

REVIEW

Open Access



# Research progress of CD73-adenosine signaling regulating hepatocellular carcinoma through tumor microenvironment

Liang Shan<sup>1,2,3,4</sup>, Mingxu Gong<sup>2,3,4</sup>, Dandan Zhai<sup>1</sup>, Xiangyun Meng<sup>1</sup>, Jianjun Liu<sup>1\*</sup>  and Xiongwen Lv<sup>2,3,4\*</sup> 

## Abstract

Adenosine signaling pathway is a kind of signal regulation hub widely existing in human body, which is involved in a series of physiological processes such as energy supply of body cells. CD73 is a highly concerned signaling protein in purine adenosine pathway, and its role in tumor development and prognosis has been paid more and more attention in recent years, especially in hepatocellular carcinoma (HCC). In this paper, the specific mechanism by which CD73-adenosine signaling regulates tumor microenvironment (TME) of liver cancer tumors was analyzed in detail, highlighting the importance of this pathway as a therapeutic target to combat tumor immunosuppression and enhance the anti-tumor immune response to prevent and treat hepatocellular carcinoma (HCC). In addition, a variety of current targeted therapeutic strategies for adenosine metabolic pathways are summarized, including the development of new drugs in the stage of preclinical research and clinical trials, and the mechanism of action, implementation possibility, and clinical effects of these therapies are discussed. By summarizing the latest scientific research results, in this review, we attempt to paint a panorama of the mechanism of adenosine action in tumor immunotherapy, with the aim to provide a solid theoretical basis and practical guidance for subsequent research and clinical application, ultimately promoting the development of more accurate and efficient tumor immunotherapy.

**Keywords** CD73-adenosine signaling pathway, Hepatocellular carcinoma, Tumor microenvironment, Adenosine, Adenosine receptor

## Background

Liver cancer, the third leading cause of cancer-related death worldwide, is a multi-factor-induced, multi-gene-involved and complex digestive system malignancy [1]. Hepatocellular carcinoma (HCC) accounts for more than 85% of primary liver cancers [2]. Immunoinflammatory microenvironment-driven tumor development and treatment are major challenges in the prevention and treatment of HCC [3]. It is of great scientific significance to deeply understand the regulatory mechanism of the tumor immune microenvironment and to discover and elucidate the characteristics and functional remodeling of immune cells in the immune microenvironment and the

\*Correspondence:

Jianjun Liu

Jianjun\_liu2020@163.com

Xiongwen Lv

Xiongwen\_lv2019@163.com

<sup>1</sup>Department of Pharmacy, The Second People's Hospital of Hefei (Hefei Hospital Affiliated to Anhui Medical University), Hefei, Anhui 230000, China

<sup>2</sup>Anhui Province Key Laboratory of Major Autoimmune Diseases, Anhui Medical University, Hefei, Anhui 230032, China

<sup>3</sup>Inflammation and Immune Mediated Diseases Laboratory of Anhui Province, Hefei, Anhui 230032, China

<sup>4</sup>The Key Laboratory of Major Autoimmune Diseases, Hefei, Anhui Province 230032, China



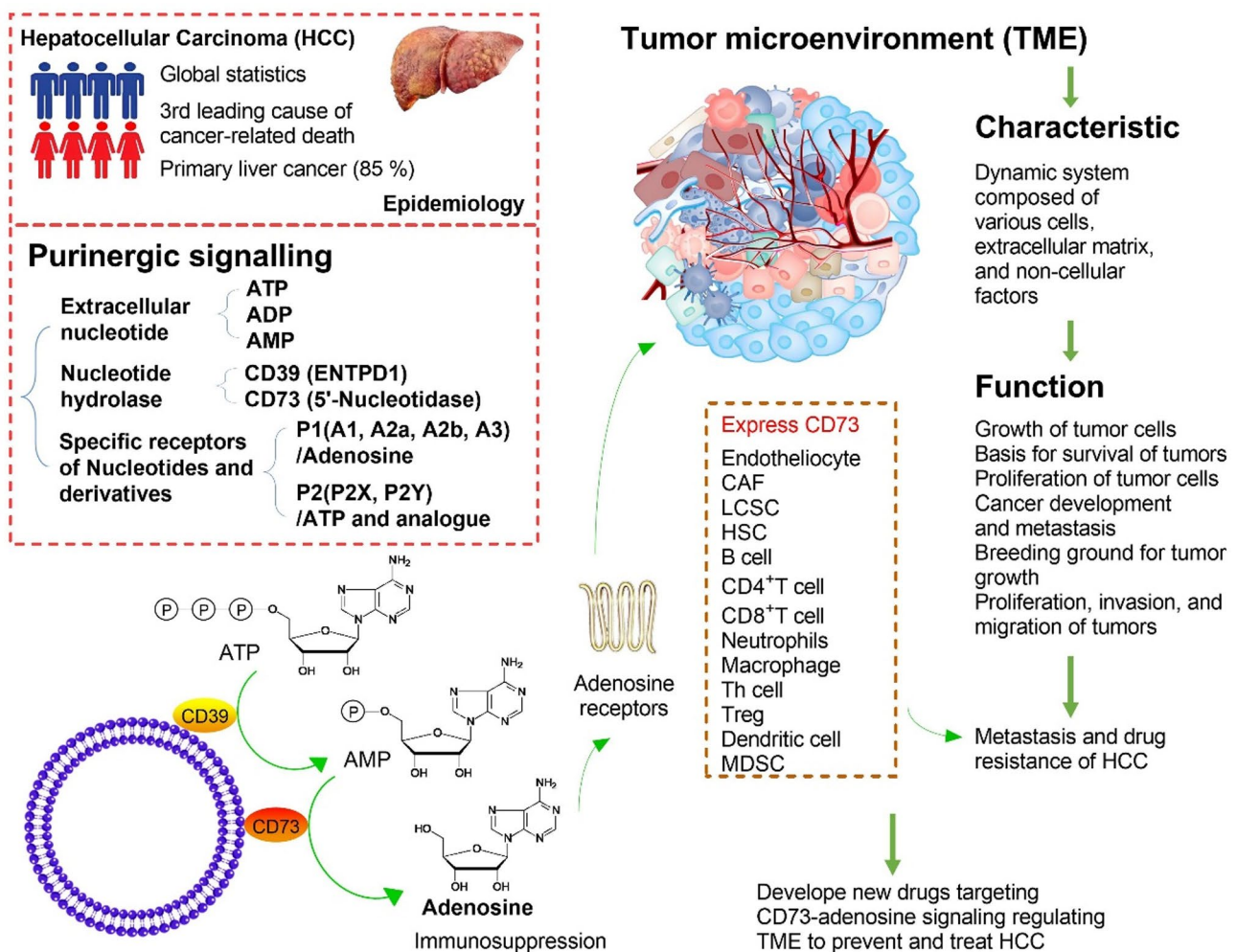
© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

mechanism of HCC progression [4]. Purinergic signaling participates in the pathophysiological regulation of many diseases, including liver disease [5]. Purinergic signaling is chiefly composed of extracellular nucleotides, nucleotide hydrolases (e.g., CD39, ENTPD1, CD73), nucleotides, and derivatives of specific receptors [6]. The purine receptors are divided into two categories, P1 receptors acting on adenosine, and P2 receptors acting on adenosine triphosphate (ATP) and its analogues [7]. Four subtypes of P1 receptor, including A1R, A2aR, A2bR, and A3R, have been cloned. P2 receptors are divided into P2X and P2Y groups [8]. CD73 plays an inhibitory role in the tumor microenvironment (TME) by mediating adenosine production [9]. Adenosine can inhibit the function of tumor-killing immune cells, such as effector T cells, natural killer (NK) cells, and dendritic cells (DCs), and enhance the function of suppressive immune cells, including regulatory T cells, myeloid suppressor cells,

and tumor-associated macrophages [10]. At present, many companies at home and abroad are developing new drugs targeting CD73-adenosine to regulate the TME to prevent and treat HCC [11]. CD73 is expressed in a variety of human tumors, especially in HCC cells, cancer-associated fibroblasts (CAFs), and endothelial cells, but also in myeloid cells, NK cells, and T cells [12]. Numerous studies have described a strong association between elevated CD73 levels and adverse clinical outcomes [13]. In this article, we review recent progress in the immunotherapy of HCC by targeting CD73-adenosine signaling to regulate the TME of HCC (Fig. 1).

### TME regulates the development and progression of HCC

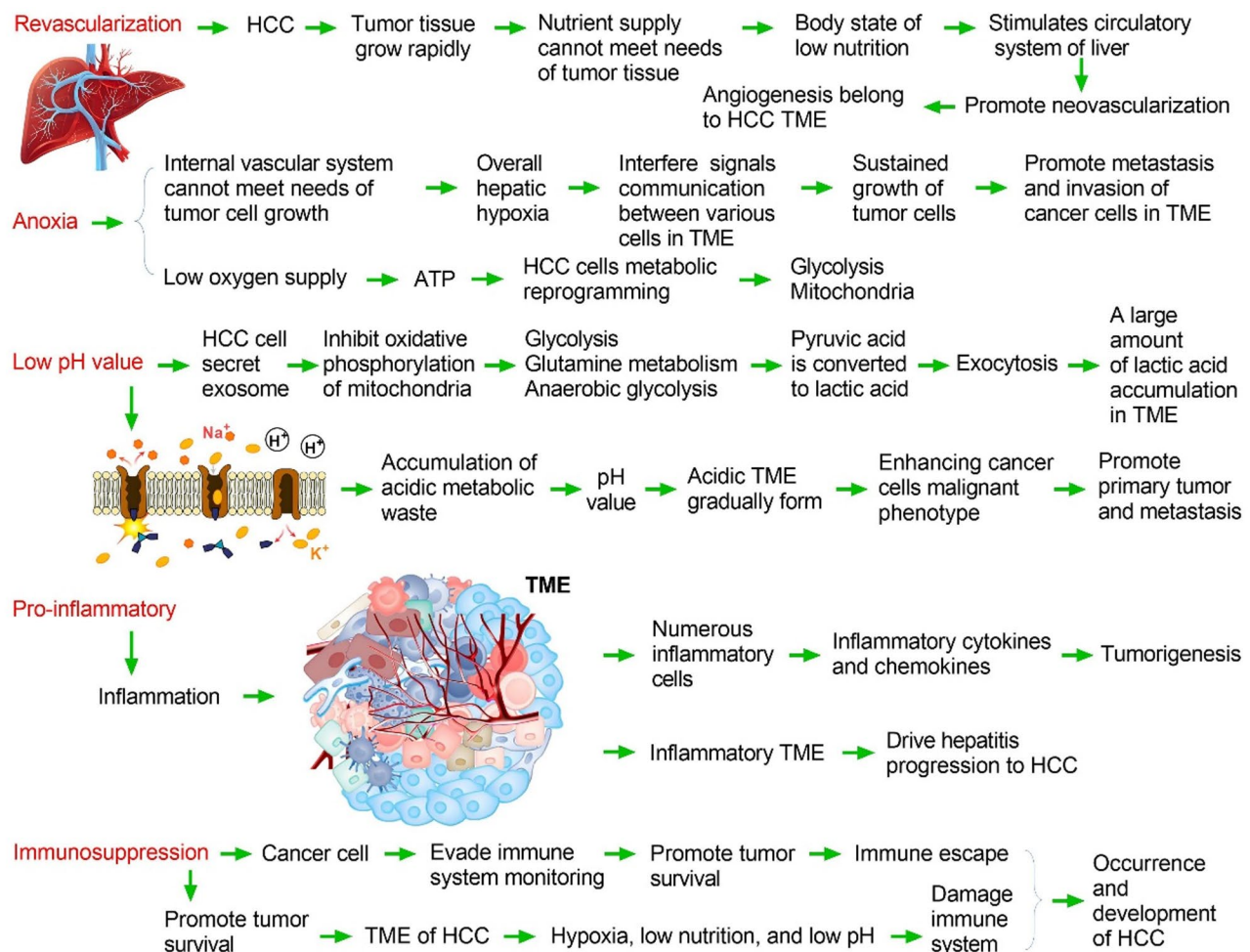
The TME refers to the local environment in which tumor cells reside [14]. It not only includes the tumor cells themselves, but also the surrounding support cells,



**Fig. 1** CD73 and adenosine receptors are highly expressed in the liver tissues of patients with hepatocellular carcinoma (HCC). CD73 and adenosine receptors are expressed in a variety of cells. The regulation of CD73-adenosine signals in the tumor microenvironment (TME) to inhibit HCC progression is a current research hotspot

extracellular matrix (ECM), blood vessels, immune cells, and a series of signaling molecules [15]. Most of the TME is occupied by the tumor, and it constitutes the mesenchyme of the tumor [16]. Low oxygen levels, high lactate levels, extracellular acidosis, and low nutrient content are notable features of the TME [17]. A variety of cells exist in the TME, including mesenchymal stem cells (MSCs), fibroblasts, endothelial cells, and immune cells. The TME can secrete cytokines and growth factors [18]. CAFs are among the most abundant cells in the TME, creating conditions for tumor growth and progression [19]. The interaction between the TME and the activation/inhibition signaling network may determine tumor progression [20]. Since 2017, other therapies to modulate the liver TME have emerged [21]. The liver immune microenvironment contains a large number of immune cells, including neutrophils, monocytes, resident macrophages (Kupffer cells [KCs]), NK cells, natural killer T

(NKT) cells, and liver transport and/or resident lymphocytes (B, CD8+ T, CD4+ T, and  $\gamma\delta$  T cells) [22]. There is an overall balance between anti-inflammatory cytokines (interleukin [IL]-10, IL-13, and TGF- $\beta$ ) and pro-inflammatory cytokines (IL-2, IL-7, IL-12, IL-15, and interferon [IFN]- $\gamma$ ), maintaining homeostasis in vivo [23]. In recent years, purine signaling pathways have emerged as important players in cancer progression, with extracellular ATP, adenosine diphosphate (ADP), and adenosine being major signaling molecules [24]. The immunosuppressive metabolite adenosine is a component of the TME [25]. As the main enzyme that catalyzes adenosine production, CD73 is critical in inhibiting adequate anti-tumor immune responses, mainly through the production of adenosine, but also by promoting cancer cell proliferation, tumor growth, angiogenesis, and metastasis, warranting further study (Fig. 2).



**Fig. 2** In the tumor microenvironment (TME) of hepatocellular carcinoma (HCC), the immune system is destroyed and rebuilt under an adverse environment, constituting hypoxia, low nutrition, low pH, and blood flow changes; this leads to the formation of a new systemic and local immune microenvironment that is suitable for tumor survival, regulating inflammation, and liver fibrosis, assisting tumors to escape immune system monitoring, leading to the occurrence and development of HCC



### TME promotes vascular regeneration at the tumor site

HCC is a highly angiogenic cancer, and angiogenesis plays an important role in tumor growth, early metastasis, and poor survival [26]. The cell components of the TME system include hepatic stellate cells (HSCs), fibroblasts, immune cells, and endothelial cells [27]. Non-cellular components include growth factors (fibroblast growth factor [FGF], hepatocyte growth factor [HGF], and vascular endothelial growth factor [VEGF]), proteolytic enzymes, ECM proteins, and inflammatory factors [28]. The activated HSCs secrete angiogenic growth factor, which, together with VEGF, stimulate angiogenesis, form a new vascular system within the TME, and provide various nutrients for tumor growth [29, 30]. At present, many angiogenesis inhibitors have been approved for the clinical treatment of liver cancer [31]. Anti-VEGF-VEGFR drugs generally include the following categories: antibody drugs directly targeting VEGF and VEGFR proteins, such as bevacizumab; tyrosine kinase signaling pathway inhibitors, such as sorafenib; and types of fusion proteins and immunomodulators [32]. Combined with clinical findings, the clinical benefits of angiogenesis inhibitors are not obvious and have therapeutic limitations, which may be due to the regulation of angiogenesis networks involving multiple proteins or signaling pathways [33]. When a protein is inhibited by a single target drug, there is a compensatory phenomenon of angiogenesis, and the structure of new blood vessels in patients with tumors is incomplete and the permeability is poor; thus, some therapeutic drugs cannot effectively reach the lesion site (Fig. 2) [34].

### Hypoxia is an important characteristic of the HCC TME

Hypoxia plays an important role in tumor cell characteristics [35]. Studies have confirmed that hypoxia is closely related to the genesis, proliferation, apoptosis, autophagy, and metabolism of tumor cells [36]. Tumor cells are in a hypoxic environment for a long time [37]. Not only can tumor cells adapt to hypoxia, but hypoxia has also become a living environment for tumor cells to grow, proliferate, and maintain their characteristics [38]. Hypoxia not only controls cell proliferation but also apoptosis [39]. During the process from the beginning of malignant transformation to the formation of highly malignant tumors, malignant cells that can adapt to the hypoxic microenvironment survive and continue to proliferate to form highly malignant tumor tissues, whereas the cells in the inappropriate environment disappear through apoptosis and other mechanisms [40]. Most of the activities of tumor tissues are thought to be related to the anoxic microenvironment [41]. Hypoxia-inducible factors (HIFs) are some of the most important endogenous transcription factors produced under hypoxia, serving as the upstream factor responsible for causing

many hypoxia reactions [42]. Among them, HIF-1 is the most widely distributed, most important, and most thoroughly studied, with HIF-1 $\alpha$  as its functional monomer [43]. The expression of HIF-1 $\alpha$  has been shown to be positively correlated with the severity of hypoxia, and the expression of HIF-1 $\alpha$  is higher in regions with more severe hypoxia [44]. In addition, within a short time after the occurrence of hypoxia, HIF-1 $\alpha$  in the cell is rapidly increased to a certain level [45]. The expression of CD73 is regulated by HIF-1, transforming growth factor (TGF)- $\beta$ , EGFR, AKT,  $\beta$ -catenin, and other molecules, especially HIF-1, which plays the function of a transcription factor [46]. Hypoxia is an important feature of the TME [47]. Hypoxia induces HIF-1 upregulation, leading to the widespread expression of CD73 in the TME [48].

Current studies have confirmed that *HIF-1 $\alpha$*  can directly regulate more than 60 target genes, and new target genes are constantly being discovered [49]. In tumor tissue, HIF-1 $\alpha$  is closely related to tumor progression [50]. HIF-1 $\alpha$  can regulate cell apoptosis and proliferation, enabling tumors to selectively multiply tumor clones that are more suitable for survival in anoxic microenvironments [51]. HIF-1 $\alpha$  regulates a key enzyme involved in glycolysis, enabling tumor cells to survive hypoxia [52]. Anticancer therapies trigger the release of ATP from tumor cells, leading to the rapid formation of adenosine by the exonucleoenzymes CD39 and CD73, which subsequently aggravates immunosuppression in the TME [53]. ATP is the energy source of various cells, and several studies have found that downregulation of metabolic reprogramming in HCC cells can inhibit their growth [53, 54]. In addition, adenosine deaminase (ADA) can metabolize adenosine to inosine (INO), which causes severe metabolic reprogramming, reducing glycolysis and increasing mitochondrial and glycolytic capacity; however, the specific mechanism still needs to be further studied [55]. In conclusion, hypoxia is common in all solid tumors, and the HIF signaling pathway is involved in the inflammatory response, immunosuppression, and activation of a variety of cancer-promoting biological effects. HIF can lead to the aggregation of regulatory T cells (Tregs) and macrophages, promote sorafenib resistance, and is a potential biomarker for the diagnosis, prognosis, and recurrence of HCC (Fig. 2).

### TME of HCC exhibits a low pH (acidity)

Under normal circumstances, the pH levels inside and outside the cells of the human body are maintained within a certain range, which is essential for maintaining the physiological function of the human body [56]. The pH of the TME is usually between 6.5 and 7.5, slightly lower than the pH of normal cells [57]. The low pH of the TME of HCC may be related to the excessive production of lactic acid or abnormal glucose metabolism

of tumor cells in an anoxic environment [58]. This is because tumor cells have certain metabolic characteristics and will produce vast amounts of lactic and carbonic acids, making the surrounding environment acidic and promoting the growth and metastasis of tumor cells [59]. The acidic condition of the TME affects cell survival and proliferation to some extent [60]. First, an acidic environment can reduce the stability of the ECM, facilitating the invasion of surrounding tissues by tumor cells [61]. Second, the acidic environment can also affect the metabolic activity of tumor cells, reducing their energy production and DNA repair ability, thereby increasing the susceptibility and mortality of cells [62]. An acidic environment can also affect the function of immune cells and inhibit the killing effect of the immune system on tumor cells [63]. Research on the pH range of the TME is still in its infancy and needs to be further explored. In future studies, pH changes in the TME can be monitored by employing non-invasive techniques such as magnetic resonance imaging and spectroscopy. In addition, the metabolic characteristics and gene expression profiles of tumor cells can be combined to further reveal the relationship between changes in the TME pH and tumor development (Fig. 2).

#### **Proinflammatory TME of HCC**

Chronic liver inflammation caused by various pathogenic factors is an important basis for the formation of HCC [64]. Immune cells are an important part of the TME, including macrophages, DCs, NK cells, bone marrow-derived suppressor cells (MDSCs), and T/B lymphocytes [65]. The main function of these immune cells is to secrete a large number of inflammatory factors, such as IL-1 and IL-6, to induce an inflammatory response, which also plays a dual role in the regulation of tumor processes [66]. The TME plays an immune monitoring role in the early stage of tumors, mainly through T/B lymphocytes and NK cells, to stimulate anti-tumor immunity and inhibit tumor progression [67]. With the development of the tumor, tumor cells develop immune tolerance under long-term inflammatory stimulation, and excessive inflammation promotes the occurrence and development of the tumor [68]. Together with hypoxia, the upregulation of IL-6 expression by the toll-like receptor signaling pathway in tumor cells promotes the inflammatory response of tumors and promotes tumor resistance, proliferation, and invasion [69]. The initial goal of the inflammatory response is to eliminate foreign invaders or damaged tissue [70]. However, the composition and function of inflammatory cells are often altered in the TME, resulting in immunosuppression and contributing to tumor immune escape [71]. For example, tumor and stromal cells promote inflammation and immunosuppression through NF- $\kappa$ B and STAT3, which promotes tumor cells

to evade immune recognition and participate in tumor cell proliferation, metastasis, drug resistance, and tumor angiogenesis [72]. Currently, many methods are being researched and developed to normalize the inflammatory TME in patients with HCC. However, there is no doubt that simultaneously targeting cancer and stromal cells is more effective and more challenging than targeting cancer cells alone (Fig. 2).

#### **TME promotes immunosuppression in HCC**

The TME is a complex internal environment network that tumor cells depend on for survival and development [73]. The immunosuppressive TME is a part of the TME that plays a role in suppressing immune function [74]. Once HCC is formed, the interaction of cellular and non-cellular components in the TME gradually forms an immunosuppressive microenvironment and promotes tumor development [75]. In recent years, the treatment of immune checkpoint blockers (ICBs), such as Nivolumab (anti-programmed cell death protein 1 [PD-1]) and Atezolizumab (anti-programmed cell death ligand-1 [PD-L1]), has revolutionized the treatment landscape of advanced HCC [76]. However, most patients with HCC have primary resistance to immunotherapy and are unable to achieve significant survival benefits [77]. Studies have shown that the key mechanism leading to poor response to ICB therapy lies in the immunosuppressive TME, with MDSCs as the core [78]. The complex interactions between HCC cells and their immunosuppressive microenvironment during HCC development and drug therapy remain to be further studied [79]. Selectively modulating the immunosuppressive regulatory networks associated with primary or secondary tumors may reprogram the microenvironment, providing an immunotherapy strategy for treating HCC [80]. Encouragingly, other molecules with immunosuppressive activity expressed by Tregs, such as T-cell immunoreceptor with Ig and ITIM domains (TIGIT), lymphocyte-activation gene 3 (LAG3), and T-cell immunoglobulin and mucin domain-containing protein 3 (TIM3), are also currently in clinical trials [81]. At present, it is urgent to systematically analyze the molecular regulatory mechanisms of the immunosuppressive TME, especially the key pathways of abnormal activation of various inhibitory immune cells, and establish precise combination therapy strategies to break through the bottleneck of the clinical treatment of HCC (Fig. 2).

#### **Key role of CD73 in HCC progression**

The cell types that most frequently express CD73 are various tumor, immune, and stromal cells [82–85]. In solid tumors, especially HCC, CD73 is a key component in the formation of the immunosuppressive microenvironment and the occurrence of tumor immune escape

[82]. Human CD73 is a multifunctional transmembrane glycoprotein composed of 523 amino acids encoded by ecto-5'-nucleotidase (*NT5E*) (located at 6q14-21), with a relative molecular mass of 70,000, which is anchored to the cell membrane by glycosylphosphatidylinositol (GPI) [83]. CD73 is widely expressed on the surface of a variety of human cells, including lymphocytes, endothelial cells, and epithelial cells, and controls a variety of physiological functions, including epithelial ion exchange, fluid transport, platelet function, tissue hypoxia, and vascular leakage [84]. CD73 is primarily involved in the following physiological effects: (1) affecting the purine nucleotide synthesis process, where adenosine, mediated by CD73, controls the production of purine nucleotides through CD39 and CD73, thereby regulating nucleotide signaling [85]; (2) catalyzing 5'-adenosine monophosphate (AMP), where the produced adenosine binds to the A1, A2a, A2b, and A3 adenosine receptors, producing different physiological effects through biological signal transduction [86]; and (3) involvement in T cell activation, where adenosine produced by CD73 hydrolysis activates immune cells, affects the proliferation of immune cells, regulates CD4+CD25+Treg cells, and reduces the immune function of T cells [87]. CD73, a cell surface enzyme that is widely expressed on the surface of human endothelial cells and lymphocytes (such as Tregs) [88], can convert immune-activating ATP into the immunosuppressant adenosine [89]. Adenosine downregulates immune activity by binding with the downstream adenosine receptor (A2aR) [90]. In the TME, hypoxia induces overexpression of CD73 on the surface of cancer cells to dephosphorylate AMP to adenosine through CD73, thereby forming an immunosuppressive TME and promoting tumor growth [91]. Therefore, CD73 inhibition may activate T cell function. Preclinical studies have shown that CD73 has a good synergistic effect with PD-L1 [92]. Considering the current global clinical development of the CD73 monoclonal antibody, the treatment options explored are mostly a combination of CD73 and PD-1 or other drugs [93]. In the future, CD73 is expected to become a good companion for PD-1 drugs and to further improve the response rate of tumor immunotherapy (Fig. 3).

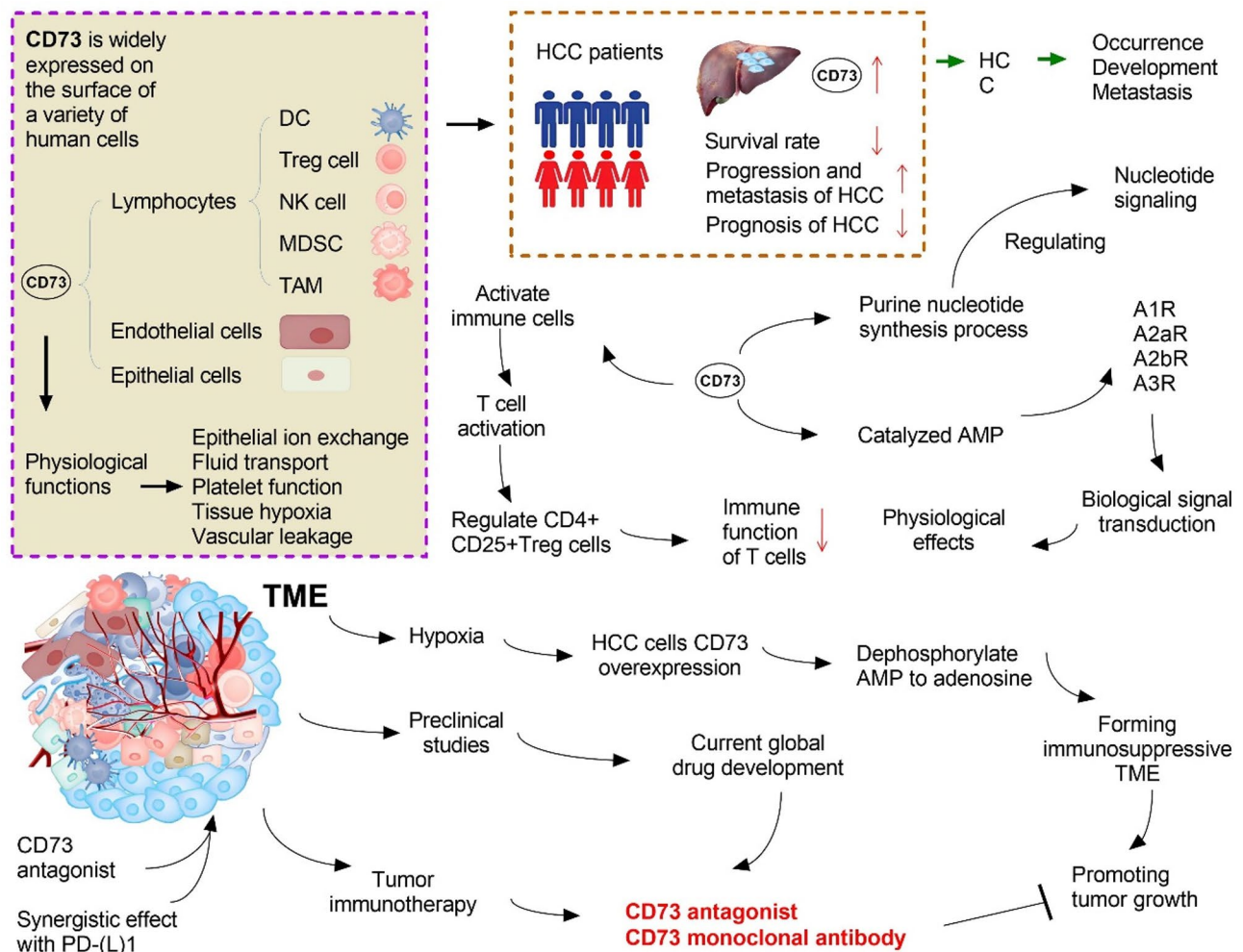
#### **CD73-adenosine signaling in the TME mediates immune escape in HCC**

Inhibition of CD73 can enhance the anti-HCC immune effect, and the mechanism is related to the inhibition of adenosine accumulation and the reversal of immune system function inhibited by adenosine [94]. The surrounding microenvironment of HCC cells is called the TME and includes various signaling molecules and ECM, including blood vessels in and around the tumor, tumor-infiltrating immune cells, fibroblasts, and PD-1/PD-L1 [95]. HCC can affect its microenvironment through the

release of cell signaling molecules, promoting tumor angiogenesis, inducing immune tolerance, and helping tumor cells escape the killing of immune cells within the tumor [96]. The CD73-adenosine pathway is an important pathway involved in tumor immune escape [97]. HCC cells generate immunosuppressive signals mainly by increasing adenosine levels in the microenvironment [98]. Adenosine binds to adenosine receptors in various immune cells to reduce inflammation and inhibit immune responses [99]. First, adenosine can inhibit or affect the maturation, differentiation, and function of various immune cells [100]. For example, adenosine can promote the migration of macrophages from the anti-tumor M1 type to the tumor-promoting M2 type [101]. For DCs, adenosine can inhibit its antigen-presenting ability while inducing DCs to secrete multiple cytokines that promote tumor growth [102]. Second, adenosine can inhibit the maturation of NK cells and significantly decrease their killing activity [103]. For T cells, adenosine inhibits the production of IL-2 by CD4+T cells, the proliferation, differentiation, and maturation of CD8+T cells, and the production of a variety of cytokines that help the immune system fight tumors, such as IFN- $\gamma$  and tumor necrosis factor (TNF)- $\alpha$ . In contrast, adenosine promotes the function of immunosuppressive cells, such as Tregs and MDSCs, to suppress the immune response [104]. Finally, adenosine can increase the expression of immune checkpoints on the surface of immune cells, including PD-1, cytotoxic T lymphocyte-associated antigen 4 (CTLA-4), and LAG3 [105]. Targeted inhibition of CD73-adenosine signaling in the TME has become a promising therapy to alter adenosine levels in the TME and fully restore the anti-cancer function of the immune system. Although drug research targeting the adenosine pathway derived from the TME is promising, it remains in its infancy and its anti-tumor effect still needs to be further confirmed by more large-sample studies (Fig. 4).

#### **Inhibition of CD73-adenosine signaling reduces inflammatory responses**

CD39 upstream of CD73 catalyzes ATP to produce AMP, and the resulting AMP can be converted into adenosine by CD73 [106]. Adenosine binds to downstream adenosine receptors (A2aR or A2bR), which exert a broad immunosuppressive effect by activating the protein kinases protein kinase A (PKA) and c-Src tyrosine kinase (CSK) while inhibiting a series of signaling pathways related to inflammation, such as mitogen-activated protein kinase (MAPK) and protein kinase C (PKC) [107]. In HCC-related inflammation, which is characterized by chronic persistent inflammation, immune cells typically do not exhibit antitumor effects [108]. On the contrary, adenosine can participate in the development, proliferation, and metastasis of HCC by releasing inflammatory

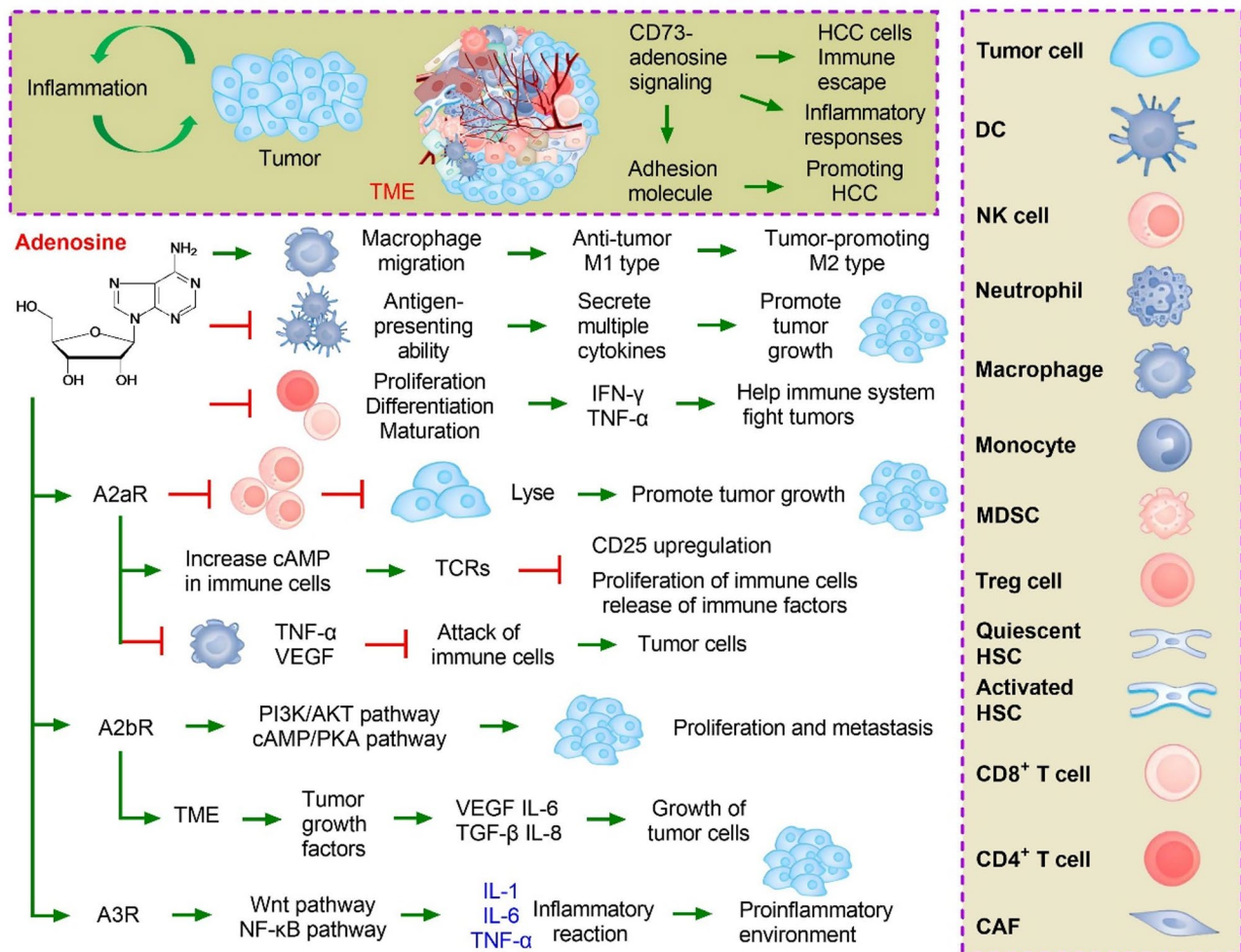


**Fig. 3** The tumor microenvironment (TME) is an important internal environment in the occurrence and development of tumor diseases. In the TME, CD73 plays an important role in the occurrence, development, and metastasis of hepatocellular carcinoma (HCC) by promoting angiogenesis through immune escape, inflammatory cancer signaling transmission, adhesion molecular function, and neovascularization

mediators and inflammatory-related cytokines into the TME [109]. However, generating an adenosine pathway-induced anti-inflammatory response to enhance the anti-tumor effect and improve the HCC-related inflammatory microenvironment has attracted much attention. Adenosine mediates anti-inflammatory effects through P1 purine receptors (A1R, A2aR, A2bR, A3R) [110]. Adenosine receptors belong to the G protein-coupled receptor family and are expressed by various cells, including immune cells [111]. Adenosine, as a key effector molecule in the regulation of innate and adaptive immunity, can bind to A2aR on the surface of various immune cells to mediate the increase in cAMP in immune cells, thereby inhibiting the upregulation of CD25 mediated by effector T cell receptors (TCRs) as well as inhibiting the proliferation of immune cells and the release of immune factors [112]. Adenosine combined with A2aR can inhibit the production of macrophages, reduce the secretion of

different pro-inflammatory mediators such as TNF- $\alpha$  and VEGF, inhibit the attack of immune cells on tumor cells, and help tumor cells to escape immune system control [113]. Adenosine can also affect the differentiation of DCs, mainly through binding with A2bRs, so that it can secrete different tumor growth factors in the immune microenvironment of the tumor, including VEGF, IL-6, IL-8, and TGF- $\beta$ , providing a good “soil” for the growth of tumor cells [114]. CD73, as one of the main enzyme activators regulating adenosine, plays an important role in the process by which adenosine regulates inflammation and the immune response, responsible for mediating immune escape [115]. In conclusion, CD73 promotes the proliferation and metastasis of HCC cells by mediating adenosine in the process of immune regulation and inflammation. However, the genetic characteristics of CD73/AR and the mechanism underlying the immunosuppressive pathway remain unclear (Fig. 4).





**Fig. 4** In the tumor microenvironment (TME), CD73 mediates the production and inhibition of inflammatory signaling molecules through the adenosine pathway. CD73 stimulates the production of inflammatory cytokines through various downstream signaling pathways, placing hepatocellular carcinoma (HCC) cells in a homeostasis environment with high expression of inflammatory cytokines, promoting the coexistence of inflammatory cells and cancer cells, and promoting the progression of HCC

#### CD73 in the TME promotes HCC via its adhesion molecule function

CD73 is a glycoprotein that exists on the surface of cell membranes of all cell types and can also be free outside cells [116]. Studies have shown that CD73 in the TME can participate in the proliferation, angiogenesis, and invasion of tumor cells and can also serve as an important adhesion signaling molecule on the cell surface to promote intercellular adhesion, migration, and cancer cell invasion [117]. High expression of CD73 promotes the progression of HCC and is associated with promoting the invasion and metastasis of HCC [118]. The CD73-adenosine signal induces increased inflammation, decreases or loses the adhesion function between normal cells, and leads to cell looseness and disconnection, thereby facilitating invasion and metastasis of HCC cells [119]. CD73 also promotes HCC cell proliferation by regulating the cell cycle, apoptosis, and signaling

pathways, such as EGFR,  $\beta$ -catenin/cyclin D1, VEGF, and AKT/extracellular signal-regulated kinase (ERK). Independent of its enzymatic function, CD73 promotes the mutual adhesion, migration, and invasion of HCC cells [120]. New studies have shown that in tumor cells, activation of CD73 can promote the adhesion of tumor cells through the EGFR pathway, produce B-cell lymphoma-2 (Bcl-2) and Bcl-xL to inhibit cell apoptosis, release matrix metalloproteinases (MMPs) to hydrolyze the ECM, and promote the remote migration of tumor cells [121]. In addition, the activation of CD73-adenosine signaling in patients with HCC has been shown to promote inflammation and induce increased levels of E-selectin, intercellular cell adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), IL-1, and TNF- $\alpha$  in the blood of patients with HCC [122]. ICAM-1, VCAM-1, and platelet endothelial cell adhesion molecule-1 (PECAM-1) are highly expressed in vascular endothelial

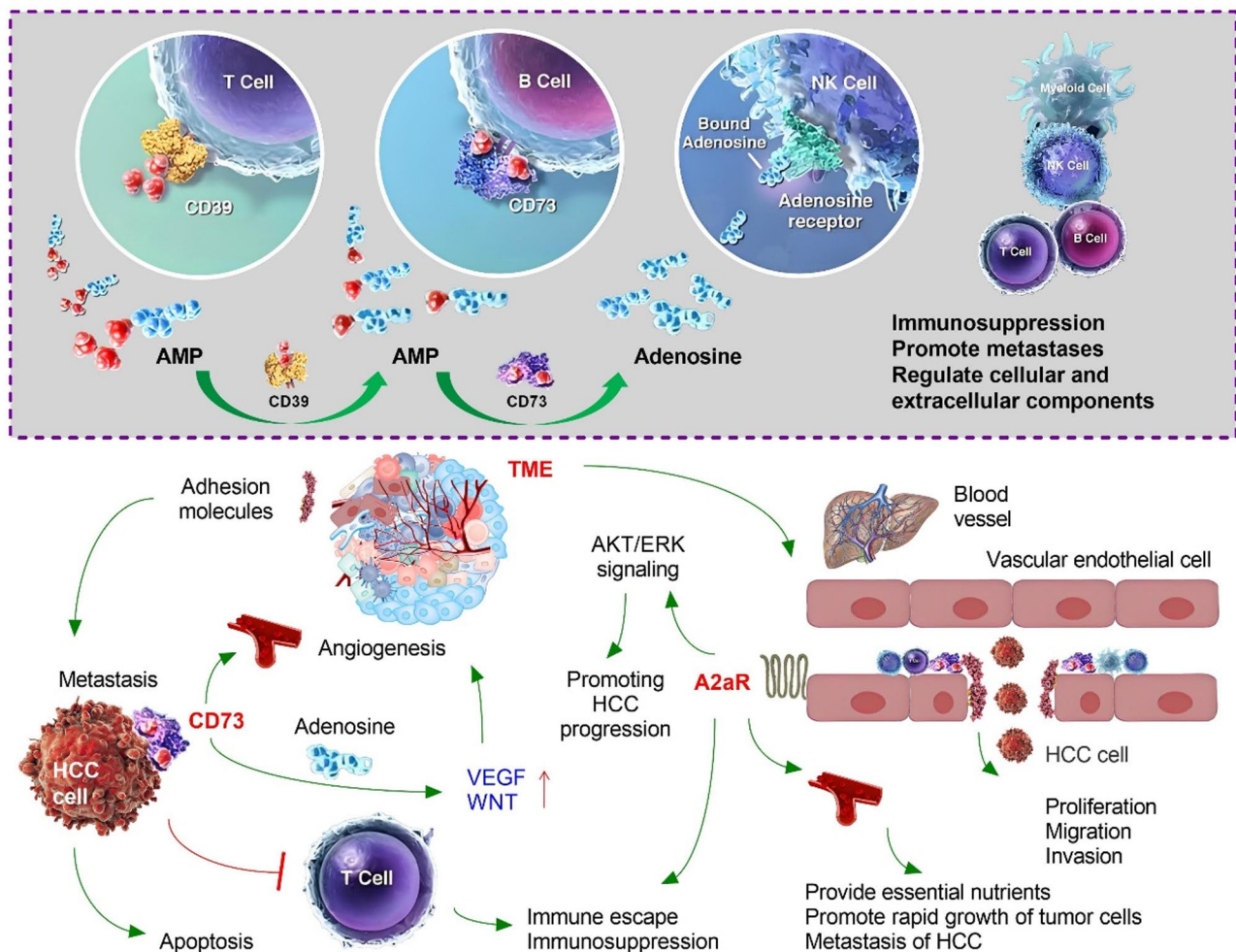


cells [123]. Currently, there are few studies on the regulatory role of CD73 in the TME of HCC, and its specific mechanism needs to be further clarified (Fig. 4).

#### CD73 in the TME promotes angiogenesis in HCC

New research has revealed that CD73 plays a key role in promoting tumor angiogenesis. As a rate-limiting enzyme for adenosine synthesis, CD73 drives tumor angiogenesis and promotes rapid tumor growth by promoting the expression of VEGF and HIF-1 $\alpha$  [124]. Some studies have found that CD73+AD-MSCs have higher paracrine activity, which can better promote angiogenesis and have an obvious therapeutic effect on myocardial infarction [125]. CD73 has a non-selective effect on promoting angiogenesis, as well as a strong effect on both normal and cancer tissues. In addition, tumor-associated macrophages (TAMs) in the TME show high expression of CD73, which can promote angiogenesis and lymphogenesis through the secretion of WNT7B, WNT5A,

WNT11, VEGF-C, VEGF-D, and other cytokines after activation [126]. TAMs can enhance tumor hypoxia and glycolysis, two important causes of angiogenesis [127]. A new study found that, in the TME, the angiogenic factor VEGF- $\alpha$  mainly exists in T3 neutrophil subsets, and its activation may promote tumor angiogenesis, indicating that T3 neutrophils may be a new target for pathological angiogenesis in tumors [128]. Moreover, HCC cells catalyze AMP to produce adenosine through CD73, which acts on the A2aR of macrophages in the TME and activates its downstream AKT/ERK signaling pathway to induce macrophage proliferation and angiogenesis, thus promoting HCC progression [129]. However, currently, the specific mechanism responsible for reducing HCC angiogenesis by inhibiting CD73 in the TME remains unclear and requires further study. In addition, the possible side effects of the widespread expression of CD73 are issues that remain to be addressed (Fig. 5).



**Fig. 5** CD73 on tumor cells can effectively promote the formation of tumor blood vessels in the TME, and high expression of CD73 can also promote the proliferation and metastasis of cancer cells. CD73 can stimulate tumor neovascularization through multiple adenosine receptor pathways, as well as protect neovascularization and promote the growth and metastasis of hepatocellular carcinoma (HCC) in vivo

### Different roles of CD73-associated adenosine receptors within the TME of HCC

During the development of HCC, the TME interacts with HCC cells to mediate the immune tolerance of HCC, with adenosine playing an important role in this process [130, 131]. CD73 is an ectonucleotidase that works with its upstream signaling molecule, CD39, to convert extracellular ATP into adenosine, which then binds to different adenosine receptors to regulate the immune system and exert anti-HCC effects; however, the associated mechanism has yet to be elucidated [130–135]. Many scholars have conducted research on the regulation of CD73 in HCC cells, especially the CD73 found in the TME of HCC [136–140]. Adenosine mediates anti-inflammatory effects through P1 purine receptors (A1R, A2aR, A2bR, A3R). Adenosine receptors belong to the G protein-coupled receptor family and are expressed by a variety of cells, including immune cells. Adenosine has been found to have a low affinity for the A2bR, and inhibits NK cell maturation via the A2aR in the innate immune system [132]. The differentiation of monocytes into macrophages has been shown to be inhibited by the A2bR, which also promotes macrophages from the pro-inflammatory M1 type to the anti-inflammatory M2 type [133]. Similarly, Adenosine acts on DCs through the A2bR, allowing them to secrete vasotropic and immunosuppressive factors that further promote tumor cell growth [134]. In the acquired immune response system, Adenosine inhibits the production of IL-17 by CD4+ T cells through A2aR and A2bR, and promotes the transformation of CD4+ T cells into immunosuppressive cells [135]. At high concentrations, adenosine has been shown to inhibit the proliferation, differentiation, maturation, and cytokine production of B cells via A2aR and A3R [136]. Adenosine also inhibits the proliferation, differentiation, and maturation of CD8+ T cells, as well as the production of cytokines such as IL-2, INF- $\gamma$ , granulocyte-macrophage colony-stimulating factor (GM-CSF), and granzyme B through A2aR and A3R [137]. On the contrary, adenosine promotes the function of immunosuppressive cells, such as CD4+ Tregs and MDSCs, through A2aR or A2bR [138]. In general, adenosine inhibits the functions of NK cells, DCs, and cytotoxic T lymphocytes (CTLs) through its receptors, while enhancing the activity of immunosuppressive cells and participating in the formation of TME immune tolerance [139]. At the same time, adenosine acts on vascular endothelial cells and stromal cells, promoting the formation of TME neovascularization and providing the necessary conditions for tumor metastasis [140]. In summary, the following strategies can be adopted to block the role of adenosine in the TME: (1) preventing the synthesis of adenosine; (2) preventing adenosine from binding to its receptor; and

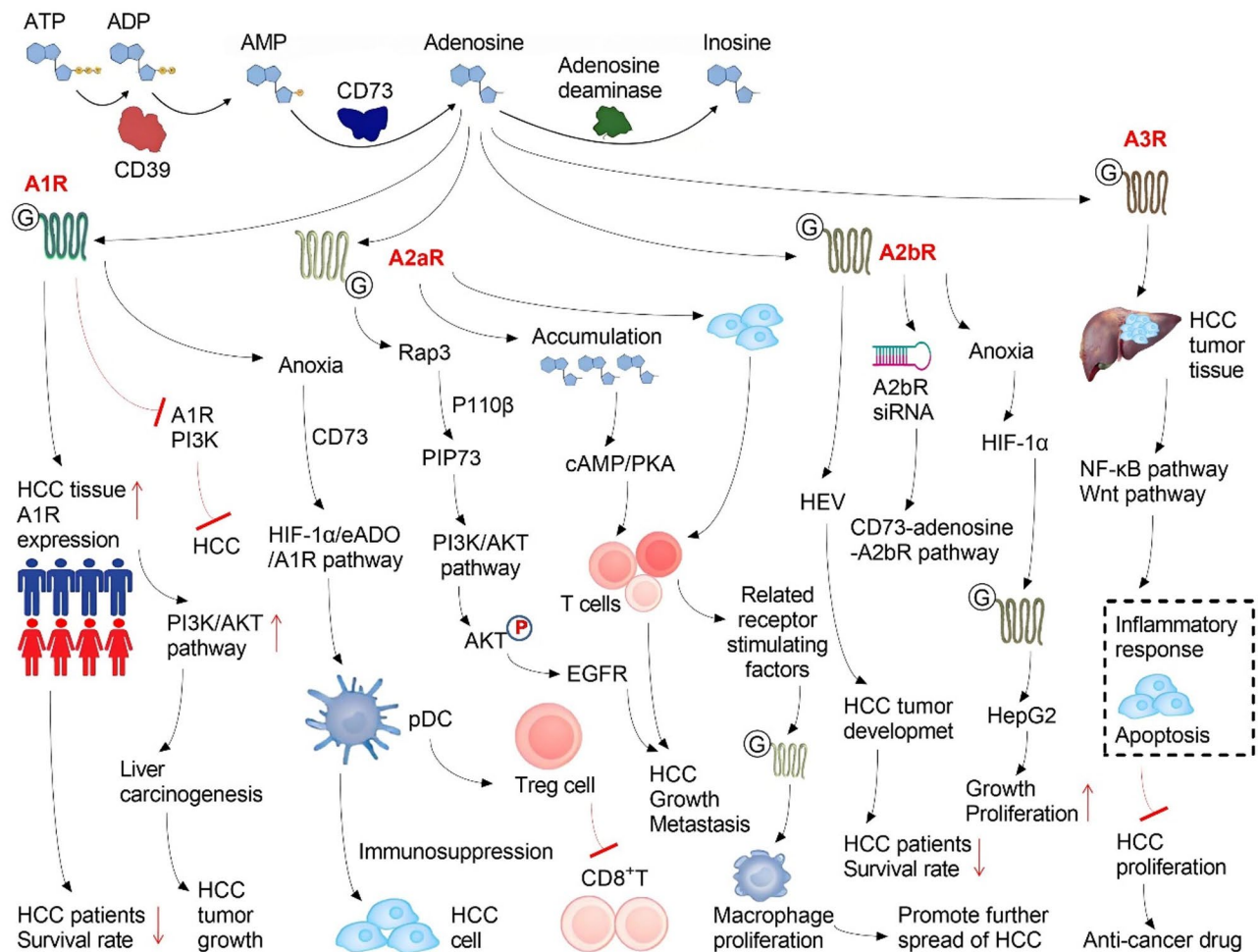
(3) preventing the degradation of high concentrations of extracellular adenosine (Fig. 6).

### A1 adenosine receptor in HCC

The hypoxic TME of HCC facilitates plasmacytoid dendritic cell (pDC) recruitment, the mechanism of which is related to activation of the HIF-1 $\alpha$ /adenosine/A1R signaling pathway [141]. Tumor-derived adenosine promotes the immunosuppressive phenotype of pDCs, leading to the depletion of cytotoxic CD8+ T cells and the amplification of Tregs [142]. Notably, in an immunoactive HCC mouse model, monoclonal antibodies or A1R inhibitors have been shown to significantly inhibit HCC growth by enhancing tumor-killing immunity [143]. Therefore, targeted inhibition of A1R to downregulate pDC recruitment may serve as a potential adjunct strategy for HCC immunotherapy [144]. The immunosuppressive TME and unsatisfactory T-cell persistence are the main causes of nutrient competition problems in the immune micro-environment. For example, researchers have engineered chimeric antigen receptor T (CAR-T) cells to express membrane-bound CD26 and cytoplasmic A1R, converting adenosine to inosine. They found that A1R, in turn, activated CAR-T cells in the autocrine response to CD3/CD26 stimulation, improving migration and resisting TGF- $\beta$ 1 inhibition. The fusion of ADA1 with anti-CD3 single chain antibody fragment (scFv) further promoted the production of inosine and reduced tumor cell uptake. In a mouse model of HCC, these researchers found that metabolically supplemented CAR-T cells showed improved tumor-reduction ability; this new approach has the potential to provide selective inosine supplementation to enhance the efficacy of CAR-T therapy for HCC [145]. In addition, A1R can trigger the signal transduction of the (phosphatidylinositol-3-kinase) PI3K/AKT/glycogen synthase kinase (GSK)-3 $\beta$ / $\beta$ -catenin intracellular carcinogenic pathway [146]. The A1R agonist 2-Chloro-N6-cyclopentyladenosine (CCPA) enhances ischemia-reperfusion (IR) damage, intracellular steatosis, and oxidative species (OS) production, thereby further increasing lipid/OS-dependent apoptosis signal regulating kinase 1 (ASK1)-c-jun N-terminal kinase (JNK) stimulation [147]. In conclusion, several studies have found that A1R deletion inhibits the growth of various tumor cell lines and tumor development in immunodeficient xenografts both in vitro and in vivo [148–150]. Therefore, A1R represents a promising research area (Fig. 6).

### A2a adenosine receptor in HCC

The inhibition of A2aR is a promising cancer immunotherapy approach that is currently being evaluated in several clinical trials [151–153]. In patients with HCC, the RNA expression of *A2aR* in the tumor tissues has been shown to be higher than that in other tumor tissues



**Fig. 6** CD73 is expressed in a variety of cells and binds to a variety of adenosine receptors (e.g., A1R, A2aR, A2bR, and A3R) on the surface of hepatocellular carcinoma (HCC) cells by catalyzing the production of adenosine in the tumor microenvironment (TME), thus activating downstream signaling pathways to play a protective role in the disease process and prognosis of HCC

[154, 155]. Adenosine signaling is regulated by the anoxic immune microenvironment in the liver. After treatment with the A2aR inhibitor SCH58261 combined with anti-PD1, peripheral blood A2aR<sup>+</sup> T cells showed high proliferation in patients with HCC. In an in situ mouse model of HCC, the combination of SCH58261 and anti-PD1 activated T cells and reduced tumor size. This suggests that A2aR blocking promotes an immunotherapy effect in mouse HCC models, highlighting the clinical benefit in patients with advanced HCC [154, 155]. Mechanistically, adenosine produced by CD73 binds to the adenosine A2aR and activates Rap1, which recruits P110β to the plasma membrane and triggers the production of phosphatidylinositol 3-phosphate (PIP3), thereby promoting the phosphorylation of AKT in HCC cells [156]. Recent studies have found that both mouse and human double-negative T (DNT) regulatory cells overexpress CD39, protect DNT from extracellular ATP-induced apoptosis, and produce adenosine together with CD73, thereby

inducing high levels of neutrophil apoptosis [157]. Notably, in human HCC, low expression of A2aR is associated with cirrhosis, liver inflammation, and poor survival, suggesting the cautious use of A2aR antagonists in patients with HCC [158]. Current research is focused on the screening of novel A2aR antagonist candidates with strong A2aR antagonist activity, good hepatic microsomal metabolic stability, and good oral bioavailability for cancer immunotherapy (Fig. 6) [159, 160].

#### A2bR adenosine receptor in HCC

Adenosine is a metabolite that suppresses the anti-tumor immune response of T and NK cells via extracellular binding to two subtypes of A2Rs [161, 162]. CD39 + T-cell infiltration and adenosine receptor A2bR expression levels in the tumor TME have been shown to be negatively correlated with overall survival in patients with HCC [163]. Three-dimensional morphological analysis of the mitochondrial network revealed that these ARs are



potential regulators of mitochondrial energy metabolism due to increased ATP production and coupling efficiency in the presence of BAY 60-6583 (A2bR agonist) [164]. Intracellular ATP is released into the extracellular space and degraded into adenosine, which acts as a signaling molecule to activate the cyclic adenosine phosphate (cAMP) pathway through the A2bR, thereby promoting the polarization of M2 macrophages [165]. The A2bR antagonist ISAM-R56A, can promote the proliferation of T and NK cells, the production of IFN- $\gamma$  and perforin, and increase the infiltration of tumor-infiltrating lymphocytes into tumor spheres, without altering the expression of adhesion molecules. The A2bR is a promising target in immunotherapy, with A2bR and A2aR/A2bR double antagonists showing similar or better results than the A2aR antagonist AZD-4635 [166]. The expression of CAF-CD73 is enhanced by A2bR-mediated feedforward circuitry triggered by tumor cell death, thereby reinforcing the CD73 checkpoint [167]. The activation of A2bR and the induction of NADPH oxidase 2 (NOX2)-dependent oxidative stress within endothelial cells promote the interaction between endothelial and stromal cells, thereby promoting tumor angiogenesis [168]. The human A2bR is a G protein-coupled receptor of class A, which has a relatively low affinity for its ligand adenosine and the A2bR agonist NECA. The two structures of A2bR bind NECA and BAY60-6583. The residues V2506.51 and N2737.36 are the key determinants of its selectivity to A2bR [169]. Single A2bR antagonists and A2aR/A2bR double antagonists with good liver microsomal stability are future research directions for HCC immunotherapy (Fig. 6) [170].

### A3R adenosine receptor in HCC

A3R is overexpressed in human HCC cells [171]. Namodenoson (CF102) is an A3R agonist that induces dysregulation of the Wnt/ $\beta$ -catenin and NF- $\kappa$ B signaling pathways leading to apoptosis in HCC cells and is currently being used in phase III trials in advanced HCC [172, 173]. Additionally, the specific A3R agonist MECA, which has antitumor effects through both A3R-dependent and -independent pathways, has shown antiproliferative effects against various tumor types, especially HCC [174]. Approaches from the opposite direction also have potential, such as the new dual A2aR/A3R nucleoside antagonists that are currently under development, which represent promising candidates for immunoncology [175]. Recent findings indicate that inhibition of A3R by a highly selective antagonist of A3R, FM101, or gene deletion alleviates inflammation and fibrosis in metabolic dysfunction-associated steatotic liver disease (MASLD) by inducing mitochondrial dysfunction and subsequent necrosis of proinflammatory monocyte-derived KCs (MoKCs) [176]. In addition, the A3R agonist

Namodenoson improves liver function/pathology in patients with non-alcoholic fatty liver disease [177]. A3R has become a target for drug development, and the identification of potent and highly selective A3R agonists is the current research direction (Fig. 6) [178–181].

### Anti-HCC agents targeting CD73-adenosine signaling

The overexpression of CD73 and the four types of adenosine receptors in a variety of tumor cells is closely related to the regulation of the TME, inhibition of the tumor immune response, tumor metastasis, drug resistance, and patient prognosis, which also provides a basis for the development of drugs targeting CD73 and adenosine receptors in clinical therapy. At present, many pharmaceutical companies worldwide are actively developing adenosine pathway drug research and development, which is still in the clinical stage. Currently, there are no approved drugs on the market, and the fastest progress of drug development for clinical phase II is currently underway. From the current clinical trials, drug combination represents the most important direction for CD73-adenosine pathway drug development.

### Anti-HCC drug candidates targeting CD73

At present, there are 18 monoclonal antibody drug candidates targeting CD73 in clinical stages worldwide, 17 of which are related to the regulation of the immune microenvironment (Table 1). The bispecific antibody targeting CD73 is represented by AK131 of Akesobio, which is also the only CD73 bispecific antibody to have entered the clinical stage worldwide. AK131 has shown strong activity in vivo and in vitro, not only effectively blocking the PD-1/PD-L1 interaction but also effectively promoting the activation of T and B cells and inducing the endocytosis of CD73 (Table 2). Currently, there are a total of five small molecule drug candidates targeting CD73 entering the clinical stage, four of which are related to HCC, and Quemliclustat is the fastest developing drug worldwide. A small-molecule inhibitor targeting CD73 inhibits AMP and extracellular adenosine-mediated tumor immunosuppression by effectively blocking adenosine production in the TME. The drug candidate was initiated in a phase III trial on September 23, 2024 (PRISM-1/NCT06608927), becoming the first small molecule CD73 inhibitor to enter phase III (Table 3). Currently, two drug candidates of the CD73-antibody-drug conjugate (CD73 ADC) are entering the clinic, both of which are in the first clinical phase. Studies have shown that, compared to uncoupled CD73 naked antibody, CD73 ADCs can induce the accumulation of pro-inflammatory macrophages and activated DCs in tumors. At the same time, CD73 ADCs also have the function of protecting effector T cells and stimulating DCs, which has the dual

**Table 1** Advances in monoclonal antibodies targeting CD73

Drug Candidate	Company	phase	Indication	Type
Oleclumab	AstroZeneca, MedImmune	III	Advanced malignant solid tumor	Antibody
Uliedlimab	I-mab Biopharma Halozyme	II/III	Advanced malignant solid tumor, Gastrointestinal tumor	Antibody
BMS-086179	Therapeutics	II	Advanced malignant solid tumor	Antibody
JAB-BX102	Jacobio Pharmaceuticals	II	Solid tumor, Advanced cancer, Advanced malignant solid tumor	Antibody
Dresbuxelimab	Akesobio		Advanced malignant solid tumor,	
HB0045	Huabo Biopharm	I/II		Antibody
Sym-024	Huaota Biopharm	I/II	Solid tumor	Antibody
Ansipastobart	Symphogen A/S, Servier Group	I/II	Advanced malignant solid tumor,	Antibody
Anti-CD73-mAb	Bioraypharm	I	Solid tumor, Tumor metastasis	Antibody
HBM-1007	BMS	I	Advanced malignant solid tumor, Solid tumor	Antibody
IBI-325	Harbour BioMed	I	Cancer	Antibody
IPH5301	Innovent	I	Cancer	Antibody
Mupadolimab	Innate Pharma	I	Advanced malignant solid tumor, Advanced cancer	Antibody
PM-1015	Corvus Pharmaceuticals	I	Advanced malignant solid tumor	Antibody
PT-199	Biotheus	I	Advanced malignant solid tumor, Advanced cancer	Antibody
Uprevstobart	Phanes Therapeutics	I	Advanced malignant solid tumor	Antibody
Recombinant anti-CD73 mAb	Incyte Corp	I	Advanced malignant solid tumor	Antibody
	Henlix Biotech	Apply	Solid tumor	Antibody

**Table 2** Bispecific antibody (BsAb) targeting CD73

Candidate	Target	Mechanism	Indication	Company	Phrase
AK-131	CD73xPD-1	CD73 antagonist, PD-1 antagonist, ADCC, T lymphocyte stimulant	Advanced malignant solid tumor, Autoimmune disease	AKEsobio	I
AK-137	CD73xLAG3	CD73 conditioning agent, LAG3 conditioning agent	Advanced cancer	AKEsobio	Apply
HB-0046	CD39xCD73	CD39 antagonist, CD73 antagonist	Advanced malignant solid tumor	Huaota Biopharm	Apply

**Table 3** Small molecule drug candidates targeting CD73

Candidate	Target	Mechanism	Indication	Company	Phrase
Quemliclustat	CD73	Blocking adenosine production in the TME inhibits AMP and extracellular adenosine-mediated tumor immunosuppression	Advanced bile duct cancer, cancer	Arcus, Gilead, Sarah Cannon	III
ABSK-051	CD73	Blocking adenosine production in the TME	Advanced malignant solid tumor, cancer	Abbisko Therapeutics	I
CB-708	CD73	Blocking adenosine production in the TME	Locally advanced/metastatic solid tumor	Antengene, Calithera	I
BPI-472,372	CD73	Blocking adenosine production in the TME	Advanced malignant solid tumor	Betta pharma	Apply

**Table 4** Drug candidates of antibody-drug conjugates (ADCs) targeting CD73

Candidate	Target	Mechanism	Indication	Company	Phrase
BB-1709	CD73	Recruit and aggregate macrophages and activated DCs in the tumor	Solid tumor, solid tumor expressing CD73	Bliss Biopharm	I
HB-0052	CD73	Recruit and aggregate macrophages and activated DCs in the tumor	Advanced malignant solid tumor, solid tumor	Huaota Biopharm	I

advantages of killing CD73-highly expressed tumors and improving the tumor immune response. However, the specific effect requires further clinical verification (Table 4).

#### Anti-HCC drug candidates targeting adenosine and its receptors

Adenosine overproduction occurs at all stages of tumorigenesis, making the adenosine pathway an attractive but

**Table 5** Immune therapeutic drug candidates targeting the adenosine pathway

Candidate	Target	Indication	Company	Phrase
CPI-444	A2aR	Combination with a PD-L1 monoclonal antibody and/or CD73 monoclonal antibody for treatment of advanced solid tumors	Roche/Corvus	I/II
PBF-509	A2aR	Combination with a PD-L1 monoclonal antibody and/or CD73 monoclonal antibody for treatment of advanced solid tumors	Novartis	I/II
CS3005	A2aR	Advanced solid tumors	Cstone Pharma	I
SHR5126	A2aR	Advanced solid tumors	Hengrui Pharma	I

challenging therapeutic target. Currently, much of the research in immuno-oncology is focused on restoring immune surveillance, primarily by blocking adenosine-producing enzymes in the TME and adenosine receptors on immune cells through single or combination drugs. The number of clinical trials targeting adenosine pathway components has increased in recent years; however, most trials are still in the early stages of development, with a few being in the clinical stage; additionally, no drugs are approved for marketing, and the most advanced drugs under development have just entered clinical phase II (i.e., NCT02655822, NCT04280328, NCT03207867, NCT04895748, NCT03549000, NCT03381274, NCT04089553, NCT02403193, NCT03720678, NCT03629756, NCT04262856, NCT04892875, NCT05060432, NCT04233060, and NCT03873883) (Table 5).

Treatment and prospects

Overactivation of CD73-adenosine pathways in the TME induces immunosuppressive signals and promotes the development and progression of HCC. The proliferation and metastasis of tumors can be promoted through an immunosuppressive pathway, a mechanism that prevents the immune system from eliminating malignant tumor cells and allows the disease to progress from an early stage to a deadly state. Since the discovery of tumor immunotherapy targeting the “immune-escape” mechanism, great breakthroughs have been made in cancer treatment. Immune checkpoint inhibitors (e.g., PD-1/PD-L1, CTLA-4) have shown excellent therapeutic effects on various tumors. Large- and small-molecule drugs targeting immune checkpoints have also become a research hotspot. Despite these successes, there remain some problems to be solved, such as the limited indications and immune resistance in some patients.

In the CD73-adenosine pathway, CD39, CD73, and A2aR are the three key molecules for the production and function of adenosine. As a biomarker of tumors, the expression level of adenosine pathway-related molecules is significantly correlated with the clinicopathological features and prognosis of tumors. In addition, the adenosine pathway has broad application prospects in the evaluation of immunotherapy effects. A variety of targeted inhibitors have been developed, including CD39

inhibitors, CD73 inhibitors, and A2aR blockers, many of which have shown exciting clinical activity, but also have low selectivity, metabolic instability, low water solubility, or high plasma protein binding degree shortcomings or deficiencies. However, drug research targeting the CD73-adenosine pathway has just begun, and its anti-HCC effect needs to be further confirmed by more large-scale studies. Given that adenosine production depends on the hypoxic environment and cell renewal, in the future, CD73-adenosine pathway inhibitors may be combined with anti-HCC agents that induce hypoxia and cell death. Synergies with other immunotherapies, such as the combination of PD-1, PD-L1, and CTLA-4 antibodies, can also be explored. In conclusion, the CD73-adenosine pathway plays an important role in tumor immunity, and the CD73-adenosine pathway will show great clinical translational potential with the continuous development of research.

The CD73-adenosine pathway is a new target for tumor immunotherapy due to immune system suppression caused by adenosine production in the TME. Through drug screening for molecular targets in the CD73-adenosine pathway, lead compounds, monoclonal antibodies, and small-molecule CD73 inhibitors have been obtained. Blocking antibodies and small molecule inhibitors targeting the CD73-adenosine pathway have shown good anti-tumor efficacy; however, poor metabolic stability, low inhibition efficiency, low selectivity, high potential for drug resistance, and binding site mutations leading to competitive inhibitor resistance, are still the main problems in drug development. CD73 is the main immunosuppressive mediator of the TME, which acts primarily through the production of extracellular adenosine. In addition to the inherent effects of CD73 in tumor cells on tumor cell proliferation, angiogenesis, invasion, and metastasis, the expression of CD73 by tumor and immune cells also weakens anti-tumor immunity by inhibiting the function of protective immune cells (e.g., effector T cells, NK cells, DCs, and B cells) while maintaining the function of regulatory immune cells (e.g., Tregs, MDSCs, TAMs, and CAFs). Improving the response rate of immunotherapy and increasing the efficacy of immunotherapy for treating tumors are the focus of many researchers at present.



## Acknowledgements

All of the figures were created by the authors. We thank LetPub ([www.letpub.com.cn](http://www.letpub.com.cn)) for its linguistic assistance during the preparation of this manuscript.

## Author contributions

LS, MXG and DDZ wrote the main manuscript text. LS prepared the Figs. 1, 2, 3, 4, 5 and 6. LS and MXG wrote the final manuscript together with XYM, JLL and XWL. All authors read and approved the final manuscript.

## Funding

This study was supported by the National Natural Science Foundation of China (Grant 82370602), the Special Doctoral Fund of Hefei Hospital Affiliated to Anhui Medical University (the 2nd People's Hospital of Hefei) (Grant 2020BSZX04), the Key Scientific Research Foundation of the Education Department of the Province Anhui (Grant 2023AH052595), and the Scientific Research Projects of Health Commission of Anhui Province in 2023 (Grant AHWJ2023BAc20099).

## Data availability

No datasets were generated or analysed during the current study.

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

All authors agree with the submission.

### Competing interests

The authors declare no competing interests.

Received: 17 January 2025 / Accepted: 10 May 2025

Published online: 26 May 2025

## References

- Cheng C, Zha Q, Sun L, et al. VCP downstream metabolite glycerol-3-phosphate (G3P) inhibits CD8+T cells function in the HCC microenvironment. *Signal Transduct Target Ther*. 2025;10(1):26.
- Gu L, Zhu Y, Nandi SP, et al. FBP1 controls liver cancer evolution from senescent MASH hepatocytes. *Nature*. 2025;637(8045):461–9.
- Lu E, Wolfreys FD, Muppidi JR, et al. S-Geranylgeranyl-L-glutathione is a ligand for human B cell-confinement receptor P2RY8. *Nature*. 2019;567(7747):244–8.
- Dudek M, Pfister D, Donakonda S, et al. Auto-aggressive CXCR6+ CD8 T cells cause liver immune pathology in NASH. *Nature*. 2021;592(7854):444–9.
- Gutiérrez-Herrero S, Fernández-Infante C, Hernández-Cano L, et al. C3G contributes to platelet activation and aggregation by regulating major signaling pathways. *Signal Transduct Target Ther*. 2020;5(1):29.
- Haffke M, Fehlmann D, Rummel G, et al. Structural basis of species-selective antagonist binding to the succinate receptor. *Nature*. 2019;574(7779):581–5.
- Chang-Graham AL, Perry JL, Engevik MA, et al. Rotavirus induces intercellular calcium waves through ADP signaling. *Science*. 2020;370(6519):eabc3621.
- Ishikawa M, Hasanali ZS, Zhao Y, et al. Bone marrow plasma cells require P2RX4 to sense extracellular ATP. *Nature*. 2024;626(8001):1102–7.
- Ni C, Fang QQ, Chen WZ, et al. Breast cancer-derived exosomes transmit LncRNA SNHG16 to induce CD73+ γδ1 Treg cells. *Signal Transduct Target Ther*. 2020;5(1):41.
- Allard D, Cormery J, Bricha S, et al. Adenosine uptake through the nucleoside transporter ENT1 suppresses antitumor immunity and T-cell pyrimidine synthesis. *Cancer Res*. 2025;85(4):692–703.
- García-Carrillo R, Molina-Pelayo FA, Zarate-Lopez D, et al. An adenosine derivative promotes mitochondrial supercomplexes reorganization and restoration of mitochondria structure and bioenergetics in a diethylnitrosamine-induced hepatocellular carcinoma model. *Sci Rep*. 2024;14(1):6348.
- Gammelgaard OL, Terp MG, Renn C, et al. Targeting two distinct epitopes on human CD73 with a bispecific antibody improves anticancer activity. *J Immunother Cancer*. 2022;10(9):e004554.
- Jin H, Li M, Wang X, et al. Purinergic signaling by TCRαβ+ double-negative T regulatory cells ameliorates liver ischemia-reperfusion injury. *Sci Bull (Beijing)*. 2025;70(2):241–54.
- Zou Z, Lin Z, Wu C, et al. Micro-Engineered Organoid-on-a-Chip based on mesenchymal stromal cells to predict immunotherapy responses of HCC patients. *Adv Sci (Weinh)*. 2023;10(27):e2302640.
- Thiel V, Renders S, Panten J et al. Characterization of single neurons reprogrammed by pancreatic cancer. *Nature*. 2025 Feb 17.
- Zhu GQ, Tang Z, Chu TH, et al. Targeting SRSF1 improves cancer immunotherapy by dually acting on CD8+T and tumor cells. *Signal Transduct Target Ther*. 2025;10(1):25.
- Kerzel T, Giacca G, Beretta S, et al. In vivo macrophage engineering reshapes the tumor microenvironment leading to eradication of liver metastases. *Cancer Cell*. 2023;41(11):1892–e191010.
- Zhang P, Chen Z, Kuang H, et al. Neuregulin 4 suppresses NASH-HCC development by restraining tumor-prone liver microenvironment. *Cell Metab*. 2022;34(9):1359–e13767.
- Zhang YP, Guo ZQ, Cai XT, et al. PAI-1-driven SFRP2high cancer-associated fibroblasts hijack the abscopal effect of radioimmunotherapy. *Cancer Cell*. 2025 Mar;125:1535. 6108(25)00076–5.
- Sun Y, Wu P, Zhang Z, et al. Integrated multi-omics profiling to dissect the spatiotemporal evolution of metastatic hepatocellular carcinoma. *Cancer Cell*. 2024;42(1):135–e15617.
- Lu Y, Liu Y, Zuo X, et al. CXCL12+ tumor-associated endothelial cells promote immune resistance in hepatocellular carcinoma. *J Hepatol*. 2025;82(4):634–48.
- Melero I, de Miguel Luken M, de Velasco G, et al. Neutralizing GDF-15 can overcome anti-PD-1 and anti-PD-L1 resistance in solid tumours. *Nature*. 2025;637(8048):1218–27.
- Martínez-Ordoñez A, Duran A, Ruiz-Martínez M, et al. Hyaluronan driven by epithelial aPKC deficiency remodels the microenvironment and creates a vulnerability in mesenchymal colorectal cancer. *Cancer Cell*. 2023;41(2):252–e2719.
- Zhou Y, Wang J, Chen Y et al. NR1R promotes immune escape in hepatocellular cancer by regulating IFNγ-induced PD-L1 expression. *J Adv Res* 2025 Feb 27:S2090-1232(25)00133-X.
- Lu Y, Sun Q, Guan Q, et al. The XOR-IDH3a axis controls macrophage polarization in hepatocellular carcinoma. *J Hepatol*. 2023;79(5):1172–84.
- Fu Y, Mackowiak B, Feng D, et al. MicroRNA-223 attenuates hepatocarcinogenesis by blocking hypoxia-driven angiogenesis and immunosuppression. *Gut*. 2023;72(10):1942–58.
- Liu Y, Zhang Q, Xing B, et al. Immune phenotypic linkage between colorectal cancer and liver metastasis. *Cancer Cell*. 2022;40(4):424–e4375.
- Lu Y, Yang A, Quan C, et al. A single-cell atlas of the multicellular ecosystem of primary and metastatic hepatocellular carcinoma. *Nat Commun*. 2022;13(1):4594.
- Zhao S, Mi Y, Zheng B, et al. Highly-metastatic colorectal cancer cell released miR-181a-5p-rich extracellular vesicles promote liver metastasis by activating hepatic stellate cells and remodelling the tumour microenvironment. *J Extracell Vesicles*. 2022;11(1):e12186.
- Liu X, Liu J, Wang X, et al. Cancer-secreted Exosomal miR-1246 promotes colorectal cancer liver metastasis by activating hepatic stellate cells. *Mol Med*. 2025;31(1):68.
- Wang K, Liu J, Hai P, et al. Novel angiogenesis inhibitors with superoxide anion radical amplification effect: surmounting the Achilles' heels of angiogenesis inhibitors and photosensitizers. *Eur J Med Chem*. 2024;272:116495.
- Montemagno C, Durivault J, Gastaldi C, et al. A group of novel VEGF splice variants as alternative therapeutic targets in renal cell carcinoma. *Mol Oncol*. 2023;17(7):1379–401.
- Zhang K, Zhang H, Zou XR, et al. An antibody-like peptidic network for anti-angiogenesis. *Biomaterials*. 2021;275:120900.
- Ko J, Hyung S, Heo YJ, et al. Patient-derived tumor spheroid-induced angiogenesis preclinical platform for exploring therapeutic vulnerabilities in cancer. *Biomaterials*. 2024;306:122504.
- Ma B, Wen S, Luo Y, et al. Targeting tumor hypoxia inhibits aggressive phenotype of dedifferentiated thyroid Cancer. *J Clin Endocrinol Metab*. 2023;108(2):368–84.
- Zhu X, Chen J, Li W, et al. Hypoxia-Responsive CAR-T cells exhibit reduced exhaustion and enhanced efficacy in solid tumors. *Cancer Res*. 2024;84(1):84–100.
- Li X, Li M, Huang M, et al. Hypoxic response patterns in lung tissue: an integrated analysis of comparative physiological and transcriptomic

- studies from *Neodon fuscus* and *Lasiopodomys brandtii*. *Sci Total Environ.* 2023;892:164537.
38. Huang CH, Chong KY, Lei KF. Analysis of the internal hypoxic environment in solid tumor tissue using a folding paper system. *ACS Appl Mater Interfaces.* 2021;13(29):33885–93.
39. Zhong C, Niu Y, Liu W, et al. S100A9 derived from Chemoembolization-Induced hypoxia governs mitochondrial function in hepatocellular carcinoma progression. *Adv Sci (Weinh).* 2022;9(30):e2202206.
40. Chen Y, Yan H, Yan L, et al. Hypoxia-induced ALDH3A1 promotes the proliferation of non-small-cell lung cancer by regulating energy metabolism reprogramming. *Cell Death Dis.* 2023;14(9):617.
41. Ma J, Al Moussawi K, Lou H, et al. Deficiency of factor-inhibiting HIF creates a tumor-promoting immune microenvironment. *Proc Natl Acad Sci U S A.* 2024;121(10):e2309957121.
42. Chen W, He C, Qiao N, et al. Dual drugs decorated bacteria irradiate deep hypoxic tumor and arouse strong immune responses. *Biomaterials.* 2022;286:121582.
43. Sun J, Zhao Z, Lu J, et al. The tumor microenvironment mediates the HIF-1 $\alpha$ /PD-L1 pathway to promote immune escape in colorectal Cancer. *Int J Mol Sci.* 2024;25(7):3735.
44. Li M, Zhang X, Wang M, et al. Activation of Piezo1 contributes to matrix stiffness-induced angiogenesis in hepatocellular carcinoma. *Cancer Commun (Lond).* 2022;42(11):1162–84.
45. Huang R, Zhang L, Jin J, et al. Bruceine D inhibits HIF-1 $\alpha$ -mediated glucose metabolism in hepatocellular carcinoma by blocking ICAT/ $\beta$ -catenin interaction. *Acta Pharm Sin B.* 2021;11(11):3481–92.
46. Salman S, Meyers DJ, Wicks EE, et al. HIF inhibitor 32-134D eradicates murine hepatocellular carcinoma in combination with anti-PD1 therapy. *J Clin Invest.* 2022;132(9):e156774.
47. Yang Z, Qiao C, Jia Q, et al. Redox dyshomeostasis modulation of the tumor intracellular environment through a metabolic intervention strategy for enhanced photodynamic therapy. *Theranostics.* 2022;12(14):6143–54.
48. Jarvis LB, Rainbow DB, Coppard V, et al. Therapeutically expanded human regulatory T-cells are super-suppressive due to HIF1A induced expression of CD73. *Commun Biol.* 2021;4(1):1186.
49. Yang Y, Lu H, Chen C, et al. HIF-1 interacts with TRIM28 and DNA-PK to release paused RNA polymerase II and activate target gene transcription in response to hypoxia. *Nat Commun.* 2022;13(1):316.
50. Baumeister J, Chatain N, Hubrich A, et al. Hypoxia-inducible factor 1 (HIF-1) is a new therapeutic target in JAK2V617F-positive myeloproliferative neoplasms. *Leukemia.* 2020;34(4):1062–74.
51. Seo J, Jeong DW, Park JW, et al. Fatty-acid-induced FABP5/HIF-1 reprograms lipid metabolism and enhances the proliferation of liver cancer cells. *Commun Biol.* 2020;3(1):638.
52. Zheng Y, Huang C, Lu L, et al. STOML2 potentiates metastasis of hepatocellular carcinoma by promoting PINK1-mediated mitophagy and regulates sensitivity to lenvatinib. *J Hematol Oncol.* 2021;14(1):16.
53. Guo W, Tan HY, Li S, et al. Glutamic-Pyruvic transaminase 1 facilitates alternative fuels for hepatocellular carcinoma Growth-A small molecule inhibitor, Berberine. *Cancers (Basel).* 2020;12(7):1854.
54. Theparambil SM, Kopach O, Braga A, et al. Adenosine signalling to astrocytes coordinates brain metabolism and function. *Nature.* 2024;632(8023):139–46.
55. Klysz DD, Fowler C, Malipatlolla M, et al. Inosine induces stemness features in CAR-T cells and enhances potency. *Cancer Cell.* 2024;42(2):266–e2828.
56. Gou K, Xin W, Lv J, et al. A pH-responsive chiral mesoporous silica nanoparticles for delivery of doxorubicin in tumor-targeted therapy. *Colloids Surf B Biointerfaces.* 2023;221:113027.
57. Yin C, Sun M, Yan Z, et al. pH-Responsive Plasmon-Enhanced persistent luminescent ZnGa<sub>2</sub>O<sub>4</sub>: Cr<sup>3+</sup> + Nanopomegranate for tumor imaging. *ACS Appl Mater Interfaces.* 2023;15(48):55323–34.
58. Yan Y, Li J, Yi X, et al. Peritumoral scaffold neutralizes tumor pH for chemotherapy sensitization and metastasis inhibition. *J Control Release.* 2022;352:747–58.
59. Jin HS, Choi DS, Ko M, et al. Extracellular pH modulating injectable gel for enhancing immune checkpoint inhibitor therapy. *J Control Release.* 2019;315:65–75.
60. Wang D, Zhou N, Zhang N, et al. Facile Preparation of pH/redox dual-responsive biodegradable polyphosphazene prodrugs for effective cancer chemotherapy. *Colloids Surf B Biointerfaces.* 2021;200:111573.
61. Ippolito L, Duatti A, Iozzo M, et al. Lactate supports cell-autonomous ECM production to sustain metastatic behavior in prostate cancer. *EMBO Rep.* 2024;25(8):3506–31.
62. Lien EC, Westermark AM, Zhang Y, et al. Low glycaemic diets alter lipid metabolism to influence tumour growth. *Nature.* 2021;599(7884):302–7.
63. Zhang J, Wang S, Guo X, et al. Arginine supplementation targeting tumor-Killing immune cells reconstructs the tumor microenvironment and enhances the antitumor immune response. *ACS Nano.* 2022;16(8):12964–78.
64. Kurahashi T, Yoshida Y, Ogura S, et al. Forkhead box M1 transcription factor drives liver inflammation linking to hepatocarcinogenesis in mice. *Cell Mol Gastroenterol Hepatol.* 2020;9(3):425–46.
65. Elia I, Rowe JH, Johnson S, et al. Tumor cells dictate anti-tumor immune responses by altering pyruvate utilization and succinate signaling in CD8+ T cells. *Cell Metab.* 2022;34(8):1137–e11506.
66. Lin L, Chen S, Wang H, et al. SPTBN1 inhibits inflammatory responses and hepatocarcinogenesis via the stabilization of SOCS1 and downregulation of p65 in hepatocellular carcinoma. *Theranostics.* 2021;11(9):4232–50.
67. Wang S, Zhu L, Li T, et al. Disruption of MerTK increases the efficacy of checkpoint inhibitor by enhancing ferroptosis and immune response in hepatocellular carcinoma. *Cell Rep Med.* 2024;5(2):101415.
68. Wei CY, Zhu MX, Zhang PF, et al. PKC $\alpha$ /ZFP64/CSF1 axis resets the tumor microenvironment and fuels anti-PD1 resistance in hepatocellular carcinoma. *J Hepatol.* 2022;77(1):163–76.
69. Chen X, Liu X, Zhang Y, et al. Methyltransferase Dot1l preferentially promotes innate IL-6 and IFN- $\beta$  production by mediating H3K79me2/3 methylation in macrophages. *Cell Mol Immunol.* 2020;17(1):76–84.
70. Tsu BV, Beierschmitt C, Ryan AP, et al. Diverse viral proteases activate the NLRP1 inflammasome. *Elife.* 2021;10:e60609.
71. Niu N, Shen X, Zhang L, et al. Tumor Cell-Intrinsic SETD2 deficiency reprograms neutrophils to foster immune escape in pancreatic tumorigenesis. *Adv Sci (Weinh).* 2023;10(2):e2202937.
72. Zhou Q, Tian W, Jiang Z, et al. A positive feedback loop of AKR1C3-Mediated activation of NF- $\kappa$ B and STAT3 facilitates proliferation and metastasis in hepatocellular carcinoma. *Cancer Res.* 2021;81(5):1361–74.
73. Hong L, Mei J, Sun X et al. Spatial single-cell proteomics landscape decodes the tumor microenvironmental ecosystem of intrahepatic cholangiocarcinoma. *Hepatology.* 2025 Feb 25.
74. Han S, Bao X, Zou Y, et al. d-lactate modulates M2 tumor-associated macrophages and remodels immunosuppressive tumor microenvironment for hepatocellular carcinoma. *Sci Adv.* 2023;9(29):eadg2697.
75. Sathe A, Mason K, Grimes SM, et al. Colorectal Cancer metastases in the liver Establish immunosuppressive Spatial networking between Tumor-Associated SPP1 + Macrophages and fibroblasts. *Clin Cancer Res.* 2023;29(1):244–60.
76. Kwong TT, Xiong Z, Zhang Y, et al. Overcoming immunotherapy resistance in hepatocellular carcinoma by targeting myeloid IL-8/CXCR2 signaling. *Mol Ther.* 2025;33(4):1659–73.
77. Shi Y, Niu Y, Yuan Y, et al. PRMT3-mediated arginine methylation of IGF2BP1 promotes oxaliplatin resistance in liver cancer. *Nat Commun.* 2023;14(1):1932.
78. Wang X, Zhang Q, Zhou J, et al. T cell-mediated targeted delivery of Tadalafil regulates immunosuppression and polyamine metabolism to overcome immune checkpoint Blockade resistance in hepatocellular carcinoma. *J Immunother Cancer.* 2023;11(2):e006493.
79. Yang T, Zhang S, Yuan H, et al. Platinum-Based TREM2 inhibitor suppresses tumors by remodeling the immunosuppressive microenvironment. *Angew Chem Int Ed Engl.* 2023;62(2):e202213337.
80. Chen Z, Zhang G, Ren X, et al. Cross-talk between myeloid and B cells shapes the distinct microenvironments of primary and secondary liver Cancer. *Cancer Res.* 2023;83(21):3544–61.
81. Kattelus R, Starskaia I, Lindén M, et al. Phenotypic profiling of human induced regulatory T cells at early differentiation: insights into distinct immunosuppressive potential. *Cell Mol Life Sci.* 2024;81(1):399.
82. Neo SY, Yang Y, Record J, et al. CD73 immune checkpoint defines regulatory NK cells within the tumor microenvironment. *J Clin Invest.* 2020;130(3):1185–98.
83. Badimon A, Strasburger HJ, Ayata P, et al. Negative feedback control of neuronal activity by microglia. *Nature.* 2020;586(7829):417–23.
84. Jin D, Fan J, Wang L, et al. CD73 on tumor cells impairs antitumor T-cell responses: a novel mechanism of tumor-induced immune suppression. *Cancer Res.* 2010;70(6):2245–55.
85. Deng Y, Chen Q, Yang X, et al. Tumor cell senescence-induced macrophage CD73 expression is a critical metabolic immune checkpoint in the aging tumor microenvironment. *Theranostics.* 2024;14(3):1224–40.
86. Zhuge A, Li S, Han S, et al. Akkermansia muciniphila-derived acetate activates the hepatic AMPK/SIRT1/PGC-1 $\alpha$  axis to alleviate ferroptosis in metabolic-associated fatty liver disease. *Acta Pharm Sin B.* 2025;15(1):151–67.

87. Dei Zotti F, Qiu A, D'Agati VD, et al. Mitigation of checkpoint inhibitor-induced autoimmune hemolytic anemia through modulation of purinergic signaling. *Blood*. 2024;144(15):1581–94.
88. Tang T, Huang X, Lu M, et al. Transcriptional control of pancreatic cancer immunosuppression by metabolic enzyme CD73 in a tumor-autonomous and -autocrine manner. *Nat Commun*. 2023;14(1):3364.
89. Faraoni EY, Singh K, Chandra V, et al. CD73-Dependent adenosine signaling through Adora2b drives immunosuppression in ductal pancreatic Cancer. *Cancer Res*. 2023;83(7):1111–27.
90. Leone RD, Sun IM, Oh MH, et al. Inhibition of the adenosine A2a receptor modulates expression of T cell coinhibitory receptors and improves effector function for enhanced checkpoint Blockade and ACT in murine cancer models. *Cancer Immunol Immunother*. 2018;67(8):1271–84.
91. Lu JC, Zhang PF, Huang XY, et al. Amplification of spatially isolated adenosine pathway by tumor-macrophage interaction induces anti-PD1 resistance in hepatocellular carcinoma. *J Hematol Oncol*. 2021;14(1):200.
92. Yu Y, Zhang C, Dong B, et al. Neutrophil extracellular traps promote immune escape in hepatocellular carcinoma by up-regulating CD73 through Notch2. *Cancer Lett*. 2024;598:217098.
93. Kim M, Min YK, Jang J, et al. Single-cell RNA sequencing reveals distinct cellular factors for response to immunotherapy targeting CD73 and PD-1 in colorectal cancer. *J Immunother Cancer*. 2021;9(7):e002503.
94. Alcedo KP, Guerrero A, Basur V, et al. Tumor-Selective altered glycosylation and functional Attenuation of CD73 in human hepatocellular carcinoma. *Hepatol Commun*. 2019;3(10):1400–14.
95. Gong Y, Cheng Y, Zeng F, et al. A self-gelling hemostatic powder boosting radiotherapy-elicited NK cell immunity to combat postoperative hepatocellular carcinoma relapse. *Biomaterials*. 2025;317:123068.
96. Liu Y, Xun Z, Ma K, et al. Identification of a tumour immune barrier in the HCC microenvironment that determines the efficacy of immunotherapy. *J Hepatol*. 2023;78(4):770–82.
97. Allard D, Cousineau I, Ma EH, et al. The CD73 immune checkpoint promotes tumor cell metabolic fitness. *Elife*. 2023;12:e84508.
98. Cheu JW, Chiu DK, Kwan KK, et al. Hypoxia-inducible factor orchestrates adenosine metabolism to promote liver cancer development. *Sci Adv*. 2023;9(18):eade5111.
99. Xia P, Zhang H, Lu H, et al. METTL5 stabilizes c-Myc by facilitating USP5 translation to reprogram glucose metabolism and promote hepatocellular carcinoma progression. *Cancer Commun (Lond)*. 2023;43(3):338–64.
100. Mao C, Yeh S, Fu J, et al. Delivery of an ectonucleotidase inhibitor with ROS-responsive nanoparticles overcomes adenosine-mediated cancer immunosuppression. *Sci Transl Med*. 2022;14(648):eabh1261.
101. Toller-Kawahisa JE, Viacava PR, Palsson-McDermott EM, et al. Metabolic reprogramming of macrophages by PKM2 promotes IL-10 production via adenosine. *Cell Rep*. 2025;44(1):115172.
102. Wen X, Xiong X, Yang G, et al. A macrophage membrane-coated mesoporous silica nanoplateform inhibiting adenosine A2AR via in situ oxygen supply for immunotherapy. *J Control Release*. 2023;353:535–48.
103. Chambers AM, Lupo KB, Wang J, et al. Engineered natural killer cells impede the immunometabolic CD73-adenosine axis in solid tumors. *Elife*. 2022;11:e73699.
104. Stirn K, Leary P, Wüst D, et al. Treg-selective IL-2 starvation synergizes with CD40 activation to sustain durable responses in lymphoma models. *J Immunother Cancer*. 2023;11(2):e006263.
105. Edmunds GL, Wong CCW, Ambler R, et al. Adenosine 2A receptor and TIM3 suppress cytolytic killing of tumor cells via cytoskeletal polarization. *Commun Biol*. 2022;5(1):9.
106. Zhang H, Feng L, de Andrade Mello P, et al. Glycoengineered anti-CD39 promotes anticancer responses by depleting suppressive cells and inhibiting angiogenesis in tumor models. *J Clin Invest*. 2022;132(13):e157431.
107. Shao B, Wang HD, Ren SH, et al. Exosomes derived from a mesenchymal-like endometrial regenerative cells ameliorate renal ischemia reperfusion injury through delivery of CD73. *Stem Cell Res Ther*. 2025;16(1):148.
108. He J, Duan P, Liu Y, et al. Unveiling the impact of hemodynamics on endothelial Inflammation-Mediated hepatocellular carcinoma metastasis using a biomimetic vascular flow model. *Adv Healthc Mater*. 2024;13(18):e2304439.
109. Zhang L, Xu J, Zhou S, et al. Endothelial DGKG promotes tumor angiogenesis and immune evasion in hepatocellular carcinoma. *J Hepatol*. 2024;80(1):82–98.
110. Luo J, Gong L, Yang Y, et al. Enhanced mitophagy driven by ADAR1-GLI1 editing supports the self-renewal of cancer stem cells in HCC. *Hepatology*. 2024;79(1):61–78.
111. Li Y, Sun J, Li D, et al. The full activation mechanism of the adenosine A1 receptor revealed by GaMD and Su-GaMD simulations. *Proc Natl Acad Sci U S A*. 2022;119(42):e2203702119.
112. Zhang X, Greve PF, Minh TTN, et al. Extracellular vesicles from seminal plasma interact with T cells in vitro and drive their differentiation into regulatory T-cells. *J Extracell Vesicles*. 2024;13(7):e12457.
113. Li R, Lei Y, Rezk A, et al. Oxidative phosphorylation regulates B cell effector cytokines and promotes inflammation in multiple sclerosis. *Sci Immunol*. 2024;9(95):eadk0865.
114. Mohammadpour H, MacDonald CR, Qiao G, et al.  $\beta$ 2 adrenergic receptor-mediated signaling regulates the immunosuppressive potential of myeloid-derived suppressor cells. *J Clin Invest*. 2019;129(12):5537–52.
115. Chen Q, Yin H, He J, et al. Tumor microenvironment responsive CD8+T cells and Myeloid-Derived suppressor cells to trigger CD73 inhibitor AB680-Based synergistic therapy for pancreatic Cancer. *Adv Sci (Weinh)*. 2023;10(33):e2302498.
116. Schneider E, Winzer R, Rissiek A, et al. CD73-mediated adenosine production by CD8 T cell-derived extracellular vesicles constitutes an intrinsic mechanism of immune suppression. *Nat Commun*. 2021;12(1):5911.
117. Miller RA, Luke JJ, Hu S, et al. Anti-CD73 antibody activates human B cells, enhances humoral responses and induces redistribution of B cells in patients with cancer. *J Immunother Cancer*. 2022;10(12):e005802.
118. Ma XL, Hu B, Tang WG, et al. CD73 sustained cancer-stem-cell traits by promoting SOX9 expression and stability in hepatocellular carcinoma. *J Hematol Oncol*. 2020;13(1):11.
119. Wang H, Wei X, Liu L, et al. Suppression of A-to-I RNA-editing enzyme ADAR1 sensitizes hepatocellular carcinoma cells to oxidative stress through regulating Keap1/Nrf2 pathway. *Exp Hematol Oncol*. 2024;13(1):30.
120. Shali S, Yu J, Zhang X, et al. Ecto-5'-nucleotidase (CD73) is a potential target of hepatocellular carcinoma. *J Cell Physiol*. 2019;234(7):10248–59.
121. Ploeg EM, Samplonius DF, Xiong X, et al. Bispecific antibody CD73xEGFR more selectively inhibits the CD73/adenosine immune checkpoint on cancer cells and concurrently counteracts pro-oncogenic activities of CD73 and EGFR. *J Immunother Cancer*. 2023;11(9):e006837.
122. Sciarra A, Monteiro I, Ménétrier-Caux C, et al. CD73 expression in normal and pathological human hepatobiliarypancreatic tissues. *Cancer Immunol Immunother*. 2019;68(3):467–78.
123. Mierzejewska P, Zabielska MA, Kutryb-Zajac B, et al. Impaired L-arginine metabolism marks endothelial dysfunction in CD73-deficient mice. *Mol Cell Biochem*. 2019;458(1–2):133–42.
124. Kwon JH, Lee J, Kim J, et al. HIF-1 $\alpha$  regulates A2B adenosine receptor expression in liver cancer cells. *Exp Ther Med*. 2019;18(6):4231–40.
125. Monguió-Tortajada M, Prat-Vidal C, Martínez-Falguera D, et al. Acellular cardiac scaffolds enriched with MSC-derived extracellular vesicles limit ventricular remodelling and exert local and systemic Immunomodulation in a myocardial infarction Porcine model. *Theranostics*. 2022;12(10):4656–70.
126. Ai L, Lin S, Huang C, et al. Simultaneous interference of SP1 and HIF1 $\alpha$  retarding the proliferation, migration, and invasion of human microvascular endothelial cells (HMEC-1) under hypoxia. *J Cell Biochem*. 2019;120(10):17912–25.
127. Wang Y, Zhang J, Shi H, et al. M2 Tumor-Associated Macrophages-Derived Exosomal MALAT1 promotes Glycolysis and gastric Cancer progression. *Adv Sci (Weinh)*. 2024;11(24):e2309298.
128. Ng MSF, Kwok I, Tan L, et al. Deterministic reprogramming of neutrophils within tumors. *Science*. 2024;383(6679):eadf6493.
129. Wang J, Wang Y, Chu Y, et al. Tumor-derived adenosine promotes macrophage proliferation in human hepatocellular carcinoma. *J Hepatol*. 2021;74(3):627–37.
130. Yi C, Chen L, Lin Z, et al. Lenvatinib targets FGF receptor 4 to enhance antitumor immune response of Anti-Programmed cell Death-1 in HCC. *Hepatology*. 2021;74(5):2544–60.
131. Gan WL, Ren X, Ng VHE, et al. Hepatocyte-macrophage crosstalk via the PGRN-EGFR axis modulates ADAR1-mediated immunity in the liver. *Cell Rep*. 2024;43(7):114400.
132. Lillo A, Raich I, Lillo J, et al. Expression of the adenosine A2A-A3 receptor heteromer in different brain regions and marked upregulation in the microglia of the Transgenic APPSw, Ind Alzheimer's disease model. *Biomedicines*. 2022;10(2):214.
133. Devi VJ, Radhika A, Biju PG. Adenosine receptor activation promotes macrophage class switching from LPS-induced acute inflammatory M1 to anti-inflammatory M2 phenotype. *Immunobiology*. 2023;228(3):152362.



134. Chen S, Akdemir I, Fan J, et al. The expression of adenosine A2B receptor on Antigen-Presenting cells suppresses CD8+T-cell responses and promotes tumor growth. *Cancer Immunol Res*. 2020;8(8):1064–74.
135. Gourdin N, Bossennec M, Rodriguez C, et al. Autocrine adenosine regulates tumor polyfunctional CD73 + CD4 + Effector T cells devoid of immune checkpoints. *Cancer Res*. 2018;78(13):3604–18.
136. Zhang T, Liu H, Jiao L, et al. Genetic characteristics involving the PD-1/PD-L1/L2 and CD73/A2aR axes and the immunosuppressive microenvironment in DLBCL. *J Immunother Cancer*. 2022;10(4):e004114.
137. Chen S, Fan J, Xie P, et al. CD8+T cells sustain antitumor response by mediating crosstalk between adenosine A2A receptor and glutathione/GPX4. *J Clin Invest*. 2024;134(8):e170071.
138. Zhang H, Han K, Li H, et al. hPMSCs regulate the level of TNF- $\alpha$  and IL-10 in Th1 cells and improve hepatic injury in a GVHD mouse model via CD73/ADO/Fyn/Nrf2 Axis. *Inflammation*. 2024;47(1):244–63.
139. Renauer P, Park JJ, Bai M, et al. Immunogenetic metabolomics reveals key enzymes that modulate CAR T-cell metabolism and function. *Cancer Immunol Res*. 2023;11(8):1068–84.
140. Gatsiou A, Tual-Chalot S, Napoli M, et al. The RNA editor ADAR2 promotes immune cell trafficking by enhancing endothelial responses to interleukin-6 during sterile inflammation. *Immunity*. 2023;56(5):979–e99711.
141. Pang L, Ng KT, Liu J, et al. Plasmacytoid dendritic cells recruited by HIF-1 $\alpha$ /eADO/ADORA1 signaling induce immunosuppression in hepatocellular carcinoma. *Cancer Lett*. 2021;522:80–92.
142. Yang M, Zhang Z, Chen J, et al. Soluble fibrinogen-like protein 2 promotes the growth of hepatocellular carcinoma via attenuating dendritic cell-mediated cytotoxic T cell activity. *J Exp Clin Cancer Res*. 2019;38(1):351.
143. El-Sayed Ebead E, Aboelnaga A, Nassar E, et al. Ultrasonic-induced synthesis of novel diverse arylidenes via Knoevenagel condensation reaction. *Antitumor, QSAR, Docking and DFT assessment*. *RSC Adv*. 2023;13(42):29749–67.
144. Sun D, Ko M, Shao H, et al. Adenosine receptor ligation tips the uveitogenic Th1 and Th17 balance towards the latter in experimental autoimmune uveitis-induced mouse. *Curr Res Immunol*. 2021;2:93–103.
145. Hu Y, Sarkar A, Song K, et al. Selective refueling of CAR T cells using ADA1 and CD26 boosts antitumor immunity. *Cell Rep Med*. 2024;5(5):101530.
146. Pan S, Liang S, Wang X. ADORA1 promotes nasopharyngeal carcinoma cell progression through regulation of PI3K/AKT/GSK-3 $\beta$ / $\beta$ -catenin signaling. *Life Sci*. 2021;278:119581.
147. Alchera E, Chandrashekar BR, Clemente N, et al. Ischemia/Reperfusion injury of fatty liver is protected by A2AR and exacerbated by A1R stimulation through opposite effects on ASK1 activation. *Cells*. 2021;10(11):3171.
148. Liu H, Kuang X, Zhang Y, et al. ADORA1 Inhibition promotes tumor immune evasion by regulating the ATF3-PD-L1 Axis. *Cancer Cell*. 2020;37(3):324–e3398.
149. Choi SJ, Ryu E, Lee S, et al. Adenosine induces EBV lytic reactivation through ADORA1 in EBV-Associated gastric carcinoma. *Int J Mol Sci*. 2019;20(6):1286.
150. Saito M, Yaguchi T, Yasuda Y, et al. Adenosine suppresses CW2 human colonic cancer growth by inducing apoptosis via A(1) adenosine receptors. *Cancer Lett*. 2010;290(2):211–5.
151. Young A, Ngiew SF, Gao Y, et al. A2AR adenosine signaling suppresses natural killer cell maturation in the tumor microenvironment. *Cancer Res*. 2018;78(4):1003–16.
152. Lyu A, Fan Z, Clark M, et al. Evolution of myeloid-mediated immunotherapy resistance in prostate cancer. *Nature*. 2025;637(8048):1207–17.
153. Nascimento DC, Viacava PR, Ferreira RG, et al. Sepsis expands a CD39+ plasmablast population that promotes immunosuppression via adenosine-mediated inhibition of macrophage antimicrobial activity. *Immunity*. 2021;54(9):2024–e20418.
154. Myojin Y, McCallen JD, Ma C, et al. Adenosine A2a receptor Inhibition increases the anti-tumor efficacy of anti-PD1 treatment in murine hepatobiliary cancers. *JHEP Rep*. 2023;6(1):100959.
155. Liu Y, Liu Y, Xu D, et al. Targeting the negative feedback of Adenosine-A2AR metabolic pathway by a tailored nanoinhibitor for photothermal immunotherapy. *Adv Sci (Weinh)*. 2022;9(14):e2104182.
156. Ma XL, Shen MN, Hu B, et al. CD73 promotes hepatocellular carcinoma progression and metastasis via activating PI3K/AKT signaling by inducing Rap1-mediated membrane localization of P110 $\beta$  and predicts poor prognosis. *J Hematol Oncol*. 2019;12(1):37.
157. Hamed A, Ghareeb D, Mohamed TM, et al. Caffeine-folic acid-loaded-chitosan nanoparticles combined with methotrexate as a novel HepG2 immunotherapy targeting adenosine A2A receptor downstream cascade. *BMC Complement Med Ther*. 2023;23(1):384.
158. Allard B, Jacobberger-Foissac C, Cousineau I, et al. Adenosine A2A receptor is a tumor suppressor of NASH-associated hepatocellular carcinoma. *Cell Rep Med*. 2023;4(9):101188.
159. Zhu C, Ze S, Zhou R, et al. Discovery of pyridinone derivatives as potent, selective, and orally bioavailable adenosine A2A receptor antagonists for Cancer immunotherapy. *J Med Chem*. 2023;66(7):4734–54.
160. Sun Y, Liu C, He L. Adenosine A2A receptor antagonist Sch58261 improves the cognitive function in Alzheimer's disease model mice through activation of Nrf2 via an Autophagy-Dependent pathway. *Antioxid Redox Signal*. 2024;41(16–18):1117–33.
161. Shin K, Park M, Kim S, et al. Novel anti-CD73-IL-2v bispecific fusion protein augments antitumor immunity by alleviating immunosuppressive adenosine pathways in CD8+T cells. *J Immunother Cancer*. 2025;13(3):e008594.
162. Schäkel L, Mirza S, Winzer R, et al. Protein kinase inhibitor ceritinib blocks ectonucleotidase CD39 – a promising target for cancer immunotherapy. *J Immunother Cancer*. 2022;10(8):e004660.
163. Liao J, Zeng DN, Li JZ, et al. Targeting adenosinergic pathway enhances the anti-tumor efficacy of Sorafenib in hepatocellular carcinoma. *Hepatol Int*. 2020;14(1):80–95.
164. Sánchez-Melgar A, Vultaggio-Poma V, Falzoni S, et al. Mitochondrial localization and function of adenosine receptors. *Int J Biol Sci*. 2025;21(5):1874–93.
165. Sun X, Li Z, Wang X, et al. Inorganic phosphate as bioenergetic messenger triggers M2-Type macrophage polarization. *Adv Sci (Weinh)*. 2024;11(13):e2306062.
166. Tay AHM, Prieto-Díaz R, Neo S, et al. A2B adenosine receptor antagonists rescue lymphocyte activity in adenosine-producing patient-derived cancer models. *J Immunother Cancer*. 2022;10(5):e004592.
167. Yu M, Guo G, Huang L, et al. CD73 on cancer-associated fibroblasts enhanced by the A2B-mediated feedforward circuit enforces an immune checkpoint. *Nat Commun*. 2020;11(1):515.
168. Angioni R, Liboni C, Herkenne S, et al. CD73 + extracellular vesicles inhibit angiogenesis through adenosine A2B receptor signalling. *J Extracell Vesicles*. 2020;9(1):1757900.
169. Chen Y, Zhang J, Weng Y, et al. Cryo-EM structure of the human adenosine A2B receptor-Gs signaling complex. *Sci Adv*. 2022;8(51):eadd3709.
170. Li Z, Kou L, Fu X, et al. Design, synthesis, and biological evaluation of triazole-pyrimidine-methylbenzonitrile derivatives as dual A2A/A2B adenosine receptor antagonists. *J Enzyme Inhib Med Chem*. 2022;37(1):1514–26.
171. Oshima HS, Ogawa A, Sano FK, et al. Structural insights into the agonist selectivity of the adenosine A3 receptor. *Nat Commun*. 2024;15(1):9294.
172. Storme J, Cannaert A, Van Craenenbroeck K, et al. Molecular dissection of the human A3 adenosine receptor coupling with  $\beta$ -arrestin2. *Biochem Pharmacol*. 2018;148:298–307.
173. Cai H, Guo S, Xu Y, et al. Cryo-EM structures of adenosine receptor A3AR bound to selective agonists. *Nat Commun*. 2024;15(1):3252.
174. Kotulova J, Lonova K, Kubickova A, et al. 2-Cl-IB-MECA regulates the proliferative and drug resistance pathways, and facilitates chemosensitivity in pancreatic and liver cancer cell lines. *Int J Mol Med*. 2022;49(3):31.
175. Kim G, Hou X, Byun WS, et al. Structure-Activity relationship of truncated 2,8-Disubstituted-Adenosine derivatives as dual A2A/A3 adenosine receptor antagonists and their Cancer immunotherapeutic activity. *J Med Chem*. 2023;66(17):12249–65.
176. Park JS, Ma YQ, Wang F, et al. A3AR antagonism mitigates metabolic dysfunction-associated steatotic liver disease by exploiting monocyte-derived Kupffer cell necroptosis and inflammation resolution. *Metabolism*. 2025;164:156114.
177. Safadi R, Braun M, Francis A, et al. Randomised clinical trial: A phase 2 double-blind study of Namodenoson in non-alcoholic fatty liver disease and steatohepatitis. *Aliment Pharmacol Ther*. 2021;54(11–12):1405–15.
178. Marucci G, Santinelli C, Buccioni M, et al. Anticancer activity study of A3 adenosine receptor agonists. *Life Sci*. 2018;205:155–63.
179. Falloot LB, Suresh RR, Fisher CL, et al. Structure-Activity studies of 1H-Imidazo[4,5-c]quinolin-4-amine derivatives as A3 adenosine receptor positive allosteric modulators. *J Med Chem*. 2022;65(22):15238–62.
180. Suresh RR, Jain S, Chen Z, et al. Design and in vivo activity of A3 adenosine receptor agonist prodrugs. *Purinergic Signal*. 2020;16(3):367–77.

181. Fisher CL, Pavan M, Salmaso V, et al. Extrahelical binding site for a 1H-Imidazo[4,5-c]quinolin-4-amine A3 adenosine receptor positive allosteric modulator on Helix 8 and distal portions of transmembrane domains 1 and 7. *Mol Pharmacol*. 2024;105(3):213–23.

**Publisher's note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.