

High Prevalence of *vanM* in Vancomycin-Resistant *Enterococcus* faecium Isolates from Shanghai, China

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The vanM gene was first found in a vancomycin-resistant *Enterococcus faecium* (VREm) isolate in Shanghai in 2006. In this study, we found that, in 70 VREm strains isolated in nine Shanghai hospitals from 2006 to 2014, vanM was more prevalent than the vanA gene (64.3% [45/70] versus 35.7% [25/70]). The vanM-type isolates showed similar antimicrobial susceptibility patterns with the vanA types. The vanM-type VREm emerged and disseminated in Shanghai.

The isolation of vancomycin-resistant enterococci (VRE) was first reported in 1988 (1, 2). During the last 2 decades, VRE have become significant nosocomial pathogens worldwide, mainly due to their adaptability in hospital environments and the limited treatment options. Nine types of glycopeptide resistance determinants (*vanA*, *vanB*, *vanC*, *vanD*, *vanE*, *vanG*, *vanL*, *vanM*, and *vanN*) have been reported and well characterized on the basis of phenotypic and genotypic criteria (3). The *vanA and vanB* genotypes predominate worldwide (3, 4).

We first reported the *vanM* gene in a vancomycin-resistant *Enterococcus faecium* (VREm) clinical isolate from a teaching hospital in Shanghai in 2006 (5). Subsequently, only a single study from Singapore has reported *vanM*-type VRE isolates (6). Epidemiology data for strains with *van* determinants other than *vanA* and *vanB* remain rare. In this study, we investigated the prevalence of *van* and virulence genes in VREm strains isolated from 9 hospitals in Shanghai. Pulsed-field gel electrophoresis (PFGE) and multilocus sequence type (MLST) were also performed to elucidate the molecular epidemiology of these strains.

Seventy consecutive and nonduplicate VREm clinical strains were collected from 9 hospitals in Shanghai between 2006 and 2014. MICs of 10 antimicrobial agents (vancomycin, teicoplanin, linezolid, fosfomycin, ampicillin, erythromycin, levofloxacin, gentamicin, minocyline, and rifampin) were determined by agar dilution. Etest (bioMérieux) was used to determine the MICs of tigecycline. Susceptibility to daptomycin was determined by broth microdilution using Mueller-Hinton II broth (cation adjusted) supplemented with calcium 50 μ g/ml. Results were interpreted using the 2012 guidelines of the Clinical and Laboratory Standards Institute (www.clsi.org; Wayne, PA, USA). Due to the lack of an acknowledged fosfomycin breakpoint for *E. faecium*, we used the breakpoints of fosfomycin for *Enterococcus faecalis* proposed by the CLSI. *E. faecalis* ATCC 29212 was used as a quality-control strain for MIC determination.

Vancomycin resistance genes were detected by PCR amplification, as previously described (5). The PCR products were sequenced to determine the particular *van* genotype. The presence of five virulence genes (*asa1*, *gelE*, *cylA*, *esp*, and *hyl*) was assayed by multiplex PCR, as described previously (7). PFGE analysis was performed using a contour-clamped homogeneous electric field (CHEF) mapper system (Bio-Rad, USA), as previously described (8). Banding patterns were analyzed with BioNumerics software, version 5.0 (Belgium). Isolates were categorized into the same PFGE pulsotype group if they shared more than 80% similarity. MLST analysis was performed as described previously (9). Alleles and sequence types (STs) were analyzed and determined via the MLST database (http://efaecium.mlst.net/). Clusters of related STs were grouped into clonal complexes (CCs) using the eBURST program, version 3 (http://efaecium.mlst.net/eburst/). Statistical analysis was performed with the chi-square test or Fisher's extract test, as appropriate, using the statistical program SPSS 22.0. A *P* value of ≤ 0.05 was considered statistically significant.

Among the 70 VREm isolates, 45 strains (64.3%) carried the *vanM* gene, and 25 isolates (35.7%) harbored *vanA*. No other *van* genes were found. The *vanM*-type VREm isolates were detected in 8 hospitals located at the center of Shanghai city. The *vanM* gene has been predominant in VREm strains in Shanghai since 2011 (Fig. 1).

The *vanM*-type *E. faecium* isolates showed similar antimicrobial susceptibility patterns to the *vanA*-type isolates. All 70 VREm isolates were resistant to vancomycin (MICs, 128 to >256 μ g/ml) and levofloxacin, and all were susceptible to linezolid, daptomycin, and tigecycline. The teicoplanin resistance rates were 71.1% (32/45) in *vanM*-type and 84.0% (21/25) in *vanA*-type VREm isolates. The gentamicin resistance rates were 64.4% and 76% in *vanM*-type and *vanA*-type isolates, respectively. No statistically significant differences in susceptibility to the 12 antimicrobial agents were observed between *vanM*- and *vanA*-type strains (Table 1).

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FIG 1 Distribution of vancomycin-resistant genes in 70 VREm strains isolated from nine hospitals, Shanghai, China, 2006 to 2014.

Five different pulsotypes were found among the 70 VREm strains, and each pulsotype included strains from at least 2 different hospitals (Fig. 2). By MLST analysis, 12 sequence types (STs) were identified, including ST 17 (n = 3), ST 18 (n = 2), ST 78 (n = 46), ST 203 (n = 2), ST 252 (n = 1), ST 262 (n = 2), ST 290 (n = 1), ST 341 (n = 1), ST 389 (n = 1), ST 555 (n = 7), ST 564 (n = 3), and ST 881 (n = 1). ST 881 is a new sequence type found in this study, and the data were uploaded to the eBURST database.

eBURST analysis showed that all of the 70 VREm isolates belonged to clonal complex (CC) 17.

The *esp* gene was present 97.8% (44/45) and 84% (21/25) of *vanM*-type and *vanA*-type isolates, respectively (P = 0.033). The *hyl* gene was detected in 17.8% (6/45) and 32% (8/25) of *vanM*-type and *vanA*-type isolates, respectively (P = 0.063). All strains were negative for the presence of *cylA*, *gelE*, and *asa1* virulence genes.

TABLE 1 Comparison of the MICs	of 12 antimicrobial agents between vanA-	 and vanM-type VREm isolates
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	<i>vanA</i> -type VREm ($n = 25$)				vanM-type VREm ($n = 45$)				
	MIC (µg/ml) data:			MIC (µg/ml) data:					
Antibacterial agent	Range	MIC ₅₀	MIC ₉₀	\mathbf{R}^{a} (%)	MIC range	MIC ₅₀	MIC ₉₀	\mathbb{R}^{a} (%)	Р
Vancomycin	128 to >256	>256	>256	100	128 to >256	>256	>256	100	NA^b
Teicoplanin	0.5 to >256	32	128	71.1	0.5->256	64	128	84	0.232
Linezolid	1 to 2	1	2	0	1 to 2	1	2	0	NA
Daptomycin	2 to 4	4	4	0	0.5 to 4	4	4	0	NA
Tigecycline	0.032 to 0.094	0.064	0.064	0	0.032 to 0.125	0.064	0.094	0	NA
Ampicillin	0.5 to >256	>256	>256	97.8	0.5 to >256	>256	>256	96	0.671
Levofloxacin	32 to >256	64	>256	100	32 to >256	64	128	100	NA
Erythromycin	0.125 to >256	>256	>256	91.1	0.125 to >256	>256	>256	92	0.899
Fosfomycin	64 to >512	64	>512	26.7	16 to >512	64	>512	24	0.808
Rifampin	2 to >256	8	16	82.2	≤ 0.06 to 16	8	16	80	0.820
Minocycline	≤ 0.06 to 32	0.125	16	15.6	≤ 0.06 to 32	0.125	16	16	0.961
Gentamicin	4 to >256	>256	>256	64.4	2 to >256	>256	>256	76	0.322

^a Resistance rate.

^b Not applicable.

- \$ 8 8 8 8 8 8 8		Isolates	Date of isolation	Hospitals	MLST	Specimens	Genotype of van	Virulence genes
		1	2006-1-27	HS	ST17	Exudate	vanA	esp. hyl
- L		7	2007-7-9	HS	ST78	Urine	vanM	
i je		9	2008-2-5	HS	ST78	Urine	vanM	1.1110-045
		19	2010-8-3	HS	ST252	Blood	vanA	esp, hyl
1 1 1		10	2008-8-5	HS	ST341	Urine	vanM	esp
		17	2010-7-7	HS	ST78	Urine	vanA	esp. hyl
_P		20	2010-11-10	HS	ST78	Urine	vanA	esp, hyl
U L		18	2010-7-15	HS	ST78	Urine	vanA	esp, hyl
卢노		6	2007-6-2	JS	ST78	Urine	vanA	esp. hyl
		25	2011-5-19	HS	ST78	Bile	vanA	eso, hu >
	11111 111010	38	2012.10.11	HS	ST17	Urice	vanA	050
		23	2012-10-11	01	OTTO	Theost cursh	vanA	
		26	2011-4-2	RJ	5170	Analauah	vanA	esp
1 1 4		24	2011-6-15	RJ	5176	Ariai swab	Manual	esp
		24	2011-4-2	RJ	51555	Throat swab	varies	esp
	ini nittimu	21	2011-1-21	RJ	ST555	Anal swab	vaniw	esp
	1.1.1.1111	22	2011-2-25	RJ	ST78	Blood	vanM	esp
1 1		2	2006-2-4	HS	ST78	Exudate	vanM	esp, hyl
1 1		5	2007-3-9	HS	ST78	Urine	vanM	,
r (48	2013-5-7	CZ	ST78	Urine	vanM	esp)
1		63	2013-12-19	JS	ST78	Blood	vanA	esp
		65	2014-1-17	JS	ST78	Bile	vanA	esp
- 1,		53	2013-7-16	JS	ST78	Stool	vanA	esp, hyl
/ł		47	2013-4-29	RX	ST78	Throat swab	vanM	esp
		61	2013.11.16	21,	ST78	Blood	vanA	esp. hvi
		64	2013-11-15	P7	STEEL	Urine	vanM	050
		03	2013-12-20	PI	ST70	Blood	vanM	eso hu
		66	2014-2-18		0170	Union	vanM	
	men house	00	2014-2-13	PCJ	5178	Onne	vanite	esp
1 1 1		45	2013-4-8	zs	5178	Biood	Varies	esp
	.11 .1111.000	12	2009-1-7	HS	ST78	Urine	vanA	esp, hyl
		60	2013-10-15	RJ	ST78	Blood	vanM	esp
		54	2013-8-26	HS	ST78	Blood	vanM	esp
[] L		62	2013-11-16	RJ	ST18	Blood	vanA	·
l r		58	2013-9-29	RJ	ST78	Throat swab	vanM	esp
		59	2013-9-29	RJ	ST78	Blood	vanM	esp
į L		57	2013-9-23	RJ	ST78	Anal swab	vanA	esp /
-	111110100	67	2014-2-13	CZ	ST78	Urine	vanM	esp >
		15	2009-5-21	HS	ST203	Urine	vanA	esp
		14	2009.3.27	HS	ST290	Urine	vanA	050
		52	2013.7.9	PI	ST78	Anal swah	vanM	050
		31	2013-7-3	LIC	OTTEL	licios	vanM	
이 그는 눈눈ㅋㅋ			2012-2-0	ns Ol	01004	Uning	Manu	and a
		10	2013-9-18	KJ	51555	Unine	vante	esp >
1 1 4 5		10	2009-11-7	HS	S117	Unne	varies	esp
		68	2014-2-14	RJ	ST78	Urine	vanM	esp
		4	2007-2-4	HS	ST203	Urine	vanA	esp, hyl
		70	2014-4-2	RJ	ST262	Anal swab	vanM	esp
		13	2009-3-1	HS	ST389	Urine	vanA	esp
		8	2008-2-2	HS	ST78	Urine	vanM	esp)
		56	2013-9-23	RJ	ST78	Anal swab	vanA	esp
		49	2013-6-14	PR	ST564	Urine	vanM	esp >
		39	2012-11-8	CZ	ST555	Urine	vanM	esp)
		44	2013-2-28	HS	ST78	Urine	VanM	esp
		40	2013 12 14	C7	STEEL	Unice	Mag	050
		33	2012-12-11	OL.	01000	Line	vanA	050
h 1 1		35	2012-6-1	10	51/8	Util0	unald	ash
		35	2012-9-1	RJ	\$178	Inroat swab	vanwi	esp
		11	2008-10-22	HS	5118	Unne	varini	nya.
		34	2012-8-1	RJ	ST555	Urine	vanA	esp
1 1		30	2012-2-1	RJ	ST78	Urine	vanM	esp
		42	2013-1-1	RJ	ST78	Urine	vanM	esp
		36	2012-9-12	HS	ST78	Exudat	vanM	esp
		27	2011-12-1	RJ	ST78	Anal swab	VanM	esp
Ph		29	2012-1-20	RJ	ST78	Exudate	vanM	esp
		50	2013-6-22	HS	ST78	Urine	vanM	esp
		37	2012-10-1	RI	ST262	Catheter	Mac	050
		28	2012-10-1	PI	STTP	Blood	vanM	050
		2	2011-12-1	ev.	0170	Line	VanM	
		5	2006-11-13	ST	51/8	Unite	upphi	050
		40	2013-4-13	02	\$1881	Unne	Varian	esp
1 6		43	2013-2-1	RJ	\$1555	Unne	vanim	650
1 L		51	2013-7-9	RJ	S178	Bde	vanM	esp
	II II IN INTERACTOR							-
		41	2012-12-19	CZ	ST78	Urine	vanM	esp

FIG 2 Strains particulars and PFGE dendrogram of the 70 VREm isolates from nine hospitals in Shanghai. Detailed information of the isolated dates, hospitals, specimen sources, MLST, *van* genotypes, and virulence genes are listed for each isolate. Pulsotypes A through E are clustered based on 80% similarity of the PFGE pattern.

Previous studies found that *vanA* is the most frequently encountered genotype of VREm in Asia, as in other countries worldwide (10-12). This study, however, showed that the *vanM* genotype has predominated in VREm clinical isolates in Shanghai since 2011. Similar to *vanA*-type VREm strains, *vanM*-type VREm strains are multidrug resistant, belong to CC17, and carry virulence genes *esp* and *hyl*, which provide these VREm strains more advantages to adapt to the hospital environment. Data from annual bacterial resistance surveillance program in Shanghai, China, showed that vancomycin resistance strains in *E. faecium* (VREm) increased from 0.33% in 2006 to 1.62% in 2011 and to 1.95% in 2014 (unpublished data). Thus, the high prevalence of *vanM* might contribute to the increasing VRE prevalence in Shanghai. PFGE analysis indicated that the *vanM* gene spread among diverse VRE strains in different hospitals instead of as a single clone.

The *vanM* gene was first found in a VREm clinical isolate from our hospital in Shanghai in 2006 (5). In 2011, Teo et al. reported a *vanM*-type *E. faecium* clinical strain in Singapore (6), thus indicating that this new vancomycin resistance gene might spread to other countries.

One of the reasons for the rarity of *vanM*-type VREm strains might be that most clinical laboratories and commercial molecular detection kits (Cepheid, Bouwel, Belgium; BD Diagnostics-GeneOhm, San Diego, CA) mainly focus on *vanA* and *vanB* genes and do not include the *vanM* gene (13, 14). In a study conducted in Mexico, one isolate of *E. faecium* demonstrated high-level resistance to vancomycin and teicoplanin, but it was classified as non-*vanA*, non-*vanB* isolate (15), which suggests that detection of new vancomycin resistance genes, such as *vanM*, might be missed based on current screening methods.

Overall, the results presented here suggest that *vanM* gene plays an important role in vancomycin resistance and dissemination in *E. faecium* strains in Shanghai. Therefore, it is necessary to screen for *vanM* in *E. faecium* strains to better control *vanM*-type VREm infection and dissemination.

New eBURST sequence type. ST 881 is a new sequence type found in this study, and the data were uploaded to the eBURST database.

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