

A personalized approach to acute myeloid leukemia therapy: current options

This article was published in the following Dove Press journal:
Pharmacogenomics and Personalized Medicine

Raveen Stephen Stallon
Illangeswaran
Saswati Das
Daniel Zechariah Paul
Vikram Mathews
Poonkuzhali
Balasubramanian

Department of Haematology, Christian
Medical College, Vellore, India

Abstract: Therapeutic options for acute myeloid leukemia (AML) have remained unchanged for nearly the past 5 decades, with cytarabine and anthracyclines and use of hypomethylating agents for less intensive therapy. Implementation of large-scale genomic studies in the past decade has unraveled the genetic landscape and molecular etiology of AML. The approval of several novel drugs for targeted therapy, including midostaurin, enasidenib, ivosidenib, gemtuzumab–ozogamicin, and CPX351 by the US Food and Drug Administration has widened the treatment options for clinicians treating AML. This review focuses on some of these novel therapies and other promising agents under development, along with key clinical trial findings in AML.

Keywords: acute myeloid leukemia, personalized medicine, chemotherapy, molecular markers

Background

Acute myeloid leukemia (AML) is characterized by blocked myeloid-lineage differentiation and accumulation of myeloid blast cells in the bone-marrow that results in catastrophic bone-marrow failure.^{1,2} AML is a highly heterogeneous disease driven by different mutations that affect signaling pathways, transcription factors, and epigenetic regulators.³ The current treatment for AML starts with induction chemotherapy (7+3), followed by a few cycles of consolidation chemotherapy or an allogeneic hematopoietic stem-cell transplantation (allo-HSCT).⁴

The disease's prognosis and outcomes in adults and the elderly, who account for the majority of AML patients, remains poor.^{5,6} The ever-growing advancement in genomic technologies allows us to have an unprecedented view of the spectrum and frequency of mutations, their clonal nature, and their evolution during progression of the disease and the epigenetic modulation of the disease. Incorporating “omics” data with ex vivo high-throughput drug screening has the potential to tailor therapy for patients.⁷ This review provides an overview on novel targeted therapies, approved drugs, and ongoing clinical trials with the aim of personalizing AML therapy (Figure 1).

A nucleoside analogue and an antibiotic

Cytarabine, a nucleoside analogue, and anthracyclines (daunorubicin, idarubicin, doxorubicin) derived from antibiotics have been an integral part of the chemotherapy regimen in treating AML for the past 5 decades. Cytarabine as a continuous infusion for 7 days with addition of anthracyclines as a short infusion in the first 3 days forms the basis of chemotherapy (7+3 regimen), drastically reducing the leukemia burden in

Correspondence: Poonkuzhali
Balasubramanian
Department of Haematology, Christian
Medical College, Vellore 632004, India
Tel +91 416 228 3476
Email bpoonkuzhali@cmcvellore.ac.in

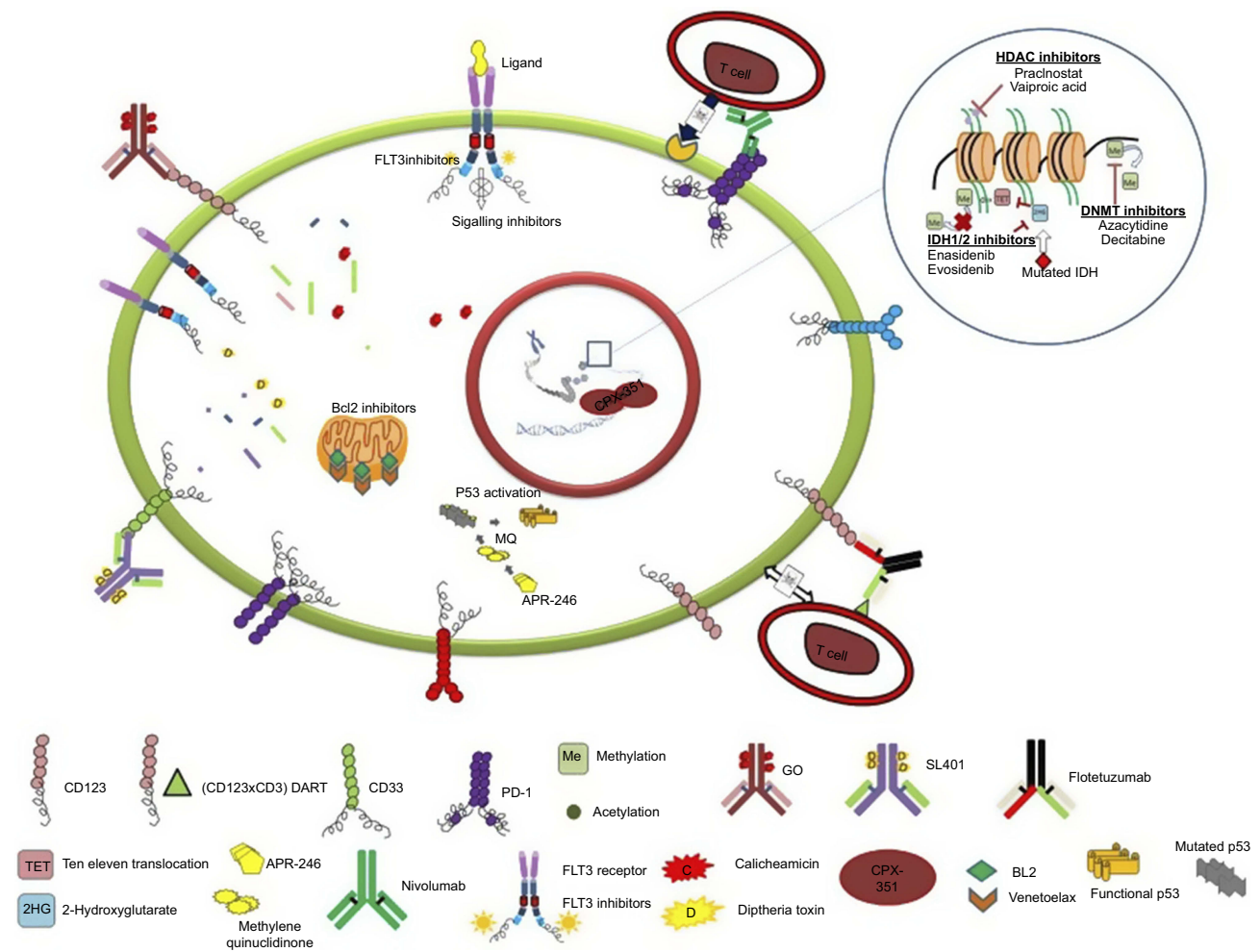


Figure 1 Illustration of recently approved and novel drugs in clinical trials targeting the different entities of leukemic cells. **Notes:** CPX351 causes enhanced DNA damage. Epigenetic drugs target aberrant methylation and acetylation patterns. Immunomodulating molecules nivolumab and flotetuzumab enhance cytotoxic activity of T cells. GO and SL401-based monoclonal antibody therapy selectively targets leukemic cells. FLT3 inhibitors obstruct proliferative signals. Venetoclax targets BCL2 and reduces mitochondrial fitness. IDH inhibitors enasidenib and ivosidenib deplete oncometabolites in leukemic cells and prevent aberrant methylation patterns. Stabilization of mutated p53 by APR246 leads to enhanced tumor-suppressor activity.

patients. This, followed by high-dose cytarabine therapy for consolidation or allo-HSCT, has remained the standard treatment for AML patients. We and others have evaluated the mechanisms of resistance to cytarabine and anthracyclines extensively, and have shown how leukemic cells evolve and adapt to become resistant to chemotherapy.⁸⁻¹⁴ In vitro and preclinical studies with novel agents in combination with standard chemotherapeutic drugs have shown to overcome drug resistance in AML.¹⁵⁻¹⁹ This could pave the way to the introduction of promising agents that would help to personalize therapy for AML patients.

CPX351

Cytarabine and daunorubicin have been the backbone of conventional chemotherapy in AML over the past four

decades. CPX351, a novel nanoliposomal formulation of cytarabine and daunorubicin in a 5:1 molar ratio, leads to enhanced intracellular uptake of the drug and reduced clearance compared to the conventional 7+3 regimen.²⁰⁻²² A multicenter phase III trial comparing CPX351 with standard induction and consolidation among elderly patients showed significantly improved overall survival (OS; 9.6 months vs 5.9 months, $P=0.005$) with CPX351 compared to the standard 7+3 arm (A group of patients receiving a specific treatment). Patients in the CPX351 arm showed a substantial increase in mean terminal half-life compared to the conventional 7+3 infusion (24.5 hours for CPX351 vs 3 hours for cytarabine), although the time to neutrophil and platelet recovery was longer. Also, more patients in the CPX351 arm were eligible for an allotransplant than the 7

+3 arm (34% versus 25%).²³ CPX351 was approved by the FDA in 2017 for fit elderly patients with treatment-related AML (t-AML) and for de novo AML with myelodysplastic syndrome (MDS).

Hypomethylating agents

The hypomethylating agents azacitidine (Aza) and decitabine are analogues of cytosine with replacement of the fifth carbon atom by a nitrogen atom.²⁴ For the past decade, they have been commonly used in AML patients who are not eligible for cytarabine-based intensive therapy. Aza has a short half-life.²⁵ The new oral formulation of Aza, CC486 is in early-stage trials testing its efficacy. CC486 could possibly enhance antileukemic activity by increasing drug-exposure time. Extended-dosing studies with CC486 have shown it to be well tolerated, with a toxicity profile comparable to Aza.²⁶ Patients who fail therapy with Aza respond well with extended dosing of CC486, indicating involvement of contrasting demethylation patterns due to prolonged exposure.²⁷ Efficacy of CC486 as maintenance therapy post-HSCT (NCT01835587) and after induction/consolidation for AML/MDS patients is under investigation.

Guadecitabine is a next-generation decitabine that is resistant to deamination by cytidine deaminase, thereby increasing its half-life and leading to prolonged exposure during the S-phase and enhanced cell kill.²⁸ Patients ineligible for intensive therapy who are treatment-naïve responded well to guadecitabine in early-phase trials, with >50% of patients achieving composite complete remission (CR).²⁹ A large multicenter phase III trial with guadecitabine or treatment of choice by the physician is under way for previously treated AML (NCT02920008).

Isocitrate dehydrogenase inhibitors

IDH catalyzes the oxidative decarboxylation of isocitrate to α -ketoglutarate while converting NAD(P)⁺ to NAD(P)H and maintaining cellular redox homeostasis.³⁰ Mutations in *IDH1* and *IDH2* are seen in approximately 6%–10% and 9%–13% of patients with AML.³¹ Mutant *IDH* starts catalyzing the α -ketoglutarate (α KG) to the oncometabolite D₂-hydroxyglutarate (D2HG). This in turn acts as a competitive antagonist for α KG and downregulates the family of α KG-dependent dioxygenases, in particular histone lysine demethylases and the Ten eleven translocation (TET) family of DNA hydroxylases, both in vitro and in vivo.^{32,33} There is a resultant elevation in histone methylation and 5-methylcytosine, leading to global

hypermethylation with lack of global gene expression, causing maturation arrest.³⁴ *IDH2* mutations produce high amounts of D2HG with increased cellular expression compared to *IDH1*-mutated cells. Studies have shown that *IDH1*-mutated cells depend on the wild-type (WT) *IDH1* allele for enhanced D2HG production.³⁵

Enasidenib (previously known as AG221) has been shown to selectively target the *IDH2* mutation. In a recent study conducted on relapsed refractory AML (RR-AML) patients with *IDH2* mutations treated with enasidenib monotherapy, the overall response rate was 40%, median response duration 5.8 months and median OS 9.3 months. Among the 34 patients who achieved CR with enasidenib, the median OS was 19.7 months.³⁶ Inhibitors of *IDH* have been shown to produce synergy with Aza, as both drugs reduce DNA methylation and evade blockade in cell differentiation.³⁷ Ivosidenib (AG120) was approved in 2018 for RR-AML with *IDH1* mutation. Ivosidenib efficacy was tested in a multicenter trial with 179 RR-AML patients. The overall response rates was 41.6%, with 21.6% achieving CR.³⁸ Treatment using enasidenib and ivosidenib induces differentiation syndrome, owing to the drugs' mechanisms of action.³⁹

FLT3 inhibitors

Approximately a third of adult patients with AML carry mutations in the FMS like tyrosine kinase 3 (FLT3) receptor.⁴⁰ *FLT3* mutation leads to aberrant downstream proliferation signals independently of ligand activation. Internal tandem duplication (ITD) mutations in the *FLT3* gene are seen in about 22% of AML patients, and is one of the driver mutations that presents with high leukemic burden and confers a poor prognosis (high incidence of relapse). The remaining 8% of mutations occur in the tyrosine-kinase domain (TKD), with unknown prognosis.^{41,42} The *FLT3*-WT vs *FLT3*-ITD allelic ratio is also an important prognostic factor. According to European LeukemiaNet guidelines, patients with an allelic ratio <0.5 or low ITD with an Nucleophosmin (*NPM1*) mutations are classified under “good prognosis”, while patients with high allelic ratio (>0.5 or high ITD) and *NPM1*-WT are grouped under the adverse-risk category.⁴³

Given the adverse clinical outcomes due to the *FLT3*-ITD mutation in AML patients, induction chemotherapy followed by HSCT is warranted, as it lowers the incidence of relapse and improves survival rates. The incidence of relapse is higher in patients with ITD mutations undergoing allo-HSCT with lower OS compared to *FLT3*-WT

patients.^{44,45} This has vastly driven attempts to target the *FLT3* mutation with kinase inhibitors (KIs) and clinical trials for *FLT3* targeted therapy in combination with induction chemotherapy or as maintenance therapy following HSCT.

Midostaurin (Mido), a multi-KI, has been shown to inhibit *FLT3* ITD–receptor activity and showed synergistic effects with the standard chemotherapeutic drugs cytarabine and daunorubicin in preclinical studies.⁴⁶ A phase II trial using Mido as monotherapy for RR-AML/MDS patients with *FLT3* mutations showed reduction in peripheral blasts in 70% of patients.⁴⁷ Subsequently, a phase III randomized trial (RATIFY) that spanned 10 years⁴⁸ comparing Mido and placebo along with standard chemotherapy in AML patients <60 years of age harbouring *FLT3* mutations (ITD, TKD) was conducted. Mido treatment resulted in significant improvements in OS (HR 0.78, *P*=0.009) and event-free survival (HR 0.78, *P*=0.002) at 4 years, despite an insignificant difference in the rate of CR. OS was also higher for patients in the Mido arm who underwent HSCT in first remission⁴⁹ As a multi KI, Mido

may also have inhibitory action on the *FLT3*-WT receptor, as shown by significant blast reduction in *FLT3*-WT AML patients, based on which a phase III study on *FLT3*-WT AML patients with Mido has been initiated (NCT003512197).⁵⁰ In April 2017, Mido in combination with conventional chemotherapy was approved for treatment in newly diagnosed AML patients with *FLT3* mutations <60 years of age by the FDA.

Second-generation *FLT3* tyrosine-kinase inhibitors

The first generation tyrosine KIs (TKIs) sorafenib, lestaurtinib, and Mido are broad spectrum multi-KIs. The next-generation TKIs quizartinib, crenolanib, and gilteritinib are more selective and potent oral drugs that are currently in clinical trials. These drugs have reduced toxicity compared to the first-generation TKIs.⁵¹ There are encouraging clinical data for these second-generation TKIs, as shown in Tables 1 and 2.^{52–57} Resistance to TKIs has been a major roadblock in AML therapy,

Table 1 Select clinical studies on second-generation TKIs with induction therapy

	Study	De novo AML	Efficacy
Gilteritinib	Phase II dose escalation with 7+3	n=66 <i>FLT3</i> mutant and WT	27 of 30 (90%) of <i>FLT3</i> -mutant patients achieved composite CR
Crenolanib	Phase I with 7+3	n=29 <i>FLT3</i> mutant	21 of 29 (72%) patients achieved CR
Quizartinib	Phase I dose escalation with 7+3	n=19 <i>FLT3</i> mutant and WT	14 of 19 achieved composite CR

Abbreviations: TKIs, tyrosine-kinase inhibitors; AML, acute myeloid leukemia; CR, complete remission.

Table 2 Select clinical studies on second-generation TKIs as monotherapy

	Study	Relapsed/Refractory AML	Efficacy
Gilteritinib	Phase I–II dose-escalation and expansion monotherapy	n=252 <i>FLT3</i> mutated and WT	100 of 249 (40%) evaluable patients achieved composite CR
Crenolanib	Phase I dose-escalation monotherapy	n=69 <i>FLT3</i> mutated	Overall response rate (ORR) 50% (nine of 18) in <i>FLT3</i> inhibitor-naïve patients ORR of 31% (11 of 36) previously treated with TKIs
Quizartinib	Phase II monotherapy	n=333 <i>FLT3</i> mutated and WT Cohort 1 (>60 years) Cohort 2 (18–60 years)	Cohort 1: 63 of 112 (56%) <i>FLT3</i> -mutated and 16 of 44 (36%) <i>FLT3</i> -WT patients achieved composite CR Cohort 2: 62 of 133 (46%) <i>FLT3</i> -mutated and 12 of 40 (30%) <i>FLT3</i> -WT achieved composite CR.

Abbreviations: TKIs, tyrosine-kinase inhibitors; AML, acute myeloid leukemia; CR, complete remission.

making these TKIs less efficient as single agents. Resistance mechanisms are diverse, and one known mechanism is acquisition of *FLT3* point mutations in TKD during therapy that may mediate resistance.⁵¹ Elevated levels of FLT3 ligands in the bone-marrow microenvironment after induction therapy leads to activation of *FLT3*-WT receptors that are sensitive to ligands but resistant to FLT3 inhibitors.⁵⁸ In addition, rewired signaling pathways and additional driver mutations in other genes also lead to TKI resistance. One strategy to overcome resistance is to employ combination therapy that targets different entities of the leukemic cell. FLT3 inhibitors in combination with the hypomethylating agent 5-Aza has been shown to alter methylation patterns in *FLT3*-mutated cells and overcome stromal protection.⁵⁹

Gemtuzumab ozogamicin

The CD33 antigen is a transmembrane receptor and myeloid-differentiation marker that is expressed in most AMLs. Its extracellular immunoglobulin-like V domain can be exploited using antibody–drug conjugate (anti-CD33-directed) therapy to target CD33-positive leukemic blasts.^{60,61} Gemtuzumab ozogamicin (GO) is an anti-CD33-directed antibody–drug conjugate comprised of a human mAb linked to calicheamicin. GO binds to the CD33 antigen present on the surface of leukemic blasts and myeloid-precursor cells and localizes into the cell.^{62,63} The complex breaks after localization and the released calicheamicin binds to DNA, resulting in DNA double-strand breaks and initiates apoptosis. GO was initially approved in 2000 as monotherapy (two doses of 9 mg/m², 14 days apart) for older patients (60 years) in first relapse with CD33-positive AML who were not eligible for chemotherapy.⁶⁴ In 2010, during the phase III SWOG S0106 study the drug was withdrawn from the market, due to increased mortality when used in combination with induction and consolidation therapy (single dose of 6 mg/m² on day 4).⁶⁵

Subsequent trials (ALFA-0701 phase III study, AML-19 phase III study, MyloFrance 1 and 2 studies) were conducted with lower doses of GO (3 mg/m² on days 1, 4, and 7, and 6 mg/m² on day 1 and 3 mg/m² on day 4) in patients with de novo AML. After a long break, GO was reapproved by the FDA in 2017 for treating CD33-positive newly diagnosed or RR-AML, with a black-box warning for hepatotoxicity and veno-occlusive disease.^{66,67} A subtype of AML known as core binding factor (CBF) AML responds well to

GO.⁶⁸ The biological significance of CBF-AML blast cells arises from matured committed leukemia-initiating cells with CD33 expression. Multi drug transporter 1 (MDR1) is known to extrude GO from leukemic cells and reduce its efficacy.⁶⁹ CBF-AML inherently has reduced expression of MDR1, allowing cells to retain GO and enhance cytotoxicity.^{70–72}

Targeting leukemic stem cells

Leukemic stem cells (LSCs) are believed to be responsible for disease relapse in AML, as they escape conventional chemotherapy.⁷⁰ Targeting LSCs in AML has been a paramount task because of the absence of a consistent surface marker that distinguishes HSCs from LSCs. Many studies have described multiple immunophenotypic differences that may distinguish LSCs from HSCs (CD25, CD44, CD96, CD93, and CD123), but a universal surface marker for LSCs is yet to be identified.⁷³ CD123 has been the only marker under constant investigation to target LSCs compared to the other surface markers. SL401 is the first CD123 monoclonal antibody fused with a catalytic unit of a diphtheria toxin introduced to target LSCs. In a phase I trial conducted on 74 patients with RR-AML or de novo AML, CR was observed in two patients, four patients showed 50% blast reduction in bone marrow, while 14 showed reduction in blast percentage from baseline.⁷⁴ Capillary-leak syndrome was one of the noted adverse events in this trial, but was manageable with early intervention.^{75,76} SL401 is now currently being investigated as remission-maintenance therapy for high-risk AML in CR1 or CR2 to improve relapse-free survival (phase II trial NCT02270463)

Flotetuzumab is a bispecific (CD123 × CD3) dual-affinity retargeting novel T cell–redirecting molecule.⁷⁷ A phase I dose-escalation study in RR-AML/MDS related AML patients demonstrated an overall response of 43% (six of 14 patients), while two patients showed consistent disease with only 20%–25% blast reduction. Consistent activation of CD4 and CD8 T cells was observed in the peripheral blood. Toxicity profiles mainly consisted of cytokine-release syndrome.⁷⁸ The study continues with cohort expansion (24 RR-AML and 24 MDS-AML [NCT02152956]).

Novel therapies

Mutant *TP53* activation

Tumour suppressor p53 (*TP53*) is one of the commonly mutated genes in malignancies. About 5%–10% of de

novo AML and 30% of t-AML harbor *TP53* mutations, conferring poor prognosis.⁷⁹ The mutation leads to misfolded formation of the transcription factor and voiding its ability to bind DNA. The prodrug APR246 spontaneously gets converted to its active compound methylene quinuclidinone at physiological pH. This compound then stabilizes the mutant protein by covalently binding to cysteine residues, leading to refolding and restoration of p53 function.⁸⁰ Promising initial results were observed from phase IB/II trials using APR246 in combination with Aza conducted on MDS/AML patients with *TP53* mutations (n=11). There was a 100% response rate by International Working Group criteria, and nine of eleven patients achieved CR.⁸¹

BCL2 inhibitor

B-cell lymphoma 2 (BCL2), an antiapoptotic protein highly expressed in AML cells, confers resistance to conventional therapy by increasing mitochondrial fitness and prevents apoptosis.⁸² BCL2 is also highly upregulated in the LSC compartment, aiding quiescence and disease relapse.⁸³ Venetoclax (Ven) is a BCL2 inhibitor that blocks its antiapoptotic activity, potentiates the effect of other partnered drugs, and enhances apoptosis. In a phase IB/II trial of Ven in combination with low-dose cytarabine in elderly AML patients unfit for intensive therapy, 62% of patients achieved CR, with median OS of 11.4 months.⁸⁴ Further promising results were observed combining Ven with Aza in elderly AML patients unfit for therapy, with CR 61% and median OS 17.5 months in early-phase trials.^{85,86} Phase III randomized trials are ongoing comparing the combination of Aza and Ven versus Aza monotherapy (NCT02993523).

Immune checkpoint inhibitors

Allogenic transplant remains the superior therapy for AML, mainly due to the graft vs. leukemia effect produced by the donor cells.^{87,88} AML cells express high levels of Programmed death ligand 1 (PDL-1), an inhibitory molecule that suppresses the cytotoxic activity of T cells, leading to immune exhaustion, and increases the chances of relapse.^{89,90} Immune modulation could effectively work in the AML context, rewiring the immune system. Early-phase trials with nivolumab (PD1 inhibitor) and Aza in RR-AML patients have demonstrated CR of 22% and median OS of 15 months in responders.⁹¹ Phase II trials combining nivolumab with conventional 7 +3 in newly diagnosed AML showed CR and CR with an

incomplete hematologic recovery rate of 72%, and nine patients were eligible for transplant.⁹² Studies evaluating the efficacy of pembrolizumab (PD1 inhibitor) with other agents are ongoing NCT02996474, NCT02845297. A list of other novel agents in early- and late-phase trials is presented in Table 3.

Standard chemotherapy to personalizing therapy: a reality?

Over the past few decades of treatment and trials, it has become clear that “one size does not fit all” in terms of treating AML. The best and first example would be acute promyelocytic leukemia. Until 1987, this was treated solely with intensive chemotherapy.⁹³ It was only after 1992 that the combination of all-*trans* retinoic acid and arsenic trioxide to induce apoptosis and differentiation created a benchmark in the treatment of acute promyelocytic leukemia.⁹⁴ Years later, the genomic landscape of AML helped us to understand that somatic mutations carry prognostic importance no less than recurrent cytogenetic abnormalities. Only after the discovery of somatic mutations did personalizing therapy for treating AML come into the limelight.⁹⁵

Though there are studies to substantiate the need and importance of novel therapies targeting different drivers of mutation, there is a setback when this is translated with genomic information alone. Even though genomic testing has been employed in a number of clinical trials on AML patients, it has been hardly used to stratify, due to extended turnaround times, which often delay the treatment.^{2,3,96} On the other hand, drug-sensitivity testing/ex-vivo high-throughput drug screening can be performed, as it is not time-consuming and a large variety of drugs can be screened within a short time.^{97–99} Clinical trials like the BEAT AML study aimed to integrate genomic profiling and drug-sensitivity testing in a large cohort of elderly AML patients, which seems promising for making personalization of the treatment regimen for each individual with the disease a reality. We believe that the combination of molecular profiling and ex-vivo drug screening would significantly improve the survival of patients with newly diagnosed and RR-AML (Table 4). Further clinical trials are recommended to substantiate the data obtained from both genomic profiling and ex-vivo high-throughput drug screening in patients. However, newer paradigms incorporating

Table 3 Novel agents in clinical trials

Category of drug target	Drug target	Drug	Trial phase	Remarks	Single agent/combination	Status	Reference
Cell surface receptors	CD123	SL401	I	Relapsed/refractory acute myeloid leukemia (AML) or high-risk myelodysplastic syndrome (MDS)	Combination with azacitidine	Recruiting	NCT03113643
		MGD006	I	Relapsed/refractory AML or intermediate-2/ MDS	Monotherapy	Recruiting	NCT02152956
	CD33	AMG330	I	Relapsed/refractory AML	Monotherapy	Recruiting	NCT02520427
		IMGN779	I	Relapsed/refractory CD33-positive AML	Monotherapy	Recruiting	NCT02674763
	CLEC12A	MCLA117	I	Adult AML patients (newly diagnosed/ relapsed/refractory)	Monotherapy	Recruiting	NCT03038230
	CTLA4	Ipilimumab	I	Relapsed or refractory MDS/AML	Combination with decitabine	Recruiting	NCT02890329
		Nivolumab	II I/II	AML in remission at high risk of relapse High-risk MDS and AML	Monotherapy Combination with idarubicin and cytarabine	Recruiting Recruiting	NCT02532231 NCT02464657
	PD1	Nivolumab	II	Refractory/relapsed or newly diagnosed AML	Combination with azacitidine with or without ipilimumab	Recruiting	NCT02397720
			I	AML and MDS	Combination with ipilimumab	Recruiting	NCT02846376
	Tyrosine-kinase pathways	FLT3	Pembrolizumab	II	Postremission treatment of patients aged ≥ 60 years with AML	Monotherapy	Recruiting
II				Relapsed/refractory AML	Monotherapy	Recruiting	NCT02768792
II			Relapsed/refractory AML patients and newly diagnosed older (≥ 65 years) AML patients	Azacitidine followed by pembrolizumab	Recruiting	NCT02845297	
I/II			Refractory/relapsed AML	Combination with 5-azacitidine/azacitidine	Recruiting	NCT02953561	
Tyrosine-kinase pathways	FLT3	Dasatinib	III	Newly diagnosed CBF-AML	Standard induction therapy and consolidation therapy with or without dasatinib	Recruiting	NCT02013648
		Quizartinib	I/II	Combination with azacitidine or cytarabine	Relapsed or refractory acute myeloid leukemia or myelodysplastic syndrome	Recruiting	NCT01892371
		Crenolanib	II	FLT3-positive AML	Monotherapy	Recruiting	NCT02400255

(Continued)

Table 3 (Continued).

Category of drug target	Drug target	Drug	Trial phase	Remarks	Single agent/combination	Status	Reference
Apoptotic targets	BCL2	Venetoclax	III	Patients with AML ineligible for standard induction therapy	Combination with azacitidine	Recruiting	NCT02993523
		Epigenetic targets	HDAC	SGI110	II	AML or MDS relapse following allogeneic hematopoietic stem-cell transplantation	SGI110 with donor-lymphocyte infusion (DLI)
	II			Elderly AML patients	Single-agent SGI110 and SGI110 with idarubicin combination regimen	Active, not recruiting	NCT02096055
Guadecitabine	III			Adults with previously treated AML	Guadecitabine (SGI110) versus treatment choice	Active, not recruiting	NCT02920008
NEDD8-activating enzyme (NAE)		Vorinostat	I	Relapsed/refractory AML	Using decitabine and vorinostat before and during chemotherapy with fludarabine, cytarabine and G-CSF (FLAG)	Recruiting	NCT03263936
		Valproic acid	II	High-risk AML or MDS	Combination with azacitidine	Recruiting	NCT02124174
		Entinostat (MS275)	I/II	Acute-leukemia relapse after allotransplantation	Combination with azacitidine, hydroxyurea, and eventually donor-leukocyte infusions	Recruiting	NCT01369368
		Panobinostat	II	Elderly patients with AML	Combination with 5AC	Recruiting	NCT01305499
			I/II	High-risk MDS or AML	Monotherapy	Active, not recruiting	NCT01451268
		Pracinostat	I	Relapsed/refractory AML	Combination with gemtuzumab-ozogamicin	Not yet recruiting	NCT03848754
			III	Newly diagnosed AML unfit for standard induction chemotherapy	Combination with azacitidine	Recruiting	NCT03151408
		Pevonedistat	I	AML and advanced MDS	Combination with low-dose cytarabine (LDAC)	Recruiting	NCT03459859
			II	Nonremission AML	Combination with azacitidine	Recruiting	NCT03709576

(Continued)

Table 3 (Continued).

Category of drug target	Drug target	Drug	Trial phase	Remarks	Single agent/combination	Status	Reference	
	DOT1L	Pinometostat	I/II	Newly diagnosed AML and MLL gene rearrangement	With standard chemotherapy	Recruiting	NCT03724084	
			I/II	Relapsed, refractory, or newly diagnosed AML with 11q23 rearrangement	Combination with azacitidine	Not yet recruiting	NCT03701295	
	IDH1/2	AG221	III	Late stage AML harboring an isocitrate dehydrogenase 2 mutation (IDHENTIFY)	AG221 versus conventional-care regimens (CCRs)	Recruiting	NCT02577406	
	Bromodomain	CPI0610	I/II	Previously treated acute leukemia, MDS, myelodysplastic/myeloproliferative neoplasms, and myelofibrosis	With and without ruxolitinib	Recruiting	NCT02158858	
		FT1101	I	AML/MDS or non-Hodgkin lymphoma (NHL)	Monotherapy	Recruiting	NCT02543879	
	CDK	Alvociclib	II	Relapsed/refractory AML with Noxa BH3 priming $\geq 40\%$ by mitochondrial profiling in bone marrow	Combination with induction therapy	Recruiting	NCT02520011	
		Palbociclib	I/II	MLL-rearranged acute leukemias AMLSG 23-14/Palbo-AL	Monotherapy	Recruiting	NCT02310243	

Table 4 Exvivo drug screening–based clinical trials

	Remarks	Status	Reference
High-Throughput Drug-Sensitivity Assay and Genomics-Guided Treatment of Patients with Relapsed or Refractory Acute Leukemia BEAT AML Core Study	Relapsed or refractory acute myeloid leukemia	Recruiting	NCT02551718
	Newly diagnosed or relapsed/refractory acute myeloid leukemia	Recruiting	NCT02927106

novel agents as monotherapy or in combination with standard care still pose a daunting task.

Acknowledgments

PB (IA/S/15/1/501842) and VM (IA/S/11/2500267) are supported by Wellcome DBT India Alliance Senior fellowships. This study was partly funded by a Centre of Excellence grant from the Department of Biotechnology India (BT/COE/34/SP13432/2015).

Disclosure

SD is supported by a junior research fellowship from the University Grants Commission (UGC), New Delhi, India. The authors report no other conflicts of interest in this work.

References

- Assi SA, Imperato MR, Coleman DJL, et al. Subtype-specific regulatory network rewiring in acute myeloid leukemia. *Nat Genet.* 2019;51(1):151–162. doi:10.1038/s41588-018-0270-1
- Ley TJ, Miller C, Ding L, et al. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. *N Engl J Med.* 2013;368(22):2059–2074. doi:10.1056/NEJMoa1301689.
- Papaemmanuil E, Gerstung M, Bullinger L, et al. Genomic classification and prognosis in acute myeloid leukemia. *N Engl J Med.* 2016;374(23):2209–2221. doi:10.1056/NEJMoa1516192
- Brinda B, Khan I, Parkin B, Konig H. The rocky road to personalized medicine in acute myeloid leukaemia. *J Cell Mol Med.* 2018;22(3):1411–1427. doi:10.1111/jcmm.13478
- De Kouchkovsky I, Abdul-Hay M. Acute myeloid leukemia: a comprehensive review and 2016 update. *Blood Cancer J.* 2016;6(7):e441. doi:10.1038/bcj.2016.50
- Rubnitz JE. Current management of childhood acute myeloid leukemia. *Paediatr Drugs.* 2017;19(1):1–10. doi:10.1007/s40272-016-0200-6
- Friedman AA, Letai A, Fisher DE, Flaherty KT. Precision medicine for cancer with next-generation functional diagnostics. *Nat Rev Cancer.* 2015;15(12):747–756. doi:10.1038/nrc4015
- Abraham A, Varatharajan S, Karathedath S, et al. RNA expression of genes involved in cytarabine metabolism and transport predicts cytarabine response in acute myeloid leukemia. *Pharmacogenomics.* 2015;16(8):877–890. doi:10.2217/pgs.15.44
- Abraham A, Devasia AJ, Varatharajan S, Karathedath S, Balasubramanian P, Mathews V. Effect of cytosine arabinoside metabolizing enzyme expression on drug toxicity in acute myeloid leukemia. *Ann Hematol.* 2015;94(5):883–885. doi:10.1007/s00277-014-2254-2

- Varatharajan S, Panetta JC, Abraham A, et al. Population pharmacokinetics of Daunorubicin in adult patients with acute myeloid leukemia. *Cancer Chemother Pharmacol.* 2016;78(5):1051–1058. doi:10.1007/s00280-016-3166-8
- Varatharajan S, Abraham A, Zhang W, et al. Carbonyl reductase 1 expression influences daunorubicin metabolism in acute myeloid leukemia. *Eur J Clin Pharmacol.* 2012;68(12):1577–1586. doi:10.1007/s00228-012-1291-9
- Elsayed AH, Cao X, Crews KR, et al. Comprehensive Ara-C SNP score predicts leukemic cell intracellular ara-CTP levels in pediatric acute myeloid leukemia patients. *Pharmacogenomics.* 2018;19(14):1101–1110. doi:10.2217/pgs-2018-0086
- Mitra AK, Crews KR, Pounds S, et al. Genetic variants in cytosolic 5'-nucleotidase II are associated with its expression and cytarabine sensitivity in HapMap cell lines and in patients with acute myeloid leukemia. *J Pharmacol Exp Ther.* 2011;339(1):9–23. doi:10.1124/jpet.111.182873
- Schneider C, Oellerich T, Baldauf H-M, et al. SAMHD1 is a biomarker for cytarabine response and a therapeutic target in acute myeloid leukemia. *Nat Med.* 2017;23(2):250–255. doi:10.1038/nm.4255
- Garzon R, Savona M, Baz R, et al. A phase 1 clinical trial of single-agent selinexor in acute myeloid leukemia. *Blood.* 2017;129(24):3165–3174. doi:10.1182/blood-2016-11-750158
- Martelli MP, Gionfriddo I, Mezzasoma F, et al. Arsenic trioxide and all-trans retinoic acid target NPM1 mutant oncoprotein levels and induce apoptosis in NPM1-mutated AML cells. *Blood.* 2015;125(22):3455–3465. doi:10.1182/blood-2014-11-611459
- Karathedath S, Rajamani BM, Aalam SMM, et al. Role of NF-E2 related factor 2 (Nrf2) on chemotherapy resistance in acute myeloid leukemia (AML) and the effect of pharmacological inhibition of Nrf2. *PLoS One.* 2017;12(5):e0177227. doi:10.1371/journal.pone.0177227
- Ganesan S, Alex AA, Chendamarai E, et al. Rationale and efficacy of proteasome inhibitor combined with arsenic trioxide in the treatment of acute promyelocytic leukemia. *Leukemia.* 2016;30(11):2169–2178. doi:10.1038/leu.2016.227
- Skrčić M, Sriskanthadevan S, Jhas B, et al. Inhibition of mitochondrial translation as a therapeutic strategy for human acute myeloid leukemia. *Cancer Cell.* 2011;20(5):674–688. doi:10.1016/j.ccr.2011.10.015
- Tardi P, Johnstone S, Harasym N, et al. In vivo maintenance of synergistic cytarabine: daunorubicin ratios greatly enhances therapeutic efficacy. *Leuk Res.* 2009;33(1):129–139. doi:10.1016/j.leukres.2008.06.028
- Lim W-S, Tardi PG, Dos Santos N, et al. Leukemia-selective uptake and cytotoxicity of CPX-351, a synergistic fixed-ratio cytarabine: daunorubicin formulation, in bone marrow xenografts. *Leuk Res.* 2010;34(9):1214–1223. doi:10.1016/j.leukres.2010.01.015
- Mayer LD, Harasym TO, Tardi PG, et al. Ratiometric dosing of anticancer drug combinations: controlling drug ratios after systemic administration regulates therapeutic activity in tumor-bearing mice. *Mol Cancer Ther.* 2006;5(7):1854–1863. doi:10.1158/1535-7163.MCT-06-0118

23. Lancet JE, Uy GL, Cortes JE, et al. CPX-351 (cytarabine and daunorubicin) liposome for injection versus conventional cytarabine plus daunorubicin in older patients with newly diagnosed secondary acute myeloid leukemia. *J Clin Oncol*. 2018;36(26):2684–2692. doi:10.1200/JCO.2017.77.6112
24. Glover AB, Leyland-Jones B. Biochemistry of azacitidine: a review. *Cancer Treat Rep*. 1987;71(10):959–964.
25. Silverman LR, McKenzie DR, Peterson BL, et al. Further analysis of trials with azacitidine in patients with myelodysplastic syndrome: studies 8421, 8921, and 9221 by the cancer and leukemia group B. *J Clin Oncol*. 2006;24(24):3895–3903. doi:10.1200/JCO.2005.05.4346
26. Savona MR, Gore SD, Kolibaba KS, et al. CC-486 (oral azacitidine) monotherapy in patients with acute myeloid leukemia (AML). *Blood*. 2015;126(23):452–452.
27. Savona MR, Kolibaba K, Conkling P, et al. Extended dosing with CC-486 (oral azacitidine) in patients with myeloid malignancies. *Am J Hematol*. 2018;93(10):1199–1206. doi:10.1002/ajh.25216
28. Roosendaal J, Wang K, Rosing H, et al. Development and validation of LC-MS/MS methods for the quantification of the novel anticancer agent guadecitabine and its active metabolite β -decitabine in human plasma, whole blood and urine. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2019;1109:132–141. doi:10.1016/j.jchromb.2019.01.011
29. Kantarjian HM, Roboz GJ, Kropf PL, et al. Guadecitabine (SGI-110) in treatment-naive patients with acute myeloid leukaemia: phase 2 results from a multicentre, randomised, phase 1/2 trial. *Lancet Oncol*. 2017;18(10):1317–1326. doi:10.1016/S1470-2045(17)30576-4
30. Lu C, Ward PS, Kapoor GS, et al. IDH mutation impairs histone demethylation and results in a block to cell differentiation. *Nature*. 2012;483(7390):474–478. doi:10.1038/nature10860
31. Medeiros BC, Fathi AT, DiNardo CD, Pollyea DA, Chan SM, Swords R. Isocitrate dehydrogenase mutations in myeloid malignancies. *Leukemia*. 2017;31(2):272–281. doi:10.1038/leu.2016.275
32. Figueroa ME, Abdel-Wahab O, Lu C, et al. Leukemic IDH1 and IDH2 mutations result in a hypermethylation phenotype, disrupt TET2 function, and impair hematopoietic differentiation. *Cancer Cell*. 2010;18(6):553–567. doi:10.1016/j.ccr.2010.11.015
33. Xu W, Yang H, Liu Y, et al. Oncometabolite 2-hydroxyglutarate is a competitive inhibitor of α -ketoglutarate-dependent dioxygenases. *Cancer Cell*. 2011;19(1):17–30. doi:10.1016/j.ccr.2010.12.014
34. Ward PS, Patel J, Wise DR, et al. The common feature of leukemia-associated IDH1 and IDH2 mutations is a neomorphic enzyme activity converting α -ketoglutarate to 2-hydroxyglutarate. *Cancer Cell*. 2010;17(3):225–234. doi:10.1016/j.ccr.2010.01.020
35. Ward PS, Lu C, Cross JR, et al. The potential for isocitrate dehydrogenase mutations to produce 2-hydroxyglutarate depends on allele specificity and subcellular compartmentalization. *J Biol Chem*. 2013;288(6):3804–3815. doi:10.1074/jbc.M112.435495
36. Stein EM, DiNardo CD, Pollyea DA, et al. Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. *Blood*. 2017;130(6):722–731. doi:10.1182/blood-2017-04-779405
37. DiNardo CD, Stein AS, Fathi AT, et al. Mutant isocitrate dehydrogenase (mIDH) inhibitors, enasidenib or ivosidenib, in combination with azacitidine (AZA): preliminary results of a Phase 1b/2 study in patients with newly diagnosed acute myeloid leukemia (AML). *Blood*. 2017;130(Suppl 1):639–639.
38. DiNardo CD, Stein EM, de Botton S, et al. Durable Remissions with Ivosidenib in IDH1-Mutated Relapsed or Refractory AML. *N Engl J Med*. 2018. doi:10.1056/NEJMoa1716984
39. Fathi AT, DiNardo CD, Kline I, et al. Differentiation syndrome associated with enasidenib, a selective inhibitor of mutant isocitrate dehydrogenase 2: analysis of a Phase 1/2 study. *JAMA Oncol*. 2018;4(8):1106–1110. doi:10.1001/jamaoncol.2017.4695
40. Kelly LM, Liu Q, Kutok JL, Williams IR, Boulton CL, Gilliland DG. FLT3 internal tandem duplication mutations associated with human acute myeloid leukemias induce myeloproliferative disease in a murine bone marrow transplant model. *Blood*. 2002;99(1):310–318. doi:10.1182/blood.V99.1.310
41. Yanada M, Matsuo K, Suzuki T, Kiyoi H, Naoe T. Prognostic significance of FLT3 internal tandem duplication and tyrosine kinase domain mutations for acute myeloid leukemia: a meta-analysis. *Leukemia*. 2005;19(8):1345–1349. doi:10.1038/sj.leu.2403838
42. Thiede C, Steudel C, Mohr B, et al. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. *Blood*. 2002;99(12):4326–4335. doi:10.1182/blood.v99.12.4326
43. Döhner H, Estey E, Grimwade D, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood*. 2017;129(4):424–447. doi:10.1182/blood-2016-08-733196
44. Brunet S, Labopin M, Esteve J, et al. Impact of FLT3 internal tandem duplication on the outcome of related and unrelated hematopoietic transplantation for adult acute myeloid leukemia in first remission: a retrospective analysis. *J Clin Oncol*. 2012;30(7):735–741. doi:10.1200/JCO.2011.36.9868
45. Deol A, Sengsayadeth S, Ahn KW, et al. Does FLT3 mutation impact survival after hematopoietic stem cell transplantation for acute myeloid leukemia? A Center for International Blood and Marrow Transplant Research (CIBMTR) analysis. *Cancer*. 2016;122(19):3005–3014. doi:10.1002/cncr.30140
46. Weisberg E, Boulton C, Kelly LM, et al. Inhibition of mutant FLT3 receptors in leukemia cells by the small molecule tyrosine kinase inhibitor PKC412. *Cancer Cell*. 2002;1(5):433–443.
47. Stone RM, DeAngelo DJ, Klimek V, et al. Patients with acute myeloid leukemia and an activating mutation in FLT3 respond to a small-molecule FLT3 tyrosine kinase inhibitor, PKC412. *Blood*. 2005;105(1):54–60. doi:10.1182/blood-2004-03-0891
48. Stone RM, Manley PW, Larson RA, Capdeville R. Midostaurin: its odyssey from discovery to approval for treating acute myeloid leukemia and advanced systemic mastocytosis. *Blood Adv*. 2018;2(4):444–453. doi:10.1182/bloodadvances.2017011080
49. Stone RM, Mandrekar SJ, Sanford BL, et al. Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. *N Engl J Med*. 2017;377(5):454–464. doi:10.1056/NEJMoa1614359
50. Fischer T, Stone RM, DeAngelo DJ, et al. Phase IIB trial of oral midostaurin (PKC412), the FMS-like tyrosine kinase 3 receptor (FLT3) and multi-targeted kinase inhibitor, in patients with acute myeloid leukemia and high-risk myelodysplastic syndrome with either wild-type or mutated FLT3. *J Clin Oncol*. 2010;28(28):4339–4345. doi:10.1200/JCO.2010.28.9678
51. Staudt D, Murray HC, McLachlan T, et al. Targeting oncogenic signaling in mutant FLT3 acute myeloid leukemia: the path to least resistance. *Int J Mol Sci*. 2018;19:10. doi:10.3390/ijms19103198
52. Pratz KW, Cherry M, Altman JK, et al. Updated results from a Phase 1 study of gilteritinib in combination with induction and consolidation chemotherapy in subjects with newly diagnosed acute myeloid leukemia (AML). *Blood*. 2018;132(Suppl1):564–564. doi:10.1182/blood-2018-99-110975
53. Wang ES, Tallman MS, Stone RM, et al. Low relapse rate in younger patients \leq 60 years old with newly diagnosed FLT3-mutated acute myeloid leukemia (AML) treated with crenolanib and cytarabine/anthracycline chemotherapy. *Blood*. 2017;130(Suppl 1):566–566.
54. Altman JK, Foran JM, Pratz KW, Trone D, Cortes JE, Tallman MS. Phase 1 study of quizartinib in combination with induction and consolidation chemotherapy in patients with newly diagnosed acute myeloid leukemia. *Am J Hematol*. 2018;93(2):213–221. doi:10.1002/ajh.24974

55. Perl AE, Altman JK, Cortes J, et al. Selective inhibition of FLT3 by gilteritinib in relapsed or refractory acute myeloid leukaemia: a multicentre, first-in-human, open-label, phase 1-2 study. *Lancet Oncol*. 2017;18(8):1061–1075. doi:10.1016/S1470-2045(17)30416-3
56. Cortes JE, Kantarjian HM, Kadia TM, et al. Crenolanib besylate, a type I pan-FLT3 inhibitor, to demonstrate clinical activity in multiply relapsed FLT3-ITD and D835 AML. *J Clin Oncol*. 2016;34(15_suppl):7008. doi:10.1200/JCO.2016.34.15_suppl.7008
57. Cortes J, Perl AE, Döhner H, et al. Quizartinib, an FLT3 inhibitor, as monotherapy in patients with relapsed or refractory acute myeloid leukaemia: an open-label, multicentre, single-arm, phase 2 trial. *Lancet Oncol*. 2018;19(7):889–903. doi:10.1016/S1470-2045(18)30240-7
58. Sato T, Yang X, Knapper S, et al. FLT3 ligand impedes the efficacy of FLT3 inhibitors in vitro and in vivo. *Blood*. 2011;117(12):3286–3293. doi:10.1182/blood-2010-01-266742
59. Al-Jamal HAN, Mat Jusoh SA, Hassan R, Johan MF. Enhancing SHP-1 expression with 5-azacytidine may inhibit STAT3 activation and confer sensitivity in lestaurtinib (CEP-701)-resistant FLT3-ITD positive acute myeloid leukemia. *BMC Cancer*. 2015;15:869. doi:10.1186/s12885-015-1695-x
60. Godwin CD, Gale RP, Walter RB. Gemtuzumab ozogamicin in acute myeloid leukemia. *Leukemia*. 2017;31(9):1855–1868. doi:10.1038/leu.2017.187
61. DeStefano CB, Hourigan CS. Personalizing initial therapy in acute myeloid leukemia: incorporating novel agents into clinical practice. *Ther Adv Hematol*. 2018;9(5):109–121. doi:10.1177/2040620718761778
62. van der Velden VHJ, Te Marvelde JG, Hoogeveen PG, et al. Targeting of the CD33-calicheamicin immunoconjugate Mylotarg (CMA-676) in acute myeloid leukemia: in vivo and in vitro saturation and internalization by leukemic and normal myeloid cells. *Blood*. 2001;97(10):3197–3204. doi:10.1182/blood.V97.10.3197
63. Bross PF, Beitz J, Chen G, et al. Approval summary: gemtuzumab ozogamicin in relapsed acute myeloid leukemia. *Clin Cancer Res*. 2001;7(6):1490–1496.
64. Larson RA, Boogaerts M, Estey E, et al. Antibody-targeted chemotherapy of older patients with acute myeloid leukemia in first relapse using Mylotarg (gemtuzumab ozogamicin). *Leukemia*. 2002;16(9):1627–1636. doi:10.1038/sj.leu.2402677
65. Sperr WR, Florian S, Hauswirth AW, Valent P. CD 33 as a target of therapy in acute myeloid leukemia: current status and future perspectives. *Leuk Lymphoma*. 2005;46(8):1115–1120. doi:10.1080/10428190500126075
66. Jen EY, Ko C-W, Lee JE, et al. FDA approval: gemtuzumab ozogamicin for the treatment of adults with newly diagnosed CD33-positive acute myeloid leukemia. *Clin Cancer Res*. 2018;24(14):3242–3246. doi:10.1158/1078-0432.CCR-17-3179
67. Raj RV, Abedin SM, Atallah E. Incorporating newer agents in the treatment of acute myeloid leukemia. *Leuk Res*. 2018;74:113–120. doi:10.1016/j.leukres.2018.10.008
68. Gottardi M, Mosna F, de Angeli S, et al. Clinical and experimental efficacy of gemtuzumab ozogamicin in core binding factor acute myeloid leukemia. *Hematol Rep*. 2017;9(3). doi:10.4081/hr.2017.7028
69. Linenberger ML, Hong T, Flowers D, et al. Multidrug-resistance phenotype and clinical responses to gemtuzumab ozogamicin. *Blood*. 2001;98(4):988–994. doi:10.1182/blood.v98.4.988
70. Thomas D, Majeti R. Biology and relevance of human acute myeloid leukemia stem cells. *Blood*. 2017;129(12):1577–1585. doi:10.1182/blood-2016-10-696054
71. Del Poeta G, Venditti A, Aronica G, et al. P-glycoprotein expression in de novo acute myeloid leukemia. *Leuk Lymphoma*. 1997;27(3–4):257–274. doi:10.3109/10428199709059682
72. Lutterbach B, Sun D, Schuetz J, Hiebert SW. The MYND motif is required for repression of basal transcription from the multidrug resistance 1 promoter by the t(8;21) fusion protein. *Mol Cell Biol*. 1998;18(6):3604–3611. doi:10.1128/mcb.18.6.3604
73. Pollyea DA, Jordan CT. Therapeutic targeting of acute myeloid leukemia stem cells. *Blood*. 2017;129(12):1627–1635. doi:10.1182/blood-2016-10-696039
74. Assi R, Kantarjian H, Ravandi F, Daver N. Immune therapies in acute myeloid leukemia: a focus on monoclonal antibodies and immune checkpoint inhibitors. *Curr Opin Hematol*. 2018;25(2):136–145. doi:10.1097/MOH.0000000000000401
75. Yalcintepe L, Frankel AE, Hogge DE. Expression of interleukin-3 receptor subunits on defined subpopulations of acute myeloid leukemia blasts predicts the cytotoxicity of diphtheria toxin interleukin-3 fusion protein against malignant progenitors that engraft in immunodeficient mice. *Blood*. 2006;108(10):3530–3537. doi:10.1182/blood-2006-04-013813
76. Sweet KL, Pemmaraju N, Lane AA, et al. Lead-in stage results of a pivotal trial of SL-401, an interleukin-3 receptor (IL-3R) targeting biologic, in patients with blastic plasmacytoid dendritic cell neoplasm (BPDCN) or acute myeloid leukemia (AML). *Blood*. 2015;126(23):3795–3795.
77. Campagne O, Delmas A, Fouliard S, et al. Integrated pharmacokinetic/pharmacodynamic model of a bispecific CD3xCD123 DART molecule in nonhuman primates: evaluation of activity and impact of immunogenicity. *Clin Cancer Res*. 2018;24(11):2631–2641. doi:10.1158/1078-0432.CCR-17-2265
78. Uy GL, Godwin J, Rettig MP, et al. Preliminary results of a Phase 1 study of flotetuzumab, a CD123 x CD3 Bispecific Dart® protein, in patients with relapsed/refractory acute myeloid leukemia and myelodysplastic syndrome. *Blood*. 2017;130(Suppl 1):637–637.
79. Ok CY, Patel KP, Garcia-Manero G, et al. TP53 mutation characteristics in therapy-related myelodysplastic syndromes and acute myeloid leukemia is similar to de novo diseases. *J Hematol Oncol*. 2015;8:45. doi:10.1186/s13045-015-0139-z
80. Lambert JMR, Gorzov P, Veprintsev DB, et al. PRIMA-1 reactivates mutant p53 by covalent binding to the core domain. *Cancer Cell*. 2009;15(5):376–388. doi:10.1016/j.ccr.2009.03.003
81. Sallman DA, DeZern A, Sweet K, et al. Abstract CT068: Phase Ib/II combination study of APR-246 and azacitidine (AZA) in patients with TP53 mutant myelodysplastic syndromes (MDS) and acute myeloid leukemia (AML). *Cancer Res*. 2018;78(13 Supplement):CT068–CT068. doi:10.1158/1538-7445.AM2018-CT068
82. Anstee NS, Bilardi RA, Ng AP, et al. Impact of elevated anti-apoptotic MCL-1 and BCL-2 on the development and treatment of MLL-AF9 AML in mice. *Cell Death Differ*. 2018;1. doi:10.1038/s41418-018-0209-1
83. Lagadinou ED, Sach A, Callahan K, et al. BCL-2 inhibition targets oxidative phosphorylation and selectively eradicates quiescent human leukemia stem cells. *Cell Stem Cell*. 2013;12(3):329–341. doi:10.1016/j.stem.2012.12.013
84. Thomas XG, Dmoszynska A, Wierzbowska A, et al. Results from a randomized phase III trial of decitabine versus supportive care or low-dose cytarabine for the treatment of older patients with newly diagnosed AML. *J Clin Oncol*. 2011;29(15_suppl):6504–6504. doi:10.1200/jco.2011.29.15_suppl.6504
85. DiNardo CD, Pratz K, Pullarkat V, et al. Venetoclax combined with decitabine or azacitidine in treatment-naïve, elderly patients with acute myeloid leukemia. *Blood*. 2019;133(1):7–17. doi:10.1182/blood-2018-08-868752
86. Pollyea DA, Stevens BM, Jones CL, et al. Venetoclax with azacitidine disrupts energy metabolism and targets leukemia stem cells in patients with acute myeloid leukemia. *Nat Med*. 2018;24(12):1859–1866. doi:10.1038/s41591-018-0233-1
87. Ganapule A, Nemani S, Korula A, et al. Allogeneic stem cell transplant for acute myeloid leukemia: evolution of an effective strategy in India. *J Glob Oncol*. 2017;3(6):773–781. doi:10.1200/JGO.2016.06650
88. Lipof JJ, Loh KP, O'Dwyer K, Liesveld JL. Allogeneic hematopoietic cell transplantation for older adults with acute myeloid leukemia. *Cancers*. 2018;10(6):179. doi:10.3390/cancers10060179

89. Brodská B, Fuchs O, Otevřelová P, Salek C, Kuželová K. PD-L1 Is frequently expressed in acute myeloid leukemia patients with leukocytosis. *Blood*. 2016;128(22):5229–5229.
90. Haroun F, Solola SA, Nassereddine S, Tabbara I. PD-1 signaling and inhibition in AML and MDS. *Ann Hematol*. 2017;96(9):1441–1448. doi:10.1007/s00277-017-3051-5
91. Daver N, Garcia-Manero G, Basu S, et al. Nivolumab (Nivo) with azacytidine (AZA) in patients (pts) with relapsed acute myeloid leukemia (AML) or frontline elderly AML. *Blood*. 2017;130(Suppl 1):1345–1345.
92. Ravandi F, Daver N, Garcia-Manero G, et al. Phase 2 study of combination of cytarabine, idarubicin, and nivolumab for initial therapy of patients with newly diagnosed acute myeloid leukemia. *Blood*. 2017;130(Suppl 1):815–815.
93. Parmar S, Tallman MS. Acute promyelocytic leukaemia: a review. *Expert Opin Pharmacother*. 2003;4(8):1379–1392. doi:10.1517/14656566.4.8.1379
94. Lo-Coco F, Avvisati G, Vignetti M, et al. Retinoic acid and arsenic trioxide for acute promyelocytic leukemia. *N Engl J Med*. 2013;369(2):111–121. doi:10.1056/NEJMoa1300874
95. Welch JS, Ley TJ, Link DC, et al. The origin and evolution of mutations in acute myeloid leukemia. *Cell*. 2012;150(2):264–278. doi:10.1016/j.cell.2012.06.023
96. Metzeler KH, Herold T, Rothenberg-Thurley M, et al. Spectrum and prognostic relevance of driver gene mutations in acute myeloid leukemia. *Blood*. 2016;128(5):686–698. doi:10.1182/blood-2016-01-693879
97. Eriksson A, Österroos A, Hassan S, et al. Drug screen in patient cells suggests quinacrine to be repositioned for treatment of acute myeloid leukemia. *Blood Cancer J*. 2015;5(4):e307. doi:10.1038/bcj.2015.31
98. Lee S-I, Celik S, Logsdon BA, et al. A machine learning approach to integrate big data for precision medicine in acute myeloid leukemia. *Nat Commun*. 2018;9(1):42. doi:10.1038/s41467-017-02465-5
99. Azzam D, Volmar C-H, Hassan -A-A, et al. A patient-specific ex vivo screening platform for personalized acute myeloid leukemia (AML) therapy. *Blood*. 2015;126(23):1352–1352.

Pharmacogenomics and Personalized Medicine

Dovepress

Publish your work in this journal

Pharmacogenomics and Personalized Medicine is an international, peer-reviewed, open access journal characterizing the influence of genotype on pharmacology leading to the development of personalized treatment programs and individualized drug selection for improved safety, efficacy and sustainability. This journal is indexed

on the American Chemical Society's Chemical Abstracts Service (CAS). The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/pharmacogenomics-and-personalized-medicine-journal>