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ORIGINAL RESEARCH

Oligosaccharide nanomedicine of alginate sodium improves therapeutic results of posterior lumbar interbody fusion with cages for degenerative lumbar disease in osteoporosis patients by downregulating serum miR-155

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Abstract: Degenerative lumbar disease (DLD) is a significant issue for public health. Posterior lumbar intervertebral fusion with cages (PLIFC) has high-level fusion rate and realignment on DLD. However, there are some complications following the surgery. Alginate oligosaccharides (AOS) have antioxidant and anti-inflammatory activities and may be suitable for infection therapy. MiR-155 is a biomarker associated with inflammatory and oxidative stress. AOS may promote PLIFC therapy by regulating miR-155. Pluronic nanoparticles and oligosaccharide nanomedicine of alginate sodium (ONAS) were prepared with ampicillin at size <200 nm. Ninety-six DLD osteoporosis patients received PLIFC and were evenly assigned into ONAS group (OG, oral administration of 100 mg ONAS daily) and control group (PG, 100 mg pluronic nanoparticles). Serum miR-155 level was measured by real-time quantitative PCR. The levels of superoxide dismutase (SOD), glutathione (GSH), aspartate aminotransaminase (AST), alanine aminotransferase (ALT), interleukin-1 β (IL-1 β), and interleukin-1 receptor antagonist (IL-1ra) were measured. Weighted mean difference (WMD), relative risk (RR), complications, surgery infection rate, fusion rate, and Japanese Orthopaedic Association (JOA) scores were used to evaluate therapeutic efficacy. After 1-month therapy, infection rates and side effects were lower in OG than those in PG (RR =0.64, 95% confidence interval [CI] [0.48, 0.84], P=0.001). The fusion rates were higher in OG than in PG (WMD =21.96, 95% CI [-0.24, 37.62], P=0.021). The JOA scores were higher in OG than in PG (RR = 0.52, 95% CI [0.33, 0.84], P = 0.007), and no significant difference was found for the visual analog scale and Oswestry Disability Index. Serum levels of miR-155, ALT, AST, and IL-1ß were lower while SOD, GSH, and IL-1ra were higher in OG than in PG. MiR-155 mimic increased the levels of ALT, AST, and IL-1 β and reduced the levels of SOD, GSH, and IL-1ra. In contrast, miR-155 inhibitor had reverse results. Therefore, ONAS has better improvement in complications and therapeutic effects on DLD by regulating serum miR-155.

Keywords: alginate oligosaccharide, antioxidant, anti-inflammatory, fusion rate, infection rate, Japanese Orthopaedic Association, Oswestry Disability Index

Introduction

Degenerative lumbar disease (DLD) is a significant issue for public health and may cause compression fractures and abnormal spinal curvature. Most osteoporosis patients with DLD have the experiences of back and leg pains. Furthermore, DLD leads to

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With the development of biomechanical research, the cage has been developed from a cylindrical form to rectangular, mesh, oval, and other forms. PLIF with cages (PLIFC) has caught much attention for its superiority in biomechanics, but high incidence of thin-layer chromatography-related complications may cause a new problem.⁵ In contrast, PLIF is a common choice in the treatment of osteoporosis patients with chronic DLD. The incidence and severity of complications related to cages remain the main problem in the surgery.

Alginate oligosaccharides (AOS) as natural products have beneficial effects on human health, including antioxidant^{6,7} and anti-inflammatory activities,8 and can inhibit the development and deterioration of the disorders caused by the surgery. DLD is often associated with inflammatory status9 and oxidative stresses.¹⁰ AOS may be beneficial for preventing DLD progression.^{11,12} Nanotechnology has provided the possibility of delivering drugs to target tissues via nanoparticles. In this way, unwanted adverse effects can be controlled because no higher-dose medicine is needed. However, the related molecular mechanisms for the functions of oligosaccharide nanomedicine of alginate sodium (ONAS) on DLD patients remain widely unknown. MiR-155, a multifunctional microRNA, has been found to be involved with inflammatory responses by mediating various genes.13 In contrast, miR-155 deletion can reduce the generation of reactive oxygen species and oxidative stress.14 AOS may exert their antioxidant and anti-inflammatory functions by affecting the levels of miR-155. Therefore, the efficacy and safety of ONAS on the posterior lumbar interbody fusion with cages were investigated in DLD treatment by investigating the changes of serum miR-155.

Methods AOS molecular weight was determined by electrospray ionisation mass spectrometry (ESI) MASS

AOS were purchased from Seebio Biotech (Shanghai) Co., Ltd. (Shanghai, China) and dissolved in ddH₂O. AOS were prepared as follows: alginate from Lonicera japonica was purchased from Shanghai Macklin Biochemical Co., Ltd. (Shanghai, China), and the mannuronic acid to guluronic acid ratio (M/G) is 2.28. Alginate lyase was prepared according to an earlier report.¹⁵ Ten grams of alginate was dissolved in 1 L 20 mM Tris-HCl buffer (pH 7.0), and 10 U alginate lyase was added. The reaction was performed at 39°C for 5 min, and then, the solution was heated to 100°C to stop the reaction. Ethanol was added to final concentration 50% (v/v) to remove the precipitate via centrifugation at $1,000 \times g$ for 10 min. The supernatant was filtered by a 0.45 µm membrane and lyophilized. To ensure the removal of trace salt, the sample was purified by using a Micro Bio-Spin 6 Column (Bio-Rad Laboratories Inc., Hercules, CA, USA) prior to MS analysis. Samples are typically analyzed at 1% (w/v) concentration.

MS data were obtained on a solariX FT-ICR Mass Spectrometer (Bruker Corporation, Billerica, MA, USA). In general, ionization is achieved by applying a potential of 1.4–1.8 kV and setting the drying gas to 120°C. Based on molecular weight of the analyzed AOS components to adjust the ion transmission radio frequency (RF) and flight time. The resulting mass spectra were externally calibrated using an ESI tuned mixture (Agilent Technologies, Santa Clara, CA, USA) and analyzed by using data analysis software (Bruker Corporation).

Normal nanoparticle preparation

Nanoparticles are typically sized in nanoscale systems at 10–1,000 nm. These systems contain drugs by embedding them in the matrix of the particles or attaching them to the matrix of the particles. The nanocapsules are blister systems in which the drug is confined in the oily or aqueous liquid core and surrounded by the polymer film. The nanospheres are spherical particles having gelatinized internal particles in which the entrapped components are physically dispersed. Nanocapsules with structured cores have also been designed and are essentially a combination of nanocapsules and nanospheres. They are usually prepared by preparing a nanosphere and subsequently forming an additional shell on the interface of the nanosphere.

Two grams of ampicillin was dissolved in 10 mL ddH₂O and mixed with pluronic-F68 10 g, phosphoprotein 2.0 g, glycerol 30 g, 70% sorbitol 21 g, and 35 mL 96% ethanol. The process can be described as follows: the phospholipid was dissolved in ethanol at room temperature. Pluronic-F68 was added and stirred at 37°C. Ampicillin was added and stirred for 10 min. Glycerol was added and stirred until the solution became clear. Finally, sorbitol was added and continued to stir until the solution was clear. The mixture was sterilized by 0.2 μ m microporous membrane, and filtered solution was lyophilized and stored at –20°C freezer.

ONAS preparation

Two grams of ampicillin was dissolved in 10 mL ddH₂O and mixed with AOS 10 g, phosphoprotein 2.0 g, glycerol 30 g, 70% sorbitol 21 g, and 35 mL 96% ethanol. The process can be described as follows: the phospholipid was dissolved in ethanol at room temperature. AOS were added and stirred at 37°C. Ampicillin was added and stirred for 10 min. Glycerol was added and stirred until the solution became clear. Finally, sorbitol was added and continued to stir until the solution was clear. The mixture was sterilized by 0.2 μ m microporous membrane, and filtered solution was lyophilized and stored at -20° C freezer.

Morphology analysis of nanoparticles by transmission electron microscopy (TEM)

The morphology of nanoparticles was observed by using a Philips CM12 TEM (FEI, Hilsboro, OR, USA) with 100 kV accelerating voltage. One drop of the suspension diluted with water was placed on a carbon-coated copper grid and dried in the air before observation.

The effects of AOS toxicity on MG-63 human osteosarcoma cells

MG-63 human osteosarcoma cells were purchased from the Cell Bank of Chinese Academy of Sciences (Shanghai, China) and cultured in DMEM with 10% fetal bovine serum (FBS) at 37°C and 5% CO₂. The cell monolayer was digested by trypsin when the cells grew up to 90% confluency. The cells were washed with fresh DMEM three times and adjusted to 1×10^6 cells/mL. Different concentrations (0, 0.1, 1, 10, 100, and 1,000 µg/mL) of AOS were added to MG-63 human osteosarcoma cells and further cultured for 3 days. Trypan blue was used to measure live cells under a microscope. Meanwhile, [³H]thymidine absorption was used to measure the activity of the cells. Two hundred microliters of cell suspension of 1.0×10^5 cells/mL was added to each well of a 96-well plate. After 1-day culture, cells were labeled with 1 Ci/well of [³H]thymidine for 3 h. Absorption values were measured with a liquid scintillation counter (model SN 6930; Shanghai Hesuo Rihuan Photoelectric Instrument Co., Shanghai, China).

Cell transient transfection

Different concentrations (0, 0.1, 1, 10, and 100 μ g/mL) of AOS were added to cells and further cultured for 3 days under the same condition. The level of miR-155 was measured. The cells were transfected with miR-155 mimic or inhibitors (single-stranded chemically modified oligonucleotides; Shanghai GenePharma Co., Ltd., Shanghai, China) by using Lipofectamine[®] 2000 (Thermo Fisher Scientific, Waltham, MA, USA).

Patient screening

Before the experiment, all procedures were approved by the Ethics Committee of The Second Hospital of Jilin University (approval no 20151213002N). Written informed consent was obtained from each patient with his/her signed name. The following inclusion criteria were used: 1) prospective, randomized, and/or nonrandomized controlled studies for DLD therapy; 2) the DLD osteoporosis patients had the symptoms of lumbar spinal stenosis, instability and spondylolisthesis (I and II degrees), and lumbar disk herniation; 3) the osteoporosis patients had low back pain, intermittent claudication, lumbar spinal canal decompression, and persistent lower extremity symptoms; 4) the osteoporosis patients with DLD were confirmed by auxiliary examination (X-ray, CT, and MRI).

The following exclusion criteria were used: 1) the osteoporosis patients had severe intervertebral space collapse with peripheral osteophyte formation; 2) the osteoporosis patients had lumbar degenerative scoliosis (Cobb angle $>10^\circ$); 3) the osteoporosis patients suffered from osteoporosis; 4) the osteoporosis patients suffered from diabetes; 5) the osteoporosis patients had serious medical diseases and contraindications; and 6) the osteoporosis patients had mental disorders.

Patient grouping

Based on the inclusion and exclusion criteria, 96 DLD patients were recruited from May 12, 2014, to February 14, 2015. All the patients received PLIFC from March 3–13, 2016, at our hospital. After surgery, all patients were evenly assigned into ONAS group (OG, daily oral administration of 500 mg ONAS) and control group (PG, daily oral administration istration of 500 mg pluronic nanoparticles).

Blood nanoparticle analysis

A blood sample was collected from each patient in tubes containing EDTA anticoagulant. Blood smears were prepared immediately from samples taken with anticoagulant. The blood smear was made on clean glass slides and observed under a light microscope (Olympus, Tokyo, Japan) at 100× magnification under oil immersion.

Data extraction and analysis

The following parameters were measured and analyzed: the number of osteoporosis patients, sex ratio, patient age, follow-up time, patient diagnosis, surgical segments, the number of osteoporosis patients who received cage fusion, intraoperative blood loss, the duration of surgery and hospital stay, preoperative and prospective visual analog scale (VAS), Japanese Orthopaedic Association (JOA), and Oswestry Disability Index (ODI) scores, fusion rates, and complication rates. Data were processed by RevMan 5.3 (Cochrane Collaboration, Oxford, UK). The data were collected and analyzed quantitatively. For the continuity index, the deviation was pooled into weighted mean difference (WMD) and the 95% confidence interval (CI) was used for the statistics. For the discontinuity index, the odds ratio risk, relative risk (RR), and 95% CI were used to evaluate the dichotomous results. The heterogeneity of the included studies was judged by the P-value, and the heterogeneity was judged according to l^2 value. If P > 0.1, $l^2 < 0.5$, they could be considered homogeneous with similar studies and a fixed-effect model would be used for meta-analysis; if P < 0.1, $I^2 \ge 0.5$, a random-effect model would be selected. If there was heterogeneity between the studies, the original data and extraction methods would be reanalyzed. A descriptive method would be considered if the data could not be analyzed by using the two methods.

Heterogeneity analysis

The sources of heterogeneity were identified and examined in sub-group and sensitivity analyses. Heterogeneity was evaluated by using I^2 values, of which <25% was regarded as low heterogeneity, 50% was regarded as mild, and 75% was regarded as high. If the effect of the merger between the patients with cage fusion and the patients without cage fusion was decreased significantly, P<0.1, and the heterogeneity could not be determined.

Quantitative RT-PCR analysis

Blood samples were obtained from DLD patients, and serum was purified within 1 h. Total RNA was isolated by using an RNA purification kit (TRI Reagent[®]; Molecular Research Center, Cincinnati, OH, USA). The level of miR-155 was normalized to total RNA. qRT-PCR was carried out on a 7500 Real-Time PCR System (Thermo Fisher Scientific) by using a universal reaction condition: 95°C for 2 min, followed by 40 cycles of 95°C for 10 s and 68°C for 30 s. After the PCR, $C_{\rm T}$ values were calculated and expressed by using 2^{-ΔG}.

Biochemical analysis

The serum activity of superoxide dismutase (SOD) was measured by formazan-WST method. The serum concentration of reduced glutathione (GSH) was determined by the dithiobis-2-nitrobenzoic acid method. The serum concentrations of aspartate aminotransaminase (AST) and alanine aminotransferase (ALT) were evaluated by biochemical analyzer (Fenghui Co., Ltd., Shanghai, China). The levels of IL-1 β and IL-1ra were measured by using ELISA kits (catologue numbers ab46052 and ab174450) from Abcam (Shanghai, China).

Statistical analysis

Student's *t*-test and chi-squared statistic tests were performed using SPSS Version 20.0. The differences were considered statistically significant if P < 0.05.

Results

Characterization of ONAS

The main components of AOS (DP20 to DP35) were confirmed by ESI MS under the conditions that produced mass spectra with $M+H^+$. Figure 1 shows that the masses predicted from DP20 (C120H142O121Na20) to DP35 (C210H247O122Na35) are 4,176 and 7,146 Da, respectively.

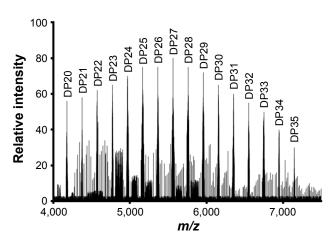


Figure 1 ESI mass spectrometry analysis of bioactive fractions from ONAS under the conditions that produced mass spectra with $M\!+\!H^*\!.$

Abbreviations: ESI, electrospray ionisation; ONAS, oligosaccharide nanomedicine of alginate sodium.

Note: Mass spectra were visualized following the separation of DP20 (M+H $^+$ = 4,177 Da) to DP35 (M+H $^+$ =7,147 Da).

Morphological characterization of nanoparticles

The TEM was used to characterize the morphology of the nanoparticles. Figure 2 shows the TEM images for two samples of nanoparticles obtained by using pluronic-F68 (Figure 2A) or AOS (Figure 2B). All nanoparticles are in tube form. These images show that the particle size of nanoparticles is <200 nm. Light microscope images revealed that blood cells were regularly spherical in shape and had a generally smooth surface with an average size of ~10 μ m. Nanoparticles could be observed in the patients with pluronic nanoparticles (Figure 2D) or ONAS nanoparticles (Figure 2E) when compared with a control without nanoparticles (Figure 2C).

The effects of AOS toxicity on MG-63 human osteosarcoma cells

To avoid the effects of AOS toxicity on MG-63 human osteosarcoma cells, a suitable concentration of AOS was determined. Trypan blue analysis indicated that exposure of the MG-63 cells to AOS, in the presence of $>100 \,\mu$ g/mL, resulted in the loss of cell viability (Figure 3A, P<0.05). The toxicity of AOS was determined via [³H]thymidine incorporation into DNA after exposure to different concentrations of AOS. There was significant reduction in [³H]thymidine uptake when the concentration was >100 μ g/mL (Figure 3B, *P*<0.05). Based on these results, we assessed that administration of 100 mg AOS daily will not cause toxicity to human cells.

AOS reduce miR-155 level in MG-63 human osteosarcoma cells

qRT-PCR analysis showed that AOS reduce miR-155 level in MG-63 human osteosarcoma cells when the concentration was $>1 \mu g/mL$ (Figure 4). Meanwhile, the level of miR-155 was further reduced with the increase in the AOS concentration (Figure 4). The results suggest that AOS reduce miR-155 level in MG-63 human osteosarcoma cells.

MiR-155 increase oxidant stress of MG-63 human osteosarcoma cells

Table 1 shows that miR-155 mimics reduced antioxidant functions by inhibiting the activities of SOD and GSH in MG-63 cells and increasing the levels of ALT (P<0.05). In contrast, miR-155 inhibitor increased antioxidant functions by improving the activities of SOD and GSH in MG-63 cells and reducing the levels of ALT (P<0.05). The results suggest that miR-155 increases oxidant stress of MG-63 cells.

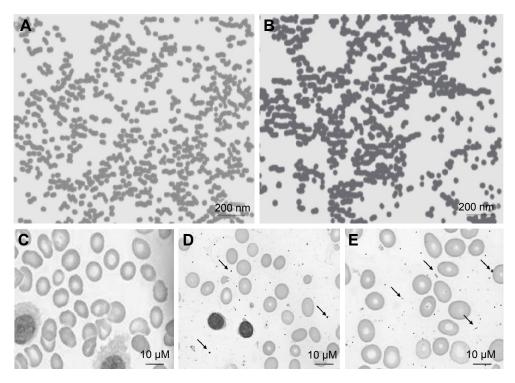


Figure 2 Images of nanoparticles.

Notes: (A) TEM images of pluronic nanoparticles. (B) TEM images of AOS nanoparticles. (C) Light micrograph of blood cells in a human blood smear without nanoparticles. (D) Light micrograph of blood cells in a human blood smear with pluronic nanoparticles. (E) Light micrograph of blood cells in a human blood smear with AOS nanoparticles. (Arrows show the nanoparticles in blood samples.

Abbreviations: AOS, alginate oligosaccharides; TEM, transmission electron microscopy.

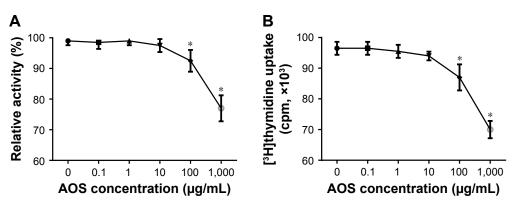


Figure 3 The effects of AOS on the viability of MG-63 cells.

Notes: (A) Cell viability of MG-63 was measured using Trypan blue. (B) Cell viability of MG-63 was measured using [${}^{2}H$]thymidine uptake. The data are presented as the mean \pm standard deviation. n=5 in each group.

Abbreviation: AOS, alginate oligosaccharides.

MiR-155 increases inflammatory activities of MG-63 human osteosarcoma cells

ELISA analysis showed that there was no statistically significant difference for serum levels of IL-1 β (Figure 5A, P>0.05) and IL-1ra (Figure 5B, P>0.05) on 0-day culture. After 3-day culture, miR-155 mimics increase the serum levels of IL-1 β (Figure 6A, P<0.05) and reduce the levels of IL-1ra (Figure 5B, P<0.05). In contrast, miR-155 inhibitors decrease the serum levels of IL-1 β (Figure 5A, P<0.05) and increase the serum levels of IL-1 β (Figure 5B, P<0.05). All the results suggest that miR-155 increases inflammatory activities of MG-63 human osteosarcoma cells.

Baseline clinical characterization of patients

A total of 96 patients were recruited, including 59 males and 37 females. All patients underwent PLIFC. Baseline characteristics of patients in the study are shown in Table 2.

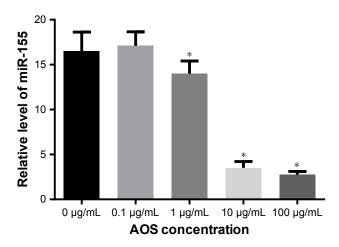


Figure 4 The effects of AOS on the level of miR-155 in MG-63 human osteosarcoma cells.

Notes: *P < 0.05 via a control group without AOS addition. There are significantly statistical differences if P < 0.05.

Abbreviation: AOS, alginate oligosaccharides.

8464 submit your manuscript | www.dovepress.com Dovepress There was no significant difference in age between the two groups of osteoporosis patients with degenerative lumbar disease at mean age 32.4 years (range 24.1–40.0 years). There was no pseudarthrosis, instrumentation failures, or significant subsidence during the surgery. Disk height was increased from 5.3 ± 2.1 mm before surgery to 9.5 ± 3.1 mm after surgery. The mean operative time was 142 ± 34 min. The mean blood loss was 716 ± 326 mL in all patients. There was no obvious difference in the baseline characters for preoperative symptoms between the two groups (P > 0.05). There were significant differences in the postoperative symptoms, infection, and back pain between the two groups (P < 0.05).

Integrated analysis of clinical efficacy

The main curative efficacy was evaluated by using VAS, JOA, and ODI scores, complications, and fusion rate.

The VAS score was used to assess the pain levels. There was no statistically significant difference in the VAS scores between the two groups in a fixed-effect model before surgery (WMD =-0.08, 95% CI [-0.39, 0.23], P=0.63). There was no statistically significant difference for VAS score in a random-effect model between the two groups at the final follow-up examination after surgery (WMD =0.22, 95% CI [-2.49, 2.93], P=0.87).

The JOA score was used to evaluate the functional recovery of lumbar vertebrae. There was no statistically significant difference in the JOA score between the two groups before surgery (WMD = 0.29, 95% CI [-0.31, 0.89], P=0.35). OG showed better improvement when compared with PG after surgery (RR = 0.52, 95% CI [0.33, 0.84], P=0.007).

The complication rates were similar in a fixed-effect model between the two groups before surgery (RR =0.85, 95% CI [0.64, 1.13], P=0.26). The fusion rates of the lumbar vertebrae were evaluated in final follow-up examination

Table I The effects of miR-155 on the antioxidant activities in MG-63 human osteosarcoma cells

Group	SOD (U/mL)	GSH (ng/L)	ALT (U/mL)	AST (U/mL)
Control	21.36±3.17	20.78±2.35	40.93±4.57	83.25±11.36
MiR-155 mimic	10.23±3.29*	8.17±1.58*	67.39±9.75*	84.33±15.28
MiR-155 inhibitor	46.13±6.48*	42.36±9.36*	26.99±8.74*	86.78±11.49

Notes: Eight blood samples were analyzed in each group. All data are presented as mean value \pm SD. *P<0.05 vs a control group.

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransaminase; GSH, glutathione; SOD, superoxide dismutase.

after surgery. The fusion rates were higher in OG than in PG (RR =0.64, 95% CI [0.48, 0.84], *P*=0.001).

The secondary clinical outcomes were evaluated by using surgical duration, the amounts of intraoperative blood loss, height of intervertebral space, ODI score, and abnormal movement of lumbar segment. The surgical duration was similar between the two groups (Table 2). There was no statistically significant difference between the two groups in a random-effect model (WMD =14.69, 95% CI [-8.24, 37.62], P=0.21). There was no statistically significant difference for the quantity of blood loss between groups in a random-effect model (WMD =21.96, 95% CI [-67.44, 111.36], P=0.63). There was a statistically significant difference for the increase of the height of intervertebral disk between the two groups in a random-effect model (WMD =1.31, 95% CI [0.78, 1.85], P<0.001).

ODI score was used to evaluate the functional recovery of the lumbar vertebrae. Three studies reported ODI score before surgery and at the final follow-up examination after surgery. There was no statistically significant difference in the ODI score in a fixed-effect model between the two groups (WMD =0.99, 95% CI [-0.48, 2.46], P=0.19). There was no statistically significant difference for ODI score in the random-effect model between the two groups at the final follow-up examination after surgery (WMD =2.52, 95% CI [-2.26, 7.29], P=0.30).

There was no statistically significant difference for abnormal movement of lumbar segment between the two groups in a fixed-effect model (WMD =-0.37, 95% CI [-1.96, 1.22], P=0.65). Similarly, there was no statistically significant difference for abnormal movement of lumbar segment between the two groups in a random-effect model (WMD =-0.05, 95% CI [-1.1, 0.02], P=0.16).

Quality assessment of included studies

Methodological Index for Non-Randomized Studies (MINORS) was used to assess bias risk of nonrandomized trials and bias risk profile. Furthermore, the comparison was performed before and after surgery for all the osteoporosis patients. All studies had low-risk publication and attrition bias. The Grading of Recommendations Assessment, Development and Evaluation (GRADE) system was used to evaluate the quality of the evidence, and the results had an average MINORS score of 15.81. Prospective studies and randomized control trials were included, and the related parameters (coherence in osteoporosis patients, key indicators, adequate

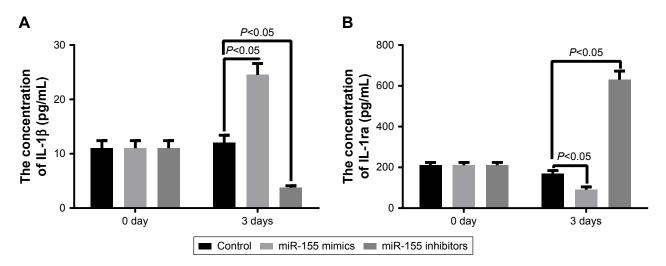


Figure 5 The effects of miR-155 on the anti-inflammatory activities in MG-63 human osteosarcoma cells.

Notes: (A) The effects of miR-155 on the level of IL-1 β in MG-63 cells. (B) The effects of miR-155 on the level of IL-1ra in MG-63 cells. There are significant statistical differences if P<0.05 vs a control group.

Abbreviations: IL-1 β , interleukin-1 β ; IL-1ra, interleukin-1 receptor antagonist.



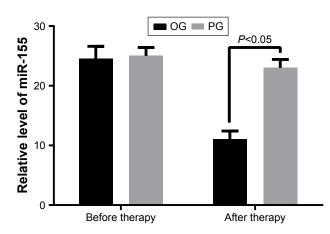


Figure 6 The effects of ONAS on the level of miR-155 in DLD patients. **Notes:** ONAS group (OG, daily oral administration of 100 mg ONAS) and control group (PG, daily oral administration of 100 mg pluronic nanoparticles). There are significant statistical differences between OG and PG groups if P<0.05. **Abbreviations:** DLD, degenerative lumbar disease; ONAS, oligosaccharide nanomedicine of alginate sodium.

follow-up time, follow-up rate, sample size, control group, baseline characters, and statistical analysis) were investigated. The results demonstrated that most studies were with low-risk bias.

Serum level of miR-155 was higher in PG than in OG

qRT-PCR analysis showed that there was no significant statistical difference for serum level of miR-155 between

 Table 2 Baseline characters of all degnerative lumbar disease

 patients

Characteristics	OG	PG	P-values
	(n=48)	(n=48)	
Male (%)	29 (58.3)	31 (64.6)	0.67ª
Mean age, years (SD)	33.2 (6.8)	31.5 (7.4)	0.08 ^b
Operative time, min (SD)	140 (30)	146 (36)	0.26 ^b
Blood loss, mL (SD)	712 (321)	735 (328)	0.49 ^b
Disk height increase, mm (SD)	4.1 (1.8)	3.9 (2.1)	0.17 ^b
Preoperative symptoms, n (%)			
Dyspnea, n (%)	18 (37.5)	16 (33.3)	0.67ª
Back pain, n (%)	20 (41.6)	21 (43.8)	0.84ª
Fatigue, decreased energy, n (%)	11 (22.9)	9 (18.8)	0.62ª
Tachycardia/palpitations, n (%)	6 (12.5)	7 (14.6)	0.77ª
Dizziness, n (%)	2 (4.2)	3 (6.3)	1.00ª
Syncope, n (%)	3 (6.3)	2 (4.2)	1.00ª
Dysphagia, n (%)	3 (6.3)	3 (6.3)	1.00ª
Postoperative complications, n (%)			
Surgery reintervention, n (%)	0 (0.0)	5 (10.4)	0.07ª
Surgery infection, n (%)	0 (0.0)	6 (12.5)	0.04ª
Chronic wound infection, n (%)	I (2.I)	8 (16.7)	0.04ª
Prolonged pain, n (%)	1 (2.1)	9 (18.8)	0.02ª

Notes: There are significantly statistical differences if P<0.05. ^aChi-squared test. ^bStudent's *t*-test.

Abbreviations: OG, alginate oligosaccharide treated; PG, placebo treated.

Antioxidant activities were higher in OG than in PG

Before nanoparticle treatment, there was no significant statistical difference for antioxidant activities between OG and PG groups according to activities of SOD, GSH, ALT and AST (Table 3, P>0.05). After 1-month therapy, serum levels of SOD and GSH were higher in OG than in PG (Table 3, P<0.05) while serum levels of ALT were lower in OG than in PG (Table 3, P<0.05). The results suggest that AOS treatment increases antioxidant activities of DLD patients.

Anti-inflammatory activities were higher in OG than in PG

ELISA analysis showed that there was no significant statistical difference for serum levels of IL-1 β and IL-1ra between OG and PG groups before nanoparticle treatment (P>0.05) (Figure 7). After 1-month therapy, serum level of IL-1 β was higher in PG than in OG (P<0.05) (Figure 7A), while serum level of IL-1ra was lower in PG than in OG (P<0.05) (Figure 7B). The results suggest that AOS treatment increases anti-inflammatory activities of DLD patients.

Discussion

PLIF has been proposed for many decades, applied to clinical trials, and developed into basic techniques for spine surgery. Cage fusion belongs to the interfacial fixation technique. Some articles report that the disadvantages of cage fusion are with long surgical duration, more hemorrhage, and nerve root injury.¹⁶ Each method has advantages and disadvantages, and the value of PLIFC still needs to be investigated. Therefore, we compared 2,138 osteoporosis patients undergoing PLIFC, and 1,037 osteoporosis patients undergoing PLIFC and without cages in DLD treatment.

VAS, JOA, and ODI scores were used to evaluate clinical outcomes of each method. There was no statistically significant difference for these scores in a fixed-effect model between the two groups before surgery and immediate examination after surgery. After the merger collected data from literature, the difference was statistically significant

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	Group	SOD (U/mL)	GSH (ng/L)	ALT (U/mL)	AST (U/mL)
Before therapy	OG	27.24±3.26	24.15±2.14	45.12±10.43	103.32±25.17
	PG	26.25±4.16	23.23±1.93	47.79±8.40	108.26±19.64
After therapy	OG	32.34±5.34*	34.38±8.02*	31.45±12.18*	106.19±25.28
	PG	27.34±2.32	24.28±1.74	48.79±12.36	109.42±24.48

Table 3 Biochemical parameters of enzyme activities for DLD patients

Notes: Eight blood samples were analyzed in each group. All data are presented as mean value \pm SD. **P*<0.05 vs PG group.

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransaminase; DLD, degenerative lumbar disease; GSH, glutathione; OG, alginate oligosaccharide treated; PG, placebo treated; SOD, superoxide dismutase.

in a fixed-effect model between the two groups (RR =0.52, 95% CI [0.33, 0.84], P=0.007). There was no heterogeneity since I^2 =34%. The results of the merger fell on the left side of the invalid line, indicating that AOS had better therapeutic results in the PLIFC.

Although the small number of samples, the heterogeneity of the study was not a reason, the results had clinical significance. Fusion rate has become one of the controversy issues in the PLIFC fusions.^{17,18} There were statistically significant differences for fusion rates in a fixed-effect model between the two groups at final follow-up examination after surgery (RR =0.64, 95% CI [0.48, 0.84], P=0.001). There was no heterogeneity in the selected articles, and the results of the merger fell on the left side of the invalid line. Therefore, the group undergoing the ONAS had a higher fusion rate than the PG group.

Surgical duration is an important quality metric for spinal fusion, and decreased surgical time will be beneficial for improving patient outcomes. Here, the present study was used to compare the surgical duration. The results demonstrated that the surgical duration was similar between the two groups in a random-effect model (WMD =14.69, 95% CI [-8.24, 37.62], P=0.21). The heterogeneity of the combined results was not significant.

Fusion surgery is the main method for the therapy of DLD, but the method is often accompanied with substantial blood loss. We investigated the differences of surgical blood loss, in the osteoporosis patients undergoing the surgery for DLD, between groups. There was no statistically significant difference for blood loss between the two groups (WMD =21.96, 95% CI [-67.44, 111.36], P=0.63).

For both groups, the abnormal motion of the surgical segment was similar in a fixed-effect model between the two groups before surgery and at the final follow-up examination after surgery. Abnormal motion may be caused by physiological loads, and thus, it is not associated with an obvious increase in motion. In statistical analysis, abnormal motion is often followed by non-Gaussian heavy-tailed behavior and other activities. It may take a long time to stimulate abnormal motion.

Alginate particles have been reported to have antioxidant activities,^{19,20} but the related molecular mechanisms remain unknown. AOS have antioxidant and anti-inflammatory activities and may be suitable for infection therapy. Thus, we want to find the microRNA associated with oxidative stress and inflammation. In a review paper, >10 microRNAs have been reported to regulate oxidative stress and/or inflammation.²¹

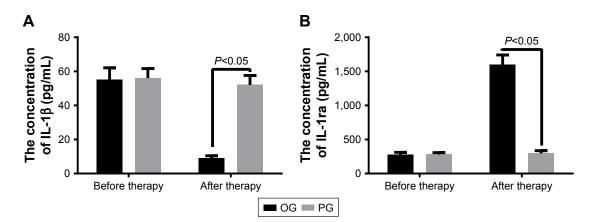


Figure 7 The effects of ONAS on the level of cytokines in DLD patients.

Notes: ONAS group (OG, daily oral administration of 100 mg ONAS) and control group (PG, daily oral administration of 100 mg pluronic nanoparticles). (**A**) The effects of ONAS on the level of IL-1 β in DLD patients. (**B**) The effects of ONAS on the level of IL-1ra in DLD patients. There are significant statistical differences between OG and PG groups if P<0.05.

Abbreviations: DLD, degenerative lumbar disease; IL-1 β , interleukin-1 β ; IL-1ra, interleukin-1 receptor antagonist; ONAS, oligosaccharide nanomedicine of alginate sodium.

However, only miR-155 has been clearly reported to be associated with both oxidative stress and inflammation.²² Thus, miR-155 may be a potential target of AOS and was chosen in the present study. Here, we find that AOS can increase the antioxidant activities of DLD patients by downregulating the serum levels of miR-155 (Figure 4 and Table 2). Meanwhile, miR-155 is a main factor for inducing inflammatory responses.^{23,24} Thus, AOS also increases antiinflammatory properties of DLD patients by controlling miR-155 level. Hence, all microRNA should be screened in future work to confirm the present results.

ONAS is mainly made of AOS, which have a mild reaction condition and excellent gel properties and are easy to purify. AOS are environmentally friendly, pH neutral, and biodegradable with fewer side effects for human health. More importantly, AOS are made from alginate, which has abundant supplies in nature. Thus, AOS can be used world wide. The main difference between OG and PG is that AOS are used in OG and pluronics are used in PG. Both nanomaterials have sparked a considerable interest as vehicles used for therapeutic agents. Pluronic block copolymers affect cellular functions such as mitochondrial respiration, ATP synthesis, apoptosis, and gene expression.²⁵ AOS and their derivatives can delay the growth of tumors and reduce tumor size. AOS may increase regulatory effects on the immune organs, such as spleen.²⁶

There are some limitations for the present systematic review: 1) prospective randomized controlled trials were lacking; 2) the sample size was limited with the available number of osteoporosis patients and the small number of cases; a larger sample size is needed to improve statistical performance; 3) homogeneity of the complications was lacking and should be investigated in the future, especially in the study of nonrandomized controlled trials and in a large multicenter randomized controlled study; and 4) solubility of nanomaterials in water is important, to avoid immune clearance in human hosts. Low-molecular-weight AOS are cationic and highly soluble, although their solubility is limited by viscosity when the concentration is >1% (v/v). High-molecular-weight AOS are insoluble, although they may have a better activity. Improving the solubility of highmolecular-weight AOS will be the main work in the future.

Conclusion

PLIFC had higher fusion rate and JOA scores with ONAS administration. AOS had better improvement in complication rates than a PG group. AOS provide a better option for DLD therapy. AOS have been shown to be superior to normal nanoparticles by reducing miR-155, which affects antioxidant and anti-inflammatory properties of DLD patients. Further work is still needed to confirm the present results in the future.

Disclosure

The authors report no conflicts of interest in this work.

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