

Study of the activation of the PI3K/Akt pathway by the motif of σA and σNS proteins of avian reovirus

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Abstract

The present study was conducted to determine whether avian reovirus (ARV) activates the phosphatidylinositol 3-kinase-dependent Akt (PI3K/Akt) pathway according to the PXXP or YXXXM motifs of σA and σNS proteins. Gene splicing by overlap extension PCR was used to change the PXXP or YXXXM motifs of the σA and σNS genes. Plasmid constructs that contain mutant σA and σNS genes were generated and transfected into Vero cells, and the expression levels of the corresponding genes were quantified according to immunofluorescence and Western blot analyses. The Akt phosphorylation (P-Akt) profile of the transfected Vero cells was examined by flow cytometry and Western blot. The results showed that the σA and σNS genes were expressed in the Vero cells, and P-Akt expression in the σA mutant groups (amino acids 110–114 and 114–117) was markedly decreased. The results indicated that the σA protein of ARV activates the PI3K/Akt pathway via the PXXP motif. The results of this study reveal the mechanisms by which ARV manipulates the cellular signal transduction pathways, which may provide new ideas for novel drug targets.

Keywords

Avian reovirus, PI3K/Akt pathway, σA protein, σNS protein, PXXP motif

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Introduction

Avian reovirus (ARV) is one of the most important avian viruses, causing clinical diseases in poultry worldwide and resulting in severe economic losses.^{1,2} ARV-affected flocks commonly suffer viral arthritis or tenosynovitis, runting-stunting syndrome (RSS), enteric disease, immunosuppression and malabsorption syndrome.^{2–5} In the early stages of ARV infection, ARV activates the phosphatidylinositol 3-kinase-dependent Akt (PI3K/Akt) signalling axis in Vero cells⁶ – a pathway that is associated with cell survival, proliferation, migration, differentiation and apoptosis.^{7,8}

PXXP or YXXXM/YXXM motifs are present in a number of viral and cellular proteins involved in PI3K signaling and form extended helices that bind to SH domains on the p85 subunit of PI3K.^{9–15} Therefore, as our previous research review¹⁶ showed that various amino acid sequence have PXXP/YXXXM/YXXM motifs, σ A, σ NS, μ A, μ B and μ NS of ARV were speculated to be involved in PI3K signalling. The results

showed that σA and σNS -expressing cells had higher P-Akt levels than the pcAGEN-expressing cells and that in the cells expressing other proteins (i.e. μA , μB and μNS), pretreatment with the PI3K inhibitor LY294002 inhibited Akt phosphorylation in σA - and σNS -expressing cells.¹⁶ These results indicate that the σA and σNS proteins can activate the PI3K/Akt pathway.

Mutant the PXXP motif of NS1 protein from Influenza A virus or mutant the YXXM motif of envelope protein of avian leukosis virus results in loss of PI3K/Akt pathway activation.^{15,17} According to amino

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Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us. sagepub.com/en-us/nam/open-access-at-sage). acid sequence analysis of the σA and σNS genes, the PXXP and YXXXM motifs are conserved between different ARV strains. The aim of the current study was to determine whether the σA and σNS proteins affect the activation of the PI3K/Akt pathway *in vivo* via the PXXP or YXXXM motifs. To accomplish this objective, we mutated the PXXP or YXXXM motifs of σA and σNS genes. Plasmid constructs containing mutant σA and σNS genes were generated and transfected into Vero cells, and the expression levels of the σA and σNS genes were quantified according to immunofluorescence and Western blot analysis. The Akt phosphorylation (P-Akt) profiles of the transfected Vero cells were examined by flow cytometry and Western blot analysis.

Materials and methods

Plasmids and primers

The plasmids σ A-pcAGEN and σ NS-pcAGEN were generated by our lab.¹⁶ Nine pairs of primers were designed to mutate the PXXP or YXXXM motifs of the σ A and σ NS genes (Table 1). The primer sequences are presented in Table 1. The red colours represent the mutant bases. The complete σ A gene (1248 bp) was amplified using the primers σ A-F (5'-gatgatctcgaggccaccatggcggtgccatatacgac-3') and σ A-R (5'-atcgcggcc gcttaggcggtaaaagtggctagaac-3'), and the complete σ NS gene (1101 bp) was amplified using the primers σ NS-F (5'-gatgatctcgaggccaccatggacaacccgtgcgtgtt-3') and σ NS-R (5'-atcgcggccgcttacgccatcctagctggagagac-3'). The underlined text indicates Kozak sequences, and the italicized text represents restriction sites (XhoI and NotI). All primers were synthesized by Takara (Dalian, PR China).

RNA extraction and RT-PCR amplification of the mutant σA and σNS genes

Genomic RNA was extracted from $200 \,\mu$ l of ARV using an EasyPure viral DNA/RNA kit (Transgen, Beijing, PR China) according to the protocol suggested by the manufacturer.

RT-PCR was performed using an RNA LA PCR kit (Takara). Gene splicing by overlap extension PCR was used to mutate σA and σNS genes. To amplify the σA -M1 gene (σA gene mutant 1), the PCR protocol consisted of three rounds of PCR amplification. The first PCR amplification used σA -F and σA -M1-R primers, with σA -pcAGEN as the template; the second PCR amplification used σA -M1-F and σA -R primers, with σA -pcAGEN as the template. The PCR products of these two amplifications were obtained via gel extraction. The third PCR amplification used σA -F and σA -R primers with the gel extraction purification product of the first and second PCR used as the template. The PCR product was then obtained via gel extraction.

We used the same method to amplify the other mutant genes: σ A-M2, σ A-M3, σ A-M4, σ A-M5, σ A-M6, σ NS-M1, σ NS-M2 and σ NS-M3.

Table 1. Primer se	quences
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Mutant gene	Primer name	Sequence (5'-3')	Amino acids	Amino acid sequence→mutated sequence
σΑ-ΜΙ	σ Α-ΜΙ- Ϝ	caacggctgaagcagcctatccaggt	55–58	PXXP→AXXA
	σA-MI-R	acctggataggctgcttcagccgttg		
σ Α-Μ2	σ Α-Μ2- Ε	atgttccaagagtctgcactccac	65–69	$YXXXM{\rightarrow}FXXXA$
	σ Α-Μ2- R	gtggagtgcagactcttggaacat		
σ Α-Μ3	σ Α-Μ3- Ε	atgctgccgctgccgcgtatcagccagc	110-114	PPXXP
	σ Α-Μ3- R	gctggctgatacgcggcggcaggggggggagcat		
σ Α-Μ4	σ Α-Μ4- Ε	atcctcccctgccgccgcgtatcaggcagc	114–117	PXXP→AXXA
	σ Α-Μ4- R	gctgcctgatacgcggcggcaggggggggggt		
σ Α-Μ5	σ Α-Μ5- Ε	acatggctgttgaggctgatg	200-203	PXXP→AXXA
	σ Α-Μ5- R	catcagcctcaacagccatgt		
σ Α-Μ6	σ Α-Μ6- Ε	atttcgatcagcaggcacgtgct	207–211	$YXXXM{\rightarrow}FXXXA$
	σ Α-Μ6- R	agcacgtgcctgctgatcgaaat		
σ NS-M I	σNS-MI-F	gctgcgagtggcgctattgact	159–162	PXXP→AXXA
	σNS-M1-R	agtcaatagcgccactcgcagc		
σ NS-M 2	σNS-M2-F	tccctttcatgcttgacgcagta	179–183	$YXXXM{\rightarrow}FXXXA$
	σ NS-M2-R	tactgcgtcaagcatgaaaggga		
σNS-M 3	σNS-M3-F	cggtcgtactcgtcttctt	333–336	PXXP→AXXA
	σNS-M3-R	aagaagacgagtacgaccg		

Recombinant plasmid construction

The mutant σA and σNS gene products were cloned into pMD18-T cloning vectors (Takara) according to the manufacturer's instructions. The constructed recombinant plasmids were designated of A-M1pMD18T, oA-M2-pMD18T, oA-M3-pMD18T, oA-M4-pMD18T, oA-M5-pMD18T, oA-M6-pMD18T, σNS-M1-pMD18T, σNS-M2-pMD18T and σNS-M3-pMD18T. The plasmids were digested with XhoI and NotI enzymes (Takara) and were then ligated into the corresponding sites of pcAGEN expression vectors before being transformed into competent Escherichia *coli* cells (DH5 α). Positive colonies, which were designated oA-M1-pcAGEN, oA-M2-pcAGEN, oA-M3pMD18T, σ A-M4-pcAGEN, σA-M5-pcAGEN, σ A-M6-pcAGEN, σ NS-M1-pcAGEN, σNS-M2pcAGEN and σ NS-M3-pcAGEN, were identified by PCR and double digestion and sequenced by Invitrogen (Guangzhou, PR China).

Expression of σA and σNS proteins

Plasmids (pcAGEN, σ A-pcAGEN, σ NS-pcAGEN, σA-M1-pcAGEN, σA-M2-pcAGEN, σA-M3pMD18T. σA-M4-pcAGEN. σA-M5-pcAGEN. σ A-M6-pcAGEN, σ NS-M1-pcAGEN, σNS-M2pcAGEN and ons-M3-pcAGEN) were extracted using a plasmid mini kit (Omega Bio-Tek, Norcross, GA). Vero cells were seeded onto 6- or 24-well cell culture plates and were then transfected with 2.5 or $0.5 \,\mu g$ appropriate expression vectors of the using Lipofectamine[®] 3000 transfection reagent (Invitrogen) according to the manufacturer's instructions.

Immunofluorescence and Western blot analysis to quantify protein expression

After the cells were seeded onto 24-well plates and allowed to adhere, Vero cells were transfected with various plasmids for 6 h and were then washed three times with PBS. The cells were then fixed with cold methanol for 10 min at room temperature, washed three times with PBS and blocked for 1 h in 10% normal goat serum (Abcam, Cambridge, UK) in PBS containing 0.5% Triton X-100 (Sigma–Aldrich, St Louis, MO). The cells were then incubated with primary Abs against ARV¹⁸ overnight at 4°C. Following three 5-min washes with PBST, the cells were incubated with fluorescently labelled secondary Abs (Alexa Fluor; Abcam) for 60 min at room temperature.

Vero cells seeded onto six-well plates were harvested, and their cell lysates were used for Western blot analysis to detect σA and σNS protein expression, as described by Xie et al.¹⁸ Proteins were visualized using an enhanced chemiluminescence reagent

Flow cytometry and Western blot analysis of P-Akt expression

For flow cytometry and Western blot analyses of P-Akt expression, Vero cells seeded onto six-well plates were harvested following transfection with the corresponding plasmids for 6 h. The cells were then subjected to flow cytometry and Western blot analysis to detect P-Akt expression.

For flow cytometry analysis, the transfected cells were detached from the culture plates via incubation with Accutase (Sigma-Aldrich) for 5 min. Then, the cells were washed with PBS and fixed and permeabilized by incubation with Cytofix/Cytoperm solution (BD Biosciences, Franklin Lakes, NJ) at 4°C for 20 min. Next, the cells were washed twice with FACS buffer (0.5% BSA, 0.01% sodium azide in DPBS) and then incubated at 4°C for 30 min with a rabbit Ab (Cell Signaling Technology, Danvers, MA) against P-Akt. After this incubation, the cells were washed twice with FACS buffer and then incubated at 4°C for 30 min with goat anti-rabbit IgG conjugated to Alexa Fluor[®] 488 (Abcam). After staining, the cells were washed twice with FACS buffer and re-suspended in FACS buffer for analysis. The samples were then analysed via flow cytometry (Beckman Coulter, Brea, CA).

Vero cells seeded onto six-well plates were harvested, and their cell lysates were subjected to Western blot analysis to detect P-Akt expression, as described by Wang et al.¹⁹

All flow cytometry and Western blot analyses were repeated three times.

Results

Recombinant plasmid construction

The recombinant plasmids σ A-M1-pcAGEN, σ A-M2-pcAGEN, σ A-M3-pMD18T, σ A-M4-pcAGEN, σ A-M5-pcAGEN, σ A-M6-pcAGEN, σ NS-M1-pcAGEN, σ NS-M2-pcAGEN and σ NS-M3-pcAGEN were first subjected to PCR amplification (Figure 1) and double digestion (Figure 2). Then, DNA sequencing was performed to ensure that the recombinant plasmids contained intact mutant σ A and σ NS genes, validating that the recombinant plasmids were successfully constructed.

σA and σNS protein expression

As shown by the immunofluorescence and Western blot analyses, the σA and σNS proteins were expressed in

Figure 1. Identification of the recombinant plasmids by PCR. Lane M: 100-bp DNA ladder; Lane 1: σ A-M1-pcAGEN; Lane 2: σ A-M2-pcAGEN; Lane 3: σ A-M3-pcAGEN; Lane 4: σ A-M4-pcAGEN; Lane 5: σ A-M5-pcAGEN; Lane 6: σ A-M6-pcAGEN; Lane 7: σ NS-M1-pcAGEN; Lane 8: σ NS-M2-pcAGEN; Lane 9: σ NS-M3-pcAGEN.



Figure 2. Identification of the recombinant plasmids by double digests. Lane M: Trans2K Plus II DNA Marker; Lane I: σ A-M1-pcAGEN; Lane 2: σ A-M2-pcAGEN; Lane 3: σ A-M3-pcAGEN; Lane 4: σ A-M4-pcAGEN; Lane 5: σ A-M5-pcAGEN; Lane 6: σ A-M6-pcAGEN; Lane 7: σ NS-M1-pcAGEN; Lane 8: σ NS-M2-pcAGEN; Lane 9: σ NS-M3-pcAGEN.

the Vero cells 6 h after transfection with the corresponding plasmids (σ A-pcAGEN, σ A-M1-pcAGEN, σ A-M2-pcAGEN, σ A-M3-pMD18T, σ A-M4-pcAGEN, σ A-M5-pcAGEN, σ A-M6-pcAGEN, σ NS-pcAGEN, σ NS-M1-pcAGEN, σ NS-M2-pcAGEN and σ NS-M3pcAGEN; Figures 3 and 4). Cells that were not transfected cells and transfected with pcAGEN served as negative controls cells (Figure 3a and b and Figure 4, lanes 1 and 2).

σA and σNS proteins activate the PI3K/Akt signalling pathway

Figure 5 shows that σ A-M3- and σ A-M4- expressing cells had the lowest levels of P-Akt compared to σ ApcAGEN, σ A-M1-pcAGEN, σ A-M2-pcAGEN, σ A-M5-pcAGEN and σ A-M6-pcAGEN. However, these cells had the same levels of P-Akt as negative control and pcAGEN-expressing cells. Figure 6 shows the same results as Figure 4. After mutation at amino acids 110–114 (PPXXP \rightarrow AAXXA) and 114–117 (PXXP \rightarrow AXXA), the σ A gene lost the capacity to increase P-Akt expression in Vero cells.

The σ NS-M1-, σ NS-M2- and σ NS-M3-expressing cells had similar P-Akt levels compared to σ NSpcAGEN, but these cells had increased levels of P-Akt expression compared to negative control cells and pcAGEN-expressing cells (data not shown). The mutations at amino acids 159–162 (PXXP \rightarrow AXXA), 179–183(YXXXM \rightarrow FXXXA) and 333–336 (PXXP \rightarrow AXXA) did not affect the capacity of the σ NS gene to increase P-Akt expression in Vero cells.

Discussion

The PI3K/Akt pathway is involved in numerous cellular processes, such as cell proliferation, differentiation and survival.^{7,8} PI3K is activated by the binding of autophosphorylated tyrosine kinase receptors or nonreceptor tyrosine kinases to the SH2 and SH3 domains of its regulatory/adaptor subunit p85.¹⁹ This binding is mediated by PXXP and YXXXM/YXXM motifs and activates the PI3K/Akt pathway.^{9–15} As reported by Shin,¹⁵ the Influenza A virus NS1 protein activates the PI3K/Akt pathway by direct interaction with the p85 subunit of PI3K (SH2-binding motif YXXXM and SH3-binding motif PXXP).

The S2-encoded protein σA , a component of the inner core shell, displays anti-IFN activity by preventing the activation of the dsRNA-dependent protein kinase PKR; this activity is likely linked to its capacity to bind and sequester dsRNA.^{20,21} The non-structural protein σNS is encoded by the ARV S4 genome segment. As a non-structural RNA-binding protein that accumulates in viral factories of ARV-infected cells, σNS is a likely candidate for playing key roles in RNA packaging and replication.²²

Our previous works showed that the σA and σNS proteins of ARV activate the PI3K/Akt pathway.¹⁶ According to amino acid sequence analysis of the σA and σNS genes, σA contains four PXXP and two YXXXM motifs, and σNS contains one YXXXM and two PXXP motifs. Mutation of the PXXP motif of NS1 protein from influenza A virus or mutation of the YXXM motif of envelope protein from avian leukosis virus will result in loss of PI3K/Akt pathway activation.^{15,17} We speculate that the mutant PXXP and YXXXM motifs for the σA and σNS genes of ARV may also affect the activation of the PI3K/Akt pathway.

Gene splicing by overlap extension PCR was used to change the PXXP or YXXXM motifs of the σA and σNS genes and to construct the mutant σA and σNS recombinant plasmids. The mutant σA and σNS



Figure 3. IFA analysis of the expression of recombinant plasmids in transfected Vero cells. (a) Negative control, (b) pcAGEN, (c) σA-M1-pcAGEN, (d) σA-M2-pcAGEN, (e) σA-M3-pcAGEN, (f) σA-M4-pcAGEN, (g) σA-M5-pcAGEN, (h) σA-M6-pcAGEN, (i) σA-pcAGEN, (j) σNS-M1-pcAGEN, (k) σNS-M2-pcAGEN, (l) σNS-M3-pcAGEN, (m) σNS-pcAGEN.



Figure 4. Western blot analysis of gene expression. Lane 1: Negative control; Lane 2: pcAGEN; Lane 3: σ NS-M1-pcAGEN; Lane 4: σ NS-M2-pcAGEN; Lane 5: σ NS-M3-pcAGEN; Lane 6: σ NS-pcAGEN; Lane 7: σ A-M1-pcAGEN; Lane 8: σ A-M2pcAGEN; Lane 9: σ A-M3-pcAGEN; Lane 10: σ A-M4-pcAGEN; Lane 11: σ A-M5-pcAGEN; Lane 12: σ A-M6-pcAGEN; Lane 13: σ A-pcAGEN.

recombinant plasmids were transfected into Vero cells. The P-Akt profile of the transfected Vero cells was examined by flow cytometry and Western blot analysis.

In this study, after amino acids 110–114 (PPXXP \rightarrow AAXXA) and 114–117 (PXXP \rightarrow AXXA) were

mutated, the σA gene lost its capacity to improve P-Akt expression in Vero cells, and the σA protein of ARV activated the PI3K/Akt pathway via the PXXP motif. PXXP motif of the σA protein of ARV is involved in PI3K signalling, and the mutant PXXP motif of σA affects PI3K/Akt pathway activation. These results are similar to those of a previous study.¹⁶ However, whether this activation was a result of the σA protein binding to the p85 subunit via interactions with PXXP motifs, as occurs in other viruses,^{11,15} requires further study.

We also demonstrated that the mutations at amino acids 159–162 (PXXP \rightarrow AXXA), 179–183 (YXXXM \rightarrow FXXXA) and 333–336 (PXXP \rightarrow AXXA) did not affect the capacity of the σ NS gene to increase P-Akt expression in Vero cells. The σ NS protein may activate the PI3K/Akt pathway via more than one motif, other motifs or σ NS interaction with the P85 subunit according to the middle protein. These hypotheses must be further studied.



Figure 5. Flow cytometry analysis of P-Akt expression levels. (a) Negative control, (b) pcAGEN, (c) σA-pcAGEN, (d) σA-M1-pcAGEN, (e) σA-M2-pcAGEN, (f) σA-M3-pcAGEN, (g) σA-M4-pcAGEN, (h) σA-M5-pcAGEN, (i) σA-M6-pcAGEN.



Figure 6. Western blot analysis of P-Akt expression levels. Lane 1: Negative control; Lane 2: pcAGEN; Lane 3: σ A-pcAGEN; Lane 4: σ A-M1-pcAGEN; Lane 5: σ A-M2-pcAGEN; Lane 6: σ A-M3pcAGEN; Lane 7: σ A-M4-pcAGEN; Lane 8: σ A-M5-pcAGEN; Lane 9: σ A-M6-pcAGEN.

Moreover, the relationship between the p110 catalytic subunit of PI3K and the PXXP or YXXXM/ YXXM motifs of the $\sigma A/\sigma NS$ genes will be further studied, and these research results will be published in the future.

Although the exact mechanisms by which PI3K/Akt regulates ARV replication and other biological functions of PI3K/Akt in virus infection remain uncharacterized, our study reveals the mechanism underlying PI3K/Akt activation and adds a novel aspect to the functions of the σA and σNS proteins of ARV.

Declaration of conflicting interests

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