

Citation: Bezamat M, Cunha EJ, Modesto AM, Vieira AR, Taboas JM (2020) Aquaporin locus (12q13.12) might contribute to susceptibility of temporomandibular joint disorder associated with periodontitis. PLoS ONE 15(3): e0229245. https://doi.org/10.1371/journal.pone.0229245

Editor: JJ Cray, Jr., Ohio State University, UNITED STATES

Received: March 26, 2019

Accepted: February 3, 2020

Published: March 4, 2020

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Data Availability Statement: All relevant data are within the paper and its Supporting Information files

Funding: This work was supported by the Dental Registry and DNA Repository and the School of Dental Medicine at the University of Pittsburgh. JMT was supported by the National Institutes of Health (https://www.nih.gov/) K01 training award AR062598. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

RESEARCH ARTICLE

Aquaporin locus (12q13.12) might contribute to susceptibility of temporomandibular joint disorder associated with periodontitis

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Abstract

Aquaporins (AQPs) are membrane channels that provide for transport of water and other small molecules across the lipid bilayer of cells. Their function is essential for physiologic processes such as cell volume regulation, chondrocyte hypertrophy during appendicular skeletal growth, water reabsorption in the kidney tubules, and water excretion by the salivary glands. The ten AQP isoforms show tissue specificity and are involved in different pathologies and inflammatory diseases. This study addresses the hypothesis that arthritis, periodontitis, and temporomandibular joint disorders (TMDs) can be influenced by variation in the AQP genes at 12q13.12 locus. Salivary samples of 688 individuals were obtained from the Dental Registry and DNA Repository project at the University of Pittsburgh. Ten polymorphisms in four AQP genes (AQP1, 2, 5, and 6) were genotyped and correlated to disease status as reported by patients. Associations were found between the single nucleotide polymorphism (SNP) rs467323 in AQP2 and TMD in both genotypic (p = 0.03) and recessive (p = 0.02) models, and between rs1996315 in AQP6 and periodontitis (p = 0.05). Combined analysis of TMD and periodontitis showed an association with rs3741559 in AQP2 (p = 0.02). When conducting haplotype analysis of rs467323 and rs10875989 in AQP2, the haplotype CT showed an association with the TMD phenotype (p = 0.007). Our results suggest that the aquaporin locus at 12q13.12 may contribute to the pathogenesis of inflammatory conditions such as periodontitis and TMD. Thus, oral and skeletal health are correlated and potential susceptibility screening strategies may be developed.

Introduction

Aquaporins (AQP) are protein membrane channels expressed in numerous cell types such as endothelial and epithelial cells [1]. There are thirteen different isoforms with varying expression across cells and organs [2]. In addition to transporting water, they facilitate transport of small neutral-charge solutes, cell migration [3] and neuronal signaling [4]. Considering their

Competing interests: The authors have declared that no competing interests exist.

expression by cells of the innate and adaptive immune system [5-7], roles in inflammation and inflammatory diseases have been elucidated, including regulation of cell volume [8] and migration of neutrophils, macrophages and T cells [9-11].

AQPs apparently play a role in periodontitis, one of the biggest threats to oral health and the major cause of tooth loss [12, 13]. Periodontitis is a chronic inflammatory disease identified by the destruction of the tooth-supporting tissues including periodontal-ligament, alveolar bone, and cementum. Aqp3 deficiency is associated with greater mortality in a murine model of bacterial periodontitis [7]. Furthermore, AQP1 expression is increased in gingival tissues of individuals who have periodontitis, and expression decreases in healed periodontal mucosa [14]. Anjomshoaa et al. reported associations between polymorphisms in AQP5 and high caries experience in four different populations [15]. Because AQP5 is expressed in serous acinar cells of salivary glands, polymorphisms may impact salivary flow. A decrease in the volume of saliva might increase the adherence of plaque to dental surfaces, and thereby caries and periodontitis incidence [16]. However, the best evidence for AQP5 on dental caries susceptibility suggests a role during dental enamel formation [15,16].

Less studied are potential associations between AQP polymorphisms and inflammatory joint diseases, namely arthritis and temporomandibular joint disorders (TMDs). AQPs are expressed by chondrocytes and synoviocytes of appendicular and temporomandibular joints [17]. These cells respond to inflammatory cytokines and arthritis with changes in expression of AQPs [18–21]. Recently, conclusive evidence that Aqp4 mediates rheumatoid arthritis severity was demonstrated in a rat model [22].

AQPs may be associated with both periodontitis and degenerative joint diseases because of the common inflammatory processes underlying these. In addition, periodontitis is associated with several other systemic inflammatory conditions [23–26], including rheumatoid arthritis [27]. We hypothesized that associations between AQPs and periodontitis, arthritis, and TMDs can be better detected when individuals concomitantly affected by these three conditions are studied. AQP2, 5 and 6 are clustered on the same locus, 12q13.12, while AQP1 is the first identified constitute of the aquaporin family and the most studied AQP gene so far [28]. Therefore, we investigated the associations between periodontitis, arthritis, and TMDs and SNPs in the aquaporin locus at 12q13.12 and AQP1.

Materials and methods

Individual samples and clinical history were obtained through the Dental Registry and DNA Repository of the School of Dental Medicine, University of Pittsburgh [29–31]. The total cohort consisted of 6,092 subjects with mean age of 59.1 years (ranging from 19.4 to 90.9 years-old). This project was approved by the University of Pittsburgh Institutional Review Board (IRB # 0606091). Written informed consent documents were obtained from all subjects. We followed the STREGA guidelines for this report.

Genotypes were generated blindly to clinical diagnosis status. Genomic DNA was extracted from whole saliva using established protocols [32]. Ten polymorphisms in four *AQP* genes (*AQP1*, 2, 5, and 6) were analyzed (Table 1). PCR reactions were carried out using Taqman chemistry [33] in volumes of 3.0 μl in an ABI PRISM Sequence Detection System 7900 (Applied Biosystems, Foster City, CA, USA). The genotyping results were analyzed using SDS software version 1.7 (Applied Biosystems). PCR reactions were repeated twice when necessary. Genotypic (2df) tests were performed comparing genotypes to phenotype between affected individuals and their respective comparison group with PLINK software [34]. Additional haplotype analyses were performed utilizing Haploview software [35]. All 10 markers were consistent with Hardy-Weinberg equilibrium (Chi-squared p-values greater than 0.001) [34].

Table 1. Details of the SNPs investigated in this study.

Chromosome	Gene	SNP marker	Base Position	Location	Consequence	Base change	
7	AQP1 rs17		30,919,387	intron	Intron Variant	CT	
12	AQP2	rs467323	rs467323 49,955,982 intron 3 Prime UTR Variant		3 Prime UTR Variant	CT	
12	AQP2	rs3741559 49,951,193 intron Intron Variant		AG			
12	AQP2	rs461872	49,951,423	intron	Intron Variant	AG	
12	AQP2	rs2878771	49,958,610 intron 3 Prime UTR Variant		CG		
12	AQP2	rs10875989	rs10875989 49,957,292 intron 3 Prime UTR Variant		CT		
12	AQP5	rs296763	49,969,231	intron	Non-Coding Transcript Variant	CG	
12	AQP5	rs3759129	49,960,654	intron	2KB Upstream Variant	CA	
12	AQP5	rs3736309	49,964,271	intron Intron Variant		GA	
12	AQP6	rs1996315	49,970,924	intron	2KB Upstream Variant	GA	

https://doi.org/10.1371/journal.pone.0229245.t001

Sample and phenotypes selection

First, we selected an arthritis affected group, which consisted of individuals who self-reported osteoarthritis, rheumatoid arthritis or arthritis in our database. We selected 137 cases (47 males and 90 females). The mean age was 59.5 years, ranging from 19.4 to 90.9 years. For comparison we selected 551 unaffected individuals (191 males and 360 females), with a ratio of affected to unaffected of approximately 1:4 [36], matched to arthritis cases by age, ethnicity, and sex. Within the arthritis total sample, we selected the periodontitis, TMD groups, and unaffected groups using the same selection and matching criteria, except that periodontitis was not self-reported, but diagnosed by a professional dentist. For the periodontitis phenotype, individuals were considered affected if presenting with either at least three teeth exhibiting sites of clinical attachment loss and probing depth equal or greater to 5 mm in two different quadrants, or if presenting with alveolar bone resorption (evaluated radiographically) and severe dental mobility. A total of 141 affected individuals (93 females and 48 males) and 228 unaffected individuals (154 females and 74 males) constituted the periodontitis phenotype groups. For the TMD affected group, we selected 184 individuals (133 females and 51 males) that reported at least one symptom in the temporomandibular joint (clicking, sounds or pain) and 476 individuals (295 females and 181 males) constituted the comparison group. One of the authors (M.B.) carried out the extraction of clinical data after being calibrated by an experienced specialist (A.R.V.). The clinical data included the complete oral conditions and treatments present in the database for each of the patients seeking care between September 2006 and May 2018. The intra-extractor agreement was assessed by a second extraction of clinical data in 10% of the sample after 2 weeks, with a kappa of 1.0. Calculating inter- or intra-examiner agreement was not possible, because each phenotype studied was recovered from a registry of clinical information derived from the dental clinics of the University of Pittsburgh.

We estimated power for the association study by using the Genetic Power Calculator tool [37], setting the frequency of the high-risk allele in the population equal to 0.2, the prevalence of the conditions as 0.1, the effect size in the heterozygote form as 2.0, D' as 1.0 (amount of linkage disequilibrium), the respective number of cases of each disease and the case-control ratio. The samples selected for arthritis, periodontitis, and TMD would have 76%, 62%, and 81% power, respectively, to detect association at an alpha of 0.05. We did not perform power calculations on the cohorts of individuals with more than one condition. One can assume that these subgroups of combined individuals with more than one disease would provide less statistical power because they are smaller, although we hope that this approach increases homogeneity.

Linkage disequilibrium of the region and multiple testing concerns

We generated a linkage disequilibrium plot of the 12q13.12 locus in this study population (Fig 1). We can conclude that the SNP rs467323 is in linkage disequilibrium with rs3736309, as well as the SNPs rs10875989, rs2878771, rs296763 and rs1996315, which mean these SNPs provide redundant information. We employed a Bonferroni correction for multiple comparisons, and set alpha at 0.01 (0.05/5 markers) to account for disequilibrium, excluding redundant information given by 4 SNPs.

Results

P-values below 0.05 of periodontitis and temporomandibular joint symptoms were found and are represented in Table 2. The affected and unaffected individuals on the genotypic model represent the number of different genotypes (e.g. CC, CT and TT) in the affected group and in the unaffected group where the first number is the homozygous for the mutated alleles, followed by the heterozygous, followed by the homozygous for the normal alleles. We used our genotyping data to recalculate power since we could derive relative risks for AA, Aa and aa directly from our results. These post-hoc calculations showed our power for the comparisons of the sample selected for arthritis, periodontitis, TMD, and TMD combined with periodontitis were 12%, 58%, 83.5%, and 68.6%, respectively.

Single disease associations

We found an association between one marker (rs1996315) in AQP6 and periodontitis (p = 0.05). Temporomandibular joint dysfunction was associated with one marker (rs467323) in AQP2 for both genotypic (p = 0.03) and recessive (p = 0.02) models (Table 2). No association was found for arthritis with the markers tested.

Dual disease associations

When individuals who reported both TMD and arthritis were analyzed, there were no statistically significant differences. However, when TMD and periodontitis were combined, a marker in AQP2 (rs3741559) showed and association (p = 0.02) (Table 2).

Triple disease associations

When the three disease phenotypes were combined (temporomandibular joint dysfunction, periodontitis, and arthritis), no associations were found.

Haplotype analysis

We performed association tests of two or more adjacent SNPs within the 12q13.12 locus. The haplotype analysis revealed an additional association not previously identified by the single marker analysis. For rs10875989 and rs467323 in AQP2, the first SNP did not show signals alone but when combined with the second SNP, the haplotype CT showed a significant association with the TMD phenotype (p = 0.007), being approximately three times more frequently found in the control group (Table 3).

Discussion

AQPs have been reported to play a role in progression and resolution of inflammatory status [38]. Thus, here we analyzed inflammatory conditions that have different causes and chronic inflammation in common. We show that variation in *AQP2* are potentially associated with

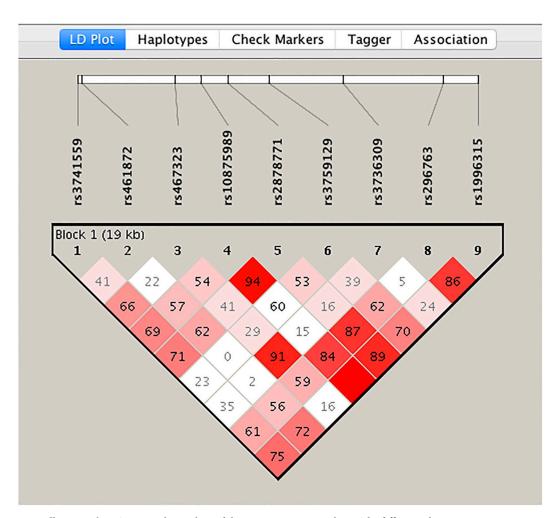


Fig 1. Illustrates the *AQP 2***, 5 and 6 markers of the 12q13.12 aquaporin locus.** The different color intensities represent a different D prime value, the stronger the red, the closer a D' value is to 1.

https://doi.org/10.1371/journal.pone.0229245.g001

temporomandibular joint disorder and periodontitis. Our previous studies have shown that a more unique population is generated when disease phenotypes are analyzed in combination, increasing the chance of identifying biologically relevant associations [39]. Our intention in combining those diseases was to verify if a pleiotropic effect is present that leads those individuals to develop different conditions and symptoms at the same time. An association was found when patients who have both TMD and periodontitis were analyzed however, when all three diseases were analyzed in combination in this study, we did not find significant results. These

Table 2. Summarizes the nominal results of both phenotypic isolated and combined analyses.

Phenotype	A1 / A2	Gene	SNP	Model	Affected	Unaffected	X ²	p value	Relative risk of carrying one copy of the risk allele
Periodontitis	G/A	AQP6	rs1996315	Dominant	106/23	150/55	3.583	0.058	1.2
Temporomandibular joint disorder	C/T	AQP2	rs467323	Genotypic	18/84/56	80/178/149	6.706	0.03	1.46
(TMD)		AQP2	rs467323	Recessive	18/140	80/327	5.421	0.02	
Periodontitis and TMD	A/G	AQP2	rs3741559	Allelic	25/177	45/171	5.355	0.02	2.15

https://doi.org/10.1371/journal.pone.0229245.t002

Table 3. Summarizes the haplotype significant association identified between rs467323 and rs10875989 block markers using the haploview software.

Haplotype Associations rs467323/rs10875989 block markers	Frequency	Case, Control Ratios	Chi Square	p value
TT	0.639	172.1: 91.9, 424.9: 245.1	0.257	0.6122
CC	0.283	77.5: 186.5, 187.1: 482.9	0.194	0.6596
CT	0.051	5.4: 258.6, 42.6: 627.4	7.212	0.0072
TC	0.026	8.9: 255.1, 15.4: 654.6	0.9	0.3428

https://doi.org/10.1371/journal.pone.0229245.t003

negative results are likely due to the small sample size of patients affected by the three conditions concomitantly (n = 9).

Even though all markers complied with Hardy Weinberg equilibrium (HWE) cutoff of p>0.001 set as default in the PLINK software [34], the initial sample selected to define a cohort of individuals with arthritis had the genotype distribution of markers rs10875989 and rs2878771 different than expected under HWE (p = 0.009 and 0.004, respectively). It has been suggested [40] that this might happen due to population stratification, when both Whites and African-Americans are included in the study. Concerned with this possible effect we ran a separate analysis excluding African-Americans and we did not see a significant difference in HWE just for Whites [rs10875989, 25CC, 225CT and 207TT for Whites (p = 0.001) versus 44CC, 278CT and 257TT for the total sample (p = 0.009)] and [rs2878771, 6CC, 166CG and 295GG (p = 0.002) versus 12CC, 211CG and 378GG (p = 0.004)]. This indicates that any possible population substructure may be due to combinations of different groups of Europeans that cannot be detected with the information available to us [41]. The aquaporin locus in particular may make these individuals more susceptible to the inflammatory conditions we are studying, in comparison to African-Americans. The other possibility is that the selection of individuals with arthritis and the subsequent matched group (by sex, age, and ethnicity) may have caused this effect because these groups have an excess of heterozygous and homozygous rare individuals. This may be the reflex of the actual effect of the aquaporin locus on the underlying conditions studied since all sample is enriched by individuals affected by TMD and periodontitis. This effect was seen before when families with a child born with cleft lip and palate were analyzed based on paternal origin and showed Hardy-Weinberg distortion [42]. This effect would not necessarily translate to significant results detected by association methods because the overall population studied is enriched by the rare allele.

When sorting the subjects who were affected by more than one disease, it was interesting to observe that most of the patients who have periodontitis are also affected by TMD (106 individuals). This phenotype-to-phenotype finding correlates with the genotypic association results reported here. A recent study analyzed disease patterns of patients with temporomandibular disorders and concluded that chronic periodontitis resulting in unilateral mastication induces temporomandibular joint changes and pain is those patients [43]. Our study showed that a genetic component might also be present, increasing the chances of having temporomandibular joint disorders associated with periodontitis.

The rs1996315 marker is located between *AQP5* and 6 and is a substitution of G for A, being reported as associated with dental caries [44]. The suggested underlying mechanism of this association with caries might be due to reduced salivary flow, and likewise could affect dental plaque retention and increase the risk for periodontitis. The *AQP2* rs467323 marker is a substitution of C for T and the clinical significance is reported as benign, which means that the variant has no deleterious effect on gene expression or function [45]. Thus, our results demonstrate for the first time a relationship and possible protective effect of *AQP2* and TMD—the CC genotype is less seen in the affected group compared to the unaffected (11% and 19% respectively for rs467323 marker). The rs3741559 *AQP2* marker did not show associations

with any of the diseases isolated but when individuals affected by both TMD and periodontitis were tested, the A allele was less present in the cases, characterizing a protective effect.

A limitation we faced in our study was that arthritis was self-reported by the patients; we did not always have a detailed description of the type of arthritis affliction. In our data, the results did not differ when rheumatoid arthritis and osteoarthritis patients are analyzed in separate and we decided to study both types together. Since we did not have an independent sample to replicate these results, independent investigations testing the aquaporin locus studied here in the inflammatory conditions we studied are warranted.

This study did not investigate all AQPs expressed by leukocytes and in the periodontium and cartilaginous tissues (e.g. AQP 4, 9). Further work is needed to delineate if the effects of AQP polymorphisms in periodontitis and degenerative joint diseases are tissue specific (e.g. changes in epithelial barrier function) or immune response specific (e.g. phagocytosis), and to define their contribution to the type and severity of immune response to tissue injury and infection.

Periodontitis has been associated with a number of conditions that include cardiovascular risks and stroke, arthritis, lupus erythematosus, infections of the upper respiratory tract, diabetes, low birth weight and premature births (23–27). Here we provided statistical evidence that the aquaporin locus may be implicated in individuals affected with TMD and a trend to associations in individuals that show concomitant TMD and periodontitis, suggesting that this may be the reason why conditions that are inflammatory in nature are found to be associated with periodontitis in human populations.

Supporting information

S1 Table. Genotyping data used for analysis of arthritis group. (XLSX)

S2 Table. Genotyping data used for analysis of periodontitis group. (XLSX)

S3 Table. Genotyping data used for analysis of temporomandibular joint disorder group. (XLSX)

S4 Table. Genotyping data used for analysis of arthritis and periodontitis combined group.

(XLSX)

S5 Table. Genotyping data used for analysis of arthritis and TMD combined group. (XLSX)

S6 Table. Genotyping data used for analysis of periodontitis and TMD combined group. (XLSX)

S7 Table. Genotyping data used for analysis of arthritis, periodontitis and TMD combined group. (XLSX)

Acknowledgments

The University of Pittsburgh Dental Registry and DNA Repository provided data and DNA samples for this study.

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References

- Ma TH, Gao HW, Fang XD, Yang H. Expression and function of aquaporins in peripheral nervous system. Acta Pharmacol Sin. 2011; 32(6):711–5. https://doi.org/10.1038/aps.2011.63 PMID: 21602841
- 2. Takata K, Matsuzaki T, Tajika Y. Aquaporins: water channel proteins of the cell membrane. Prog Histochem Cytochem. 2004; 39(1):1–83. https://doi.org/10.1016/j.proghi.2004.03.001 PMID: 15242101
- Papadopoulos MC, Saadoun S, Verkman AS. Aquaporins and cell migration. Pflugers Arch. 2008; 456 (4):693–700. https://doi.org/10.1007/s00424-007-0357-5 PMID: 17968585
- Verkman AS. Mammalian aquaporins: diverse physiological roles and potential clinical significance. Expert Rev Mol Med. 2008; 10:e13. https://doi.org/10.1017/S1462399408000690 PMID: 18482462
- Ishibashi K, Kuwahara M, Gu Y, Tanaka Y, Marumo F, Sasaki S. Cloning and functional expression of a new aquaporin (AQP9) abundantly expressed in the peripheral leukocytes permeable to water and urea, but not to glycerol. Biochem Biophys Res Commun. 1998; 244(1):268–74. https://doi.org/10. 1006/bbrc.1998.8252 PMID: 9514918
- Moon C, Rousseau R, Soria JC, Hoque MO, Lee J, Jang SJ, et al. Aquaporin expression in human lymphocytes and dendritic cells. Am J Hematol. 2004; 75(3):128–33. https://doi.org/10.1002/ajh.10476
 PMID: 14978691
- Zhu N, Feng X, He C, Gao H, Yang L, Ma Q, et al. Defective macrophage function in aquaporin-3 deficiency. FASEB J. 2011; 25(12):4233–9. https://doi.org/10.1096/fj.11-182808
- Compan V, Baroja-Mazo A, Lopez-Castejon G, Gomez AI, Martinez CM, Angosto D, et al. Cell volume regulation modulates NLRP3 inflammasome activation. Immunity. 2012; 37(3):487–500. https://doi.org/ 10.1016/j.immuni.2012.06.013 PMID: 22981536
- Loitto VM, Forslund T, Sundqvist T, Magnusson KE, Gustafsson M. Neutrophil leukocyte motility requires directed water influx. J Leukoc Biol. 2002; 71(2):212–22. PMID: 11818441
- Tyteca D, Nishino T, Debaix H, Van Der Smissen P, N'Kuli F, Hoffmann D, et al. Regulation of macrophage motility by the water channel aquaporin-1: crucial role of M0/M2 phenotype switch. PLoS One. 2015; 10(2):e0117398. https://doi.org/10.1371/journal.pone.0117398 PMID: 25719758
- Hara-Chikuma M, Chikuma S, Sugiyama Y, Kabashima K, Verkman AS, Inoue S, et al. Chemokine-dependent T cell migration requires aquaporin-3-mediated hydrogen peroxide uptake. J Exp Med. 2012; 209(10):1743–52. https://doi.org/10.1084/jem.20112398 PMID: 22927550
- Zhao X, Liu Z, Shu D, Xiong Y, He M, Xu S, et al. Association of Periodontitis with Rheumatoid Arthritis and the Effect of Non-Surgical Periodontal Treatment on Disease Activity in Patients with Rheumatoid Arthritis. Med Sci Monit. 2018; 24:5802–10. https://doi.org/10.12659/MSM.909117 PMID: 30124222
- Benjamin RM. Oral health: the silent epidemic. Public Health Rep. 2010; 125(2):158–9. https://doi.org/10.1177/003335491012500202 PMID: 20297740

- Buffoli B, Dalessandri M, Favero G, Mensi M, Dalessandri D, Di Rosario F, et al. AQP1 expression in human gingiva and its correlation with periodontal and peri-implant tissue alterations. Acta Histochem. 2014; 116(5):898–904. https://doi.org/10.1016/j.acthis.2014.02.010 PMID: 24698289
- Anjomshoaa I, Briseno-Ruiz J, Deeley K, Poletta FA, Mereb JC, Leite AL, et al. Aquaporin 5 Interacts with Fluoride and Possibly Protects against Caries. PLoS One. 2015; 10(12):e0143068. https://doi.org/ 10.1371/journal.pone.0143068 PMID: 26630491
- Pedersen AML, Sorensen CE, Proctor GB, Carpenter GH, Ekstrom J. Salivary secretion in health and disease. J Oral Rehabil. 2018; 45(9):730–46. https://doi.org/10.1111/joor.12664 PMID: 29878444
- Trujillo E, Gonzalez T, Marin R, Martin-Vasallo P, Marples D, Mobasheri A. Human articular chondrocytes, synoviocytes and synovial microvessels express aquaporin water channels; upregulation of AQP1 in rheumatoid arthritis. Histol Histopathol. 2004; 19(2):435–44. https://doi.org/10.14670/HH-19.435 PMID: 15024704
- Mobasheri A, Trujillo E, Bell S, Carter SD, Clegg PD, Martin-Vasallo P, et al. Aquaporin water channels AQP1 and AQP3, are expressed in equine articular chondrocytes. Vet J. 2004; 168(2):143–50. https://doi.org/10.1016/j.tvjl.2003.08.001 PMID: 15301762
- Meng JH, Ma XC, Li ZM, Wu DC. Aquaporin-1 and aquaporin-3 expressions in the temporo-mandibular joint condylar cartilage after an experimentally induced osteoarthritis. Chin Med J (Engl). 2007; 120 (24):2191–4.
- Nagahara M, Waguri-Nagaya Y, Yamagami T, Aoyama M, Tada T, Inoue K, et al. TNF-alpha-induced aquaporin 9 in synoviocytes from patients with OA and RA. Rheumatology (Oxford). 2010; 49(5):898– 906.
- Gao H, Ren G, Xu Y, Jin C, Jiang Y, Lin L, et al. [Correlation between expression of aquaporins 1 and chondrocyte apoptosis in articular chondrocyte of osteoarthritis]. Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi. 2011; 25(3):279–84. PMID: 21500577
- Cai L, Lei C, Li R, Chen WN, Hu CM, Chen XY, et al. Overexpression of aquaporin 4 in articular chondrocytes exacerbates the severity of adjuvant-induced arthritis in rats: an in vivo and in vitro study. J Inflamm (Lond). 2017; 14:6.
- 23. Chatzopoulos GS, Cisneros A, Sanchez M, Wolff LF. Systemic medical conditions and periodontal status in older individuals. Spec Care Dentist. 2018.
- Stanko P, Izakovicova Holla L. Bidirectional association between diabetes mellitus and inflammatory periodontal disease. A review. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub. 2014; 158 (1):35–8. https://doi.org/10.5507/bp.2014.005 PMID: 24509898
- Lockhart PB, Bolger AF, Papapanou PN, Osinbowale O, Trevisan M, Levison ME, et al. Periodontal disease and atherosclerotic vascular disease: does the evidence support an independent association?: a scientific statement from the American Heart Association. Circulation. 2012; 125(20):2520–44. https://doi.org/10.1161/CIR.0b013e31825719f3 PMID: 22514251
- 26. Ayed MS, Shafiq SS, Diab HM, Alahmari AD, Divakar DD. Assessing periapical dental radiographs as a screening parameter for early indications of osteoporosis in postmenopausal periodontal patients and root surface evaluation using spectrochemical analysis. Saudi Med J. 2018; 39(7):719–24. https://doi.org/10.15537/smj.2018.7.22453 PMID: 29968896
- 27. Golub LM, Payne JB, Reinhardt RA, Nieman G. Can systemic diseases co-induce (not just exacerbate) periodontitis? A hypothetical "two-hit" model. J Dent Res. 2006; 85(2):102–5. https://doi.org/10.1177/154405910608500201 PMID: 16434727
- Wei M, Yu H, Zhang Y, Zeng J, Cai C, Shi R. Decreased expression of aquaporin 1 correlates with clinicopathological features of patients with cervical cancer. Onco Targets Ther. 2019; 12:2843–51. https:// doi.org/10.2147/OTT.S194650 PMID: 31118662
- Vieira AR, Hilands KM, Braun TW. Saving more teeth-a case for personalized care. J Pers Med. 2015; 5(1):30–5. https://doi.org/10.3390/jpm5010030 PMID: 25758360
- Anjomshoaa I, Cooper ME, Vieira AR. Caries is Associated with Asthma and Epilepsy. Eur J Dent. 2009; 3(4):297–303. PMID: 19826602
- 31. Vieira AR, Silva MB, Souza KKA, Filho AVA, Rosenblatt A, Modesto A. A Pragmatic Study Shows Failure of Dental Composite Fillings Is Genetically Determined: A Contribution to the Discussion on Dental Amalgams. Front Med (Lausanne). 2017; 4:186.
- Aidar M, Line SR. A simple and cost-effective protocol for DNA isolation from buccal epithelial cells. Braz Dent J. 2007; 18(2):148–52. https://doi.org/10.1590/s0103-64402007000200012 PMID: 17982556
- Ranade K, Chang MS, Ting CT, Pei D, Hsiao CF, Olivier M, et al. High-throughput genotyping with single nucleotide polymorphisms. Genome Res. 2001; 11(7):1262–8. PMID: 11435409

- Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for wholegenome association and population-based linkage analyses. Am J Hum Genet. 2007; 81(3):559–75. https://doi.org/10.1086/519795 PMID: 17701901
- **35.** Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics. 2005; 21(2):263–5. https://doi.org/10.1093/bioinformatics/bth457 PMID: 15297300
- Hennessy S, Bilker WB, Berlin JA, Strom BL. Factors influencing the optimal control-to-case ratio in matched case-control studies. Am J Epidemiol. 1999; 149(2):195–7. https://doi.org/10.1093/ oxfordjournals.aje.a009786 PMID: 9921965
- Purcell S, Cherny SS, Sham PC. Genetic Power Calculator: design of linkage and association genetic mapping studies of complex traits. Bioinformatics. 2003; 19(1):149–50. https://doi.org/10.1093/bioinformatics/19.1.149 PMID: 12499305
- Meli R, Pirozzi C, Pelagalli A. New Perspectives on the Potential Role of Aquaporins (AQPs) in the Physiology of Inflammation. Front Physiol. 2018; 9:101. https://doi.org/10.3389/fphys.2018.00101 PMID: 29503618
- Bezamat M, Deeley K, Khaliq S, Letra A, Scariot R, Silva RM, et al. Are mTOR and Endoplasmic Reticulum Stress Pathway Genes Associated with Oral and Bone Diseases? Caries Res. 2018; 53(3):235–41. https://doi.org/10.1159/000492675 PMID: 30205378
- 40. Heiman GA, Hodge SE, Gorroochurn P, Zhang J, Greenberg DA. Effect of population stratification on case-control association studies. I. Elevation in false positive rates and comparison to confounding risk ratios (a simulation study). Hum Hered. 2004; 58(1):30–9. https://doi.org/10.1159/000081454 PMID: 15604562
- **41.** Hinrichs AL, Larkin EK, Suarez BK. Population stratification and patterns of linkage disequilibrium. Genet Epidemiol. 2009; 33 Suppl 1:S88–92.
- Prescott NJ, Winter RM, Malcolm S. Maternal MTHFR genotype contributes to the risk of non-syndromic cleft lip and palate. J Med Genet. 2002; 39(5):368–9. https://doi.org/10.1136/jmg.39.5.368 PMID: 12011160
- Jeon HM, Ahn YW, Jeong SH, Ok SM, Choi J, Lee JY, et al. Pattern analysis of patients with temporomandibular disorders resulting from unilateral mastication due to chronic periodontitis. J Periodontal Implant Sci. 2017; 47(4):211–8. https://doi.org/10.5051/jpis.2017.47.4.211 PMID: 28861285
- 44. Wang X, Willing MC, Marazita ML, Wendell S, Warren JJ, Broffitt B, et al. Genetic and environmental factors associated with dental caries in children: the Iowa Fluoride Study. Caries Res. 2012; 46(3):177–84. https://doi.org/10.1159/000337282 PMID: 22508493
- **45.** Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015; 17(5):405–24. https://doi.org/10.1038/gim.2015.30 PMID: 25741868