### MITOGENOME ANNOUNCEMENT

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# The complete mitochondrial genome of Rhinolophus affinis himalayanus

Yuting Ding<sup>a</sup>, Wenli Chen<sup>b</sup> and Xiuguang Mao<sup>b</sup>

<sup>a</sup>Institute of Estuarine and Coastal Research, East China Normal University, Shanghai, China; <sup>b</sup>School of Ecological and Environmental Sciences, East China Normal University, Shanghai, China

#### ABSTRACT

Here we generated the complete mitochondrial genome of one subspecies of *R. affinis (R. affinis hima-layanus)* using next generation sequencing and Sanger sequencing. The length of the complete mitochondrial genome was 16,886 bp, containing 13 protein-coding genes, 22 tRNAs, 2 rRNAs, and a non-coding control region. A maximum-likelihood tree based on the 13 concatenated mitochondrial protein-coding genes of 16 *Rhinolophus* taxon and one outgroup *Hipposideros armiger* indicates that *R. affinis* shows a closer relationship with *R. sinicus* complex than with other taxa.

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The intermediate horseshoe bat, *Rhinolophus affinis*, is distributed throughout Southeast Asia and nine subspecies have been recognized (Csorba et al. 2019). There are three subspecies of *R. affinis* in China, *R. a. himalayanus*, *R. a. macrurus* and *R. a. hainanus* (Mao et al. 2010). Previous studies on this species have identified a hybrid zone between *R. a. himalayanus* and *R. a. macrurus* in the eastern region of China (Mao et al. 2013; Mao and Rossiter 2020). In this study we generated the complete mitochondrial genome of *R. a. himalayanus* (GenBank Accession number: MT845219).

An adult male of R. a. himalayanus was captured with nets from Anhui province, China (30°26′51.82″N, 118°26′54.16″E) and stored in 95% ethanol at -80°C laboratory freezer in East China Normal University (Voucher No. AH17). Our sampling procedure was approved by the National Animal Research Authority, East China Normal University (approval ID bf20190301). Muscle tissue was used to extract genomic DNA using DNeasy Blood & Tissue Kit (Tiangen, China). Then a DNA library was constructed with insert fragments of  $\sim$ 350bp and sequenced on the Illumina HiSeg 4000 sequencer (150 bp paired-end). A total of 688,763,636 reads were generated and processed using TRIMMOMATIC-0.36 (Bolger et al. 2014) with a sliding window of 4 bp, minimum average PHRED quality score of 20 and minimum reads length of 50 bp. Over six hundred million filtered reads were obtained and mapped to the complete mitochondrion genome of Rhinolophus sinicus sinicus (GenBank accession no. KP257597) using BWA v. 0.7.12-r1039 (Li and Durbin 2009) and SAMTOOL v. 1.9 (Li and Durbin 2009) was used to retrieve mapped mitochondrial reads of R. affinis. Then the retrieved reads were used to assemble mitochondrial genome by A5-miseg version 20160825 with default parameters (Coil et al. 2015). We amplified the noncoding control region (D-loop) using traditional Sanger sequencing. PCR primers have been described previously (Sun et al. 2009).

The complete mitochondrial genome of R. affinis is 16,886 bp in length and was annotated based on published mitochondrion genome of Rhinolophus sinicus sinicus. It includes 13 protein-coding genes, 22 tRNA genes, 2 rRNA genes and a non-coding control region (D-loop). Gene number and order are identical with other Chiroptera mitogenomes (e.g. Xing and Mao 2016; Wang et al. 2020). The overall base composition of the whole mitochondrial genome is 30.98% A, 25.13% T, 28.96% C and 14.92% G. All genes were initiated with ATN (nd1, cox1, cox2, atp8, atp6, cox3, nd4, nd6 and cytb with ATG; nd2, nd3 and nd5 with ATA) except for nd4l with GTG. Eight coding genes use TAN as the termination codons (cox1, atp8, atp6, nd3, nd4l, nd5, and nd6 with TAA; cox2 with TAG), and only one gene (cytb) is stopped with AGA. Incomplete stop codon (T-- or TA-) was found in nd1, nd2, cox3, and nd4.

A maximum-likelihood tree was constructed using RAxML v.8.2.11 (Stamatakis 2014) based on the 13 concatenated mitochondrial protein-coding genes of 16 *Rhinolophus* taxa and one outgroup *Hipposideros armiger* (Figure 1) with GTRGAMMA model and bootstraps 1000 setting. The ML tree indicates that *R. affinis* shows a closer relationship with *R. sinicus* complex (*R. sinicus* and *R. thomasi*) (Figure 1).

#### Disclosure statement

No potential conflict of interest was reported by the authors.

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CONTACT Wenli Chen 🐼 51193904033@stu.ecnu.edu.cn 😰 School of Ecological and Environmental Sciences, East China Normal University, Shanghai, China © 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

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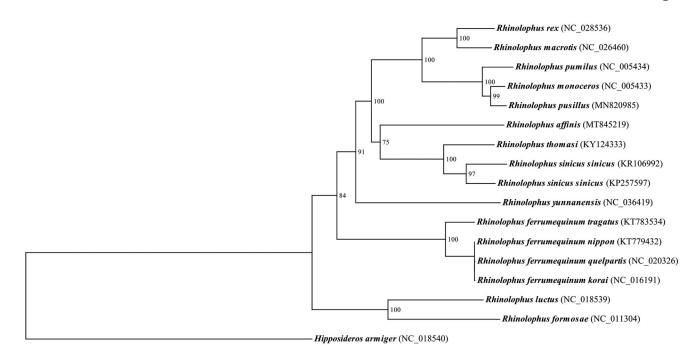


Figure 1. A maximum-likelihood tree reconstructed based on the 13 concatenated mitochondrial protein-coding genes from 17 bat species. *Hipposideros ammiger* was used as an outgroup. Numbers at the nodes indicated bootstrap support values. GenBank accession number for each bat mitogenome is indicated in bracket.

### Data availability statement

The data that support the findings of this study are openly available in NCBI at https://www.ncbi.nlm.nih.gov/, GenBank Accession number: MT845219.

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## References

- Bolger A, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics. 30(15):2114–2120.
- Coil D, Jospin G, Darling AE. 2015. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. Bioinformatics. 31(4):587–589.
- Csorba G, Hutson AM, Rossiter SJ, Burgin C. 2019. Family Rhinolophidae (Horseshoe Bats). In: Mittermeier RA, Wilson DE, editors. Handbook of the mammals of the World: 9. Chiroptera. Barcelona (Spain): Lynx Ediciones; p. 1–22.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. Bioinformatics. 25(14):1754–1760.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R. 2009. The sequence alignment/map format and SAMtools. Bioinformatics. 25(16):2078–2079.

- Mao X, He G, Hua P, Jones G, Zhang S, Rossiter SJ. 2013. Historical introgression and the persistence of ghost alleles in the intermediate horseshoe bat (*Rhinolophus affinis*). Mol Ecol. 22(4):1035–1050.
- Mao X, Rossiter SJ. 2020. Genome-wide data reveal discordant mitonuclear introgression in the intermediate horseshoe bat (*Rhinolophus affinis*). Mol Phylogenet Evol. 150:106886.
- Mao X, Zhu G, Zhang S, Rossiter SJ. 2010. Pleistocene climatic cycling drives intra-specific diversification in the intermediate horseshoe bat (Rhinolophus affinis) in Southern China . Mol Ecol. 19(13):2754–2769.
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 30(9): 1312–1313.
- Sun K, Feng J, Jin L, Liu Y, Shi L, Jiang T. 2009. Structure, DNA sequence variation and phylogenetic implications of the mitochondrial control region in horseshoe bats. Mamm Biol. 74(2):130–144.
- Xing Y, Mao X. 2016. The complete mitochondrial genome of the Thomas's horseshoe bat (*Rhinolophus thomasi*) using next-generation sequencing and Sanger sequencing. Mitochondrial DNA Part B. 1(1): 964–965.
- Wang J, Zhao A, Sun H. 2020. The complete mitochondrial genome of the least horseshoe bat (*Rhinolophus pusillus*). Mitochondrial DNA Part B. 5(1):881–882.