Research Article

Modulation of Colorectal Cancer Risk by Polymorphisms in 51Gln/His, 64Ile/Val, and 148Asp/Glu of APEX Gene; 23Gly/Ala of XPA Gene; and 689Ser/Arg of ERCC4 Gene

L. Dziki,¹ A. Dziki,¹ M. Mik,¹ I. Majsterek,² and J. Kabzinski²

¹Department of General and Colorectal Surgery, Medical University of Lodz, Lodz, Poland ²Department of Clinical Chemistry and Biochemistry, Medical University of Lodz, Lodz, Poland

Correspondence should be addressed to I. Majsterek; ireneusz.majsterek@umed.lodz.pl

Received 21 September 2016; Revised 1 December 2016; Accepted 7 December 2016; Published 12 March 2017

Academic Editor: Nicola Silvestris

Copyright © 2017 L. Dziki et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Polymorphisms in DNA repair genes may affect the activity of the BER (base excision repair) and NER (nucleotide excision repair) systems. Using DNA isolated from blood taken from patients (n = 312) and a control group (n = 320) with CRC, we have analyzed the polymorphisms of selected DNA repair genes and we have demonstrated that genotypes 51Gln/His and 148Asp/Glu of APEX gene and 23Gly/Ala of XPA gene may increase the risk of colorectal cancer. At the same time analyzing the gene-gene interactions, we suggest the thesis that the main factor to be considered when analyzing the impact of polymorphisms on the risk of malignant transformation should be intergenic interactions. Moreover, we are suggesting that some polymorphisms may have impact not only on the malignant transformation but also on the stage of the tumor.

1. Introduction

Currently, we are observing an increase of the incidence of colorectal cancer (CRC). In 2012, according to GLOBCAN, there were 1360000 new CRC cases, which with 9.7% made it the third most common cancer after lung and breast cancers [1, 2]. While causes of CRC remain unknown, it is estimated that about 20% of cancer cases are familial and approximately 3% are caused by mutations of strongly predisposed genes [3, 4]. Studies have shown that individual predispositions for developing this cancer may depend on genetic changes, including changes in genes involved in the process of DNA repair, which is responsible for dealing with DNA damages [5-7]. Several single-nucleotide polymorphisms (SNPs) have been associated with colorectal cancer susceptibility; most of them are part of mismatch DNA repair system (MMR) [8-10]. However, besides MMR system in mammalian cells, there are three more basic mechanisms of DNA repair: BER (base excision repair), NER (nucleotide excision repair), and DSB (double-strand brakes), which are currently under strong investigation in terms of connection with an increased risk of colorectal cancer [11-13].

In this paper, we study the selected polymorphisms of nucleotide excision repair (NER) and base excision repair (BER) pathways and their impact on modulating risk of colorectal cancer occurrence. Among the known polymorphisms of the DNA repair genes, the polymorphisms of ERCC4 and XPA genes from NER pathway have been repeatedly studied as potentially connected with susceptibility to the occurrence of various cancers [14-17]. NER is a particularly important excision mechanism that removes DNA damage induced by ultraviolet light (UV). UV DNA damage results in bulky DNA adducts-these adducts are mostly thymine dimers and 6,4-photoproducts. The importance of NER is evidenced by the severe human diseases that result from in-born genetic mutations of NER proteins such as xeroderma pigmentosum and Cockayne's syndrome [18, 19]. The second studied pathway-BER-is a DNA repair system that operates on small lesions such as oxidized or reduced bases. A single damaged base is removed by base-specific DNA glycosylases and apurinic/apyrimidinic sites are created (that can occur also by spontaneous hydrolysis or by DNA damaging agents). AP sites are premutagenic lesions that can prevent normal DNA replication and therefore need to be identified

TABLE 1: The refSNP and thermal conditions used in the PCR reaction.

Gene	APEX	APEX	APEX	XPA	ERCC4
Polymorphism	51Gln/His	64Ile/Val	148Asp/Glu	23Gly/Ala	689Ser/Arg
refSNP	rs1048945	rs2307486	rs1130409	rs1800975	rs149364215
Thermal conditions			 (1) 95°C, 10 min (2) 92°C, 15 sec (3) 60°C, 1 min (4) Steps 2&3, 45× 		
Dyes			ROX, HEX, FAM		
Ref dye	ROX				

His

and repaired. Whole process is initiated by the major AP endonuclease in human cells coded by *APEX* gene, whose polymorphisms have been so far connected to several types of cancer [20–22]. Moreover, our goal was to evaluate the mutual action of those two DNA repair systems on modulating CRC risk.

2. Materials and Methods

DNA for genotyping was isolated from lymphocytes of the peripheral blood. The blood samples were taken from 312 unrelated patients hospitalized in the Military Medical Academy University Teaching Hospital-Central Veterans' Hospital in Lodz. Each patient had histopathologically confirmed colorectal cancer. The studied group included 178 men and 134 women (average age 63 years ± 8 years). The stage of the tumors was established according to the TNM scale. The control group included 320 individuals not diagnosed with cancer and with ages corresponding to the age of the studied group (p < 0.05). Permission to conduct research was granted by the Bioethics Committee of the Medical University of Lodz.

DNA isolation was carried out with a commercial kit QIAamp DNA Blood Mini Kit for isolation of highmolecular weight DNA (Qiagen).

The occurrence of polymorphic variants in 51Gln/His, 64Ile/Val, and 148Asp/Glu of *APEX* gene; 23Gly/Ala of *XPA* gene; and 689Ser/Arg of *ERCC4* gene was studied with the TaqMan technique. Briefly, $25 \,\mu$ l of reaction mixture was used for analysis, containing $1 \,\mu$ l of genomic DNA solution, $1 \,\mu$ l of probes designed specifically for each polymorphism, $13 \,\mu$ l of premix with polymerase, and $10 \,\mu$ l of water. The PCR reaction was performed in a Stratagene Mx3005P real-time PCR thermocycler. The RS numbers for polymorphisms and thermal conditions of reaction are shown in Table 1. For 10% of the randomly selected samples, genotyping was repeated to confirm reproducibility. Cases and controls were genotyped randomly and researchers were blinded to the case/control status during genotyping.

3. Results

3.1. Genotyping. The genotyping results indicate that Gln/His genotype of 51Gln/His polymorphism of *APEX* gene (Table 2) increases the risk of CRC (OR 1.706 (1.174–2.480); p = 0.005). The same effect was observed in the case of

control group.				
Genotype/allele	Patients $(n = 311)$	Controls $(n = 302^*)$	OR (95% CI)	p
Gln/Gln	69	92	1 (ref)	_
Gln/His	206	161	1.706 (1.174–2.480)	0.005
His/His	36	49	0.979 (0.576–1.667)	0.920
Gln	344	345	1 (ref)	_

1.077

(0.859 - 1.349)

0.522

TABLE 2: The distribution of genotypes and allele frequencies and

the analysis of the odds ratio (OR) for 51Gln/His polymorphism of *APEX* gene in patients with colorectal cancer (CRC) and the

*Genotype distribution in the Hardy-Weinberg equilibrium, $\chi^2 = 0.125$. Values set in italics denote statistical significance.

259

278

TABLE 3: The distribution of genotypes and allele frequencies and the analysis of the odds ratio (OR) for 148Asp/Glu polymorphism of *APEX* gene in patients with colorectal cancer (CRC) and the control group.

Genotype/allele	Patients $(n = 309)$	Controls $(n = 301^*)$	OR (95% CI)	Р
Asp/Asp	51	88	1 (ref)	—
Asp/Glu	237	158	2.588 (1.736–3.859)	<0.0001
Glu/Glu	21	55	0.659 (0.358–1.212)	0.179
Asp	339	334	1 (ref)	—
Glu	279	268	1.026 (0.819–1.285)	0.823

*Genotype distribution in the Hardy-Weinberg equilibrium, $\chi^2 = 0.277$. Values set in italics denote statistical significance.

Asp/Glu genotype (Table 3) of 148Asp/Glu polymorphism of *APEX* gene (OR 2.588 (1.736–3.859); p < 0.0001) and Gly/Ala genotype (Table 4) of 23Gly/Ala polymorphism of *XPA* gene (OR 5.373 (3.418–8.446); p < 0.0001). We did not find any significant influence of 64Ile/Val polymorphism of APEX gene (Table 5) and 689Ser/Arg polymorphism of ERCC4 gene (Table 6) on modulation of CRC risk.

3.2. Gene-Gene Interactions. In order to investigate the interaction of the polymorphisms of the studied genes and to

TABLE 4: The distribution of genotypes and allele frequencies and the analysis of the odds ratio (OR) for 23Gly/Ala polymorphism of *XPA* gene in patients with colorectal cancer (CRC) and the control group.

Genotype/allele	Patients $(n = 310)$	Controls $(n = 304^*)$	OR (95% CI)	P
Gly/Gly	31	95	1 (ref)	_
Gly/Ala	263	150	5.373 (3.418–8.446)	<0.0001
Ala/Ala	16	59	0.831 (0.419–1.649)	0.597
Gly	325	340	1 (ref)	_
Ala	295	268	1.152 (0.920–1.442)	0.218

*Genotype distribution in the Hardy-Weinberg equilibrium, $\chi^2 = 0.988$. Values set in italics denote statistical significance.

TABLE 5: The distribution of genotypes and allele frequencies and the analysis of the odds ratio (OR) for 64Ile/Val polymorphism of *APEX* gene in patients with colorectal cancer (CRC) and the control group.

Genotype/allele	Patients $(n = 307)$	Controls $(n = 304^*)$	OR (95% CI)	Р
Ile/Ile	93	78	1 (ref)	_
Ile/Val	148	159	0.781 (0.537–1.136)	0.195
Val/Val	66	67	0.826 (0.525-1.301)	0.409
Ile	334	315	1 (ref)	_
Val	280	293	0.901 (0.720-1.128)	0.365

*Genotype distribution in the Hardy-Weinberg equilibrium, $\chi^2 = 0.408$.

TABLE 6: The distribution of genotypes and allele frequencies and the analysis of the odds ratio (OR) for 689Ser/Arg polymorphism of *ERCC4* gene in patients with colorectal cancer (CRC) and the control group.

Genotype/allele	Patients $(n = 309)$	Controls $(n = 304^*)$	OR (95% CI)	P
Ser/Ser	93	101	1 (ref)	_
Ser/Arg	155	160	1.052 (0.736–1.505)	0.777
Arg/Arg	61	43	1.541 (0.952–2.493)	0.078
Ser	341	362	1 (ref)	_
Arg	277	246	1.195 (0.953–1.499)	0.123

*Genotype distribution in the Hardy-Weinberg equilibrium, $\chi^2 = 0.107$.

evaluate their mutual influence on the risk of colorectal cancer, gene-gene interactions were analyzed. Results are presented in Table 7; only pairs that modulate the risk at a statistically significant level are shown. For the full set of results showing all pairs of gene-gene interactions,

please refer to the tables in Supplementary Material available online at https://doi.org/10.1155/2017/3840243. It has been revealed that genotype pair Gln/His-Val/Val for 51Gln/His APEX-64Ile/Val APEX increases the risk of CRC. For 51Gln/His APEX-148Asp/Glu APEX, we can observe increased risk in the case of Gln/His-Asp/Glu pair, but at the same time, coincidence of genotypes Gln/His-Glu/Glu and His/His-Asp/Asp decreases the risk. For pair 51Gln/His APEX-23Gly/Ala XPA, we observed increased risk in the case of genotypes Gln/His-Gly/Ala and His/His-Gly/Ala and decreased risk for Gln/His-Gly/Gly. Moreover, increased risk of colorectal cancer was revealed for pairs Gln/His-Arg/Arg (51Gln/His APEX-689Ser/Arg ERCC4), Val/Val-Asp/Glu (64Ile/Val APEX-148Asp/Glu APEX), Val/ Val-Gly/Ala (64Ile/Val APEX-23Gly/Ala XPA), and Asp/ Glu-Gly/Ala (148Asp/Glu APEX-23Gly/Ala XPA); while at the same time, risk was decreased for pairs Ile/Val-Asp/Asp (64Ile/Val APEX-148Asp/Glu APEX), Ile/Val-Gly/Gly, and Ile/Val-Ala/Ala (64Ile/Val APEX-23Gly/Ala XPA) as well as Asp/Asp-Gly/Ala and Asp/Glu-Gly/Gly (148Asp/Glu APEX-23Gly/Ala XPA). In addition, worth noticing is the major impact of Asp/Glu genotype of 148Asp/Glu APEX gene when paired with all genotypes of 689Ser/Arg ERCC4, and similarly, Gly/Ala genotype again paired with all genotypes of 689Ser/ Arg ERCC4.

3.3. Influence on Tumor Progression. In addition, we wanted to investigate potential correlation of our results with clinical data; therefore impact of presence of studied polymorphisms on progression of stage of tumor was tested, by a correlation of the distribution of genotypes with the state of tumor by the American Joint Committee on Cancer classification. Results are presented in Table 8. We found that 148Asp/Glu polymorphism of *APEX* gene and 23Gly/Ala polymorphism of XPA gene are increasing the risk of cancer in the second degree of advancement in relation to the first degree.

4. Discussion

All cells of the human body are constantly exposed to damaging agents, which can cause changes in the DNA. These changes, if not repaired, may lie at the basis of the process of carcinogenesis. To cope with those damages, the human body has developed a number of DNA repair mechanisms, including BER and NER systems. One of the key elements of BER is APEX gene product-class II AP endonuclease. Endonuclease cleaves the phosphodiester backbone 5' to the AP site, thereby initiating a repair process [23]. Polymorphisms in APEX gene have been for a long time a subject of interest in the area of modulating risk of malignant transformation and many of them have been connected to several types of cancers such as lung cancer, breast cancer, or bladder cancer [24–28]. In the case of colorectal cancer, it has been estimated that APEX Asp148Glu is involved in increasing CRC risk [21, 29] which is consistent with our results. However, some researchers suggest that there is no association between increased cancer risk and the APEX Asp148Glu polymorphisms [20] or even that its occurrence decreases

TABLE 7: The distribution of genotypes and the analysis of the odds ratio (OR) for gene-gene interactions in analyzed polymorphisms in patients with colorectal cancer (CRC) and the control group. Shown are only pairs that modulate the risk at a statistically significant level. All partial results for the gene-gene interaction are shown in the supplementary materials.

Gene-gene interaction	Genotype	Patients	Controls	OR (95% CI)	P
51Gln/His APEX-64Ile/Val APEX	Gln/His-Val/Val	55	32	2.266 (1.120-4.585)	0.022
	Gln/His-Asp/Glu	174	102	2.003 (1.091-3.676)	0.023
51Gln/His APEX-148Asp/Glu APEX	Gln/His-Glu/Glu	8	26	0.361 (0.137-0.951)	0.036
	His/His-Asp/Asp	6	28	0.252 (0.089-0.714)	0.007
	Gln/His-Gly/Gly	6	35	0.220 (0.078-0.622)	0.003
51Gln/His APEX-23Gly/Ala XPA	Gln/His-Gly/Ala	193	98	2.532 (1.362-4.707)	0.003
	His/His-Gly/Ala	30	8	4.821 (1.835-12.670)	0.001
51Gln/His APEX-689Ser/Arg ERCC4	Gln/His-Arg/Arg	39	24	2.464 (1.247-4.870)	0.009
	Ile/Val-Asp/Asp	10	27	0.403 (0.170-0.955)	0.036
6411e/Val APEX-148Asp/Glu APEX	Val/Val-Asp/Glu	59	18	3.567 (1.765-7.211)	0.0003
	Ile/Val-Gly/Gly	9	37	0.269 (0.103-0.700)	0.006
64Ile/Val APEX-23Gly/Ala XPA	Ile/Val-Ala/Ala	6	23	0.288 (0.097-0.859)	0.022
	Val/Val-Gly/Ala	62	14	4.895 (2.093-11.445)	0.0001
	Asp/Asp-Gly/Ala	26	58	0.399 (0.176-0.902)	0.025
148Asp/Glu APEX-23Gly/Ala XPA	Asp/Glu-Gly/Gly	8	46	0.155 (0.056-0.424)	0.0001
	Asp/Glu-Gly/Ala	218	81	2.392 (1.164-4.915)	0.015
	Asp/Asp-Ser/Arg	21	48	0.844 (0.370-1.924)	0.689
148Asp/Glu APEX-689Ser/Arg ERCC4	Asp/Asp-Arg/Arg	13	13	1.929 (0.707-5.263)	0.198
	Asp/Glu-Ser/Ser	69	49	2.716 (1.293-5.704)	0.007
	Gly/Ala-Ser/Ser	76	57	2.333 (1.061-5.131)	0.032
23Gly/Ala XPA-689Ser/Arg ERCC4	Gly/Ala-Ser/Arg	135	77	3.068 (1.431-6.577)	0.003
	Gly/Ala-Arg/Arg	47	16	5.141 (2.073-12.749)	0.0003

the risk—Brevik et al. report that carriers of the APEX codon 51 Gln/His genotype had a reduced CRC risk compared with carriers of the Gln/Gln genotype [30]. In contrast to these reports, we in our study found that Gln/His genotype increases the CRC risk. Similar differences can be observed in the case of NER system genes studied by us-23Gly/Ala of XPA gene is suggested to have no influence on risk of colorectal cancer [31, 32], while our results indicate the opposite (odds ratio (OR) 5.373 (3.418–8.446); *p* < 0.0001; Table 4). We believe that the cause of such divergence may lie in different selections of the study group (presented studies were carried out on the Japanese and Turkish populations while our research was on the Polish population) as well as bias in the selection because of the diet (eating red meat) and smoking tobacco, since it has been proven that ethnic group as well as other factors have a significant impact on the modulation of the risk of particular diseases [33, 34]. However, in our opinion, the main factor that could cause individual differences in modulation of risk by the same polymorphisms is gene-gene interactions. Several studies have confirmed that the polymorphisms of individual genes can significantly change the level of risk in case of coexistence with other specific polymorphisms. This phenomenon is observed even in cases when those polymorphisms do not have a significant effect on the modulation of the cancer risk when studied without mentioned coexistence [35-39]. Therefore, in the second part of this work, we have made calculations of gene-gene interactions to test the impact of a joint action

examined in earlier polymorphisms. Results are shown in Table 7. The first thing worth noting is the increased risk of CRC in the case of co-occurrence of genotype 51Gln/His of APEX gene with 64Val/Val of APEX gene when compared to risk associated only with 51Gln/His (OR 2.266 (1.120-4.585); p = 0.022 versus 1.706 (1.174–2.480); p = 0.005) and similar increased risk of CRC in the case of co-occurrence of genotype 51Gln/His of APEX gene with 689Arg/Arg ERCC4 (OR 2.464 (1.247–4.870); p = 0.009 versus 1.706 (1.174–2.480); p = 0.005). Similar increased risk can also be seen in the case of pairs 51Gln/His APEX-148Asp/Glu APEX and 51Gln/His APEX-23Gly/Ala XPA; however, 148Asp/Glu APEX and 23Gly/Ala XPA also considered individually increased risk of CRC. That is why we want to pay special attention to the first two pairs (51Gln/His APEX-64Val/Val APEX and 51Gln/His APEX-689Arg/Arg ERCC4) in which only 51Gln/His APEX increases the risk, but in the case of coexistence with the aforementioned polymorphisms, this risk becomes even greater. In our opinion, this clearly indicates a much more advanced system of impact of polymorphisms on the risk of cancer than the effect of a SNP. It can also indicate the interaction of BER and NER systems in removing damage which has been suggested by other researchers [40]. Confirmation of the thesis of common effect of polymorphisms of different genes to modulate the risk of cancer is also observed by us as protective effect in form of reducing the risk of CRC for genotype pairs (51Gln/His APEX-148Glu/Glu APEX and 51Gln/His APEX-23Gly/Gly

TABLE 8: Analysis of correlation of selected polymorphisms with the state of tumor according to classification of American Join	it Committee
on Cancer.	

					51Gln/His poly	morphisr	n of APEX gene				
Genotype	I	Patients	(<i>n</i> = 31	1)	II° versus I°		$III^{\circ} + IV^{\circ}$ versus I° $III^{\circ} + IV^{\circ}$		$III^{\circ} + IV^{\circ}$ versus	II°	
	I°	II°	$\operatorname{III}^{\circ}$	IV°	OR (95% CI)	P	OR (95% CI)	p	OR (95% CI)	p	
Gln/Gln	24	21	19	5	1 (ref)	_	1 (ref)	_	1 (ref)	_	
Gln/His	57	94	48	7	1.885 (0.963-3.690)	0.062	0.965 (0.491-1.898)	0.920	0.512 (0.261-1.004)	0.049	
His/His	19	15	2	0	0.902 (0.369-2.209)	0.823	_	_	_	_	
					64Ile/Val po	lymorphi	ism of APEX				
Genotype	I	Patients	(<i>n</i> = 30)7)	II° versus I°		III° + IV° versus	I°	$III^{\circ} + IV^{\circ}$ versus II°		
	I°	II°	$\operatorname{III}^{\circ}$	IV°	OR (95% CI)	р	OR (95% CI)	p	OR (95% CI)	p	
Ile/Ile	41	32	19	1	1 (ref)	_	1 (ref)	_	1 (ref)	—	
Ile/Val	64	51	28	5	1.021 (0.566-1.843)	1.000	1.057 (0.536-2.086)	0.862	1.035 (0.509-2.105)	0.920	
Val/Val	33	17	14	2	0.660 (0.313-1.391)	0.273	0.994 (0.446-2.215)	1.000	1.506 (0.623-3.638)	0.362	
					148Asp/Glu p	olymorp	hism of APEX				
Genotype	I	Patients	(<i>n</i> = 30)9)	II° versus I°		$III^{\circ} + IV^{\circ}$ versus I°		$III^{\circ} + IV^{\circ}$ versus II°		
	I°	II°	III°	IV°	OR (95% CI)	Р	OR (95% CI)	Р	OR (95% CI)	Р	
Asp/Asp	29	17	5	0	1 (ref)	_	1 (ref)	_	1 (ref)	_	
Asp/Glu	98	133	5	1	2.315 (1.205–4.449)	0.01	—	—	_	—	
Glu/Glu	16	5	0	0	0.533 (0.166-1.716)	0.288	—	—	—		
					23Gly/Ala poly	ymorphis	m of XPA gene				
Genotype	I	Patients	(<i>n</i> = 31	0)	II° versus I°		$III^{\circ} + IV^{\circ}$ versus	I°	$III^{\circ} + IV^{\circ}$ versus II°		
	I°	II°	$\operatorname{III}^{\circ}$	IV°	OR (95% CI)	p	OR (95% CI)	р	OR (95% CI)	P	
Gly/Gly	17	9	5	0	1 (ref)	_	1 (ref)	_	1 (ref)	_	
Gly/Ala	87	165	9	2	3.582 (1.533-8.370)	0.002	_	_	_	_	
Ala/Ala	11	5	0	0	0.859 (0.227-3.248)	0.823	_	_	_		
					689Ser/Arg p	olymorph	ism of ERCC4				
Genotype	F	Patients	(<i>n</i> = 30)9)	II° versus I°		III° + IV° versus	$III^{\circ} + IV^{\circ}$ versus I°		$III^{\circ} + IV^{\circ}$ versus II°	
	I°	II°	$\operatorname{III}^{\circ}$	IV°	OR (95% CI)	P	OR (95% CI)	p	OR (95% CI)	p	
Ser/Ser	49	31	9	4	1 (ref)	_	1 (ref)	_	1 (ref)	_	
Ser/Arg	83	52	16	4	0.990 (0.561-1.747)	1.000	0.908 (0.415-1.986)	0.806	0.795 (0.356-1.776)	0.578	
Arg/Arg	32	24	3	2	1.186 (0.592–2.374)	0.632	0.589 (0.192-1.811)	0.351	0.431 (0.137-1.352)	0.143	

Values set in italics denote statistical significance.

XPA) where one of the polymorphisms previously increased the risk, so we can see a reversal of the trend. It is also worth noting that the protective effect may have a pair in which none of the polymorphisms, when considered individually, showed no previous effect on the risk of CRC increasing nor decreasing it (64Ile/Val APEX-23Ala/ Ala XPA and 148Asp/Glu APEX-23Gly/Gly XPA). Finally, we want to draw attention to the fact that genotype 148Asp/Glu APEX which itself increases the risk of CRC (OR 2.588 (1.736–3.859); *p* < 0.0001), when considered along with any polymorphism of 689Ser/Arg ERCC4 (Ser/Ser, Ser/ Arg, or Arg/Arg), will always show a greater risk than alone (OR, resp., 2.716 (1.293-5.704), p=0.007, 2.963 (1.467-5.985), p = 0.002, and 2.643 (1.171–5.965), p = 0.018). In our opinion, this confirms the thesis pronounced earlier that the gene-gene interactions are the main factor that may influence individual differences in the predisposition to the occurrence of cancer.

Given such a complicated set of factors and their mutual interactions that may cause malignant transformation, it should be also suspected that there can be whole set of various factors contributing to the progression of already formed tumor. In order to evaluate the impact of polymorphisms of analyzed genes on the progress of colorectal cancer, we have correlated the distribution of genotypes with the progress of the tumor according to the classification of the American Joint Committee on Cancer. Increased risk of CRC in the II° of advancement in relation to the I° was observed in the case of 148Asp/Glu polymorphism of *APEX* gene and 23Gly/Ala polymorphism of XPA gene (Table 8). The observed effect is consistent with the effect of the polymorphisms in the increased risk of disease onset, which may suggest the relationship between specific genotypes and initiation and promotion of carcinogenesis as well as the progression of cancer and metastasis. This is in line with our earlier studies in which we presented similar thesis [38]. In our opinion, this allows not only to identify patients with an increased risk of cancer but also to identify within the group of patients already diagnosed with the disease to predict potential tumor growth and thus allows a more comprehensive approach to treating patients.

5. Conclusions

Genotypes 51Gln/His and 148Asp/Glu of APEX gene and 23Gly/Ala of XPA gene may increase the risk of CRC, while polymorphisms of 64Ile/Val of APEX gene and 689Ser/Arg of ERCC4 gene have no effect on modulating the risk. At the same time, gene-gene interactions may completely change the risk level; therefore, we advocate that they should be considered as very important factors when calculating the risk factor. Moreover, polymorphisms of BER and NER systems may not only initiate malignant transformation but also be able to contribute to tumor growth. We believe that our results are promising, yet further studies are needed on this subject to establish an incontestable link between a given polymorphism and its phenotypic effect in the modulation of BER and NER activity and thus its impact on carcinogenesis.

Conflicts of Interest

The authors declare no competing financial interests.

Acknowledgments

This work has been supported by Polish Ministry of Science Grant: 507/5-108-05/507-50-013 and UMED in Lodz Grant: 502-03/5-108-05/502-54-158.

References

- J. Ferlay, I. Soerjomataram, R. Dikshit et al., "Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012," *International Journal of Cancer*, vol. 136, no. 5, pp. E359–E386, 2015.
- [2] International Agency for Research on Cancer, WHO: GLOBCAN, *Estimated Cancer Incidence, Mortality and Prevalence Worldwide: Colorectal Cancer*, 2012.
- [3] L. Aaltonen, L. Johns, H. Järvinen, J. P. Mecklin, and R. Houlston, "Explaining the familial colorectal cancer risk associated with mismatch repair (MMR)-deficient and MMR-stable tumors," *Clinical Cancer Research*, vol. 13, no. 1, pp. 356–361, 2007.
- [4] W. M. Abdel-Rahman and P. Peltomaki, "Lynch syndrome and related familial colorectal cancers," *Critical Reviews in Oncogenesis*, vol. 14, no. 1, 2008.
- [5] S. N. Thibodeau, G. Bren, and D. Schaid, "Microsatellite instability in cancer of the proximal colon," *Science*, vol. 260, no. 5109, pp. 816–819, 1993.
- [6] L. A. Aaltonen, P. Peltomäki, F. S. Leach et al., "Clues to the pathogenesis of familial colorectal cancer," *Science*, vol. 260, no. 5109, pp. 812–816, 1993.

- [7] N. Seguí, L. B. Mina, C. Lázaro et al., "Germline mutations in FAN1 cause hereditary colorectal cancer by impairing DNA repair," *Gastroenterology*, vol. 149, no. 3, pp. 563–566, 2015.
- [8] B. Liu, R. Parsons, N. Papadopoulos et al., "Analysis of mismatch repair genes in hereditary non-polyposis colorectal cancer patients," *Nature Medicine*, vol. 2, no. 2, pp. 169– 174, 1996.
- [9] R. Fishel, M. K. Lescoe, M. R. Rao et al., "The human mutator gene homolog MSH2 and its association with hereditary nonpolyposis colon cancer," *Cell*, vol. 75, no. 5, pp. 1027–1038, 1993.
- [10] A. K. Win, J. P. Young, N. M. Lindor et al., "Colorectal and other cancer risks for carriers and noncarriers from families with a DNA mismatch repair gene mutation: a prospective cohort study," *Journal of Clinical Oncology*, vol. 30, no. 9, pp. 958–964, 2012.
- [11] Y. Li, S. Li, Z. Wu et al., "Polymorphisms in genes of APE1, PARP1, and XRCC1: risk and prognosis of colorectal cancer in a northeast Chinese population," *Medical Oncology*, vol. 30, no. 2, pp. 1–7, 2013.
- [12] M. G. Dunlop, S. E. Dobbins, S. M. Farrington et al., "Common variation near CDKN1A, POLD3 and SHROOM2 influences colorectal cancer risk," *Nature Genetics*, vol. 44, no. 7, pp. 770–776, 2012.
- [13] A. D. Beggs, E. Domingo, M. McGregor et al., "Loss of expression of the double strand break repair protein ATM is associated with worse prognosis in colorectal cancer and loss of Ku70 expression is associated with CIN," *Oncotarget*, vol. 3, no. 11, pp. 1348–1355, 2012.
- [14] X. Zhang, J. Yeo, E. L. Crawford, and J. C. Willey, "Investigation of C/EBPG transcription factor role in regulation of ERCC4 and ERCC5 in human lung cancer cells," *Cancer Research*, vol. 74, 19 Supplement, pp. 3381–3381, 2014.
- [15] S. Kohlhase, N. V. Bogdanova, P. Schürmann et al., "Mutation analysis of the ERCC4/FANCQ gene in hereditary breast cancer," *PloS One*, vol. 9, no. 1, article e85334, 2014.
- [16] B. Lu, J. Li, Q. Gao, W. Yu, Q. Yang, and X. Li, "Laryngeal cancer risk and common single nucleotide polymorphisms in nucleotide excision repair pathway genes ERCC1, ERCC2, ERCC3, ERCC4, ERCC5 and XPA," *Gene*, vol. 542, no. 1, pp. 64–68, 2014.
- [17] Y. Lou, R. Li, Y. Zhang et al., "XPA gene rs1800975 single nucleotide polymorphism and lung cancer risk: a metaanalysis," *Tumor Biology*, vol. 35, no. 7, pp. 6607–6617, 2014.
- [18] S. C. Koch, N. Simon, C. Ebert, and T. Carell, "Molecular mechanisms of xeroderma pigmentosum (XP) proteins," *Quarterly Reviews of Biophysics*, vol. 49, article e5, 2016.
- [19] J. de Boer and J. H. Hoeijmakers, "Nucleotide excision repair and human syndromes," *Carcinogenesis*, vol. 21, no. 3, pp. 453–460, 2000.
- [20] R. J. Hung, J. Hall, P. Brennan, and P. Boffetta, "Genetic polymorphisms in the base excision repair pathway and cancer risk: a HuGE review," *American Journal of Epidemiology*, vol. 162, no. 10, pp. 925–942, 2005.
- [21] M. Kasahara, K. Osawa, K. Yoshida et al., "Association of MUTYH Gln324His and APEX1 Asp148Glu with colorectal cancer and smoking in a Japanese population," *Journal of Experimental & Clinical Cancer Research*, vol. 27, no. 1, p. 1, 2008.
- [22] C. Ryk, R. Kumar, R. K. Thirumaran, and S. M. Hou, "Polymorphisms in the DNA repair genes XRCC1, APEX1,

XRCC3 and NBS1, and the risk for lung cancer in never-and ever-smokers," *Lung Cancer*, vol. 54, no. 3, pp. 285–292, 2006.

- [23] C. D. Mol, D. J. Hosfield, and J. A. Tainer, "Abasic site recognition by two apurinic/apyrimidinic endonuclease families in DNA base excision repair: the 3' ends justify the means," *Mutation Research/DNA Repair*, vol. 460, no. 3, pp. 211–229, 2000.
- [24] M. Shen, S. I. Berndt, N. Rothman et al., "Polymorphisms in the DNA base excision repair genes APEX1 and XRCC1 and lung cancer risk in Xuan Wei, China," *Anticancer Research*, vol. 25, no. 1B, pp. 537–542, 2005.
- [25] C. Kiyohara, K. Takayama, and Y. Nakanishi, "Association of genetic polymorphisms in the base excision repair pathway with lung cancer risk: a meta-analysis," *Lung Cancer*, vol. 54, no. 3, pp. 267–283, 2006.
- [26] M. Yin, Z. Liao, Z. Liu et al., "Functional polymorphisms of base excision repair genes XRCC1 and APEX1 predict risk of radiation pneumonitis in patients with non-small cell lung cancer treated with definitive radiation therapy," *International Journal of Radiation Oncology Biology Physics*, vol. 81, no. 3, pp. e67–e73, 2011.
- [27] R. Gangawar, D. Ahirwar, A. Mandhani, and R. D. Mittal, "Impact of nucleotide excision repair ERCC2 and base excision repair APEX1 genes polymorphism and its association with recurrence after adjuvant BCG immunotherapy in bladder cancer patients of North India," *Medical Oncology*, vol. 27, no. 2, pp. 159–166, 2010.
- [28] Y. Zhang, P. A. Newcomb, K. M. Egan et al., "Genetic polymorphisms in base-excision repair pathway genes and risk of breast cancer," *Cancer Epidemiology Biomarkers & Prevention*, vol. 15, no. 2, pp. 353–358, 2006.
- [29] E. Canbay, B. Cakmakoglu, U. Zeybek et al., "Association of APE1 and hOGG1 polymorphisms with colorectal cancer risk in a Turkish population," *Current Medical Research and Opinion*, vol. 27, no. 7, pp. 1295–1302, 2011.
- [30] A. Brevik, A. D. Joshi, R. Corral et al., "Polymorphisms in base excision repair genes as colorectal cancer risk factors and modifiers of the effect of diets high in red meat," *Cancer Epidemiology Biomarkers & Prevention*, vol. 19, no. 12, pp. 3167–3173, 2010.
- [31] R. D. Hansen, M. Sørensen, A. Tjønneland et al., "XPA A23G, XPC Lys939Gln, XPD Lys751Gln and XPD Asp312Asn polymorphisms, interactions with smoking, alcohol and dietary factors, and risk of colorectal cancer," *Mutation Research/Fundamental and Molecular Mechanisms of Muta*genesis, vol. 619, no. 1, pp. 68–80, 2007.
- [32] J. Liu, Z. Zhang, X. L. Cao et al., "XPA A23G polymorphism and susceptibility to cancer: a meta-analysis," *Molecular Biology Reports*, vol. 39, no. 6, pp. 6791–6799, 2012.
- [33] F. Kamangar, G. M. Dores, and W. F. Anderson, "Patterns of cancer incidence, mortality, and prevalence across five continents: defining priorities to reduce cancer disparities in different geographic regions of the world," *Journal of Clinical Oncology*, vol. 24, no. 14, pp. 2137–2150, 2006.
- [34] M. Manuguerra, F. Saletta, M. R. Karagas et al., "XRCC3 and XPD/ERCC2 single nucleotide polymorphisms and the risk of cancer: a HuGE review," *American Journal of Epidemiology*, vol. 164, no. 4, pp. 297–302, 2006.
- [35] J. Kabzinski, B. Mucha, M. Cuchra et al., "Efficiency of base excision repair of oxidative DNA damage and its impact on the risk of colorectal cancer in the polish population," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 3125989, 9 pages, 2016.

- [36] L. W. Hahn, M. D. Ritchie, and J. H. Moore, "Multifactor dimensionality reduction software for detecting gene-gene and gene-environment interactions," *Bioinformatics*, vol. 19, no. 3, pp. 376–382, 2003.
- [37] E. L. Goode, C. M. Ulrich, and J. D. Potter, "Polymorphisms in DNA repair genes and associations with cancer risk," *Cancer Epidemiology Biomarkers & Prevention*, vol. 11, no. 12, pp. 1513–1530, 2002.
- [38] J. Kabzinski, K. Przybylowska, L. Dziki, A. Dziki, and I. Majsterek, "An association of selected ERCC2 and ERCC5 genes polymorphisms, the level of oxidative DNA damage and its repair efficiency with a risk of colorectal cancer in polish population," *Cancer Biomarkers*, vol. 15, no. 4, pp. 413–423, 2015.
- [39] D. G. Cox, R. M. Tamimi, and D. J. Hunter, "Gene×gene interaction between MnSOD and GPX-1 and breast cancer risk: a nested case-control study," *BMC Cancer*, vol. 6, no. 1, p. 1, 2006.
- [40] B. Plosky, L. Samson, B. P. Engelward et al., "Base excision repair and nucleotide excision repair contribute to the removal of N-methylpurines from active genes," *DNA Repair*, vol. 1, no. 8, pp. 683–696, 2002.