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Insight

The symbiosome – a transient organelle in evolution

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This article comments on:

Casaes PA, Ferreira dos Santos JM, Silva VC, Rhem MFK, Teixeira Cota MM, de Faria SM, Rando JG, James EK, Gross E. 2024. The radiation of nodulated *Chamaecrista* species from the rainforest into more diverse habitats has been accompanied by a reduction in growth form and a shift from fixation threads to symbiosomes. Journal of Experimental Botany **75**, 3643–3662.

The efficiency of nitrogen-fixing root nodule symbiosis is greatly dependent on the manner in which the symbiont is intracellularly accommodated. Rhizobia can reside either in cell wall-bound fixation threads (FTs) or in membrane-bound, organelle-like structures termed symbiosomes (SYMs). Casaes et al. (2024) investigated the evolution of Chamaecrista (a legume genus belonging to the Caesalpinioideae, a sister subfamily to the Papilionoideae), focusing on a possible relationship between the plant's growth habitat and the rhizobial housing mechanism. They identified tree species with FTs, (sub) shrubs with SYMs, and, notably, several shrub species displaying an intermediate FT-SYM phenotype. The presence of multiple rhizobia housing mechanisms within a single evolutionary genus, in combination with the presence of a possibly unique intermediate form of rhizobia housing, opens up the opportunity to unravel the genetic adaptations leading towards SYM release, and could potentially shed more light on organelle evolution.

Symbiosomes: a crucial evolutionary adaptation

Both FT- and SYM-type structures enable nutrient exchange between the host plant and the N_2 -fixing rhizobium in its symbiotic form, the bacteroid. Three key features distinguish

the FTs from SYMs (Fig. 1A-C). Firstly, the FT is derived from invaginations of the host cell wall of the infection thread (IT) and remains cell wall bound at all times (Brewin, 1998; Fonseca et al., 2012). Conversely, SYM release involves the breakdown of the plant cell wall at the tip of the IT, the release of the rhizobia into the host cytoplasm, but with the retention of the symbiont only in a thin host-derived (peribacteroid) membrane (Gavrin et al., 2016). A second key difference is that SYMs typically contain only one to at most a few rhizobia. Combined with the removal of the host-derived cell wall, this effectively reduces the amount of space occupied by each individual symbiont, and, as a consequence, allows the host to accommodate a far greater number of symbionts per cell (de Faria et al., 2022; Casaes et al., 2024). Akin to chloroplasts, a greater number of smaller organelles is believed to be more efficient than a few larger FTs (Schumpp et al., 2009; Xiong et al., 2017). A third, and probably crucial key difference, is that the SYM fully encloses the symbiont by only the peribacteroid membrane, thereby maximizing the surface area available for nutrient exchange while probably at the same time minimizing any impediment to diffusion of metabolites and nutrients. This more efficient plant-microbe interaction probably confers a competitive advantage to the host plant. It is therefore not surprising that the SYM-type symbiosis is the predominant symbiont housing strategy among all nodulating Fabaceae species (de Faria et al., 2022).

The currently prevailing hypothesis on the origin of nodulation is a single gain followed by massive parallel losses (Griesmann *et al.*, 2018; van Velzen *et al.*, 2018). Stable retention of the nodulation trait seemingly correlates with the ability to develop SYM-type nodules (de Faria *et al.*, 2022). Within the N₂-fixing clade, the vast majority of nodulation species are found within the *Fabaceae* family (Doyle, 2011; Griesmann *et al.*, 2018). Yet even here the nodulation trait is unequally distributed. Of the over ~20 000 *Fabaceae*, most nodulating species are members of either the *Papilionoideae* subfamily or

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the mimosoid clade in the *Caesalpinioideae* subfamily (de Faria *et al.*, 2022). With the exception of a few basal *Papilionoideae* clades, all of the aforementioned nodulation-competent species exclusively develop SYM-type nodules. The phylogenetic distribution of these species suggests the FT to be an ancestral trait. Consequently, SYMs must have been acquired independently through convergent evolution (Sprent *et al.*, 2013; Ardley and Sprent, 2021; de Faria *et al.*, 2022) (Fig. 2). It thus appears that the development of SYMs is pivotal, as failure to do so may ultimately lead to the loss of the ability to nodulate.

Chamaecrista as a novel experimental system to investigate symbiosome evolution

Despite being a crucial adaption, the genetic changes underlying a transition from FT- to SYM-type rhizobia housing have remained largely elusive. Previous studies on SYM evolution predominantly focused on SYM-developing species (Gavrin *et al.*, 2016; Libourel *et al.*, 2023), presumably limited by the large evolutionary gap between FT-type and



Fig. 1. Schematic overview of the different rhizobial housing mechanisms. (A) A fixation thread (FT) derived from invagination of the plant cell wall (black) and surrounded by a plant-derived membrane (red). (B) An intermediate FT–symbiosome (FT–SYM) housing type. Note the thinning of the cell wall and protrusion of the symbionts. (C) SYMs surrounded by a plant-derived membrane and lacking a plant cell wall.



Fig. 2. Simplified phylogeny of the nitrogen-fixing clade with emphasis on the rhizobial housing mechanism and the legume family. Symbiosomes (SYMs; red); fixation threads (FTs; blue); and fixation thread–symbiosome intermediate (FT–SYM; green). IRLC=inverted repeat lacking clade. Time scales are approximate.

SYM-type Papilionoideae. The comparison between basal FT-type Papilionoideae clades and SYM-forming Papilionoideae is complicated by >50 million years of evolutionary separation, thereby introducing significant noise within the analysis (Sprent et al., 2017). However, the recent study by Casaes et al. (2024) positions the non-mimosoid Caesalpinioideae Chamaecrista genus as a particularly attractive system to study SYM evolution. Chamaecrista stands out as having independently acquired SYMs and being the only known genus to date to contain both SYM- and FT-type nodules. Within the Chamaecrista genus, tree species of the basal Apoucouita section develop FTs, whereas shrubby members of the Absus and Chamaecrista sections develop SYMs (Naisbitt et al., 1992; Casaes et al., 2024). Notably, Casaes et al. (2024) pinpoint several species seemingly in transition from FT- to SYM-type nodules, further suggesting an ongoing evolutionary development towards SYM acquisition in the Chamaecrista genus (Fig. 1B). The most recent FT-SYM to SYM transition event in this genus occurred relatively recently, ~17 million years ago, while a comparison between *Chamaecrista* species employing FT-type and SYM-type strategies is separated by ~35 million years of evolution (Fig. 2). Combined, this means that a comparative analysis within the *Chamaecrista* genus holds promise for identifying the crucial adaptations that could be causal in facilitating a transition towards SYM housing mechanisms of N₂-fixing rhizobia.

The aim of such comparative analyses would be to derive a blueprint of the genetic constraints needed to enable SYM formation. The ultimate proof of concept, and validation, of such a blueprint is to engineer SYMs on an FT-type species. Moving beyond *in silico* analyses towards practical application necessitates developing a diversity of *Chamaecrista* species as suitable models for *in planta* research. The SYM-type shrubby species *Chamaecrista fasciculata* and *C. mimosoides* have already been used in laboratory settings, their genomes have been sequenced, and they are amenable to hairy root transformations via *Rhizobium rhizogenes* (formerly *Agrobacterium rhizogenes*) (Griesmann *et al.*, 2018; Wardhani, 2020). However, the

Box 1. Terminally differentiated bacteroids - a temporary organelle

Following symbiosome release, certain species impose terminal differentiation upon their symbiont. While primarily observed in members of the inverted repeat lacking clade (IRLC), a form of terminal differentiation is also observed in stem nodules of the *Aeschynomene* genus (Mergaert *et al.*, 2006; Czernic *et al.*, 2015). Thus, like symbiosomes, this trait too appears to be an example of convergent evolution.

Terminal differentiation is characterized by the enlargement of the symbiont, endoreplication of its genomic content, increased membrane permeability and the irreversible loss of autonomy (Fig. 3).

In *Medicago truncatula* (an IRLC member), the induction of terminal differentiation is dependent on host-produced nodule-specific cysteine-rich (NCR) peptides (Van de Velde *et al.*, 2010). The NCR family of peptides comprises >600 members, few of which have been characterized. The mature peptides are transported towards the symbiosome membrane where they induce terminal differentiation (Wang *et al.*, 2010; Montiel *et al.*, 2017; Yang *et al.*, 2023). Although the genetic regulation of terminal differentiation in the *Aeschynomene* genus is unknown, it too appears to utilize NCR peptides (Czernic *et al.*, 2015). Terminal differentiation is believed to further enhance the efficiency of the nitrogen-fixing symbiosis (Oono and Denison, 2010), a claim corroborated by the apparent convergent evolution of the trait.





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biggest challenge probably lies with the FT-type *Chamaecrista*. FT-type *Chamaecrista* species are all tropical trees, which generally suffer from relatively long, sexually incompetent, juvenile stages. Consequently, utilizing tree species in laboratory conditions is a non-trivial task. Nevertheless, the successful utilization of the nodulating tree species *Parasponia andersonii* indicates that establishing a model tree species is feasible (Wardhani *et al.*, 2019). Comparative analyses involving *P andersonii* have already provided valuable insights into the evolution of nodulation (vanVelzen *et al.*, 2018; Libourel *et al.*, 2023; Zhang *et al.*, 2023). The *Chamaecrista* genus holds similar potential to provide crucial insights into the evolution of SYMs. However, to truly establish *Chamaecrista* as a model system for SYM evolution, significant efforts must be directed towards establishing FT, SYM, and FT–SYM species as additional experimental models.

The evolution of a transient organelle

SYMs not only boost the efficiency of the symbioses, but also enable the host plant to exert greater levels of control over its symbiont. Consider the inverted repeat lacking clade (IRLC) within the Papilionoideae subfamily of legumes for instance. Here, following SYM release, the symbiont is terminally differentiated during the nodulation process, which further optimizes-exploits-the symbiotic interaction. During terminal differentiation, host-produced peptides trigger the symbiont to lose its capacity to function as a free-living organism, essentially becoming a transient organelle (Box 1; Fig. 3) (Mergaert et al., 2006; Van de Velde et al., 2010). While members of the Mimosoid clade in the Caesalpinioideae do not appear to impose terminal differentiation upon their symbionts (Marchetti et al., 2011; Libourel et al., 2023), it is currently unknown if such terminal differentiation could occur within the SYM-type nodules of Chamaecrista species.

During terminal differentiation, the symbiont seems to have relinquished all control to the host. However, one outstanding question that remains is whether the host or the symbiont is in control of SYM formation. The current hypothesis is that the host plant controls the party (Ferguson *et al.*, 2019).

The cross-nodulation experiments performed by Casaes and colleagues (2024) identified strains capable of nodulating on a SYM-type shrub species, though their primary host is an FT-type tree. An investigation on the structure of these nodules would be warranted. Such an analysis of SYM-type nodules with a rhizobial strain generally associated with FT-type nodules, and vice versa, would be able to test the current dogma on host control of bacterial release.

Terminally differentiated or not, SYMs bear a significant resemblance to an organelle; they are enclosed in host-derived membranes, an import–export mechanism is established, and in some cases the symbiont is stripped of its autonomy. This study by Casaes and colleagues (2024) now positions the *Chamaecrista* genus as an interesting system for comparative evolutionary analyses to study SYM evolution. This will not only provide valuable insights for nodule engineering efforts but will also shed light onto the acquisition of a new organelle.

Conflict of interest

The authors declare no conflicts of interest.

Keywords: Chamaecrista, evolution, fixation thread, symbiosome.

References

Ardley J, Sprent J. 2021. Evolution and biogeography of actinorhizal plants and legumes: a comparison. Journal of Ecology **109**, 1098–1121.

Brewin NJ. 1998. Tissue and cell invasion by rhizobium: the structure and development of infection threads and symbiosomes. In: Spaink HP, Kondorosi A, Hooykaas PJJ, eds. The Rhizobiaceae: molecular biology of model plant-associated bacteria. Dordrecht: Springer Netherlands, 417–429.

Casaes PA, Ferreira dos Santos JM, Silva VC, Rhem MFK, Cota MMT, de Faria SM, Rando JG, James EK, Gross E. 2024. The radiation of nodulated *Chamaecrista* species from the rainforest into more diverse habitats has been accompanied by a reduction in growth form and a shift from fixation threads to symbiosomes. Journal of Experimental Botany **75**, 3643–3662.

Czernic P, Gully D, Cartieaux F, et al. 2015. Convergent evolution of endosymbiont differentiation in dalbergioid and inverted repeat-lacking clade legumes mediated by nodule-specific cysteine-rich peptides. Plant Physiology **169**, 1254–1265.

de Faria SM, Ringelberg JJ, Gross E, et al. 2022. The innovation of the symbiosome has enhanced the evolutionary stability of nitrogen fixation in legumes. New Phytologist **235**, 2365–2377.

Doyle JJ. 2011. Phylogenetic perspectives on the origins of nodulation. Molecular Plant-Microbe Interactions **24**, 1289–1295.

Ferguson BJ, Mens C, Hastwell AH, Zhang M, Su H, Jones CH, Chu X, Gresshoff PM. 2019. Legume nodulation: the host controls the party. Plant, Cell & Environment 42, 41–51.

Fonseca MB, Peix A, de Faria SM, et al. 2012. Nodulation in *Dimorphandra wilsonii* Rizz. (Caesalpinioideae), a threatened species native to the Brazilian Cerrado. PLoS One 7, e49520.

Gavrin A, Chiasson D, Ovchinnikova E, Kaiser BN, Bisseling T, Fedorova EE. 2016. VAMP721a and VAMP721d are important for pectin dynamics and release of bacteria in soybean nodules. New Phytologist **210**, 1011–1021.

Griesmann M, Chang Y, Liu X, et al. 2018. Phylogenomics reveals multiple losses of nitrogen-fixing root nodule symbiosis. Science **361**, eaat1743.

Libourel C, Keller J, Brichet L, *et al*. 2023. Comparative phylotranscriptomics reveals ancestral and derived root nodule symbiosis programmes. Nature Plants **9**, 1067–1080.

Marchetti M, Catrice O, Batut J, Masson-Boivin C. 2011. *Cupriavidus taiwanensis* bacteroids in *Mimosa pudica* indeterminate nodules are not terminally differentiated. Applied and Environmental Microbiology **77**, 2161–2164.

Mergaert P, Uchiumi T, Alunni B, et al. 2006. Eukaryotic control on bacterial cell cycle and differentiation in the *Rhizobium*–legume symbiosis. Proceedings of the National Academy of Sciences, USA **103**, 5230–5235.

Montiel J, Downie JA, Farkas A, Bihari P, Herczeg R, Bálint B, Mergaert P, Kereszt A, Kondorosi E. 2017. Morphotype of bacteroids in different legumes correlates with the number and type of symbiotic NCR peptides. Proceedings of the National Academy of Sciences, USA **114**, 5041–5046. **Naisbitt T, James EK, Sprent JI.** 1992. The evolutionary significance of the legume genus *Chamaecrista*, as determined by nodule structure. New Phytologist **122**, 487–492.

Oono R, Denison RF. 2010. Comparing symbiotic efficiency between swollen versus nonswollen rhizobial bacteroids. Plant Physiology **154**, 1541–1548.

Schumpp O, Crèvecoeur M, Broughton WJ, Deakin WJ. 2009. Delayed maturation of nodules reduces symbiotic effectiveness of the *Lotus japonicus–Rhizobium* sp. NGR234 interaction. Journal of Experimental Botany **60**, 581–590.

Sprent JI, Ardley JK, James EK. 2013. From North to South: a latitudinal look at legume nodulation processes. South African Journal of Botany **89**, 31–41.

Sprent JI, Ardley J, James EK. 2017. Biogeography of nodulated legumes and their nitrogen-fixing symbionts. New Phytologist **215**, 40–56.

Van de Velde W, Zehirov G, Szatmari A, et al. 2010. Plant peptides govern terminal differentiation of bacteria in symbiosis. Science **327**, 1122–1126.

van Velzen R, Holmer R, Bu F, et al. 2018. Comparative genomics of the nonlegume Parasponia reveals insights into evolution of nitrogen-fixing rhizobium symbioses. Proceedings of the National Academy of Sciences, USA **115**, E4700–E4709.

Wang D, Griffitts J, Starker C, Fedorova E, Limpens E, Ivanov S, Bisseling T, Long S. 2010. A nodule-specific protein secretory pathway required for nitrogen-fixing symbiosis. Science **327**, 1126–1129.

Wardhani TAK. 2020. Divergence of rhizobium-induced cytokinin signalling in nodulating species. PhD thesis, Wageningen University.

Wardhani TAK, Roswanjaya YP, Dupin S, Li H, Linders S, Hartog M, Geurts R, van Zeijl A. 2019. Transforming, genome editing and phenotyping the nitrogen-fixing tropical Cannabaceae tree *Parasponia andersonii*. Journal of Visualized Experiments **150**, 1–17. doi:10.3791/59971

Xiong D, Huang J, Peng S, Li Y. 2017. A few enlarged chloroplasts are less efficient in photosynthesis than a large population of small chloroplasts in *Arabidopsis thaliana*. Scientific Reports **7**, 5782.

Yang J, Zhai N, Chen Y, Wang L, Chen R, Pan H. 2023. A signal peptide peptidase is required for ER–symbiosome proximal association and protein secretion. Nature Communications **14**, 4355.

Zhang Y, Fu Y, Xian W, *et al.* 2023. Comparative phylogenomics and phylotranscriptomics provide insights into the genetic complexity of nitrogenfixing root-nodule symbiosis. Plant Communications **5**, 100671.