

# Longitudinal Dopamine D2 Receptor Changes and Cerebrovascular Health in Aging

Nina Karalija, PhD, Jarkko Johansson, PhD, Goran Papenberg, PhD, Anders Wåhlin, PhD, Alireza Salami, PhD, Ylva Köhncke, PhD, Andreas M. Brandmaier, PhD, Micael Andersson, MSc, Jan Axelsson, PhD, Katrine Riklund, MD, PhD, Martin Lövdén, PhD, Ulman Lindenberger, PhD, Lars Bäckman, PhD, and Lars Nyberg, PhD

## Correspondence

Dr. Karalija  
nina.karalija@umu.se

*Neurology*® 2022;99:e1278-e1289. doi:10.1212/WNL.0000000000200891

## Abstract

### Background and Objectives

Cross-sectional studies suggest marked dopamine (DA) decline in aging, but longitudinal evidence is lacking. The aim of this study was to estimate within-person decline rates for DA D2-like receptors (DRD2) in aging and examine factors that may contribute to individual differences in DRD2 decline rates.

### Methods

We investigated 5-year within-person changes in DRD2 availability in a sample of older adults. At both occasions, PET with <sup>11</sup>C-raclopride and MRI were used to measure DRD2 availability in conjunction with structural and vascular brain integrity.

### Results

Longitudinal analyses of the sample (baseline: n = 181, ages: 64–68 years, 100 men and 81 women; 5-year follow-up: n = 129, 69 men and 60 women) revealed aging-related striatal and extrastriatal DRD2 decline, along with marked individual differences in rates of change. Notably, the magnitude of striatal DRD2 decline was ~50% of past cross-sectional estimates, suggesting that the DRD2 decline rate has been overestimated in past cross-sectional studies. Significant DRD2 reductions were also observed in select extrastriatal regions, including hippocampus, orbitofrontal cortex (OFC), and anterior cingulate cortex (ACC). Distinct profiles of correlated DRD2 changes were found across several associative regions (ACC, dorsal striatum, and hippocampus) and in the reward circuit (nucleus accumbens and OFC). DRD2 losses in associative regions were associated with white matter lesion progression, whereas DRD2 losses in limbic regions were related to reduced cortical perfusion.

### Discussion

These findings provide the first longitudinal evidence for individual and region-specific differences of DRD2 decline in older age and support the hypothesis that cerebrovascular factors are linked to age-related dopaminergic decline.

---

From the Departments of Radiation Sciences, Diagnostic Radiology (N.K., J.J., K.R., L.N.) and Radiation Physics (A.W., J.A.), Department of Applied Physics and Electronics (A.W.), and Umeå Center for Functional Brain Imaging (UFBI) (N.K., J.J., A.W., A.S., M.A., J.A., K.R., L.N.), Umeå University; Aging Research Center (G.P., A.S., L.B.), Karolinska Institutet & Stockholm University; Department of Integrative Medical Biology (A.S., M.A., L.N.), and Wallenberg Center for Molecular Medicine (A.S., L.N.), Umeå University, Sweden; Center for Lifespan Psychology (Y.K., A.M.B., U.L.), Max Planck Institute for Human Development; Max Planck UCL Centre for Computational Psychiatry and Ageing Research (A.M.B., U.L.), Berlin, Germany and London, UK; and Department of Psychology (M.L.), University of Gothenburg, Sweden.

Go to [Neurology.org/N](https://www.neurology.org/N) for full disclosures. Funding information and disclosures deemed relevant by the authors, if any, are provided at the end of the article.

The Article Processing Charge was funded by the authors.

This is an open access article distributed under the terms of the Creative Commons Attribution License 4.0 (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Glossary

**ACC** = anterior cingulate cortex; **ANOVA** = analysis of variance; **BMI** = body mass index; **BP<sub>ND</sub>** = DRD2 binding potential; **COBRA** = Cognition, Brain, and Aging; **DA** = dopamine; **DRD2** = DA D2-like receptor; **GM** = gray matter; **MMSE** = Mini-Mental State Examination; **MRTM** = multilinear reference tissue model; **OFC** = orbitofrontal cortex; **PCA** = principal component analysis; **pcASL** = pseudocontinuous arterial spin labeling; **PVE** = partial volume effect; **ROI** = region of interest; **WM** = white matter.

Neurocognitive impairment compromises health for many older adults. Owing to a globally aging population, efforts are directed at identifying modifiable brain changes within key pathologic aging routes.<sup>1,2</sup> In this context, the age sensitivity of the dopamine (DA) pathways has been highlighted in various studies.<sup>3</sup> Cross-sectional studies suggest that the availability of DA receptors and transporters is reduced by 8%–14% per decade across the lifespan.<sup>4</sup> Most studies assessed DA D2-like receptor (DRD2) availability because of their central role for cognition and in neurodegenerative conditions.<sup>3,5</sup>

A major shortcoming is that most human *in vivo* DA studies to date relied on cross-sectional designs with small samples (median: 21 participants).<sup>4</sup> Cohort effects in cross-sectional studies can result in overestimation or underestimation of the extent of within-person brain and behavioral changes.<sup>6–8</sup> Accordingly, findings from cross-sectional studies may diverge from longitudinal results in patterns of brain-behavior change-change associations.<sup>9</sup> The few existing longitudinal DRD2 studies were conducted with a few days or months between sessions and revealed little change.<sup>10,11</sup> Longitudinal assessments over longer periods are key to understand the magnitude of individual differences in DA losses and to delineate factors that contribute to DA losses in aging.<sup>12</sup>

Vascular changes occur early in the adult lifespan, are part of dementia pathologies,<sup>13,14</sup> and may modulate DA integrity. Ischemic strokes are followed by DA changes,<sup>15</sup> and Parkinson disease is associated with an elevated risk of cardiovascular disease.<sup>16</sup> Associations among markers of cerebral small vessel disease and DA integrity have also been observed in healthy older samples.<sup>17</sup> Hence, maintained vascular health may constitute one predictor of preserved DA integrity in aging, but this relation has not been probed using longitudinal data.

We present the first long-term longitudinal investigation of aging-related DRD2 changes, with data from the Cognition, Brain, and Aging (COBRA) study.<sup>18</sup> In COBRA, healthy adults (>60 years) have undergone PET with <sup>11</sup>C-raclopride, MRI, and assessment of health, lifestyle, and cognition at 2 occasions separated by 5 years. The relatively large sample size (*n* = 181 and 129 at baseline and follow-up) permits analyses of trends for DRD2 change across regions, individual differences in change, and associations between DRD2 and other brain changes.

First, we assessed longitudinal rates of DRD2 change to corroborate findings of aging-related DRD2 differences in cross-

sectional studies (~8% per decade in the striatum<sup>4</sup>). Second, we investigated whether the regional DRD2 decline rates correlated within striatal compartments and across the mesolimbic and mesocortical DA pathways.<sup>19,20</sup> Finally, we tested whether individual differences in DRD2 decline were related to reduced cerebrovascular health, using white matter (WM) lesion progression and reductions in cerebral perfusion as indicators of vascular health.<sup>17</sup> We expected the vascular-DRD2 link to be strongest for the hippocampus and basal ganglia because of these regions' sensitivity to vascular insult.<sup>21,22</sup>

## Methods

### Standard Protocol Approvals, Registrations, and Patient Consents

This study was approved by the Swedish Ethical Review Authority (Umeå, Sweden; registration number: 2012-57-31M) and conducted in accordance with the Declaration of Helsinki. Written informed consent was obtained from all participants before any testing.

### Sample

The original sample consisted of 181 healthy older adults (100 men, 81 women; ages: 64–68 years, mean: 66.2 ± 1.2) randomly selected from the population registry in Umeå (in northern Sweden) who were offered participation. Eligible participants were free from disorders and conditions that can alter brain and cognitive functioning. Exclusion criteria were neurologic and psychiatric disorders, epilepsy, previous brain trauma, intellectual disability, a Mini-Mental State Examination (MMSE) score of <27, structural brain abnormalities (inspection performed by neuroradiologists), cancer, diabetes, severe auditory and visual impairments, claustrophobia, and metal implants. The assessment battery included brain imaging with <sup>11</sup>C-raclopride-PET and MRI, cognitive testing, and collection of demographics, health, and lifestyle data and blood samples. The baseline sample, assessment battery, and exclusion criteria, and details for the a priori statistical power analyses, have been described previously.<sup>18</sup> The statistical power analyses revealed that for a sample size of 180, a 5-year separation between test waves, and an estimated attrition rate of 20%, the expected power to detect significant individual differences in change over 5 years was 83% for the cognitive measures (which were deemed to have lower reliability than the brain measures). The attrition rate was based on longitudinal data for 60–70-year-old adults from a separate study.<sup>8</sup> All participants were offered to undergo the

assessments for a second time at the 5-year follow-up, when 129 returned (69 men, 60 women; ages: 69–73 years, mean:  $71.2 \pm 1.2$ ). The duration between test waves was  $60.1 \pm 0.6$  months for PET and  $60.0 \pm 0.4$  months for MRI. The scheduled time between MRI and PET was 2 days, but technical issues, traveling, and disease caused longer duration between sessions for some (baseline: mean =  $3.9 \pm 5.7$  days, max: 44 days; follow-up: mean =  $3.8 \pm 7.6$  days, max: 72 days).

## Brain Imaging

MRI was performed with a 3 tesla Discovery MR 750 scanner (General Electric, Milwaukee, WI), and PET data were acquired with a Discovery PET/CT 690 (General Electric) at both time points.

## Regional Volumes

T1-weighted images were obtained with echo time 3.2 milliseconds, flip angle  $12^\circ$ , repetition time 8.19 milliseconds, 176 slices with thickness 1.0 mm, and field of view 25.0 cm with resolution 0.98 mm upsampled to 0.49 mm. The longitudinal image processing pipeline in FreeSurfer, version 6.0, was used to process T1-weighted images and derive estimates of gray matter (GM), WM, and lateral ventricle size. Subcortical GM segmentations and cortical parcellations<sup>23</sup> (see eMethods, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)) were used to define regions of interest (ROIs) for DRD2 and perfusion assessment.

## DRD2 Availability

A 55-minute, 18-frame dynamic PET scan was acquired during rest after IV bolus injection of approximately 250 MBq <sup>11</sup>C-raclopride (baseline:  $263.5 \pm 19.0$  MBq; follow-up:  $260.2 \pm 15.0$  MBq;  $t(123) = 1.8$ ;  $p = 0.076$ ). The range for the injected mass of raclopride was larger at baseline (0.17–9.48  $\mu\text{g}$ ) than at follow-up (0.11 and 2.67  $\mu\text{g}$ ;  $t(124) = 4.6$ ;  $p < 0.001$ ). An attenuation CT scan (20 mA, 120 kV, 0.8 seconds/revolution) preceded ligand injection. Attenuation- and decay-corrected images (47 slices, field of view = 25 cm,  $256 \times 256$ -pixel transaxial images, voxel size =  $0.977 \times 0.977 \times 3.27$  mm<sup>3</sup>) were reconstructed with the iterative algorithm VUE Point HD-SharpIR (GE; 6 iterations, 24 subsets, 3.0 mm postfiltering; full width at half maximum: 3.2 mm). PET images were motion corrected and coregistered with the structural T1-weighted images from the corresponding session (baseline and follow-up) using the Statistical Parametric Mapping software (SPM12). As a source for coregistration, the mean of the first 5 frames was used. PET images from both time points were coregistered with the baseline T1 image for 3 participants (no MRI at follow-up). Two individuals declined to undergo PET at follow-up.

DRD2 binding potential ( $\text{BP}_{\text{ND}}$ ) was estimated with and without correction for partial volume effects (PVEs). Regional PVE correction was conducted using the symmetric geometric transfer matrix implemented in FreeSurfer.<sup>24</sup> An incremental PVE-correction approach was used in which (1) the initial correction was achieved using resolution modeling in the iterative image reconstruction procedure (SHARP-IR),

and (2) the remnant PVE was controlled for using the ROI-based geometric transfer matrix approach. The size of the secondary correction kernel was estimated empirically (point spread function of 2.5 mm; isotropic) to achieve a similar level of correction as earlier.<sup>25</sup> FreeSurfer segmentations and pre-processed PET data were used to estimate PVE-corrected regional radioactivity concentrations per ROI and time frame. PVE-corrected  $\text{BP}_{\text{ND}}$  estimates were calculated with the multilinear reference tissue model (MRTM) on dynamic PVE-corrected data, with cerebellar GM radioactivity as an indirect input function.

<sup>11</sup>C-raclopride  $\text{BP}_{\text{ND}}$  values without PVE correction were estimated using Logan analysis<sup>26</sup> from time activity curves within T1-segmented ROIs (median of ROI voxel values from time frames between 18 and 55 minutes). The cerebellar GM served as the reference area. The 2 pipelines (MRTM vs Logan) allowed for comparisons of the robustness of DRD2 decline patterns (Table 1). The analyses that appear in Results were conducted with PVE-corrected data. Notably, analyses were replicated with  $\text{BP}_{\text{ND}}$  values without PVE correction, including assessments of change over time (Table 1, and for a more extensive set of regions in eTable 2, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)), associations for DRD2 changes across regions (eTable 1), and DRD2-GM volume associations (eTable 3).

## Perfusion

Three-dimensional pseudocontinuous arterial spin labeling (pcASL) was acquired with background suppression and spiral readout. Labeling time = 1.5 seconds, postlabeling delay time = 1.5 seconds, field of view = 24 cm, slice thickness = 4 mm, and acquisition resolution =  $8 \times 512$  (arms  $\times$  data points), with the number of averages set at 3. The total scanning time was approximately 5 minutes. This sequence provided whole-brain perfusion in mL/100 g/min. The reconstructed voxel size was  $1.875 \times 1.875 \times 4$  mm<sup>3</sup>. Quantitative perfusion maps were calculated using a postprocessing tool installed on the scanner by the manufacturer. Mean GM perfusion was computed for FreeSurfer-segmented ROIs as the average of the individual perfusion estimates weighted by volume.

## WM Lesions

WM hyperintensities were segmented from a fluid-attenuated inversion recovery image (48 slices, slice thickness = 3 mm, TE = 120 milliseconds, TR = 8,000 milliseconds, and field of view =  $24 \times 24$  cm) with the lesion growth algorithm,<sup>27</sup> as implemented in the LST toolbox version 2.0.14 for SPM12<sup>28</sup> (eMethods, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)).

## Cognition

Participants performed the MMSE, in which a minimum of 27 points (max: 30 points) was required for inclusion at baseline. A digit-symbol coding subtest from the Wechsler Adult Intelligence Scale was also administered in which participants received 1 point per correct item coding (duration: 90 seconds). Semantic knowledge was assessed with a vocabulary test. For each word, 1 correct synonym of 5 possible

**Table 1** Descriptive Data for Baseline and Follow-up Along With 5-Year Brain Changes

	Baseline	Follow-up	Change	<i>r</i>
<b>DRD2 availability</b>	Mean	Mean	Mean %	
<b>Putamen</b>	<b>3.74 (0.36)</b>	<b>3.68 (0.38)</b>	<b>-2.1 (5.2)**</b>	<b>0.84</b>
Without PVC	3.35 (0.26)	3.29 (0.30)	-1.8 (5.1)***	0.84
<b>Caudate</b>	<b>2.67 (0.38)</b>	<b>2.63 (0.41)</b>	<b>-2.5 (10.4)*</b>	<b>0.71</b>
Without PVC	2.20 (0.28)	2.11 (0.33)	-4.7 (9.2)***	0.77
<b>Nucleus accumbens</b>	<b>2.72 (0.40)</b>	<b>2.60 (0.41)</b>	<b>-5.4 (11.6)***</b>	<b>0.54</b>
Without PVC	2.17 (0.29)	2.09 (0.27)	-3.3 (11.2)**	0.58
<b>Hippocampus</b>	<b>0.25 (0.05)</b>	<b>0.23 (0.06)</b>	<b>-7.5 (17.1)***</b>	<b>0.72</b>
Without PVC	0.27 (0.05)	0.25 (0.05)	-9.1 (13.3)***	0.79
<b>ACC</b>	<b>0.26 (0.07)</b>	<b>0.24 (0.07)</b>	<b>-8.4 (19.0)***</b>	<b>0.79</b>
Without PVC	0.25 (0.05)	0.23 (0.05)	-7.3 (13.9)***	0.82
<b>OFC</b>	<b>0.33 (0.06)</b>	<b>0.31 (0.09)</b>	<b>-5.9 (22.4)***</b>	<b>0.56</b>
Without PVC	0.26 (0.04)	0.25 (0.04)	-4.6 (13.5)***	0.62
<b>Frontal cortex</b>	<b>0.18 (0.06)</b>	<b>0.18 (0.06)</b>	<b>-1.2 (30.8)</b>	<b>0.52</b>
Without PVC	0.19 (0.04)	0.18 (0.04)	-2.1 (15.7)*	0.80
<b>Temporal cortex</b>	<b>0.32 (0.04)</b>	<b>0.32 (0.04)</b>	<b>-0.2 (8.7)</b>	<b>0.75</b>
Without PVC	0.29 (0.03)	0.28 (0.04)	-1.2 (8.1)	0.81
<b>WM lesions</b>	Sum	Sum	Mean difference	
<b>Total volume (cm<sup>3</sup>)</b>	2.0 (1.7)	3.7 (3.6)	+1.4 (1.4)***	0.93
<b>No. of lesions</b>	27.8 (13.6)	33.9 (16.2)	+4.1 (6.6)***	0.88
<b>Perfusion (mL/100 g/min)</b>	Mean	Mean	Mean %	
<b>Basal ganglia</b>	44.3 (6.5)	41.4 (6.8)	-6.5 (13.2)***	0.59
<b>Hippocampus</b>	40.9 (7.4)	37.6 (8.0)	-9.1 (13.1)***	0.70
<b>Frontal cortex</b>	44.7 (10.4)	41.1 (11.1)	-8.0 (19.0)***	0.61
<b>Temporal cortex</b>	38.7 (8.3)	35.4 (8.8)	-9.0 (15.9)***	0.72
<b>Mean GM</b>	42.0 (8.8)	38.5 (9.3)	-9.1 (15.1)***	0.69
<b>Volumes (cm<sup>3</sup>)</b>	Sum	Sum	Mean %	
<b>Basal ganglia</b>	21.2 (2.4)	20.9 (2.4)	-1.4 (2.5)***	0.97
<b>Hippocampus</b>	7.8 (0.8)	7.6 (0.8)	-2.7 (3.6)***	0.94
<b>Frontal cortex</b>	106.7 (11.3)	105.4 (11.3)	-1.3 (1.8)***	0.99
<b>Temporal cortex</b>	105.5 (10.5)	103.4 (10.6)	-2.1 (2.4)***	0.97
<b>WM</b>	440.0 (55.1)	426.8 (54.6)	-3.1 (1.3)***	0.99
<b>Lateral ventricles</b>	30.0 (13.0)	35.2 (15.5)	+18.7 (9.9)***	0.98

Abbreviations: ACC = anterior cingulate cortex; GM = gray matter; OFC = orbitofrontal cortex; *r* = Pearson's correlation coefficient; WM = white matter. Sizeable reductions were found for DRD2 availability in select brain regions and for brain-wide vascular measures. Correlations (*r*) between baseline and follow-up measures are also presented. DRD2 availability was estimated with 2 separate pipelines, with correction for partial-volume effects (PVC; bold font) or not (italics font). \**p* < 0.05, \*\**p* < 0.01, and \*\*\**p* < 0.001 for within-person comparisons of baseline vs follow-up with paired-samples *t* tests. Basal ganglia represent putamen, caudate, nucleus accumbens, and globus pallidus. The Bonferroni-adjusted  $\alpha$  level is 0.002 (0.05/29).



alternatives was to be selected (30 words; 1 point per correct answer).

## Lifestyle and Health

Hours per week spent on various social, intellectual, and physical activities were assessed through self-report questionnaires. History of known disorders, medicine intake, and nicotine consumption was documented. Blood pressure was measured in a sitting position. The body mass index (BMI) was calculated from height and weight. Ten-year cardiovascular disease risk (%) was calculated from hypertension and diabetes diagnosis, systolic blood pressure, BMI, smoking, age, and sex, according to an established model.<sup>17</sup> ApoE ε4 allele status was assessed from blood samples.<sup>29</sup>

## Statistical Analyses

Analyses were performed with SPSS (version 26) and *Ω*nyx.<sup>30,31</sup> Descriptive data are presented with mean or sum (over the left and right hemispheres for brain data), SDs, or frequencies. The significance level for all tests was 5%.

Univariate outliers were defined as >3.29 SD from the mean and excluded as pairwise deletions per ROI and modality. For WM lesions, this procedure was performed twice because of the remaining outliers after the first round of exclusions. Multivariate outliers across ROIs per brain domain (i.e., ROIs listed in eTable 2, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)) were identified according to the Mahalanobis distance ( $p < 0.001$ ) and excluded as listwise deletions in analyses of baseline ( $n = 0$  for DRD2 data;  $n = 1$  for volumes;  $n = 1$  for perfusion), follow-up ( $n = 0$  for DRD2;  $n = 2$  for volumes;  $n = 1$  for perfusion), and change ( $n = 2$  for perfusion;  $n = 7$  for DRD2; and  $n = 0$  for volumes). The effective sample for analyses ranged between 120 and 129 depending on the ROI and modality at baseline and follow-up and between 111 and 126 for estimates of change. After exclusions, values for DRD2s, volumes, and perfusion and the number of WM lesions were normally distributed (skewness: -0.73 to 0.90; kurtosis: -0.68 to 1.67) and slightly skewed for lesion volume (skewness: 1.49 to 1.86; kurtosis: 3.00 to 3.69). Accordingly, linear associations are illustrated with the Spearman's rank-order correlation ( $r_s$ ) for lesions and with Pearson's correlation coefficient ( $r$ ) for normally distributed data.

Attrition (Table 2) was assessed through the comparison of returnees and dropouts at baseline with independent sample  $t$  tests and total selectivity<sup>32</sup>:

$$\text{Selectivity} = \frac{\text{Mean}_{\text{returnees}} - \text{Mean}_{\text{parent sample}}}{\text{SD}_{\text{parent sample}}}$$

Change (%) for DRD2 and structural measures (Table 1) was calculated as:

$$\text{Change (\%)} = \frac{\text{Follow-up} - \text{Baseline}}{\text{Baseline}} \times 100$$

For lesions, change is presented as differences (follow-up minus baseline) because of large variation in baseline levels.

Paired-sample  $t$  tests (Table 1, Figure 1A) and repeated-measures analysis of variance (ANOVA) with select covariates (e.g., GM volume on DRD2 change) were conducted to assess statistical significance for brain changes over 5 years and hemispheric differences in change. Regional differences in the magnitude of DRD2 change, and DRD2 change as a function of age, sex, and education, were assessed with univariate or multivariate ANOVA and multivariate regressions. Multiple group comparisons were assessed through Bonferroni post hoc tests.

Univariate and bivariate difference score models were estimated with the *Ω*nyx software to assess individual differences in change and for tests of baseline-change and change-change associations (Figure 1B and Figure 2, B and D). Baseline data were entered for the full sample to increase the precision of the mean and variance at the first measurement occasion. In the graphical representations of these models, observed (measured) variables are represented by a rectangle, a constant by a triangle, and change by a circle. The obtained parameters include mean levels at baseline (1-sided arrow from constant to baseline), mean change (1-sided arrow from constant to change variable), individual differences in change (2-sided arrow above change variable), and standardized covariances between baseline DRD2 and change.  $Z$  values >1.96 or <-1.96 indicate statistical significance at an alpha level of  $p < 0.05$ . The models reported here are re-expressions of simple difference scores, that is, saturated models that always yield perfect fit to the data. This is why we do not report measures of goodness of fit.

Significant variance in DRD2 change was tested by a 2-df likelihood-ratio test between the original model (Figure 1B) and a corresponding null model, in which the variance for the change variable and its covariance with baseline was set to zero. The interpretation of a significant likelihood-ratio test ( $p < 0.001$ ) is that the variance in change and the covariance cannot be assumed to be zero without losing model fit, hence indicating statistically significant individual differences in rates of change.<sup>33</sup>

Regions with significant mean DRD2 change and individual differences in change (i.e., regions in Figure 1, A and B) were entered into a principal component analysis (PCA) to analyze the patterns of interregional DRD2 change (Table 3). An oblique rotation method (oblimin with Kaiser normalization) was used under the assumption that DRD2 components may be correlated.<sup>19,20</sup> Components with eigenvalues >1 are reported. With regard to the sample size, standardized loadings >0.50 are considered significant. Factor scores were exported for the DRD2 components and used as dependent variables in multivariate regression analyses, with vascular parameters as independent variables (Figure 2, A and C). The slight skewness of lesion volume data did not affect lesion-DRD2 associations, and a comparable correlation coefficient was achieved with parametric and nonparametric tests (Pearson vs Spearman; Figure 2A). The analysis of cortical vs

**Table 2** Comparisons of Cognitive, Lifestyle, Health, and Brain Measures Between Returnees, Dropouts, and the Entire Sample at Baseline

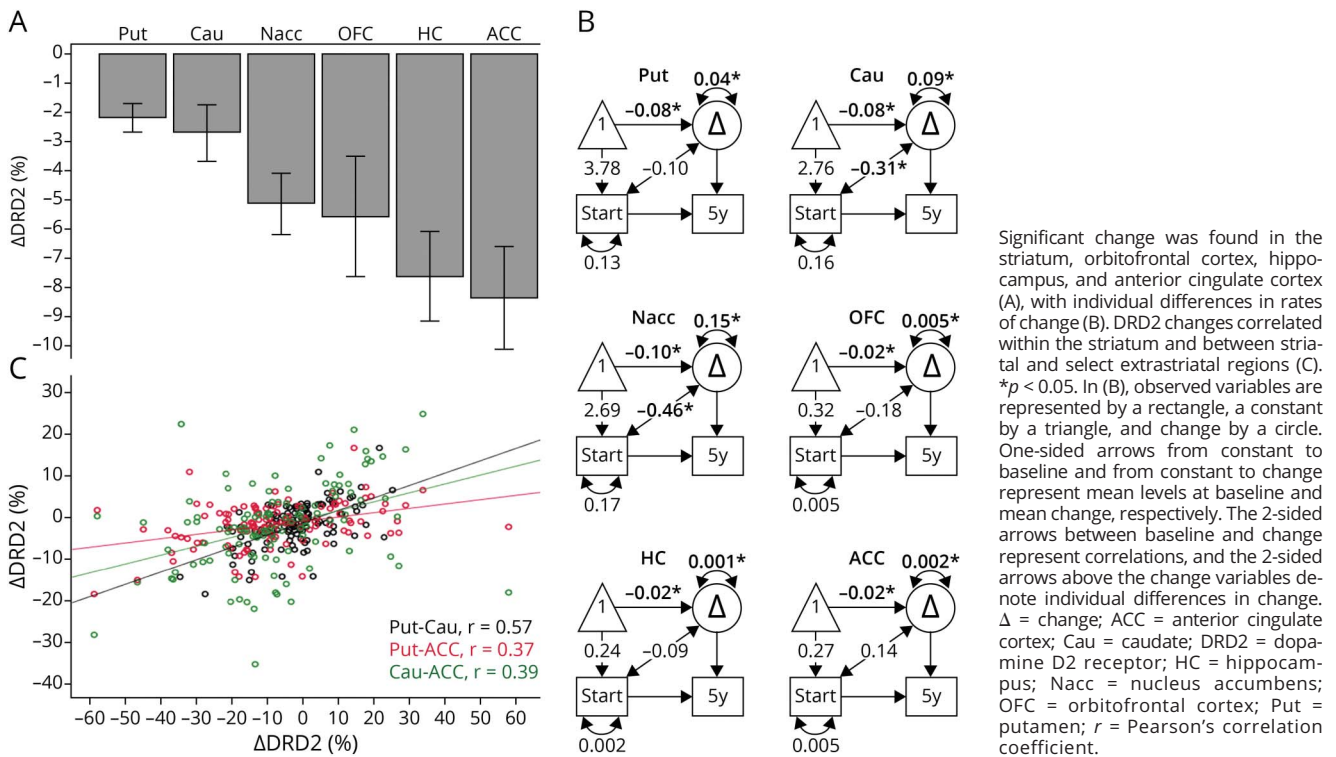
	Returnees	Dropouts	p Value	Selectivity
<b>Sex, men, women</b>	69, 60	31, 21	0.453	0.04
<b>Education, y</b>	13.4 (3.5)	12.9 (3.6)	0.372	0.04
<b>Age, y</b>	66.2 (1.2)	66.3 (1.2)	0.607	-0.02
<b>Retirement</b>	66.7%	84.6%	0.015	-0.11
<b>Performance</b>				
<b>Vocabulary</b>	23.6 (3.5)	22.1 (4.0)	0.016	0.11
<b>Digit-symbol coding</b>	38.7 (7.9)	34.9 (8.1)	0.004	0.14
<b>MMSE</b>	29.3 (0.7)	29.1 (1.0)	0.036	0.10
<b>Activity levels, h/wk</b>				
<b>Intellectual</b>	30.8 (13.2)	25.8 (11.2)	0.024	0.11
<b>Social</b>	32.8 (14.8)	28.9 (14.7)	0.117	0.08
<b>Physical</b>	12.2 (7.3)	11.4 (7.7)	0.491	0.03
<b>Health</b>				
<b>Smoking status</b>	6.2%	13.5%	0.109	-0.08
<b>Hypertension diagnosis</b>	29.5%	42.3%	0.097	-0.08
<b>Hyperlipidemia diagnosis</b>	14%	21%	0.232	-0.05
<b>Systolic, mm Hg</b>	140.9 (16.1)	144.1 (20.3)	0.269	-0.05
<b>Diastolic, mm Hg</b>	84.9 (9.7)	85.5 (10.3)	0.705	-0.02
<b>BMI</b>	26.2 (3.4)	25.7 (3.2)	0.365	0.04
<b>ApoE ε4-allele carriers</b>	25.8%	36.5%	0.149	-0.07
<b>Cardiovascular disease risk (%)</b>	23.0 (10.0)	27.6 (13.3)	0.026	-0.12
<b>Brain</b>				
<b>Caudate DRD2</b>	2.67 (0.38)	2.91 (0.44)	0.000	-0.16
<b>Putamen DRD2</b>	3.74 (0.36)	3.85 (0.37)	0.062	-0.09
<b>Hippocampal DRD2</b>	0.25 (0.05)	0.24 (0.05)	0.387	0.04
<b>Cortical perfusion</b>	41.7 (9.2)	42.1 (10.7)	0.816	-0.01
<b>Subcortical perfusion</b>	43.5 (7.0)	43.6 (7.3)	0.921	0.00
<b>Lesion volume, mm<sup>3</sup></b>	2.0 (1.9)	2.95 (2.47)	0.020	-0.13
<b>No. of lesions</b>	28.2 (14.2)	34.5 (14.2)	0.009	-0.13
<b>Brain per intracranial ratio, %</b>	0.74 (0.03)	0.76 (0.06)	0.021	-0.14
<b>Lateral ventricles, cm<sup>3</sup></b>	30.7 (14.1)	33.5 (13.6)	0.232	-0.06

Abbreviations: BMI = body mass index; DRD2 = dopamine D2-like receptor; MMSE = Mini-Mental State Examination. Values are presented with means, SDs, frequencies (%), and selectivity. Selectivity represents the mean value for returnees at baseline, standardized to the mean value and SD for the parent sample at baseline. DRD2 values are corrected for partial volume effects. Group differences were estimated with paired-sample *t* tests for continuous variables and  $\chi^2$  tests for dichotomous variables. The Bonferroni-adjusted  $\alpha$  level is 0.002 (0.05/27).

subcortical perfusion was motivated by observations of aging-related perfusion reductions being higher in several frontal and temporal regions as compared to subcortical regions<sup>34</sup> and may thus have different bearings on DRD2 losses in these

regions. Next, bivariate difference score models were estimated to assess the vascular-DRD2 association at the indicator level (i.e., per DRD2 region; Figure 2, B and D). The models included associations between vascular parameters

**Figure 1** Five-Year Changes for In Vivo DRD2 Availability in Older Adults



and DRD2 at baseline, change-change correlations, and standardized covariances between baseline levels in variable a and change in variable b (and vice versa).<sup>35</sup>

### Data Availability

The anonymized datasets generated and analyzed during the current study are available from the corresponding author on reasonable request from a qualified investigator. Prerequisites encompass approval of a formal project outline and data sharing agreement and ethical permission for the outlined research questions.

## Results

### Attrition and Longitudinal Selectivity

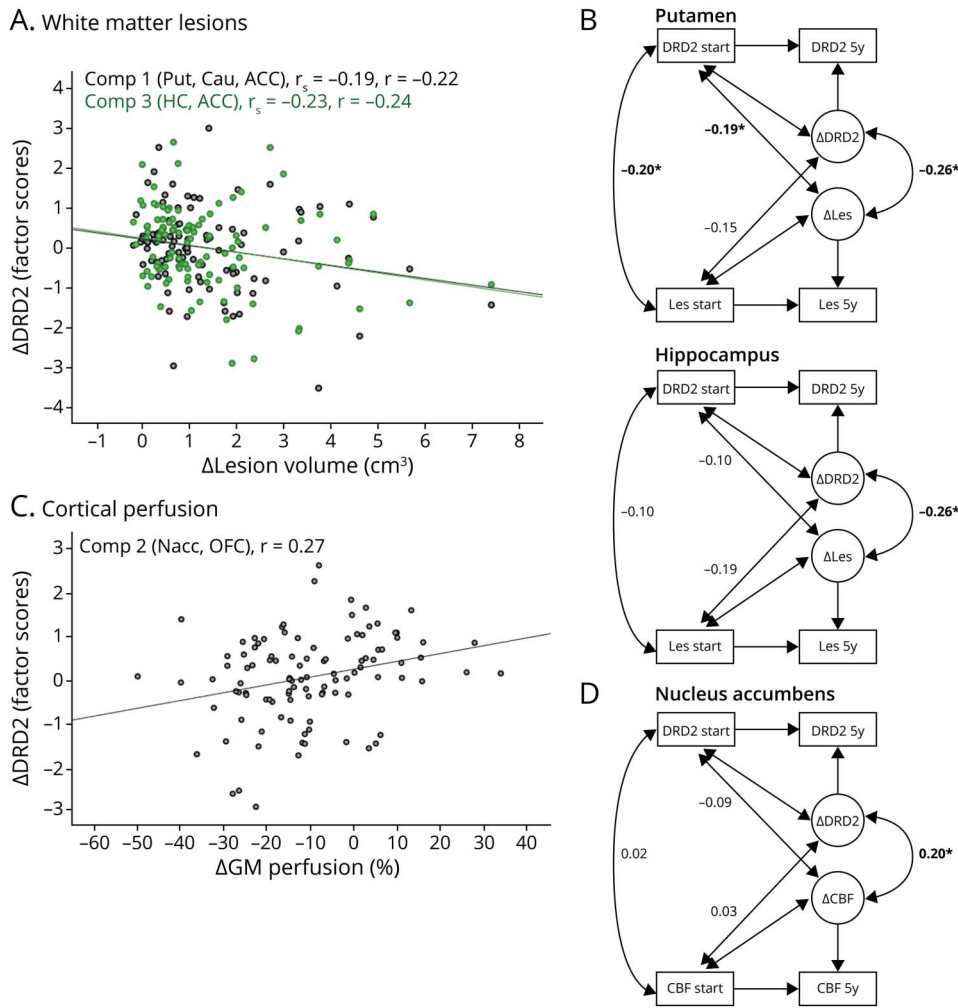
To assess the degree of nonrandom dropout as a possible threat to generalization,<sup>32</sup> we investigated longitudinal selectivity. Selectivity was relatively low (0.10–0.16 SDs; Table 2). Participants not returning for the follow-up session were somewhat more likely to be retired, to show lower cognitive baseline performance and activity levels, having poorer vascular health, and having higher caudate DRD2 levels at baseline. Three individuals died between test waves. Elevated caudate DRD2s for dropouts reflect high dropout (43%) from a previously identified subgroup characterized by low cognition but high DRD2 levels.<sup>29</sup> Dropout was 21% for a subgroup with high cognition and high DRD2 levels and 33% for a group with low cognition and low DRD2 levels ( $p = 0.032$ ; eFigure 1, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)).

For the returnees, significant 5-year changes included reduced systolic ( $140.9 \pm 16.1$  and  $136.5 \pm 17.2$ ,  $t(128) = 2.9$ ,  $p = 0.004$ ) and diastolic ( $84.9 \pm 9.7$  and  $82.4 \pm 10.1$ ,  $t(127) = 2.9$ ,  $p = 0.005$ ) blood pressure, likely reflecting additional medications at follow-up (number of medicines:  $1.26 \pm 1.7$  at baseline and  $2.0 \pm 1.9$  at follow-up;  $t(128) = 3.2$ ,  $p = 0.001$ ). The frequency of medication increased from 30% to 46% for hypertension and from 14% to 29% for hyperlipidemia. The BMI remained unchanged over time ( $26.1 \pm 3.3$  and  $26.3 \pm 3.6$ ,  $t(126) = -1.2$ ,  $p = 0.225$ ). Five participants were diagnosed with diabetes mellitus between waves, which was an exclusion criterion at baseline but not at follow-up. No cases of neurologic disorders (e.g., Parkinson disease) were noted at follow-up. The frequency of retirement was 67% at baseline and 91% at follow-up.

### Five-Year Changes in DRD2 Availability

The PVE-corrected DRD2 measures revealed significant 5-year DRD2 reductions within the striatum (putamen:  $t(118) = 4.49$ , caudate:  $t(118) = 2.47$ , and nucleus accumbens:  $t(118) = 3.67$ ; Table 1 and Figure 1A) and also in select extrastriatal regions (anterior cingulate cortex [ACC]:  $t(116) = 4.89$ , hippocampus:  $t(118) = 5.33$ , and orbitofrontal cortex [OFC]:  $t(117) = 3.10$ ). Difference score models confirmed significant individual differences in DRD2 change for these regions (see double-headed arrow above the change factor; Figure 1B). The magnitude of DRD2 loss (%) was comparable among striatal regions but higher extrastrially ( $F(5, 740) = 3.4$ ,  $p = 0.004$ ; Figure 1A). Specifically,

**Figure 2** Cerebrovascular Changes Are Associated With DRD2 Decline



Increased white matter lesion burden was associated with exacerbated DRD2 decline in the dorsal striatum, hippocampus, and anterior cingulate cortex (A). Bivariate difference score models revealed change-change associations for the putamen and hippocampus (B). Reductions in cortical perfusion were associated with DRD2 losses in the nucleus accumbens and orbitofrontal cortex (C). Change-change links were especially evident for the nucleus accumbens (D). In (B) and (D), observed variables are represented by a rectangle and change by a circle. The 2-sided arrows represent correlations. Note that mean values have been omitted for visual clarity. \* $p < 0.05$ .  $\Delta$  = change; ACC = anterior cingulate cortex; Cau = caudate; CBF = cerebral blood flow; Comp = component; DRD2 = dopamine D2 receptor availability; GM = gray matter; HC = hippocampus; Les = lesions; Nacc = nucleus accumbens; OFC = orbitofrontal cortex; Put = putamen;  $r_s$  = Spearman's rank-order correlation coefficient;  $r$  = Pearson's correlation coefficient.

between-region comparisons revealed that DRD2 decline was significantly larger in the ACC, as compared to the putamen ( $p = 0.014$ ) and caudate ( $p = 0.045$ ). Furthermore, significant DRD2 reductions remained after adjustment for GM loss in each region ( $F$  values with/without covariates: putamen: 40.2/20.1, caudate: 10.7/6.1, nucleus accumbens: 7.1/13.4, hippocampus: 8.9/28.4, ACC: 18.8/20.1, and OFC: 8.8/9.6;  $p < 0.05$  for all). It should be noted that a significant DRD2 decline was found within these regions also when analyzing DRD2  $BP_{ND}$  values derived from the pipeline without PVE correction (without PVC in Table 1). Reliable DRD2 change was not observed in other extrastriatal regions (eTable 2, [links.lww.com/WNL/C161](https://www.lww.com/WNL/C161)).

Within-person regional DRD2 change did not differ across hemispheres ( $t(111-117) = 0.2-1.1$ ;  $p > 0.05$ ), nor as a function of age ( $F(6, 104) = 0.9$ ,  $p = 0.523$ ) or level of educational attainment ( $F(6, 104) = 1.0$ ,  $p = 0.446$ ). We have previously reported sex differences for striatal DRD2 levels at baseline.<sup>18,36</sup> No sex differences were found for DRD2 decline in the dorsal striatum or in the

ACC, hippocampus, or OFC ( $p > 0.05$ ); however, women showed steeper DRD2 decline rates than men in the nucleus accumbens ( $-8.0\%$  vs  $-3.0\%$ ;  $F(1,110) = 5.8$ ,  $p = 0.018$ ). Analyses with values derived from the pipeline without PVE correction revealed no association between DRD2 change in relation to age ( $F(6, 116) = 1.2$ ,  $p = 0.304$ ), level of educational attainment ( $F(6, 116) = 0.7$ ,  $p = 0.624$ ), or sex ( $F(6,116) = 2.0$ ,  $p = 0.071$ ). However, exacerbated right-sided DRD2 reductions were found for the nucleus accumbens ( $t(125) = 3.2$ ,  $p = 0.002$ ), OFC ( $t(125) = 5.0$ ,  $p < 0.001$ ), and hippocampus ( $t(125) = 2.1$ ,  $p = 0.041$ ).

### Patterns of Striatal and Extrastriatal DRD2 Changes

A PCA was conducted to investigate the structure of regionally correlated DRD2 changes across those regions showing a significant mean change and variance in change (i.e., the regions in Figure 1, A and B). Three principal components were significant in a rotated solution (model 1 in Table 3). The first component (36% of variance explained) included the caudate, putamen, and ACC. The nucleus



**Table 3** Patterns of DRD2 Change

Components	1	2	3	4	5
<b>Model 1: DRD2 change</b>					
Variance explained	36%	28%	17%		
Putamen	<b>0.88</b>	0.18	0.09		
Caudate	<b>0.86</b>	-0.43	0.12		
Nucleus accumbens	0.06	<b>0.87</b>	-0.15		
Hippocampus	0.12	0.11	<b>0.93</b>		
ACC	<b>0.66</b>	-0.02	<b>0.58</b>		
OFC	-0.16	<b>0.88</b>	0.20		
<b>Model 2: DRD2 and volume changes</b>					
Variance explained	23%	19%	14%	11%	9%
Putamen DRD2	-0.07	<b>0.80</b>	0.19	-0.48	0.02
Caudate DRD2	-0.09	<b>0.85</b>	-0.41	-0.31	0.02
Nucleus accumbens DRD2	0.12	0.01	<b>0.86</b>	-0.02	0.16
Hippocampus DRD2	0.12	0.20	0.01	-0.03	<b>-0.93</b>
ACC DRD2	0.08	<b>0.77</b>	0.00	0.25	-0.35
OFC DRD2	0.03	-0.12	<b>0.88</b>	0.02	-0.21
Putamen volume	0.05	-0.09	-0.08	<b>0.85</b>	-0.10
Caudate volume	0.21	-0.23	0.12	<b>0.77</b>	0.12
Nucleus accumbens volume	<b>0.94</b>	0.08	0.08	0.10	-0.04
Hippocampus volume	<b>0.58</b>	0.17	0.17	-0.05	-0.36
ACC volume	<b>0.94</b>	0.08	0.08	0.10	-0.04
OFC volume	<b>0.65</b>	-0.05	-0.05	0.35	-0.21

Abbreviations: ACC = anterior cingulate cortex; DRD2 = dopamine D2-like receptor; OFC = orbitofrontal cortex.

Principal component analyses revealed that DRD2 changes correlated across the dorsal striatum and ACC; ventral striatum and OFC; and hippocampus and ACC (model 1). Similar DRD2 change patterns were found when including volumes (model 2).

DRD2 values are corrected for partial volume effects. Components with eigenvalues >1 are shown. Coefficients <-0.50 and >0.50 are marked with bold font.

accumbens and OFC loaded strongly on the second component (28% of variance), whereas the hippocampus and ACC loaded onto a third component (17% of variance). Pairwise correlations for DRD2 change in regions loading onto the first component are illustrated in Figure 1C. However, no between-component correlations were found (component 1 vs 2:  $r = -0.06$ ,  $p = 0.546$ ; 1 vs 3:  $r = 0.16$ ,  $p = 0.092$ ; 2 vs 3:  $r = 0.01$ ,  $p = 0.943$ ), indicating the presence of distinct patterns of DRD2 change. A similar pattern was found when analyzing the BP<sub>ND</sub> values without PVE correction (eTable 1, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)).

The potential influence of GM atrophy to DRD2 change was assessed through a second PCA where DRD2 change

and volume change per ROI was entered. This model demonstrated correlations between ACC-striatal, as compared to ACC-hippocampal, DRD2 changes (Table 3, model 2). Furthermore, correlations were found for volume changes and DRD2 change for the dorsal striatum. Significant zero-order correlations for DRD2 vs volume change were found for the striatum across both DRD2 pipelines, but not for extrastriatal regions (eTable 3, [links.lww.com/WNL/C161](https://links.lww.com/WNL/C161)).

### Relationships Between Cerebrovascular Health and DRD2 Decline

Sizable perfusion reductions were observed across 5 years (approx. -6 to -10%; Table 1). WM lesion burden varied greatly, and individuals with the largest lesion diameters at baseline demonstrated larger 5-year increases in lesion burden ( $0.5 \pm 0.5 \text{ cm}^3$ ,  $1.41 \pm 1.1 \text{ cm}^3$ , and  $3.9 \pm 2.0 \text{ cm}^3$  for individuals with lesion diameters of <10, 10–20, and >20 mm at baseline, respectively;  $F(2, 118) = 43.7$ ,  $p < 0.001$ ). GM perfusion was negatively associated with WM lesion burden at baseline (cortical perfusion:  $r_s = -0.22$ ,  $p = 0.019$ ; subcortical perfusion:  $r_s = -0.17$ ,  $p = 0.061$ ). However, no change-change associations were found for these variables ( $r_s = 0.08$  and  $0.09$  for cortical and subcortical perfusion, respectively;  $p$  values >0.05).

No association was found among WM lesion volume and cortical or subcortical GM volume at baseline ( $r_s = 0.15$  and  $0.11$ , respectively,  $p$ 's > 0.05). WM lesion volume change was associated with cortical ( $r = -0.21$ ,  $p = 0.026$ ), but not subcortical ( $r_s = 0.12$ ,  $p > 0.05$ ) volume change. No perfusion-volume associations were found for cortical ( $r = -0.03$ ,  $p > 0.05$ ) or subcortical GM ( $r = -0.02$ ,  $p > 0.05$ ) at baseline or for changes among these measures ( $r_s = 0.09$  and  $-0.05$ ,  $p$ 's > 0.05).

Factor scores were extracted for the 3 distinct dimensions of DRD2 losses (Table 3) and entered as dependent variables in a multivariate regression analysis with (1) change in WM lesion volume, (2) change in frontotemporal GM perfusion, and (3) subcortical GM perfusion as independent variables. DRD2 change was associated with changes in lesion volume ( $F(3,94) = 3.7$ ,  $p = 0.015$ ) and cortical perfusion ( $F(3,94) = 3.3$ ,  $p = 0.024$ ). Specifically, increases in WM lesion volume were associated with more pronounced DRD2 loss in regions of component 1 (caudate, putamen, and ACC;  $F(1,96) = 5.1$ ,  $p = 0.026$ ) and component 3 (hippocampus and ACC;  $F(1,96) = 6.0$ ,  $p = 0.016$ ; Figure 2A), whereas reduced frontotemporal GM perfusion was associated with DRD2 loss in regions of component 2 (nucleus accumbens and OFC;  $F(1,96) = 8.1$ ,  $p = 0.006$ ; Figure 2C).

Bivariate difference score models demonstrated that baseline lesion burden was associated with baseline DRD2 levels in the putamen (Figure 2B), but also in the caudate ( $r = -0.34$ ,  $p < 0.05$ ) and ACC ( $r = -0.28$ ,  $p < 0.05$ ). Significant change-change associations were found for lesions and DRD2 levels in the putamen and hippocampus (Figure 2B). Moreover, baseline DRD2 levels in the putamen (Figure 2B),

caudate ( $r = -0.26, p < 0.05$ ), and ACC ( $r = -0.18, p < 0.05$ ) were associated with change in lesion burden. Change-change associations were found for cortical GM perfusion and DRD2 in the nucleus accumbens (Figure 2D), but not the OFC ( $r = 0.14$ ).

## Discussion

This work provides the first longitudinal evidence for aging-related DRD2 losses and paves the way for a mechanistic understanding for such reductions. The magnitude of striatal DRD2 reductions was about half of previous cross-sectional estimates of age differences across the adult lifespan ( $\sim 4\%$  vs  $\sim 8\%$  per decade).<sup>4</sup> This demonstrates once again differences in cross-sectional approximations vs true longitudinal measurements of age-related changes. Thus, previous statements about the magnitude of average DA decline,<sup>3,4</sup> at least for the age segment of  $\sim 65$ – $75$  years, need to be revised. Cohort effects and infrequent use of PVE correction may have contributed to the higher magnitude of the cross-sectional estimates.<sup>25</sup> For instance, caudate DRD2 reductions were twice as large for uncorrected, as compared to PVE-corrected, values. DRD2 losses correlated across striatal and select extrastriatal regions and were associated with changes in cerebrovascular parameters. DRD2 losses were primarily observed in regions particularly susceptible to vascular insults, such as basal ganglia and hippocampus.<sup>21,37</sup>

The current study demonstrates that within-person DRD2 changes in aging are particularly marked in several associative and limbic regions, such as caudate, nucleus accumbens, hippocampus, and ACC. These are also regions with prominent dopaminergic losses when cognitive impairments and dementia emerge.<sup>38</sup> Cross-sectional studies have revealed mixed regional rank orders for age differences in DRD2s. Using <sup>18</sup>F-fallypride/PET, the largest age differences were demonstrated for frontal and temporal DRD2s ( $-6\%$  to  $-16\%$  per decade), followed by striatal ( $-3\%$  to  $-5\%$ ) and hippocampal DRD2s ( $0\%$  to  $-2\%$ ).<sup>39</sup> <sup>11</sup>C-FLB 457/PET revealed similar cortical and hippocampal DRD2 reductions as reported in the current study ( $-12\%$  to  $-14\%$  per decade).<sup>40</sup> Insufficient statistical power, cohort effects, and methodologic choices (e.g., ligand characteristics) may underlie the diverging age effects on hippocampal DRD2s in previous studies.<sup>12,25,39</sup>

Atrophy can influence ligand-binding estimates, with overestimated aging effects as a result.<sup>25</sup> Notably, analyses on PVE-corrected and uncorrected DRD2 values revealed similar regional patterns, with lower DRD2 reductions in a few regions after PVE correction. The patterns of change remained after controlling for within-person atrophy, underscoring that DRD2 decline was not driven by volume losses. This is in line with experimental work conducted at the same scanner and with the same reconstruction methods.<sup>41</sup> Other sources of variation across DRD2 studies are differences in ligand characteristics. Despite low accumulation and a signal-to-

noise ratio in extrastriatal regions,<sup>42</sup> a growing number of studies demonstrate reliability and validity for extrastriatal <sup>11</sup>C-raclopride binding.<sup>11,20,43,44</sup>

There is a distinct topography of midbrain DA neuron projections.<sup>45,46</sup> DRD2 losses were correlated within DA pathways, as indicated previously,<sup>19,20,47</sup> but also between DA pathways (striatum-ACC). The distinct pattern of DRD2 decline in interconnected structures of the reward circuit (ventral striatum and OFC),<sup>46</sup> and across associative and motor regions (dorsal striatum, hippocampus, and ACC), with links to reduced frontotemporal perfusion and lesion progression, respectively, is noteworthy. The relationship between cerebral perfusion and lesion manifestation in aging is unclear.<sup>48</sup> Perfusion data, as quantified with pcASL, have a fair amount of noise, which was minimized here through averaging of voxels in relatively large ROIs. The distinct links between dorsal vs ventral striatal DRD2 levels and the vascular parameters may stem from separate microvascular territories within the striatum, giving rise to differential susceptibility to ischemic events.<sup>49</sup> Another factor could be proximity to the epicenter where vascular injury typically manifests.

A conceivable mechanism that might explain the observed associations is that vascular dysfunction exerts insult in target regions of DA nerve fibers. Still, it is not possible to firmly delineate the direction of causality between vascular and DA alterations. As shown here, baseline DRD2 status was associated with lesion progression. There is a vasomotor response after provision of DA agonists,<sup>50</sup> and patients with Parkinson disease are at higher risk for cardiovascular disease.<sup>16</sup> Hence, the relationship between these cascades may be reciprocal. Generalizability of our findings is limited by the narrow age range of the COBRA sample, and that dropout may have led to selection bias. Consequently, we cannot exclude the possibility that DRD2 decline rates may differ in younger vs older age segments. Furthermore, the DRD2-vascular link may be underestimated in older persons because compromised vascular health predicted dropout. DRD2 reductions in younger age segments (30 years and onward<sup>4</sup>) may be attributable to factors other than vascular health.

This longitudinal study provides new knowledge on how aging affects DRD2 levels and suggests that individual differences in rates of DRD2 change are related to vascular health. Future work may consider conducting extensive mapping of several cerebrovascular events<sup>48</sup> to further examine the DA-vascular link in aging.

## Study Funding

This study was funded by the Swedish Research Council (grant numbers: 421-2012-648 [2012–2016] and 2017-02217 [2017–2020]), Umeå University, Umeå University–Karolinska Institute Strategic Neuroscience Program, the Knut and Alice Wallenberg Foundation (grant number: 2015.0277), the Torsten and Ragnar Söderberg Foundation,

an Alexander von Humboldt Research Award, a donation from the Jochnick Foundation, Swedish Brain Power, the Swedish Brain Foundation, Västerbotten County Council, Innovation Fund of the Max Planck Society, and Gottfried Wilhelm Leibniz Research Award 2010 of the German Research Foundation (DFG). The FreeSurfer analyses were performed on resources provided by the Swedish National Infrastructure for Computing (SNIC) at HPC2N in Umeå, partially funded by the Swedish Research Council through grant agreement no. 2018-05973.

## Disclosure

The authors report no relevant disclosures. Go to Neurology.org/N for full disclosures.

## Publication History

Received by *Neurology* December 14, 2021. Accepted in final form May 11, 2022. Submitted and externally peer reviewed. The handling editor was Linda Hershey, MD, PhD, FAAN.

## Appendix Authors

Name	Location	Contribution
<b>Nina Karalija, PhD</b>	Department of Radiation Sciences, Diagnostic Radiology, and Umeå Center for Functional Brain Imaging (UFBI), Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content; study concept or design; and analysis or interpretation of data
<b>Jarkko Johansson, PhD</b>	Department of Radiation Sciences, Diagnostic Radiology, and Umeå Center for Functional Brain Imaging (UFBI), Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Goran Papenberg, PhD</b>	Aging Research Center, Karolinska Institutet & Stockholm University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Anders Wåhlin, PhD</b>	Department of Radiation Sciences, Radiation Physics, Department of Applied Physics and Electronics, and Umeå Center for Functional Brain Imaging (UFBI), Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Alireza Salami, PhD</b>	Umeå Center for Functional Brain Imaging (UFBI), Umeå University; Aging Research Center, Karolinska Institutet & Stockholm University; Department of Integrative Medical Biology, and Wallenberg Center for Molecular Medicine, Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Ylva Köhncke, PhD</b>	Center for Lifespan Psychology, Max Planck Institute for Human Development, Berlin, Germany	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data

## Appendix (continued)

Name	Location	Contribution
<b>Andreas M. Brandmaier, PhD</b>	Center for Lifespan Psychology, Max Planck Institute for Human Development, Berlin, Germany; Max Planck UCL Centre for Computational Psychiatry and Ageing Research, Berlin, Germany, and London, United Kingdom	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Micael Andersson, MSc</b>	Umeå Center for Functional Brain Imaging (UFBI), and Department of Integrative Medical Biology, Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Jan Axelsson, PhD</b>	Department of Radiation Sciences, Radiation Physics, and Umeå Center for Functional Brain Imaging (UFBI), Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content, and analysis or interpretation of data
<b>Katrine Riklund, MD, PhD</b>	Department of Radiation Sciences, Diagnostic Radiology, and Umeå Center for Functional Brain Imaging (UFBI), Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and study concept or design
<b>Martin Lövdén, PhD</b>	Department of Psychology, University of Gothenburg, Sweden	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and study concept or design
<b>Ulman Lindenberger, PhD</b>	Center for Lifespan Psychology, Max Planck Institute for Human Development; Max Planck UCL Centre for Computational Psychiatry and Ageing Research, Berlin, Germany, and London, United Kingdom	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and study concept or design
<b>Lars Bäckman, PhD</b>	Aging Research Center, Karolinska Institutet & Stockholm University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and study concept or design
<b>Lars Nyberg, PhD</b>	Department of Radiation Sciences, Diagnostic Radiology, Umeå Center for Functional Brain Imaging (UFBI), Department of Integrative Medical Biology, and Wallenberg Center for Molecular Medicine, Umeå University, Sweden	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and study concept or design

## References

- Nyberg L, Lövdén M, Riklund K, Lindenberger U, Bäckman L. Memory aging and brain maintenance. *Trends Cogn Sci*. 2012;16(5):292-305.
- Lindenberger U. Human cognitive aging: corrigere la fortune? *Science*. 2014; 346(6209):572-578.
- Bäckman L, Nyberg L, Lindenberger U, Li SC, Farde L. The correlative triad among aging, dopamine, and cognition: current status and future prospects. *Neurosci Biobehav Rev*. 2006;30(6):791-807.



4. Karrer TM, Josef AK, Mata R, Morris ED, Samanez-Larkin GR. Reduced dopamine receptors and transporters but not synthesis capacity in normal aging adults: a meta-analysis. *Neurobiol Aging*. 2017;57:36-46.
5. Cools R. Chemistry of the adaptive mind: lessons from dopamine. *Neuron*. 2019;104(1):113-131.
6. Raz N, Lindenberger U, Rodrigue KM, et al. Regional brain changes in aging healthy adults: general trends, individual differences and modifiers. *Cereb Cortex*. 2005;15(11):1676-1689.
7. Nyberg L, Salami A, Andersson M, et al. Longitudinal evidence for diminished frontal cortex function in aging. *Proc Natl Acad Sci USA*. 2010;107(52):22682-22686.
8. Rönnlund M, Nyberg L, Bäckman L, Nilsson LG. Stability, growth, and decline in adult life span development of declarative memory: cross-sectional and longitudinal data from a population-based study. *Psychol Aging*. 2005;20(1):3-18.
9. Lindenberger U, von Oertzen T, Ghisletta P, Hertzog C. Cross-sectional age variance extraction: what's change got to do with it? *Psychol Aging*. 2011;26(1):34-47.
10. Jonasson LS, Nyberg L, Kramer AF, Lundquist A, Riklund K, Boraxbekk CJ. Aerobic exercise intervention, cognitive performance, and brain structure: results from the physical influences on brain in aging (PHIBRA) study. *Front Aging Neurosci*. 2017;8:336.
11. Alakurtti K, Johansson JJ, Joutsa J, et al. Long-term test-retest reliability of striatal and extrastriatal dopamine D2/3 receptor binding: study with [(11)C]raclopride and high-resolution PET. *J Cereb Blood Flow Metab*. 2015;35(7):1199-1205.
12. Raz N, Lindenberger U. Only time will tell: cross-sectional studies offer no solution to the age-brain-cognition triangle: comment on Salthouse (2011). *Psychol Bull*. 2011;137(5):790-795.
13. Iturria-Medina Y, Sotero RC, Toussaint PJ, Mateos-Pérez JM, Evans AC. Early role of vascular dysregulation on late-onset Alzheimer's disease based on multifactorial data-driven analysis. *Nat Commun*. 2016;7:11934.
14. Juttukonda MR, Li B, Almaktoom R, et al. Characterizing cerebral hemodynamics across the adult lifespan with arterial spin labeling MRI data from the Human Connectome Project-Aging. *Neuroimage*. 2021;230:117807.
15. Gower A, Tiberi M. The intersection of central dopamine system and stroke: potential avenues aiming at enhancement of motor recovery. *Front Synaptic Neurosci*. 2018;10:18.
16. Park JH, Kim DH, Park YG, et al. Association of Parkinson disease with risk of cardiovascular disease and all-cause mortality. *Circulation*. 2020;141(14):1205-1207.
17. Karalija N, Wählin A, Ek J, et al. Cardiovascular factors are related to dopamine integrity and cognition in aging. *Ann Clin Transl Neurol*. 2019;6(11):2291-2303.
18. Nevalainen N, Riklund K, Andersson M, et al. COBRA: a prospective multimodal imaging study of dopamine, brain structure and function, and cognition. *Brain Res*. 2015;1612:83-103.
19. Zald DH, Woodward ND, Cowan RL, et al. The interrelationship of dopamine D2-like receptor availability in striatal and extrastriatal brain regions in healthy humans: a principal component analysis of [18F]Fallypride binding. *Neuroimage*. 2010;51(1):53-62.
20. Papenberg G, Jonasson L, Karalija N, et al. Mapping the landscape of human dopamine D2/3 receptors with [11C]raclopride. *Brain Struct Funct*. 2019;224(8):2871-2882.
21. Wählin A, Nyberg L. At the heart of cognitive functioning in aging. *Trends Cogn Sci*. 2019;23(9):717-720.
22. Spence JD. Blood pressure gradients in the brain: their importance to understanding pathogenesis of cerebral small vessel disease. *Brain Sci*. 2019;9:21.
23. Desikan RS, Ségonne F, Fischl B, et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage*. 2006;31(3):968-980.
24. Greve DN, Salat DH, Bowen SL, et al. Different partial volume correction methods lead to different conclusions: an (18)F-FDG-PET study of aging. *Neuroimage*. 2016;132:334-343.
25. Smith CT, Crawford JL, Dang LC, et al. Partial-volume correction increases estimated dopamine D2-like receptor binding potential and reduces adult age differences. *J Cereb Blood Flow Metab*. 2019;39(5):822-833.
26. Logan J, Fowler JS, Volkow ND, Wang GJ, Ding YS, Alexoff DL. Distribution volume ratios without blood sampling from graphical analysis of PET data. *J Cereb Blood Flow Metab*. 1996;16(5):834-840.
27. Schmidt P, Gaser C, Arsic M, et al. An automated tool for detection of FLAIR-hyperintense white-matter lesions in Multiple Sclerosis. *Neuroimage*. 2012;59(4):3774-3783.
28. [statisticalmodelling.de/1st.html](http://statisticalmodelling.de/1st.html).
29. Lövdén M, Karalija N, Andersson M, et al. Latent-profile analysis reveals behavioral and brain correlates of dopamine-cognition associations. *Cereb Cortex*. 2018;28(11):3894-3907.
30. von Oertzen T, Brandmaier AM, Tsang S. Structural equation modeling with  $\Omega$ nyx. *Struct Equ Modeling*. 2015;22(1):148-161. <https://onyx-sem.com/>.
31. Lindenberger U, Singer T, Baltes PB. Longitudinal selectivity in aging populations: separating mortality-associated versus experimental components in the Berlin Aging Study (BASE). *J Gerontol B Psychol Sci Soc Sci*. 2002;57(6):P474-P482.
32. Brandmaier AM, von Oertzen T, Ghisletta P, Lindenberger U, Hertzog C. Precision, reliability, and effect size of slope variance in latent growth curve models: implications for statistical power analysis. *Front Psychol*. 2018;9:294.
33. Chen JJ, Rosas HD, Salat DH. Age-associated reductions in cerebral blood flow are independent from regional atrophy. *Neuroimage*. 2011;55(2):468-478.
34. Kievit RA, Brandmaier AM, Ziegler G, et al. Developmental cognitive neuroscience using latent change score models: a tutorial and applications. *Dev Cogn Neurosci*. 2018;33:99-117.
35. Karalija N, Papenberg G, Wählin A, et al. Sex differences in dopamine integrity and brain structure among healthy older adults: relationships to episodic memory. *Neurobiol Aging*. 2021;105:272-279.
36. Smith EE, Saposnik G, Biessels GJ, et al. Prevention of stroke in patients with silent cerebrovascular disease: a scientific statement for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke*. 2017;48(2):e44-e71.
37. Sala A, Caminiti SP, Presotto L, et al. In vivo human molecular neuroimaging of dopaminergic vulnerability along the Alzheimer's disease phases. *Alzheimers Res Ther*. 2021;13(1):187.
38. Seaman KL, Smith CT, Juarez EJ, et al. Differential regional decline in dopamine receptor availability across adulthood: linear and nonlinear effects of age. *Hum Brain Mapp*. 2019;40(10):3125-3138.
39. Inoue M, Suhara T, Sudo Y, et al. Age-related reduction of extrastriatal dopamine D2 receptor measured by PET. *Life Sci*. 2001;69(9):1079-1084.
40. Jonasson LS, Axelsson J, Riklund K, Boraxbekk CJ. Simulating effects of brain atrophy in longitudinal PET imaging with an anthropomorphic brain phantom. *Phys Med Biol*. 2017;62(13):S213-S227.
41. Farde L, Pauli S, Hall H, et al. Stereoselective binding of 11C-raclopride in living human brain—a search for extrastriatal central D2-dopamine receptors by PET. *Psychopharmacology (Berl)*. 1988;94(4):471-478.
42. Karalija N, Jonasson L, Johansson J, et al. High long-term test-retest reliability for extrastriatal 11C-raclopride binding in healthy older adults. *J Cereb Blood Flow Metab*. 2019;40(9):1859-1868.
43. Lippert RN, Cremer AL, Edwin Thanarajah S, et al. Time-dependent assessment of stimulus-evoked regional dopamine release. *Nat Commun*. 2019;10(1):336.
44. Engelhard B, Finkelstein J, Cox J, et al. Specialized coding of sensory, motor and cognitive variables in VTA dopamine neurons. *Nature*. 2019;570(7762):509-513.
45. Haber SN, Knutson B. The reward circuit: linking primate anatomy and human imaging. *Neuropsychopharmacology*. 2010;35(1):4-26.
46. Nyberg L, Karalija N, Salami A, et al. Dopamine D2 receptor availability is linked to hippocampal-caudate functional connectivity and episodic memory. *Proc Natl Acad Sci USA*. 2016;113(28):7918-7923.
47. Wardlaw JM, Smith C, Dichgans M. Small vessel disease: mechanisms and clinical implications. *Lancet Neurol*. 2019;18(7):684-696.
48. Feekes JA, Cassell MD. The vascular supply of the functional compartments of the human striatum. *Brain*. 2006;129(Pt 8):2189-2201.
49. Krimer LS, Muly EC III, Williams GV, Goldman-Rakic PS. Dopaminergic regulation of cerebral cortical microcirculation. *Nat Neurosci*. 1998;1(4):286-289.