



Emerging Functional Imaging Biomarkers of Tumour Responses to Radiotherapy

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Abstract: Tumour responses to radiotherapy are currently primarily assessed by changes in size. Imaging permits non-invasive, whole-body assessment of tumour burden and guides treatment options for most tumours. However, in most tumours, changes in size are slow to manifest and can sometimes be difficult to interpret or misleading, potentially leading to prolonged durations of ineffective treatment and delays in changing therapy. Functional imaging techniques that monitor biological processes have the potential to detect tumour responses to treatment earlier and refine treatment options based on tumour biology rather than solely on size and staging. By considering the biological effects of radiotherapy, this review focusses on emerging functional imaging techniques with the potential to augment morphological imaging and serve as biomarkers of early response to radiotherapy.

Keywords: radiotherapy; functional imaging; magnetic resonance imaging; positron emission tomography; hyperpolarised ¹³C; diffusion-weighted imaging; treatment response

1. Introduction

The introduction of cross-sectional anatomical imaging using computed tomography (CT) and magnetic resonance imaging (MRI) in the 1970s revolutionised clinical oncology by permitting non-invasive determination of tumour burden that is essential for diagnosis and staging and assessing treatment responses. To standardise characterisation of tumour responses that were frequently incomplete and heterogeneous and for comparison between clinical trials the World Health Organisation defined the first response criteria in 1979 [1]. Although subsequently refined the criteria used today, most commonly in the form of the Response Evaluation Criteria in Solid Tumours (RECIST) [2,3], are recognisably similar (Table 1). Measurements of tumour burden are often an excellent determinant of disease progression or response but changes usually manifest themselves slowly and can sometimes be misleading [4]. In the forty years since the introduction of size response criteria, functional imaging techniques have emerged that are capable of reporting many aspects of tumour biology. In response to therapy, biochemical changes precede anatomical changes, sometimes by many months [5]. Earlier determination of response to treatment would facilitate modification of treatment before significant disease progression and reduce the physical, psychological and financial costs of ineffective or unnecessary therapy. Of the fourteen million people diagnosed with cancer worldwide every year, more than half receive radiation therapy [6]. Identification of radio-resistant tumour regions at a pre- or early therapy stage could be used for localised or global dose escalation or initiation of concomitant chemotherapy. Following treatment, functional imaging is a



potentially powerful tool where the conclusions of morphological imaging can be limited, for example, differentiation of radiation necrosis and residual disease in the brain.

	WHO	RECIST 1.0	RECIST 1.1
Measurement	Sum of maximal perpendicular diameters of all measured lesions	Sum of long axis of up to 10 target lesions	Sum of long axis of up to 5 target lesions (short axis for lymph nodes)
Complete response	Disappearance of all known disease	Disappearance of all target lesions	Disappearance of all target lesions
Partial response	\geq 50% decrease in lesion size and no new lesions	≥30% decrease in sum of target lesion diameters	≥30% decrease in sum of target lesion diameters
No change/ Stable disease	Neither partial response or progressive disease	Neither partial response or progressive disease	Neither partial response or progressive disease
Progressive disease	\geq 25% increase in lesion size or \geq 1 new lesion	≥20% increase in the sum of target lesions (no minimum size increase)	≥20% increase in the sum of target lesions (≥5 mm absolute increase)
Functional imaging	None	None	¹⁸ F-FDG-PET can be used to complement CT

Table 1. Comparison between the most frequently used imaging response criteria.

WHO, World Health Organisation; RECIST, Response evaluation criteria in solid tumours; CT, computed tomography; FDG-PET, positron emission tomography with 2-¹⁸F-fluoro-2-deoxy-D-glucose.

In this review we will focus on emerging functional imaging techniques that exploit the biological changes in tumours following radiation therapy and have the potential to improve the early detection of treatment response. Many of the functional imaging techniques discussed can also be applied to delivering intensity-modulated or stereotactic body radiation therapy using increasingly sophisticated methods, a subject that is beyond the scope of this review but has been extensively reviewed elsewhere [7,8].

2. Biological Effects of Radiation

Ionising radiation refers to particles that have sufficient energy to release electrons from an atom. The most significant biological target of ionising radiation is DNA that can be ionised directly or indirectly by free radicals (e.g., hydroxyl ($^{\circ}$ OH), superoxide (O_2^{-}) and hydrogen peroxide (H_2O_2)) produced by ionisation of adjacent molecules [9,10]. DNA ionisation can result in damage to any part of the molecule. Base damage and single-stranded breaks occur frequently but efficient repair mechanisms limit the biological effect. In contrast, double-stranded breaks and DNA crosslinking are less frequent events but are also less likely to be repaired effectively resulting in either genomic mutation or repair failure and subsequent cell death. Radiation-induced cell death can result from activation of cellular senescence or apoptosis, the latter predominantly via the intrinsic pathway [11]. However, particularly in tumour cells where cell cycle checkpoint controls, DNA repair and apoptotic pathways are frequently perturbed, cell death predominantly occurs from mitotic catastrophe, a result of premature induction of mitosis before S and G₂ phase completion that ultimately results in cell necrosis [9,12].

Radiobiological effects are dependent on external factors such as the dose and type of radiation used. For example, protons and alpha particles have a high linear energy transfer and are more likely to induce complex DNA damage with a higher probability of lethality [13]. Additionally, biological variables result in heterogeneous radiation sensitivity between and within tissues. Hypoxia and low rates of proliferation tend to promote radio-resistance and cancer stem cells may be more resistant than the bulk of tumour cells [14].

The effects of radiation on tumours are not limited to cell death with virtually every aspect of the tumour microenvironment responding to the insult. In the acute phase necrosis and vascular disruption leads to hypoperfusion, oedema and an inflammatory response that begins in the first few hours following acute radiation injury. Chronic activation of the inflammatory response results in dysregulated tissue remodelling characterised by decreased vascularity and fibrosis [14].

3. Imaging Apoptosis and Necrosis

Accurate determination of cell death would find application in a wide range of conditions including stroke, myocardial infarction and cancer. Several probes have been designed to assess biochemical events that occur during cell death. Phosphatidylserine, an anionic phospholipid and a major component of the inner leaflet of the cell membrane, is externalised by stressed or dying cells and is a target for phagocytosis [15,16]. Annexin-V binds to externalised phosphatidylserine with low nanomolar affinity and has been radiolabelled with ¹⁸F and ^{99m}Tc for positron emission tomography (PET) and single photon emission computed tomography (SPECT) imaging, respectively [17]. ^{99m}Tc-annexin-V has progressed to clinical trials, where increases in labelling of 20–30% in the first 72 h following chemotherapy or radiotherapy were associated with treatment response in lung, breast, lymphoma and head and neck cancers [18]. Unfortunately, annexin-V is limited by slow pharmacokinetics and high levels of non-specific binding, particularly to the abdominal organs [17]. An alternative is using the C2A domain of synaptotagmin-I which binds to anionic phospholipids and has been labelled with ^{99m}Tc and ¹¹¹In for SPECT [19,20], ¹⁸F for PET [21] and gadolinium chelates for magnetic resonance imaging (MRI) in preclinical in vivo studies [22]. To improve the biodistribution, simplify labelling and improve pharmacokinetics (which are often slow when using peptide-based tracers), a smaller modified C2A protein (C2Am) has been labelled with ^{99m}Tc for SPECT (Figure 1) [20] and a near-infrared fluorophore for multispectral optoacoustic tomography (MSOT) [23], demonstrating high sensitivity and specificity for cell death. Tumour uptake of ^{99m}Tc-duramycin which binds to phosphatidylethanolamine, another phospholipid externalised during cell death, has demonstrated improved sensitivity in detecting early treatment response compared to ¹⁸F-FDG in preclinical studies [24].

Several small-molecule imaging probes that detect cell or mitochondrial membrane depolarisation and/or acidification of apoptotic cells have also been developed. In patients with intracranial tumours, the change in ¹⁸F-ML-10 uptake from before to 48 h after CyberKnife stereotactic radiotherapy correlated with the decrease in tumour volume measured at 2–4 months after treatment [25]. Similar correlations have been made in patients with brain metastases imaged before and nine days after whole-brain radiotherapy [26].

Cell membrane changes are not specific to apoptosis and increased binding and uptake are also seen in autophagy, necroptosis and necrosis. Several PET radiotracers have been designed to detect cleaved caspase 3 and 7, components of the final common pathway of apoptosis that have greater specificity for apoptosis. Of these, ¹⁸F-ICMT-11 has recently been used in breast and lung cancer patients, although low tumour uptake explained by low cleaved caspase 3 expression before and after treatment limited the conclusions [27].



Figure 1. Single photon emission computed tomography (SPECT) imaging of cell death in vivo in $E\mu$ -*Myc* tumours. Imaging of (^{99m}Tc)-labelled C2Am at 2 h after probe administration and 24 h after drug treatment. SPECT/CT of representative $E\mu$ -*Myc* mice before (BT, left) and after (AT, right) cyclophosphamide treatment. Top—maximum intensity projections (MIPs); middle—axial section through the cervical tumour; bottom—axial section through the axillary and mediastinal tumours. Tumours are indicated with arrowheads. Reproduced with permission from Neves et al. © SNMMI [20].

4. Imaging Changes in Vasculature

Radiation therapy results in acute endothelial cell dysfunction, apoptosis and disruption of blood vessels. Above doses of 8–10 Gy endothelial cell apoptosis is induced by activation of the acid sphingomyelinase (ASMase)/ceramide signalling pathway [28–30]. Therefore, activation of this pathway does not occur with the lower doses delivered in fractionated radiotherapy, only with the higher single doses delivered with stereotactic radiotherapy [31]. Capillaries increase in permeability and become thrombosed due to platelet aggregation and microthrombus formation with subsequent hypoperfusion causing further tumour necrosis [12,32]. This suggests that imaging changes in perfusion have potential for early detection of tumour responses to radiotherapy.

4.1. Dynamic Contrast-Enhanced (DCE) CT

DCE-CT following an intravenous bolus of iodinated contrast agent is a highly reproducible imaging technique that permits relatively simple absolute quantification of blood flow, blood volume, permeability–surface area product, mean transit time and extravascular volume [33]. Correlation of DCE-CT metrics with histological determination of microvessel density and vascular endothelial growth factor (VEGF) expression has been possible in some studies [34,35]. Reductions in blood flow, blood volume, mean transit time and permeability–surface area product have been demonstrated in patients with rectal cancer, head and neck cancer and brain metastases following radiotherapy \pm chemotherapy [35–40]. However, in patients with cervical cancer increases in tumour blood volume were observed three weeks into chemoradiotherapy which were predictive of complete metabolic response at three months [41]. The conflicting findings may reflect heterogeneity between tumour types and responses to treatment but also differences in timing of the post-treatment study.

4.2. Perfusion MRI

Following injection, paramagnetic contrast agents (typically low-molecular-weight gadolinium (Gd^{3+}) chelates) are distributed via the blood and diffuse freely into the interstitial space but do not cross the cell membrane. Paramagnetic contrast agents cause magnetic field inhomogeneities that reduce the T_1 , T_2 and T_2^* relaxation times of nearby protons resulting in temporal changes in MR signal intensity and can provide information on the concentration of the injected contrast agent, microvessel density, perfusion and vessel permeability [42–44]. The most commonly used techniques are DCE-MRI and dynamic susceptibility contrast MRI, which exploit the T_1 and T_2^* effects of paramagnetic contrast agents, respectively. In addition to subjective visual analysis of the rate, total amount and decrease (washout) of contrast enhancement in lesions, semi-quantitative and quantitative parameters can be derived similar to those of DCE-CT, although the post-processing is complicated by a nonlinear relationship between contrast agent concentration and change in signal intensity [33]. The use of an exogenous contrast agent can be avoided by using arterial spin labelling (ASL) in which blood water protons are magnetically labelled. This suffers from low temporal and spatial resolution and low signal-to-noise ratio but has greatly reduced post-processing requirements when compared to imaging exogenous contrast agents [45].

In patients with cervical cancer high contrast enhancement before and in the first few weeks after the initiation of chemoradiotherapy is a better predictor of response than tumour volume measurements [46–48]. Other semi-quantitative and quantitative measures, particularly higher rates of K^{trans} (the volume transfer constant between plasma and the extravascular extracellular space) and plasma flow, have also been shown to be predictive of response [47,49,50]. Similar results have been obtained for rectal cancer and head and neck cancer where a high K^{trans} before chemoradiotherapy and a large decrease or low K^{trans} after therapy are generally associated with good response [51,52]. In high-grade gliomas and cerebral metastases, reductions in K^{trans} and changes in tumour blood volume and flow (from DSC-MRI and ASL) have detected response to stereotactic radiosurgery or whole-brain irradiation as early as one week after treatment [53–55]. Correlations between high

perfusion on DCE-MRI and radio-sensitivity have frequently been attributed to decreased hypoxia in well-perfused tumours [46]. However, it has also been reported that higher microvessel density and increased angiogenesis correlates with greater metastatic potential and poorer outcome [56].

In addition to prognostication, another potential application of perfusion MRI is the differentiation between radiation necrosis and recurrence in high-grade glioma, which often appear similar using conventional contrast-enhanced MRI [57]. A meta-analysis concluded that sensitivity and specificity for tumour recurrence was 90% and 88%, respectively, using DSC-MRI and 89% and 85% with DCE-MRI [58]. Initial studies using ASL have also demonstrated its ability to differentiate disease recurrence from radiation necrosis with a high degree of accuracy (Figure 2) [59–62].



Figure 2. Perfusion magnetic resonance imaging (MRI) with pulsed arterial spin labelling (pASL) of a patient with a frontal high-grade oligodendroglioma with a focus of hyperperfusion (white arrow) identifying recurrent disease post-surgery and chemotherapy. Axial sections have the following sequences: (**a**) T_1 -weighted contrast-enhanced and (**b**) pASL. Images courtesy of Dr Harpreet Hyare, University College London Hospital.

4.3. Ultrasound and Optical Imaging

Radiation-induced changes in vasculature can also be imaged using dynamic contrast-enhanced ultrasound (CEUS). Low solubility, gas-containing microbubbles have different acoustic properties and can be used to image microvascular density and perfusion. This has been used to predict response of a number of different cancer types to chemotherapy [63–65], while decreased vascular density following radiation therapy has been used as an early marker of response in preclinical tumour models [66,67]. Furthermore, using antibodies conjugated to the surface of microbubbles could permit tumour targeting. Microbubbles targeting the angiogenesis regulators $\alpha_v\beta_3$ integrin and ICAM-1 that are upregulated in response to radiotherapy were increased in a rat prostate tumour model so treated [68].

Optical coherence tomography (OCT) is a non-invasive imaging technique that can produce 3D in vivo images at a resolution of a few micrometres by measuring the interference pattern of back-scattered light [69]. Although OCT has unrivalled spatial resolution, the scattering of light within biological tissues limits the imaging depth to a few millimetres. OCT is well established for high-resolution 3D retinal imaging and, more recently, functional imaging of the microvasculature of tumours has been demonstrated using speckle variance OCT [70]. In pancreatic human tumour xenografts irradiated with \geq 10 Gy, the vascular volume density decreased by 26% just 30 min post-radiotherapy. Early changes were predominantly seen in small vessels <30 µm in diameter

and were transient, potentially indicating rapid microthrombus formation following radiotherapy [71]. Maximal reductions in vascular volume density were seen after 2–4 weeks, depending on delivered dose, and preceded reductions in tumour volume by several weeks [70].

MSOT uses an ultrasound transducer to measure acoustic waves generated in response to localised thermoelastic expansion of tissue induced by pulses of laser light [72]. In addition to detection of exogenous contrast agents (see Section 3), endogenous biomarkers of perfusion and hypoxia can be derived due to the different light absorption spectra of oxy- and deoxyhaemoglobin [73]. In patient-derived head and neck squamous cell carcinoma (HNSCC) xenografts early changes in haemoglobin oxygen saturation following radiotherapy correlated with subsequent changes in tumour volume [74]. Although the MSOT spatial resolution of around 500 μ m is inferior to OCT a tissue depth of up to 7 cm is possible [75]. Despite depth limitation both techniques have great potential for non-invasive and endoscopic imaging of a wide range of tumours.

4.4. PET Imaging of Perfusion

Several PET tracers have been developed to measure perfusion, of which ¹⁵O-H₂O has been the most extensively used. ¹⁵O-H₂O is an inert PET tracer that freely diffuses across cell membranes and allows absolute quantification of tumour blood flow with a reproducibility comparable to other imaging modalities [76,77]. High tumour blood flow on ¹⁵O-H₂O PET before treatment was predictive of poor response to radiotherapy in head and neck cancer [78]. Unfortunately, the short half-life (2 min) of ¹⁵O that requires an onsite cyclotron has limited the widespread use of the technique.

5. Hypoxia Imaging

Hypoxia is an important biological determinant of radio-sensitivity and is well characterised having first been recognised in the early part of the 20th century [79]. Dysregulated tumour proliferation and angiogenesis, the latter resulting in the formation of structurally and functionally abnormal neovasculature, combine to increase the distance between cells and a sufficient blood supply resulting in chronic hypoxia and nutrient depletion. The abnormal vasculature is also prone to transient occlusion and hypoperfusion, causing acute, fluctuating hypoxia. Both sources of hypoxia contribute to radio-resistance and the transcriptional regulation of many genes associated with tumour growth and survival, notably hypoxia inducible factor 1 (HIF-1) [80]. This has led to great efforts to minimise tumour hypoxia particularly prior to radiotherapy, with variable levels of clinical success [79]. Imaging modalities that are sensitive to hypoxia could be prognostic and potentially improve outcomes by permitting dose and treatment modification or dose painting.

5.1. PET Imaging of Hypoxia

¹⁸F-labelled 2-nitroimidazole-based markers have been widely used for PET imaging of hypoxia. In an anoxic environment reduction of the NO₂ moiety of 2-nitroimidazole by nitroreductases produces highly reactive intermediates which bind to many macromolecules and also undergo glutathione conjugation [81]. ¹⁸F-fluoromisonidazole (¹⁸F-FMISO) was the first PET tracer for hypoxic imaging to be developed and has subsequently been the most extensively used, with accumulation having been demonstrated in human glioma, head and neck squamous cell carcinomas (HNSCC), breast, lung and renal tumours. In HNSCC patients, high baseline and ongoing ¹⁸F-FMISO uptake in the first two weeks of uptake was significantly associated with loco-regional recurrence and was used as a rationale for radiation dose escalation [82,83]. The feasibility of dose painting based on hypoxic and nonhypoxic tumour subvolumes to improve local tumour control has also been demonstrated [84]. Similarly, in locally advanced non-small cell lung cancer, ¹⁸F-FMISO uptake on baseline scans is strongly associated with poor prognosis. Although dose escalation was possible in this study without excessive toxicity it was not shown to improve outcome [85].

A major limitation of ¹⁸F-FMISO is slow clearance due to its lipophilicity and, to achieve acceptable signal-to-background ratios, static imaging is performed at delayed timepoints

(typically up to four hours post-injection). ¹⁸F-fluoroazomycinarabinoside (¹⁸F-FAZA) and 3^{-18} F-fluoro-2-(4-((2- nitro-1*H*-imidazol-1-yl)methyl)-1*H*-1,2,3,-triazol-1-yl)-propan-1-ol (¹⁸F-HX4) are hydrophilic molecules and thus have improved pharmacokinetics for PET imaging [86]. The prognostic value and feasibility of dose escalation guided by imaging with ¹⁸F-FAZA has been shown in HNSCC and NSCLC patients [87–91]. In sarcomas, hypoxia identification with ¹⁸F-FAZA was associated with radio-resistance and recurrence. However, toxicity limited attempts to combine radiotherapy with sunitinib [92]. An alternative PET tracer for hypoxia imaging is Cu(II)-diacetyl-*bis*(N4-methylthiosemicarbazone (Cu-ATSM) which is believed to be reduced from Cu²⁺ to Cu¹⁺ and trapped in hypoxic cells [93]. As a predictive marker, Cu-ATSM has shown potential in cervical, rectal, lung and head and neck cancer [93–97].

The inherent sensitivity of PET imaging makes it an attractive modality for hypoxia imaging where detection of relatively small changes in oxygen concentration is required and the initial studies as a prognostic marker have been mostly positive. However, the resolution of clinical PET images is typically around 5 mm, which may lead to a lack of sensitivity when subvoxel hypoxia variation exists and is a potential barrier to dose painting based on hypoxia imaging [93]. It should also be recognised that no imaging modality is sensitive to hypoxia alone and even specific PET tracer uptake is dependent to a certain degree on perfusion, cellularity and other biological variables.

5.2. MRI Imaging of Hypoxia

Hypoxia imaging is also possible with MRI because dissolved oxygen and deoxyhaemoglobin are paramagnetic and decrease T_1 and T_2^* relaxation. Methods that exploit the effects of these molecules on T_1 relaxation are termed oxygen-enhanced (OE) or tumour oxygenation level-dependent (TOLD) MRI, while imaging of T_2^* effects is termed blood oxygenation level-dependent (BOLD) MRI [98]. In a typical study, baseline imaging is performed with the patient breathing room air followed by imaging while the patient inhales oxygen to create arterial hyperoxia with the difference in signal between the two images corresponding to the effect of oxygen inhalation. These assays have been shown to correlate with tumour pO₂ [98–100] and several studies have used the techniques to detect or predict response to radiotherapy. In animal prolactinoma and fibrosarcoma tumour models, BOLD was able to predict growth response after a single radiation dose [101] and the technique has now entered early clinical trials in head and neck cancer patients [102]. OE-MRI is still in preclinical development. Nevertheless, in rats with subcutaneous prostate tumours improved oxygenation of tumours after radiotherapy correlated with response and OE-MRI measurements offered better prognostication than BOLD [103]. Additionally, OE-MRI has also been used in mouse models to differentiate radiation necrosis from glioma [104].

6. Imaging Changes in Tissue Structure

6.1. Diffusion-Weighted MRI

Diffusion-weighted imaging (DWI) is an MRI technique that measures the random, or Brownian, movement of water molecules in tissues that can be quantified. The simplest and most commonly used quantifiable metric used in DWI is the apparent diffusion coefficient (ADC) but more complex models such as VERDICT (vascular, extracellular and restricted diffusion for cytometry in tumours) can extract additional data related to cell size, vascular, intra- and extracellular volume fractions and perfusion effects, which may lead to improved detection of early treatment response [105]. Additionally, numerous imaging techniques have evolved from DWI. For example, diffusion tensor imaging (DTI) and diffusion kurtosis imaging (DKI) can provide information on diffusion directionality and tissue microstructure, respectively [106,107]. Recently, filter-exchange imaging (FEXI) has been used to determine the exchange rate of water across the cell membrane [108,109].

In tissues, Brownian motion of water is limited by membranes and macromolecules, giving a lower ADC value (indicating restricted diffusion) for intracellular water than extracellular water [110].

ADC has been shown to have a strong inverse correlation with tumour cellularity in glioma, lung and ovarian tumours, but the relationship is less significant for other tumour types [111]. Following radiotherapy, ADC can transiently decrease due to cellular swelling before increasing due to cell death, the latter being associated with decreasing cellularity and a response to treatment in most studies. At later stages, reductions in ADC can occur due to inflammation and fibrosis [112,113]. Unfortunately, many of these processes coexist, resulting in conflicting effects on diffusion imaging and potentially limiting the early predictive value of the technique. Few studies have looked at the time of assessment with DWI, but in the longitudinal assessment of brain metastases from a range of primary sites treated with whole-brain external beam radiotherapy, the optimal timepoint for prediction of response was after seven fractions on day seven to nine [114]. Similarly, in HNSCC patients an increase in ADC one week after radiotherapy was predictive of response with a sensitivity of 86% and specificity of 83% [115] However, in cervical cancer, although diffusion imaging could detect treatment response upon completion of chemoradiotherapy, reimaging performed in the first two weeks of treatment was unable to differentiate complete, partial and non-responders [116]. In rectal cancer, although DWI alone is not sufficiently accurate for prediction of early response following chemoradiotherapy [117], a combination of DWI, ¹⁸F-FDG-PET/CT and T_2 -weighted volumetry permitted early response prediction with a sensitivity and specificity of 75% and 94%, respectively [118]. In primary glioblastoma, DWI can help to differentiate progression from pseudoprogression (Figure 3) [119]. Recently a combination of twelve multi-parametric imaging features (including ADC) differentiated pseudoprogression from progression with a sensitivity and specificity of 71% and 90%, respectively, versus 100% and 20% for ADC alone [120]. Furthermore, a positive correlation has been reported between necrosis and ADC following treatment and, in most studies, tumour recurrence has generally been found to have a lower ADC than radiation necrosis [121–124].

6.2. Chemical Exchange Saturation Transfer MRI

The MRI techniques discussed so far all image the protons (¹H) of water molecules, the abundance of which in biological tissues (60–80 M) facilitates imaging at high temporal and spatial resolution. In the presence of a magnetic field, nuclei with spin (e.g., ¹H and ¹³C) resonate at a frequency that is partly dependent on the electronic environment of the molecule they are part of, for example, amide protons resonate at a different frequency to water protons. This phenomenon is known as chemical shift and means that MR spectroscopy (MRS) can non-invasively detect the presence and relative concentration of multiple metabolites in vivo [125]. However, the low concentration of these metabolites makes MRS a technique with low temporal and spatial resolution. Chemical exchange saturation transfer (CEST) MRI, described in detail elsewhere [126], is a technique that allows the indirect detection of molecules containing exchangeable protons via attenuation of the water signal. This indirect detection offers greatly enhanced sensitivity, facilitating high-resolution imaging. Amide proton transfer (APT), a CEST technique that detects exchangeable amide protons present in mobile peptides and proteins, has been used to detect the higher concentration of proteins present in tumours corresponding to a higher APT signal than surrounding normal tissue (Figure 4) [127]. In neuro-oncology the technique has promise for the differentiation of progression and radiation necrosis in particular and has already been demonstrated in a study of patients with brain metastases [128].



Figure 3. Diffusion-weighted MRI of a patient with glioblastoma multiforme. The necrotic core shows facilitated diffusion with a surrounding ring of restricted diffusion indicative of progressive disease. Axial sections have the following sequences: (a) T_1 -weighted contrast-enhanced; (b) T_2 FLAIR (fluid-attenuated inversion recovery); (c) diffusion-weighted imaging (DWI); (d) apparent diffusion coefficient (ADC) map. Images courtesy of Dr Harpreet Hyare, University College London Hospital.



Figure 4. Detection of recurrence in two patients using ¹⁸F-choline and amide proton transferchemical exchange saturation transfer (APT-CEST). Patient 1 (**a**–**d**): WHO grade I pilocytic astrocytoma initially treated with radiotherapy. Imaging performed four years later when the patient re-presented with seizures. Patient 2 (**e**–**h**): WHO grade IV glioblastoma multiforme treated with surgery and chemoradiotherapy. Imaging surveillance performed eighteen months after completing initial treatment. Tumours indicated with white arrowheads. Abbreviations: T_2 W FLAIR, T_2 -weighted fluid-attenuated inversion recovery; T_1 W CE, T_1 -weighted contrast enhanced. Images courtesy of Dr Harpreet Hyare, University College London Hospital.

7. Imaging Changes in Metabolism

Aberrant nutrient uptake and subsequent metabolism is a feature of malignant tumours that results from the increased demand for the synthesis of proteins, nucleic acids, fatty acids and other macromolecules required for increased growth and proliferation. Aerobic glycolysis, whereby glucose is reduced to lactate even when oxygen is abundant, was first described by Otto Warburg nearly a century ago and has subsequently been observed in many malignant tumours [129,130]. Following treatment, a decrease in tumour metabolic activity precedes changes in structure and volume, making metabolic imaging attractive for detecting early treatment response [125].

7.1. Imaging Changes in Glycolysis and TCA Cycle Metabolism

2-(¹⁸F-fluoro)-2-deoxy-D-glucose (¹⁸F-FDG) is a glucose analogue that is transported into cells and phosphorylated, trapping the tracer intracellularly and allowing identification of glucose-avid tissues upon subsequent PET imaging. ¹⁸F-FDG-PET is the most commonly used PET tracer and is used as an adjunct to morphological imaging in the follow-up of many tumours following treatment (Figure 5). Its widespread use and standardisation of acquisition has meant that ¹⁸F-FDG is currently the only functional imaging technique to be (semi)quantified for use in response evaluation criteria, most notably in PERCIST 1.0 and the EORTC guidelines [131,132]. In HNSCC, the negative predictive value for primary and nodal disease with ¹⁸F-FDG-PET/CT was 99–100% four months after chemoradiotherapy [133,134]. ¹⁸F-FDG is also useful in several cancers, for example, nonsmall cell lung cancer, for differentiating recurrence from radiation necrosis [135]. However, attempts to shorten the interval scanning time have produced mixed results [136], with a lack of early response often attributed to inflammation and macrophage infiltration, although the evidence for this mechanistically is limited [137].



Figure 5. A T4N0M0 rectal adenocarcinoma treated with neoadjuvant chemoradiotherapy. (**b** and **c**) Prior to treatment, the lesion is ¹⁸F-FDG-avid; (**g** and **h**) after chemoradiation, there is minimal residual uptake despite a minimal change in tumour volume. At subsequent resection, no residual disease remained and the patient was down-staged to ypT0N0M0. (**a** and **f**) axial CT; (**b** and **g**) axial ¹⁸F-FDG-PET; (**c** and **h**) PET/CT fusion; (**d** and **i**) axial T_2 W MRI; (**e** and **j**) coronal T_2 W MRI. The tumour is indicated by arrowheads and dashed outline. All ¹⁸F-FDG-PET images are scaled between an standardised uptake value (SUV) of 0–5.

As discussed earlier, the low concentration of biological metabolites and low sensitivity of NMR limits the temporal and spatial resolution of MRS in vivo. The method of dynamic nuclear polarisation (DNP) of ¹³C-labelled substrates is a technique that can increase the signal-to-noise ratio of ¹³C MR spectroscopy and imaging by >10⁴ in vivo [138]. Hyperpolarised (1-¹³C)pyruvate has been the most widely used substrate due to its high polarisation levels, long polarisation lifetime and its position in the glycolytic pathway. Following injection, hyperpolarised (1-¹³C)pyruvate enters cells via monocarboxylate transporters and, in tumours, is predominantly reduced to lactate by lactate dehydrogenase [139]. Compared to ¹⁸F-FDG, hyperpolarised (1-¹³C)pyruvate has improved specificity for indicating the Warburg effect and may better differentiate inflammation from tumour progression/recurrence [140]. A reduction of (1-¹³C)lactate production following hyperpolarised (1-¹³C)pyruvate injection after antiandrogen therapy has been observed in a prostate cancer patient [141]. In an orthotopic rat glioma model, a reduction in label flux from (1-¹³C)pyruvate to (1-¹³C)lactate was seen in all animals in the first 96 h after radiotherapy despite increases in tumour size, suggesting that (1-¹³C)pyruvate may be useful to differentiate progression and pseudoprogression (Figure 6) [142].

The other readily translatable hyperpolarised substrate is $(1,4-{}^{13}C_2)$ fumarate, which is hydrated to malate by fumarase. During cell death an increase in membrane permeability results in leakage of fumarase into the extracellular space and an increased rate of malate production following hyperpolarised $(1,4-{}^{13}C_2)$ fumarate injection which, in preclinical studies, has been shown to be a sensitive indicator of cell death [143,144].



Figure 6. Representative images following injection of hyperpolarised $(1^{-13}C)$ pyruvate in a C6 glioma-bearing rat before (top, PreTx) and 96 h after radiotherapy (bottom, PostTx). Images of $(1^{-13}C)$ pyruvate $((1^{-13}C)$ pyr) and $(1^{-13}C)$ lactate $((1^{-13}C)$ lac) overlaid on coronal T_1W MR images. A reduction in $(1^{-13}C)$ lactate production was observed in all tumours after 15 Gy irradiation without decreases in tumour volume. Adapted with permission from Day et al. [142].

7.2. Imaging Proliferation

Several PET tracers have been designed as biomarkers of proliferation. 3'-deoxy-3'-¹⁸F-fluorothymidine (¹⁸F-FLT) is taken into cells and phosphorylated by thymidine kinase, the first step of the thymidine salvage pathway essential for DNA synthesis. Thus, ¹⁸F-FLT preferentially accumulates in cells undergoing proliferation with potentially greater tumour specificity than ¹⁸F-FDG. Several systematic reviews have concluded that (¹⁸F-FLT has potential as a marker of early response and shown that a change in uptake correlated well with progression-free and disease-free survival [145,146]. In HNSCC, a comparison of ¹⁸F-FLT and ¹⁸F-FDG during radiotherapy showed the overall accuracy of ¹⁸F-FLT to be significantly higher (74 vs. 30%) [147]. However, following chemoradiotherapy in rectal cancer, despite correlations with disease-free survival, decreases in ¹⁸F-FLT uptake did not correlate with pathological response, a discrepancy attributed to changes in perfusion following radiotherapy [148].

7.3. PET Imaging of Brain Tumours

Lack of tumour specificity of ¹⁸F-FDG is a particular problem in neuroradiology where there is high background uptake from normal brain tissue and radiation necrosis is often hypermetabolic [149]. Numerous tracers have been designed that are superior to ¹⁸F-FDG for detection of recurrence and differentiation from pseudoprogression. Brain tumours often have increased uptake of amino acids relative to normal brain and several have been labelled with ¹¹C and ¹⁸F for PET imaging [150]. ¹¹C-methionine has been the most widely used amino acid PET tracer. It can differentiate tumours from normal brain with an accuracy of 94% versus 80% for ¹⁸F-FDG [151] and is also more sensitive for differentiating recurrence from radiation necrosis following radiotherapy [152]. However, the application of ¹¹C-labelled substrates will always be limited by the short half-life (20 min) requiring onsite production of the tracer. Therefore, several alternatives have been developed including ¹⁸F-fluoro-ethyl-tyrosine, an artificial amino acid that is not incorporated into proteins but has increased uptake into tumours [153,154]. Decreased uptake in the first 10 d following

chemoradiotherapy was predictive of progression-free survival with an accuracy of 75% [155]. Other tracers that have demonstrated improved performance over ¹⁸F-FDG for distinguishing recurrence from radiation necrosis include ¹¹C- and ¹⁸F-labelled choline (Figure 4), surrogate measures of the rate of phospholipid membrane synthesis, and ¹⁸F-dihydroxyphenylalanine (¹⁸F-DOPA), an analog of the dopamine precursor L-DOPA (Figure 7) [156–158].



Figure 7. ¹⁸F-dihydroxyphenylalanine (¹⁸F-DOPA) uptake in a patient with glioblastoma multiforme treated with chemoradiotherapy before (top row) and after 12 weeks (bottom row). From left to right: Post-gadolinium T_1 W axial MRI; T_2 W axial MRI; fusion PET/MRI with ¹⁸F-DOPA.

8. Conclusions and Future Perspectives

There has been an explosion in the number of functional imaging techniques that can non-invasively report on multiple biological characteristics of the tumour microenvironment with great potential to guide therapy and improve outcomes as personalised therapy in oncology becomes realistic. Several functional imaging techniques have already been clinically translated, including ¹⁸F-FDG-PET and DWI-MRI. Detection of early treatment response remains challenging but, as highlighted in this review, there are numerous functional imaging biomarkers that are sensitive to the early biological effects of radiation therapy and can provide prognostic information and guide future treatment.

There are several common limitations that affect many imaging studies. Most studies are technically challenging and expensive and therefore recruit a small number of patients (typically <50). Quantification is seen as a major strength of functional imaging but a lack of consensus over the vast number of imaging biomarkers to use significantly limits the comparison of findings and meta-analysis. Unfortunately, in clinical practice, quantitative metrics do not necessarily perform better than simple qualitative analysis [159]. Furthermore, few studies prospectively define cutoff points or perform multicentre or external validation and variation between scanners is a significant barrier to quantitative analysis. Reference to the imaging biomarker roadmap should help to address these limitations and facilitate the translation of functional imaging biomarkers into clinical practice [160].

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Abbreviations

CT	computed tomography
DNP	dynamic nuclear polarisation
MRI	magnetic resonance imaging
MSOT	multispectral optoacoustic tomography
OCT	optical coherence tomography
PET	positron emission tomography
SPECT	single photon emission computed tomography

References

- 1. World Health Organization (WHO). *WHO Handbook for Reporting Results of Cancer Treatment;* WHO: Geneva, Switzerland, 1979.
- 2. Eisenhauer, E.A.; Therasse, P.; Bogaerts, J.; Schwartz, L.H.; Sargent, D.; Ford, R.; Dancey, J.; Arbuck, S.; Gwyther, S.; Mooney, M.; et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). *Eur. J. Cancer* **2009**, *45*, 228–247. [CrossRef] [PubMed]
- Therasse, P.; Arbuck, S.G.; Eisenhauer, E.A.; Wanders, J.; Kaplan, R.S.; Rubinstein, L.; Verweij, J.; Van Glabbeke, M.; van Oosterom, A.T.; Christian, M.C.; et al. New Guidelines to Evaluate the Response to Treatment in Solid Tumors. *J. Natl. Cancer Inst.* 2000, *92*, 205–216. [CrossRef] [PubMed]
- Abdulla, S.; Saada, J.; Johnson, G.; Jefferies, S.; Ajithkumar, T. Tumour progression or pseudoprogression? A review of post-treatment radiological appearances of glioblastoma. *Clin. Radiol.* 2015, 70, 1299–1312. [CrossRef] [PubMed]
- Stroobants, S.; Goeminne, J.; Seegers, M.; Dimitrijevic, S.; Dupont, P.; Nuyts, J.; Martens, M.; van den Borne, B.; Cole, P.; Sciot, R.; et al. 18FDG-Positron emission tomography for the early prediction of response in advanced soft tissue sarcoma treated with imatinib mesylate (Glivec). *Eur. J. Cancer* 2003, *39*, 2012–2020. [CrossRef]
- Barton, M.B.; Jacob, S.; Shafiq, J.; Wong, K.; Thompson, S.R.; Hanna, T.P.; Delaney, G.P. Estimating the demand for radiotherapy from the evidence: A review of changes from 2003 to 2012. *Radiother. Oncol.* 2014, *112*, 140–144. [CrossRef] [PubMed]
- 7. Thorwarth, D. Functional imaging for radiotherapy treatment planning: Current status and future directions-a review. *Br. J. Radiol.* 2015, *88*, 20150056. [CrossRef]
- Bhatnagar, P.; Subesinghe, M.; Patel, C.; Prestwich, R.; Scarsbrook, A.F. Functional imaging for radiation treatment planning, response assessment, and adaptive therapy in head and neck cancer. *Radiographics* 2013, 33, 1909–1929. [CrossRef] [PubMed]
- 9. Maier, P.; Hartmann, L.; Wenz, F.; Herskind, C. Cellular Pathways in Response to Ionizing Radiation and Their Targetability for Tumor Radiosensitization. *Int. J. Mol. Sci.* **2016**, *17*, 102. [CrossRef]
- 10. Baskar, R.; Dai, J.; Wenlong, N.; Yeo, R.; Yeoh, K.W. Biological response of cancer cells to radiation treatment. *Front. Mol. Biosci.* **2014**, *1*, 24. [CrossRef]
- 11. Eriksson, D.; Stigbrand, T. Radiation-induced cell death mechanisms. Tumour Biol. 2010, 31, 363–372. [CrossRef]
- 12. Jonathan, E.C.; Bernhard, E.J.; McKenna, W.G. How does radiation kill cells? *Curr. Opin. Chem. Biol.* **1999**, *3*, 77–83. [CrossRef]
- 13. Lorat, Y.; Timm, S.; Jakob, B.; Taucher-Scholz, G.; Rube, C.E. Clustered double-strand breaks in heterochromatin perturb DNA repair after high linear energy transfer irradiation. *Radiother. Oncol.* **2016**, *121*, 154–161. [CrossRef] [PubMed]
- 14. Barker, H.E.; Paget, J.T.; Khan, A.A.; Harrington, K.J. The tumour microenvironment after radiotherapy: Mechanisms of resistance and recurrence. *Nat. Rev. Cancer* **2015**, *15*, 409–425. [CrossRef] [PubMed]
- 15. Krysko, O.; De Ridder, L.; Cornelissen, M. Phosphatidylserine exposure during early primary necrosis (oncosis) in JB6 cells as evidenced by immunogold labeling technique. *Apoptosis* **2004**, *9*, 495–500. [CrossRef] [PubMed]

- 16. Neves, A.A.; Brindle, K.M. Imaging cell death. J. Nucl. Med. 2014, 55, 1–4. [CrossRef] [PubMed]
- Nguyen, Q.D.; Challapalli, A.; Smith, G.; Fortt, R.; Aboagye, E.O. Imaging apoptosis with positron emission tomography: 'bench to bedside' development of the caspase-3/7 specific radiotracer [¹⁸F]ICMT-11. *Eur. J. Cancer* 2012, 48, 432–440. [CrossRef]
- Belhocine, T.Z.; Blankenberg, F.G.; Kartachova, M.S.; Stitt, L.W.; Vanderheyden, J.L.; Hoebers, F.J.; Van de Wiele, C. ^{99m}Tc-Annexin A5 quantification of apoptotic tumor response: A systematic review and meta-analysis of clinical imaging trials. *Eur. J. Nucl. Med. Mol. Imaging* **2015**, *42*, 2083–2097. [CrossRef]
- Wang, F.; Fang, W.; Zhao, M.; Wang, Z.; Ji, S.; Li, Y.; Zheng, Y. Imaging paclitaxel (chemotherapy)-induced tumor apoptosis with ^{99m}Tc C2A, a domain of synaptotagmin I: A preliminary study. *Nucl. Med. Biol.* 2008, 35, 359–364. [CrossRef]
- Neves, A.A.; Xie, B.; Fawcett, S.; Alam, I.S.; Witney, T.H.; de Backer, M.M.; Summers, J.; Hughes, W.; McGuire, S.; Soloviev, D.; et al. Rapid Imaging of Tumor Cell Death In Vivo Using the C2A Domain of Synaptotagmin-I. J. Nucl. Med. 2017, 58, 881–887. [CrossRef]
- Wang, F.; Fang, W.; Zhang, M.R.; Zhao, M.; Liu, B.; Wang, Z.; Hua, Z.; Yang, M.; Kumata, K.; Hatori, A.; et al. Evaluation of chemotherapy response in VX2 rabbit lung cancer with ¹⁸F-labeled C2A domain of synaptotagmin I. *J. Nucl. Med.* 2011, *52*, 592–599. [CrossRef]
- Krishnan, A.S.; Neves, A.A.; de Backer, M.M.; Hu, D.E.; Davletov, B.; Kettunen, M.I.; Brindle, K.M. Detection of cell death in tumors by using MR imaging and a gadolinium-based targeted contrast agent. *Radiology* 2008, 246, 854–862. [CrossRef] [PubMed]
- 23. Xie, B.; Tomaszewski, M.R.; Neves, A.A.; Ros, S.; Hu, D.E.; McGuire, S.; Mullins, S.R.; Tice, D.; Sainson, R.C.A.; Bohndiek, S.E.; et al. Optoacoustic Detection of Early Therapy-Induced Tumor Cell Death Using a Targeted Imaging Agent. *Clin. Cancer Res.* **2017**, *23*, 6893–6903. [CrossRef] [PubMed]
- Elvas, F.; Boddaert, J.; Vangestel, C.; Pak, K.; Gray, B.; Kumar-Singh, S.; Staelens, S.; Stroobants, S.; Wyffels, L. ^{99m}Tc-Duramycin SPECT Imaging of Early Tumor Response to Targeted Therapy: A Comparison with ¹⁸F-FDG PET. J. Nucl. Med. 2017, 58, 665–670. [CrossRef]
- Sun, L.; Zhou, K.; Wang, W.; Zhang, X.; Ju, Z.; Qu, B.; Zhang, Z.; Wang, J.; Ling, Z.; Yu, X.; et al. [¹⁸F]ML-10 Imaging for Assessment of Apoptosis Response of Intracranial Tumor Early after Radiotherapy by PET/CT. *Contrast Media Mol. Imaging* 2018, 2018, 9365174. [CrossRef] [PubMed]
- Allen, A.M.; Ben-Ami, M.; Reshef, A.; Steinmetz, A.; Kundel, Y.; Inbar, E.; Djaldetti, R.; Davidson, T.; Fenig, E.; Ziv, I. Assessment of response of brain metastases to radiotherapy by PET imaging of apoptosis with ¹⁸F-ML-10. *Eur. J. Nucl. Med. Mol. Imaging* **2012**, *39*, 1400–1408. [CrossRef] [PubMed]
- Dubash, S.R.; Merchant, S.; Heinzmann, K.; Mauri, F.; Lavdas, I.; Inglese, M.; Kozlowski, K.; Rama, N.; Masrour, N.; Steel, J.F.; et al. Clinical translation of [¹⁸F]ICMT-11 for measuring chemotherapy-induced caspase 3/7 activation in breast and lung cancer. *Eur. J. Nucl. Med. Mol. Imaging* 2018, 45, 2285–2299. [CrossRef] [PubMed]
- Garcia-Barros, M.; Paris, F.; Cordon-Cardo, C.; Lyden, D.; Rafii, S.; Haimovitz-Friedman, A.; Fuks, Z.; Kolesnick, R. Tumor response to radiotherapy regulated by endothelial cell apoptosis. *Science* 2003, 300, 1155–1159. [CrossRef]
- Garcia-Barros, M.; Thin, T.H.; Maj, J.; Cordon-Cardo, C.; Haimovitz-Friedman, A.; Fuks, Z.; Kolesnick, R. Impact of stromal sensitivity on radiation response of tumors implanted in SCID hosts revisited. *Cancer Res.* 2010, 70, 8179–8186. [CrossRef]
- 30. Fuks, Z.; Kolesnick, R. Engaging the vascular component of the tumor response. *Cancer Cell* **2005**, *8*, 89–91. [CrossRef]
- 31. Corre, I.; Guillonneau, M.; Paris, F. Membrane signaling induced by high doses of ionizing radiation in the endothelial compartment. Relevance in radiation toxicity. *Int. J. Mol. Sci.* **2013**, *14*, 22678–22696. [CrossRef]
- 32. Miyatake, S.; Nonoguchi, N.; Furuse, M.; Yoritsune, E.; Miyata, T.; Kawabata, S.; Kuroiwa, T. Pathophysiology, diagnosis, and treatment of radiation necrosis in the brain. *Neurol. Med. Chir.* **2015**, *55*, 50–59. [CrossRef] [PubMed]
- 33. O'Connor, J.P.; Tofts, P.S.; Miles, K.A.; Parkes, L.M.; Thompson, G.; Jackson, A. Dynamic contrast-enhanced imaging techniques: CT and MRI. *Br. J. Radiol.* **2011**, *84*, S112–S120. [CrossRef] [PubMed]
- 34. Yi, C.A.; Lee, K.S.; Kim, E.A.; Han, J.; Kim, H.; Kwon, O.J.; Jeong, Y.J.; Kim, S. Solitary pulmonary nodules: Dynamic enhanced multi-detector row CT study and comparison with vascular endothelial growth factor and microvessel density. *Radiology* **2004**, *233*, 191–199. [CrossRef] [PubMed]

- Bellomi, M.; Petralia, G.; Sonzogni, A.; Zampino, M.G.; Rocca, A. CT perfusion for the monitoring of neoadjuvant chemotherapy and radiation therapy in rectal carcinoma: Initial experience. *Radiology* 2007, 244, 486–493. [CrossRef] [PubMed]
- Sahani, D.V.; Kalva, S.P.; Hamberg, L.M.; Hahn, P.F.; Willett, C.G.; Saini, S.; Mueller, P.R.; Lee, T.Y. Assessing tumor perfusion and treatment response in rectal cancer with multisection CT: Initial observations. *Radiology* 2005, 234, 785–792. [CrossRef] [PubMed]
- Ursino, S.; Faggioni, L.; Guidoccio, F.; Ferrazza, P.; Seccia, V.; Neri, E.; Cernusco, L.N.; Delishaj, D.; Morganti, R.; Volterrani, D.; et al. Role of perfusion CT in the evaluation of functional primary tumour response after radiochemotherapy in head and neck cancer: Preliminary findings. *Br. J. Radiol.* 2016, *89*, 20151070. [CrossRef]
- Coolens, C.; Driscoll, B.; Foltz, W.D.; Jaffray, D.A.; Chung, C. Early Detection of Tumor Response Using Volumetric DCE-CT and DCE-MRI in Metastatic Brain Patients Treated with Radiosurgery. *Int. J. Radiat. Oncol. Biol. Phys.* 2015, 93, S7. [CrossRef]
- Surlan-Popovic, K.; Bisdas, S.; Rumboldt, Z.; Koh, T.S.; Strojan, P. Changes in perfusion CT of advanced squamous cell carcinoma of the head and neck treated during the course of concomitant chemoradiotherapy. *Am. J. Neuroradiol.* 2010, *31*, 570–575. [CrossRef]
- Kino, A.; Shaffer, J.; Maturen, K.E.; Schmiedeskamp, H.; Koong, A.C.; Chang, D.T.; Fleischmann, D.; Kamaya, A. Perfusion CT measurements predict tumor response in rectal carcinoma. *Abdom. Radiol.* 2017, 42, 1132–1140. [CrossRef]
- 41. Banks, T.I.; von Eyben, R.; Hristov, D.; Kidd, E.A. Pilot study of combined FDG-PET and dynamic contrast-enhanced CT of locally advanced cervical carcinoma before and during concurrent chemoradiotherapy suggests association between changes in tumor blood volume and treatment response. *Cancer Med.* **2018**, *7*, 3642–3651. [CrossRef]
- 42. Zahra, M.A.; Hollingsworth, K.G.; Sala, E.; Lomas, D.J.; Tan, L.T. Dynamic contrast-enhanced MRI as a predictor of tumour response to radiotherapy. *Lancet Oncol.* **2007**, *8*, 63–74. [CrossRef]
- 43. Padhani, A.R.; Husband, J.E. Dynamic contrast-enhanced MRI studies in oncology with an emphasis on quantification, validation and human studies. *Clin. Radiol.* **2001**, *56*, 607–620. [CrossRef] [PubMed]
- 44. Khalifa, F.; Soliman, A.; El-Baz, A.; Abou El-Ghar, M.; El-Diasty, T.; Gimel'farb, G.; Ouseph, R.; Dwyer, A.C. Models and methods for analyzing DCE-MRI: A review. *Med. Phys.* **2014**, *41*, 124301. [CrossRef] [PubMed]
- 45. Petcharunpaisan, S.; Ramalho, J.; Castillo, M. Arterial spin labeling in neuroimaging. *World J. Radiol.* **2010**, *2*, 384–398. [CrossRef] [PubMed]
- 46. Mayr, N.A.; Yuh, W.T.; Magnotta, V.A.; Ehrhardt, J.C.; Wheeler, J.A.; Sorosky, J.I.; Davis, C.S.; Wen, B.C.; Martin, D.D.; Pelsang, R.E.; et al. Tumor perfusion studies using fast magnetic resonance imaging technique in advanced cervical cancer: A new noninvasive predictive assay. *Int. J. Radiat. Oncol. Biol. Phys.* **1996**, *36*, 623–633. [CrossRef]
- Zahra, M.A.; Tan, L.T.; Priest, A.N.; Graves, M.J.; Arends, M.; Crawford, R.A.; Brenton, J.D.; Lomas, D.J.; Sala, E. Semiquantitative and quantitative dynamic contrast-enhanced magnetic resonance imaging measurements predict radiation response in cervix cancer. *Int. J. Radiat. Oncol. Biol. Phys.* 2009, 74, 766–773. [CrossRef]
- Yuh, W.T.; Mayr, N.A.; Jarjoura, D.; Wu, D.; Grecula, J.C.; Lo, S.S.; Edwards, S.M.; Magnotta, V.A.; Sammet, S.; Zhang, H.; et al. Predicting control of primary tumor and survival by DCE MRI during early therapy in cervical cancer. *Investig. Radiol.* 2009, 44, 343–350. [CrossRef]
- Dickie, B.R.; Rose, C.J.; Kershaw, L.E.; Withey, S.B.; Carrington, B.M.; Davidson, S.E.; Hutchison, G.; West, C.M.L. The prognostic value of dynamic contrast-enhanced MRI contrast agent transfer constant K^{trans} in cervical cancer is explained by plasma flow rather than vessel permeability. *Br. J. Cancer* 2017, *116*, 1436–1443. [CrossRef] [PubMed]
- 50. Semple, S.I.; Harry, V.N.; Parkin, D.E.; Gilbert, F.J. A combined pharmacokinetic and radiologic assessment of dynamic contrast-enhanced magnetic resonance imaging predicts response to chemoradiation in locally advanced cervical cancer. *Int. J. Radiat. Oncol. Biol. Phys.* **2009**, *75*, 611–617. [CrossRef]
- 51. Dijkhoff, R.A.P.; Beets-Tan, R.G.H.; Lambregts, D.M.J.; Beets, G.L.; Maas, M. Value of DCE-MRI for staging and response evaluation in rectal cancer: A systematic review. *Eur. J. Radiol.* **2017**, *95*, 155–168. [CrossRef]
- 52. King, A.D.; Thoeny, H.C. Functional MRI for the prediction of treatment response in head and neck squamous cell carcinoma: Potential and limitations. *Cancer Imaging* **2016**, *16*, 23. [CrossRef] [PubMed]

- 53. Jakubovic, R.; Sahgal, A.; Soliman, H.; Milwid, R.; Zhang, L.; Eilaghi, A.; Aviv, R.I. Magnetic resonance imaging-based tumour perfusion parameters are biomarkers predicting response after radiation to brain metastases. *Clin. Oncol.* **2014**, *26*, 704–712. [CrossRef] [PubMed]
- 54. Cao, Y.; Tsien, C.I.; Nagesh, V.; Junck, L.; Ten Haken, R.; Ross, B.D.; Chenevert, T.L.; Lawrence, T.S. Survival prediction in high-grade gliomas by MRI perfusion before and during early stage of RT. *Int. J. Radiat. Oncol. Biol. Phys.* **2006**, *64*, 876–885. [CrossRef] [PubMed]
- 55. Wang, P.; Li, J.; Diao, Q.; Lin, Y.; Zhang, J.; Li, L.; Yang, G.; Fang, X.; Li, X.; Chen, Y.; et al. Assessment of glioma response to radiotherapy using 3D pulsed-continuous arterial spin labeling and 3D segmented volume. *Eur. J. Radiol.* **2016**, *85*, 1987–1992. [CrossRef] [PubMed]
- 56. Hawighorst, H.; Weikel, W.; Knapstein, P.G.; Knopp, M.V.; Zuna, I.; Schonberg, S.O.; Vaupel, P.; van Kaick, G. Angiogenic activity of cervical carcinoma: Assessment by functional magnetic resonance imaging-based parameters and a histomorphological approach in correlation with disease outcome. *Clin. Cancer Res.* 1998, 4, 2305–2312. [PubMed]
- 57. Bisdas, S.; Naegele, T.; Ritz, R.; Dimostheni, A.; Pfannenberg, C.; Reimold, M.; Koh, T.S.; Ernemann, U. Distinguishing recurrent high-grade gliomas from radiation injury: A pilot study using dynamic contrast-enhanced MR imaging. *Acad. Radiol.* **2011**, *18*, 575–583. [CrossRef] [PubMed]
- 58. Patel, P.; Baradaran, H.; Delgado, D.; Askin, G.; Christos, P.; John Tsiouris, A.; Gupta, A. MR perfusion-weighted imaging in the evaluation of high-grade gliomas after treatment: A systematic review and meta-analysis. *Neuro-Oncology* **2017**, *19*, 118–127. [CrossRef]
- Ozsunar, Y.; Mullins, M.E.; Kwong, K.; Hochberg, F.H.; Ament, C.; Schaefer, P.W.; Gonzalez, R.G.; Lev, M.H. Glioma recurrence versus radiation necrosis? A pilot comparison of arterial spin-labeled, dynamic susceptibility contrast enhanced MRI, and FDG-PET imaging. *Acad. Radiol.* 2010, *17*, 282–290. [CrossRef]
- 60. Ye, J.; Bhagat, S.K.; Li, H.; Luo, X.; Wang, B.; Liu, L.; Yang, G. Differentiation between recurrent gliomas and radiation necrosis using arterial spin labeling perfusion imaging. *Exp. Ther. Med.* **2016**, *11*, 2432–2436. [CrossRef]
- 61. Razek, A.; El-Serougy, L.; Abdelsalam, M.; Gaballa, G.; Talaat, M. Differentiation of residual/recurrent gliomas from postradiation necrosis with arterial spin labeling and diffusion tensor magnetic resonance imaging-derived metrics. *Neuroradiology* **2018**, *60*, 169–177. [CrossRef]
- 62. Nyberg, E.; Honce, J.; Kleinschmidt-DeMasters, B.K.; Shukri, B.; Kreidler, S.; Nagae, L. Arterial spin labeling: Pathologically proven superiority over conventional MRI for detection of high-grade glioma progression after treatment. *Neuroradiol. J.* **2016**, *29*, 377–383. [CrossRef] [PubMed]
- 63. Lee, Y.J.; Kim, S.H.; Kang, B.J.; Kim, Y.J. Contrast-Enhanced Ultrasound for Early Prediction of Response of Breast Cancer to Neoadjuvant Chemotherapy. *Ultraschall Med.* **2018**. [CrossRef] [PubMed]
- 64. Kim, Y.; Kim, S.H.; Song, B.J.; Kang, B.J.; Yim, K.I.; Lee, A.; Nam, Y. Early Prediction of Response to Neoadjuvant Chemotherapy Using Dynamic Contrast-Enhanced MRI and Ultrasound in Breast Cancer. *Korean J. Radiol.* **2018**, *19*, 682–691. [CrossRef] [PubMed]
- Lassau, N.; Koscielny, S.; Albiges, L.; Chami, L.; Benatsou, B.; Chebil, M.; Roche, A.; Escudier, B.J. Metastatic renal cell carcinoma treated with sunitinib: Early evaluation of treatment response using dynamic contrast-enhanced ultrasonography. *Clin. Cancer Res.* 2010, *16*, 1216–1225. [CrossRef] [PubMed]
- Kasoji, S.K.; Rivera, J.N.; Gessner, R.C.; Chang, S.X.; Dayton, P.A. Early Assessment of Tumor Response to Radiation Therapy using High-Resolution Quantitative Microvascular Ultrasound Imaging. *Theranostics* 2018, *8*, 156–168. [CrossRef]
- Arteaga-Marrero, N.; Mainou-Gomez, J.F.; Brekke Rygh, C.; Lutay, N.; Roehrich, D.; Reed, R.K.; Olsen, D.R. Radiation treatment monitoring with DCE-US in CWR22 prostate tumor xenografts. *Acta Radiol.* 2018. [CrossRef]
- Abdollahi, A.; Griggs, D.W.; Zieher, H.; Roth, A.; Lipson, K.E.; Saffrich, R.; Grone, H.J.; Hallahan, D.E.; Reisfeld, R.A.; Debus, J.; et al. Inhibition of α_vβ₃ integrin survival signaling enhances antiangiogenic and antitumor effects of radiotherapy. *Clin. Cancer Res.* 2005, *11*, 6270–6279. [CrossRef]
- 69. Zysk, A.M.; Nguyen, F.T.; Oldenburg, A.L.; Marks, D.L.; Boppart, S.A. Optical coherence tomography: A review of clinical development from bench to bedside. *J. Biomed. Opt.* **2007**, *12*, 051403. [CrossRef]
- Demidov, V.; Maeda, A.; Sugita, M.; Madge, V.; Sadanand, S.; Flueraru, C.; Vitkin, I.A. Preclinical longitudinal imaging of tumor microvascular radiobiological response with functional optical coherence tomography. *Sci. Rep.* 2018, *8*, 38. [CrossRef]

- 71. Maeda, A.; Leung, M.K.; Conroy, L.; Chen, Y.; Bu, J.; Lindsay, P.E.; Mintzberg, S.; Virtanen, C.; Tsao, J.; Winegarden, N.A.; et al. In vivo optical imaging of tumor and microvascular response to ionizing radiation. *PLoS ONE* 2012, 7, e42133. [CrossRef]
- 72. McNally, L.R.; Mezera, M.; Morgan, D.E.; Frederick, P.J.; Yang, E.S.; Eltoum, I.E.; Grizzle, W.E. Current and Emerging Clinical Applications of Multispectral Optoacoustic Tomography (MSOT) in Oncology. *Clin. Cancer Res.* **2016**, *22*, 3432–3439. [CrossRef] [PubMed]
- 73. Tomaszewski, M.R.; Gehrung, M.; Joseph, J.; Quiros-Gonzalez, I.; Disselhorst, J.A.; Bohndiek, S.E. Oxygen-Enhanced and Dynamic Contrast-Enhanced Optoacoustic Tomography Provide Surrogate Biomarkers of Tumor Vascular Function, Hypoxia, and Necrosis. *Cancer Res.* 2018, 78, 5980–5991. [CrossRef] [PubMed]
- 74. Rich, L.J.; Seshadri, M. Photoacoustic monitoring of tumor and normal tissue response to radiation. *Sci. Rep.* **2016**, *6*, 21237. [CrossRef] [PubMed]
- 75. Zackrisson, S.; van de Ven, S.; Gambhir, S.S. Light in and sound out: Emerging translational strategies for photoacoustic imaging. *Cancer Res.* **2014**, *74*, 979–1004. [CrossRef] [PubMed]
- 76. Lodge, M.A.; Jacene, H.A.; Pili, R.; Wahl, R.L. Reproducibility of tumor blood flow quantification with 15O-water PET. J. Nucl. Med. 2008, 49, 1620–1627. [CrossRef] [PubMed]
- 77. De Langen, A.J.; Lubberink, M.; Boellaard, R.; Spreeuwenberg, M.D.; Smit, E.F.; Hoekstra, O.S.; Lammertsma, A.A. Reproducibility of tumor perfusion measurements using 15O-labeled water and PET. *J. Nucl. Med.* 2008, 49, 1763–1768. [CrossRef] [PubMed]
- 78. Lehtio, K.; Eskola, O.; Viljanen, T.; Oikonen, V.; Gronroos, T.; Sillanmaki, L.; Grenman, R.; Minn, H. Imaging perfusion and hypoxia with PET to predict radiotherapy response in head-and-neck cancer. *Int. J. Radiat. Oncol. Biol. Phys.* **2004**, *59*, 971–982. [CrossRef]
- 79. Rockwell, S.; Dobrucki, I.T.; Kim, E.Y.; Marrison, S.T.; Vu, V.T. Hypoxia and radiation therapy: Past history, ongoing research and future promise. *Curr. Mol. Med.* **2009**, *9*, 442–458. [CrossRef]
- 80. Harris, A.L. Hypoxia—A key regulatory factor in tumour growth. Nat. Rev. Cancer 2002, 2, 38–47. [CrossRef]
- Masaki, Y.; Shimizu, Y.; Yoshioka, T.; Nishijima, K.I.; Zhao, S.; Higashino, K.; Numata, Y.; Tamaki, N.; Kuge, Y. FMISO accumulation in tumor is dependent on glutathione conjugation capacity in addition to hypoxic state. *Ann. Nucl. Med.* 2017, *31*, 596–604. [CrossRef]
- Lock, S.; Perrin, R.; Seidlitz, A.; Bandurska-Luque, A.; Zschaeck, S.; Zophel, K.; Krause, M.; Steinbach, J.; Kotzerke, J.; Zips, D.; et al. Residual tumour hypoxia in head-and-neck cancer patients undergoing primary radiochemotherapy, final results of a prospective trial on repeat FMISO-PET imaging. *Radiother. Oncol.* 2017, 124, 533–540. [CrossRef] [PubMed]
- 83. Welz, S.; Monnich, D.; Pfannenberg, C.; Nikolaou, K.; Reimold, M.; La Fougere, C.; Reischl, G.; Mauz, P.S.; Paulsen, F.; Alber, M.; et al. Prognostic value of dynamic hypoxia PET in head and neck cancer: Results from a planned interim analysis of a randomized phase II hypoxia-image guided dose escalation trial. *Radiother. Oncol.* 2017, 124, 526–532. [CrossRef] [PubMed]
- Chang, J.H.; Wada, M.; Anderson, N.J.; Lim Joon, D.; Lee, S.T.; Gong, S.J.; Gunawardana, D.H.; Sachinidis, J.; O'Keefe, G.; Gan, H.K.; et al. Hypoxia-targeted radiotherapy dose painting for head and neck cancer using ¹⁸F-FMISO PET: A biological modeling study. *Acta Oncol.* **2013**, *52*, 1723–1729. [CrossRef]
- 85. Vera, P.; Thureau, S.; Chaumet-Riffaud, P.; Modzelewski, R.; Bohn, P.; Vermandel, M.; Hapdey, S.; Pallardy, A.; Mahe, M.A.; Lacombe, M.; et al. Phase II Study of a Radiotherapy Total Dose Increase in Hypoxic Lesions Identified by ¹⁸F-Misonidazole PET/CT in Patients with Non-Small Cell Lung Carcinoma (RTEP5 Study). J. Nucl. Med. **2017**, 58, 1045–1053. [CrossRef]
- Chen, L.; Zhang, Z.; Kolb, H.C.; Walsh, J.C.; Zhang, J.; Guan, Y. ¹⁸F-HX4 hypoxia imaging with PET/CT in head and neck cancer: A comparison with ¹⁸F-FMISO. *Nucl. Med. Commun.* 2012, 33, 1096–1102. [CrossRef] [PubMed]
- 87. Saga, T.; Inubushi, M.; Koizumi, M.; Yoshikawa, K.; Zhang, M.R.; Obata, T.; Tanimoto, K.; Harada, R.; Uno, T.; Fujibayashi, Y. Prognostic value of PET/CT with ¹⁸F-fluoroazomycin arabinoside for patients with head and neck squamous cell carcinomas receiving chemoradiotherapy. *Ann. Nucl. Med.* **2016**, *30*, 217–224. [CrossRef] [PubMed]
- Servagi-Vernat, S.; Differding, S.; Sterpin, E.; Hanin, F.X.; Labar, D.; Bol, A.; Lee, J.A.; Gregoire, V. Hypoxia-guided adaptive radiation dose escalation in head and neck carcinoma: A planning study. *Acta Oncol.* 2015, 54, 1008–1016. [CrossRef]

- Bollineni, V.R.; Koole, M.J.; Pruim, J.; Brouwer, C.L.; Wiegman, E.M.; Groen, H.J.; Vlasman, R.; Halmos, G.B.; Oosting, S.F.; Langendijk, J.A.; et al. Dynamics of tumor hypoxia assessed by 18F-FAZA PET/CT in head and neck and lung cancer patients during chemoradiation: Possible implications for radiotherapy treatment planning strategies. *Radiother. Oncol.* 2014, *113*, 198–203. [CrossRef]
- Servagi-Vernat, S.; Differding, S.; Hanin, F.X.; Labar, D.; Bol, A.; Lee, J.A.; Gregoire, V. A prospective clinical study of ¹⁸F-FAZA PET-CT hypoxia imaging in head and neck squamous cell carcinoma before and during radiation therapy. *Eur. J. Nucl. Med. Mol. Imaging* **2014**, *41*, 1544–1552. [CrossRef]
- Trinkaus, M.E.; Blum, R.; Rischin, D.; Callahan, J.; Bressel, M.; Segard, T.; Roselt, P.; Eu, P.; Binns, D.; MacManus, M.P.; et al. Imaging of hypoxia with 18F-FAZA PET in patients with locally advanced non-small cell lung cancer treated with definitive chemoradiotherapy. *J. Med. Imaging Radiat. Oncol.* 2013, 57, 475–481. [CrossRef]
- 92. Lewin, J.; Khamly, K.K.; Young, R.J.; Mitchell, C.; Hicks, R.J.; Toner, G.C.; Ngan, S.Y.; Chander, S.; Powell, G.J.; Herschtal, A.; et al. A phase Ib/II translational study of sunitinib with neoadjuvant radiotherapy in soft-tissue sarcoma. *Br. J. Cancer* 2014, *111*, 2254–2261. [CrossRef] [PubMed]
- 93. Horsman, M.R.; Mortensen, L.S.; Petersen, J.B.; Busk, M.; Overgaard, J. Imaging hypoxia to improve radiotherapy outcome. *Nat. Rev. Clin. Oncol.* **2012**, *9*, 674–687. [CrossRef] [PubMed]
- 94. Nyflot, M.J.; Kruser, T.J.; Traynor, A.M.; Khuntia, D.; Yang, D.T.; Hartig, G.K.; McCulloch, T.M.; Wiederholt, P.A.; Gentry, L.R.; Hoang, T.; et al. Phase 1 trial of bevacizumab with concurrent chemoradiation therapy for squamous cell carcinoma of the head and neck with exploratory functional imaging of tumor hypoxia, proliferation, and perfusion. *Int. J. Radiat. Oncol. Biol. Phys.* 2015, *91*, 942–951. [CrossRef] [PubMed]
- 95. Minagawa, Y.; Shizukuishi, K.; Koike, I.; Horiuchi, C.; Watanuki, K.; Hata, M.; Omura, M.; Odagiri, K.; Tohnai, I.; Inoue, T.; et al. Assessment of tumor hypoxia by ⁶²Cu-ATSM PET/CT as a predictor of response in head and neck cancer: A pilot study. *Ann. Nucl. Med.* **2011**, *25*, 339–345. [CrossRef] [PubMed]
- 96. Dietz, D.W.; Dehdashti, F.; Grigsby, P.W.; Malyapa, R.S.; Myerson, R.J.; Picus, J.; Ritter, J.; Lewis, J.S.; Welch, M.J.; Siegel, B.A. Tumor hypoxia detected by positron emission tomography with ⁶⁰Cu-ATSM as a predictor of response and survival in patients undergoing Neoadjuvant chemoradiotherapy for rectal carcinoma: A pilot study. *Dis. Colon Rectum* **2008**, *51*, 1641–1648. [CrossRef] [PubMed]
- Grassi, I.; Nanni, C.; Cicoria, G.; Blasi, C.; Bunkheila, F.; Lopci, E.; Colletti, P.M.; Rubello, D.; Fanti, S. Usefulness of ⁶⁴Cu-ATSM in head and neck cancer: A preliminary prospective study. *Clin. Nucl. Med.* 2014, 39, e59–e63. [CrossRef] [PubMed]
- O'Connor, J.; Robinson, S.; Waterton, J. Imaging tumour hypoxia with oxygen-enhanced MRI and BOLD MRI. *Br. J. Radiol.* 2018. [CrossRef] [PubMed]
- O'Connor, J.P.; Boult, J.K.; Jamin, Y.; Babur, M.; Finegan, K.G.; Williams, K.J.; Little, R.A.; Jackson, A.; Parker, G.J.; Reynolds, A.R.; et al. Oxygen-Enhanced MRI Accurately Identifies, Quantifies, and Maps Tumor Hypoxia in Preclinical Cancer Models. *Cancer Res.* 2016, *76*, 787–795. [CrossRef] [PubMed]
- Dewhirst, M.W.; Birer, S.R. Oxygen-Enhanced MRI Is a Major Advance in Tumor Hypoxia Imaging. *Cancer Res.* 2016, 76, 769–772. [CrossRef] [PubMed]
- 101. Rodrigues, L.M.; Howe, F.A.; Griffiths, J.R.; Robinson, S.P. Tumor R₂* is a prognostic indicator of acute radiotherapeutic response in rodent tumors. *J. Magn. Reson. Imaging* **2004**, *19*, 482–488. [CrossRef] [PubMed]
- Rumley, C.N.; Lee, M.T.; Holloway, L.; Rai, R.; Min, M.; Forstner, D.; Fowler, A.; Liney, G. Multiparametric magnetic resonance imaging in mucosal primary head and neck cancer: A prospective imaging biomarker study. *BMC Cancer* 2017, *17*, 475. [CrossRef] [PubMed]
- White, D.A.; Zhang, Z.; Li, L.; Gerberich, J.; Stojadinovic, S.; Peschke, P.; Mason, R.P. Developing oxygen-enhanced magnetic resonance imaging as a prognostic biomarker of radiation response. *Cancer Lett.* 2016, 380, 69–77. [CrossRef] [PubMed]
- 104. Beeman, S.C.; Shui, Y.B.; Perez-Torres, C.J.; Engelbach, J.A.; Ackerman, J.J.; Garbow, J.R. O₂-sensitive MRI distinguishes brain tumor versus radiation necrosis in murine models. *Magn. Reson. Med.* 2016, 75, 2442–2447. [CrossRef] [PubMed]
- 105. Panagiotaki, E.; Walker-Samuel, S.; Siow, B.; Johnson, S.P.; Rajkumar, V.; Pedley, R.B.; Lythgoe, M.F.; Alexander, D.C. Noninvasive quantification of solid tumor microstructure using VERDICT MRI. *Cancer Res.* 2014, 74, 1902–1912. [CrossRef] [PubMed]
- 106. Steven, A.J.; Zhuo, J.; Melhem, E.R. Diffusion kurtosis imaging: An emerging technique for evaluating the microstructural environment of the brain. *Am. J. Roentgenol.* **2014**, 202, W26–W33. [CrossRef]

- Soares, J.M.; Marques, P.; Alves, V.; Sousa, N. A hitchhiker's guide to diffusion tensor imaging. *Front. Neurosci.* 2013, 7, 31. [CrossRef] [PubMed]
- 108. Schilling, F.; Ros, S.; Hu, D.E.; D'Santos, P.; McGuire, S.; Mair, R.; Wright, A.J.; Mannion, E.; Franklin, R.J.; Neves, A.A.; et al. MRI measurements of reporter-mediated increases in transmembrane water exchange enable detection of a gene reporter. *Nat. Biotechnol.* 2017, 35, 75–80. [CrossRef]
- 109. Lampinen, B.; Szczepankiewicz, F.; van Westen, D.; Englund, E.; Sundgren, P.C.; Lätt, J.; Ståhlberg, F.; Nilsson, M. Optimal experimental design for filter exchange imaging: Apparent exchange rate measurements in the healthy brain and in intracranial tumors. *Magn. Reson. Med.* 2017, 77, 1104–1114. [CrossRef]
- 110. Sehy, J.V.; Ackerman, J.J.; Neil, J.J. Evidence that both fast and slow water ADC components arise from intracellular space. *Magn. Reson. Med.* **2002**, *48*, 765–770. [CrossRef]
- 111. Surov, A.; Meyer, H.J.; Wienke, A. Correlation between apparent diffusion coefficient (ADC) and cellularity is different in several tumors: A meta-analysis. *Oncotarget* **2017**, *8*, 59492–59499. [CrossRef]
- 112. Afaq, A.; Andreou, A.; Koh, D.M. Diffusion-weighted magnetic resonance imaging for tumour response assessment: Why, when and how? *Cancer Imaging* **2010**, *10*, S179–S188. [CrossRef] [PubMed]
- 113. Chenevert, T.L.; Stegman, L.D.; Taylor, J.M.; Robertson, P.L.; Greenberg, H.S.; Rehemtulla, A.; Ross, B.D. Diffusion magnetic resonance imaging: An early surrogate marker of therapeutic efficacy in brain tumors. J. Natl. Cancer Inst. 2000, 92, 2029–2036. [CrossRef]
- 114. Mahmood, F.; Johannesen, H.H.; Geertsen, P.; Hansen, R.H. Repeated diffusion MRI reveals earliest time point for stratification of radiotherapy response in brain metastases. *Phys. Med. Biol.* 2017, 62, 2990–3002. [CrossRef] [PubMed]
- 115. Kim, S.; Loevner, L.; Quon, H.; Sherman, E.; Weinstein, G.; Kilger, A.; Poptani, H. Diffusion-weighted magnetic resonance imaging for predicting and detecting early response to chemoradiation therapy of squamous cell carcinomas of the head and neck. *Clin. Cancer Res.* **2009**, *15*, 986–994. [CrossRef]
- 116. Schreuder, S.M.; Lensing, R.; Stoker, J.; Bipat, S. Monitoring treatment response in patients undergoing chemoradiotherapy for locally advanced uterine cervical cancer by additional diffusion-weighted imaging: A systematic review. J. Magn. Reson. Imaging 2015, 42, 572–594. [CrossRef] [PubMed]
- 117. Joye, I.; Deroose, C.M.; Vandecaveye, V.; Haustermans, K. The role of diffusion-weighted MRI and ¹⁸F-FDG PET/CT in the prediction of pathologic complete response after radiochemotherapy for rectal cancer: A systematic review. *Radiother. Oncol.* **2014**, *113*, 158–165. [CrossRef] [PubMed]
- 118. Joye, I.; Debucquoy, A.; Deroose, C.M.; Vandecaveye, V.; Cutsem, E.V.; Wolthuis, A.; D'Hoore, A.; Sagaert, X.; Zhou, M.; Gevaert, O.; et al. Quantitative imaging outperforms molecular markers when predicting response to chemoradiotherapy for rectal cancer. *Radiother. Oncol.* 2017, *124*, 104–109. [CrossRef]
- 119. Chu, H.H.; Choi, S.H.; Ryoo, I.; Kim, S.C.; Yeom, J.A.; Shin, H.; Jung, S.C.; Lee, A.L.; Yoon, T.J.; Kim, T.M.; et al. Differentiation of true progression from pseudoprogression in glioblastoma treated with radiation therapy and concomitant temozolomide: Comparison study of standard and high-b-value diffusion-weighted imaging. *Radiology* **2013**, *269*, 831–840. [CrossRef]
- 120. Kim, J.Y.; Park, J.E.; Jo, Y.; Shim, W.H.; Nam, S.J.; Kim, J.H.; Yoo, R.E.; Choi, S.H.; Kim, H.S. Incorporating diffusion- and perfusion-weighted MRI into a radiomics model improves diagnostic performance for pseudoprogression in glioblastoma patients. *Neuro-Oncology* **2018**. [CrossRef]
- 121. Mannelli, L.; Kim, S.; Hajdu, C.H.; Babb, J.S.; Clark, T.W.; Taouli, B. Assessment of tumor necrosis of hepatocellular carcinoma after chemoembolization: Diffusion-weighted and contrast-enhanced MRI with histopathologic correlation of the explanted liver. *Am. J. Roentgenol.* **2009**, *193*, 1044–1052. [CrossRef]
- 122. Hein, P.A.; Eskey, C.J.; Dunn, J.F.; Hug, E.B. Diffusion-weighted imaging in the follow-up of treated high-grade gliomas: Tumor recurrence versus radiation injury. *Am. J. Neuroradiol.* 2004, 25, 201–209. [PubMed]
- 123. Asao, C.; Korogi, Y.; Kitajima, M.; Hirai, T.; Baba, Y.; Makino, K.; Kochi, M.; Morishita, S.; Yamashita, Y. Diffusion-weighted imaging of radiation-induced brain injury for differentiation from tumor recurrence. *Am. J. Neuroradiol.* 2005, 26, 1455–1460. [PubMed]
- 124. Zeng, Q.S.; Li, C.F.; Liu, H.; Zhen, J.H.; Feng, D.C. Distinction between recurrent glioma and radiation injury using magnetic resonance spectroscopy in combination with diffusion-weighted imaging. *Int. J. Radiat. Oncol. Biol. Phys.* 2007, 68, 151–158. [CrossRef] [PubMed]
- 125. Brindle, K. New approaches for imaging tumour responses to treatment. *Nat. Rev. Cancer* 2008, *8*, 94–107. [CrossRef] [PubMed]

- 126. Van Zijl, P.C.; Yadav, N.N. Chemical exchange saturation transfer (CEST): What is in a name and what isn't? *Magn. Reson. Med.* **2011**, *65*, 927–948. [CrossRef] [PubMed]
- 127. Zhou, J.; Zhu, H.; Lim, M.; Blair, L.; Quinones-Hinojosa, A.; Messina, S.A.; Eberhart, C.G.; Pomper, M.G.; Laterra, J.; Barker, P.B.; et al. Three-dimensional amide proton transfer MR imaging of gliomas: Initial experience and comparison with gadolinium enhancement. *J. Magn. Reson. Imaging* **2013**, *38*, 1119–1128. [CrossRef]
- Mehrabian, H.; Desmond, K.L.; Soliman, H.; Sahgal, A.; Stanisz, G.J. Differentiation between Radiation Necrosis and Tumor Progression Using Chemical Exchange Saturation Transfer. *Clin. Cancer Res.* 2017, 23, 3667–3675. [CrossRef] [PubMed]
- 129. Warburg, O.; Negelein, E. Über den stoffwechsel der carcinomzelle. Biochem. Z. 1924, 152, 319–344. [CrossRef]
- 130. Hensley, C.T.; Faubert, B.; Yuan, Q.; Lev-Cohain, N.; Jin, E.; Kim, J.; Jiang, L.; Ko, B.; Skelton, R.; Loudat, L.; et al. Metabolic Heterogeneity in Human Lung Tumors. *Cell* **2016**, *164*, 681–694. [CrossRef]
- 131. Wahl, R.L.; Jacene, H.; Kasamon, Y.; Lodge, M.A. From RECIST to PERCIST: Evolving Considerations for PET response criteria in solid tumors. *J. Nucl. Med.* **2009**, *50* (Suppl. 1), 122S–150S. [CrossRef]
- 132. Young, H.; Baum, R.; Cremerius, U.; Herholz, K.; Hoekstra, O.; Lammertsma, A.A.; Pruim, J.; Price, P. Measurement of clinical and subclinical tumour response using [¹⁸F]-fluorodeoxyglucose and positron emission tomography: Review and 1999 EORTC recommendations. European Organization for Research and Treatment of Cancer (EORTC) PET Study Group. *Eur. J. Cancer* **1999**, 35, 1773–1782. [CrossRef]
- Slevin, F.; Subesinghe, M.; Ramasamy, S.; Sen, M.; Scarsbrook, A.F.; Prestwich, R.J. Assessment of outcomes with delayed ¹⁸F-FDG PET-CT response assessment in head and neck squamous cell carcinoma. *Br. J. Radiol.* 2015, *88*, 20140592. [CrossRef]
- Prestwich, R.J.; Subesinghe, M.; Gilbert, A.; Chowdhury, F.U.; Sen, M.; Scarsbrook, A.F. Delayed response assessment with FDG-PET-CT following (chemo) radiotherapy for locally advanced head and neck squamous cell carcinoma. *Clin. Radiol.* 2012, 67, 966–975. [CrossRef]
- 135. Hicks, R.J.; Mac Manus, M.P.; Matthews, J.P.; Hogg, A.; Binns, D.; Rischin, D.; Ball, D.L.; Peters, L.J. Early FDG-PET imaging after radical radiotherapy for non-small-cell lung cancer: Inflammatory changes in normal tissues correlate with tumor response and do not confound therapeutic response evaluation. *Int. J. Radiat. Oncol. Biol. Phys.* 2004, 60, 412–418. [CrossRef] [PubMed]
- Hentschel, M.; Appold, S.; Schreiber, A.; Abramyuk, A.; Abolmaali, N.; Kotzerke, J.; Baumann, M.; Zophel, K. Serial FDG-PET on patients with head and neck cancer: Implications for radiation therapy. *Int. J. Radiat. Biol.* 2009, *85*, 796–804. [CrossRef] [PubMed]
- 137. Kubota, R.; Yamada, S.; Kubota, K.; Ishiwata, K.; Tamahashi, N.; Ido, T. Intratumoral distribution of fluorine-18-fluorodeoxyglucose in vivo: High accumulation in macrophages and granulation tissues studied by microautoradiography. J. Nucl. Med. 1992, 33, 1972–1980. [PubMed]
- 138. Ardenkjaer-Larsen, J.H.; Fridlund, B.; Gram, A.; Hansson, G.; Hansson, L.; Lerche, M.H.; Servin, R.; Thaning, M.; Golman, K. Increase in signal-to-noise ratio of >10,000 times in liquid-state NMR. *Proc. Natl. Acad. Sci. USA* 2003, 100, 10158–10163. [CrossRef]
- 139. Hesketh, R.L.; Brindle, K.M. Magnetic resonance imaging of cancer metabolism with hyperpolarized ¹³C-labeled cell metabolites. *Curr. Opin. Chem. Biol.* **2018**, 45, 187–194. [CrossRef]
- 140. Gutte, H.; Hansen, A.E.; Henriksen, S.T.; Johannesen, H.H.; Ardenkjaer-Larsen, J.; Vignaud, A.; Hansen, A.E.; Borresen, B.; Klausen, T.L.; Wittekind, A.M.; et al. Simultaneous hyperpolarized ¹³C-pyruvate MRI and ¹⁸F-FDG-PET in cancer (hyperPET): Feasibility of a new imaging concept using a clinical PET/MRI scanner. *Am. J. Nucl. Med. Mol. Imaging* **2015**, *5*, 38–45.
- Aggarwal, R.; Vigneron, D.B.; Kurhanewicz, J. Hyperpolarized 1-[¹³C]-Pyruvate Magnetic Resonance Imaging Detects an Early Metabolic Response to Androgen Ablation Therapy in Prostate Cancer. *Eur. Urol.* 2017, 72, 1028–1029. [CrossRef]
- 142. Day, S.E.; Kettunen, M.I.; Cherukuri, M.K.; Mitchell, J.B.; Lizak, M.J.; Morris, H.D.; Matsumoto, S.; Koretsky, A.P.; Brindle, K.M. Detecting response of rat C6 glioma tumors to radiotherapy using hyperpolarized [1-¹³C]pyruvate and ¹³C magnetic resonance spectroscopic imaging. *Magn. Reson. Med.* 2011, 65, 557–563. [CrossRef] [PubMed]
- 143. Gallagher, F.A.; Kettunen, M.I.; Hu, D.E.; Jensen, P.R.; Zandt, R.I.; Karlsson, M.; Gisselsson, A.; Nelson, S.K.; Witney, T.H.; Bohndiek, S.E.; et al. Production of hyperpolarized [1,4-¹³C₂]malate from [1,4-¹³C₂]fumarate is a marker of cell necrosis and treatment response in tumors. *Proc. Natl. Acad. Sci. USA* 2009, 106, 19801–19806. [CrossRef] [PubMed]

- 144. Duwel, S.; Durst, M.; Gringeri, C.V.; Kosanke, Y.; Gross, C.; Janich, M.A.; Haase, A.; Glaser, S.J.; Schwaiger, M.; Schulte, R.F.; et al. Multiparametric human hepatocellular carcinoma characterization and therapy response evaluation by hyperpolarized ¹³C MRSI. *NMR Biomed.* **2016**, *29*, 952–960. [CrossRef] [PubMed]
- 145. Bollineni, V.R.; Kramer, G.M.; Jansma, E.P.; Liu, Y.; Oyen, W.J. A systematic review on [¹⁸F]FLT-PET uptake as a measure of treatment response in cancer patients. *Eur. J. Cancer* **2016**, *55*, 81–97. [CrossRef] [PubMed]
- 146. Sanghera, B.; Wong, W.L.; Sonoda, L.I.; Beynon, G.; Makris, A.; Woolf, D.; Ardeshna, K. FLT PET-CT in evaluation of treatment response. *Indian J. Nucl. Med.* **2014**, *29*, 65–73. [CrossRef] [PubMed]
- 147. Kishino, T.; Hoshikawa, H.; Nishiyama, Y.; Yamamoto, Y.; Mori, N. Usefulness of 3'-deoxy-3'-18F-fluorothymidine PET for predicting early response to chemoradiotherapy in head and neck cancer. *J. Nucl. Med.* 2012, 53, 1521–1527. [CrossRef] [PubMed]
- 148. Dehdashti, F.; Grigsby, P.W.; Myerson, R.J.; Nalbantoglu, I.; Ma, C.; Siegel, B.A. Positron emission tomography with [¹⁸F]-3'-deoxy-3'fluorothymidine (FLT) as a predictor of outcome in patients with locally advanced resectable rectal cancer: A pilot study. *Mol. Imaging Biol.* **2013**, *15*, 106–113. [CrossRef]
- 149. Shah, R.; Vattoth, S.; Jacob, R.; Manzil, F.F.; O'Malley, J.P.; Borghei, P.; Patel, B.N.; Cure, J.K. Radiation necrosis in the brain: Imaging features and differentiation from tumor recurrence. *Radiographics* 2012, *32*, 1343–1359. [CrossRef]
- 150. Leung, K. L-[methyl-11C[Methionine. In *Molecular Imaging and Contrast Agent Database (MICAD)*; National Center for Biotechnology Information: Bethesda, MD, USA, 2005.
- 151. Zhao, C.; Zhang, Y.; Wang, J. A meta-analysis on the diagnostic performance of ¹⁸F-FDG and ¹¹C-methionine PET for differentiating brain tumors. *Am. J. Neuroradiol.* **2014**, *35*, 1058–1065. [CrossRef]
- 152. Sharma, R.; D'Souza, M.; Jaimini, A.; Hazari, P.P.; Saw, S.; Pandey, S.; Singh, D.; Solanki, Y.; Kumar, N.; Mishra, A.K.; et al. A comparison study of ¹¹C-methionine and ¹⁸F-fluorodeoxyglucose positron emission tomography-computed tomography scans in evaluation of patients with recurrent brain tumors. *Indian J. Nucl. Med.* **2016**, *31*, 93–102. [CrossRef]
- Langen, K.J.; Hamacher, K.; Weckesser, M.; Floeth, F.; Stoffels, G.; Bauer, D.; Coenen, H.H.; Pauleit, D. O-(2-[¹⁸F]fluoroethyl)-L-tyrosine: Uptake mechanisms and clinical applications. *Nucl. Med. Biol.* 2006, 33, 287–294. [CrossRef] [PubMed]
- 154. Dunet, V.; Rossier, C.; Buck, A.; Stupp, R.; Prior, J.O. Performance of ¹⁸F-fluoro-ethyl-tyrosine (¹⁸F-FET) PET for the differential diagnosis of primary brain tumor: A systematic review and Metaanalysis. *J. Nucl. Med.* **2012**, *53*, 207–214. [CrossRef] [PubMed]
- 155. Galldiks, N.; Langen, K.J.; Holy, R.; Pinkawa, M.; Stoffels, G.; Nolte, K.W.; Kaiser, H.J.; Filss, C.P.; Fink, G.R.; Coenen, H.H.; et al. Assessment of treatment response in patients with glioblastoma using O-(2-¹⁸F-fluoroethyl)-L-tyrosine PET in comparison to MRI. *J. Nucl. Med.* **2012**, *53*, 1048–1057. [CrossRef] [PubMed]
- 156. Takenaka, S.; Asano, Y.; Shinoda, J.; Nomura, Y.; Yonezawa, S.; Miwa, K.; Yano, H.; Iwama, T. Comparison of ¹¹C-methionine, ¹¹C-choline, and ¹⁸F-fluorodeoxyglucose-PET for distinguishing glioma recurrence from radiation necrosis. *Neurol. Med. Chir.* **2014**, *54*, 280–289. [CrossRef]
- 157. Calabria, F.F.; Barbarisi, M.; Gangemi, V.; Grillea, G.; Cascini, G.L. Molecular imaging of brain tumors with radiolabeled choline PET. *Neurosurg. Rev.* **2018**, *41*, 67–76. [CrossRef] [PubMed]
- 158. Cicone, F.; Minniti, G.; Romano, A.; Papa, A.; Scaringi, C.; Tavanti, F.; Bozzao, A.; Maurizi Enrici, R.; Scopinaro, F. Accuracy of F-DOPA PET and perfusion-MRI for differentiating radionecrotic from progressive brain metastases after radiosurgery. *Eur. J. Nucl. Med. Mol. Imaging* **2015**, *42*, 103–111. [CrossRef] [PubMed]
- 159. Lapela, M.; Eigtved, A.; Jyrkkio, S.; Grenman, R.; Kurki, T.; Lindholm, P.; Nuutinen, J.; Sutinen, E.; Solin, O.; Bjornskov, I.; et al. Experience in qualitative and quantitative FDG PET in follow-up of patients with suspected recurrence from head and neck cancer. *Eur. J. Cancer* 2000, *36*, 858–867. [CrossRef]
- 160. O'Connor, J.P.; Aboagye, E.O.; Adams, J.E.; Aerts, H.J.; Barrington, S.F.; Beer, A.J.; Boellaard, R.; Bohndiek, S.E.; Brady, M.; Brown, G.; et al. Imaging biomarker roadmap for cancer studies. *Nat. Rev. Clin. Oncol.* 2017, 14, 169–186. [CrossRef]



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