## **Supplemental information**

Hypoxic and pharmacological activation

of HIF inhibits SARS-CoV-2

infection of lung epithelial cells

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## **Supplemental Figures**

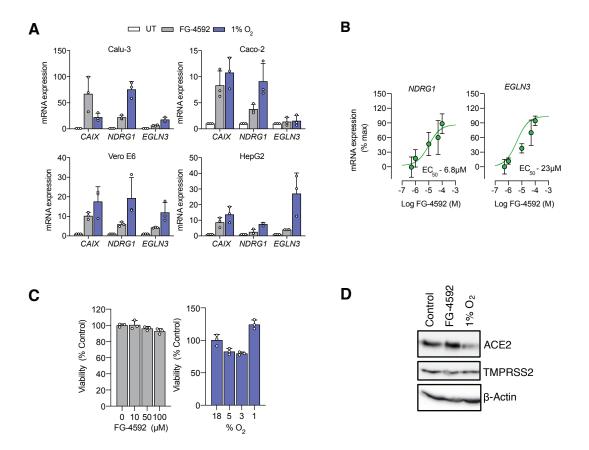
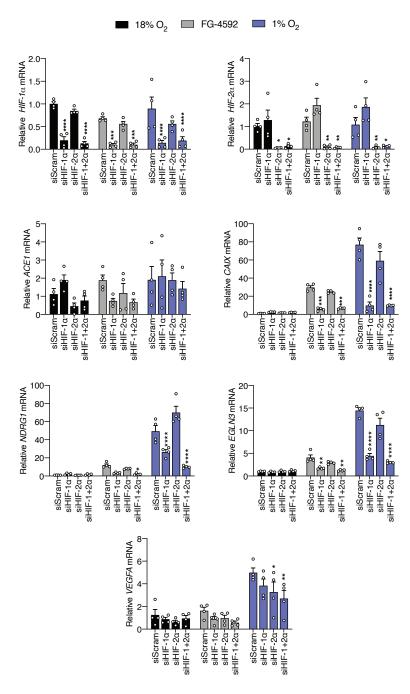


Fig.S1 Hypoxic gene induction in ACE2 expressing cells. [Related to Fig.1] (A) Calu-3, Caco-2, Vero E6 and HepG2 cells were treated with FG-4592 (50μM) or 1% O<sub>2</sub> for 24h with *CAIX*, *NDRG1*, and *EGLN3* mRNA assessed by qPCR. Data are expressed relative to normoxic untreated (UT) cells as mean  $\pm$  S.D. from n=3 biological replicates. (B) HepG2 cells were treated with increasing concentrations of FG-4592 for 24h with *NDRG1* and *EGLN3* mRNA quantified. Half-maximal effective concentration (EC<sub>50</sub>) values for both genes in response to FG-4592 treatment were calculated. (C) The impact of either FG-4592 or hypoxic incubation on the viability of Calu-3 cells was assessed through quantification of extracellular lactate dehydrogenase (LDH) 24h post-treatment. (D) HepG2 cells were treated with FG-4592 (50μM) or 1% O<sub>2</sub> for 24h and ACE2/TMPRSS2 protein expression assessed by immunoblot. β-Actin was used to show equal protein loading.



**Fig.S2:** Validating siRNA silencing of HIF-1α and HIF-2α in Calu-3 cells. [Related to Fig.2]. siRNAs targeting either HIF-1 or  $2\alpha$  were delivered into Calu-3 cells either individually or in combination along with a control scrambled siRNA. 48h post-transfection the cells were treated with FG-4592 (50μM) or 1% O<sub>2</sub> for 24h and total cellular RNA extracted. siRNA knock-down was confirmed by qPCR quantification of *HIF-1α*, *HIF-2α*, *ACE1*, *CAIX*, *NDRG1*, *EGLN3* and *VEGFA* mRNA levels. Bars represent mean  $\pm$  S.D. from n=4 biological replicates and data plotted relative to siScram at 18% O<sub>2</sub> with statistical significance determined by two-way ANOVA, \* p<0.05 \*\* p<0.01, \*\*\*\* p<0.001.

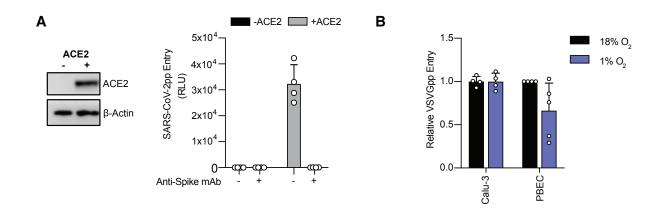


Fig.S3: SARS-CoV-2pp entry is ACE2 dependent. [Related to Fig.2]. (A) Human embryonic kidney 293T cells were transfected with a control or human ACE2 overexpression plasmid and infected with SARS-CoV-2 pseudoparticles (pp) 48h post-transfection. ACE2 expression was confirmed by immunoblot. SARS-CoV-2pp were pre-treated with or without an anti-Spike mAb Fl-3A (1µg/ml) for 30min prior to infection. Data is mean  $\pm$  S.D. from n=4 biological replicates. (B) Calu-3 or PBECs were cultured under 18% or 1%  $O_2$  for 24h before infection with viral pseudoparticles expressing vesicular stomatitis virus glycoprotein (VSV-G). Infection was assessed 48h later by quantification of luciferase activity. Data is expressed relative to the normoxic samples and is the mean  $\pm$  S.D. of n=4 biological replicates.

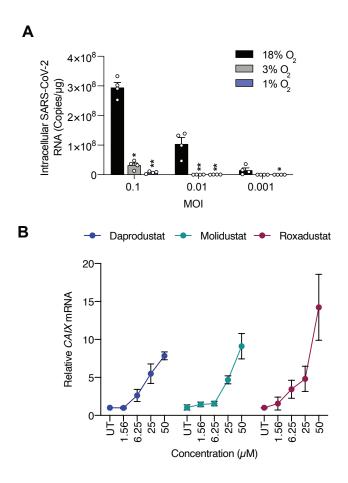
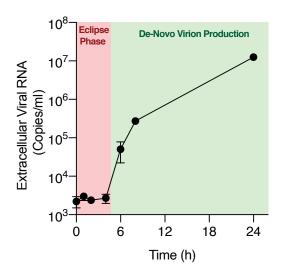


Fig.S4: Dose dependent Inhibition of SARS-CoV-2 by hypoxia and HIF prolyl hydroxylase inhibitors. [Related to Fig.2]. (A) Calu-3 cells were incubated at 18%, 3% or  $1\% O_2$  for 24h prior to infection with SARS-CoV-2 at the indicated MOIs. Viral RNA was quantified from infected cells 24h post infection. (B) Calu-3 cells were treated increasing concentrations of Daprodustat (GSK1278863), Molidustat (Bay 85-3934) or Roxadustat (FG-4592), infected with SARS-CoV-2 and *CAIX* mRNA quantified by qPCR. All data is n=4 biological replicates and presented as mean  $\pm$  S.D. with statistical significance determined by two-way ANOVA, \* p<0.05 \*\* p<0.01.



**Fig.S5. Single step growth curve of SARS-CoV-2.** [Related to Fig.4]. Calu-3 cells were inoculated with SARS-CoV-2 for 1h at an MOI of 1, unbound virus removed by washing and cells cultured in 18%  $O_2$ . At the indicated times extracellular samples were collected and SARS-CoV-2 RNA quantified by qPCR. Data is presented as mean  $\pm$  range of n=2 biological replicates.

## Supplementary Table 1: Probe sequences for quantification of SARS-CoV-2 RNA by smFISH [Related to STAR METHODS]

Positive gRNA probes
TAGATCGGCGCCGTAACTAT
TCCTTTATTACCGTTCTTAC
AGAAGAACCTTGCGGTAAGC
TACTGAATGCCTTCGAGTTC
AGCATCCGAACGTTTGATGA
TAGTAGTTGTCTGATTGTCC
GTCTTGTTGACCAACAGTTT
CTCATATTGAGTTGATGGCT
AGTAGTATGTAGCCATACTC
TCTAAATCAATGCCCAGTGG
GTAATTCAGATACTGGTTGC
CCTTTGAGTGTGAAGGTATT
GAGCAACATAAGCCCGTTAA
AGGTTGTTCTAATGGTTGTA
CATAGGGCTGTTCAAGTTGA
GCTTTTAGAGGCATGAGTAG
TGCGTGACAAATGTTTCACC
AAGGCTTTAAGTTTAGCTCC
CCCAACCGTCTCTAAGAAAC
AAGCCAATCAAGGACGGGTT
TTAGTTAGCCACTGCGAAGT
ACTGAACAACACCACCTGTA
GTAGGCCATTACAACTAGAT
AGTAGCCAAATCAGATGTGA
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TTGACGTGCCTCTGATAAGA
TGCGGGAGAAAATTGATCGT
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GGTTGTCATTAAGACCTTCG
ACAACCTATGTTAGCGCTAG
ATAGGCACACTTGTTATGGC
TCCAAAGGCAATAGTGCGAC
AAGACTATGCTCAGGTCCTA
AGTAACCACAAGTAGTGGCA
TCACACTTCATGAGAGTTGA
GCACATTTGGTTGCATTCAT
CAAAGCCACGTACGAGCACG
GGTGACGCAACTGGATAGAC
CCTTGGTTGAATAGTCTTGA
TTTCAGAACGTTCCGTGTAC
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