizes were compared, and spatial patterns of variations were evaluated using four sets of consensus maps including: mean absolute effect size maps, significance maps, variance maps, and principal component analysis. Our findings across genetic variants increasing the risk for NPDs align with twin and SNP heritability estimates¹⁴, which are higher for surface area and sensorimotor regions, suggesting that these are general properties of the genetic architecture of the cerebral cortex. Overall, our study suggests that the neuroimaging alterations observed in NPDs are distinct from those observed across genetic variants increasing the risk for NPDs.

Brain and cortex maps were generated using the *ggseg* package in R⁷⁸. Common and rare variant illustrations are from the NIAID NIH BIOART Source (https://bioart.niaid.nih.gov/bioart/170 and https://bioart.niaid.nih.gov/bioart/204)

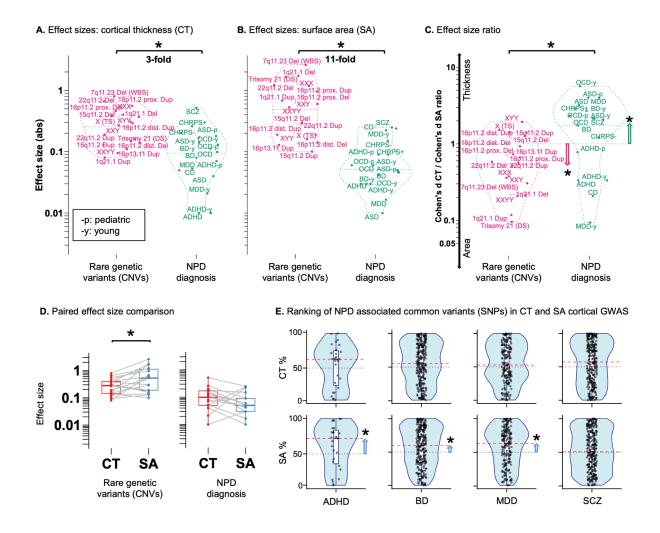


Figure 2: Global cortical differences across NPDs and associated genetic variants.

Legend: **A-D**) Comparison between neurodevelopmental and psychiatric disorders (NPDs) associated rare genetic liability (CNVs) and diagnosis on **A**) mean cortical thickness (CT) effect sizes; **B**) total surface area (SA) effect sizes; **C**) the ratio of CT and SA effect sizes; and **D**) paired CT and SA effect sizes. Case-control differences were adjusted for age, sex, and site. *: FDR significant (q<0.05), across all pairs of comparisons. Absolute effect sizes (Y-axis) are plotted on a log10 scale. **E**) Effects on CT and SA of common variants

associated with NPDs. We tested if NPD genome-wide significant SNPs were enriched in SNPs associated with SA or CT. We ranked independent NPD-associated SNPs based on their p-value association with CT and SA. Median rankings are indicated using dotted red lines. * and arrows represent significant (FDR q<0.05) median ranking compared to

permutation-based null distribution.

Abbreviations: Adult-Adolescence-Pediatric sample abbreviations: -y=young; -p=pediatric; Abs=absolute; ADHD=attention deficit hyperactivity disorder; ASD=autism spectrum disorder; BD=bipolar disorder; CD: conduct disorder; CHRPS: clinical high risk for psychosis; CHRPSn: CHR who did not develop a psychotic disorder; CHRPSp: CHR who later developed a psychotic disorder; CNV=copy number variant; CT=cortical thickness; Del=deletion; Dup=duplication; GWAS: genome-wide association study; MDD=major depressive NPD=neurodevelopmental psychiatric disorders; disorder; and OCD=obsessive-compulsive disorder; prox.=proximal; SA=surface area; SCZ=schizophrenia; SNP=single nucleotide polymorphism; TS=Turner WBS=Williams-Beuren syndrome;

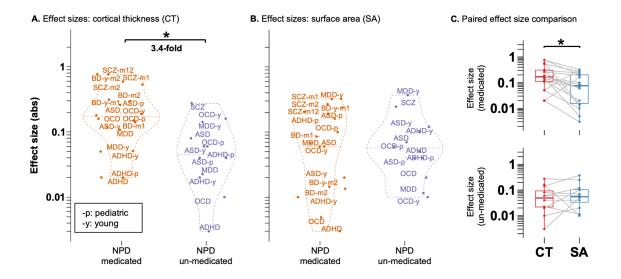


Figure 3: Global cortical differences across medication sub-groups.

Legend: **A-C**) Comparing the effect sizes on mean cortical thickness (CT) and total surface area (SA) of neurodevelopmental and psychiatric disorders (NPDs) diagnosis sub-groups with and without medications using A-B) violin plots; and C) paired boxplots. Case-control differences were adjusted for age, sex, and site. *: FDR significant (q<0.05), across all pairs of comparisons. Absolute effect sizes (Y-axis) are plotted on a log10 scale.

Abbreviations: Adult-Adolescence-Pediatric sample abbreviations: -y=young; -p=pediatric; Abs=absolute; ADHD=attention deficit hyperactivity disorder; ASD=autism spectrum disorder; BD=bipolar disorder; CD: conduct disorder; CHRPS: clinical high risk for psychosis; CHRPSn: CHR who did not develop a psychotic disorder; CHRPSp: CHR who later developed a psychotic disorder; CT=cortical thickness; Del=deletion; Dup=duplication; DZ=di-zygotic; MDD=major depressive disorder; MZ=mono-zygotic; NPD=neurodevelopmental and psychiatric disorders; OCD=obsessive-compulsive disorder; SA=surface area; SCZ=schizophrenia;

Medication abbreviations: -m=medicated; -u=un-medicated; BD-m1=lithium medication;

BD-m2=antiepileptics medication; SCZ-m1=1st generation; SCZ-m2=2nd generation; SCZ-m12=1st & 2nd generation;

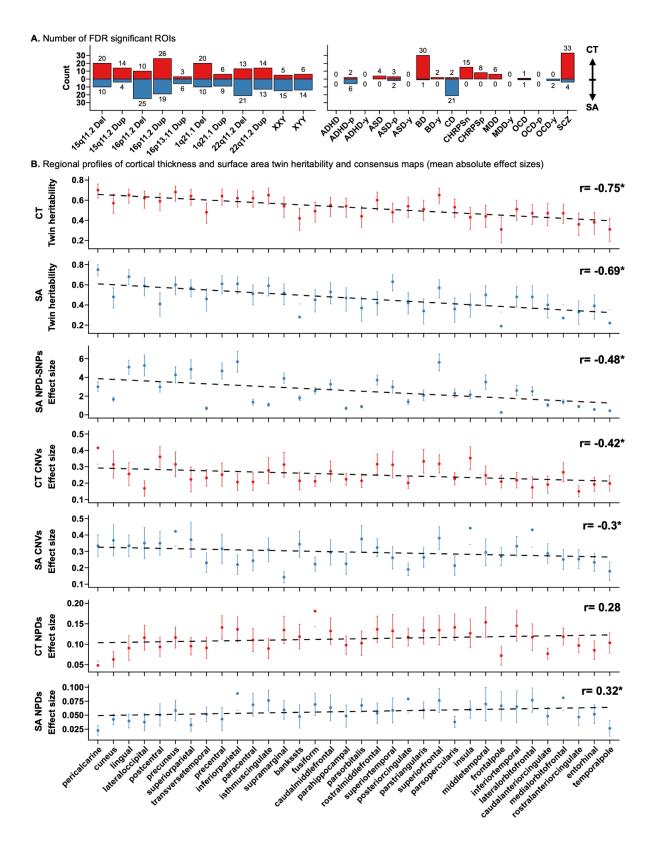


Figure 4. Consensus maps of regional cortical differences.

Legend: A) Number of FDR significant cortical regions (out of 34) per CNV/NPD for

cortical thickness (up, red) and surface area (down, blue). B) Regional profiles of twin

heritability, and mean absolute effect sizes across common and rare genetic variants and

NPDs for cortical thickness and surface area across 34 Desikan cortical regions. Each point

represents: i) First two rows: the twin heritability and 95% CI; ii) third row: mean estimate

from linear regression for NPD associated common variants (SA NPD-SNPs); and iii) bottom

four rows: mean absolute effect size (Cohen's d), with error bars showing the standard error

of the mean. Y-axis: heritability estimates or effect sizes. X-axis: cortical regions ordered

according to the cortical gradient from sensorimotor to association regions. Dotted line:

correlation with the cortical gradient. Each panel displays Pearson correlation and

*:spin-permutation significant, p-spin < 0.05.

Abbreviations, Abs=absolute; ADHD=attention deficit hyperactivity disorder; ASD=autism

spectrum disorder; BD=bipolar disorder; CD: conduct disorder; CHRPS: clinical high risk for

psychosis; CHRPSn: CHR who did not develop a psychotic disorder; CHRPSp: CHR who

later developed a psychotic disorder; CNV=copy number variant; CT=cortical thickness;

Del=deletion; Dup=duplication; MDD=major depressive disorder; NPD=neurodevelopmental

and psychiatric disorders; OCD=obsessive-compulsive disorder; SA=surface area;

33

SCZ=schizophrenia; TS=Turner syndrome;

Adult-Adolescence-Pediatric abbreviations: -y=adolescence/young; -p=pediatric).

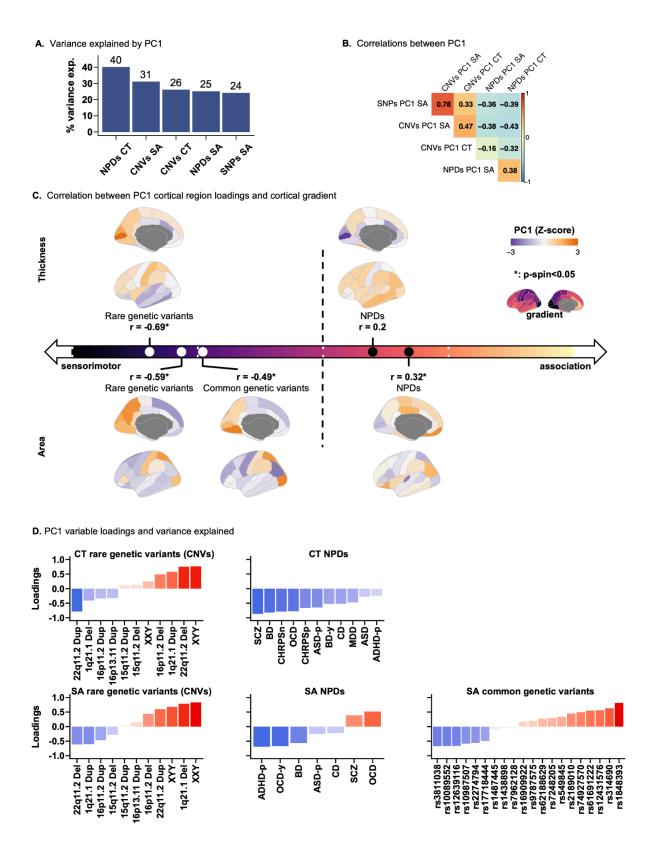


Figure 5. Latent dimensions of regional cortical differences.

Legend: **A)** The variance explained by the first principal component (PC1) for neurodevelopmental and psychiatric disorders (NPDs), their associated common and rare genetic variants. **B)** Correlation between latent dimensions. Pairwise spatial correlations between PC1 for CT and SA across NPDs, and associated common and rare genetic variants. **C)** Correlation between the cortical gradient and the latent dimension of cortical differences. Plots are arranged along the sensorimotor-association axis³⁸ based on correlation with cortical gradient (AHBA gene expression principal component). Positive and negative correlation values indicate greater similarity with association and sensorimotor cortical regions, respectively. Each plot displays Pearson correlation and *:spin-permutation significant, p-spin < 0.05. **D)** PC1 variable loadings and variance explained for NPDs and genetic variants.

Abbreviations, Abs=absolute; ADHD=attention deficit hyperactivity disorder; ASD=autism spectrum disorder; BD=bipolar disorder; CD: conduct disorder; CHRPS: clinical high risk for psychosis; CHRPSn: CHR who did not develop a psychotic disorder; CHRPSp: CHR who later developed a psychotic disorder; CNV=copy number variant; CT=cortical thickness; Del=deletion; Dup=duplication; MDD=major depressive disorder; NPD=neurodevelopmental and psychiatric disorders; OCD=obsessive-compulsive disorder; PC: principal component; p-spin: spin permutation based p-value; r: Pearson correlation; SA: surface area; SCZ=schizophrenia; SNPs: single nucleotide polymorphism; TS=Turner syndrome; % variance exp.= percentage of variance explained;

References

- 1. Brainstorm Consortium *et al.* Analysis of shared heritability in common disorders of the brain. *Science* **360**, (2018).
- 2. Sullivan, P. F., Yao, S. & Hjerling-Leffler, J. Schizophrenia genomics: genetic complexity and functional insights. *Nature Reviews Neuroscience* 1–14 (2024) doi:10.1038/s41583-024-00837-7.
- 3. American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*(DSM-5 (R)). (American Psychiatric Association Publishing, Arlington, TX, 2013).
 doi:10.1176/appi.books.9780890425596.
- 4. Grotzinger, A. D. *et al.* The landscape of shared and divergent genetic influences across 14 psychiatric disorders. *medRxiv* 2025.01.14.25320574 (2025) doi:10.1101/2025.01.14.25320574.
- 5. Polderman, T. J. C. *et al.* Meta-analysis of the heritability of human traits based on fifty years of twin studies. *Nat Genet* **47**, 702–709 (2015).
- 6. Grotzinger, A. D. *et al.* Genetic architecture of 11 major psychiatric disorders at biobehavioral, functional genomic and molecular genetic levels of analysis. *Nat. Genet.* **54**, 548–559 (2022).
- 7. Sanders, S. J. *et al.* Insights into Autism Spectrum Disorder Genomic Architecture and Biology from 71 Risk Loci. *Neuron* **87**, 1215–1233 (2015).
- 8. Marshall, C. R. *et al.* Contribution of copy number variants to schizophrenia from a genome-wide study of 41,321 subjects. *Nat. Genet.* **49**, 27–35 (2017).
- 9. Green, E. K. et al. Copy number variation in bipolar disorder. Mol. Psychiatry 21, 89–93 (2016).
- Kendall, K. M. *et al.* Association of Rare Copy Number Variants With Risk of Depression. *JAMA Psychiatry* 76, 818–825 (2019).
- 11. Halvorsen, M. *et al.* A Burden of Rare Copy Number Variants in Obsessive-Compulsive Disorder. *Res Sq* (2024) doi:10.21203/rs.3.rs-3749504/v1.
- 12. Sánchez, X. C. et al. Associations of psychiatric disorders with sex chromosome aneuploidies in

- the Danish iPSYCH2015 dataset: a case-cohort study. Lancet Psychiatry 10, 129–138 (2023).
- 13. Vaez, M. *et al.* Population-Based Risk of Psychiatric Disorders Associated With Recurrent Copy Number Variants. *JAMA Psychiatry* (2024) doi:10.1001/jamapsychiatry.2024.1453.
- 14. Grasby, K. L. et al. The genetic architecture of the human cerebral cortex. Science 367, (2020).
- Raznahan, A., Won, H., Glahn, D. C. & Jacquemont, S. Convergence and Divergence of Rare Genetic Disorders on Brain Phenotypes: A Review. *JAMA Psychiatry* (2022) doi:10.1001/jamapsychiatry.2022.1450.
- Liao, Z. et al. Copy number variants and the tangential expansion of the cerebral cortex. Nat.
 Commun. 16, 1–12 (2025).
- 17. Warrier, V. *et al.* Genetic insights into human cortical organization and development through genome-wide analyses of 2,347 neuroimaging phenotypes. *Nat. Genet.* 1–11 (2023) doi:10.1038/s41588-023-01475-y.
- 18. Makowski, C. *et al.* Discovery of genomic loci of the human cerebral cortex using genetically informed brain atlases. *Science* **375**, 522–528 (2022).
- 19. Kumar, K. *et al.* 295. Rare variant genetic architecture of the human cortical MRI phenotypes in general population. *Biol. Psychiatry* **95**, S220–S221 (2024).
- Kopal, J. et al. High-effect gene-coding variants impact cognition, mental well-being, and neighborhood safety substrates in brain morphology. bioRxiv (2024) doi:10.1101/2024.05.21.24307729.
- 21. Rakic, P. Specification of cerebral cortical areas. Science 241, 170–176 (1988).
- 22. Selemon, L. D. *et al.* Distinct abnormalities of the primate prefrontal cortex caused by ionizing radiation in early or midgestation. *J. Comp. Neurol.* **521**, 1040–1053 (2013).
- Paus, T. Population Neuroscience: Principles and advances. Curr. Top. Behav. Neurosci. 68, 3–34 (2024).
- 24. Bethlehem, R. A. I. et al. Brain charts for the human lifespan. Nature 604, 525–533 (2022).
- 25. Raznahan, A. et al. How does your cortex grow? J. Neurosci. 31, 7174–7177 (2011).

- 26. Zhiqiang Sha *et al*. The overlapping genetic architecture of psychiatric disorders and cortical brain structure. *bioRxiv* 2023.10.05.561040 (2023) doi:10.1101/2023.10.05.561040.
- 27. Stauffer, E.-M. *et al.* The genetic relationships between brain structure and schizophrenia. *Nat. Commun.* **14**, 7820 (2023).
- 28. Cheng, W. *et al.* Genetic Association Between Schizophrenia and Cortical Brain Surface Area and Thickness. *JAMA Psychiatry* (2021) doi:10.1001/jamapsychiatry.2021.1435.
- 29. Lin, B. D. *et al.* Dissecting causal relationships between cortical morphology and neuropsychiatric disorders: a bidirectional Mendelian randomization study. *bioRxiv* 2024.09.05.24313146 (2024) doi:10.1101/2024.09.05.24313146.
- 30. Larivière, S. *et al.* The ENIGMA Toolbox: multiscale neural contextualization of multisite neuroimaging datasets. *Nat. Methods* **18**, 698–700 (2021).
- 31. Thompson, P. M. *et al.* ENIGMA and global neuroscience: A decade of large-scale studies of the brain in health and disease across more than 40 countries. *Transl. Psychiatry* **10**, 100 (2020).
- 32. Opel, N. *et al.* Cross-Disorder Analysis of Brain Structural Abnormalities in Six Major

 Psychiatric Disorders: A Secondary Analysis of Mega- and Meta-analytical Findings From the

 ENIGMA Consortium. *Biol. Psychiatry* **88**, 678–686 (2020).
- 33. Hettwer, M. D. *et al.* Coordinated cortical thickness alterations across six neurodevelopmental and psychiatric disorders. *Nat. Commun.* **13**, 6851 (2022).
- 34. Patel, Y. *et al.* Virtual histology of cortical thickness and shared neurobiology in 6 psychiatric disorders. *JAMA Psychiatry* **78**, 47–63 (2021).
- 35. Patel, Y. *et al.* Virtual Ontogeny of Cortical Growth Preceding Mental Illness. *Biol. Psychiatry* (2022) doi:10.1016/j.biopsych.2022.02.959.
- 36. Cao, Z. *et al.* Cortical profiles of numerous psychiatric disorders and normal development share a common pattern. *Mol. Psychiatry* (2022) doi:10.1038/s41380-022-01855-6.
- 37. Levitis, E. *et al.* The variegation of human brain vulnerability to rare genetic disorders and convergence with behaviorally defined disorders. *Biol. Psychiatry* (2023)

- doi:10.1016/j.biopsych.2023.07.008.
- 38. Sydnor, V. J. *et al.* Neurodevelopment of the association cortices: Patterns, mechanisms, and implications for psychopathology. *Neuron* Preprint at https://doi.org/10.1016/j.neuron.2021.06.016 (2021).
- 39. Markello, R. D. *et al.* neuromaps: structural and functional interpretation of brain maps. *Nat. Methods* **19**, 1472–1479 (2022).
- 40. Burt, J. B. *et al.* Hierarchy of transcriptomic specialization across human cortex captured by structural neuroimaging topography. *Nat. Neurosci.* **21**, 1251–1259 (2018).
- 41. Modenato, C. *et al.* Effects of eight neuropsychiatric copy number variants on human brain structure. *Transl. Psychiatry* **11**, 399 (2021).
- 42. Sønderby, I. E. *et al.* Effects of copy number variations on brain structure and risk for psychiatric illness: Large-scale studies from the ENIGMA working groups on CNVs. *Hum. Brain Mapp.* **43**, 300–328 (2021).
- 43. Modenato, C. *et al.* Lessons learnt from neuroimaging studies of Copy Number Variants, a systematic review. *Biol. Psychiatry* (2021) doi:10.1016/j.biopsych.2021.05.028.
- 44. Kopal, J. *et al.* Rare CNVs and phenome-wide profiling highlight brain structural divergence and phenotypical convergence. *Nat Hum Behav* 7, 1001–1017 (2023).
- 45. Sun, D. *et al.* Large-scale mapping of cortical alterations in 22q11.2 deletion syndrome: Convergence with idiopathic psychosis and effects of deletion size. *Mol. Psychiatry* **25**, 1822–1834 (2020).
- 46. Caseras, X. *et al.* Effects of genomic copy number variants penetrant for schizophrenia on cortical thickness and surface area in healthy individuals: analysis of the UK Biobank. *Br. J. Psychiatry* **218**, 104–111 (2021).
- 47. Hoogman, M. *et al.* Brain Imaging of the Cortex in ADHD: A Coordinated Analysis of Large-Scale Clinical and Population-Based Samples. *Am. J. Psychiatry* **176**, 531–542 (2019).
- 48. van Rooij, D. et al. Cortical and subcortical brain morphometry differences between patients

- with autism spectrum disorder and healthy individuals across the lifespan: Results from the ENIGMA ASD working group. *Am. J. Psychiatry* **175**, 359–369 (2018).
- 49. Boedhoe, P. S. W. *et al.* Subcortical Brain Volume, Regional Cortical Thickness, and Cortical Surface Area Across Disorders: Findings From the ENIGMA ADHD, ASD, and OCD Working Groups. *Am. J. Psychiatry* **177**, 834–843 (2020).
- 50. Hibar, D. P. *et al.* Cortical abnormalities in bipolar disorder: an MRI analysis of 6503 individuals from the ENIGMA Bipolar Disorder Working Group. *Mol. Psychiatry* **23**, 932–942 (2018).
- 51. ENIGMA Clinical High Risk for Psychosis Working Group *et al.* Association of structural magnetic resonance imaging measures with psychosis onset in individuals at Clinical High Risk for developing psychosis: An ENIGMA working group mega-analysis: An ENIGMA working group mega-analysis. *JAMA Psychiatry* **78**, 753–766 (2021).
- 52. Gao, Y., Staginnus, M. & ENIGMA-Antisocial Behavior Working Group. Cortical structure and subcortical volumes in conduct disorder: a coordinated analysis of 15 international cohorts from the ENIGMA-Antisocial Behavior Working Group. *Lancet Psychiatry* 11, 620–632 (2024).
- 53. Schmaal, L. *et al.* Cortical abnormalities in adults and adolescents with major depression based on brain scans from 20 cohorts worldwide in the ENIGMA Major Depressive Disorder Working Group. *Mol. Psychiatry* **22**, 900–909 (2017).
- 54. Boedhoe, P. S. W. *et al.* Cortical abnormalities associated with pediatric and adult obsessive-compulsive disorder: Findings from the ENIGMA obsessive-compulsive disorder working group. *Am. J. Psychiatry* **175**, 453–462 (2018).
- 55. van Erp, T. G. M. *et al.* Cortical Brain Abnormalities in 4474 Individuals With Schizophrenia and 5098 Control Subjects via the Enhancing Neuro Imaging Genetics Through Meta Analysis (ENIGMA) Consortium. *Biol. Psychiatry* **84**, 644–654 (2018).
- 56. O'Connell, K. S. *et al.* Genomics yields biological and phenotypic insights into bipolar disorder.

 Nature 1–12 (2025) doi:10.1038/s41586-024-08468-9.
- 57. Genome-wide analyses of ADHD identify 27 risk loci, refine the genetic architecture and

- implicate several cognitive domains. *Nature* https://www.nature.com/articles/s41588-022-01285-8.
- 58. Meng, X. *et al.* Multi-ancestry genome-wide association study of major depression aids locus discovery, fine mapping, gene prioritization and causal inference. *Nat. Genet.* **56**, 222–233 (2024).
- 59. Trubetskoy, V. *et al.* Mapping genomic loci implicates genes and synaptic biology in schizophrenia. *Nature* 1–13 (2022) doi:10.1038/s41586-022-04434-5.
- 60. Zimmerman, M., Morgan, T. A. & Stanton, K. The severity of psychiatric disorders: World Psychiatry. *World Psychiatry* **17**, 258–275 (2018).
- 61. de Zwarte, S. M. C. *et al.* The association between familial risk and brain abnormalities is disease specific: An ENIGMA-relatives study of schizophrenia and bipolar disorder. *Biol. Psychiatry* **86**, 545–556 (2019).
- 62. Voineskos, A. N. *et al.* Effects of antipsychotic medication on brain structure in patients with major depressive disorder and psychotic features: Neuroimaging findings in the context of a randomized placebo-controlled clinical trial: Neuroimaging findings in the context of a randomized placebo-controlled clinical trial. *JAMA Psychiatry* 77, 674–683 (2020).
- 63. Norbom, L. B. *et al.* New insights into the dynamic development of the cerebral cortex in childhood and adolescence: Integrating macro- and microstructural MRI findings. *Prog. Neurobiol.* **204**, 102109 (2021).
- 64. Paus, T., Keshavan, M. & Giedd, J. N. Why do many psychiatric disorders emerge during adolescence? *Nat. Rev. Neurosci.* **9**, 947–957 (2008).
- 65. Parker, N. et al. Assessment of Neurobiological Mechanisms of Cortical Thinning During Childhood and Adolescence and Their Implications for Psychiatric Disorders. JAMA Psychiatry 77, 1127–1136 (2020).
- 66. Gandal, M. J. *et al.* Broad transcriptomic dysregulation occurs across the cerebral cortex in ASD.

 Nature 1–8 (2022) doi:10.1038/s41586-022-05377-7.

- 67. Kumar, K. *et al.* Subcortical Brain Alterations in Carriers of Genomic Copy Number Variants. *Am. J. Psychiatry* **180**, 685–698 (2023).
- 68. Lenroot, R. K., Lee, N. R. & Giedd, J. N. Effects of sex chromosome aneuploidies on brain development: evidence from neuroimaging studies. *Dev. Disabil. Res. Rev.* **15**, 318–327 (2009).
- 69. Desikan, R. S. *et al.* An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage* **31**, 968–980 (2006).
- 70. Benjamini, Y. & Hochberg, Y. Controlling the false discovery rate: A practical and powerful approach to multiple testing. *J. R. Stat. Soc.* **57**, 289–300 (1995).
- 71. Hansen, J. Y. *et al.* Local molecular and global connectomic contributions to cross-disorder cortical abnormalities. *Nat. Commun.* **13**, 4682 (2022).
- 72. Raznahan, A. *et al.* Globally Divergent but Locally Convergent X- and Y-Chromosome Influences on Cortical Development. *Cereb. Cortex* **26**, 70–79 (2016).
- 73. Seidlitz, J. *et al.* Transcriptomic and cellular decoding of regional brain vulnerability to neurogenetic disorders. *Nat. Commun.* **11**, 1–14 (2020).
- 74. Green, T. *et al.* Surface-based morphometry reveals distinct cortical thickness and surface area profiles in Williams syndrome. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **171B**, 402–413 (2016).
- 75. Sønderby, I. E. *et al.* Dose response of the 16p11.2 distal copy number variant on intracranial volume and basal ganglia. *Mol. Psychiatry* **25**, 584–602 (2020).
- Markello, R. D. & Misic, B. Comparing spatial null models for brain maps. *Neuroimage* 236, 118052 (2021).
- 77. Alexander-Bloch, A. F. *et al.* On testing for spatial correspondence between maps of human brain structure and function. *Neuroimage* **178**, 540–551 (2018).
- 78. Mowinckel, A. M. & Vidal-Piñeiro, D. Visualization of brain statistics with R packages ggseg and ggseg3d. *Adv. Methods Pract. Psychol. Sci.* **3**, 466–483 (2020).
- 79. Lê, S., Josse, J. & Husson, F. FactoMineR: AnRPackage for Multivariate Analysis. J. Stat. Softw.

25, 1–18 (2008).

80. Thompson, D. J. *et al.* UK Biobank release and systematic evaluation of optimised polygenic risk scores for 53 diseases and quantitative traits. *Genetic and Genomic Medicine* (2022).