S6K in geroconversion

Olga V Leontieva*, Zoya N Demidenko, and Mikhail V Blagosklonny* Department of Cell Stress Biology; Roswell Park Cancer Institute; Buffalo, NY USA

Keywords: cell cycle, senescence, aging, cancer, growth, rapalogs

Submitted: 06/20/2013

Revised: 08/15/2013

Accepted: 08/21/2013

http://dx.doi.org/10.4161/cc.26248

*Correspondence to: Olga V Leontieva; Email: olga.leontieva@roswellpark.org; Mikhail V Blagosklonny; Email: Blagosklonny@oncotarget.com; mikhail.blagosklonny@roswellpark.org

Markers of cellular senescence depend in part on the MTOR (mechanistic target of rapamycin) pathway. MTOR participates in geroconversion, a conversion from reversible cell cycle arrest to irreversible senescence. Recently we demonstrated that hyperinduction of cyclin D1 during geroconversion was mostly dependent on MEK, whereas rapamycin only partially inhibited cyclin D1 accumulation. Here we show that, while not affecting cyclin D1, siRNA for p70S6K partially prevented loss of RP (replicative/regenerative potential) during p21-induced cell cycle arrest. Similarly, an inhibitor of p70 S6 kinase (PF-4708671) partially inhibited phosphorylation of S6 and preserved RP, while only marginally prevented cyclin D1 induction. Thus S6K and MEK play different roles in geroconversion.

Cell cycle arrest is not yet senescence.¹ To induce senescence in cell culture, cells first need to be arrested by different means such as telomere shortening, DNA damage, cytotoxic stresses, as well as strong oncogenic stimulation (Ras and Raf), which induce cell cycle arrest by induction of CDK inhibitors such as p21 and p16.2-14 Importantly, cells become arrested in growth-promoting conditions (in the presence of serum, nutrients, and oxygen, which all activate MTOR, like in proliferating cells). At first, arrested cells are not senescent. Yet, still active MTOR initiates the conversion to senescence, named gerogenic conversion or geroconversion.¹ Under the pressure of MTOR, cells acquire markers of senescence: hypertrophy (a large, flat cell morphology), cellular hyper-functions, including hyper secretion of cytokines, hyper-elevated levels of cyclins D1 and E and loss of regenerative/

replicative potential (RP), i.e., the ability to resume proliferation when cell cycle is released (Fig. 1A). The process of geroconversion is a proper target for suppression of senescence without abrogating cell cycle arrest. Inhibition of MTOR by rapamycin, p53, hypoxia, and MEK inhibitors suppresses geroconversion, preserving RP (Fig. 1B) and preventing other markers of senescence (in cell type-dependent manner, in the case of hypoxia, p53, and MEK inhibitors).15-25 Recently, we demonstrated that MEK inhibitors completely prevented induction of cyclin D1, even when MTOR remained fully activated.²⁶ In contrast, the effect of rapamycin on cyclin D1 was modest compared with the complete elimination of cyclin D1 by MEK inhibition. The MTOR pathway was mostly responsible for loss of RP and hypertrophy.²⁶ p70 S6 kinase 1 (S6K1) is a crucial substrate of MTOR given that knockdown of S6K1 extends lifespan in mice.²⁷ Here we compared consequences of inhibition of MEK²⁶ with inhibition of S6K1, using RNAi technology (Fig. 2).

siRNA for S6K1 decreased level of phosphorylated p70S6K1 (**Fig. 2A**). Both siRNA for MEK1 and S6K1 decreased acidification of cell culture medium as evident by reddish color compared with yellow medium in control cells (**Fig. 2B**), reflecting inhibition of lactic acid production.²⁸ siRNA for MEK1 and S6K1 also decreased cell size, as was measured by the amount of protein per cell (**Fig. 2B**). This effect was especially prominent with siRNA for S6K1 (**Fig. 2B**). Finally, both siRNAs for MEK and S6K1 preserved RP in HT–p21 cells treated with IPTG (**Fig. 2C**).

We next used small-molecule kinase inhibitors (Fig. 3). As expected, both



Figure 1. How to measure geroconversion and gerosuppression. (**A**) Geroconversion (conversion from arrest to senescence). In proliferating cells, the MTOR pathway is active (especially in malignant cells used as a model). When the cell cycle is arrested, MTOR drives geroconversion (during 3–5 days in cell culture conditions). Senescent cells cannot proliferate after abrogation of cell cycle arrest (release). As a particular example, cells expressing ectopic IPTG-inducible p21 can be arrested by addition of IPTG.57 When IPTG is removed, then the cells are released. (**B**) Gerosuppression. Inhibition of the MTOR pathway suppresses geroconversion. Cells resume proliferation, when cell cycle is released. A number of colonies or cells may serve as a quantification of gerosuppression (determined by dividing a number of colonies [or cells] in (**B**) by respective numbers in panel (**A**). B/A = regenerative or replicative potential (RP).



Figure 2. Effects of siRNA for MEK and S6K1 on senescence. (**A**) HT–p21cells transfected with siRNA for MEK1 or p70S6K1 or with lipofectamine alone were lysed 4 days after transfection and immunoblotted with the indicated antibodies. (**B**) HT–p21 cells were transfected with siRNA for MEK1 or S6K1 or with lipofectamine alone (Mock). Next day cells were trypsinized and plated at low density. After 6 days in culture, wells were photographed for the color of the media, trypsinized, and counted, then lysed. Protein amount per cell was determined (shown as pg/cell). (**C**) Regenerative/ replicative potential (RP). HT–p21 cells, transfected with siRNA for MEK1 or S6K1 or with lipofectamine alone (Mock), were split 4 days after transfection and treated with IPTG for 3 days. (Note: IPTG causes cell cycle arrest in HT–p21 cells by inducing ectopic p2157). Then IPTG was washed out, and colonies were grown for 10 days and stained with Crystal Violet. A number of colonies is presented as mean \pm SD.

rapamycin and everolimus prevented loss of RP in HT-p21 cells (Fig. 3A). This potent gerossupressive effect can explain life-extending and anti-aging effects of rapamycin in diverse species,²⁹ including yeast,³⁰ worm,³¹ flies,³²⁻³⁴ and

mice.35-45 Treatment with inhibitors of S6K (PF-478671)⁴⁶ and MEK (PD184352), especially at concentration 10 µM, preserved RP of IPTG-treated HT-p21 cells (Fig. 3A). These results were consistent with the effects of siRNAs for S6K1 and MEK (Fig. 2C). In addition, we tested inhibitors of several related pathways: p90/RSK (SL 0101-1 and BRD7389), phospholipase D2 or PLD2 (halopemide), and JNK (SP600125). PLD2 is known to activate the MTOR/S6K pathway.47-50 P90/RSK was chosen as a target, because it phosphorylates S6 independently from the MTOR pathway. Lastly, JNK is involved in aging and age-related pathology in Drosophila.51-54 However, effects of these inhibitors on gerosuppression were insignificant (data not shown).

In agreement with our recent work,²⁸ MEK inhibitors (PD184352 and U0126) eliminated cyclin D1, whereas effect of rapamycin on cyclin D1 accumulation was incomplete, and cyclin D1 was still visible on the longer exposured blot (Fig. 3B). Inhibitor of S6K (PF478671) only slightly decreased levels of cyclin D1, indicating that effects of rapamycin on cyclin D1 accumulation may involve different pathways. In fact, it was shown that MTOR increases cyclin D1 through inactivation of 4EBP1.55,56 We also investigated 2 inhibitors of p90 RSK (SL 0101-1 and BRD7389). These inhibitors did not affect MTOR pathway and just slightly decreased cyclin D1 (Fig. 3B). Compared with SL 0101-1, BRD7389 exerted a stronger effect on cyclin D1, which could be due to its toxicity at concentration 10 µM. Halopemide affected neither phosphorylation of S6 nor cyclin D1 levels. Thus, we identified p70 S6K as a target for gerosuppression, yet an inhibitor of p70 S6K did not decrease cyclin D1 levels. In contrast, MEK inhibitor was extremely effective in prevention of cyclin D1 accumulation, confirming our conclusion that markers of senescence can be dissociated. At standard concentration of 10 µM, inhibitors of p90 ribosomal S6 kinase or RSK (SL 0101-1 and BRD7389), Jun N-terminal kinase (SP600125), and phospholipase D2 (halopemide) failed to suppress geroconversion in this preliminary assessment. It is possible that, while ineffective as



Figure 3. Effects of kinase inhibitors on senescence. (**A**) RP. HT–p21 cells were treated with IPTG and with indicated drugs. Rapa, rapamycin 500 nM; Eve, Everolimus 500 nM; PF, PF4708671; PD, PD184352: 10; 30; 100 μ M. After 3 days, drugs were washed out and cells were allowed to grow. A number of colonies represents RP. (**B**) Immunoblot. HT–p21 cells were treated as indicated for 24 h and then lysed. Immunoblotting was performed with the indicated antibodies. R – rapamycin 500 nM; All other drugs were used at 10 μ M. U, U0126; PF, PF4708671; PD, PD184352; BRD, BRD7389; SL, SL 0101-1; Ha, Halopemide.

single drugs in this cell line, SL 0101-1, and SP600125 might potentiate effects of other gerosupressants and be effective in drug combinations as a cocktail of inhibitors at low doses. Finally, the effects of gerosuppressants may be cell type-specific and detailed study is under way.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

This "Extra View" is an addition to our recent publication in *Cell Death Differen-tiation*, as specified by the instruction to authors of *Cell Cycle*.

References

- Blagosklonny MV. Cell cycle arrest is not yet senescence, which is not just cell cycle arrest: terminology for TOR-driven aging. Aging (Albany NY) 2012; 4:159-65; PMID:22394614
- Serrano M, Lin AW, McCurrach ME, Beach D, Lowe SW. Oncogenic ras provokes premature cell senescence associated with accumulation of p53 and p16INK4a. Cell 1997; 88:593-602; PMID:9054499; http://dx.doi.org/10.1016/S0092-8674(00)81902-9
- Lin AW, Barradas M, Stone JC, van Aelst L, Serrano M, Lowe SW. Premature senescence involving p53 and p16 is activated in response to constitutive MEK/MAPK mitogenic signaling. Genes Dev 1998; 12:3008-19; PMID:9765203; http://dx.doi. org/10.1101/gad.12.19.3008
- Morisaki H, Ando A, Nagata Y, Pereira-Smith O, Smith JR, Ikeda K, Nakanishi M. Complex mechanisms underlying impaired activation of Cdk4 and Cdk2 in replicative senescence: roles of p16, p21, and cyclin D1. Exp Cell Res 1999; 253:503-10; PMID:10585273; http://dx.doi.org/10.1006/ excr.1999.4698

- Ferbeyre G, de Stanchina E, Lin AW, Querido E, McCurrach ME, Hannon GJ, Lowe SW. Oncogenic ras and p53 cooperate to induce cellular senescence. Mol Cell Biol 2002; 22:3497-508; PMID:11971980; http://dx.doi.org/10.1128/ MCB.22.10.3497-3508.2002
- Serrano M, Blasco MA. Putting the stress on senescence. Curr Opin Cell Biol 2001; 13:748-53; PMID:11698192; http://dx.doi.org/10.1016/ S0955-0674(00)00278-7
- Itahana K, Dimri G, Campisi J. Regulation of cellular senescence by p53. Eur J Biochem 2001; 268:2784-91; PMID:11358493; http://dx.doi. org/10.1046/j.1432-1327.2001.02228.x
- Ben-Porath I, Weinberg RA. When cells get stressed: an integrative view of cellular senescence. J Clin Invest 2004; 113:8-13; PMID:14702100
- Collado M, Blasco MA, Serrano M. Cellular senescence in cancer and aging. Cell 2007; 130:223-33; PMID:17662938; http://dx.doi.org/10.1016/j. cell.2007.07.003
- Serrano M. Dissecting the role of mTOR complexes in cellular senescence. Cell Cycle 2012; 11:2231-2; PMID:22714590; http://dx.doi.org/10.4161/ cc.21065
- Deschênes-Simard X, Gaumont-Leclerc MF, Bourdeau V, Lessard F, Moiseeva O, Forest V, Igelmann S, Mallette FA, Saba-El-Leil MK, Meloche S, et al. Tumor suppressor activity of the ERK/MAPK pathway by promoting selective protein degradation. Genes Dev 2013; 27:900-15; PMID:23599344; http://dx.doi.org/10.1101/gad.203984.112
- Lopez MF, Tollervey J, Krastins B, Garces A, Sarracino D, Prakash A, Vogelsang M, Geesman G, Valderrama A, Jordan IK, et al. Depletion of nuclear histone H2A variants is associated with chronic DNA damage signaling upon drug-evoked senescence of human somatic cells. Aging (Albany NY) 2012; 4:823-42; PMID:23235539
- Kolesnichenko M, Hong L, Liao R, Vogt PK, Sun P. Attenuation of TORC1 signaling delays replicative and oncogenic RAS-induced senescence. Cell Cycle 2012; 11:2391-401; PMID:22627671; http://dx.doi. org/10.4161/cc.20683
- Pospelova TV, Leontieva OV, Bykova TV, Zubova SG, Pospelov VA, Blagosklonny MV. Suppression of replicative senescence by rapamycin in rodent embryonic cells. Cell Cycle 2012; 11:2402-7; PMID:22672902; http://dx.doi.org/10.4161/cc.20882
- Demidenko ZN, Blagosklonny MV. Growth stimulation leads to cellular senescence when the cell cycle is blocked. Cell Cycle 2008; 7:3355-61; PMID:18948731; http://dx.doi.org/10.4161/ cc.7.21.6919
- Demidenko ZN, Zubova SG, Bukreeva EI, Pospelov VA, Pospelova TV, Blagosklonny MV. Rapamycin decelerates cellular senescence. Cell Cycle 2009; 8:1888-95; PMID:19471117; http://dx.doi. org/10.4161/cc.8.12.8606
- Demidenko ZN, Korotchkina LG, Gudkov AV, Blagosklonny MV. Paradoxical suppression of cellular senescence by p53. Proc Natl Acad Sci U S A 2010; 107:9660-4; PMID:20457898; http://dx.doi. org/10.1073/pnas.1002298107
- Korotchkina LG, Leontieva OV, Bukreeva EI, Demidenko ZN, Gudkov AV, Blagosklonny MV. The choice between p53-induced senescence and quiescence is determined in part by the mTOR pathway. Aging (Albany NY) 2010; 2:344-52; PMID:20606252
- Leontieva OV, Blagosklonny MV. DNA damaging agents and p53 do not cause senescence in quiescent cells, while consecutive re-activation of mTOR is associated with conversion to senescence. Aging (Albany NY) 2010; 2:924-35; PMID:21212465

- Leontieva OV, Natarajan V, Demidenko ZN, Burdelya LG, Gudkov AV, Blagosklonny MV. Hypoxia suppresses conversion from proliferative arrest to cellular senescence. Proc Natl Acad Sci U S A 2012; 109:13314-8; PMID:22847439; http://dx.doi. org/10.1073/pnas.1205690109
- Leontieva OV, Lenzo F, Demidenko ZN, Blagosklonny MV. Hyper-mitogenic drive coexists with mitotic incompetence in senescent cells. Cell Cycle 2012; 11:4642-9; PMID:23187803; http:// dx.doi.org/10.4161/cc.22937
- Leontieva OV, Blagosklonny MV. Hypoxia and gerosuppression: the mTOR saga continues. Cell Cycle 2012; 11:3926-31; PMID:22987149; http://dx.doi. org/10.4161/cc.21908
- 23. Iglesias-Bartolome R, Gutkind SJ. Exploiting the mTOR paradox for disease prevention. Oncotarget 2012; 3:1061-3; PMID:23165441
- Halicka HD, Zhao H, Li J, Lee YS, Hsieh TC, Wu JM, Darzynkiewicz Z. Potential anti-aging agents suppress the level of constitutive mTOR- and DNA damage- signaling. Aging (Albany NY) 2012; 4:952-65; PMID:23363784
- Leontieva OV, Demidenko ZN, Blagosklonny MV. MEK drives cyclin D1 hyperelevation during geroconversion. Cell Death Differ 2013; 20:1241-9; PMID:23852369; http://dx.doi.org/10.1038/ cdd.2013.86
- Selman C, Tullet JM, Wieser D, Irvine E, Lingard SJ, Choudhury AI, Claret M, Al-Qassab H, Carmignac D, Ramadani F, et al. Ribosomal protein S6 kinase 1 signaling regulates mammalian lifespan. Science 2009; 326:140-4; PMID:19797661; http://dx.doi. org/10.1126/science.1177221
- Leontieva OV, Blagosklonny MV. Yeast-like chronological senescence in mammalian cells: phenomenon, mechanism and pharmacological suppression. Aging (Albany NY) 2011; 3:1078-91; PMID:22156391
- Blagosklonny MV. Rapamycin and quasi-programmed aging: four years later. Cell Cycle 2010; 9:1859-62; PMID:20436272; http://dx.doi. org/10.4161/cc.9.10.11872
- Powers RW 3rd, Kaeberlein M, Caldwell SD, Kennedy BK, Fields S. Extension of chronological lifespan in yeast by decreased TOR pathway signaling. Genes Dev 2006; 20:174-84; PMID:16418483; http://dx.doi.org/10.1101/gad.1381406
- Robida-Stubbs S, Glover-Cutter K, Lamming DW, Mizunuma M, Narasimhan SD, Neumann-Haefelin E, Sabatini DM, Blackwell TK. TOR signaling and rapamycin influence longevity by regulating SKN-1/ Nrf and DAF-16/FoxO. Cell Metab 2012; 15:713-24; PMID:22560223; http://dx.doi.org/10.1016/j. cmet.2012.04.007
- Moskalev AA, Shaposhnikov MV. Pharmacological inhibition of phosphoinositide 3 and TOR kinases improves survival of Drosophila melanogaster. Rejuvenation Res 2010; 13:246-7; PMID:20017609; http://dx.doi.org/10.1089/rej.2009.0903
- Bjedov I, Toivonen JM, Kerr F, Slack C, Jacobson J, Foley A, Partridge L. Mechanisms of lifespan extension by rapamycin in the fruit fly Drosophila melanogaster. Cell Metab 2010; 11:35-46; PMID:20074526; http://dx.doi.org/10.1016/j.cmet.2009.11.010
- Partridge L, Alic N, Bjedov I, Piper MD. Ageing in Drosophila: the role of the insulin/Igf and TOR signalling network. Exp Gerontol 2011; 46:376-81; PMID:20849947; http://dx.doi.org/10.1016/j. exger.2010.09.003

- Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature 2009; 460:392-5; PMID:19587680
- 36. Anisimov VN, Zabezhinski MA, Popovich IG, Piskunova TS, Semenchenko AV, Tyndyk ML, Yurova MN, Antoch MP, Blagosklonny MV. Rapamycin extends maximal lifespan in cancer-prone mice. Am J Pathol 2010; 176:2092-7; PMID:20363920; http:// dx.doi.org/10.2353/ajpath.2010.091050
- Longo VD, Fontana L. Intermittent supplementation with rapamycin as a dietary restriction mimetic. Aging (Albany NY) 2011; 3:1039-40; PMID:22147496
- Khanna A, Kapahi P. Rapamycin: killing two birds with one stone. Aging (Albany NY) 2011; 3:1043-4; PMID:22170738
- 39. Miller RA, Harrison DE, Astle CM, Baur JA, Boyd AR, de Cabo R, Fernandez E, Flurkey K, Javors MA, Nelson JF, et al. Rapamycin, but not resveratrol or simvastatin, extends lifespan of genetically heterogeneous mice. J Gerontol A Biol Sci Med Sci 2011; 66:191-201; PMID:20974732; http://dx.doi. org/10.1093/gerona/glq178
- Wilkinson JE, Burmeister L, Brooks SV, Chan CC, Friedline S, Harrison DE, Hejtmancik JF, Nadon N, Strong R, Wood LK, et al. Rapamycin slows aging in mice. Aging Cell 2012; 11:675-82; PMID:22587563; http://dx.doi.org/10.1111/j.1474-9726.2012.00832.x
- Anisimov VN, Zabezhinski MA, Popovich IG, Piskunova TS, Semenchenko AV, Tyndyk ML, Yurova MN, Rosenfeld SV, Blagosklonny MV. Rapamycin increases lifespan and inhibits spontaneous tumorigenesis in inbred female mice. Cell Cycle 2011; 10:4230-6; PMID:22107964; http://dx.doi. org/10.4161/cc.10.24.18486
- Komarova EA, Antoch MP, Novototskaya LR, Chernova OB, Paszkiewicz G, Leontieva OV, Blagosklonny MV, Gudkov AV. Rapamycin extends lifespan and delays tumorigenesis in heterozygous p53+/- mice. Aging (Albany NY) 2012; 4:709-14; PMID:23123616
- Comas M, Toshkov I, Kuropatwinski KK, Chernova OB, Polinsky A, Blagosklonny MV, Gudkov AV, Antoch MP. New nanoformulation of rapamycin Rapatar extends lifespan in homozygous p53-/- mice by delaying carcinogenesis. Aging (Albany NY) 2012; 4:715-22; PMID:23117593
- Donehower LA. Rapamycin as longevity enhancer and cancer preventative agent in the context of p53 deficiency. Aging (Albany NY) 2012; 4:660-1; PMID:23128359
- Spong A, Bartke A. Rapamycin slows aging in mice. Cell Cycle 2012; 11:845; PMID:22356747; http:// dx.doi.org/10.4161/cc.11.5.19607
- Pearce LR, Alton GR, Richter DT, Kath JC, Lingardo L, Chapman J, Hwang C, Alessi DR. Characterization of PF-4708671, a novel and highly specific inhibitor of p70 ribosomal S6 kinase (S6K1). Biochem J 2010; 431:245-55; PMID:20704563; http://dx.doi. org/10.1042/BJ20101024

- Lehman N, Ledford B, Di Fulvio M, Frondorf K, McPhail LC, Gomez-Cambronero J. Phospholipase D2-derived phosphatidic acid binds to and activates ribosomal p70 S6 kinase independently of mTOR. FASEB J 2007; 21:1075-87; PMID:17242159; http:// dx.doi.org/10.1096/fj.06-6652com
- Chen Y, Zheng Y, Foster DA. Phospholipase D confers rapamycin resistance in human breast cancer cells. Oncogene 2003; 22:3937-42; PMID:12813467; http://dx.doi.org/10.1038/sj.onc.1206565
- Kam Y, Exton JH. Role of phospholipase D1 in the regulation of mTOR activity by lysophosphatidic acid. FASEB J 2004; 18:311-9; PMID:14769825; http://dx.doi.org/10.1096/fj.03-0731com
- Hornberger TA, Chu WK, Mak YW, Hsiung JW, Huang SA, Chien S. The role of phospholipase D and phosphatidic acid in the mechanical activation of mTOR signaling in skeletal muscle. Proc Natl Acad Sci U S A 2006; 103:4741-6; PMID:16537399; http://dx.doi.org/10.1073/pnas.0600678103
- Biteau B, Karpac J, Hwangbo D, Jasper H. Regulation of Drosophila lifespan by JNK signaling. Exp Gerontol 2011; 46:349-54; PMID:21111799; http:// dx.doi.org/10.1016/j.exger.2010.11.003
- Jasper H, Kennedy BK. Niche science: the aging stem cell. Cell Cycle 2012; 11:2959-60; PMID:22926561; http://dx.doi.org/10.4161/cc.21558
- Biteau B, Karpac J, Supoyo S, Degennaro M, Lehmann R, Jasper H. Lifespan extension by preserving proliferative homeostasis in Drosophila. PLoS Genet 2010; 6:e1001159; PMID:20976250; http:// dx.doi.org/10.1371/journal.pgen.1001159
- Biteau B, Hochmuth CE, Jasper H. JNK activity in somatic stem cells causes loss of tissue homeostasis in the aging Drosophila gut. Cell Stem Cell 2008; 3:442-55; PMID:18940735; http://dx.doi. org/10.1016/j.stem.2008.07.024
- Averous J, Fonseca BD, Proud CG. Regulation of cyclin D1 expression by mTORC1 signaling requires eukaryotic initiation factor 4E-binding protein 1. Oncogene 2008; 27:1106-13; PMID:17724476; http://dx.doi.org/10.1038/sj.onc.1210715
- Cohen JD, Gard JM, Nagle RB, Dietrich JD, Monks TJ, Lau SS. ERK crosstalks with 4EBP1 to activate cyclin D1 translation during quinol-thioetherinduced tuberous sclerosis renal cell carcinoma. Toxicol Sci 2011; 124:75-87; PMID:21813464; http://dx.doi.org/10.1093/toxsci/kfr203
- 57. Chang BD, Broude EV, Fang J, Kalinichenko TV, Abdryashitov R, Poole JC, Roninson IB. p21Waf1/ Cip1/Sdi1-induced growth arrest is associated with depletion of mitosis-control proteins and leads to abnormal mitosis and endoreduplication in recovering cells. Oncogene 2000; 19:2165-70; PMID:10815808; http://dx.doi.org/10.1038/sj.onc.1203573