

Expanding roles for CD4⁺ T cells in immunity to viruses

Susan L. Swain, K. Kai McKinstry and Tara M. Strutt

Abstract | Viral pathogens often induce strong effector CD4⁺ T cell responses that are best known for their ability to help B cell and CD8⁺ T cell responses. However, recent studies have uncovered additional roles for CD4⁺ T cells, some of which are independent of other lymphocytes, and have described previously unappreciated functions for memory CD4⁺ T cells in immunity to viruses. Here, we review the full range of antiviral functions of CD4⁺ T cells, discussing the activities of these cells in helping other lymphocytes and in inducing innate immune responses, as well as their direct antiviral roles. We suggest that all of these functions of CD4⁺ T cells are integrated to provide highly effective immune protection against viral pathogens.

Pattern-recognition receptors

(PRRs). Host receptors that can detect pathogen-associated molecular patterns and initiate signalling cascades, leading to an innate immune response. Examples include Toll-like receptors (TLRs) and NOD-like receptors (NLRs). PRRs can be membrane-bound receptors (as in the case of TLRs) or soluble cytoplasmic receptors (as in the case of NLRs, retinoic acid-inducible gene I (RIG-I) and melanoma differentiation-associated protein 5 (MDA5)).

Viruses can enter the body by diverse routes, infect almost every type of host cell and mutate to avoid immune recognition. Destroying rapidly dividing viruses efficiently requires the coordination of multiple immune effector mechanisms. At the earliest stages of infection, innate immune mechanisms are initiated in response to the binding of pathogens to pattern-recognition receptors (PRRs), and this stimulates the antiviral activities of innate immune cells to provide a crucial initial block on viral replication. Innate immune responses then mobilize cells of the adaptive immune system, which develop into effector cells that promote viral clearance.

Activation through PRRs causes professional antigen-presenting cells (APCs) to upregulate co-stimulatory molecules and promotes the migration of these cells to secondary lymphoid organs. Here, they present virus-derived peptides on MHC class II molecules to naive CD4⁺ T cells and deliver co-stimulatory signals, thereby driving T cell activation. The activated CD4⁺ T cells undergo extensive cell division and differentiation, giving rise to distinct subsets of effector T cells (BOX 1). The best characterized of these are T helper 1 (T_H1) and T_H2 cells, which are characterized by their production of interferon- γ (IFN γ) and interleukin-4 (IL-4), respectively¹. Specialized B cell helpers, known as follicular helper T (T_{FH}) cells, and the pro-inflammatory T_H17 cell subset also develop, along with regulatory T (T_{Reg}) cells, which are essential for avoiding over-exuberant immune responses and associated immunopathology².

A key role of CD4⁺ T cells is to ensure optimal responses by other lymphocytes. CD4⁺ T cells are necessary as helpers to promote B cell antibody production and are often required for the generation of cytotoxic and

memory CD8⁺ T cell populations. Recent studies have defined additional roles for CD4⁺ T cells in enhancing innate immune responses and in mediating non-helper antiviral effector functions. We discuss what is known about the T cell subsets that develop following acute viral infection and how different subsets contribute to viral control and clearance.

Following a rapid and effective antiviral response, infection is resolved and the majority of effector CD4⁺ T cells die, leaving a much smaller population of memory CD4⁺ T cells that persists long-term^{3,4}. Memory CD4⁺ T cells have unique functional attributes and respond more rapidly and effectively during viral re-infection. A better understanding of the functions of memory CD4⁺ T cells will allow us to evaluate their potential contribution to immunity when they are induced by either infection or vaccination. We describe the antiviral roles of CD4⁺ T cells during the first encounter with a virus and also following re-infection.

Generation of antiviral CD4⁺ T cells

To develop into effector populations that combat viral infections, naive CD4⁺ T cells need to recognize peptide antigens presented by MHC class II molecules on activated APCs. PRR-mediated signalling activates APCs to upregulate their expression of MHC class II molecules, co-stimulatory molecules (such as CD80 and CD86) and pro-inflammatory cytokines (such as type I IFNs, tumour necrosis factor (TNF), IL-1, IL-6 and IL-12)⁵. When the activated APCs migrate to draining lymph nodes, they prime naive virus-specific CD4⁺ T cells, which then differentiate into antiviral effectors (FIG. 1). The priming environment can vary

Department of Pathology,
University of Massachusetts
Medical School, 55 Lake
Avenue N, Worcester,
Massachusetts 01655, USA.
Correspondence to S.L.S.
e-mail:

Susan.Swain@umassmed.edu
doi:10.1038/nri3152

Published online
20 January 2012

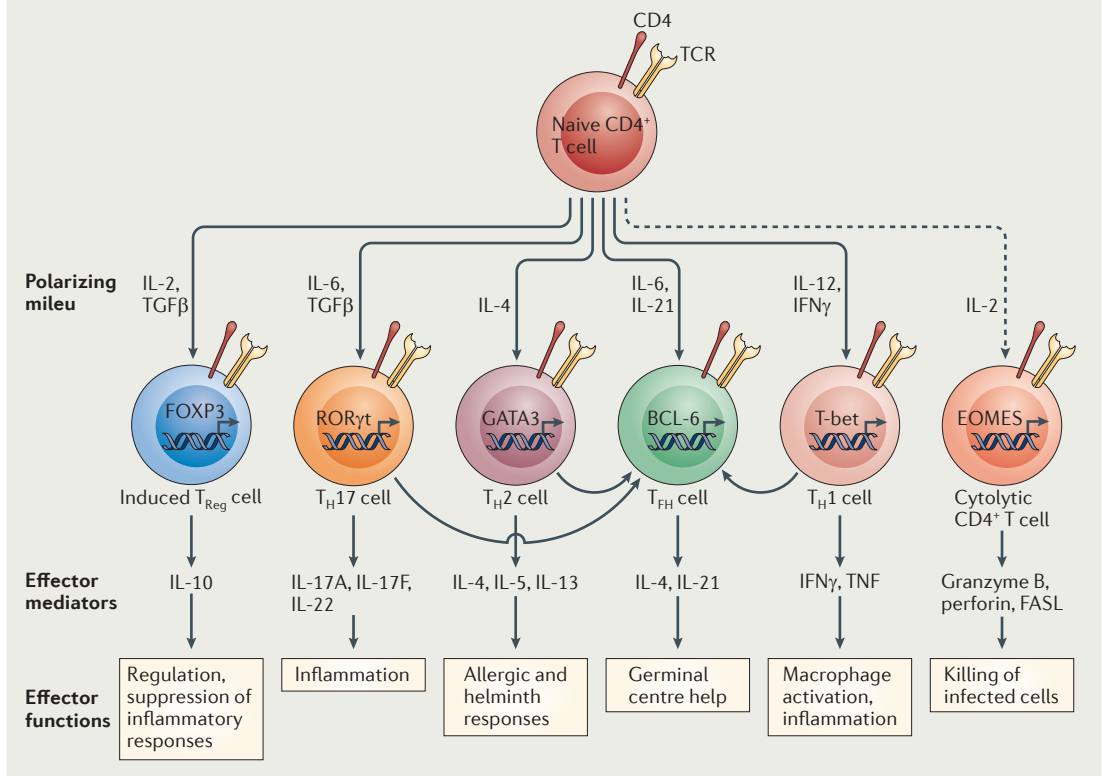
Box 1 | Subsetting CD4⁺ T cell responses based on T_H cell polarization

Following recognition of a specific antigen presented by an appropriately activated antigen-presenting cell, naive CD4⁺ T cells undergo several rounds of division and can become polarized into distinct effector T helper (T_H) cell subsets that differentially orchestrate protective immune responses (see the figure). The differentiation of polarized effector T cells is controlled by unique sets of transcription factors, the expression of which is determined by multiple signals but particularly by soluble factors that act on responding CD4⁺ T cells during their activation. The elucidation of the crucial cytokines that govern the differentiation of distinct T_H cell subsets has allowed researchers to examine the protective capacities of differently polarized CD4⁺ T cell subsets in several models of infectious disease.

Although the production of the signature cytokines interferon-γ (IFNγ), interleukin-4 (IL-4) and IL-17 is routinely used to assign subsets (T_H1, T_H2 and T_H17, respectively) to responding CD4⁺ T cells, it is increasingly clear that considerable plasticity exists within T_H cell subsets *in vivo*, especially during responses to pathogens. Moreover, certain cytokines (for example, IL-10) can be produced by subpopulations of cells within multiple effector subsets. Finally, the successful clearance of viral pathogens in particular often depends on complex CD4⁺ T cell responses that encompass multiple T_H cell subsets. Together, these T cell subsets are capable of mediating direct antiviral functions, of providing help for B cells, of regulating immunopathology and of mediating cytotoxic killing of virus-infected cells. CD4⁺ T cells with cytotoxic activity have been described in several models of viral disease as well as in the clinic. Following re-stimulation, memory CD4⁺ T cells retain their previous effector functions and rapidly produce effector cytokines. This property of primed CD4⁺ T cells represents a key advantage of the memory state.

Certain T_H cell subsets — including follicular helper T (T_{FH}) cells and regulatory T (T_{Reg}) cells — are often defined less by their cytokine profile and more by their functional attributes. Further studies will be required to determine whether populations of CD4⁺ T cells with specialized functions can include cytokine-polarized cells from several subsets.

BCL-6, B cell lymphoma 6; EOMES, eomesodermin, FASL, FAS ligand; FOXP3, forkhead box P3; GATA3, GATA-binding protein 3; RORγt, retinoic acid receptor-related orphan receptor-γt; TCR, T cell receptor; TGFβ, transforming growth factor-β; TNF, tumour necrosis factor.



T-bet
A member of the T-box family of transcription factors. T-bet is a master switch in the development of T helper 1 (T_H1) cell responses through its ability to regulate the expression of the interleukin-12 receptor, inhibit signals that promote T_H2 cell development and promote the production of interferon-γ.

dramatically during different viral infections and is influenced by many variables, including the route of infection, the viral dose and the organ or cell types targeted. As has been extensively reviewed elsewhere, the extent of T cell proliferation and the determination of T cell subset specialization are affected by the specific subset of APCs that is activated^{6,7}, the antigen load and the duration of antigen presentation⁸, and the patterns and amounts of cytokines produced by different APCs⁹.

Effector T_H cell subsets in viral infection

In contrast to T_H cell populations that are generated *in vitro*, effector T cells that are found *in vivo* are often characterized by plasticity and heterogeneity in terms of their cytokine-producing potential. Nevertheless, the CD4⁺ T cells that are generated in response to viral infection mainly have a T_H1-type phenotype and produce large amounts of IFNγ and express T-bet. This phenotype classically depends on the exposure of T cells to high levels of IL-12, type I IFNs and IFNγ in

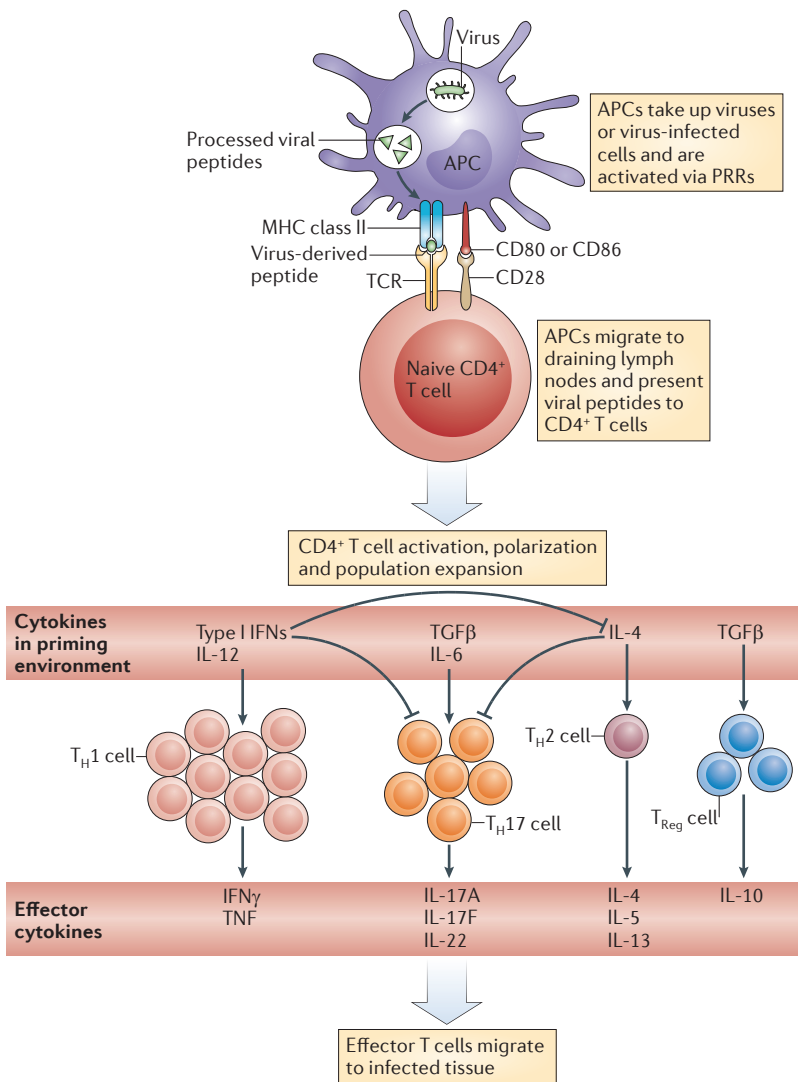


Figure 1 | Generation of antiviral effector CD4⁺ T cells. The crucial initial steps in generating primary antiviral T cell responses are the uptake of viral antigens by antigen-presenting cells (APCs) in infected tissue, the activation of APCs by pattern-recognition receptor (PRR) ligation, and the migration of these cells to draining lymph nodes. The nature of the viral infection, as well as PRR ligation, can influence the activation status of antigen-bearing APCs and the T helper (T_H) cell-polarizing environment. The recognition of antigens on activated APCs by naive T cells during viral infection predominately results in the generation of T_H1 cells owing to the presence of type I interferons (IFNs) and interleukin-12 (IL-12). However, T_H17, T_H2 and regulatory T (T_{Reg}) cell populations are also generated to some degree in certain viral infections. TCR, T cell receptor; TGFβ, transforming growth factor-β; TNF, tumour necrosis factor.

the priming milieu¹⁰, although T_H1 cell responses are generated in response to certain viruses independently of IL-12 or type I IFNs^{11–13}, suggesting that other factors can also contribute to T_H1 cell polarization. IL-12 and type I IFNs promote T_H1 cell differentiation both directly and indirectly (by repressing the development of other T_H cell subsets), and can even influence effector T cells that are already polarized. For example, polarized T_H2 cells that were transferred to hosts infected with lymphocytic choriomeningitis virus (LCMV) acquired a mixed T_H1/T_H2 cell phenotype, which was

characterized by high levels of IFNγ production and diminished IL-4 production. Interestingly, most IFNγ-producing cells in the T_H1/T_H2 cell population co-produced IL-4 or IL-13, and IL-12 and type I IFNs were shown to drive this mixed T_H1/T_H2 cell phenotype¹⁴. *In vivo*, IL-12 can also reprogramme *in vitro*-polarized T_H17 cells to a T_H1-type phenotype^{15,16}.

Originally, it was thought that IL-4-producing T_H2 cells were needed to drive optimal humoral immune responses. Thus, the predominance of T_H1 cells over T_H2 cells during viral infection was somewhat surprising, given the important role of neutralizing antibodies in viral clearance and in providing long-term immunity to re-infection. However, adoptive transfer of either T_H1 cells or T_H2 cells was shown to provide efficient help for the generation of neutralizing antiviral IgG responses^{17,18}. The signature T_H1-type cytokine, IFNγ, enhances IgG2a class switching, and this explains why IgG2a is usually the dominant isotype in IgG responses generated against viruses¹⁹. In fact, several studies have found that, far from promoting antiviral responses, T_H2 cell-associated mediators (and IL-4 in particular) have a strong negative impact on immune protection and drive immunopathology during infection with many viruses, including influenza virus^{20,21}, respiratory syncytial virus (RSV)²², herpes simplex virus (HSV)²³ and vaccinia virus²⁴. Instead, it is now clear that IL-4-producing T_{FH} cells provide much of the help required for IgG1 production (see below).

The roles of T_H17-type effector responses during viral infection are not well understood, but virus-specific IL-17-producing CD4⁺ T cells have been detected in mice following infection with mouse cytomegalovirus (MCMV)²⁵, HSV²⁶, vaccinia virus²⁷, Theiler's murine encephalomyelitis virus²⁸ or influenza virus¹⁶, although at levels lower than those of T_H1 cells. The generation of polarized T_H17 cells during viral infection has been correlated with high levels of IL-6 and may also be influenced by transforming growth factor-β (TGFβ)²⁸. T_H17 cells are implicated in driving harmful inflammation during autoimmunity, and IL-17 may contribute to immunopathology during responses against viruses, as demonstrated in studies using influenza virus or vaccinia virus^{27,29}. However, in some cases, T_H17 cells contribute to host protection against viruses¹⁶. One protective mechanism mediated by T_H17 cells might be the promotion of enhanced neutrophil responses at sites of infection. IL-17 upregulates CXC-chemokines that promote neutrophil recruitment³⁰, and neutrophils can contribute to protection against certain viruses, including influenza virus³¹. We expect that other roles for T_H17 cells will be identified in future studies, as T_H17 cells often produce significant levels of IL-21 and IL-22 in addition to IL-17 (REF. 16). Although not extensively studied in viral systems, IL-22 production by T_H17 cells could be involved in regulating the expression of defensins, and could also play an important part in tissue repair (reviewed in REF. 32). IL-21 production may also regulate aspects of the innate immune response during viral infection. Moreover, IL-21 is involved in sustaining CD8⁺ T cell responses during chronic viral infection, as discussed below.

Roles of CD4⁺ T cells during primary infection

Although the best-studied pathways of CD4⁺ T cell-mediated help are those that promote antibody production by B cells, CD4⁺ T cells also enhance effector CD8⁺ T cell responses during certain viral infections and contribute to the maintenance of a functional memory CD8⁺ T cell pool (FIG. 2). Furthermore, effector CD4⁺ T cells regulate the inflammatory response and can directly mediate viral clearance.

CD4⁺ T cell-mediated help for B cells. Current licensed vaccines directed against viral pathogens are evaluated almost exclusively on their ability to generate strong neutralizing antibody responses. Antibody-mediated protection can be extraordinarily long-lived³³, and neutralizing antibodies present at the time of pathogen encounter can prevent rather than combat infection, thereby achieving 'sterilizing' immunity. Thus, it is crucial to understand the mechanisms by which CD4⁺ T cells help B cells during viral infection in order to define what is required for effective vaccination (FIG. 2a). CD4⁺ T cells that enter B cell follicles and provide help to B cells, resulting in germinal centre formation, are now referred to as T_{FH} cells. The generation and functions of T_{FH} cells have been expertly reviewed elsewhere^{34,35}, so we focus only on the roles of these cells during viral infection.

Following viral infection, the expression of SLAM-associated protein (SAP) by T_{FH} cells is necessary to direct the formation of germinal centres^{36,37}, where T_{FH} cells promote the generation of B cell memory and long-lived antibody-producing plasma cells^{36,38}. Thus, T_{FH} cells are likely to be important for generating long-lived antibody responses and protective immunity to most, if not all, viruses. Indeed, CD4⁺ T cells have been shown to be required for the generation of optimal antibody responses following infection with coronavirus³⁹, vaccinia virus⁴⁰, yellow fever virus⁴¹ or vesicular stomatitis virus (VSV)⁴². It is not yet clear how distinct cytokine-polarized CD4⁺ T cell subsets influence the B cell response during primary viral infection. One possibility is that distinct T_H cell subsets, such as T_H1, T_H2 and T_H17 cells, can each develop into T_{FH} cells and provide efficient help for B cells. This hypothesis is supported by a recent study demonstrating that such polarized subsets can be reprogrammed to express T_{FH} cell characteristics *in vitro*⁴³. As different T_H cell subsets have been associated with distinct antibody class-switching responses, the broad range of protective antibody isotypes that is often found in individuals with immunity to a virus favours such a model.

Several co-stimulatory ligands expressed by CD4⁺ T cells contribute to the promotion of B cell activation and antibody production. One notable ligand is CD40 ligand (CD40L). Interactions between CD40 (which is expressed on B cells) and CD40L (which is expressed on activated CD4⁺ T cells) are crucial for generating optimal humoral responses against several viral pathogens, including LCMV, Pichinde virus, VSV, HSV and influenza virus^{44–46}. The expression of inducible T cell co-stimulator (ICOS) by T_{FH} cells has also been shown to

be important for germinal centre formation³⁴, and ICOS expression is required for optimal induction of humoral responses against LCMV, VSV and influenza virus⁴⁷. The roles during viral infection of other co-stimulatory molecules expressed by T_{FH} cells are less clear. For example, OX40-deficient mice generate normal levels of class-switched antibodies during infection with LCMV, VSV or influenza virus⁴⁸. Further studies are required to determine the importance of additional signals that pass between T_{FH} cells and B cells in generating protective antibody responses during viral infection.

CD4⁺ T cell-mediated help for CD8⁺ T cells. The mechanisms by which CD4⁺ T cells promote CD8⁺ T cell effector and memory responses are less well understood than B cell help. As in B cell help, CD40L–CD40 interactions between CD4⁺ T cells and APCs are crucial (FIG. 2). One possible mechanism — the 'licensing' of APCs by CD4⁺ T cells — may be of only minor importance during viral infection, as APCs can be activated effectively by viruses through PRRs^{49,50}, thus obviating the need for this process^{50–52}. However, it is unclear whether APCs that are activated directly through PRRs are functionally similar to those licensed by CD4⁺ T cells⁵³. An absence of CD4⁺ T cells has been shown to compromise the generation of primary cytotoxic T lymphocyte (CTL) responses against vaccinia virus⁵⁴ and HSV⁵⁵ (and influenza virus in some, but not all, studies^{56–58}), suggesting that the two modes of APC activation are distinct. Whether CD4⁺ T cell-mediated help is required for the generation of optimal antiviral CD8⁺ T cell responses probably depends on which elements of the innate immune response are triggered following infection by a virus and to what extent. For example, the strong type I IFN response that is induced by administration of polyinosinic–polycytidylic acid (polyI:C) can bypass the otherwise obligate requirement for CD4⁺ T cell-mediated help in the generation of CD8⁺ T cell responses following vaccinia virus infection⁵⁹.

It is not clear whether different T_H cell subsets have distinct roles in helping CD8⁺ T cells. During the primary response, the licensing of APCs by CD4⁺ T cells probably occurs before the full polarization of effector CD4⁺ T cells, but fully polarized T_H cells might also promote efficient CD8⁺ T cell responses against viral pathogens through mechanisms other than APC licensing. For example, chemokines produced following antigen-specific interactions between APCs and CD4⁺ T cells can actively attract CD8⁺ T cells to the activated APCs⁶⁰, and CD40L–CD40 interactions between CD4⁺ T cells and APCs can protect APCs from CTL-mediated death⁶¹, perhaps leading to more-efficient CD8⁺ T cell priming.

In addition, CD4⁺ T cells facilitate the development of functional, pathogen-specific memory CD8⁺ T cells that can respond following re-infection^{51,52,62,63}. One mechanism by which CD4⁺ T cells promote this process involves the downregulation of TNF-related apoptosis-inducing ligand (TRAIL) expression on responding CD8⁺ T cells (FIG. 2). It is thought that CD8⁺ T cells that are helped by CD4⁺ T cells downregulate

Class switching

The process by which proliferating B cells rearrange their DNA to switch from expressing the heavy-chain constant region of IgM (or another class of immunoglobulin) to expressing that of a different immunoglobulin class, thereby producing antibodies with different effector functions. The decision of which isotype to generate is strongly influenced by the specific cytokine milieu and by other cells, such as T helper cells.

Germinal centre

A highly specialized and dynamic microenvironment that gives rise to secondary B cell follicles during an immune response. Germinal centres are the main site of B cell maturation, which leads to the generation of memory B cells and plasma cells that produce high-affinity antibodies.

Polyinosinic–polycytidylic acid

(PolyI:C). A substance that is used as a mimic of viral double-stranded RNA.

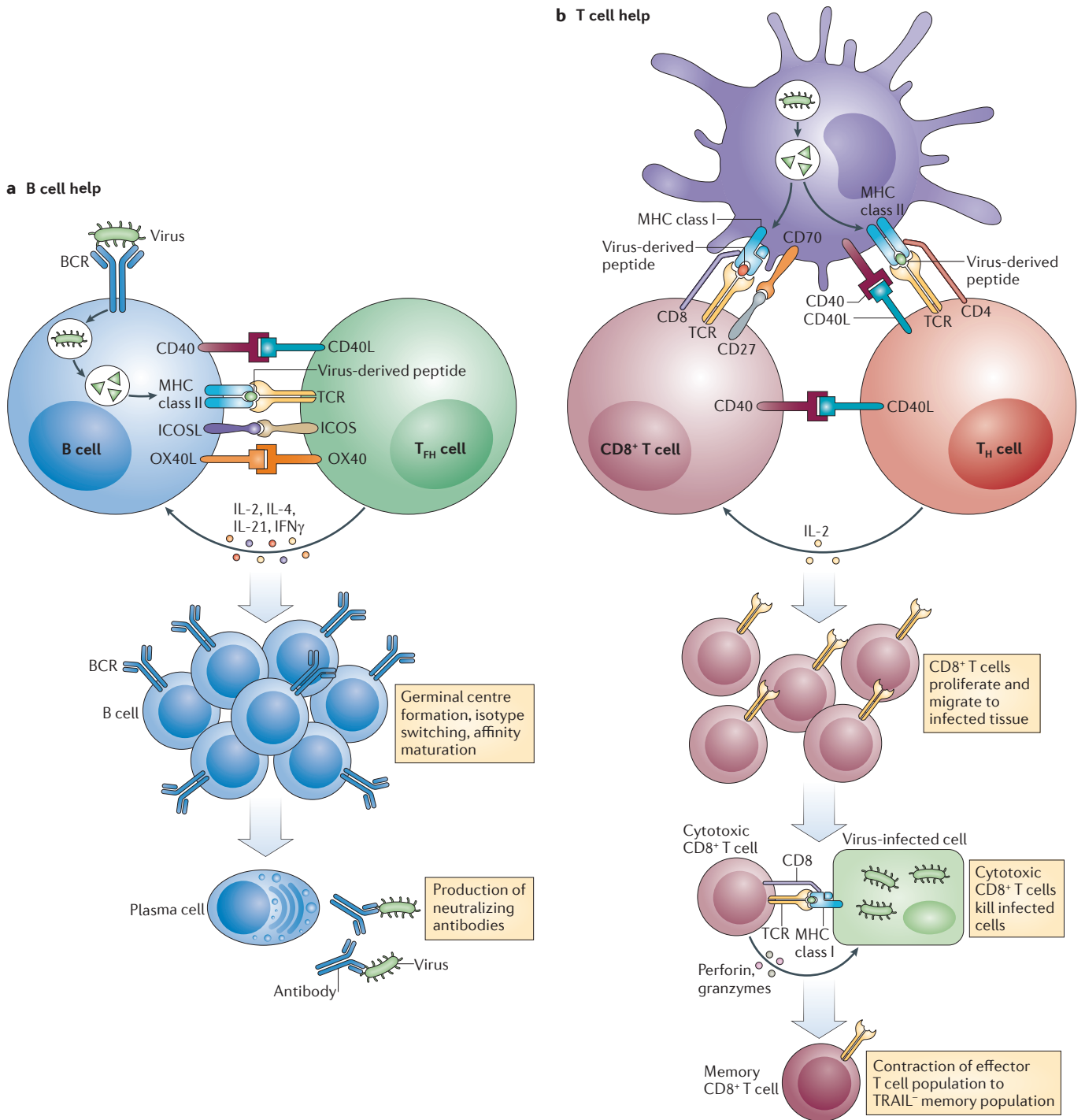


Figure 2 | Helper functions of CD4⁺ T cells. a | The canonical function of CD4⁺ T cells is the provision of help for B cells in germinal centre formation, isotype switching and affinity maturation of antibody responses. Follicular helper T (T_{FH}) cells are a specialized subset of CD4⁺ T cells that provide help to B cells through both cell–cell interactions (most notably CD40L–CD40 interactions) and the release of cytokines. The generation of neutralizing antibodies is a crucial component of protection against many viral pathogens and the goal of most vaccine strategies. **b** | The best-characterized pathway of CD4⁺ T cell-mediated help in the generation of CD8⁺ T cell effectors involves the provision of interleukin-2 (IL-2) and the activation, known as ‘licensing’, of antigen-presenting cells (APCs) via CD40L–CD40 interactions. It is often unclear whether CD4⁺ T cell-mediated help has a role in the initial generation of antiviral CD8⁺ T cell effector responses, presumably because viruses can trigger pattern-recognition receptors and independently activate APCs. However, a clear role for CD4⁺ T cell-mediated help in the generation of functional memory CD8⁺ T cells has been demonstrated during viral infection. Downregulation of TNF-related apoptosis-inducing ligand (TRAIL) expression on CD8⁺ T cells is a prominent feature of CD8⁺ T cells that have been helped and at least in part facilitates their robust recall response during secondary infection. BCR, B cell receptor; ICOS, inducible T cell co-stimulator; ICOSL, ICOS ligand; TCR, T cell receptor; T_H, T helper.

TRAIL expression and become less susceptible⁶⁴, or have delayed susceptibility⁶⁵, to TRAIL-mediated apoptosis. By contrast, CD8⁺ T cells that have not been helped undergo enhanced TRAIL-mediated apoptosis following antigen re-exposure. CD4⁺ T cell-mediated help also controls the expression of other molecules. For example, CD4⁺ T cells downregulate the expression of programmed cell death protein 1 (PD1) on CD8⁺ T cells, and this can enhance the function of pathogen-specific memory CD8⁺ T cells^{66–68}.

CD4⁺ T cells may promote the generation of effector and memory CD8⁺ T cell populations through many possible pathways. One such pathway involves enhancing the APC-mediated production of cytokines that augment initial CD8⁺ T cell responses; these cytokines include IL-1, IL-6, TNF and IL-15 (REF. 69). Paracrine IL-2 produced by CD4⁺ T cells during the initial priming of CD8⁺ T cells in LCMV infection dramatically improves the CD8⁺ T cell recall response potential⁷⁰. Furthermore, CD4⁺ T cells have been shown to upregulate the expression of CD25 (also known as IL-2R α) on CD8⁺ T cells during infection with vaccinia virus or VSV⁷¹. At later stages of the response, CD4⁺ T cells produce additional cytokines, such as IL-21, which appears to be a crucial signal for downregulating TRAIL expression on CD8⁺ T cells responding to vaccinia virus⁷². Finally, evidence suggests that direct ligation of CD40 on naive CD8⁺ T cells by CD40L on CD4⁺ T cells can enhance the generation of memory CD8⁺ T cells⁷³ (FIG. 2).

CD4⁺ T cells seem to be particularly important for maintaining memory CD8⁺ T cell populations⁷⁴, and the presence of CD4⁺ T cells during priming may influence the homing pattern and, ultimately, the tissue distribution of memory CD8⁺ T cells⁷⁵. Whereas a specialized T cell subset (namely, T_{FH} cells) provides help for B cells, no analogous helper subset for CD8⁺ T cells has been identified to date. Defining the conditions that lead to the generation of CD4⁺ T cells with potent CD8⁺ T cell helper activity could be important for developing better vaccines against several viral pathogens.

More than just lymphocyte helpers

In addition to activating cells of the innate immune system and providing potent help to promote the functions of B cells and CD8⁺ T cells during viral infection, CD4⁺ T cells develop into populations of effector T cells that migrate to sites of infection⁷⁶. Accumulating evidence suggests that effector CD4⁺ T cells have potent protective roles during viral infection that are independent of their helper activities. Strong immune protection mediated by CD4⁺ T cells has been described in animal models of infection by rotavirus⁷⁷, Sendai virus^{78,79}, gamma-herpesviruses^{80,81}, West Nile virus (WNV)⁸², HSV^{83,84}, influenza virus^{85,86}, dengue virus⁸⁷, Venezuelan equine encephalitis virus⁸⁸, coronavirus⁸⁹, VSV⁹⁰ and Friend virus^{91,92}. In many cases, the direct protective mechanism used by CD4⁺ T cells has not been defined. However, effector CD4⁺ T cells have, in general, been shown to protect against viral pathogens through two distinct mechanisms: first, through the production of cytokines, most notably IFN γ and TNF^{81,82,85,87,89,90,92}; and second,

through direct cytolytic activity^{78,81,82,87,92} mediated by both perforin and FAS (also known as CD95)^{84,86} (FIG. 3). Cytotoxic CD4⁺ T cells have also been observed following infection with LCMV⁹³.

The cytotoxic activity of CD4⁺ T cell effectors does not depend on T_H1 cell polarization⁹⁴, and expression of the transcription factor eomesodermin, but not T-bet, may be crucial in driving the development of cytotoxic CD4⁺ T cells *in vivo*⁹⁵. Thus, CD4⁺ T cells with cytotoxic activity could be considered a separate functional T cell subset. The fact that MHC class II expression is largely restricted to professional APCs under steady-state conditions may limit the protective potential of virus-specific cytotoxic CD4⁺ T cells. However, cells other than professional APCs are capable of upregulating MHC class II expression following pathogen challenge and could therefore become targets of cytotoxic CD4⁺ T cells during viral infection. For example, epithelial cells that are activated by infection⁹⁶ or IFN γ -mediated signals⁹⁷ strongly upregulate their expression of MHC class II molecules.

Although immune protection mediated by cytotoxic CD4⁺ T cells requires direct recognition of virus-infected cells, soluble factors released by other effector CD4⁺ T cells can act more broadly. For example, IFN γ can promote the establishment of an antiviral state in surrounding tissue and can also activate several innate immune cell populations, most notably macrophages, to mediate antiviral activity⁹⁸. Thus, effector CD4⁺ T cells that migrate to infected tissues probably promote viral clearance through both cytotoxic and cytokine-dependent mechanisms.

Immunoregulation by effector CD4⁺ T cells

Effector CD4⁺ T cells are capable of potent immunoregulation at sites of infection. A subset of T_H1 cells has been found to transiently produce both IFN γ and IL-10 at the peak of the effector CD4⁺ T cell response in many models of infectious disease⁹⁹. Several signals have been found to stimulate the generation of IL-10-producing effector CD4⁺ T cells, including high levels of antigen, soluble factors such as IL-12 and IL-27, and co-stimulatory signals such as ICOS (reviewed in REF. 99). IL-10 is a pleiotropic cytokine that is most often associated with anti-inflammatory or inhibitory functions (FIG. 3). The impact of IL-10 production by effector CD4⁺ T cells is complicated and can be variable, especially in situations in which strong immune responses (which are capable of serious immunopathology) are required to eliminate a rapidly replicating pathogen. For instance, the absence of IL-10 during influenza virus infection can lead, on the one hand, to improved host survival, owing to enhanced T cell¹⁶ and antibody¹⁰⁰ responses, but also, on the other hand, to exacerbated inflammation and immunopathology, which result in increased mortality¹⁰¹. Similarly, ablating *Il10* has been found to enhance protection against WNV¹⁰² and vaccinia virus¹⁰³, but to cause increased pathology following infection with RSV¹⁰⁴ and death following infection with mouse hepatitis virus¹⁰⁵ or coronavirus¹⁰⁶. Finally, as well as producing IL-10 themselves, effector CD4⁺ T cells can also promote IL-10 production by effector CD8⁺ T cells during viral infection¹⁰⁷.

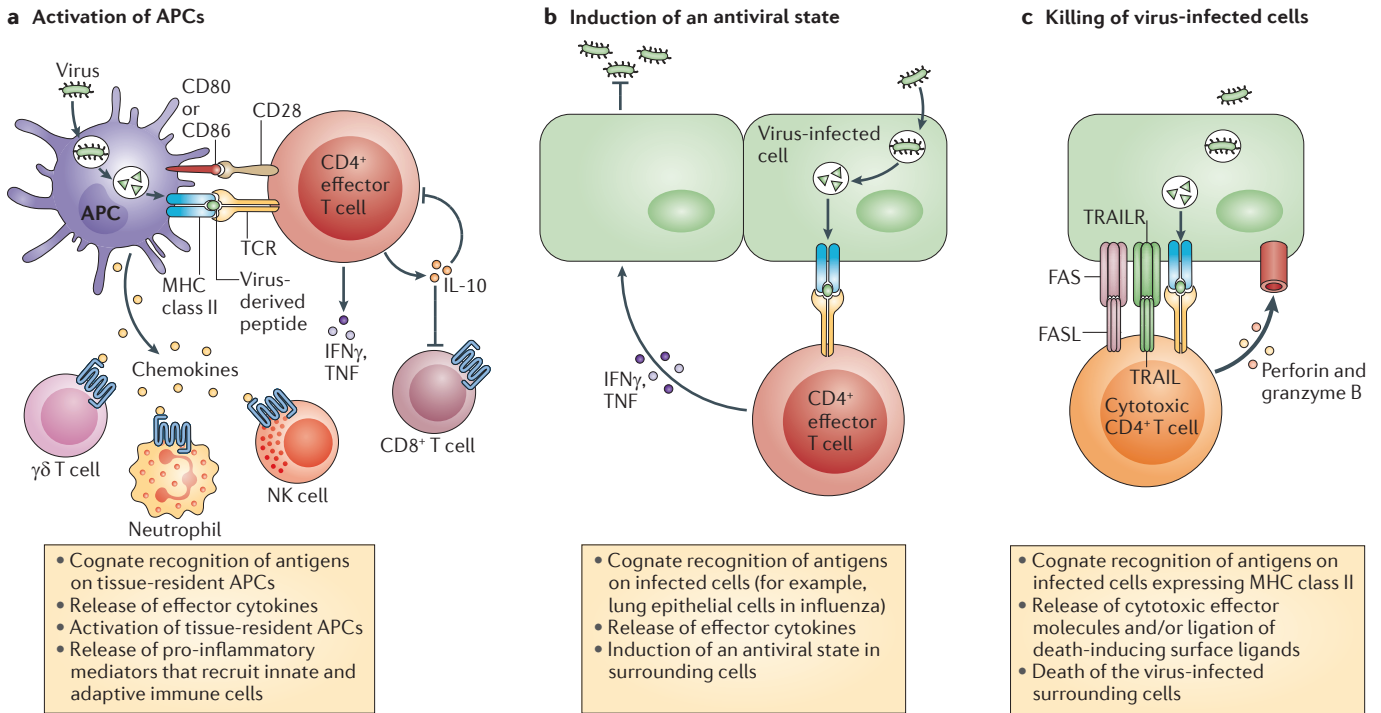


Figure 3 | Antiviral functions of CD4⁺ T cells that are independent of their lymphocyte helper functions.
a | After migrating to sites of infection, effector CD4⁺ T cells that recognize antigens on antigen-presenting cells (APCs) produce an array of effector cytokines that contribute to the character of the inflammatory responses in the tissue. Some products of highly activated effector CD4⁺ T cells, such as interleukin-10 (IL-10), dampen inflammation and regulate immunopathology, whereas others, such as interferon- γ (IFN γ), are pro-inflammatory and activate macrophages, which in turn drive further inflammation. The production of IL-10 by effector CD4⁺ T cells can have a profound impact on the outcome of a viral infection. **b** | IFN γ , tumour necrosis factor (TNF) and other cytokines produced by CD4⁺ T cells help to coordinate an antiviral state in infected tissues. **c** | Cytotoxic CD4⁺ T cells can directly lyse infected cells through diverse mechanisms, including FAS-dependent and perforin-dependent killing. FASL, FAS ligand; NK, natural killer; TCR, T cell receptor; TRAIL, TNF-related apoptosis-inducing ligand; TRAILR, TRAIL receptor.

Regulatory CD4⁺ T cells

Increased frequencies of CD4⁺ T_{Reg} cells (of both the forkhead box P3 (FOXP3)⁺ and FOXP3⁻ populations) have been observed in numerous human and animal studies of viral infection. Most studies that have assessed the impact of T_{Reg} cells during viral infection have concentrated on models of chronic infection. In such settings, T_{Reg} cells have been found, depending on the particular pathogen, to have both beneficial roles, such as limiting collateral tissue damage, and detrimental roles, including diminishing the overall magnitude of antiviral immune responses¹⁰⁸. The antigen specificity of T_{Reg} cells in the context of infectious disease has not been addressed often, but it is likely that T_{Reg} cell populations contain at least some virus-specific cells and that these populations are generated in concert with antiviral effector T cell responses. Indeed, this has recently been shown in a coronavirus infection model¹⁰⁹. The accumulation of virus-specific FOXP3⁺ T_{Reg} cells during viral infection could be due to the expansion of pre-existing populations of thymus-derived ‘natural’ T_{Reg} cells that are specific for viral antigens, or could also reflect the *de novo* generation of ‘induced’ T_{Reg} cells from naive virus-specific CD4⁺ T cells. An important factor in the generation of induced T_{Reg} cells appears to be TGF β ^{110,111},

although how induced T_{Reg} cells affect viral infection is not yet well understood. For example, some viral pathogens may promote the development of T_{Reg} cell populations to limit the host antiviral response. Evidence for this hypothesis has been found in several models, mainly of chronic viral infection^{112,113}. In acute viral infection, which has not been studied as fully as chronic infection, T_{Reg} cells have been shown to limit pathology during infection with WNV¹¹⁴ or RSV^{115,116} and in a model of influenza A virus infection¹¹⁷. A detailed account of T_{Reg} cell induction and responses during viral infection is available elsewhere¹¹⁸.

CD4⁺ T cells and chronic viral infection

Compared with our understanding of CD8⁺ T cells¹¹⁹, much less is known about how chronic infection affects CD4⁺ T cell phenotype and function. However, the impact of persistent viral infection on CD4⁺ T cell function and the importance of CD4⁺ T cells during chronic viral infection are receiving increasing attention. During persistent infection with LCMV clone 13, responding CD4⁺ T cells lose the ability to produce T_H1-type effector cytokines and to function optimally following viral rechallenge¹²⁰. The loss of function of CD4⁺ T cells responding to persistent antigen is probably driven by

Box 2 | CD4⁺ T cell help for CD8⁺ T cells during chronic viral infection

The best-characterized function of CD4⁺ T cells during persistent viral infection is the maintenance of competent CD8⁺ T cells that retain robust effector functions long-term¹⁵³. Although this has been most-rigorously studied in models of lymphocytic choriomeningitis virus (LCMV) infection, CD4⁺ T cells have also been found to affect CD8⁺ T cell responses to varying degrees in other models of chronic infection, including mouse cytomegalovirus¹⁵⁴, mouse polyomavirus¹⁵⁵ and gammaherpesvirus¹⁵⁶ infection. Recent studies using LCMV infection suggest that interleukin-21 (IL-21) production by CD4⁺ T cells during chronic infection is crucial for maintaining functional CD8⁺ T cells that are able to contain the infection^{157–159}. Importantly, clinical evidence also correlates the presence of higher numbers of IL-21-producing CD4⁺ T cells with improved CD8⁺ T cell function and improved control of HIV infection^{160,161}. These observations suggest new avenues that could improve vaccination strategies and adoptive transfer therapies¹⁶² through the generation of CD4⁺ T cell populations specifically geared towards the provision of maximal help in the context of chronic infection.

high levels of antigen following the priming phase¹²¹ and seems not to be regulated by the intrinsic changes in APCs that are caused by chronic viral pathogens¹²⁰.

Chronic infection may not lead to irreversible exhaustion of responding CD4⁺ T cells. For example, functionally impaired CD4⁺ T cells have been observed in patients with HIV, and treatment with antibodies that stimulate CD28 (REF. 122), block T cell immunoglobulin domain and mucin domain protein 3 (TIM3)¹²³ or block PD1 signalling¹²⁴ dramatically increased the proliferative potential of these T cells *in vitro*. Also, a recent study by Fahey *et al.* found that during chronic LCMV infection responding CD4⁺ T cells progressively adopt a functional T_{HH} cell phenotype¹²⁵. That CD4⁺ T cells may retain specific functions during chronic infection helps to explain earlier observations that persistent LCMV infection is eventually cleared through mechanisms that are dependent on CD4⁺ T cells (BOX 2). For instance, interactions between exhausted CD8⁺ T cells and CD4⁺ T cells may restore CD8⁺ T cell function during chronic infection with LCMV¹²⁶. These findings and others (reviewed in REFS 127, 128) have important implications for the design of vaccines against viruses that cause chronic infection in humans (such as HIV, hepatitis B and hepatitis C).

Memory CD4⁺ T cell responses against viruses

Following the resolution of infection, or after successful vaccination, most virus-specific effector CD4⁺ T cells die. This leaves a small population of memory T cells, which ensures that the frequency of virus-specific T cells is greater than it was before priming. The population of CD4⁺ memory T cells diminishes with time and may require boosting. It is unclear which responding effector CD4⁺ T cells make the transition to a memory phenotype, but a recent study suggests that those with lower expression of LY6C and T-bet have a greater potential to do so¹²⁹. A quantitative gain in antigen-specific cells represents one important advantage of the memory state, but memory T cells also differ from naive T cells by broad functional criteria. Compared with naive T cells, memory CD4⁺ T cells respond much faster, respond to lower antigen doses, require less co-stimulation and proliferate more vigorously following pathogen challenge¹³⁰. In addition, subpopulations of memory CD4⁺ T cells have wider trafficking patterns and some are retained at or near sites of previous infection¹³¹, and this contributes to their ability to be rapidly activated following local re-infection. Tissue tropism and/or retention of tissue-resident memory CD4⁺ T cells may depend on specific interactions between adhesion molecules and their receptors (BOX 3).

Memory CD4⁺ T cells enhance early innate immune responses following viral infection. Memory CD4⁺ T cell-mediated recognition of antigens presented by APCs following re-infection has rapid consequences. Within 48 hours of intranasal infection with influenza virus, antigen-specific memory CD4⁺ T cells cause an enhanced inflammatory response in the lung, and this is characterized by the upregulation of a wide array of pro-inflammatory mediators¹³² (FIG. 4). In infected tissues, transferred T_H1-type and T_H17-type memory cells, as well as memory CD4⁺ T cells generated by previous infections, upregulate the expression of pro-inflammatory cytokines and chemokines, including

Box 3 | Tissue-resident memory T cells

Following the resolution of primary immune responses, most effector T cells die by apoptosis, leaving behind a small population of long-lived memory cells. Recent studies have demonstrated that, in addition to recirculating through lymphoid and non-lymphoid tissues, some memory cells reside at sites of infection. Although most of these studies have concentrated on CD8⁺ T cell memory, CD4⁺ T cells also appear to survive for long periods in peripheral tissues^{163,164}.

Often, the expression of distinct surface proteins distinguishes tissue-specific memory cells from conventional lymphoid memory populations. These molecules are likely to have a crucial role in the retention of memory T cells at different sites through specific interactions with ligands that are expressed in particular tissues. For example, the expression of $\alpha 1\beta 1$ integrin (also known as VLA1) by airway-resident memory CD4⁺ T cells in the lungs can facilitate binding to collagen¹⁶⁴, and high levels of CD103 expression by brain- or skin-resident memory CD8⁺ T cells facilitates their interaction with cells that express E-cadherin^{165,166}.

Tissue-resident memory T cells act as a first line of defence. Combined with the ability of memory T cells, but not naive T cells, to be activated through the recognition of antigens in peripheral tissues^{132,167}, the location of memory CD4⁺ T cell populations at potential sites of re-infection represents a powerful advantage of the memory state over the naive state, in which a lag of several days precedes the influx of antigen-specific cells into infected sites. Tissue-resident memory cells may facilitate more-rapid recruitment and activation of innate cell populations that are capable of controlling initial viral titres. Moreover, these memory cells may simultaneously accelerate the development of pathogen-specific effector populations of B and T cells by promoting the earlier activation of antigen-presenting cells. Elucidating the important cues that drive the development of long-lived tissue-resident memory cells is likely to become an important area of research, especially as this understanding may lead to the design of improved vaccination strategies.

IL-1 α , IL-1 β , IL-6, IL-12p40, CC-chemokine ligand 2 (CCL2), CXC-chemokine ligand 9 (CXCL9) and CXCL10. By contrast, T_H2-type and non-polarized (T_H0) memory T cells have a minimal impact on pulmonary inflammation following influenza virus challenge. To induce innate inflammatory responses, memory CD4⁺ T cells must recognize antigens on CD11c⁺ APCs. These APCs become activated during interactions with memory CD4⁺ T cells, and this results

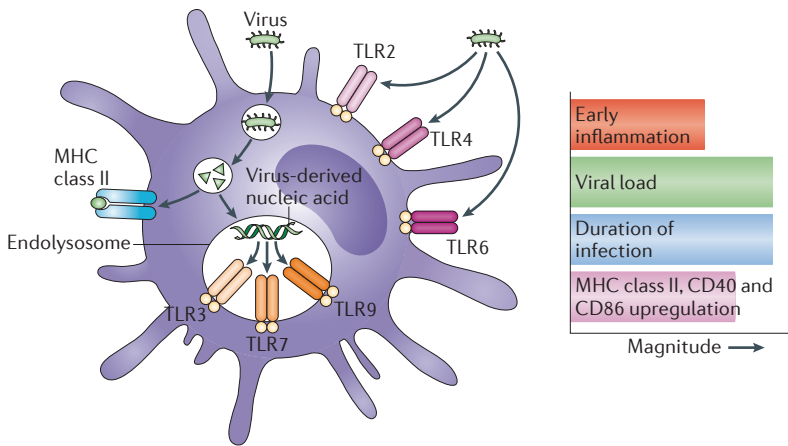
in the upregulation of MHC class II and co-stimulatory molecule expression by the APCs. Activated APCs contribute to the subsequent innate inflammatory response through the production of pro-inflammatory cytokines, such as IL-6 and IL-1 β . By contrast, the naive CD4⁺ T cell response *in vivo* does not have an appreciable impact on the tissue inflammatory response at 48 hours post-infection¹³². In other studies, naive CD4⁺ T cells could actually downregulate tissue inflammation following mouse hepatitis virus infection¹³³. The ability of T_H1-type and T_H17-type memory CD4⁺ T cells, but not T_H2-type or non-polarized memory T cells, to promote an early inflammatory response may help to explain why transfer of virus-specific T_H1 and T_H17 effector cells, but not transfer of T_H2 or non-polarized T cells, promotes early control of viral loads in mice infected with influenza virus^{16,86,132}. A similar impact of memory CD4⁺ T cells on early viral control during secondary influenza infection was recently reported by Chapman *et al.*¹³⁴, and this also correlated with a dramatic enhancement of innate immune responses against the virus. However, this effect has not yet been studied in other models of viral infection.

We suggest that the early activation of innate immune mechanisms by memory CD4⁺ T cells serves to lessen the ‘stealth phase’ of infection, during which titres of the virus are still too low to generate robust inflammatory responses¹³⁵. This would prevent viruses from gaining a ‘foothold’ in the host by infecting and replicating in cells at a level that is below immune detection. Importantly, the induction of early innate immune responses by memory CD4⁺ T cells does not require PRR activation¹³² and could therefore be important for enhancing protection against viral pathogens — such as vaccinia virus and influenza virus — that can actively antagonize key components of innate recognition pathways, such as dsRNA-dependent protein kinase (PKR)¹³⁶.

One intriguing finding is that memory CD4⁺ T cells specific for ovalbumin enhance antiviral immunity when ovalbumin is co-administered with an influenza virus that does not express ovalbumin¹³². Thus, we hypothesize that the induction of innate immune responses by memory CD4⁺ T cells could have an adjuvant effect, which could be exploited (and substituted for PRR stimulation¹³²) to promote immune responses to vaccines that do not contain live viruses. For example, a vaccine could activate the innate immune system through antigen-specific stimulation of memory CD4⁺ T cells that are known to be widespread in the human population (for example, most individuals have memory CD4⁺ T cells specific for tetanus toxin). This method could possibly enhance the immunogenicity of ‘weak’ vaccines.

Heterologous memory responses. Heterologous viral immunity occurs when memory T cells that were generated in response to a particular virus cross-react with epitopes expressed by other, unrelated viruses. It has recently become clear that this is quite a widespread phenomenon that is likely to be important in the human population, as humans are exposed to numerous antigens as a result of infection and vaccination¹³⁷.

a PRR-driven antiviral response



b Memory CD4⁺ T cell-driven antiviral response

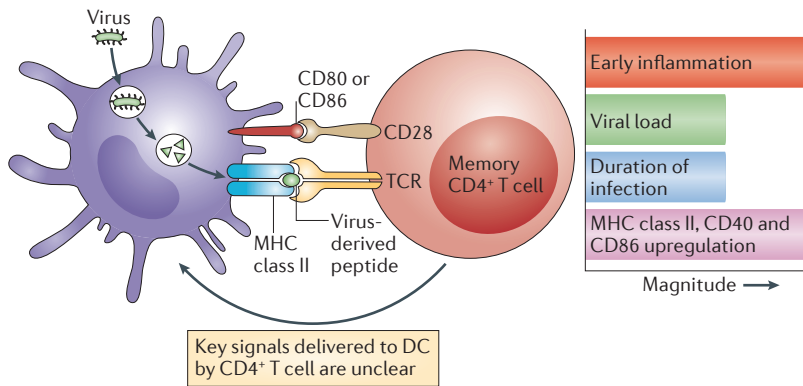


Figure 4 | Activation of APCs through PRRs and through the recognition of antigens by memory CD4⁺ T cells.

a | Several classes of pattern-recognition receptors (PRRs), such as Toll-like receptors (TLRs), sense the presence of viral pathogens, and the triggering of these receptors leads to the activation of antigen-presenting cells (APCs), including dendritic cells (DCs). Activated APCs upregulate their expression of MHC molecules and co-stimulatory molecules, which are important for the priming of naive virus-specific T cells. Triggering of PRRs at the site of infection induces local inflammation, which involves the activation of several populations of innate immune cells that can control viral titres and establish chemokine gradients to attract further antiviral effector cells. The efficiency of PRR triggering can determine whether viral replication or protective immunity gains the upper hand. **b** | Virus-specific memory CD4⁺ T cells can directly activate DCs through the recognition of antigens presented by MHC class II molecules, even in the absence of co-stimulation delivered via PRR-mediated signalling. The crucial signals delivered by memory CD4⁺ T cells to DCs in this process are unclear and could involve both cell-surface interactions and cytokine signals. The outcome of DC activation and the initiation of inflammatory responses are similar whether triggered through PRRs or memory CD4⁺ T cells but, in situations of infection with a rapidly replicating virus (such as influenza virus), memory CD4⁺ T cell-mediated enhancement of innate immunity is substantially quicker and more effective than that provided by PRR triggering. TCR, T cell receptor.

Although heterologous immune responses can be protective, they may in some cases be deleterious and can result in dramatic immunopathology¹³⁸. It will be important to determine to what extent the ability of memory CD4⁺ T cells to induce innate immunity contributes to both beneficial and deleterious heterologous responses.

Helper functions of memory CD4⁺ T cells. Several studies indicate that memory CD4⁺ T cells are superior to naive T cells in providing help for B cells, and they have been shown to promote earlier B cell proliferation, higher antibody levels and earlier class-switching responses compared with naive CD4⁺ T cells^{139–141}. In many cases, when pre-existing circulating antibodies are able to recognize the virus, re-infection may never occur. Faster antibody production could be particularly important after re-infection with rapidly mutating viruses (such as influenza virus), as the generation of neutralizing antibodies specific for new variants that evade previously generated antibodies could be necessary for immunity.

That memory CD4⁺ T cells are superior helpers for B cells compared with naive T cells is suggested by multiple criteria, and many mechanisms may contribute. Memory CD4⁺ T cells contain preformed stores of CD40L, an important signal in CD4⁺ T cell-mediated help for antibody production¹⁴². The location of memory CD4⁺ T cells may also be an advantage. For example, antigen-specific memory CD4⁺ T cells with a T_{HH} cell phenotype are retained in the draining lymph nodes of mice for over 6 months following immunization¹⁴³. The increased levels and polarized profile of cytokines produced by memory CD4⁺ T cells, as compared with those of naive T cells, is likely to promote a more-robust B cell antibody response and to dictate the antibody isotype, which has a key role in the efficacy of antibodies specific for viruses such as Ebola virus¹⁴⁴, WNV¹⁴⁵ and influenza virus¹⁴⁶. Recent observations suggest that IL-4 and IFN γ produced by T_{HH} cells have a central role in driving not only immunoglobulin class switching in the germinal centre, but also B cell affinity maturation¹⁴⁷.

Whether memory CD4⁺ T cells are superior to naive CD4⁺ T cells at providing help for primary or secondary CD8⁺ T cell responses during viral infection has not been rigorously tested. However, the features of memory cells described here suggest that memory CD4⁺ T cells could promote an accelerated response by naive CD8⁺ T cells, both through more-rapid licensing of APCs and through faster and more-robust production of IL-2 and possibly other cytokines; these possibilities need to be explored. In support of this concept, a study using *Listeria monocytogenes* found that T_{HH}1-type but not T_{HH}2-type memory CD4⁺ T cells enhanced primary CD8⁺ T cell responses in terms of both magnitude and cytokine production, although the mechanism of memory CD4⁺ T cell-mediated help was not determined¹⁴⁸. Whatever the pathways involved, the fact that memory CD4⁺ T cells enhance the generation of CD8⁺ T cell memory and that this process has a greater dependence

on CD4⁺ T cell-mediated help when PRR stimulation is limited suggests that CD4⁺ T cell-mediated help will be of particular importance in achieving optimal CD8⁺ T cell memory responses with vaccines that do not contain live, replication-competent viruses.

Finally, we should point out that the impact of memory CD4⁺ T cells on the early innate immune response may also affect subsequent antigen-specific immune responses against viruses, owing to alterations in the expression of chemokines. For example, by upregulating local production of CC-chemokine receptor 5 (CCR5) ligands at the site of infection, memory CD4⁺ T cells can promote the recruitment of memory CD8⁺ T cells that contribute to early viral control in influenza virus infection¹⁴⁹. In HSV-2 infection, IFN γ expression by CD4⁺ T cells is required to induce chemokines that recruit effector CD8⁺ T cells to the infected vaginal tissue¹⁵⁰. Similar roles for CD4⁺ T cells in promoting CD8⁺ T cell recruitment may occur during infections with other viruses, although it is often difficult to separate the roles of CD4⁺ T cells in generating effector CD8⁺ T cells from their roles in modifying CD8⁺ T cell trafficking^{39,85,151}.

Secondary effector T cells

The presence of memory CD4⁺ T cells that are capable of producing multiple cytokines has been correlated with superior protective capacity in numerous studies¹⁵², but how such cells achieve enhanced protection has not been addressed. One possibility is that 'secondary' effector CD4⁺ T cells that arise from memory precursors may be much more efficient in directly combating pathogens than primary effector CD4⁺ T cells that arise from naive cells. Our recent experiments that directly compared such primary and secondary effectors during influenza virus infection support such a view (T.M.S., K.K.M., L. M. Bradley and S.L.S., unpublished observations). We found that following adoptive transfer of either antigen-specific memory CD4⁺ T cells or an equal number of antigen-specific naive CD4⁺ T cell precursors, the secondary effector populations that developed from the memory CD4⁺ T cells showed greater expansion and contained higher frequencies of T cells that secrete multiple cytokines. Furthermore, although high levels of IL-10 were produced by primary effector CD4⁺ T cells during influenza virus infection, secondary effector cells produced much less IL-10. Thus, memory CD4⁺ T cells seem to give rise to secondary effector T cells that are distinct from and superior to the primary effectors derived from naive CD4⁺ T cells. We suggest that this results in more-protective recall responses, and we predict that further functions of secondary effectors will be discovered as additional comparative analyses are carried out.

Summary

Here, we have reviewed how CD4⁺ T cells contribute to protective immunity to viruses, during both primary and secondary infections. Several key principles emerge. Distinct CD4⁺ T cell subsets — including T_{HH}1 cells, T_{HH}17 cells, T_{HH} cells and CD4⁺ T cells with cytotoxic

functions — have important roles in the antiviral response. Key among these roles is the provision of help to B cells, but CD4⁺ T cells also contribute to the antiviral response by producing cytokines and chemokines, by enhancing CD8⁺ T cells responses and through direct cytotoxic effects on virus-infected cells.

Memory CD4⁺ T cells have additional protective functions compared with naive cells. They induce early innate inflammatory responses in the tissue that contribute to viral control. Importantly, memory CD4⁺ T cells provide more-rapid help to B cells, and probably to CD8⁺ T cells, thereby contributing to a faster and more-robust antiviral immune response. Finally, secondary effectors derived from memory CD4⁺ T cell precursors are likely to be more capable of mediating direct antiviral activity than primary effectors derived from naive CD4⁺ T cells.

The picture that emerges is one in which CD4⁺ T cells carry out an impressive variety of functions at different times and in different sites, and we suggest that the synergy of these distinct mechanisms can provide an extraordinarily high level of viral control. Depending on the particular pathogen and the level of infection, these mechanisms may often be redundant, but they are likely to each make key contributions following exposure to high doses of rapidly replicating viruses or to viruses that have evolved mechanisms to evade specific immune pathways. We point out that the study of the immune response to viral infections has revealed new mechanisms by which CD4⁺ T cells function to protect against pathogens, and we predict that additional mechanisms will be identified in the future. Thus, this area of study will lead to a better understanding of how vaccines can be designed to harness the power of CD4⁺ T cell memory.

1. Mosmann, T. R., Cherwinski, H., Bond, M. W., Giedlin, M. A. & Coffman, R. L. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J. Immunol.* **136**, 2348–2357 (1986).
2. Lund, J. M., Hsing, L., Pham, T. T. & Rudensky, A. Y. Coordination of early protective immunity to viral infection by regulatory T cells. *Science* **320**, 1220–1224 (2008).
3. Williams, M. A., Ravkov, E. V. & Bevan, M. J. Rapid culling of the CD4⁺ T cell repertoire in the transition from effector to memory. *Immunity* **28**, 533–545 (2008).
This work shows that the expansion of effector CD4⁺ T cell populations that contract to give rise to memory populations is affected by the strength of antigenic signals received during differentiation.
4. McKinstry, K. K., Strutt, T. M. & Swain, S. L. Regulation of CD4⁺ T-cell contraction during pathogen challenge. *Immunol. Rev.* **236**, 110–124 (2010).
5. Takeuchi, O. & Akira, S. Pattern recognition receptors and inflammation. *Cell* **140**, 805–820 (2010).
6. Iwasaki, A. & Medzhitov, R. Toll-like receptor control of the adaptive immune responses. *Nature Immunol.* **5**, 987–995 (2004).
7. Pulendran, B., Palucka, K. & Banchereau, J. Sensing pathogens and tuning immune responses. *Science* **293**, 253–256 (2001).
8. Constant, S. L. & Bottomly, K. Induction of Th1 and Th2 CD4⁺ T cell responses: the alternative approaches. *Annu. Rev. Immunol.* **15**, 297–322 (1997).
9. Pulendran, B. Modulating vaccine responses with dendritic cells and Toll-like receptors. *Immunol. Rev.* **199**, 227–250 (2004).
10. Cousens, L. P. *et al.* Two roads diverged: interferon α/β - and interleukin 12-mediated pathways in promoting T cell interferon γ responses during viral infection. *J. Exp. Med.* **189**, 1315–1328 (1999).
11. Schijns, V. E. *et al.* Mice lacking IL-12 develop polarized Th1 cells during viral infection. *J. Immunol.* **160**, 3958–3964 (1998).
12. Xing, Z., Zganiacz, A., Wang, J., Divangahi, M. & Nawaz, F. IL-12-independent Th1-type immune responses to respiratory viral infection: requirement of IL-18 for IFN- γ release in the lung but not for the differentiation of viral-reactive Th1-type lymphocytes. *J. Immunol.* **164**, 2575–2584 (2000).
13. Oxenius, A., Karrer, U., Zinkernagel, R. M. & Hengartner, H. IL-12 is not required for induction of type 1 cytokine responses in viral infections. *J. Immunol.* **162**, 965–973 (1999).
14. Hegazy, A. N. *et al.* Interferons direct Th2 cell reprogramming to generate a stable GATA-3⁺ Tbet⁺ cell subset with combined Th2 and Th1 cell functions. *Immunity* **32**, 116–128 (2010).
15. Lee, Y. K. *et al.* Late developmental plasticity in the T helper 17 lineage. *Immunity* **30**, 92–107 (2009).
In this study, the *in vivo* plasticity of the Th17 cell lineage is shown to be dependent on STAT4 and Tbet.
16. McKinstry, K. K. *et al.* IL-10 deficiency unleashes an influenza-specific Th17 response and enhances survival against high-dose challenge. *J. Immunol.* **182**, 7353–7363 (2009).
17. Mahon, B. P. *et al.* Poliovirus-specific CD4⁺ Th1 clones with both cytotoxic and helper activity mediate protective humoral immunity against a lethal poliovirus infection in transgenic mice expressing the human poliovirus receptor. *J. Exp. Med.* **181**, 1285–1292 (1995).
18. Maloy, K. J. *et al.* CD4⁺ T cell subsets during virus infection. Protective capacity depends on effector cytokine secretion and on migratory capability. *J. Exp. Med.* **191**, 2159–2170 (2000).
19. Coutelier, J. P., van der Logt, J. T., Heessen, F. W., Warnier, G. & Van Snick, J. IgG2a restriction of murine antibodies elicited by viral infections. *J. Exp. Med.* **165**, 64–69 (1987).
20. Graham, M. B., Braciale, V. L. & Braciale, T. J. Influenza virus-specific CD4⁺ T helper type 2 T lymphocytes do not promote recovery from experimental virus infection. *J. Exp. Med.* **180**, 1273–1282 (1994).
21. Moran, T. M., Isobe, H., Fernandez-Sesma, A. & Schulman, J. L. Interleukin-4 causes delayed virus clearance in influenza virus-infected mice. *J. Virol.* **70**, 5230–5235 (1996).
22. Alwan, W. H., Kozłowska, W. J. & Openshaw, P. J. Distinct types of lung disease caused by functional subsets of antiviral T cells. *J. Exp. Med.* **179**, 81–89 (1994).
23. Ikemoto, K., Pollard, R. B., Fukumoto, T., Morimatsu, M. & Suzuki, F. Small amounts of exogenous IL-4 increase the severity of encephalitis induced in mice by the intranasal infection of herpes simplex virus type 1. *J. Immunol.* **155**, 1326–1333 (1995).
24. Matsui, M., Moriya, O., Yoshimoto, T. & Akatsuka, T. Tbet is required for protection against vaccinia virus infection. *J. Virol.* **79**, 12798–12806 (2005).
25. Arens, R. *et al.* Cutting edge: murine cytomegalovirus induces a polyfunctional CD4 T cell response. *J. Immunol.* **180**, 6472–6476 (2008).
26. Suryawanshi, A. *et al.* Role of IL-17 and Th17 cells in herpes simplex virus-induced corneal immunopathology. *J. Immunol.* **187**, 1919–1930 (2011).
27. Oyoshi, M. K. *et al.* Vaccinia virus inoculation in sites of allergic skin inflammation elicits a vigorous cutaneous IL-17 response. *Proc. Natl Acad. Sci. USA* **106**, 14954–14959 (2009).
28. Hou, W., Kang, H. S. & Kim, B. S. Th17 cells enhance viral persistence and inhibit T cell cytotoxicity in a model of chronic virus infection. *J. Exp. Med.* **206**, 313–328 (2009).
29. Crowe, C. R. *et al.* Critical role of IL-17RA in immunopathology of influenza infection. *J. Immunol.* **183**, 5301–5310 (2009).
30. Ye, P. *et al.* Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. *J. Exp. Med.* **194**, 519–527 (2001).
31. Tate, M. D. *et al.* Neutrophils ameliorate lung injury and the development of severe disease during influenza infection. *J. Immunol.* **183**, 7441–7450 (2009).
32. Sonnenberg, G. F., Fouser, L. A. & Artis, D. Border patrol: regulation of immunity, inflammation and tissue homeostasis at barrier surfaces by IL-22. *Nature Immunol.* **12**, 383–390 (2011).
33. Amanna, I. J., Carlson, N. E. & Slifka, M. K. Duration of humoral immunity to common viral and vaccine antigens. *N. Engl. J. Med.* **357**, 1903–1915 (2007).
34. Crotty, S. Follicular helper CD4 T cells (T_{fh}). *Annu. Rev. Immunol.* **29**, 621–663 (2011).
35. Fazilleau, N., Mark, L., McHeyzer-Williams, L. J. & McHeyzer-Williams, M. G. Follicular helper T cells: lineage and location. *Immunity* **30**, 324–335 (2009).
36. Crotty, S., Kersh, E. N., Cannons, J., Schwartzberg, P. L. & Ahmed, R. SAP is required for generating long-term humoral immunity. *Nature* **421**, 282–287 (2003).
37. McCausland, M. M. *et al.* SAP regulation of follicular helper CD4 T cell development and humoral immunity is independent of SLAM and Fyn kinase. *J. Immunol.* **178**, 817–828 (2007).
38. Kamperschroer, C., Dibble, J. P., Meents, D. L., Schwartzberg, P. L. & Swain, S. L. SAP is required for Th cell function and for immunity to influenza. *J. Immunol.* **177**, 5317–5327 (2006).
39. Chen, J. *et al.* Cellular immune responses to severe acute respiratory syndrome coronavirus (SARS-CoV) infection in senescent BALB/c mice: CD4⁺ T cells are important in control of SARS-CoV infection. *J. Virol.* **84**, 1289–1301 (2010).
40. Sette, A. *et al.* Selective CD4⁺ T cell help for antibody responses to a large viral pathogen: deterministic linkage of specificities. *Immunity* **28**, 847–858 (2008).
41. Liu, T. & Chambers, T. J. Yellow fever virus encephalitis: properties of the brain-associated T-cell response during virus clearance in normal and γ interferon-deficient mice and requirement for CD4⁺ lymphocytes. *J. Virol.* **75**, 2107–2118 (2001).
42. Thomsen, A. R. *et al.* Cooperation of B cells and T cells is required for survival of mice infected with vesicular stomatitis virus. *Int. Immunol.* **9**, 1757–1766 (1997).
43. Lu, K. T. *et al.* Functional and epigenetic studies reveal multistep differentiation and plasticity of *in vitro*-generated and *in vivo*-derived follicular T helper cells. *Immunity* **35**, 622–632 (2011).
44. Borrow, P. *et al.* CD40L-deficient mice show deficits in antiviral immunity and have an impaired memory CD8⁺ CTL response. *J. Exp. Med.* **183**, 2129–2142 (1996).
45. Edelman, K. H. & Wilson, C. B. Role of CD28/CD80–86 and CD40/CD154 costimulatory interactions in host defense to primary herpes simplex virus infection. *J. Virol.* **75**, 612–621 (2001).
46. Sangster, M. Y. *et al.* An early CD4⁺ T cell-dependent immunoglobulin A response to influenza infection in the absence of key cognate T–B interactions. *J. Exp. Med.* **198**, 1011–1021 (2003).

47. Bertram, E. M. *et al.* Role of ICOS versus CD28 in antiviral immunity. *Eur. J. Immunol.* **32**, 3376–3385 (2002).
48. Kopf, M. *et al.* OX40-deficient mice are defective in Th cell proliferation but are competent in generating B cell and CTL responses after virus infection. *Immunity* **11**, 699–708 (1999).
49. Lee, B. O., Hartson, L. & Randall, T. D. CD40-deficient, influenza-specific CD8 memory T cells develop and function normally in a CD40-sufficient environment. *J. Exp. Med.* **198**, 1759–1764 (2003).
50. Johnson, S. *et al.* Selected Toll-like receptor ligands and viruses promote helper-independent cytotoxic T cell priming by upregulating CD40L on dendritic cells. *Immunity* **30**, 218–227 (2009).
51. Shedlock, D. J. & Shen, H. Requirement for CD4 T cell help in generating functional CD8 T cell memory. *Science* **300**, 337–339 (2003).
52. Sun, J. C. & Bevan, M. J. Defective CD8 T cell memory following acute infection without CD4 T cell help. *Science* **300**, 339–342 (2003).
- References 51 and 52 highlight the requirement for CD4⁺ T cell help during the priming of CD8⁺ T cells to generate functional memory cells.**
53. Hamilton-Williams, E. E. *et al.* Cutting edge: TLR ligands are not sufficient to break cross-tolerance to self-antigens. *J. Immunol.* **174**, 1159–1163 (2005).
54. Novy, P., Quigley, M., Huang, X. & Yang, Y. CD4 T cells are required for CD8 T cell survival during both primary and memory recall responses. *J. Immunol.* **179**, 8243–8251 (2007).
55. Smith, C. M. *et al.* Cognate CD4⁺ T cell licensing of dendritic cells in CD8⁺ T cell immunity. *Nature Immunol.* **5**, 1143–1148 (2004).
56. Riberdy, J. M., Christensen, J. P., Branum, K. & Doherty, P. C. Diminished primary and secondary influenza virus-specific CD8⁺ T-cell responses in CD4-depleted Ig⁺ mice. *J. Virol.* **74**, 9762–9765 (2000).
57. Tripp, R. A., Sarawar, S. R. & Doherty, P. C. Characteristics of the influenza virus-specific CD8⁺ T cell response in mice homozygous for disruption of the H-2IAb gene. *J. Immunol.* **155**, 2955–2959 (1995).
58. Belz, G. T., Wodarz, D., Diaz, G., Nowak, M. A. & Doherty, P. C. Compromised influenza virus-specific CD8⁺-T-cell memory in CD4⁺-T-cell-deficient mice. *J. Virol.* **76**, 12388–12393 (2002).
59. Wiesel, M., Kratky, W. & Oxenius, A. Type I IFN substitutes for T cell help during viral infections. *J. Immunol.* **186**, 754–763 (2011).
60. Castellino, F. *et al.* Chemokines enhance immunity by guiding naive CD8⁺ T cells to sites of CD4⁺ T cell–dendritic cell interaction. *Nature* **440**, 890–895 (2006).
- This work shows that chemokine-guided recruitment enables naive CD8⁺ T cells to locate cognate antigen-presenting cells and CD4⁺ T cells.**
61. Mueller, S. N. *et al.* CD4⁺ T cells can protect APC from CTL-mediated elimination. *J. Immunol.* **176**, 7379–7384 (2006).
62. Janssen, E. M. *et al.* CD4⁺ T cells are required for secondary expansion and memory in CD8⁺ T lymphocytes. *Nature* **421**, 852–856 (2003).
63. Northrop, J. K., Thomas, R. M., Wells, A. D. & Shen, H. Epigenetic remodeling of the IL-2 and IFN- γ loci in memory CD8 T cells is influenced by CD4 T cells. *J. Immunol.* **177**, 1062–1069 (2006).
64. Janssen, E. M. *et al.* CD4⁺ T cell help controls CD8⁺ T cell memory via TRAIL-mediated activation-induced cell death. *Nature* **434**, 88–93 (2005).
65. Badovinac, V. P., Messingham, K. A., Griffith, T. S. & Harty, J. T. TRAIL deficiency delays, but does not prevent, erosion in the quality of “helpless” memory CD8 T cells. *J. Immunol.* **177**, 999–1006 (2006).
66. Sacks, J. A. & Bevan, M. J. TRAIL deficiency does not rescue impaired CD8⁺ T cell memory generated in the absence of CD4⁺ T cell help. *J. Immunol.* **180**, 4570–4576 (2008).
67. Fuse, S. *et al.* Recall responses by helpless memory CD8⁺ T cells are restricted by the up-regulation of PD-1. *J. Immunol.* **182**, 4244–4254 (2009).
68. Intlekofer, A. M. *et al.* Requirement for T-bet in the aberrant differentiation of unhelped memory CD8⁺ T cells. *J. Exp. Med.* **204**, 2015–2021 (2007).
69. Oh, S. *et al.* IL-15 as a mediator of CD4⁺ help for CD8⁺ T cell longevity and avoidance of TRAIL-mediated apoptosis. *Proc. Natl Acad. Sci. USA* **105**, 5201–5206 (2008).
70. Williams, M. A., Tyznik, A. J. & Bevan, M. J. Interleukin-2 signals during priming are required for secondary expansion of CD8⁺ memory T cells. *Nature* **441**, 890–893 (2006).
- This work shows a previously unappreciated role for IL-2 during the programming of immune responses in shaping the development of CD8⁺ memory T cells capable of full secondary expansion.**
71. Obar, J. J. *et al.* CD4⁺ T cell regulation of CD25 expression controls development of short-lived effector CD8⁺ T cells in primary and secondary responses. *Proc. Natl Acad. Sci. USA* **107**, 193–198 (2010).
72. Barker, B. R., Gladstone, M. N., Gillard, G. O., Panas, M. W. & Letvin, N. L. Critical role for IL-21 in both primary and memory anti-viral CD8⁺ T-cell responses. *Eur. J. Immunol.* **40**, 3085–3096 (2010).
73. Bourgeois, C., Rocha, B. & Tanchot, C. A role for CD40 expression on CD8⁺ T cells in the generation of CD8⁺ T cell memory. *Science* **297**, 2060–2063 (2002).
74. Sun, J. C., Williams, M. A. & Bevan, M. J. CD4⁺ T cells are required for the maintenance, not programming, of memory CD8⁺ T cells after acute infection. *Nature Immunol.* **5**, 927–933 (2004).
75. Azadniv, M., Bowers, W. J., Topham, D. J. & Crispe, I. N. CD4⁺ T cell effects on CD8⁺ T cell location defined using bioluminescence. *PLoS ONE* **6**, e16222 (2011).
76. Roman, E. *et al.* CD4 effector T cell subsets in the response to influenza: heterogeneity, migration, and function. *J. Exp. Med.* **196**, 957–968 (2002).
77. Kushnir, N. *et al.* B2 but not B1 cells can contribute to CD4⁺ T-cell-mediated clearance of rotavirus in SCID mice. *J. Virol.* **75**, 5482–5490 (2001).
78. Hou, S., Doherty, P. C., Zijlstra, M., Jaenisch, R. & Katz, J. M. Delayed clearance of Sendai virus in mice lacking class I MHC-restricted CD8⁺ T cells. *J. Immunol.* **149**, 1319–1325 (1992).
79. Hogan, R. J. *et al.* Protection from respiratory virus infections can be mediated by antigen-specific CD4⁺ T cells that persist in the lungs. *J. Exp. Med.* **193**, 981–986 (2001).
80. Sparks-Thissen, R. L., Braaten, D. C., Kreher, S., Speck, S. H. & Virgin, H. W. An optimized CD4 T-cell response can control productive and latent gammaherpesvirus infection. *J. Virol.* **78**, 6827–6835 (2004).
81. Stuller, K. A., Cush, S. S. & Flano, E. Persistent γ -herpesvirus infection induces a CD4 T cell response containing functionally distinct effector populations. *J. Immunol.* **184**, 3850–3856 (2010).
82. Brien, J. D., Uhrlaub, J. L. & Nikolich-Zugich, J. West Nile virus-specific CD4 T cells exhibit direct antiviral cytokine secretion and cytotoxicity and are sufficient for antiviral protection. *J. Immunol.* **181**, 8568–8575 (2008).
83. Johnson, A. J., Chu, C. F. & Milligan, G. N. Effector CD4⁺ T-cell involvement in clearance of infectious herpes simplex virus type 1 from sensory ganglia and spinal cords. *J. Virol.* **82**, 9678–9688 (2008).
84. Ishikawa, T. *et al.* Protective role of Fas–FasL signaling in lethal infection with herpes simplex virus type 2 in mice. *J. Virol.* **83**, 11777–11783 (2009).
85. Teijaro, J. R., Verhoeven, D., Page, C. A., Turner, D. & Farber, D. L. Memory CD4 T cells direct protective responses to influenza virus in the lungs through helper-independent mechanisms. *J. Virol.* **84**, 9217–9226 (2010).
86. Brown, D. M., Dilzer, A. M., Meents, D. L. & Swain, S. L. CD4 T cell-mediated protection from lethal influenza: perforin and antibody-mediated mechanisms give a one-two punch. *J. Immunol.* **177**, 2888–2898 (2006).
87. Yauch, L. E. *et al.* CD4⁺ T cells are not required for the induction of dengue virus-specific CD8⁺ T cell or antibody responses but contribute to protection after vaccination. *J. Immunol.* **185**, 5405–5416 (2010).
88. Brooke, C. B., Deming, D. J., Whitmore, A. C., White, L. J. & Johnston, R. E. T cells facilitate recovery from Venezuelan equine encephalitis virus-induced encephalomyelitis in the absence of antibody. *J. Virol.* **84**, 4556–4568 (2010).
89. Savarin, C., Bergmann, C. C., Hinton, D. R., Ransohoff, R. M. & Stohman, S. A. Memory CD4⁺ T-cell-mediated protection from lethal coronavirus encephalomyelitis. *J. Virol.* **82**, 12432–12440 (2008).
90. Oxenius, A. *et al.* CD40–CD40 ligand interactions are critical in T–B cooperation but not for other anti-viral CD4⁺ T cell functions. *J. Exp. Med.* **183**, 2209–2218 (1996).
91. Pike, R. *et al.* Race between retroviral spread and CD4⁺ T-cell response determines the outcome of acute Friend virus infection. *J. Virol.* **83**, 11211–11222 (2009).
92. Iwashiro, M., Peterson, K., Messer, R. J., Stromnes, I. M. & Hasenkrug, K. J. CD4⁺ T cells and interferon in the long-term control of persistent Friend retrovirus infection. *J. Virol.* **75**, 52–60 (2001).
93. Jellison, E. R., Kim, S. K. & Welsh, R. M. Cutting edge: MHC class II-restricted killing *in vivo* during viral infection. *J. Immunol.* **174**, 614–618 (2005).
94. Brown, D. M., Kamperschroer, C., Dilzer, A. M., Roberts, D. M. & Swain, S. L. IL-2 and antigen dose differentially regulate perforin- and FasL-mediated cytolytic activity in antigen specific CD4⁺ T cells. *Cell. Immunol.* **257**, 69–79 (2009).
95. Qui, H. Z. *et al.* CD134 plus CD137 dual costimulation induces Eomesodermin in CD4 T cells to program cytotoxic Th1 differentiation. *J. Immunol.* **187**, 3555–3564 (2011).
96. Debbabi, H. *et al.* Primary type II alveolar epithelial cells present microbial antigens to antigen-specific CD4⁺ T cells. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **289**, L274–L279 (2005).
97. Cunningham, A. C., Zhang, J. G., Moy, J. V., Ali, S. & Kirby, J. A. A comparison of the antigen-presenting capabilities of class II MHC-expressing human lung epithelial and endothelial cells. *Immunology* **91**, 458–463 (1997).
98. Schroder, K., Hertzog, P. J., Ravasi, T. & Hume, D. A. Interferon- γ : an overview of signals, mechanisms and functions. *J. Leukoc. Biol.* **75**, 163–189 (2004).
99. Jankovic, D., Kugler, D. G. & Sher, A. IL-10 production by CD4⁺ effector T cells: a mechanism for self-regulation. *Mucosal Immunol.* **3**, 239–246 (2010).
100. Sun, K., Torres, L. & Metzger, D. W. A detrimental effect of interleukin-10 on protective pulmonary humoral immunity during primary influenza A virus infection. *J. Virol.* **84**, 5007–5014 (2010).
101. Sun, J., Madan, R., Karp, C. L. & Braciale, T. J. Effector T cells control lung inflammation during acute influenza virus infection by producing IL-10. *Nature Med.* **15**, 277–284 (2009).
102. Bai, F. *et al.* IL-10 signaling blockade controls murine West Nile virus infection. *PLoS Pathog.* **5**, e1000610 (2009).
103. van Den Broek, M. *et al.* IL-4 and IL-10 antagonize IL-12-mediated protection against acute vaccinia virus infection with a limited role of IFN- γ and nitric oxide synthase 2. *J. Immunol.* **164**, 371–378 (2000).
104. Weiss, K. A., Christiaansen, A. F., Fulton, R. B., Meyerholz, D. K. & Varga, S. M. Multiple CD4⁺ T cell subsets produce immunomodulatory IL-10 during respiratory syncytial virus infection. *J. Immunol.* **15 Aug 2011** (doi:10.4049/jimmunol.1100764).
105. Lin, M. T., Hinton, D. R., Parra, B., Stohman, S. A. & van der Veen, R. C. The role of IL-10 in mouse hepatitis virus-induced demyelinating encephalomyelitis. *Virology* **245**, 270–280 (1998).
106. Trandem, K., Zhao, J., Fleming, E. & Perlman, S. Highly activated cytotoxic CD8 T cells express protective IL-10 at the peak of coronavirus-induced encephalitis. *J. Immunol.* **186**, 3642–3652 (2011).
107. Sun, J., Dodd, H., Moser, E. K., Sharma, R. & Braciale, T. J. CD4⁺ T cell help and innate-derived IL-27 induce Blimp-1-dependent IL-10 production by antiviral CTLs. *Nature Immunol.* **12**, 327–334 (2011).
108. Rouse, B. T., Sarangi, P. P. & Suvas, S. Regulatory T cells in virus infections. *Immunol. Rev.* **212**, 272–286 (2006).
109. Zhao, J., Fett, C., Trandem, K., Fleming, E. & Perlman, S. IFN- γ and IL-10-expressing virus epitope-specific Foxp3⁺ T reg cells in the central nervous system during encephalomyelitis. *J. Exp. Med.* **208**, 1571–1577 (2011).
110. Curotto de Lafaille, M. A. & Lafaille, J. J. Natural and adaptive Foxp3⁺ regulatory T cells: more of the same or a division of labor? *Immunity* **30**, 626–635 (2009).
111. Amarnath, S., Dong, L., Li, J., Wu, Y. & Chen, W. Endogenous TGF- β activation by reactive oxygen species is key to Foxp3 induction in TCR-stimulated and HIV-1-infected human CD4⁺CD25⁺ T cells. *Retrovirology* **4**, 57 (2007).
112. Robertson, S. J. & Hasenkrug, K. J. The role of virus-induced regulatory T cells in immunopathology. *Springer Semin. Immunopathol.* **28**, 51–62 (2006).
113. Punkosdy, G. A. *et al.* Regulatory T-cell expansion during chronic viral infection is dependent on endogenous retroviral superantigens. *Proc. Natl Acad. Sci. USA* **108**, 3677–3682 (2011).
114. Lanteri, M. C. *et al.* Tregs control the development of symptomatic West Nile virus infection in humans and mice. *J. Clin. Invest.* **119**, 3266–3277 (2009).

115. Lee, D. C. *et al.* CD25⁺ natural regulatory T cells are critical in limiting innate and adaptive immunity and resolving disease following respiratory syncytial virus infection. *J. Virol.* **84**, 8790–8798 (2010).
116. Liu, J. *et al.* Epitope-specific regulatory CD4 T cells reduce virus-induced illness while preserving CD8 T-cell effector function at the site of infection. *J. Virol.* **84**, 10501–10509 (2010).
117. Antunes, I. & Kassiotis, G. Suppression of innate immune pathology by regulatory T cells during influenza A virus infection of immunodeficient mice. *J. Virol.* **84**, 12564–12575 (2010).
118. Belkaid, Y. & Tarbell, K. Regulatory T cells in the control of host–microorganism interactions. *Annu. Rev. Immunol.* **27**, 551–589 (2009).
119. Wherry, E. J. T cell exhaustion. *Nature Immunol.* **12**, 492–499 (2011).
120. Brooks, D. G., Teyton, L., Oldstone, M. B. & McGavern, D. B. Intrinsic functional dysregulation of CD4 T cells occurs rapidly following persistent viral infection. *J. Virol.* **79**, 10514–10527 (2005).
121. Han, S., Asoyan, A., Rabenstein, H., Nakano, N. & Obst, R. Role of antigen persistence and dose for CD4⁺ T-cell exhaustion and recovery. *Proc. Natl Acad. Sci. USA* **107**, 20453–20458 (2010).
122. McNeil, A. C. *et al.* High-level HIV-1 viremia suppresses viral antigen-specific CD4⁺ T cell proliferation. *Proc. Natl Acad. Sci. USA* **98**, 13878–13883 (2001).
123. Jones, R. B. *et al.* Tim-3 expression defines a novel population of dysfunctional T cells with highly elevated frequencies in progressive HIV-1 infection. *J. Exp. Med.* **205**, 2763–2779 (2008).
124. Day, C. L. *et al.* PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature* **443**, 350–354 (2006).
125. Fahey, L. M. *et al.* Viral persistence redirects CD4 T cell differentiation toward T follicular helper cells. *J. Exp. Med.* **208**, 987–999 (2011).
This study shows that prolonged T cell receptor stimulation during chronic infection progressively redirects CD4⁺ T cell development towards a T follicular helper cell phenotype.
126. Aubert, R. D. *et al.* Antigen-specific CD4 T-cell help rescues exhausted CD8 T cells during chronic viral infection. *Proc. Natl Acad. Sci. USA* **12** December 2011 (doi:10.1073/pnas.1118450109).
127. Frebel, H., Richter, K. & Oxenius, A. How chronic viral infections impact on antigen-specific T-cell responses. *Eur. J. Immunol.* **40**, 654–663 (2010).
128. Klenerman, P. & Hill, A. T cells and viral persistence: lessons from diverse infections. *Nature Immunol.* **6**, 873–879 (2005).
129. Marshall, H. D. *et al.* Differential expression of Ly6C and Tbet distinguish effector and memory Th1 CD4⁺ cell properties during viral infection. *Immunity* **35**, 633–646 (2011).
130. McKinstry, K. K., Strutt, T. M. & Swain, S. L. The effector to memory transition of CD4 T cells. *Immunol. Res.* **40**, 114–127 (2008).
131. Sallusto, F., Lenig, D., Forster, R., Lipp, M. & Lanzavecchia, A. Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. *Nature* **401**, 708–712 (1999).
This study was instrumental in the division of human memory T cells into distinct functional subsets based on CCR7 surface expression.
132. Strutt, T. M. *et al.* Memory CD4⁺ T cells induce innate responses independently of pathogen. *Nature Med.* **16**, 558–564 (2010).
133. Kim, K. D. *et al.* Adaptive immune cells temper initial innate responses. *Nature Med.* **13**, 1248–1252 (2007).
134. Chapman, T. J., Lambert, K. & Topham, D. J. Rapid reactivation of extralymphoid CD4 T cells during secondary infection. *PLoS ONE* **6**, e20493 (2011).
135. Moltedo, B. *et al.* Cutting edge: stealth influenza virus replication precedes the initiation of adaptive immunity. *J. Immunol.* **183**, 3569–3573 (2009).
136. Langland, J. O., Cameron, J. M., Heck, M. C., Jancovich, J. K. & Jacobs, B. L. Inhibition of PKR by RNA and DNA viruses. *Virus Res.* **119**, 100–110 (2006).
137. Welsh, R. M., Che, J. W., Brehm, M. A. & Selin, L. K. Heterologous immunity between viruses. *Immunol. Rev.* **235**, 244–266 (2010).
138. Selin, L. K. *et al.* Heterologous immunity: immunopathology, autoimmunity and protection during viral infections. *Autoimmunity* **44**, 328–347 (2011).
139. MacLeod, M. K. *et al.* Memory CD4 T cells that express CXCR5 provide accelerated help to B cells. *J. Immunol.* **186**, 2889–2896 (2011).
140. Bradley, L. M., Duncan, D. D., Yoshimoto, K. & Swain, S. L. Memory effectors: a potent, IL-4-secreting helper T cell population that develops *in vivo* after restimulation with antigen. *J. Immunol.* **150**, 3119–3130 (1993).
141. Croft, M. & Swain, S. L. Recently activated naive CD4 T cells can help resting B cells, and can produce sufficient autocrine IL-4 to drive differentiation to secretion of T helper 2-type cytokines. *J. Immunol.* **154**, 4269–4282 (1995).
142. Koguchi, Y., Thauland, T. J., Slifka, M. K. & Parker, D. C. Preformed CD40 ligand exists in secretory lysosomes in effector and memory CD4⁺ T cells and is quickly expressed on the cell surface in an antigen-specific manner. *Blood* **110**, 2520–2527 (2007).
143. Fazilleau, N. *et al.* Lymphoid reservoirs of antigen-specific memory T helper cells. *Nature Immunol.* **8**, 753–761 (2007).
144. Wilson, J. A. *et al.* Epitopes involved in antibody-mediated protection from Ebola virus. *Science* **287**, 1664–1666 (2000).
145. Hofmeister, Y. *et al.* Human IgG subclasses: *in vitro* neutralization of and *in vivo* protection against West Nile virus. *J. Virol.* **85**, 1896–1899 (2011).
146. Huber, V. C. *et al.* Distinct contributions of vaccine-induced immunoglobulin G1 (IgG1) and IgG2a antibodies to protective immunity against influenza. *Clin. Vaccine Immunol.* **13**, 981–990 (2006).
147. Reinhardt, R. L., Liang, H. E. & Locksley, R. M. Cytokine-secreting follicular T cells shape the antibody repertoire. *Nature Immunol.* **10**, 385–393 (2009).
148. Krawczyk, C. M., Shen, H. & Pearce, E. J. Memory CD4 T cells enhance primary CD8 T-cell responses. *Infect. Immun.* **75**, 3556–3560 (2007).
149. Kohlmeier, J. E. *et al.* The chemokine receptor CCR5 plays a key role in the early memory CD8⁺ T cell response to respiratory virus infections. *Immunity* **29**, 101–113 (2008).
150. Nakanishi, Y., Lu, B., Gerard, C. & Iwasaki, A. CD8⁺ T lymphocyte mobilization to virus-infected tissue requires CD4⁺ T-cell help. *Nature* **462**, 510–513 (2009).
The work reveals a previously unappreciated function of CD4⁺ T cells in guiding effector CTL migration into otherwise restricted peripheral sites of infection.
151. Stohlman, S. A., Bergmann, C. C., Lin, M. T., Cua, D. J. & Hinton, D. R. CTL effector function within the central nervous system requires CD4⁺ T cells. *J. Immunol.* **160**, 2896–2904 (1998).
152. Seder, R. A., Darrah, P. A. & Roederer, M. T-cell quality in memory and protection: implications for vaccine design. *Nature Rev. Immunol.* **8**, 247–258 (2008).
153. Zajac, A. J. *et al.* Viral immune evasion due to persistence of activated T cells without effector function. *J. Exp. Med.* **188**, 2205–2213 (1998).
154. Snyder, C. M. *et al.* CD4⁺ T cell help has an epitope-dependent impact on CD8⁺ T cell memory inflation during murine cytomegalovirus infection. *J. Immunol.* **183**, 3932–3941 (2009).
155. Kemball, C. C. *et al.* The antiviral CD8⁺ T cell response is differentially dependent on CD4⁺ T cell help over the course of persistent infection. *J. Immunol.* **179**, 1113–1121 (2007).
156. Cardin, R. D., Brooks, J. W., Sarawar, S. R. & Doherty, P. C. Progressive loss of CD8⁺ T cell-mediated control of a γ -herpesvirus in the absence of CD4⁺ T cells. *J. Exp. Med.* **184**, 863–871 (1996).
157. Yi, J. S., Du, M. & Zajac, A. J. A vital role for interleukin-21 in the control of a chronic viral infection. *Science* **324**, 1572–1576 (2009).
158. Frohlich, A. *et al.* IL-21R on T cells is critical for sustained functionality and control of chronic viral infection. *Science* **324**, 1576–1580 (2009).
159. Elsaesser, H., Sauer, K. & Brooks, D. G. IL-21 is required to control chronic viral infection. *Science* **324**, 1569–1572 (2009).
160. Williams, L. D. *et al.* Interleukin-21-producing HIV-1-specific CD8 T cells are preferentially seen in elite controllers. *J. Virol.* **85**, 2316–2324 (2011).
161. Chevalier, M. F. *et al.* HIV-1-specific interleukin-21⁺ CD4⁺ T cell responses contribute to durable viral control through the modulation of HIV-specific CD8⁺ T cell function. *J. Virol.* **85**, 733–741 (2011).
162. Hunziker, L., Klenerman, P., Zinkernagel, R. M. & Ehl, S. Exhaustion of cytotoxic T cells during adoptive immunotherapy of virus carrier mice can be prevented by B cells or CD4⁺ T cells. *Eur. J. Immunol.* **32**, 374–382 (2002).
163. Purwar, R. *et al.* Resident memory T cells (T_{RM}) are abundant in human lung: diversity, function, and antigen specificity. *PLoS ONE* **6**, e16245 (2011).
164. Chapman, T. J. & Topham, D. J. Identification of a unique population of tissue-memory CD4⁺ T cells in the airways after influenza infection that is dependent on the integrin VLA-1. *J. Immunol.* **184**, 3841–3849 (2010).
165. Wakim, L. M., Woodward-Davis, A. & Bevan, M. J. Memory T cells persisting within the brain after local infection show functional adaptations to their tissue of residence. *Proc. Natl Acad. Sci. USA* **107**, 17872–17879 (2010).
166. Gebhardt, T. *et al.* Memory T cells in nonlymphoid tissue that provide enhanced local immunity during infection with herpes simplex virus. *Nature Immunol.* **10**, 524–530 (2009).
167. Wakim, L. M., Waithman, J., van Rooijen, N., Heath, W. R. & Carbone, F. R. Dendritic cell-induced memory T cell activation in nonlymphoid tissues. *Science* **319**, 198–202 (2008).
This study provides evidence that not only effector CD8⁺ T cell functions but initiation of CD8⁺ memory T cell responses can occur within extra-lymphoid tissues through interaction with CD4⁺ T cells and recently recruited DCs.

Acknowledgements

We apologize to the many authors whose papers could not be cited owing to space limitations. We are grateful to R. W. Dutton for critical reading of the manuscript and insightful discussions. The authors are supported by grants from the US National Institutes of Health.

Competing interests statement

The authors declare no competing financial interests.

FURTHER INFORMATION

Susan L. Swain's homepage: <http://profiles.umassmed.edu/profiles/ProfileDetails.aspx?From=SE&Person=1360>

ALL LINKS ARE ACTIVE IN THE ONLINE PDF