

RESEARCH ARTICLE

Vaccinia and other viruses with available vaccines show marked homology with the HIV-1 envelope glycoprotein: The prospect of using existing vaccines to stem the AIDS pandemic

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Abstract

Cross-reactive immunity occurs when infection with or vaccination against one virus protects against another related family member. A search for homologues of the HIV-1 envelope glycoprotein revealed that it is composed of thousands of intercalating and overlapping viral matches of pentapeptide or longer gapped consensi, belonging to over 70% of the currently sequenced virome, infecting all kingdoms from bacteria to man. It was also highly homologous to proteins from the Visna/Maedi and other ovine viruses, while other proteins (nef/tat/gag/pol) were homologous to proteins from the equine infectious anaemia virus and HTLV-2/HTLV-3 viruses. This phenomenon suggests that horizontal gene transfer from coinfecting RNA and DNA viruses to retroviruses is extensive, providing a route for the subsequent insertion of non-retroviral genes into human and other genomes via retroviral integration. This homology includes all viruses for which vaccines already exist. Cross-reactive immunity may be operative in AIDS, as Vaccinia vaccination decreases viral replication in HIV-1 infected patients' cells, for the CCR5 tropic form. Measles, Dengue virus, or GB virus C infections also decrease the HIV-1 viral load. A resumption of Vaccinia/smallpox vaccination might be expected to have a significant effect on the AIDS pandemic, and a careful study of the potential uses of other existing viral and bacterial vaccines merits close attention. This phenomenon may also be relevant to other recalcitrant viruses, bacteria, and parasites for which no vaccine exists and the armory of existing vaccines may have a role to play in diseases other than those for which they were designed.

Keywords: AIDS, HIV-1, smallpox, vaccine, vaccinia

Introduction

The birth of immunology, over 200 years ago, noted that smallpox could be prevented by inoculation with cowpox,⁽¹⁾ a principle of immunity leading to the development of vaccines that have eliminated smallpox⁽²⁾ and which combat many other viral and bacterial diseases. Many viruses are however, recalcitrant to vaccination, particularly the AIDS virus, HIV-1.⁽³⁾ However it has recently been shown that Vaccinia virus vaccination reduces CCR5 tropic HIV-1 replication of the cells of infected patients.⁽⁴⁾ In HIV-1 infected patients the viral load has also been reported to be reduced in patients infected with measles or Dengue fever.^(5,6) The suppression of HIV-1 replication by measles infection

is concurrent with intense immune activation.⁽⁷⁾ It has also been shown that GB virus type C infection prolongs the survival of HIV-1 infected patients and that this effect is related to antibodies raised to the GB virus envelope protein, that cross-react with HIV-1 particles.⁽⁸⁾ This latter effect suggests cross-reactive immunity. These apparent protective effects of other viral infections could also be related to a general activation of defense networks such as the protein kinase R or retinoic acid inducible gene (RIG-1) pathways leading to interferon production and the activation of antiviral signaling programs, although some viruses, including herpes simplex and influenza are able to subvert these and other pathways.^(9,10)

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If cross-reactive immunity is also involved in such effects, one would expect a degree of homology between HIV-1 and other viral proteins, within antigenic regions. In an attempt to find homologous viruses that might serve as the cowpox equivalent to HIV-1, the HIV-1 envelope glycoprotein (*env*) was compared to all other viral proteomes. Short contiguous amino acid stretches (pentapeptides or longer gapped sequences) belonging to proteins from almost the entire current virome are encased within the *env* protein and include those for which vaccines are available. These could perhaps play a role in the development of cross-reactive immunity to HIV-1.

Methods

B-cell epitopes for the HIV-1 *env* glycoprotein (P04578: Human immunodeficiency virus type 1 group M subtype B (isolate HXB2)) were retrieved from the BepiPred server⁽¹¹⁾ (<http://www.cbs.dtu.dk/services/BepiPred/>) and examples of immunogenic regions compared to all viral proteomes using the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) server (BLASTp). This *env* sequence was derived from the reference *env* gene in the NCBI gene database (NC_001802.1). To detect small intraprotein consensi, the *E* value was set to 100,000. HIV-1, HIV-2, and other immunodeficiency viruses (Bovine, Feline, and Simian, the HIV-like cancer virus or the Aids-associated retrovirus, and the Murine AIDS virus-related provirus) were eliminated from the search due to evident homology. A list of available viral vaccines was obtained from the Center for disease control website at <http://www.cdc.gov/vaccines/vpd-vac/vaccines-list.htm>. BLASTs against these specific viruses (Table 1) were also undertaken. The *env* protein epitopes registered in the Immune epitope database (<http://www.immuneepitope.org>)⁽¹²⁾ were also compared to these viruses. In some cases, the amino acid sequences of these *env* protein epitopes differed from that of the chosen example, due to viral strain differences. Viral matches (vatches) of five contiguous amino acids or more or longer gapped sequences were identified by eye and copied to a table in the appropriate position relative to the HIV-1 *env* amino acid sequence (Supplementary Table 1). The entire protein was not processed, but the many results illustrated the principles involved. To the author's knowledge or ability, there is currently no way of automating this process (every single pentapeptide of the *env* glycoprotein tested shares similarity with several other viruses) and it is hoped that this illustration will stimulate work in this direction, which is also applicable to millions of vatches within the human proteome. Finally, the *env* protein from various HIV-1 strains was screened by BLAST analysis (BlastP) versus various Vaccinia viruses. The common peptides identified were analyzed for potential B cell immunogenicity using the BepiPred server. The server-set index of 0.35 was used as the immunogenicity index threshold.

Results

The *env* glycoprotein shows significant *overall* homology with proteins from four other viruses, the *env* proteins of the Caprine arthritis encephalitis virus ($E=3e-12$), the small ruminant lentivirus ($E=6e-10$), the visna/maedi virus ($E=6e-06$) and the ovine lentivirus ($E=4e-04$) (Figure 1). The HIV-1 *nef* protein showed significant overall homology with an ORF protein from HTLV-2 ($E=2e-45$), while the HIV-1 *tat* protein showed significant overall homology with the HTLV-3 *tat* protein ($E=1e-35$) (not shown). The HIV-1 *gag* protein is highly homologous to a protein from the puma lentivirus ($E=1E-98$) and to a *gag* protein from the equine infectious anaemia virus ($E=3e-42$) (Figure 1).

The results in relation to other viruses are shown in supplementary Table 1, where viral vatches are aligned with the *env* sequence, which is also characterized in relation to the B cell epitope index. Even though only 70% of the *env* protein was processed, HIV-1 vatches were observed in 1827 RNA and DNA viruses and phages, known to infect all kingdoms from bacteria to man. These were majoritarily species rather than strains. At the time of writing, there are 3753 reference sequences for 2565 viral genomes in the NCBI Entrez Genomes database, and the viruses containing HIV-1 *env* sequences account for 72% of the known current virome. Examples of such alignments, for viruses where vaccines are available, are shown in Table 1. All of these viruses contain HIV-1 vatches in both B cell epitope and non-epitope regions and within epitopes that have been experimentally verified.

A BLAST analysis of the *env* protein from several HIV-1 viral strains compared with Vaccinia viruses revealed a further layer of complexity. While certain identical Vaccinia/HIV-1 sequences were maintained across several HIV-1 viral strains, for example, the hexapeptides GAAGST or VVKIEP, these were often in differing positions of the *env* protein (e.g. GAAGST at positions 386, 510, 512, 524, 529, or 531). Otherwise, the profile of matching peptides derived from this sweep appears to be distinct for each strain of the HIV-1 virus. The viral matches shown in Tables 1 and 2 were predominantly pentapeptides, but longer contiguous or gapped sequences as well as frequent tetrapeptides were also observed (see supplementary Table 1).

Discussion

The close homology of the *env*, *nef*, *tat*, and *gag/pol* proteins with caprine, ovine, visna/Maedi, equine, and small ruminant viruses and particularly with HTLV-2 and HTLV-3 is of evolutionary interest as it suggests a source of the AIDS virus and its relatives, prior to simian integration and passage to man. However this is not the subject of this article.

In terms of cross-reactive immunity, no vaccines for HTLV-2 or HTLV-3 yet exist,⁽¹³⁾ although interestingly,

Table 1. Examples of viral vatches within the HIV-1 envelope protein, for viruses where vaccines are available.

Viruses and Alignments with the HIV-1 <i>env</i> protein All are within a predicted B cell epitope region (or within an experimentally described IEDB epitope)				
Chicken pox (Human Herpesvirus 3)	Hepatitis A	Hepatitis B	Influenza A virus (many different strains)	Japanese encephalitis virus
73: A +PTDP+	38: VYGV	39: Y VPV WK	32: DT+VHN	59: K YDTE
108: IIS W+	477: +NWRS+	74: CVPTD	42: VPVW	110: SLWD
121: KLTP LC TL	+KI	75: VPTDP P	108: I SLWDQ and IISLW	240: T++STV
142: SSSGR	492: EPLGV	108: IS W SL and IISL	110: SLWDQ	241: NVSTV
214: PIHY APA	575: QL VLA	141: NSSSG	121: K T PLCV L	306: RKRI+
252: RP V +LL	608: VP NAS	142: SSSGR	124: PLCV L	497: APTKA
276: NFT NA		214: PIH CA	125: LCVTL	526: AGST+G A S TL R
305: KR R IG		218: CAPA F	131: CTDLK	576: L ARV Y LK
307: KRIH and KR RI		237: GPCT+	139: NTNSS	688: IVGGL L I
313: P RAF+ P GRA		252: RP V QL	142: SSSGR	690: GGLV
413: TITLP		253: PIVST	167: GKVQK	742: RDRSI
500: KAKRRV		255: ST+LL	205: CPKVS	829: VIEVL R and VIE
502: KRRVV		293: B +INCT	207: KI FEP IP and KI SFE IP	VLQR
573: GI QLQ		307: KR H P GRA	214: P+HYC	830: IEVLQR
574: IK QLQA		314: GR AFYT	234: NGTGP	
690: GGLVG		359: QSS GD	252: R IVSTQ and RPIV Q	
		362: KQSSG	254: IVSTQ	
		441: GQ RCS S I	263: GSLAE+	
		494: LGVAP	294: INCTR	
		495: GVAPT	298: RPNNN	
		575: QL ARV	302: NNTRK and NYNK KRI I and NYNKR	
		583: VE YLKD ++L LG GC KL+C	303: YNK KR	
		584: E YLKD	304: TRKRI	
		607: AV WNA and AV WN S	305: KRIRI and KR R+ I PG and KRKR	
		678: W LW +I IF	312: GPGR F+ and GPG F+	
		685: FI++V	349: LREQF	
		690: GGL GL	350: REQFG	
		708: VRQ YS LS	356: NKTII	
		712: YSPLS	357: KTIIF	
		802: YW QEL	358: TIIFK	
		803: W QELK	362: KQSSG	
			364: SSGGD	
			369: PEIVT	
			370: EIVTH	
			371: IVTHS	
			372: VTHSF	
			373: THSFN	
			405: SNNTE	
			407: NTEGS	
			412: DTITL	
			445: CSSNI	
			490: KIEPL	
			493: PLGVA	
			571: VWGI AR	
			573: QL ARV	
			576: L+ARVL	
			605: TTAVP	
			608: +PW NASW	
			679: LWYI K F and LW IKI	
			685: FI+ IV GLV	
			686: IMIV	
			687: MIV G V L and MI+GG	
			710: QG YS LSFQ	
			711: GYS LSF	
			725: RGPDR	
			730: PEG+EE	
			744: RDRSI	
			805: +ELKN	
			806: ELKNS and ELK+ AV	
			807: LKNS V	
			825: G DRVI	
			828: RV E LQR and RV E+LQR	
			830: IEVLQ	

(Continued)

Table 1. Examples of viral vatches within the HIV-1 envelope protein, for viruses where vaccines are available. (Continued)

Viruses and Alignments with the HIV-1 <i>env</i> protein All are within a predicted B cell epitope region (or within an experimentally described IEDB epitope)				
Measles virus (repeat motifs in Bold)	Mumps virus	Papillomavirus(several strains)	Poliovirus (1,2 and 3)	Human Rotavirus A
55: ASDAKA	112: W DQSL	35: WV V YGV	36: V VYYG	63: TEVHN
109: ISL WD SL	135: N NT SSS	36: VTV PV	252: RP + TQ	121: KL LCV
110: SLWD	140: NT SSS	58: AYDT+	255: VSTQ	133: DLKND
252: RPI S QL	254: I STQL	60: AY+T HN+	314: GRA YT	208: ISF P +Y
253: PI S QL	305: KRI IG	70: ATHAC	336: AKW++ and AK NN	252: A R I VSTQ
263: GSLA EE	443: QI CS NI	74: CVPTD P P	529: TMGAA	303: YNKR
304: NKRK	493: PLGVA	77: TDPNP	531: GAAS+	307: KRIH and KR RI
308: RIH IGPG	573: IK QLQA	82: QE+VLV	836: AC I IP IRQG	308: A RIHI
312: GPGRA	577: QAR LA	142: SSSGR		313: PG AF+
314: GRAF T	688: IV GLV	208: ISF+P		337: KW +TL
401: STEGS	823:AEG RVI	218: CAPA F		369: PEIVT
493: PLGVA		234: NGTGP		413: TITLP
573: IKQL QA V		238: PCTNV		576: LQ V L VE YLK
581: LAVE LK		239: CT N STVQC		583: VER L D QLL I G and VE YLK and VE Y+K
647: E SQ+QQ+		240: TNVST		686: IM V GL V L
685: F+M LVGL		253: PI STQ		691: GLVG+
688: IVGG V		264: SLAEE		822: VAEGT+
706: NRVQR		300: NNNTR		829: VIEVL
707: RVRQG		301: NNTRK		831: EV LQRA
715: L F+ LPTPR		302: NTRKR		
716: NRVQR		305: KR RI I and KRKR+		
719: THLPT		349: LREQF		
721: LPTPR		363: QSSGG		
732: GI EE+G + R DRDR		364: SSGGD		
801: +Y SQEL		401: STEGS		
828: R EVVQ		406: NNTEG		
		407: NTEGS		
		410: GSDTI		
		413: TITLP		
		440: SGQIR		
		493: PLGVA		
		497: APTKA		
		500: KAKRR		
		502: KRRVV		
		570: VWGIK L+		
		574: KQLQ		
		576: LQ VLA		
		605: TTAVP		
		633: REIN Y S		
		635: I+NYTS		
		644: SLIEES		
		658: QELLE		
		688: IVGG		
		690: GG+VG		
		721: LPTP GP		
		730: PEG ++EGG		
		731: EG+EEE		
		734: EEEG E +R D S R		
		799: LL W QEL		
		806: ELKNS V		
		807: LKNSA		
		820: IAVAE D IE		
		828: R IEVL		
Rabies virus	Rubella	Vaccinia virus or Vaccinia virus	Yellow fever virus	
122: L LC+TL	70: AT ACV	Tian Tan	108: IIS DQ	
218: CAPA F	PTD	34: KW+TV	131: KC L D SSS	
220: PAG AI	738: GGE DR	35: LW YYGV	231: KTF GTG CT	
337: KWNN	825: G DRV+ V Q	36: VT+ Y GVPV	259: LLLN E+ SV T N T I + S E NC PN R	
354: GNNKT		37: T+YYG	276: N D KTI V L T P	
584: ER+LK		55: V LNAT IA	354: GNNKT	
687: MI GGL L		57: DAKAY	417: PCRI I+	
800: LQ WSQ		82: Q VVLV	484: YK KVVK+ L AP KA V+ R+ R G	
805: QELKN		108: II LW+	632: + EI NY S H	

(Continued)

Table 1. Examples of viral vatches within the HIV-1 envelope protein, for viruses where vaccines are available. (Continued)

Viruses and Alignments with the HIV-1 <i>env</i> protein All are within a predicted B cell epitope region (or within an experimentally described IEDB epitope)		
	112: W DQSL	687: M VGG V L
	122: LTPL V	691: GLVG
	143: SSGRM	826: TDR IE V+G
	153: EIKNC	842: H RIR GL+
	267: EEEVV	
	276: NFTD A	
	312: GPGR	
	313: A PG AF+	
	355: NNKTI	
	359: IIFKQ	
	360: IFKQS	
	524: GAAGST	
	577: QARV AV	
	579: RVL A+ R and RV AVE and RVL AV RY	
	587: L +QQ LL	
	635: I NYTS	
	645: LI EE +QE E Q L+E	
	646: +EE N+++ K EQELL	
	712: YSPLS	
	714: P SFQT	
	719: TH P T + PE I	
	720: HL TP GP	
	741: DR+R IR	
	804: SQELK	
	807: LK+SA+	
Viruses that modulate HIV-1 infection (for measles see above)		
Dengue virus (1, 2 or 3)	GB virus C	
36: VT Y+GV H and VT Y GV V WK	70: ATHAC D P+++ 122: LTPL CV	
108: IIS DQ	140: T SSSG EK	
208: I EPIP	209: SFE IP	
242: VSTVQ	213: IPI AG A	
252: RP V LL	252: RP+VS	
254: IVST LL	253: PIVS	
256: STQLL	306: RKR + PG	
264: SL EE+	468: FR GGG D W	
264: SLAEE	521: FLG T A LT L G V	
265: LAE EV	523: LGAAG TM A M	
302: NTRKR	531: GA S+T T QA	
302: NKRKR	574: K L ARVL	
313: PGR TT	581: LAVE LK	
335: RAK NT	582: AVE LK	
348: ESQ +QE	610: VPW AS	
363: QSSGG	686: IMI GL	
402: TEGSN	689: VG LVG and VG L GL	
523: LG GSTM	729: R G GER DR	
570: VWGI AR	738: GER IRLV	
580: VLAVE	807: LK VSLLNA	
606: TA PWN	828: RVIE	
685: F +VGG VG	830: IE QRA	
686: IM V GLV L	831: EVL RA	
691: VGGL		
722: PTPRG		
Position within env protein	Epitope from IEDB	Type
33	KLWVTVYYGV	MHC binding
36	VTVYYGVPVWK	T cell/MHC binding
108	IISLWDQSL	MHC binding
121	KLTPLCVTL	T cell/MHC binding
206	PKISFEPPIHYCAPAGFA	MHC binding

(Continued)

Table 1. Examples of viral matches within the HIV-1 envelope protein, for viruses where vaccines are available. (Continued)

Viruses and Alignments with the HIV-1 <i>env</i> protein All are within a predicted B cell epitope region (or within an experimentally described IEDB epitope)		
252	RPIVSTQLL	MHC binding
302	NYNKRKRIHIGPGRAFYTTKNII	B cell
311	IGPGRAFHT	T cell
312	GPGRAFYTT	MHC binding
335	RAKWNNTLK	MHC binding
570	VWGIKQLQARVLAVERYLKD	MHC binding
606	TAVPWNASW	MHC binding
678	WLWYIKIFI	MHC binding
685	FIMIVGGLV	MHC binding
686	IMIVGGLVGL	MHC binding
799	LLQYWSQEL	MHC binding
828	RVIEVLQRA	MHC binding

Their start position (within the *env* protein of 856 amino acids) is marked as is their position with respect to predicted B-cell epitopes within the *env* protein (these are all within regions with an antigenicity index of greater than the server-set threshold of 0.35: see supplementary Table 1). Spaces within the sequences indicate nonidentical amino acids and + signs an amino acid with similar physicochemical properties. The gray shaded sequences are within sequences that have been described as epitopes in experimental studies (B cell, T cell, or MHC binding from IEDB: The amino acid sequences of these experimentally verified epitopes are appended at the bottom of the table). Note that these sequences often overlap within consecutive regions of the *env* protein. In the majority of cases shown, contiguous sequences were of pentapeptides, although longer gapped sequences are also illustrated.

HTLV-2 infection appears to have a protective influence on HIV-1 infection.⁽¹⁴⁾ Should HTLV vaccines be developed, they may also have a role to play in relation to HIV-1.

As regards the shorter contiguous sequences and matches, the extensive homology of a single HIV-1 protein (*env*) with numerous phage and viral proteins (~72% of the currently sequenced virome) suggests that horizontal partial gene transfer from coinfecting DNA and RNA viruses to retrovirus, and/or vice versa, has proceeded on a massive scale during the evolutionary history of the AIDS virus and its ancestors. These include sequences from viruses infecting all kingdoms (e.g. bacteria, amoeba, fungi, plants, molluscs, insects, invertebrates, fish, birds, reptiles, and mammals) suggesting that these have at some time hosted the HIV-1 virus or its ancestors, along with other viruses, whose partial gene sequences have somehow been incorporated into the HIV-1 viral genome. There is no reason to suppose that this is not a feature of other retroviruses. As such sequences can subsequently be transferred to other genomes via retroviral insertion, this may partly explain the presence of phage and viral partial gene sequences within the genomes of plants, arthropods, fungi, nematodes, protozoa,⁽¹⁵⁾ mammals and man.⁽¹⁶⁻¹⁸⁾ The human proteome also contains multiple peptide consensi from bacterial, plant, and animal viruses.⁽¹⁹⁾

Horizontal gene transfer from virus to retrovirus does not appear to have been specifically studied in the laboratory. However, gene exchange is common between viruses,^(20,21) and also between retroviruses⁽²²⁾ where, for example, recombination can lead to the development of novel HIV-1 viral strains.⁽²³⁾ However, horizontal gene transfer has been reported from phages to bacteria,⁽²⁴⁾ between bacteria,⁽²⁵⁾ or from man to bacteria⁽²⁶⁾ and indeed appears to be a common feature

of all living matter.⁽²⁷⁾ The acquisition of genomic DNA or RNA from infected higher species, by viruses, has also been proposed as a driving force in the evolution of viruses in general.⁽²⁸⁾ Plant, arthropod, fungal, nematode, and protozoan⁽¹⁵⁾ as well as animal and human genomes also contain multiple retroviral and non-retroviral sequences.^(16,18) Clearly, this provides many potential routes for an interviral melange of genomic material. The direction or route of transfer cannot be imputed from a simple bioinformatics alignment, and the reasons for this homology require further laboratory testing. Again this evolutionary aspect is not the central theme of this analysis, and does not alter the implications ensuing from this homology.

All of the viruses for which vaccines are available, or which are known to favorably modulate HIV-1 viral load (Vaccinia, Dengue viruses, GB virus C, and measles) contain sequences matching those of the *env* protein. It is not possible to predict whether any particular sequence would potentially create cross-reactive anti-HIV-1 antibodies, but the Vaccinia virus as well as Dengue viruses, measles, and GB virus C contain several matches in B cell epitope regions of the *env* protein. Field work is necessary to define whether any of these epitopes are able to modify HIV-1 infection. In addition, theoretical T cell epitopes were not examined and are likely to reveal a yet more complex picture that may also depend upon the HLA genetic composition of the host. However, many of the matching sequences are within epitopes known to be able to label the AIDS virus in experimental studies, as cataloged by the immune epitope database. In addition, it is unlikely that all possible epitopes have been reported or characterized. While many of the viral matches were of pentapeptides or greater, multiple tetrapeptide matches were also observed. Antibodies are quite capable of recognizing

Protein comparisons	Alignment: between HIV-1 and other viral proteins
HIV-1 Tat versus AAA45459.1] tat-III protein [Human T-lymphotropic virus 3] E= 6e-71	HIV-1 1 MEPVDFRLEPWKHPGSPKTACTACTNCCYCKCCFHCQVCFITKALGISYGRKKRRQRRRAHQ 60 HTLV3 1 MEPVDFRLEPWKHPGSPKTACTACTNCCYCKCCFHCQVCFITKALGISYGRKKRRQRRRPPQ 60 HIV-1 61 NSQTHQASLSKQPTSQPRGDPTGPKKE 86 HTLV3 61 GSQTHQVLSKQPTSQSRGDPTGPKKE 86
HIV-1 Nef versus AAA45419.1] ORF [Human T-lymphotropic virus 2] E=4e-163	HIV-1 25 PAADRVGAASRDLEKHGAITSSNTAATNAACAWLEAQEEVEEVPFVTPVPLRPMTYKAA 84 HTLV-2 1 PAADRVGAASRDLEKHGAITSSNTAATNAACAWLEAQEEVEEVPFVTPVPLRPMTYKAA 60 HIV-1 85 VDLSHFLKEKGLBGLHLSQRRQQLDLMIYHTQGYFPDQWYTPGQVRYPLTFGWCTK 144 HTLV-2 61 VDLSHFLKEKGLBGLHLSQRRQQLDLMIYHTQGYFPDQWYTPGQVRYPLTFGWCTK 120 HIV-1 145 LVFVPEPKIEEANKGENTSLHPVSLHGMDDPEREVLWRFDSRLAFHHVARELHPPEYFKNC 206 HTLV-2 121 LVFVPEPKIEEANKGENTSLHPVSLHGMDDPEREVLWRFDSRLAFHHVARELHPPEYFKNC 182
HIV-1 env versus ACY78388.1] envelope glycoprotein [Caprine arthritis encephalitis virus] Length=935 (Capri) E= 2e-12	HIV-1 483 LKYYKVKI-----EPLGVAPTKAKRRVQR-----EKRAGV--IGALFLGFLGAA 526 Capri 588 LQKYQVIVRAYTYGVIEMPEYAKTRINRRKRELSHRKKRGVGLVIMLVAIAAAA 647 HIV-1 527 GSTMGAAS--MLTVQARQLSGIVQQNNLRAIEAQHLLQLTVWGIKQLQARILAV 583 Capri 648 GASLGVANAQQSYTKAAVOTLANATAAQQDALEATYA--MVQHVAGVRILEARVARV 704 HIV-1 584 ERYLKDQ---QLLIGWCSGKLICTTAVPNWASNNKSLEQIWNHTTWMEWREINNYT 639 Capri 705 E-AITDRIMLYQELDCHWY--HQYCVTSTRADVA-KYINMTRFKDNTWQWERELQGDV 760 HIV-1 640 SLIHSLSIESQSQKNEQELLEL-DKNASLWNNFNINMLWYIKIFIMIVGLVGLRIV 698 Capri 761 GNLTMLRESARQTALAEQVRIIPDVWSEKVEFVDSWGFWSLKYIPIIIVGLVGCILI 820 HIV-1 699 FAVLSIVNRVQGYSPSFTQLPTPRGPRPEGIEEBGGE 739 Capri 821 RAVICVQCPLVQIYRILTSTPTQYRTVIMEKRAVDAGENQD 861
HIV-1 env versus AA97908.1] envelope glycoprotein [Visna/Maedi virus] E=6e-06	HIV-1 484 YKYYKVKI-EPLGVAPTKAKR--RVQREKRAVIGALFLGFLGAAAGSTMGAAAMTLTVQ 540 Visna 632 YTYGVEMPOQYMEAGMKNKRSRRLQRKKRIGL-VLVLAIMAI AAGAGLGVANAVQ 690 HIV-1 541 -----ARQLLSGIVQQNNLRAIEAQHLLQLTVWGIKQLQARILAVERYLKDQQL-- 592 Visna 691 QSYTRTAVQSLANATAAQDVL---EASVAMVQIAGIIRILEARVARVE-ALVDRMMYI 746 HIV-1 593 --LGLWCSGKLICTA---VPWNASWNNKSLEQIWNHTTWMEWREINNYTSLIHSLSI 646 Visna 747 HELDCWHY--QHYCVTSTKSEVANYVNW---RFKDNCTWQWEREIEQHEHNLSQLL 799 HIV-1 647 EESNQEQEKNEQELLEL-DKNASL--WNFNINMLWYIKIFIMIVGLVGLRIVFAVLSI 704 Visna 800 REAALQVHIAQRDASRIPDVWALQEAFDNSWSSWMLKYIPMIIMGLIICFRILCMVISM 861
HIV-1 env versus AF479638.6 env polyprotein [Ovine lentivirus] E= 4e-04	HIV-1 484 YKYYKVKI-EPLGVAPTKAKRRVQREKRAVIGALFLGFLGAAAGSTMGAAAMTLTVQ-- 540 Ovine 630 YTYGVDMPKAYSEKKRQPSLQRRKRIGLVILVAIMAI AAGAGLGVANAVQSYT 689 HIV-1 541 --ARQLLSGIVQQNNLRAIEAQHLLQLTVWGIKQLQARILAVERYLKDQQLLGIWGC 598 Ovine 690 RTAVQSLANATAAQNVLEATYA--MVQHVAGVRILEARVARVEIVDRMMLYHELDC 746 HIV-1 599 SG-KLICIT---AVPWNASWNNKSLEQIWNHTTWMEWREINNYTSLIHSLSIESQSQ 653 Ovine 747 WHYQHYCVTSTKSEVANYVNW---DNCTWQWEREIEQHEHNLSQLREALQV 801 HIV-1 654 EKNEQELLEL-DKNASL--WNFNINMLWYIKIFIMIVGLVGLRIVFAVLSI 704 Ovine 802 HIAQRDASRIPDVWALQEAFDNSWSSWMLKYIPMIIMGLIICFRILCMVISM 856
HIV-1 env versus ADM23860.1] env polyprotein [Small ruminant lentivirus] (Rumin) E=6e-10	HIV-1 484 YKYYKVKI-EPLGVAPTKAKRRVQREKRAVIGALFLGFLGAAAGSTMGAAAS-- 534 Rumin 581 YTYGVIEB-PRYBQINIKRRELSHRKKRGVGLVIMLVAIAAAGAGLGVANAVQ 639 HIV-1 535 MLTVQARQLSGIVQQNNLRAIEAQHLLQLTVWGIKQLQARILAVERYLKDQQL-- 590 Rumin 640 QSYTRTAVQSLANATAAQDVL---EASVAMVQIAGIIRILEARVARVE-AITDRMMYI 695 HIV-1 591 QLLIGWCSGKLICTA---VPWNASWNNKSLEQIWNHTTWMEWREINNYTSLIHSLSI 647 Rumin 696 QELDCWHYQYCISTRTEVAKYINWT---RFKDNCTWQWEREIQHEHNLSQLL 750 HIV-1 648 ESNQEQEKNEQELLEL-DKNASLWNNFNINMLWYIKIFIMIVGLVGLRIVFAVLSI 704 Rumin 751 SARINQEQEKNEQELLEL-DKNASLWNNFNINMLWYIKIFIMIVGLVGLRIVFAVLSI 704
HIV-1 Gag-Pol versus AAG7168.1 pol polyprotein [Puma lentivirus] E= 8e-100	HIV-1 589 IS---PIETVVKLKGPM---GPKVKQWPLTEEIKKALVEICTE-MEKKGIKIGIPEN 641 Puma 64 ISTKIPI--VKAKL---VDPNKPKIKQWPLTEEIKKALVEICTE-MEKKGIKIGIPEN 117 HIV-1 642 PYNTPVFAIKKDDKTKRKLVDPRFNKRTQDFW---EVQLGIPHPAGLKKKSVTVLD 697 Puma 118 PWNTPIFCIKK-SGKWRMLDPRFNKRTQDFW---EVQLGIPHPAGLKKKSVTVLD 172 HIV-1 698 VGDYFVPLDEDPRKRYTAFIIPSNINNETPGIRYQYV--LPQGWSPFAIPQSSMTKI 754 Puma 173 LQDVFYPLDQDFYFAPFLPKRQGPQRY---VNCSPQGWVLSPLVQSTLKI 229 HIV-1 755 LEPFRKQNDPIVYQVMDLYGSD---LE---IQHRTKIIEELRQHLRWGLTPPKKH 808 Puma 230 LQWPKRYKPNIDVYQVMDLYGSDFSRLEHEKI---IQELRDLIIFWGFPEPKDL 283 HIV-1 809 QKEPPFLMWGELHPDKWTVQ-----PIVLPEKDSWTVNDIQKLWKLWASQYPIKVRQLKLLRGTKALTE 878 Puma 284 QQEPYKMGYTLYPNWTIKQTKLQDPEV-P-----TLNQLQKLAGVINWATQVGGIKI-----KALTE 343
HIV-pol versus ADK35834.1 pol polyprotein [Equine infectious anemia virus] E= 1e-98	HIV-1 511 LLDTGADDTLEMSLP-----GRWKPK-----MIGGIG--F-----I 542 Equine 91 LLDTGADDTLEMSLP-----GRWKPK-----MIGGIG--F-----I 144 HIV-1 543 KVRQYQDLIEICGKHAIGTVLGPFTVNIIGNRLLTOIGCTLANP---ISPETVIV 597 Equine 145 KTR-----MLVADIPVTILGRDILQELGAQLTMAQLSKAITPRE--I 184 HIV-1 598 KLGPMGDGPKVQWPLTEEK-----IKALVEICTE-MEKKGIKIGIPENYTPVFAI 650 Equine 185 KLTGTVGPVKPQWPLTEEKLLGAKIVKLLD-----EGKISEASDNPYNPFI 237 HIV-1 651 KKKSTKWRKLVDFRELNKRKQDFWVQLG-----IPHAGLKKKSVTVLDVGDAYS 704 Equine 238 KKK-SGWRLLQDLRELK-----VYQVTEISROLPHQGLKCNHMTLDIGDAYFT 290 HIV-1 705 VFLDEDPRKYTAFIIPSNINNETPGIRYQVYVPLQGNKSPAIPOSSMTKILEPFRKQND 764 Equine 291 IPLDPRKRYTAFIIPSNINNETPGIRYQVYVPLQGNKSPAIPOSSMTKILEPFRKQND 350 HIV-1 765 IVIQYQMDLTVGSDLE---IQHRTKIIEELRQHLL-RWGLTTPDKHKQKPPFLMWGYE 820 Equine 351 VQLYQYMDLTVGSDLE---EQRKELVEELRAILLEK-GFEMPEDKLQEBAPYNLWGY 406 HIV-1 821 LHPDKTVQPIVPLFE-KDSWTVNDIQKLWQKLNW-ASQYPIKVRQL-----CKLLR 871 Equine 407 LSPDNMKVQKML-ELVPE-TLNDVQKLMGNITWSSGV-PGLTVQOIAATTGK-L- 460 HIV-1 872 GTKALTE-VIPLTEEALELAR-NREILKEPVHVG-YDPSKDLIAEQ----- 917 Equine 461 ---DLNQKVV-WTEEAQLEBNNKKI---QEAQQLQYNNPEBVEICEIETKNYATYII 514 HIV-1 918 KQQQ-QWYIYQYEPFKNLKTKYAR-MRGAHT--NDVKQLTEAVQKITTESIV-IWGL 972 Equine 515 KQSQILW-----AGK-KIMR-ANGLWASAKNMLLLQHVATESIVRI-GT 557 HIV-1 973 TPKFKLPIQKE--TWET---HWTEYQATWIP-----EWFNTPPLVLWLYQL 1016 Equine 558 CPKFKVPTKEQKMETEKHGV--Y---SWLPMYIYHQVWHDH-----RL-KL--V 602 HIV-1 1017 EKRP---IVGAEYFVYDGAANRETLLGK-----AGYVTRNG--RQK---VVTLDTIN 1061 Equine 603 E-QPASGI---TIYTD-----GKNEGVAAYVTNGKTQKRLGVP-----TH 643 HIV-1 1062 QKTELQAIYLALQDSGLE--VNI VTDQY-----ALG-----IIQAQPDQS 1100 Equine 644 QTAERIAQMALED--EETLWNIYVDS-YCWNKNTBGLGEGPSPWMPPIQ----- 694 HIV-1 1101 ESELWQILIEQIK-KEKYLAHVFAHKIIGQEVQD 1136 Equine 695 ----N-----IRAKEMVYFANVPGHKIYGNQLAD 720

Figure 1. Significant overall homologies of HIV-1 viral proteins with proteins from other viruses. Consensi and e values are shown in bold.

Table 2. Examples of Vaccinia virus homologues (from BLASTS of the relevant HIV-1 *env* proteins versus Ankara, GLV-1h68, Tian Tian, and L-IPV Vaccinia strains) compared with the *env* glycoprotein from a selection of HIV-1 viral strains (various subtypes from groups M, N, and O).

Group M subtype A (isolate Z321) (Accession # P05881)	Group M subtype B (isolate BRU/LAI) (Accession # P03377)	Group M subtype B (isolate BH10) (Accession # P03375)	Group M subtype C (isolate ETH2220) (Accession # Q75008)
340: DTLSKV	187: CSFNIS S	524: GAAGST	154: CSFNI
388: TSGLF	529: GAAGST		391: LELFN
524: GAAGST	303: KLDII ID		429: GIIMC
488: VVKIEP	493: VVKIEP		512: GAAGST
617: KSQSD	705: A LSIVN		574: HLRDQ
644: NLIEE			629: IYINL
702: LL LSIIN			690: LSIVN
703: SIINR			685: IIFAV
841: LNIPR			
Group M subtype D (isolate Z84) (Accession # P05882)	Group M subtype F1 (isolate VI850) (Accession # Q9QSQ7)	Group M subtype G (isolate 92NG083) (Accession # O41803)	Group M subtype H (isolate 90CF056) (Accession # O70902)
63: EAHNI	377: TSGLF	329: NVSRI	661: WFDIS
197: NTNYT Y	386: SNNGT	352: NKNIT	690: LSIVN
291: NNVKTHI	678: LSIVN	383: TSGLF	752: LSLFS
364: LNQTT	696: LIPSP	392: SNINN	
495: VV IEP	753: IAARI	473: KTVK+K	
531: GAAGST	768: ALKYL	510: GAAGST	
	771: YLGNL	688: LSIVN	
	817: LNIPR		
Group M subtype J (isolate SE9173) (Accession # Q9WC69)	Group M subtype K (isolate 96CM-MP535) (Accession # Q9QBY2)	Group N (isolate YBF106) (Accession # Q9IDV2)	Group O (isolate ANT70) (Accession # Q77377)
146: SPEIM N	187: NNSST	74: LLTNV	254: QLILN
180: INSDN	448: NTHNE	79: TEYFN	544: HTLLK
194: TSVIK	510: GAAGST	134: +RTEDL	696: RVIMI
482: VVELEP	732: VRLVS	156: RDRKK	698: IMIVL
615: DIWEN	797: AISLL	250: QLILN	704: IVKNIR+G
691: IIFAV		476: VSREK	
		513: RTLLS	
		787: LKDSAI	

Identical peptides (HIV-1 = Vaccinia) were analyzed for B cell antigenicity using the BepiPred server and those predicted as epitopes are highlighted in bold.

such short sequences⁽²⁹⁾ A further point to be considered is that this homology may enable different viruses to share the same binding partners in relation to the host proteome. Viruses also demonstrate this type of homology with human proteins,^(18,30,31) an ability that no doubt enables them to compete with their human counterparts as binding partners in the numerous host/pathogen interactomes that they use during their life cycles.^(17,19) HIV-1 and pox viruses both use the CCR5 chemokine receptor and such sharing may also influence the outcome of co-infection.⁽⁴⁾

The differing matching peptide profiles for different HIV-1 viral strains also highlights the underlying complexity and shows that matching sequences will depend upon the HIV-1 strain, and presumably also the strain of the homologous virus.

In relation to AIDS, this homology may have clinical application as infection or vaccination in relation to these viral homologues might be expected, in some cases, to confer cross-reactive immunity to HIV-1. There is indeed some evidence that this may be operative. For example, Vaccinia vaccination in HIV-1 infected subjects has been shown to inhibit HIV-1 viral replication in subsequent *in vitro* tests, but only in the CCR5 tropic HIV-1 Major M strain.⁽⁴⁾ These authors noted that the increase in the

incidence of AIDS correlated with the successful eradication of smallpox and cessation of the use of Vaccinia vaccination. However, even within the M group of HIV-1 viruses, which displays tropism for the CCR5 chemokine receptor,⁽³²⁾ considerable variation exists between the Vaccinia/HIV-1 peptide matches.

In a small study (four patients), hyperimmunization with the killed poliomyelitis (Salk) vaccine was also shown to increase the T cell count and to improve symptoms in HIV-1 infected patients.⁽³³⁾ Influenza vaccination in non-HIV-1 patients also results in the suppression of HIV-1 replication *in vitro*. However, this was not observed in HIV-1 infected patients, and influenza vaccination has also been reported to increase HIV-1 replication in some patients,⁽³⁴⁾ perhaps due to the ability of the influenza virus to inhibit viral defense pathways.⁽¹⁰⁾ Both measles or GB virus C infection are also known to decrease the HIV-1 viral load in infected patients^(7,8) although other coinfections may perhaps worsen the effects of each other.

Vaccination can be a double-edged sword. For example, a lower titre of hepatitis B antibodies has been observed in several autoimmune disorders, including multiple sclerosis, suggesting a protective effect of infection.⁽³⁵⁾ However, hepatitis B vaccination can have the opposite effect and provoke demyelinating lesions in certain cases.⁽³⁶⁾ It has

been shown that the HIV-1 proteome displays a similar type of homology with the human proteome and the problems of autoimmunity in relation to certain of these vaccines need to be addressed.⁽³¹⁾ Nevertheless, Vaccinia virus vaccination does reduce the HIV-1 viral load for the common CCR5 tropic strain, *in vitro*, and a resumption of smallpox vaccination might be expected to be of benefit in certain cases, as already suggested.⁽⁴⁾ It would be premature to suggest the immediate use of other available vaccines as preventive agents without further research into the question. A more in-depth analysis of the viral homology of the *env* glycoprotein and of other HIV-1 proteins and strains is also necessary, and, given the scale of the phenomenon, which also applies to millions of viral/human and bacterial/human short consensi.^(17,37) It is clear that the development of powerful algorithms is necessary for this purpose. However, the results with the Vaccinia virus are promising and suggest that this homology may be harnessed to good effect. Whether other available vaccines could confer cross-reactive immunity remains to be assessed. These are often used in HIV-1 positive patients, once HIV-1 is present,⁽³⁸⁾ but their use as potential preventive agents, given prior to HIV-1 infection, merits further study.

A further point to consider is the microbiome in AIDS patients. If so many viruses, and probably also bacteria and other pathogens, resemble HIV-1 viral proteins, it is possible that certain species could exert beneficial (or deleterious) effects. Sequencing of the various microbiota in AIDS resistant and nonresistant patients may, thus, be of value as such analyses may well be able to identify protective viral or bacterial strains. Microbiomes can exert a powerful influence on disease. For example, the intestinal microbiome is able to influence obesity, cardiovascular disease, and inflammatory bowel disease,⁽³⁹⁾ and its manipulation in relation to HIV-1 is already attracting attention.⁽⁴⁰⁾ Indeed, probiotic yoghurt containing *Lactobacillus rhamnosus Fiti* is able to increase the CD4+ cell count in HIV-1 infected patients.⁽⁴¹⁾

HIV-1 vaccine development using attenuated Ankara Vaccinia strains, containing HIV-1 proteins, is already under development.⁽⁴²⁾ The most immediately relevant conclusion of this study is that the beneficial effects of unmodified Vaccinia vaccination in HIV-1 infected patients, *in vitro*, may well be related to cross-reactive immunity due to Vaccinia/HIV-1 homology, and that, as previously suggested⁽⁴⁾ a resumption of Vaccinia/smallpox vaccination might have a significant effect on the AIDS pandemic, even if only effective against certain strains.

Clearly further work is needed, both *in vitro* and *in vivo* to analyze these effects. Rather than suggest specific proposals for vaccine development or the use of already available vaccines, the main purpose of this article is to draw attention to this extensive protein homology, which may have far-reaching implications in this and other diseases. A similar bioinformatics approach may be relevant to other recalcitrant viruses, bacteria, and pathogens and the current treasury of available vaccines may well find

uses in diseases other than those for which they were designed. Other HIV-1 proteins and numerous strains of both the HIV-1 and other viruses also require analysis perhaps enabling the construction of more effective epitopes. The wheel has turned full circle since Edward Jenner's observation over 200 years ago that cowpox prevented smallpox, as, if this is effective, the same phenomenon and the same viruses may have a role to play in relation to today's viral scourge.

Declaration of interest

The author declares no conflict of interest.

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