DATABASE ANALYSIS

e-ISSN 1643-3750 © Med Sci Monit, 2021; 27: e934522 DOI: 10.12659/MSM.934522

Accepted Available online Published	l: 2021.10.05 2021.10.27 l: 2021.12.09		SLC26A4 Prognos Infiltrate	Antis Antis tic Bio es in B	ense l marke reast	RNA 1 r and Cance	Is an Independent Correlate of Immune
Authors S Da Statist Data In Manuscript Liter Fund	s' Contribution: itudy Design A ta Collection B ical Analysis C terpretation D Preparation E ature Search F ds Collection G	ABCEF 1 BC 2 BC 3 BC 4 BC 1 BC 5 ABCEF 1	Weiwei Yi Haiqing Shen Dexi Sun Yongchao Xu Yujie Feng Dongbing Li Caixia Wang			1 C F 2 C P 3 C P 4 C P 5 C P	Department of Oncology, Shandong Provincial Hospital Affiliated to Shandong irst Medical University, Jinan, Shandong, PR China Department of Intervention, The Fourth People's Hospital of Zibo, Zibo, Shandon R China Department of Oncology, The Fourth People's Hospital of Zibo, Zibo, Shandong, R China Department of Oncology, Guanxian Central Hospital, Liaocheng, Shandong, R China Department of Medicine, MyGene Diagnostics Co., Ltd., Guangzhou, Guangdong R China
-	Correspondin Financia Conflict of	g Author: l support: f interest:	Caixia Wang, e-mail: None declared None declared	slyywcx@163.co	m		
	Back Material/N	ground: Nethods: Results:	Aberrant expression tant role in some cer (BC) and the p Statistical analysis characteristics. The tween SLC26A4-AS1 PAM50 (P<0.001), all survival (OS) (I cific survival (OSS 0.154-0.579; P<0.0 CYP2E1 reactions, osmotic coupling, dosomal sorting c ciated with some SLC26A4-AS1 exp with BC. It may be	on of long non- cancer types. I ossible regula s was used to e Kaplan-Meie S1 expression d to investigat expression in and menopar hazard ratio [H) (HR: 0.57; 95 D01) was indep protein export budding and n omplex require types of immu ression was si e a promising p	coding RNA (li However, the tory mechanis assess the con- er method and and prognosi- te the possible n BC was ass- use status (P- HR]: 0.56; 959 5% CI: 0.37-0.4 pendently cor- t, mitochondri maturation of ed for transpo- une infiltrating ignificantly as prognostic bio	ncRNA) SLC2 clinical signi sms of SLC2 rrelation bet l Cox regress is. Gene set e regulatory ociated with (0.001). Low 6 confidence 38; P=0.011) related with al_ciii_asser HIV virion, o rt, and histo g cells. sociated with marker for E	16A4 antisense RNA 1 (SLC26A4-AS1) plays an impor- ficance of SLC26A4-AS1 in patients with breast can- 6A4-AS1 are unclear. ween SLC26A4-AS1 expression and patients' clinical sion analysis were used to assess the correlation be- enrichment analysis (GSEA) and immuno-infiltration mechanisms of SLC26A4-AS1. age (P <0.001), estrogen-receptor status (P <0.001), YSLC26A4-AS1 expression predicted a poorer over- e interval [CI]: 0.40-0.78; P =0.001) and disease-spe- 0. Also, SLC26A4-AS1 expression (HR: 0.298; 95% CI: OS in patients with BC. SLC26A4-AS1 was related to mbly, formation of adenosine triphosphate by chemi- cristae formation, biocarta proteasome pathway, en- one modification. SLC26A4-AS1 expression was asso- th poor survival and immune infiltration in patients 3C.
	Ke	ywords:	Biomarkers, Tum	or • Breast N	eoplasms • P	rognosis	
	Abbrev	viations:	IncRNA – long no cer; TCGA – The (DSS – disease-sp	ncoding RNA; Cancer Genom ecific survival	s SLC26A4-AS ne Atlas; GSE A l	51 – SLC26A A – gene set	4 antisense RNA 1; BC – breast can- t enrichment analysis; OS – overall survival;
	Full-t	ext PDF:	https://www.med	scimonit.com/	/abstract/inde	ex/idArt/934	1522
			2717	5	1 2 7	2 32	2

Low Expression of Long Noncoding RNA



MEDICAL SCIENCE

MONITOR

Received: 2021.08.25

Background

Breast cancer (BC) is the second leading cause of cancer death in women [1] with 2.1 million new cases and 627 000 deaths yearly, making it the most common malignancy and the second leading cause of cancer-related deaths in women [2]. Although significant progress has been made in the personalized treatment of BC, the 5-year overall survival (OS) rate remains relatively low due to the heterogeneity of tumors [3]. Although screening for BC, such as through mammograms and serum tumor markers, is becoming increasingly popular, limited health service infrastructure is still a problem in some areas, hindering early diagnosis [4]. Therefore, new prognostic biomarkers of BC and their molecular mechanisms need to be further explored.

Long noncoding RNAs (lncRNAs), RNA molecules with transcripts longer than 200 nucleotides, have limited coding potential [5]. They are emerging as a new class of important regulators of cell biological behavior and are indispensable players involved in tumorigenesis, progression, and metastasis [6]. For easy access and detection, lncRNA can be encapsulated in membrane vesicles, such as exosomes and apoptotic bodies, or stably bound to specific proteins and released into body fluids, such as serum and urine [7]. The aberrant expression of lncRNAs is associated with the development and progression of BC [8-10]. Therefore, screening for the lncRNAs associated with BC prognosis is important for patients with BC.

The lncRNA SLC26A4-AS1 inhibits the ability of pancreatic cancer to migrate, invade, and metastasize [11]. Overexpression of SLC26A4-AS1 inhibits the aggressiveness of glioma cells and their pro-angiogenic capacity [12]. Downregulated SLC26A4-AS1 is significantly associated with lower OS in patients with BC [13]. The upregulation of SLC26A4-AS1 was associated with its promoter hypermethylation in BC [14]. However, the clinical significance of SLC26A4-AS1 in patients with breast cancer and the possible regulatory mechanisms of SLC26A4-AS1 are unclear.

The tumor microenvironment (TME) includes cellular and noncellular components [15]. Inflammatory cells, including neutrophils and myeloid-derived suppressor cells, suppress the beneficial immune function of TME, preventing normal immune cells from attacking tumor cells and promoting tumor growth [16]. Infiltration of the TME by immune cells is a strategy used by tumor cells to evade immune-mediated killing [17]. The BC TME is rich in immune infiltrates with different functions [18]. However, the role of SLC26A4-AS1 in BC tumor immunity is unclear.

In this study, the clinical significance of SLC26A4-AS1 was analyzed based on data from The Cancer Genome Atlas (TCGA) database, and the possible regulatory network of SLC26A4-AS1 was investigated based on gene set enrichment analysis (GSEA) and immune infiltration analysis. This study may provide promising prognostic biomarkers for patients with BC.

Material and Methods

Differences in SLC26A4-AS2 Expression

For molecule SLC26A4-AS1 [ENSG00000227199], the following analyses were conducted as described in the literature: patient baseline information [19]; unpaired samples [20]; paired samples [20]; and receiver operating characteristics analysis [19].

Correlation Between SLC26A4-AS2 and Clinical Characteristics

Analysis of clinical relevance for molecule SLC26A4-AS1 was performed as described in the literature [20]. The clinical variables included were age, progesterone-receptor (PR) status, estrogen-receptor (ER) status, PAM50, and menopause status. RNAseq data and clinical data in level 3 HTSeq-FPKM format were from The Cancer Genome Atlas Breast Invasive Carcinoma (TCGA-BRCA) project.

With SLC26A4-AS1 as the dependent variable, logistics analysis was performed as described in the literature [20].

Correlation Between SLC26A4-AS2 and Prognosis

Kaplan-Meier curve was performed for SLC26A4-AS1 according to data analysis methods described in the literature [19,21]. Prognosis included OS and disease-specific survival (DSS).

Cox regression analysis was performed as described in the literature [20,21] and included the variables tumor (T) stage, lymph node (N) stage, pathologic stage, race, age, histological type, PR status, ER status, human epidermal growth factor receptor 2 (HER2) status, PAM50, menopause status, anatomic neoplasm subdivisions, radiation therapy, and SLC26A4-AS1.

GSEA Analysis

Single-gene differential analysis of SLC26A4-AS1 was performed as described in the literature [20,22]. GSEA analysis was performed as described in the literature [20,23,24].

Immune Infiltration Analysis by Single-Sample GSEA

Data analysis by single-sample GSEA of SLC26A4-AS1 was performed by methods described in the literature [20,25,26].

Characteristic	Levels	0	verall	
n		1083		
T stage, n (%)	T1	277	(25.6%)	
	T2	629	(58.2%)	
	Т3	139	(12.9%)	
	T4	35	(3.2%)	
N stage, n (%)	NO	514	(48.3%)	
	N1	358	(33.6%)	
	N2	116	(10.9%)	
	N3	76	(7.1%)	
M stage, n (%)	MO	902	(97.8%)	
	M1	20	(2.2%)	
Pathologic stage,	Stage I	181	(17.1%)	
n (%)	Stage II	619	(58.4%)	
	Stage III	242	(22.8%)	
	Stage IV	18	(1.7%)	
Race, n (%)	Asian	60	(6%)	
	Black or African American	181	(18.2%)	
	White	753	(75.8%)	
Age, n (%)	≤60	601	(55.5%)	
	>60	482	(44.5%)	
Histological type, n (%)	Infiltrating ductal carcinoma	772	(79%)	
	Infiltrating lobular carcinoma	205	(21%)	

Table 1. The Cancer Genome Atlas-based characterization of patients with breast cancer.

Results

Clinical Characteristics

A total of 1083 patients were included in the present study (**Table 1**). The T stages of the patients were as follows: 277 were T1 (25.6%), 629 were T2 (58.2%), 139 were T3 (12.9%), and 35 were T4 (3.2%). The N stage of the patients were as follows: 514 were N0 (48.3%), 358 were N1 (33.6%), and were 116 N2 (10.9%). There were 902 patients with metastasis (M) stage 0 (97.8%) and 20 with stage M1 (2.2%). The pathologic stages were as follows: 181 in stage I (17.1%), 619 in stage II (58.4%), 242 in stage III (22.8%), and 18 in stage IV (1.7%). Regarding race, 753 patients were White, 60 patients were Asian, and 181 patients were Black or African American. The

Characteristic	Levels	0	verall
PR status, n (%)	Negative	342	(33.1%)
	Indeterminate	4	(0.4%)
	Positive	688	(66.5%)
ER status, n (%)	Negative	240	(23.2%)
	Indeterminate	2	(0.2%)
	Positive	793	(76.6%)
HER2 status, n (%)	Negative	558	(76.8%)
	Indeterminate	12	(1.7%)
	Positive	157	(21.6%)
PAM50, n (%)	Normal	40	(3.7%)
	LumA	562	(51.9%)
	LumB	204	(18.8%)
	Her2	82	(7.6%)
	Basal	195	(18%)
Menopause status,	Pre	229	(23.6%)
n (%)	Peri	40	(4.1%)
	Post	703	(72.3%)
Anatomic neoplasm	Left	563	(52%)
subdivisions, n (%)	Right	520	(48%)
Radiation therapy,	No	434	(44%)
n (%)	Yes	553	(56%)
Age, median (IQR)		58	(48.5, 67)

age range was 48.5 to 67 years, with a median of 58 years; 601 patients were \leq 60 years (55.5%) and 482 patients were >60 years (44.5%). Histological types were 772 infiltrating ductal carcinoma (79%) and 205 infiltrating lobular carcinoma (21%). PR status was negative in 342 (33.1%) patients, indeterminate in 4 (0.4%) patients, and positive in 688 (66.5%) patients. ER status was negative in 240 (23.2%) patients, indeterminate in 2 (0.2%) patients, and positive in 793 (76.6%) patients. HER2 status was negative in 558 (76.8%) patients, indeterminate in 12 (1.7%) patients, and positive in 157 (21.6%) patients. The PAM50 was normal in 40 (3.7%) patients, luminal A in 562 (51.9%) patients, and basal in 195 (18%) patients. A total of 229 (23.6%) patients were in pre-menopause, 40 (4.1%) were in peri-menopause, and 703 (72.3%) were in menopause. The

This work is licensed under Creative Common Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0)

e934522-3



Figure 1. Expression of SLC26A4-AS1 in breast cancer (BC) tissues compared to normal tissues. (A) Differential expression of SLC26A4-AS1 in BC tissues and normal breast tissues. (B) Differential expression of SLC26A4-AS1 in BC tissues and matched normal breast tissues. (C) receiver operating characteristic curves showed the efficiency of SLC26A4-AS1 expression levels in distinguishing BC tissues from non-tumor tissues. Significance markers: *** P<0.001.

anatomic neoplasm subdivisions were 563 left (52%) and 520 right (48%). A total of 553 (56%) patients underwent radiation therapy and 434 (44%) did not.

Low Expression of SLC26A4-AS1 Predicted Poor Clinical Characteristics in Patients with BC

SLC26A4-AS1 was found to be significantly less expressed in BC tissues than in normal breast tissues (0.267±0.015 vs 0.808±0.062, *P*<0.001), based on the TCGA database of 1109 BC tissues and 113 normal breast tissues (**Figure 1A**). The expression of SLC26A4-AS1 in 113 BC tissues and their matched normal breast tissues showed that BC tissues had a low expression of SLC26A4-AS1 (0.231±0.036 vs 0.808±0.062, *P*<0.001, **Figure 1B**). The area under curve (AUC) of SLC26A4-AS1 was 0.805 (**Figure 1C**). SLC26A4-AS1 could be used to differentiate between BC and normal breast tissue. SLC26A4-AS1 expression was associated with age (*P*<0.001), PR status (*P*<0.001), ER status (*P*<0.001), PAM50 (*P*<0.001), and menopause status (*P*=0.002) (**Table 2**). SLC26A4-AS1 was significantly related to age (*P*<0.001), ER status (*P*<0.001), PAM50 (*P*<0.001), and menopause status (*P*<0.001) (**Figure 2, Table 3**).

Correlation Between SLC26A4-AS1 Expression and Patient Prognosis

SLC26A4-AS1 expression was significantly associated with poor OS (hazard ratio [HR]: 0.56; 95% confidence interval [CI]: 0.40-0.78; P=0.001) and DSS (HR: 0.57; 95% CI: 0.37-0.88; P=0.011) of patients with BC (**Figure 3**). Low SLC26A4-AS1 expression (HR: 0.560; 95% CI: 0.403-0.777; P<0.001) was associated with T stage (HR: 1.482; 95% CI: 1.007-2.182; P=0.046), N stage (HR: 2.239; 95% CI: 1.567-3.199; P<0.001), pathologic stage (HR:

2.391; 95% CI: 1.703-3.355; *P*<0.001), age (HR: 2.020; 95% CI: 1.465-2.784; *P*<0.001), menopause status (HR: 0.426; 95% CI: 0.259-0.700; *P*<0.001), and radiation therapy (HR: 0.576; 95% CI: 0.394-0.841; *P*=0.004) (**Table 4**). SLC26A4-AS1 expression (HR: 0.298; 95% CI: 0.154-0.579; *P*<0.001), pathologic stage (HR: 5.876; 95% CI: 2.271-15.205, *P*<0.001), and radiation therapy (HR: 0.448; 95% CI: 0.243-0.823, *P*=0.004) were independent prognostic factors for patients with BC (**Table 4**).

SLC26A4-AS1-Associated Pathway

A total of 124 datasets showed significant differential enrichment in SLC26A4-AS1 high-expression phenotypes, while 50 datasets showed significant differential enrichment in SLC26A4-AS1 low-expression phenotypes. The top 9 data sets with a low *P* value included CYP2E1 reactions, protein export, mitochondrial_ciii_assembly, formation of adenosine triphosphate (ATP) by chemiosmotic coupling, histone modification, budding and maturation of HIV virion, cristae formation, biocarta proteasome pathway, and endosomal sorting complex required for transport (**Table 5, Figure 4**).

Correlation of SLC26A4-AS1 Expression with Immune Infiltration

For aDC, the mean level of the SLC26A4-AS1 low-expression group (0.284 \pm 0.109) was significantly lower than that of the high-expression group (0.314 \pm 0.126) (*P*<0.001) (**Figure 5A**). The results (r=0.150, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and aDC (**Figures 6A and 7**). For B cells, the mean level of the SLC26A4-AS1 low-expression group (0.167 \pm 0.084) was significantly lower than that of the high-expression group (0.199 \pm 0.091, *P*<0.001) (**Figure 5B**). The

Table 2. Expression of SLC26A4-AS1 in breast cancer is associated with clinical characteristics.

Characteristic	Low expression of SLC26A4-AS1	High expression of SLC26A4-AS1	р	Statistic	Method
n	541	542			
T stage, n (%)			0.909	0.54	Chisq.test
T1	140 (13%)	137 (12.7%)			
T2	309 (28.6%)	320 (29.6%)			
T3	71 (6.6%)	68 (6.3%)			
T4	19 (1.8%)	16 (1.5%)			
N stage, n (%)			0.536	2.18	Chisq.test
NO	245 (23%)	269 (25.3%)			
N1	179 (16.8%)	179 (16.8%)			
N2	61 (5.7%)	55 (5.2%)			
N3	42 (3.9%)	34 (3.2%)			
M stage, n (%)			0.836	0.04	Chisq.test
MO	452 (49%)	450 (48.8%)			
M1	11 (1.2%)	9 (1%)			
Pathologic stage, n (%)			0.736	1.27	Chisq.test
Stage I	87 (8.2%)	94 (8.9%)			
Stage II	306 (28.9%)	313 (29.5%)			
Stage III	123 (11.6%)	119 (11.2%)			
Stage IV	11 (1%)	7 (0.7%)			
Race, n (%)			0.945	0.11	Chisq.test
Asian	30 (3%)	30 (3%)			
Black or African American	89 (9%)	92 (9.3%)			
White	363 (36.5%)	390 (39.2%)			
Age, n (%)			<0.001	11.49	Chisq.test
≤60	272 (25.1%)	329 (30.4%)			
>60	269 (24.8%)	213 (19.7%)			
Histological type, n (%)			0.940	0.01	Chisq.test
Infiltrating ductal carcinoma	387 (39.6%)	385 (39.4%)			
Infiltrating lobular carcinoma	104 (10.6%)	101 (10.3%)			
PR status, n (%)			<0.001		Fisher.test
Negative	141 (13.6%)	201 (19.4%)			
Indeterminate	2 (0.2%)	2 (0.2%)			
Positive	375 (36.3%)	313 (30.3%)			

e934522-5

Characteristic	Low expression of SLC26A4-AS1	High expression of SLC26A4-AS1	р	Statistic	Method
ER status, n (%)			<0.001		Fisher.test
Negative	89 (8.6%)	151 (14.6%)			
Indeterminate	1 (0.1%)	1 (0.1%)			
Positive	428 (41.4%)	365 (35.3%)			
HER2 status, n (%)			0.624	0.94	Chisq.test
Negative	274 (37.7%)	284 (39.1%)			
Indeterminate	5 (0.7%)	7 (1%)			
Positive	71 (9.8%)	86 (11.8%)			
PAM50, n (%)			<0.001	82.17	Chisq.test
Normal	7 (0.6%)	33 (3%)			
LumA	291 (26.9%)	271 (25%)			
LumB	139 (12.8%)	65 (6%)			
Her2	48 (4.4%)	34 (3.1%)			
Basal	56 (5.2%)	139 (12.8%)			
Menopause status, n (%)			0.002	12.79	Chisq.test
Pre	91 (9.4%)	138 (14.2%)			
Peri	17 (1.7%)	23 (2.4%)			
Post	372 (38.3%)	331 (34.1%)			
Anatomic neoplasm subdivisions, n (%)			0.236	1.4	Chisq.test
Left	271 (25%)	292 (27%)			
Right	270 (24.9%)	250 (23.1%)			
Radiation therapy, n (%)			0.576	0.31	Chisq.test
No	219 (22.2%)	215 (21.8%)			
Yes	268 (27.2%)	285 (28.9%)			
Age, median (IQR)	60 (51, 69)	55 (47, 65)	<0.001	171329	Wilcoxon

Table 2 continued. Expression of SLC26A4-AS1 in breast cancer is associated with clinical characteristics.

results (r=0.220, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and B cells (**Figures 6B and 7**). For CD8 T cells, the mean level of the SLC26A4-AS1 low-expression group (0.522 \pm 0.018) was significantly lower than that of the high-expression group (0.523 \pm 0.023) (*P*=0.003) (**Figure 5C**). The results (r=0.067, *P*=0.027) indicated a positive correlation between SLC26A4-AS1 and CD8 T cells (**Figures 6C and 7**). For cytotoxic cells, the mean level of the SLC26A4-AS1 low-expression group (0.319 \pm 0.083) was significantly lower than that of the high-expression group (0.337 \pm 0.09) (*P*<0.001) (**Figure 5D**). The results (r=0.130, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and CD8 T cells (**Figures 6D and 7**). For DC, the mean level of the SLC26A4-AS1 low-expression group (0.281±0.089) was significantly lower than that of the high-expression group (0.303±0.098) (P<0.001) (**Figure 5E**). The results (r=0.160, P<0.001) indicated a positive correlation between SLC26A4-AS1 and CD8 T cells (**Figures 6E and 7**). For macrophages, the mean level of the SLC26A4-AS1 low-expression group (0.445±0.054) was significantly lower than that of the high-expression group (0.456±0.052) (P<0.001) (**Figure 5F**). The results (r=0.140, P<0.001) indicated a positive correlation between SLC26A4-AS1 and macrophages (**Figures 6F and 7**). For neutrophils, the mean level of the SLC26A4-AS1 low-expression group (0.231±0.045) was significantly lower than that of



Figure 2. SLC26A4-AS1 expression is associated with the clinical characteristics of patients with breast cancer. (A) Age, (B) estrogenreceptor status, (C) PAM50, (D) menopause. Significance markers: *** P<0.001.

the high-expression group (0.239±0.053) (*P*=0.005) (**Figure 5G**). The results (r=0.130, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and neutrophils (**Figures 6G and 7**). For NK CD56bright cells, the mean level of the SLC26A4-AS1 low-expression group (0.348±0.050) was significantly higher than that of the high-expression group (0.331±0.056) (*P*=0.005) (**Figure 5H**). The results (r=-0.170, *P*<0.001) indicated a negative correlation between SLC26A4-AS1 and NK CD56bright cells (**Figures 6H and 7**). For NK CD56dim cells, the mean level of the SLC26A4-AS1 low-expression group (0.180±0.061) was significantly lower than that of the high-expression group (0.193±0.072) (*P*<0.001) (**Figure 5I**). The results (r=0.130, *P*<0.001) indicated a positive correlation between

SLC26A4-AS1 and NK CD56dim cells (Figures 6I and 7). For T cells, the mean level of the SLC26A4-AS1 low-expression group (0.312 \pm 0.110) was significantly lower than that of the high-expression group (0.339 \pm 0.116) (*P*<0.001) (Figure 5J). The results (r=0.160, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and T cells (Figures 6J and 7). For T helper cells, the mean level of the SLC26A4-AS1 low-expression group (0.554 \pm 0.023) was significantly lower than that of the high-expression group (0.559 \pm 0.023) (*P*<0.001) (Figure 5K). The results (r=0.120, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and T helper cells (Figures 6K and 7). For Tcm, the mean level of the SLC26A4-AS1 low-expression group (0.356 \pm 0.028) was significantly lower than that of the Table 3. Correlation between SLC26A4-AS1 expression and clinical characteristics in patients with breast cancer (logistic regression).

Characteristics	Total (N)	Odds ratio (OR)	P value
T stage (T2 & T3 & T4 vs T1)	1080	1.035 (0.787-1.360)	0.807
N stage (N1 & N2 & N3 vs N0)	1064	0.866 (0.680-1.101)	0.240
M stage (M1 vs M0)	922	0.822 (0.328-2.004)	0.666
Pathologic stage (Stage III & Stage IV vs Stage I & Stage II)	1060	0.908 (0.686-1.201)	0.499
Race (Black or African American & White vs Asian)	994	1.066 (0.631-1.802)	0.809
Age (>60 vs ≤60)	1083	0.655 (0.514-0.833)	<0.001
Histological type (Infiltrating Lobular Carcinoma vs Infiltrating Ductal Carcinoma)	977	0.976 (0.717-1.329)	0.878
PR status (Indeterminate vs Negative)	346	0.701 (0.083-5.902)	0.725
ER status (Positive & Indeterminate vs Negative)	1035	0.503 (0.373-0.675)	<0.001
HER2 status (Indeterminate & Positive vs Negative)	727	1.181 (0.836-1.671)	0.346
PAM50 (LumA & LumB & Her2 vs Basal)	1043	0.312 (0.221-0.435)	<0.001
Menopause status (Post vs Pre & Peri)	972	0.597 (0.448-0.793)	<0.001
Anatomic neoplasm subdivisions (Right vs Left)	1083	0.859 (0.677-1.091)	0.213
Radiation therapy (Yes vs No)	987	1.083 (0.842-1.393)	0.533



Figure 3. Low expression of SLC26A4-AS1 is associated with poor overall survival and disease-specific survival in patients with breast cancer. (A) Overall survival. (B) Disease-specific survival.

high-expression group (0.364 ± 0.031) (*P*<0.001) (Figure 5L). The results (r=0.150, *P*<0.001) indicated a positive correlation between SLC26A4-AS1 and Tcm (Figures 6L and 7). For Tem, the mean level of the SLC26A4-AS1 low-expression group (0.362±0.026, P=0.008) was significantly lower than the mean level of the high-expression group (0.366±0.029, *P*=0.008)

(P=0.008) (Figure 5M). The results (r=0.100, P=0.001) indicated a positive correlation between SLC26A4-AS1 and Tem (Figures 6M and 7). For TFH, the mean level of the SLC26A4-AS1 low-expression group (0.345±0.029) was significantly lower than that of the high-expression group (0.350±0.033) (P<0.001) (Figure 5N). The results (r=0.120, P<0.001) indicated a positive

Channa stanistica	T-4-1 (AI)	Univariate ana	lysis	Multivariate analysis		
Characteristics	Total (N)	HR (95% CI)	P value	HR (95% CI)	P value	
T stage (T2 & T3 & T4 vs T1)	1079	1.482 (1.007-2.182)	0.046	1.355 (0.620-2.958)	0.446	
N stage (N1 & N2 & N3 vs N0)	1063	2.239 (1.567-3.199)	<0.001	0.660 (0.251-1.733)	0.399	
Pathologic stage (Stage III & Stage IV vs Stage I & Stage II)	1059	2.391 (1.703-3.355)	<0.001	5.876 (2.271-15.205)	<0.001	
Race (Black or African American & White vs Asian)	993	1.362 (0.432-4.289)	0.598			
Age (>60 vs ≤60)	1082	2.020 (1.465-2.784)	<0.001	1.900 (0.948-3.811)	0.071	
Histological type (Infiltrating Lobular Carcinoma vs Infiltrating Ductal Carcinoma)	977	0.827 (0.526-1.299)	0.410			
PR status (Indeterminate & Positive vs Negative)	1033	0.733 (0.525-1.025)	0.070	0.712 (0.303-1.676)	0.437	
ER status (Indeterminate & Positive vs Negative)	1034	0.725 (0.505-1.041)	0.082	0.494 (0.190-1.283)	0.147	
HER2 status (Indeterminate & Positive vs Negative)	727	1.519 (0.927-2.488)	0.097	0.638 (0.300-1.355)	0.242	
PAM50 (LumA & LumB & Her2 vs Basal)	1042	0.961 (0.642-1.439)	0.848			
Menopause status (Pre & Peri vs Post)	971	0.426 (0.259-0.700)	<0.001	0.413 (0.155-1.096)	0.076	
Anatomic neoplasm subdivisions (Right vs Left)	1082	0.766 (0.554-1.057)	0.105			
Radiation therapy (Yes vs No)	986	0.576 (0.394-0.841)	0.004	0.496 (0.263-0.936)	0.030	
SLC26A4-AS1 (High vs Low)	1082	0.560 (0.403-0.777)	<0.001	0.298 (0.154-0.579)	<0.001	

Table 4. Correlation between overall survival and clinical characteristics in patients with breast cancer (Cox regression).

Table 5. Enrichment of SLC26A4-AS1-related pathways (gene set enrichment analysis).

Gene set name	NES	P adjust	FDR
REACTOME_CYP2E1_REACTIONS	-2.1883	0.0015	0.3926
KEGG_PROTEIN_EXPORT	-2.0546	0.0018	0.3926
WP_MITOCHONDRIAL_CIII_ASSEMBLY	-2.1200	0.0020	0.3926
REACTOME_FORMATION_OF_ATP_BY_CHEMIOSMOTIC_COUPLING	-2.1414	0.0021	0.3926
WP_HISTONE_MODIFICATIONS	1.4327	0.0022	0.3926
REACTOME_BUDDING_AND_MATURATION_OF_HIV_VIRION	-2.2759	0.0024	0.3926
REACTOME_CRISTAE_FORMATION	-2.4379	0.0026	0.3926
BIOCARTA_PROTEASOME_PATHWAY	-2.0874	0.0027	0.3926
REACTOME_ENDOSOMAL_SORTING_COMPLEX_REQUIRED_FOR_TRANSPORT_ESCRT_	-2.2125	0.0029	0.3926



Figure 4. Enrichment plots for gene set enrichment analysis. (A) CYP2E1 reactions, (B) protein export, (C) mitochondrial_ciii_assembly,
 (D) formation of ATP by chemiosmotic coupling, (E) histone modification, (F) budding and maturation of HIV virion, (G) cristae formation, (H) biocarta proteasome pathway, and (I) endosomal sorting complex required for transport. NES – normalized enrichment score; FDR – false discovery rate

correlation between SLC26A4-AS1 and TFH (**Figures 6N and 7**). For Th1 cells, the mean level of the SLC26A4-AS1 low-expression group (0.304 ± 0.048) was significantly lower than that of the high-expression group (0.317 ± 0.05) (P<0.001) (**Figure 50**). The results (r=0.180, P<0.001) indicated a positive correlation between SLC26A4-AS1 and Th1 cells (**Figures 60 and 7**).

Discussion

A growing number of studies have shown that lncRNAs play an important role in tumorigenesis and development [4]. The lncRNA MALAT1 is used as a potential indicator for early diagnosis and prognosis prediction in patients with BC [8]. The serum lncRNA ATB is a noninvasive diagnostic marker in the early stages of BC [9]. INC00978 is an oncogene in BC and can



Figure 5. Expression of SLC26A4-AS1 correlated with immune cells in patients with breast cancer (grouped comparison chart). (A) aDC, (B) B cells, (C) CD8 T cells, (D) cytotoxic cells, (E) DC, (F) macrophages, (G) neutrophils, (H) NK CD56bright cells, (I) NK CD56dim cells, (J) T cells, (K) T helper cells, (L) Tcm, (M) Tem, (N) TFH, and (O) Th1 cells.

e934522-11



Figure 6. Correlation of SLC26A4-AS1 expression with 24 immune cells in patients with breast cancer (scatter plot). (A) aDC, (B) B cells, (C) CD8 T cells, (D) cytotoxic cells, (E) DC, (F) macrophages, (G) neutrophils, (H) NK CD56bright cells, (I) NK CD56dim cells, (J) T cells, (K) T helper cells, (L) Tcm, (M) Tem, (N) TFH, and (O) Th1 cells.

e934522-12



Figure 7. Correlation of SLC26A4-AS1 expression with 24 immune cells in patients with breast cancer (lollipop chart).

be used as a potential prognosis biomarker for patients with BC [10]. Low expression of LINC00944 is associated with poor prognosis in patients with BC [27]. Therefore, it is crucial to explore promising novel lncRNAs as prognostic biomarkers for BC.

The low expression of SLC26A4-AS1 in thyroid cancer was significantly associated with poor patient prognosis [11]. SLC26A4-AS1 is downregulated in human glioma tissues and cells [12]. In the present study, low expression of SLC26A4-AS1 in patients with BC was correlated with age (P < 0.001), ER status (P < 0.001), PAM50 (P<0.001), and menopause status (P<0.001). BC tissue expressed less SLC26A4-AS1 than did adjacent normal breast tissue, especially in patients with older age (>60), indicating that SLC26A4-AS1 might inhibit oncogenesis and promote proliferation. Positive ER status and luminal A/luminal B/HER2 status might be the consequence of a bulky tumor burden due to low expression of SLC26A4-AS1. Low SLC26A4-AS1 expression predicted a poorer OS (HR: 0.56; 95% CI: 0.40-0.78; P=0.001) and DSS (HR: 0.57; 95% CI: 0.37-0.88; P=0.011). Also, SLC26A4-AS1 expression (HR: 0.298; 95% CI: 0.154-0.579; P<0.001) was an independent prognostic factor for poor OS in patients with BC.

SLC26A4-AS1 interacts with DDX5 and the E3 ligase TRIM25 to promote DDX5 degradation via the ubiquitin-proteasome pathway [11]. SLC26A4-AS1 is downregulated in human glioma tissues and cells [12]. SLC26A4-AS1 promotes the transcriptional activity of NPTX1 through the recruitment of NFKB1, thereby exerting an anti-angiogenic effect on glioma cells [12]. Overexpression of lncRNA SLC26A4-AS1 exerts anticancer

effects on papillary thyroid carcinoma through inactivation of the MAPK pathway [28]. In the present study, SLC26A4-AS1 was related to the following pathways: CYP2E1 reactions, protein export, mitochondrial_ciii_assembly, formation of ATP by chemiosmotic coupling, histone modification, budding and maturation of HIV virion, cristae formation, biocarta proteasome pathway, and endosomal sorting complex required for transport.

The TME, consisting of tumor-infiltrating lymphocytes and other immune cells, is an important part of tumor development [29,30]. The number and proportion of mesenchymal and immune cells in tumor tissue are associated with clinical features and prognosis [31]. Screening valuable genes from immune and stromal cells in tumor tissues may be a target for cancer therapy [32]. Therefore, there is a need to gain insight into the significance of genes associated with the TME for accurate assessment and treatment. The present study investigated the relationship between SLC26A4-AS1 expression and the level of multiple immune infiltrations in BC. The results showed modest correlations between SLC26A4-AS1 expression and infiltration levels of aDC, B cells, CD8 T cells, cytotoxic cells, DC, eosinophils, macrophages, neutrophils, NK CD56bright cells, NK CD56dim cells, T cells, T helper cells, Tcm, Tem, TFH, Tgd, Th1 cells, and TReg in BC. The present findings suggest that SLC26A4-AS1 may mediate the development and progression of BC by negatively regulating eosinophils and NK CD56bright cells and positively regulating aDC, B cells, CD8 T cells, cytotoxic cells, DCs, macrophages, neutrophils, NK CD56dim cells, T cells, T helper cells, Tcm, Tem, TFH, and Th1 cells.

This study is the first to examine the relationship between SLC26A4-AS1 and BC. However, there are some limitations. This study was conducted based on the TCGA database and bioinformatics analysis, but the specific mechanism by which SLC26A4-AS1 mediates the development and progression of BC could not be clarified. In the real world setting, the clinical significance and specific mechanisms of SLC26A4-AS1 in patients with BC need to be further investigated.

Conclusions

SLC26A4-AS1 expression was significantly lower in BC tissue than in normal breast tissue. Low expression of SLC26A4-AS1 was associated with poor OS and DSS in patients with BC. SLC26A4-AS1 may be related to the following pathways: CYP2E1 reactions, protein export, mitochondrial_ciii_assembly, formation of ATP by chemiosmotic coupling, histone modification,

References:

- 1. Zhao E, Lan Y, Quan F, et al. Identification of a six-IncRNA signature with prognostic value for breast cancer patients. Front Genet. 2020;11:673
- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. Cancer J Clin. 2018;68(6):394-424 [Erratum in: Cancer J Clin. 2020;70(4):313]
- Li Y, Liang Y, Ma T, Yang Q. Identification of DGUOK-AS1 as a prognostic factor in breast cancer by bioinformatics analysis. Front Oncol. 2020;10:1092
- Zhou T, Lin K, Nie J, et al. LncRNA SPINT1-AS1 promotes breast cancer proliferation and metastasis by sponging let-7 a/b/i-5p, Pathol Res Pract. 2021;217:153268
- 5. Hu RH, Zhang ZT, Wang HX, et al. LncRNA ST7-AS1, by regulating miR-181b-5p/KPNA4 axis, promotes the malignancy of lung adenocarcinoma. Cancer Cell Int. 2020;20:568-68
- Xu M, Chen X, Lin K, et al. The long noncoding RNA SNHG1 regulates colorectal cancer cell growth through interactions with EZH2 and miR-154-5p. Mol Cancer. 2018;17:141
- 7. Shi T, Gao G, Cao Y. Long Noncoding RNAs as novel biomarkers have a promising future in cancer diagnostics. Dis Markers. 2016;2016:9085195
- Sun Z, Liu J, Liu J. The expression of IncRNA-MALAT1 in breast cancer patients and its influences on prognosis, Cell Mol Biol (Noisy-Le-Grand). 2020;66:72-78
- El-Ashmawy NE, Hussien FZ, El-Feky OZ, et al. Serum LncRNA-ATB and FAM83H-AS1 as diagnostic/prognostic non-invasive biomarkers for breast cancer. Life Sci. 2020;259:118193
- Deng LL, Chi YY, Liu L, et al. LINC00978 predicts poor prognosis in breast cancer patients. Sci Rep. 2016;6:37936
- 11. Yuan J, Song Y, Pan W, et al. LncRNA SLC26A4-AS1 suppresses the MRN complex-mediated DNA repair signaling and thyroid cancer metastasis by destabilizing DDX5. Oncogene. 2020;39:6664-76
- Li H, Yan R, Chen W, et al. Long non coding RNA SLC26A4-AS1 exerts antiangiogenic effects in human glioma by upregulating NPTX1 via NFKB1 transcriptional factor. FEBS J. 2021;288:212-28
- Gao S, Lu X, Ma J, et al. Comprehensive analysis of lncRNA and miRNA regulatory network reveals potential prognostic non-coding RNA involved in breast cancer progression. Front Genetic. 2021;12:621809
- 14. Zhao H, Liu X, Yu L, et al. Comprehensive landscape of epigenetic-dysregulated lncRNAs reveals a profound role of enhancers in carcinogenesis in BC subtypes. Mol Ther Nucleic Acids. 2021;23:667-81
- 15. Dysthe M, Parihar R. Myeloid-derived suppressor cells in the tumor microenvironment. Adv Exp Med Biol. 2020;1224:117-40

budding and maturation of HIV virion, cristae formation, biocarta proteasome pathway, and endosomal sorting complex required for transport. SLC26A4-AS1 expression was associated with several immune infiltrating cells. This study provides a promising prognostic biomarker for patients with BC.

Acknowledgements

The authors thank The Cancer Genome Atlas for providing the data. The authors thank MyGene Diagnostics Co., Ltd. for technology support.

Declaration of Figures' Authenticity

All figures submitted have been created by the authors who confirm that the images are original with no duplication and have not been previously published in whole or in part.

- Topalian SL, Drake CG, Pardoll DM. Immune checkpoint blockade: A common denominator approach to cancer therapy. Cancer Cell. 2015;27:450-61
- 17. De Palma M, Biziato D, Petrova TV. Microenvironmental regulation of tumour angiogenesis. Nat Rev Cancer. 2017;17:457-74
- Burugu S, Asleh-Aburaya K, Nielsen TO. Immune infiltrates in the breast cancer microenvironment: Detection, characterization and clinical implication. Breast Cancer. 2017;24:3-15
- Lu X, Li G, Liu S, et al. Bioinformatics analysis of KIF1A expression and gene regulation network in ovarian carcinoma. Int J Gen Med. 2021;14:3707-17
- Chen T, Zhu C, Wang X, Pan Y. LncRNA ELF3-AS1 is a prognostic biomarker and correlated with immune infiltrates in hepatocellular carcinoma. Can J Gastroenterol Hepatol. 2021;2021:8323487
- Liu J, Lichtenberg T, Hoadley KA, et al. An integrated TCGA pan-cancer clinical data resource to drive high-quality survival outcome analytics. Cell. 2018;173:400-416.e411
- Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. Genome Biol. 2014;15:550
- 23. Yu G, Wang LG, Han Y, He QY. clusterProfiler: An R package for comparing biological themes among gene clusters. OMICS. 2012;16:284-87
- Subramanian A, Tamayo P, Mootha VK, et al. Mesirov, gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles, Proc Natl Acad Sci USA. 2005;102:15545-50
- Hänzelmann S, Castelo R, Guinney J. GSVA: Gene set variation analysis for microarray and RNA-seq data. BMC Bioinformatics. 2013;14:7
- Bindea G, Mlecnik B, Tosolini M, et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. Immunity. 2013;39:782-95
- de Santiago PR, Blanco A, Morales F, et al. Immune-related IncRNA LINC00944 responds to variations in ADAR1 levels and it is associated with breast cancer prognosis. Life Sci. 2021;268:118956
- Wang DP, Tang XZ, Liang QK et al. Overexpression of long noncoding RNA SLC26A4-AS1 inhibits the epithelial-mesenchymal transition via the MAPK pathway in papillary thyroid carcinoma. J Cell Physiol. 2020;235:2403-13
- 29. Junttila MR, de Sauvage FJ. Influence of tumour micro-environment heterogeneity on therapeutic response. Nature. 2013;501:346-54
- Wang Z, Song K, Zhao W, Zhao Z. Dendritic cells in tumor microenvironment promoted the neuropathic pain via paracrine inflammatory and growth factors. Bioengineered. 2020;11:661-78
- Zhou ZJ, Xin HY, Li J, et al. Intratumoral plasmacytoid dendritic cells as a poor prognostic factor for hepatocellular carcinoma following curative resection. Cancer Immunol Immunother. 2019;68:1223-33
- Ren H, Hu D, Mao Y, Su X. Identification of genes with prognostic value in the breast cancer microenvironment using bioinformatics analysis. Med Sci Monit. 2020;26:e920212