



T618I-Mutated Colony Stimulating Factor 3 Receptor in Chronic Neutrophilic Leukemia and Chronic Myelomonocytic Leukemia Patients who Underwent Allogeneic Stem Cell Transplantation

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Dear Editor

Recently, the mutations within the colony-stimulating factor 3 receptor gene (*CSF3R*) have been reported as a specific marker of chronic neutrophilic leukemia (CNL) and atypical CML (aCML) [1, 2]. The current WHO system classifies CNL as a *BCR-ABL1*-negative myeloproliferative neoplasm (MPN). However, in routine clinical practice, it is difficult to clearly distinguish CNL from aCML, chronic myelomonocytic leukemia (CMML), juvenile myelomonocytic leukemia, and MDS/MPN, unclassifiable under the MDS/MPN umbrella [3]. Therefore, molecular characteristics (e.g., *CSF3R* mutation) need to be included in the WHO diagnostic criteria. Here, we describe two *CSF3R* T618I-mutated patients with CNL and CMML, respectively, who underwent allogeneic stem cell transplantation (allo-SCT).

A 40-yr-old man presented in 2012 with marked neutrophilia. His total white blood cell count was $77.24 \times 10^9/L$ with a differential of 80% neutrophils, 3% band forms, 3% metamyelocytes, 1% myelocytes, 1% promyelocytes, 1% blasts, 3% lymphocytes, and 10% monocytes (Fig. 1A). He had a 23-cm splenomegaly. His bone marrow (BM) aspirate and biopsy showed hypercellu-

larity with granulocytic hyperplasia (Fig. 1B), with a normal karyotype and no evidence of *BCR-ABL1*, *PDGFRA*, or *PDGFRB* rearrangement when examined by FISH. Molecular studies demonstrated the absence of *JAK2* V617F and *BCR-ABL1* transcripts. CNL was diagnosed in accordance with the WHO diagnostic criteria, and the patient was treated with hydroxyurea. However, his neutrophilia persisted in the peripheral blood as anemia and thrombocytopenia developed. Repeated BM examinations showed hypercellular BM with granulocytic hyperplasia and decreased megakaryocytes, and a clonal chromosomal abnormality of 46,XY,der(18:21)(q10;q10),+21[7]/47,+21[5]/46,XY[8]. The *CSF3R* T618I mutation was detected (Fig. 2A). He then underwent myeloablative allo-SCT, using cells from an unrelated donor. At day 35 following the allo-SCT, BM examination showed 70% cellularity with 100% donor chimerism. The T618I mutation was not detected in the BM aspirates through sequencing (Fig. 2C).

A 60-yr-old man who presented with recurrent purpura on his extremities was referred to our hospital. He had constitutional symptoms including fatigue and weight loss. A computed tomog-

Received: September 29, 2014

Revision received: October 9, 2014

Accepted: January 28, 2015

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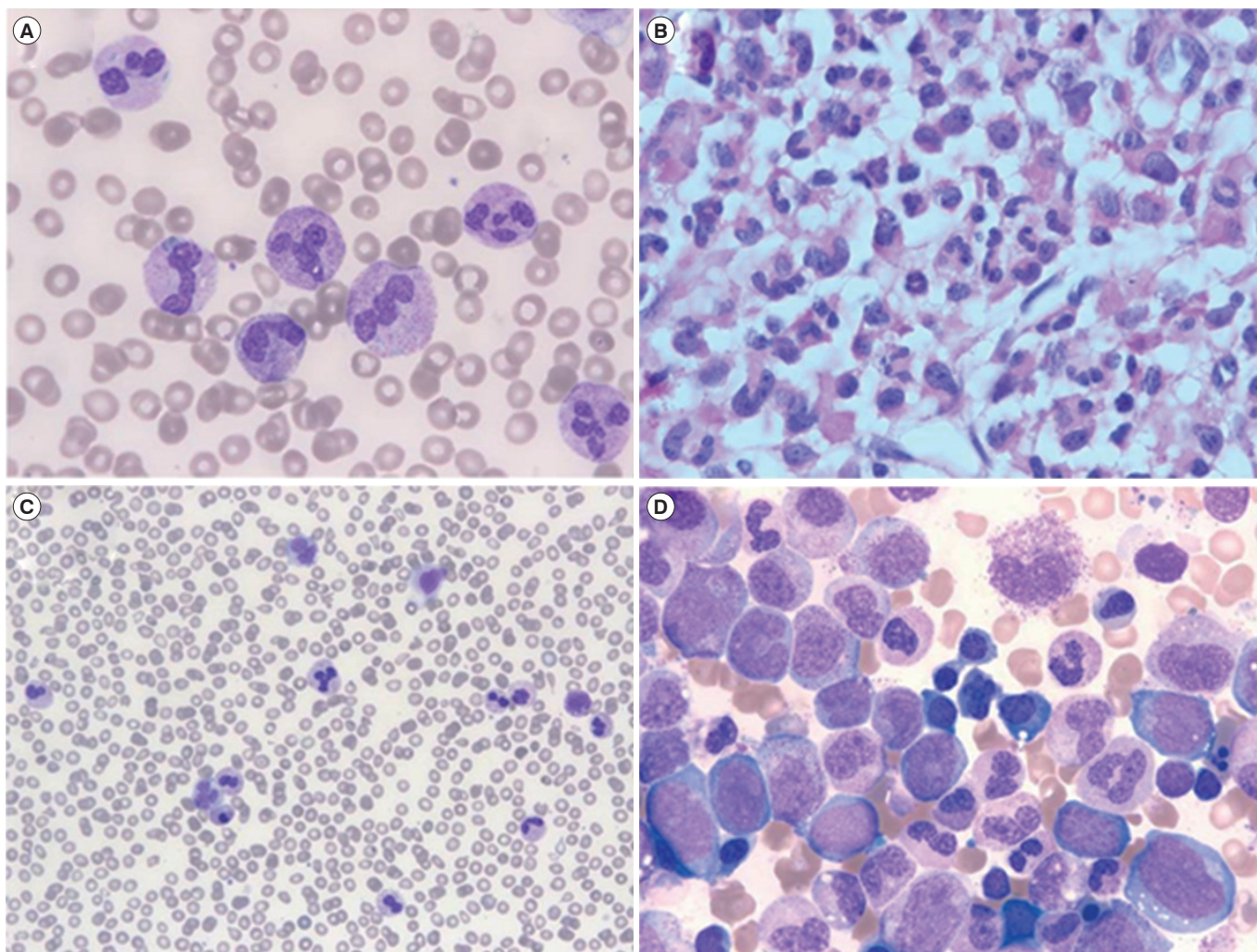


Fig. 1. Chronic neutrophilic leukemia; (A) peripheral blood showing neutrophilia with toxic granulation (Wright-Giemsa staining, $\times 1,000$) and (B) bone marrow biopsy showing a markedly elevated myeloid:erythroid ratio. Chronic myelomonocytic leukemia (Hematoxylin and eosin staining, $\times 1,000$); (C) peripheral blood with monocytosis and neutrophilia (Wright-Giemsa staining, $\times 400$) and (D) bone marrow aspirate with erythroid and granulocytic dysplasia (Wright-Giemsa staining, $\times 1,000$).

raphy scan of the abdomen demonstrated 13.5-cm mild splenomegaly. His peripheral blood (Fig. 1C) and BM findings (Fig. 1D) were consistent with CMML-1. The patient had a normal karyotype with no evidence of *BCR-ABL1*, *PDGFRA*, or *PDGFRB* rearrangement following FISH examinations. Molecular studies demonstrated the absence of *JAK2 V617F* and *BCR-ABL1* transcripts. The *CSF3R* T618I mutation was identified through direct sequencing (Fig. 2B). An initial azacitidine treatment failed to achieve any response. Thus, he underwent allo-SCT using cells from a sibling donor. At day 30 following the allo-SCT, a BM examination showed 30% cellularity with 97% donor chimerism. The *CSF3R* T618I mutation was not detected in the BM aspirates (Fig. 2D).

Since high-frequency of *CSF3R* mutations in CNL (89%) and, to a lesser extent, in aCML (40%) were discovered [1, 4], Pardanani *et al.* [2] has confirmed that the *CSF3R* T618I mutation was detected exclusively in cases of WHO-defined CNL, with a mutational frequency of 83%. Thus, *CSF3R* T618I mutation should be a diagnostic criterion for CNL.

Previous studies showed discordant frequencies in the *CSF3R* mutations in CMML and aCML. Maxson *et al.* [1] reported that eight of 20 aCML (40%) cases exhibited *CSF3R* mutations, whereas Pardanani *et al.* [2] did not find the *CSF3R* mutation in any case of aCML. Although *CSF3R* mutations have been observed in 4% of patients with CMML, they are distinct from membrane proximal mutations; the T618I mutation was identi-

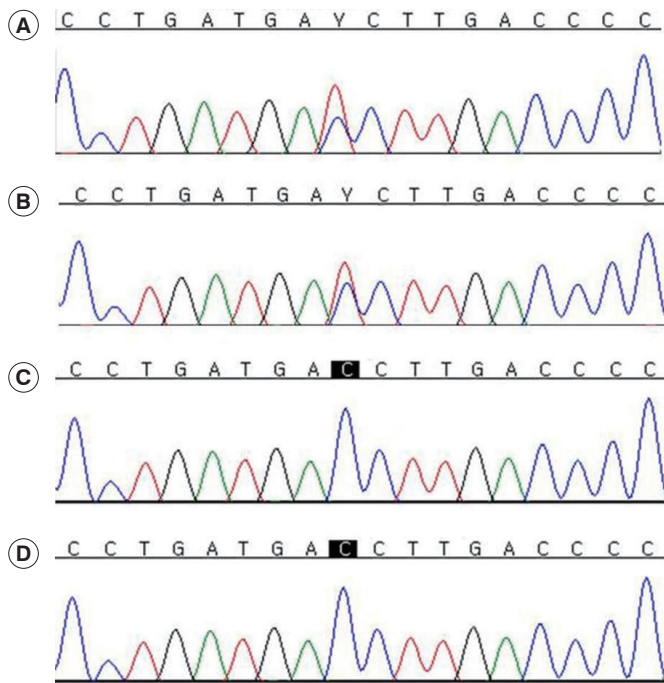


Fig. 2. Sequencing of *genomic* DNA isolated from peripheral blood leukocytes, demonstrating the *CSF3R* c.1853C>T (p.Thr618Ile) mutation. The *CSF3R* T618I mutation was detected in patient #1 with CNL, pre-allo-SCT (A) and in patient #2 with CMML at diagnosis (B). After allo-SCT, the mutation was not detected in patient #1 (C) and in patient #2 (D).

Abbreviations: CNL, chronic neutrophilic leukemia; allo-SCT, allogeneic stem cell transplantation; CMML, chronic myelomonocytic leukemia.

fied in a majority of patients with CNL [5]. However, we observed one CMML patient harboring the *CSF3R* T618I mutation. To incorporate *CSF3R* mutations into the diagnostic criteria of these disorders, the frequency, the location, and the specificity of the *CSF3R* mutations need to be determined.

Although there is no current standard of care for CNL or for aCML, allo-SCT may be applicable to young patients with potential for blast transformation and progressive refractory neutrophilia [6-9]. In our two CNL and CMML patients harboring the *CSF3R* T618I mutation, and who had undergone allo-SCT, the *CSF3R* T618I mutation was not detected following allo-SCT.

This suggests a predictive role of this mutation in post-transplant relapse, supported by prior studies showing a correlation between post-transplant relapse and the persistence of the *CSF3R* T618I in aCML [10]. Thus, testing for the *CSF3R* mutation may lead to genetically informed therapy and useful diag-

nostic approach. The influence of the *CSF3R* mutations on genotype-phenotype associations, disease prognosis, and the efficacy of the therapeutic inhibition of *CSF3R*-related signaling needs to be clarified.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

Acknowledgments

The present study was supported by a grant from the Korea Health Technology R&D Project, Ministry of Health & Welfare, Republic of Korea (grant no. A120175).

REFERENCES

1. Maxson JE, Gotlib J, Pollyea DA, Fleischman AG, Agarwal A, Eide CA, et al. Oncogenic *CSF3R* mutations in chronic neutrophilic leukemia and atypical CML. *N Engl J Med* 2013;368:1781-90.
2. Pardanani A, Lasho TL, Laborde RR, Elliott M, Hanson CA, Knudson RA, et al. *CSF3R* T618I is a highly prevalent and specific mutation in chronic neutrophilic leukemia. *Leukemia* 2013;27:1870-3.
3. Tefferi A, Thiele J, Vannucchi AM, Barbui T. An overview on *CALR* and *CSF3R* mutations and a proposal for revision of WHO diagnostic criteria for myeloproliferative neoplasms. *Leukemia* 2014;28:1407-13.
4. Gotlib J, Maxson JE, George TI, Tyner JW. The new genetics of chronic neutrophilic leukemia and atypical CML: implications for diagnosis and treatment. *Blood* 2013;122:1707-11.
5. Kosmider O, Itzykson R, Chesnais V, Lasho T, Laborde R, Knudson R, et al. Mutation of the colony-stimulating factor-3 receptor gene is a rare event with poor prognosis in chronic myelomonocytic leukemia. *Leukemia* 2013;27:1946-9.
6. Reilly JT. Chronic neutrophilic leukaemia: a distinct clinical entity? *Br J Haematol* 2002;116:10-8.
7. Breccia M, Biondo F, Latagliata R, Carosino I, Mandelli F, Alimena G. Identification of risk factors in atypical chronic myeloid leukemia. *Haematologica* 2006;91:1566-8.
8. Elliott MA, Hanson CA, Dewald GW, Smoley SA, Lasho TL, Tefferi A. WHO-defined chronic neutrophilic leukemia: a long-term analysis of 12 cases and a critical review of the literature. *Leukemia* 2005;19:313-7.
9. Hernández JM, del Cañizo MC, Cuneo A, García JL, Gutiérrez NC, González M, et al. Clinical, hematological and cytogenetic characteristics of atypical chronic myeloid leukemia. *Ann Oncol* 2000;11:441-4.
10. Langabeer SE, McCarron SL, Haslam K, O'Donovan MT, Conneally E. The *CSF3R* T618I mutation as a disease-specific marker of atypical CML post allo-SCT. *Bone Marrow Transplant* 2014;49:843-4.