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High minimum inhibitory concentration of imipenem as a predictor of fatal outcome in patients with carbapenem non-susceptible *Klebsiella pneumoniae*

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Carbapenem resistance in *Klebsiella pneumoniae* is important because of its increasing prevalence and limited therapeutic options. To investigate the clinical and microbiological characteristics of patients infected or colonized with carbapenem non-susceptible *K. pneumoniae* (CnsKP) in Taiwan, we conducted a retrospective study at Taipei Veterans General Hospital from January 2012 to November 2013. Carbapenem non-susceptibility was defined as a minimum inhibitory concentration (MIC) of ≥ 2 mg/L for imipenem or meropenem. A total of 105 cases with CnsKP were identified: 49 patients with infection and 56 patients with colonization. Thirty-one isolates had genes that encoded carbapenemases (29.5%), including *K. pneumoniae* carbapenemase (KPC)-2 (n = 27), KPC-3 (n = 1), VIM-1 (n = 1) and IMP-8 (n = 2). The in-hospital mortality among patients with CnsKP was 43.8%. A MIC for imipenem ≥ 16 μ g/mL, nasogastric intubation and Acute Physiology and Chronic Health Evaluation II score were independent risk factors for in-hospital mortality for all patients with CnsKP. A MIC for imipenem ≥ 16 μ g/mL was also an independent risk factor for 14-day mortality in patients with CnsKP. In conclusion, a positive culture for CnsKP was associated with high in-hospital mortality. A high imipenem MIC of CnsKP can predispose a patient to a poor prognosis.

Klebsiella pneumoniae is an important causative agent of nosocomial and community-acquired Gram-negative bacteremia. It can cause various infections, including pneumonia, urinary tract infections and intra-abdominal infections^{1,2}. *K. pneumoniae* is also the major pathogenic organism of community-onset pyogenic infection in Taiwan and other Asian countries^{3–6}. *K. pneumoniae* strains possessing extended spectrum β -lactamases (ESBL) conferring multidrug resistance have been reported worldwide. Carbapenems have been used extensively for severe infections arising from ESBL-producing *Enterobacteriaceae*, thereby imposing selection pressure for carbapenem resistance⁷. One of the major carbapenem resistance mechanisms of *K. pneumoniae* is the acquisition of carbapenemase genes that encode enzymes that are able to hydrolyze carbapenems. Another mechanism is the deficiency of outer-membrane porin expression with an overexpression of β -lactamases that possess very weak affinity for carbapenems⁸.

Over the past decade, the prevalence of carbapenem resistance among *K. pneumoniae* has increased dramatically worldwide⁹. The mortality attributable to carbapenem-resistant *K. pneumoniae* infection varies between 26% and 44%, with the highest mortality reported in patients with bacteremia¹⁰. Studies focusing on

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carbapenem-resistant *K. pneumoniae* have derived primarily from carbapenemase-producing *K. pneumoniae*, especially *K. pneumoniae* carbapenemase (KPC)-producing *K. pneumoniae*^{7,10}. The features of patients with non-carbapenemase producing carbapenem-resistant *K. pneumoniae* have rarely been reported¹¹. In clinical practice, distinguishing colonization from infection is difficult, especially when isolates are collected from non-sterile sites such as the respiratory tract, urine or wounds. The clear features and outcomes of patients (whether infection or colonization) from whom carbapenem-resistant *K. pneumoniae* was isolated, regardless of the resistance mechanisms, have also rarely been reported.

To address this gap in the knowledge, we conducted an observational study to investigate the clinical and microbiological features of patients with carbapenem non-susceptible *K. pneumoniae* (CnsKP) in Taiwan. Our attention was particularly focused on the mortality related to the minimum inhibitory concentration (MIC) levels of imipenem.

Methods

Setting, study design and patients. This retrospective study was conducted at Taipei Veterans General Hospital (a 2900-bed and tertiary-care teaching hospital in Taiwan) from January 2012 to November 2013. All isolates were from clinical cultures during the study period. Surveillance cultures were not conducted during this period. During this 23-month study period, clinical information was collected from all consecutive patients in whom different specimens revealed isolated *K. pneumoniae* that showed non-susceptibility to carbapenem. Resistance to carbapenem was defined as a MIC of ≥ 2 mg/L for imipenem or meropenem according to the interpretative criteria from the Clinical and Laboratory Standards Institute (CLSI) guidelines¹². Patients under the age of 20 and those with incomplete medical records were excluded. The protocol was approved by the Institution Review Board of Taipei Veterans General Hospital, and informed consent was waived by Institutional Review Board.

Definitions. Infection or colonization and the probable infectious source were determined on the basis of microbiological results and on the judgments of two infectious disease specialists. The infection and colonization of CnsKP were defined as previously described¹³. In summary, an infection was confirmed by a positive culture for CnsKP isolated from blood or any other sterile source. Positive respiratory cultures were defined according to the criteria by the American Thoracic Society and the Infectious Diseases Society of America¹⁴. The criteria outlined by the Centers for Disease Control and Prevention National Healthcare Safety Network were used for patients with positive cultures from urine or surgical wounds¹⁵. Positive cultures for CnsKP from non-surgical wounds were deemed to be infected only if infection was documented in their medical records. All other culture episodes were defined as colonizations. Isolates from index cultures that had been collected more than 48 h after admission to hospital were defined as “hospital-acquired”. The isolates were defined as “healthcare-associated” if the patients lived in a nursing home or long-term care facility, had ever been admitted to a hospital for ≥ 2 days during the previous 90 days, had ever undergone renal dialysis in a hospital or clinic during the previous 30 days or had ever received intravenous therapy at home or in an outpatient clinic during the previous 30 days³. The Acute Physiology and Chronic Health Evaluation II (APACHE II) score at the time of onset of infection was used to assess the severity of illness, as described previously¹⁶. For patients with colonization by CnsKP, the APACHE II score was also calculated within 24 h before the positive cultures. An “immunocompromised” condition included the presence of neutropenia, human immunodeficiency virus infection or steroid therapy (≥ 20 mg of prednisone or equivalent per day for ≥ 1 month) or other immunosuppressive therapy during the 30 days before the isolation of CnsKP. Treatment with at least one antibiotic agent for ≥ 48 h to which the isolate was susceptible *in vitro* according to the interpretative criteria from the CLSI guidelines¹² was defined as “appropriate therapy”. Tigecycline was considered “appropriate therapy” for bloodstream or urinary tract infection if the tigecycline MIC was ≤ 0.5 mg/L with standard dosing¹⁷.

Predictors of mortality. We compared the clinical variables between survivors and non-survivors to investigate the risk factors for mortality in patients with CnsKP. The variables reviewed included infection or colonization, demographic characteristics, MIC value for imipenem in CnsKP, history of various underlying diseases, immunocompromised state, presence of indwelling devices at the time of CnsKP isolation, any type of surgery in the previous 30 days, APACHE II score, length of stay in the intensive care unit (ICU) at the time of isolation of CnsKP and previous hospitalization. The appropriate use of antibiotics was analyzed in patients with infection.

Microbiologic methods. The identification and antimicrobial susceptibility testing of bacterial strains was performed using a Vitek 2 automated system (bioMérieux, Marcy l’Etoile, France). MICs were classified according to the CLSI breakpoints¹², except those for colistin and tigecycline. For colistin, susceptibility was defined based on the European Committee on Antimicrobial Susceptibility Testing (susceptible, MIC ≤ 2 mg/L; resistant, MIC > 2 mg/L) as described previously¹⁸. Susceptibility to tigecycline was defined according to the recommendation by the US Food and Drug Administration (FDA) (susceptible, MIC ≤ 2 mg/L; resistant, MIC ≥ 8 mg/L), as described previously¹⁸.

Polymerase chain reaction (PCR) was used for all isolates to detect carbapenemase genes (encoding Ambler class-A families NMC, IMI, SME, KPC and GES; Ambler class-B families IMP, VIM, GIM, SIM, SPM and NDM; and Ambler class-D family OXA-48-type), ESBL genes (encoding CTX-M, SHV and TEM) and plasmid-borne AmpC-like genes (encoding CMY and DHA) as described previously^{18,19}. Identification of outer-membrane porins (OMP35 and OMP36) was carried out as described previously¹⁸.

Statistical analyses. Discrete variables were analyzed using the chi-square test or Fisher’s exact test. Continuous variables were compared using Student’s *t*-test or Mann–Whitney rank sum test. The independent risk factors for 14-day and in-hospital mortality were explored by logistic regression models. Univariate analyses

| Antimicrobial agent | MIC range ($\mu\text{g/mL}$) | MIC ₅₀ ^a ($\mu\text{g/mL}$) | MIC ₉₀ ^b ($\mu\text{g/mL}$) | No. (%) of isolates susceptible |
|-------------------------|--------------------------------|---|---|---------------------------------|
| Ciprofloxacin | ≤ 0.25 to ≥ 4 | ≥ 4 | ≥ 4 | 4 (3.8) |
| Levofloxacin | ≤ 0.12 to ≥ 8 | ≥ 8 | ≥ 8 | 4 (3.8) |
| Ampicillin-sulbactam | ≥ 32 | ≥ 32 | ≥ 32 | 0 (0) |
| Piperacillin-tazobactam | ≥ 128 | ≥ 128 | ≥ 128 | 0 (0) |
| Cefazolin | ≥ 64 | ≥ 64 | ≥ 64 | 0 (0) |
| Ceftriaxone | 2 to ≥ 64 | ≥ 64 | ≥ 64 | 0 (0) |
| Ceftazidime | 16 to ≥ 64 | ≥ 64 | ≥ 64 | 0 (0) |
| Cefepime | ≤ 1 to ≥ 64 | ≥ 64 | ≥ 64 | 12 (11.4) |
| Amikacin | ≤ 2 to ≥ 64 | ≤ 2 | ≥ 64 | 84 (80) |
| Gentamicin | ≤ 1 to ≥ 16 | ≤ 1 | ≥ 16 | 59 (56.2) |
| Ertapenem | 2 to ≥ 8 | ≥ 8 | ≥ 8 | 0 (0) |
| Imipenem | 1 to ≥ 16 | 8 | ≥ 16 | 1 (0.9) |
| Colistin | ≤ 0.5 to ≥ 16 | ≤ 0.5 | 2 | 93 (88.6) |
| Tigecycline | ≤ 0.5 to ≥ 8 | 2 | 4 | 86 (81.9) |

Table 1. *In vitro* activities of the antimicrobial agents tested against 105 CnsKP isolates. CnsKP: carbapenem non-susceptible *Klebsiella pneumoniae*; MIC: minimum inhibitory concentration. ^aMIC₅₀: MIC for 50% of isolates. ^bMIC₉₀: MIC for 90% of isolates.

were performed separately for each risk-factor variable to determine the odds ratio (OR) and 95% confidence interval (CI). All biologically plausible variables with $p \leq 0.20$ in the univariate analysis were included in the logistic regression model for the multivariate analysis. A backward selection process was utilized.

Results

Microbiological characteristics of CnsKP isolates. During the study period, 105 patients had isolates of a CnsKP strain in the following clinical samples: sputum ($n = 41$), pleural effusion ($n = 1$), urine ($n = 39$), central venous catheter tip ($n = 3$), blood ($n = 10$), wound ($n = 6$) and bile ($n = 5$). Thirty-one isolates (29.5%) had genes that encoded carbapenemases, including KPC-2 ($n = 27$), KPC-3 ($n = 1$), VIM-1 ($n = 1$) and IMP-8 ($n = 2$). The most common mechanism of carbapenem resistance, as identified in 74 cases, was the production of AmpC-mediated β -lactamases or ESBL plus porin defects. Among the 74 isolates, 55 exhibited Amp-C β -lactamase (all DHA-1). Among the isolates with genes encoding ESBLs, 38 isolates exhibited the CTX-M-9 group, and 12 isolates exhibited the CTX-M-1 group. Forty-nine isolates harbored the SHV-type ESBL genes (SHV-2, SHV-5, SHV-12, SHV-31 and SHV-120). The clinical data suggested no epidemiological links between these isolates.

The MIC ranges, MIC₅₀ values and MIC₉₀ values of various antimicrobial agents against the CnsKP isolates are listed in Table 1. The MIC of imipenem was $\geq 16 \mu\text{g/mL}$ for 44 (41.9%) isolates, $8 \mu\text{g/mL}$ for 18 (17.1%) isolates, $4 \mu\text{g/mL}$ for 23 (21.9%) isolates and $2 \mu\text{g/mL}$ for 19 (18.6%) isolates. Most isolates were susceptible to colistin ($n = 93$, 88.6%), tigecycline ($n = 86$, 81.9%), and amikacin ($n = 84$, 80%), but they showed only moderate susceptibility to gentamicin ($n = 59$, 56.2%).

Characteristics of patients with CnsKP. A total of 105 cases with CnsKP were identified during the study period. The 14-day mortality associated with CnsKP was 20%. The overall in-hospital mortality associated with CnsKP was 43.8%. Infection was diagnosed in 49 patients, and colonization was established in the remaining 56 patients. The 14-day mortality was significantly higher in patients with infection than in those with colonization (34.7% vs. 7.1%, $p < 0.001$). The overall in-hospital mortality was also significantly higher in patients with infection than in those with colonization (63.3% vs. 26.8%, $P < 0.001$).

We further compared the clinical features between patients infected with CnsKP and those colonized with CnsKP (Table 2). Patients with infection had a greater prevalence of fatal outcomes than did those without infection. The Charlson Comorbidity Index and APACHE II scores were significantly higher for patients with infection than for those with colonization. Patients with colonization had a higher rate of previous antimicrobial therapy than did those with infection. For the antimicrobial susceptibility profiles, the prevalence of imipenem MIC $\geq 16 \mu\text{g/mL}$ tended to be higher in the infection group (51.0%) than in the colonization group (33.9%) ($p = 0.077$).

Risk factors for mortality in patients with CnsKP. To clarify the influence of clinical and microbiological factors on both 14-day and in-hospital mortality, multivariate logistic regression analyses were conducted for the entire cohort to identify independent risk factors for mortality (Tables 3 and 4). All biologically plausible variables with $p \leq 0.20$ in the univariate analysis were considered for inclusion in the logistic regression model for multivariate analysis. Septic shock, appropriate antibiotics and clinical syndromes were not included because we enrolled patients with infection and colonization. Among the 105 patients, isolation of a CnsKP with imipenem MIC $\geq 16 \mu\text{g/mL}$ (OR, 3.17; 95% CI, 1.11–9.05; $p = 0.032$) and APACHE II score (OR, 1.24; 95% CI, 1.13–1.36; $p < 0.001$) were independent risk factors for 14-day mortality (Table 3). Isolation of a CnsKP with imipenem MIC $\geq 16 \mu\text{g/mL}$ (OR, 5.60; 95% CI, 1.39–22.50; $p = 0.015$), nasogastric intubation (OR, 6.19; 95% CI, 1.07–9.05; $p = 0.042$) and APACHE II score (OR, 1.13; 95% CI, 1.06–1.21; $p < 0.001$) were independent risk factors for in-hospital mortality (Table 4).

| Variable | CnsKP infection (n = 49) | CnsKP colonization (n = 56) | p |
|---|-----------------------------|-----------------------------------|--------|
| Demographics | | | |
| Age, years, mean \pm SD | 79.7 \pm 12.0 | 75.8 \pm 14.7 | 0.140 |
| Male sex | 33 (67.3) | 39 (69.6) | 0.800 |
| Nosocomial-acquired isolate | 37 (75.5) | 48 (85.7) | 0.184 |
| Microbiological characteristics of CnsKP | | | |
| Carbapenemase-producing isolates | 17 (34.7) | 14 (25.0) | 0.277 |
| Imipenem MIC \geq 4 | 39 (79.6) | 46 (82.1) | 0.740 |
| Imipenem MIC \geq 8 | 30 (61.2) | 32 (57.1) | 0.671 |
| Imipenem MIC \geq 16 | 25 (51.0) | 19 (33.9) | 0.077 |
| Polymicrobial isolation | 22 (44.9) | 19 (33.9) | 0.250 |
| Comorbidities | | | |
| Diabetes mellitus | 27 (55.1) | 23 (41.1) | 0.151 |
| COPD | 33 (67.3) | 31 (55.4) | 0.724 |
| Heart failure | 14 (28.6) | 16 (28.6) | 1.000 |
| Cerebrovascular disease | 16 (32.7) | 11 (19.6) | 0.128 |
| Chronic renal failure | 18 (58.1) | 46 (62.2) | 0.695 |
| Liver cirrhosis | 2 (4.1) | 5 (8.9) | 0.321 |
| Malignancy | 16 (32.7) | 11 (19.6) | 0.128 |
| Immunocompromised state | 8 (16.3) | 13 (23.2) | 0.379 |
| Charlson Comorbidity Index, mean \pm SD | 4.9 \pm 2.5 | 3.6 \pm 2.5 | 0.009 |
| Indwelling central venous catheter | 20 (40.8) | 18 (32.1) | 0.356 |
| Indwelling urinary catheter | 31 (63.3) | 34 (60.7) | 0.788 |
| Nasogastric tube | 38 (77.6) | 42 (75.0) | 0.759 |
| Surgical drainage | 8 (16.3) | 11 (19.6) | 0.660 |
| Mechanically ventilated at isolation | 17 (34.7) | 14 (25.0) | 0.277 |
| Renal dialysis at isolation | 13 (26.5) | 8 (14.3) | 0.118 |
| Previous surgery ^a | 15 (30.6) | 16 (28.6) | 0.819 |
| Prior use of antimicrobials ^b | | | |
| Any | 43 (87.8) | 55 (98.2) | 0.048 |
| β -lactams plus β -lactamase inhibitors | 13 (26.5) | 19 (33.9) | 0.411 |
| Fluoroquinolones | 20 (40.8) | 26 (46.4) | 0.563 |
| Carbapenems | 20 (40.8) | 28 (50.0) | 0.346 |
| Third- and fourth-generation cephalosporins | 19 (38.8) | 20 (35.7) | 0.746 |
| Glycopeptides | 20 (40.8) | 16 (28.6) | 0.187 |
| Aminoglycosides | 1 (2.0) | 2 (3.6) | 1.000 |
| Metronidazole | 7 (14.3) | 8 (14.3) | 1.000 |
| Colistin | 10 (20.4) | 11 (19.6) | 0.922 |
| Tigecycline | 15 (30.6) | 11 (19.6) | 0.194 |
| APACHE II score, mean \pm SD | 25.4 \pm 9.4 | 17.6 \pm 8.2 | <0.001 |
| 14-day mortality ^c | 17 (34.7) | 4 (7.1) | <0.001 |
| In-hospital death | 31 (63.3) | 15 (26.8) | <0.001 |

Table 2. Characteristics of patients with CnsKP infection or colonization. Data are the number (%) unless specified otherwise. APACHE, Acute Physiology and Chronic Health Evaluation; BSI, bloodstream infection; CI, confidence interval; CnsKP: carbapenem non-susceptible *Klebsiella pneumoniae*; COPD, chronic obstructive pulmonary disease; IQR, interquartile range; LOS, length of hospital stay; OR, odds ratio; SD, standard deviation. ^aDuring the 3 months preceding BSI onset. ^bDuring the 30 days preceding BSI onset. ^cDeath within 14 days of the isolation of CnsKP.

We further evaluated the 49 patients infected with CnsKP. The 14-day mortality for cases with infections was 34.7% (17/49). All biologically plausible variables with $P \leq 0.20$ in the univariate analysis were considered for inclusion in the logistic regression model for multivariate analysis. Multivariate logistic regression analyses showed that MIC for imipenem $\geq 16 \mu\text{g/mL}$ (OR, 21.75; 95% CI, 1.47–321.55; $p = 0.025$), bacteremia (OR, 15.09; 95% CI, 1.24–183.56; $p = 0.033$) and APACHE II score (OR, 1.35; 95% CI, 1.10–1.65; $p = 0.005$) were independent risk factors for 14-day mortality (Table 5).

| Variable | Univariate analysis | | Multivariable analysis | |
|--------------------------------------|---------------------|----------|------------------------|----------|
| | OR (95% CI) | <i>p</i> | OR (95% CI) | <i>p</i> |
| Infection | 6.90 (2.13–22.36) | 0.001 | | |
| Carbapenemase-producing CnsKP | 2.73 (1.02–7.33) | 0.047 | | |
| Imipenem MIC ≥ 4 | 5.85 (0.74–46.44) | 0.095 | | |
| Imipenem MIC ≥ 8 | 2.64 (0.89–7.88) | 0.081 | | |
| Imipenem MIC ≥ 16 | 3.60 (1.31–9.90) | 0.013 | 5.60 (1.39–22.50) | 0.015 |
| Diabetes mellitus | 2.06 (0.77–5.50) | 0.147 | | |
| Chronic renal failure | 2.40 (0.86–7.66) | 0.117 | | |
| Charlson Comorbidity Index | 1.15 (0.96–1.39) | 0.136 | | |
| Indwelling central venous catheter | 3.84 (1.42–10.40) | 0.008 | | |
| Nasogastric tube | 3.58 (0.77–16.60) | 0.103 | | |
| Mechanically ventilated at isolation | 2.73 (1.02–7.33) | 0.047 | | |
| Renal dialysis at isolation | 2.50 (0.86–7.31) | 0.094 | | |
| APACHE II score | 1.21 (1.12–1.31) | <0.001 | 1.24 (1.13–1.36) | <0.001 |

Table 3. Logistic regression analysis of predictors for 14-day mortality among 105 patients with CnsKP isolates. Data are the number (%) unless specified otherwise. APACHE, Acute Physiology and Chronic Health Evaluation; CI, confidence interval; CnsKP: carbapenem non-susceptible *Klebsiella pneumoniae*; OR, odds ratio.

Discussion

The current study is one of the largest series of infection or colonization of CnsKP in adult patients. Carbapenemase producers accounted for $\approx 29.5\%$ of the CnsKP isolates. The 14-day mortality among patients with CnsKP was 20%. The overall in-hospital mortality associated with CnsKP was 43.8%. A MIC for imipenem $\geq 16\mu\text{g/mL}$ was an independent risk factor for both 14-day and in-hospital mortality for all patients with CnsKP.

The literature suggests that patients with carbapenem-resistant *Enterobacteriaceae* (CRE) isolated from any site, regardless of whether infection exists, are associated with poor outcomes²⁰. Won *et al.* reported that the odds of fatal outcomes for patients with infection were higher than the odds for those with colonization with regard to isolation with KPC-producing *Enterobacteriaceae* (OR, 3.8), but the difference was not significant²¹. Consistent with the literature, we found high in-hospital mortality among patients with CnsKP, regardless of infection or colonization. We also found that both 14-day and in-hospital mortality were significantly higher in infected patients than in those colonized with CnsKP. This finding suggests that positive cultures for CnsKP in clinical samples represent an emerging challenge for physicians.

It is notable that non-carbapenemase-mediated mechanisms account for the majority of *in vitro* resistance mechanisms responsible for CnsKP in Taiwan, which is consistent with previous results¹⁸. It is difficult to distinguish between infection and colonization if CRE isolates are obtained from non-sterile sites in clinical practice. Therefore, the current investigation evaluated the detailed clinical features of all patients from whom CnsKP was isolated. The current study found that more than half of the patients were colonized with CnsKP. The higher Charlson Comorbidity Index among patients with infection suggested the opportunistic nature of CnsKP and that underlying comorbidity played a major role in CnsKP infection. We found that the infection and colonization groups had both had heavy exposure to antibiotics within the previous month. This finding might suggest that antimicrobial stewardship is important in the control of CnsKP.

The significance of positive cultures for CnsKP in clinical samples is not addressed clearly. In this study, we found that positive culture for CnsKP was associated with high in-hospital mortality, regardless of colonization or infection. Because both colonization and infection were common problems in clinical practice, we tried to analyze the risk factors for mortality among these patients. In the literature, the risk factors for mortality in all patients with positive cultures for CnsKP have never been reported^{7,20}. Several studies have indicated that patients who were infected by Gram-negative bacteria with higher carbapenem MICs and who received carbapenems were associated with poorer clinical outcomes^{22–24}. Compared to previous studies, we tried to analyze all patients with positive cultures of CnsKP in this study regardless of infection or colonization. We were the first to demonstrate that an imipenem MIC of $\geq 16\mu\text{g/mL}$ was an independent risk factor for both 14-day and in-hospital mortality among these patients. This result suggested that a positive culture for CnsKP with an imipenem MIC $\geq 16\mu\text{g/mL}$ in clinical samples is of clinical significance and may suggest that the resistance level of carbapenem is as important as the presence of carbapenemase among CnsKP. It also displays that isolation of CnsKP with a high MIC value is a marker of severe underlying conditions that are associated with poor outcomes. High MIC values would also predict the limited efficacy of those drugs against CnsKP infections. Given the notable predictor for mortality observed among CnsKP isolates with high MIC values, these data suggest that MIC may play an important part in predicting outcome. One recent study suggested that antimicrobial MICs and/or molecular markers of resistance may be useful in identifying the strains that are most likely to respond to some types of antibiotics²⁵. These findings imply that the microbiological features of CnsKP play an important role in clinical outcomes.

The limitations of our study are its inherently retrospective design and the relatively limited number of occurrences of this disease. The limited numbers of cases also reduced the power of the analysis of risk factors for mortality. Evaluation of the relative efficacy of appropriate antibiotics in cases with infection was limited because

| Variable | Univariate analysis | | Multivariable analysis | |
|--------------------------------------|---------------------|----------|------------------------|----------|
| | OR (95% CI) | <i>p</i> | OR (95% CI) | <i>p</i> |
| Infection | 6.90 (2.13–22.36) | 0.001 | | |
| Nosocomial-acquired isolates | 2.80 (0.93–8.38) | 0.066 | | |
| Imipenem MIC ≥ 4 | 5.80 (1.58–21.27) | 0.008 | | |
| Imipenem MIC ≥ 8 | 2.21 (0.98–4.96) | 0.055 | | |
| Imipenem MIC ≥ 16 | 3.51 (1.56–7.92) | 0.002 | 3.17 (1.11–9.05) | 0.032 |
| Diabetes mellitus | 2.06 (0.87–4.14) | 0.108 | | |
| Chronic renal failure | 2.74 (1.19–6.30) | 0.018 | | |
| Charlson Comorbidity Index | 1.31 (1.11–1.56) | 0.002 | | |
| Indwelling central venous catheter | 2.93 (1.29–6.68) | 0.010 | | |
| Nasogastric tube | 8.52 (2.36–30.77) | 0.001 | 6.19 (1.07–9.05) | 0.042 |
| Mechanically ventilated at isolation | 4.12 (1.68–10.06) | 0.002 | | |
| Renal dialysis at isolation | 2.14 (0.80–5.73) | 0.131 | | |
| APACHE II score | 1.17 (1.10–1.25) | <0.001 | 1.13 (1.06–1.21) | <0.001 |

Table 4. Logistic regression analysis of predictors for in-hospital mortality among 105 patients with CnsKP isolates. Data are the number (%) unless specified otherwise. APACHE, Acute Physiology and Chronic Health Evaluation; CI, confidence interval; CnsKP: carbapenem non-susceptible *Klebsiella pneumoniae*; OR, odds ratio.

| Variable | Univariate analysis | | Multivariable analysis | |
|--------------------------------------|---------------------|----------|------------------------|----------|
| | OR (95% CI) | <i>p</i> | OR (95% CI) | <i>p</i> |
| Carbapenemase-producing CnsKP | 2.27 (0.67–7.73) | 0.189 | | |
| Imipenem MIC ≥ 4 | 6.26 (0.72–54.41) | 0.096 | | |
| Imipenem MIC ≥ 8 | 2.87 (0.77–10.77) | 0.117 | | |
| Imipenem MIC ≥ 16 | 3.51 (0.99–12.36) | 0.051 | 21.75 (1.47–321.55) | 0.025 |
| Pneumonia | 3.65 (1.06–12.56) | 0.040 | | |
| Urinary tract infection | 0.31 (0.08–1.31) | 0.112 | | |
| Bacteremia | 3.81 (0.90–16.19) | 0.069 | 15.09 (1.24–183.56) | 0.033 |
| Septic shock | 8.57 (2.25–32.68) | 0.002 | | |
| Polymicrobial infection | 0.37 (0.11–1.29) | 0.118 | | |
| Mechanically ventilated at isolation | 2.27 (0.67–7.73) | 0.189 | | |
| APACHE II score | 1.21 (1.08–1.36) | 0.001 | 1.35 (1.10–1.65) | 0.005 |
| Appropriate antibiotics use | 0.37 (0.11–1.26) | 0.113 | | |

Table 5. Logistic regression analysis of predictors for 14-day mortality among 49 patients infected with isolation of CnsKP. Data are the number (%) unless specified otherwise. APACHE, Acute Physiology and Chronic Health Evaluation; CI, confidence interval; CnsKP: carbapenem non-susceptible *Klebsiella pneumoniae*; OR, odds ratio; SD, standard deviation.

of the small number of patients. Validation of our findings requires future studies at other centers and in larger populations. In addition, we tried to address the significance of positive cultures for CnsKP in clinical samples in this study. However, periodical surveillance cultures were not performed among these patients. Therefore, whether a case with CnsKP infection had previous CnsKP colonization was not clear.

In conclusion, the major mechanism of the CnsKP phenotype in Taiwan was the deficiency of outer-membrane porins in association with β -lactamases such as the AmpC enzyme or ESBLs. A positive culture for CnsKP was associated with high in-hospital mortality. We were the first to demonstrate that an imipenem MIC of $\geq 16 \mu\text{g/mL}$ was an independent risk factor for both 14-day and in-hospital mortality among patients with CnsKP. Clinicians should thus consider the imipenem MIC when they manage cases with CnsKP.

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Author Contributions

Y.-T.L. contributed to study design, and data interpretation. P.-F.W. and Y.-T.L. contributed to manuscript drafting. P.-F.W., C.C., C.-F.S., Y.-T.L., Y.-J.C. and F.-D.W. contributed to data acquisition and analysis. Y.-C.C., L.K.S. and C.-P.F. contributed to the supervision of this investigation. All of the authors were involved in writing the manuscript, critically revising it for important intellectual content and approving the final version submitted for publication.

Additional Information

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