# Integrated bioinformatics analysis reveals role of the LINC01093/miR-96-5p/ZFAND5/NF-κB signaling axis in hepatocellular carcinoma

YAHUI ZHENG<sup>\*</sup>, KANGKANG YU<sup>\*</sup>, CHONG HUANG, LU LIU, HAO ZHAO, MEISI HUO and JUBO ZHANG

Department of Infectious Diseases, Huashan Hospital, Fudan University, Shanghai 200040, P.R. China

Received February 24, 2019; Accepted August 19, 2019

DOI: 10.3892/etm.2019.8046

Abstract. Hepatocellular carcinoma (HCC) is a significant health burden worldwide and its pathogenesis remains to be fully elucidated. One of the means by which long non-coding (lnc)RNAs regulate gene expression is by interacting with micro (mi)RNAs and acting as competing endogenous (ce)RNAs. IncRNAs have important roles in various diseases. The aim of the present study was to examine the potential roles of lncRNAs in HCC. The RNA expression profiles of 21 paired tissues of HCC and adjacent non-tumor tissues were obtained from the Gene Expression Omnibus database. The differentially expressed RNAs were analyzed using the DESeq package in R. Expression validation and survival analysis of selected RNAs were performed using Gene Expression Profile Interactive Analysis and/or Kaplan-Meier Plotter. The target genes of the miRNAs were predicted using lncBase or TargetScan. Functional analyses were performed using the Database for Annotation, Visualization and Integrated Discovery, and regulatory networks were determined using Cytoscape. Long intergenic non-protein coding RNA 1093 (LINC01093) was identified as one of the most significantly downregulated lncRNAs in HCC tissues. Downregulated expression of LINC01093 was associated with poor prognosis. A ceRNA network involving LINC01093, miR-96-5p and zinc finger AN1-type containing 5 (ZFAND5) was established. According to functional analyses, NF-KB signaling was implicated in the regulatory network for HCC. The present study

*Correspondence to:* Dr Jubo Zhang, Department of Infectious Diseases, Huashan Hospital, Fudan University, 12 Middle Urumqi Road, Shanghai 200040, P.R. China E-mail: drzhangjubo@163.com

\*Contributed equally

revealed that a LINC01093/miR-96-5p/ZFAND5/NF- $\kappa$ B signaling axis may have an important role in the pathogenesis of HCC, and further investigation of this axis may provide novel insight into the development and progression of HCC.

#### Introduction

Liver cancer was estimated to be the sixth most common cancer type and the fourth leading cause of cancer-associated death worldwide in 2018, and it has been reported that hepatocellular carcinoma (HCC) is the most common subtype, accounting for 75-85% of all primary cases of liver cancer (1). However, the underlying molecular mechanisms of HCC development remain to be fully elucidated, and the high incidence of metastasis and recurrence frequently results in poor prognosis for patients with HCC, even following curative therapy (2). Therefore, it is imperative to identify novel diagnostic and prognostic biomarkers and efficacious therapeutic targets.

Abnormalities in gene transcription and translation serve important roles in the development and progression of HCC. Developments in biotechnology, including genome sequencing and bioinformatics, have facilitated the study of the tumor transcriptome and proteome, which has advanced the current understanding of the underlying molecular mechanisms of HCC. Genome sequencing has indicated that the human genome is comprised of <2% protein-coding genes and >90% of the genome is transcribed as non-coding RNA (ncRNA) (3). Numerous classes of ncRNA have been identified and demonstrated to be associated with cancer, including long-ncRNAs (lncRNAs), microRNAs (miRNAs), small nucleolar RNAs and PIWI-interacting RNAs (4-7), among which lncRNAs and miRNAs have been most extensively studied. lncRNAs are >200 nucleotides long and participate in multiple biological functions, including epigenetics, nuclear import, alternative splicing, RNA decay and translation (5). miRNAs are small ncRNAs of 18-25 nucleotides in length that regulate the expression of multiple mRNAs by reducing the stability and inhibiting the translation of mRNAs at the post-transcriptional level (8). miRNAs restrain the expression of target genes, whereas lncRNAs may competitively combine

*Key words:* hepatocellular carcinoma, long non-coding RNA, microRNA, The Cancer Genome Atlas, competing endogenous RNA

with miRNAs to promote the expression of target genes; in this interaction, they are commonly referred to as competing endogenous RNAs (ceRNAs) (9). Although previous studies have reported on the roles of lncRNAs and miRNAs in HCC (10-14), the specific underlying mechanisms associated with the initiation and progression of HCC remain to be fully elucidated (15).

In the present study, profiles of differentially expressed lncRNAs between HCC tissues and adjacent normal tissues were determined from high-throughput RNA sequencing data.

Bioinformatics and survival analysis using integrated mining of data from The Cancer Genome Atlas (TCGA) was performed (16). Simultaneously, the regulatory mechanisms and signaling pathways in HCC were predicted and their differential diagnostic performance was analyzed to provide a potentially valuable reference for the early diagnosis, effective treatment and prognosis of patients with HCC.

## Materials and methods

*RNA-sequencing (seq) data retrieval and processing.* The GSE94660 dataset was obtained from the Gene Expression Omnibus (GEO), which contains the RNA sequencing data from 21 pairs of tumor and non-neoplastic liver tissues from patients with hepatitis B virus-associated HCC. The normalized gene expression dataset GSE94660 was downloaded from the GEO database (https://www.ncbi.nlm.nih.gov/geo/) (17). To identify the differentially expressed genes between tumor and non-tumor tissues, threshold values of a fold change of 11.51 and P<0.05 were used.

*Expression level analysis*. The expression of lncRNA and mRNA in patients were analyzed using either Gene Expression Profiling Interactive Analysis (GEPIA) (18) or from expression data downloaded from TCGA (16). Expression data of lncRNA in normal tissues were obtained from BioProject (accession no. PRJEB4337; linc01093) (19).

Target prediction and network establishment. To predict potential miRNAs that target candidate lncRNAs, lncBase version 2 (http://carolina.imis.athena-innovation.gr/diana\_ tools/web/index.php?r=lncbasev2/index) (20) was used with a prediction score of  $\geq 0.95$  as the threshold value. The miRNAs identified were further inputted into TargetScan (http://www. targetscan.org/vert\_72/) (21) to identify their potential target genes. Targets with a cumulative weighted context score  $\leq -0.6$ that interacted with a differentially expressed gene identified in the GSE94660 dataset were selected to establish an interaction network with candidate miRNAs and lncRNA using Cytoscape 3.40 (22).

Interactome identification and functional enrichment analysis. Proteins that physically interacted with zinc finger AN1-type containing 5 (ZFAND5) were identified from BioGRID (https://thebiogrid.org/) (23), an interaction repository containing 1,623,645 protein and genetic interactions in a number of species. Gene ontology (GO) (24) and reactome pathway enrichment analyses of the ZFAND5 interactome were performed using the Database for Annotation, Visualization and Integrated Discovery (DAVID; http://david.abcc.ncifcrf.gov/) (25), which provides a comprehensive set of functional annotation tools for researchers to understand the biological contexts surrounding a large number of interacting genes.

Survival analysis. To assess the prognostic value of the lncRNAs and genes identified, GEPIA and Kaplan-Meier plotter (http://kmplot.com/analysis/) (26) were used. Overall survival of patients with HCC was analyzed using a Kaplan-Meier plot based on the median expression levels of each gene. The hazard ratio (HR) and 95% confidence intervals were calculated and a log-rank test was used to determine whether survival was significantly different between patients with high and low expression of each gene in their HCC tissues.

# Results

Downregulated expression of LINC01093 in HCC tissues is associated with poor prognosis. The GSE94660 dataset from GEO database was analyzed, and heatmaps were generated to present the differentially expressed genes (Fig. 1A) and lncRNAs (Fig. 1B) with a fold change  $\geq |1.5|$  and P<0.05. LINC01093 (27,28) was the most significantly downregulated lncRNA (Fig. 1C). The expression profile of LINC01093 in normal tissues was obtained from BioProject, and LINC01093 was almost exclusively expressed in normal liver tissue in humans (Fig. 1D). To validate these results, the expression of LINC01093 in HCC tissues and adjacent non-tumor tissues was analyzed using the GEPIA database. The results indicated that LINC01093 was significantly downregulated in tumor tissues compared with that in normal tissues (Fig. 1E). Survival analysis using Kaplan-Meier plotter suggested that low expression levels of LINC01093 in patients with HCC predicted a less favorable prognosis (Fig. 1F).

LINC01093 is a potential target of miR-96-5p. lncRNAs primarily function as ceRNAs; thus, low expression levels of LINC01093 in tumor tissues lead to high expression of target miRNAs. To identify potential miRNAs that interact with LINC01093, the IncBase database was used. By implementing a cutoff prediction score of  $\geq 0.95$ , five potential miRNAs were identified: miR-30b-3p, miR-627-3p, miR-676-5p, miR-3689d and miR-96-5p. The potential target genes of these miRNAs were predicted using TargetScan and screened with a total context score of  $\leq 0.6$  used as the cutoff. Genes identified in TargetScan, which were also differentially expressed in the GSE94660 dataset, were selected as candidate targets. A IncRNA-miRNA-gene network was established (Fig. 2A) with the aforementioned diverse bioinformatics tools (Fig. 2B). The expression levels of the five aforementioned miRNAs were validated using data from TCGA, revealing that three of the miRNAs were significantly differentially expressed between tumor tissues and adjacent non-tumor tissues; among these miRs, miR-96-5p was upregulated in tumor tissues, whereas miR-30b-3p and miR-627-3p were significantly downregulated in tumor tissues (Fig. 2C). The Kaplan-Meier plotter was used to perform survival analysis, and the results indicated that low expression levels of miR-96-5p predicted a favorable prognosis for patients with HCC (Fig. 2D). Although the expression



Figure 1. LINC01093 is downregulated in HCC tissues and positively associated with patient prognosis. Heat maps for (A) mRNAs and (B) lncRNAs deregulated in HCC vs. adjacent non-tumor tissues. (C) Expression of LINC01093 in HCC tissues and adjacent non-tumor tissues. (D) Expression of LINC01093 across normal organs and tissues. (E) Validation of LINC01093 expression in HCC tissues (n=369) and adjacent non-tumor tissues (n=50) using data from The Cancer Genome Atlas. The tumor tissues are presented on the left and the non-tumor tissues on the right. \*P<0.05. (F) Kaplan-Meier analysis of overall survival for patients with HCC with high vs. low LINC01093 expression. Dotted lines stand for the 95% CI for the HR. HCC, hepatocellular carcinoma; lncRNA, long non-coding RNA; LINC01093, long intergenic non-protein coding RNA 1093; T, tumor samples; NT, non-tumor tissue samples; HR, hazard ratio; RPKM, reads per kilobase per million mapped reads; TPM, transcripts per million.

levels were negatively associated with patient prognosis, miR-627-3p was not statistically considered a potential candidate miRNA (Fig. 2E). There was no significant association between the expression of miR-30b-3p and survival outcome (Fig. 2F). These results indicate that miR-96-5p potentially targets LINC01093. The results indicated that miR-96-5p has a tumor suppressor role in HCC by inhibiting LINC01093, and its upregulation is associated with a favorable prognosis regarding survival.

ZFAND5 is a candidate target of miR-96-5p. miRNA regulates gene expression in a negative manner; thus, high expression levels of miR-96-5p in the tumor tissue theoretically result in decreased expression of the target gene. To identify the potential targets, expression of previously predicted miR-96-5p target genes were analyzed and the results suggested that the expression levels of netrin 4 (NTN4) and ZFAND5 were lower in HCC tissues compared with those in the adjacent non-tumor tissues (Fig. 3A). LIM domain containing preferred translocation partner in lipoma (LPP) was significantly differentially expressed; however, its expression was upregulated in tumor tissues (Fig. 3A). These results were further validated using data from TCGA, confirming that NTN4 and ZFAND5 were downregulated in HCC tissues (Fig. 3B). NTN4 and ZFAND5 were further subjected to survival analysis, and the results demonstrated that only high expression of ZFAND5 predicted a favorable prognosis for patients with HCC (Fig. 3C), whereas high expression of NTN4 was not predicted to be significantly



Figure 2. LINC01093 is a potential target of miR-96-5p. (A) Regulatory network of LINC01093, miRNAs and mRNAs: Blue nodes, LINC01093; green nodes, miRNA; pink nodes, mRNA. (B) Flow chart for the *in silico* prediction of LINC01093-miRNA-mRNA pairs. (C) Validation of miRNA expression using data from The Cancer Genome Atlas. Kaplan-Meier analysis of overall survival for patients with hepatocellular carcinoma with high vs. low expression of (D) miR-96-5p, (E) miR-627-3p and (F) miR-30b-3p. miRNA, microRNA; hsa, *Homo sapiens*; LINC01093, long intergenic non-protein coding RNA 1093; T, tumor samples; NT, non-tumor tissue samples; HR, hazard ratio. NTN4, nerve guidance factor 4; APOC3, apolipoprotein C3; CCL16, C-C motif chemokine ligand 16; CYP1A2, cytochrome P450 1A2; FAM83F, family with sequence similarity 83; MSMO1, methylsterol monoxygenase 1; SIGLEC1, sialic acid binding Ig like lectin 1; SLC19A3, solute carrier family 19 member 3; SULT1E1, sulfotransferase family 1E member 1; TBXA2R, thromboxane A2 receptor; TMEM154, transmembrane protein 154; ZFAND5, zinc finger AN1-type containing 5; SMAD5, SMAD family member 5; POLR2D, RNA polymerase II subunit D; ZNF385, zinc finger protein 385A, HIF3A, hypoxia inducible factor 3 subunit α; ZNF286B, zinc finger protein 286B; B3GALT5, β-1,3-galactosyltransferase 5; BLOC1S3, biogenesis of lysosomal organelles complex 1 subunit 3; DST, dystonin; LPP, LIM domain containing preferred translocation partner in lipoma.

associated with improved prognosis (Fig. 3D). Therefore, ZFAND5 was selected for further analysis.

Functional analysis of the ZFAND5 interactome indicates the significance of NF-κB signaling. To further explore its function, the interactome of ZFAND5 was determined using BioGRID. This led to the identification of a total of 14 physical interactors with various experimental systems (Fig. 4A). Functional analysis demonstrated that the top 10 enriched GO terms were 'Fc-epsilon receptor signaling pathway', 'stimulatory C-type lectin receptor signaling pathway', 'nucleotide-binding oligomerization domain containing signaling pathway', 'T cell receptor signaling pathway', 'protein polyubiquitination', 'JNK cascade', 'inhibitor of NF-κB kinase (IKK)/NF-κB signaling', 'Wnt signaling pathway' and 'activation of mitogen-activated protein kinase (MAPK) activity' (Fig. 4B); while the most significantly enriched reactome pathways

were tumor necrosis factor receptor associated factor 6 (TRAF6)-mediated NF-kB activation'; MAPK kinase kinase 7 (TAK1) activates NF-κB by phosphorylation and activation of IKKs complex, interleukin-1 family signaling, C-type lectin domain containing 7A (CLEC7A) signaling, nerve growth factor receptor (p75NTR) recruits signaling complexes, NF-KB is activated and signals survival, neurotrophin receptor-interacting factor (NRIF) signals cell death from the nucleus, interleukin 1 receptor associated kinase 1 (IRAK1) recruits IKK complex upon TLR7/8 or -9 stimulation, IRAK1 recruits IKK complex, and downstream T-cell receptor (TCR) signaling (Fig. 4C). A network consisting of the genes involved in the top 10 GO terms was visualized using Cytoscape (Fig. 4D). Similarly, the reactome pathway network is presented in Fig. 4E. Colored nodes represent different GO terms or pathways and the node color represents enrichment significance. Combined GO and pathway analysis indicated that NF-KB signaling had a prominent significance.



Figure 3. ZFAND5 is a candidate target of miR-96-5p. (A) Expression of selected miR-96-5p target genes. (B) The aberrant expression of the target genes was validated using The Cancer Genome Atlas dataset. Kaplan-Meier analysis of overall survival for patients with hepatocellular carcinoma with high vs. low expression of (C) ZFAND5 and (D) NTN4. ZFAND5, zinc finger AN1-type containing 5; miRNA, microRNA; T, tumor samples; NT, non-tumor tissue samples; HR, hazard ratio; NTN4, netrin 4; LPP, LIM domain containing preferred translocation partner in lipoma.

# Discussion

HCC is notoriously difficult to treat successfully and is one of the leading causes of cancer-associated death worldwide. To date, the mechanisms underlying the initiation and progression of HCC have remained to be fully elucidated. IncRNAs have been indicated to regulate gene expression and have been implicated in various biological processes and human diseases. RNA-seq is a high-throughput sequencing technique that may reveal the presence and quantity of specific RNAs in a sample, which allows for the analysis of gene expression, alternative gene splicing and gene fusion. RNA-seq has been widely used to identify pathogenesis-associated genes, and to determine potential therapeutic targets in various diseases.

In the present study, the mRNA and lncRNA expression profiles between HCC tissues and adjacent non-tumor tissues from the GSE94660 dataset were compared and it was demonstrated that LINC01093 was significantly downregulated in tumor tissues. Analysis with BioProject indicated that LINC01093 was almost exclusively expressed in the normal adult liver. The expression of LINC01093 was then validated using data from TCGA, which confirmed its downregulation in HCC, and further survival analysis demonstrated that high expression levels of LINC01093 in HCC tissues were associated with a favorable clinical prognosis.

To understand the role of LINC01093 in hepatocellular carcinogenesis, miRNAs that target LINC01093 were predicted and screened. miR-30b-3p, miR-627-3p, miR-676-5p, miR-3689d and miR-96-5p were identified as potential candidates. The expression levels of these five miRNAs were validated using TCGA, and the results demonstrated that miR-30b-3p, miR-627-3p and miR-96-5p were significantly differentially expressed between tumor tissues and non-tumor tissues, but only miR-96-5p was upregulated in tumor tissues. Survival analysis of the three miRNAs was also performed and the results demonstrated that low expression levels of miR-96-5p predicted a favorable prognosis of patients. miR-96-5p has previously been recognized as an oncogenic miRNA in different types of cancer. In bladder cancer, studies revealed that miR-96-5p promoted tumor cell migration and invasion (29). In prostate cancer, it has been reported that miR-96-5p governed tumor progression and disease outcome by targeting a retinoic acid receptor  $\gamma$  network (30). In colorectal cancer, miR-96-5p was indicated to promote tumor invasion through inhibition of reversion-inducing cysteine-rich



Figure 4. Functional analyses of the ZFAND5 interactome. (A) ZFAND5 interactors identified by different experimental systems. (B) Gene Ontology analysis of ZFAND5 interactors in the category Biological Process. (C) Reactome pathway analysis of ZFAND5 interactors. Visualization of genes involved in (D) Biological Process terms and (E) reactome pathways. The grey nodes indicate genes and other colored nodes represent different GO terms or pathways. The node size and color represent enrichment significance; the larger the node, the higher enrichment significance of the pathway. ZFAND5, zinc finger AN1-type containing 5. SQSTM1, sequestosome 1; PSMD6, proteasome 26S subunit non-ATPase 6; IKBKG, inhibitor of nuclear factor  $\kappa$ B kinase regulatory subunit  $\gamma$ ; TRAF6, TNF receptor associated factor 6; UBC, ubiquitin C; APP, amyloid  $\beta$  precursor protein; TOMM34, translocase of outer mitochondrial membrane 34; SRP19, signal recognition particle 19; MAP3K1, mitogen-activated protein kinase kinase kinase 1; CETN1, centrin 1; SMURF1, SMAD specific E3 ubiquitin protein ligase 1; UBLCP1, ubiquitin like domain containing CTD phosphatase 1; ATP6V1H, ATPase H<sup>+</sup> transporting V1 subunit H; WIPF2, WAS/WASL interacting protein family member 2.

protein expression (31). In the present study, the results also suggested that miR-96-5p may promote HCC progression and to be associated with poor disease outcome.

Target genes of miR-96-5p were then predicted and screened with a cumulative weighted context score of  $\leq -0.6$ , and LPP, NTN4 and ZFAND5 were the identified as target genes among the differentially expressed genes identified in the GSE94660 dataset. These genes were therefore selected as candidate genes. miRNA regulates gene expression in a negative manner, and since miR-96-5p was upregulated in tumor tissues, downregulation of target genes was expected. The expression of the three candidate genes was analyzed, revealing that ZFAND5 and NTN4 had lower expression in tumor tissues compared with that in adjacent non-tumor tissues, whereas LPP was upregulated in HCC tissues, and these results were further confirmed in the TCGA dataset. ZFAD5 and NTN4 were subjected to survival analysis, which revealed that high mRNA expression levels of ZFAND5 were a favorable predictor of patient prognosis.

ZFAND5 is a member of the ZFAND family (28), and studies have revealed that ZFAND5 enhances protein degradation by activating the ubiquitin-proteasome system (32,33). To understand the potential role of ZFAND5 in HCC, proteins that physically interacted with ZFAND5 were obtained from BioGRID, which included 14 interactors identified by 7 different experimental systems. The ZFAND5-interactome was subsequently subjected to functional enrichment analysis. GO analysis revealed that the top 10 biological processes were 'Fc-epsilon receptor signaling pathway', 'stimulatory C-type lectin receptor signaling pathway', 'nucleotide-binding oligomerization domain containing signaling pathway', 'T cell receptor signaling pathway', 'MyD88-dependent TLR signaling pathway', 'protein polyubiquitination', 'JNK cascade', 'IKK/NF-κB signaling', 'Wnt signaling pathway' and 'activation of MAPK activity', while the most significantly enriched reactome pathways were 'TRAF6 mediated NF-KB activation', 'TAK1 activates NF-kB by phosphorylation and activation of IKKs complex', 'interleukin-1 family signaling', 'CLEC7A signaling', 'p75NTR recruits signaling complexes', 'NF-κB is activated and signals survival', 'NRIF signals cell death from the nucleus', 'IRAK1 recruits IKK complex upon TLR7/8 or -9 stimulation', 'IRAK1 recruits IKK complex' and 'downstream TCR signaling'. These results demonstrate that the NF-kB signaling is an important downstream pathway of ZFAND5.

NF-κB serves a well-known function in immune regulation and inflammatory responses, but growing evidence has additionally demonstrated its role in tumorigenesis (34,35). Therefore, NF-κB is frequently considered as the critical link between inflammation and cancer (35-37). It was reported that ZFAND5 inhibits NF-κB activation by interacting with IKK $\gamma$  (38). Therefore, the present study indicated the role of a LINC01093/miR96-5p/ZFAND5/NF-κB axis in the regulation of HCC development and progression, and further investigations are required to verify these results. Lack of experimental data is a limitation of the present study, and in a further study, *in vitro* and *in vivo* experiments will be performed to verify the conclusions that were obtained through the bioinformatics analysis. Through the use of bioinformatics analysis and *in vitro* and *in vivo* experiments, novel insight into the development of potential treatments for patients with HCC may be gained.

# Acknowledgements

Not applicable.

# Funding

This work was supported by the National Natural Science Foundation of China (grant nos. 81372654 and 81672848).

#### Availability of data and materials

The datasets analyzed during the current study are available in the GEO repository (https://www.ncbi.nlm.nih. gov/geo/query/acc.cgi?acc=GSE94660) and the BioProject repository (https://www.ncbi.nlm.nih.gov/bioproject?term=P RJEB4337&cmd=DetailsSearch).

## Authors' contributions

JZ designed the study and revised the manuscript. YZ and KY performed the GEO database analysis, analysed the data and wrote the manuscript. CH and LL performed bioinformatics analysis. HZ and MH assisted with the collection and analysis of other data. All authors read and approved the final manuscript.

## Ethics approval and consent to participate

Not applicable.

#### **Patient consent for publication**

Not applicable.

### **Competing interests**

The authors declare that they have no competing interests.

#### References

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 68: 394-424, 2018.
- Bodzin AS and Busuttil RW: Hepatocellular carcinoma: Advances in diagnosis, management, and long-term outcome. World J Hepatol 7: 1157-1167, 2015.
- Shi X, Sun M, Liu H, Yao Y and Song Y: Long non-coding RNAs: A new frontier in the study of human diseases. Cancer Lett 339: 159-166, 2013.
- Berindan-Neagoe I and Calin GA: Molecular pathways: MicroRNAs, cancer cells, and microenvironment. Clin Cancer Res 20: 6247-6253, 2014.
- Bartonicek N, Maag JL and Dinger ME: Long noncoding RNAs in cancer: Mechanisms of action and technological advancements. Mol Cancer 15: 43, 2016.
- Ng KW, Anderson C, Marshall EA, Minatel BC, Enfield KS, Saprunoff HL, Lam WL and Martinez VD: Piwi-interacting RNAs in cancer: Emerging functions and clinical utility. Mol Cancer 15: 5, 2016.
- Mannoor K, Liao J and Jiang F: Small nucleolar RNAs in cancer. Biochim Biophys Acta 1826: 121-128, 2012.

- 8. Holoch D and Moazed D: RNA-mediated epigenetic regulation of gene expression. Nat Rev Genet 16: 71-84, 2015.
- 9. Zhu C, Cheng D, Qiu X, Zhuang M and Liu Z: Long noncoding RNA SNHG16 promotes cell proliferation by sponging MicroRNA-205 and upregulating ZEB1 expression in osteosarcoma. Cell Physiol Biochem 51: 429-440, 2018.
- 10. Lan T, Ma W, Hong Z, Wu L, Chen X and Yuan Y: Long non-coding RNA small nucleolar RNA host gene 12 (SNHG12) promotes tumorigenesis and metastasis by targeting miR-199a/b-5p in hepatocellular carcinoma. J Exp Clin Cancer Res 36: 11, 2017.
- Dong J, Teng F, Guo W, Yang J, Ding G and Fu Z: IncRNA 11. SNHG8 promotes the tumorigenesis and metastasis by sponging miR-149-5p and predicts tumor recurrence in hepatocellular carcinoma. Cell Physiol Biochem 51: 2262-2274, 2018.
- 12. Tu J, Zhao Z, Xu M, Chen M, Weng Q, Wang J and Ji J: LINC00707 contributes to hepatocellular carcinoma progression via sponging miR-206 to increase CDK14. J Cell Physiol 234: 10615-10624, 2019.
- 13. Wang YG, Liu J, Shi M and Chen FX: IncRNA DGCR5 represses the development of hepatocellular carcinoma by targeting the miR-346/KLF14 axis. J Cell Physiol 234: 572-580, 2018.
- Chen W, You J, Zheng Q and Zhu YY: Downregulation of 14 IncRNA OGFRP1 inhibits hepatocellular carcinoma progression by AKT/mTOR and Wnt/ $\beta$ -catenin signaling pathways. Cancer Manag Res 10: 1817-1826, 2018.
- 15. Takahashi K, Yan I, Haga H and Patel T: Long noncoding RNA in liver diseases. Hepatology 60: 744-753, 2014.
- Cancer Genome Atlas Research Network, Weinstein JN, 16. Collisson EA, Mills GB, Shaw KR, Ozenberger BA, Ellrott K, Shmulevich I, Sander C and Stuart JM: The cancer genome atlas pan-cancer analysis project. Nat Genet 45: 1113-1120, 2013.
- 17. Edgar R, Domrachev M and Lash AE: Gene expression omnibus: NCBI gene expression and hybridization array data repository. Nucleic Acids Res 30: 207-210, 2002.
- 18. Tang Z, Li C, Kang B, Gao G, Li C and Zhang Z: GEPIA: A web server for cancer and normal gene expression profiling and interactive analyses. Nucleic Acids Res 45: W98-W102, 2017.
- Ota T, Suzuki Y, Nishikawa T, Otsuki T, Sugiyama T, Irie R, 19 Wakamatsu A, Hayashi K, Sato H, Nagai K, et al: Complete sequencing and characterization of 21,243 full-length human cDNAs. Nat Genet 36: 40-45, 2004.
- 20. Paraskevopoulou MD, Vlachos IS, Karagkouni D, Georgakilas G, Kanellos I, Vergoulis T, Zagganas K, Tsanakas P, Floros E, Dalamagas T and Hatzigeorgiou AG: DIANA-LncBase v2: Indexing microRNA targets on non-coding transcripts. Nucleic Acids Res 44: D231-D238, 2016.
- 21. Lewis BP, Burge CB and Bartel DP: Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. Cell 120: 15-20, 2005.
- 22. Smoot ME, Ono K, Ruscheinski J, Wang PL and Ideker T: Cytoscape 2.8: New features for data integration and network visualization. Bioinformatics 27: 431-432, 2011.
- 23. Oughtred R, Stark C, Breitkreutz BJ, Rust J, Boucher L, Chang C, Kolas N, O'Donnell L, Leung G, McAdam R, et al: The BioGRID interaction database: 2019 update. Nucleic Acids Res 47: D529-D541, 2019.
- 24. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, et al: Gene ontology: Tool for the unification of biology. The gene ontology consortium. Nat Genet 25: 25-29, 2000.

- 25. Huang DW, Sherman BT, Tan Q, Collins JR, Alvord WG, Roayaei J, Stephens R, Baseler MW, Lane HC and Lempicki RA: The DAVID gene functional classification tool: A novel biological module-centric algorithm to functionally analyze large gene lists. Genome Biol 8: R183, 2007.
- 26. Hou GX, Liu P, Yang J and Wen S: Mining expression and prognosis of topoisomerase isoforms in non-small-cell lung cancer by using Oncomine and Kaplan-Meier plotter. PLoS One 12: e0174515, 2017.
- 27. Mou Y, Wang D, Xing R, Nie H, Mou Y, Zhang Y and Zhou X: Identification of long noncoding RNAs biomarkers in patients with hepatitis B virus-associated hepatocellular carcinoma. Cancer Biomark 23: 95-106, 2018.
- 28. Dai M, Chen S, Wei X, Zhu X, Lan F, Dai S and Qin X: Diagnosis, prognosis and bioinformatics analysis of lncRNAs in hepatocellular carcinoma. Oncotarget 8: 95799-95809, 2017
- 29. He C, Zhang Q, Gu R, Lou Y and Liu W: miR-96 regulates migration and invasion of bladder cancer through epithelial-mesenchymal transition in response to transforming growth factor-β1. J Cell Biochem 119: 7807-7817, 2018.
- 30. Long MD, Singh PK, Russell JR, Llimos G, Rosario S, Rizvi A, van den Berg PR, Kirk J, Sucheston-Campbell LE, Smiraglia DJ and Campbell MJ: The miR-96 and RARy signaling axis governs androgen signaling and prostate cancer progression. Oncogene 38: 421-444, 2019.
- 31. Iseki Y, Shibutani M, Maeda K, Nagahara H, Fukuoka T, Matsutani S, Hirakawa K and Ohira M: MicroRNA-96 promotes tumor invasion in colorectal cancer via RECK. Anticancer Res 38: 2031-2035, 2018.
- 32. Lee D, Takayama S and Goldberg AL: ZFAND5/ZNF216 is an activator of the 26S proteasome that stimulates overall protein degradation. Proc Natl Acad Sci USA 115: E9550-E9559, 2018.
- 33. Hishiya A, Iemura S, Natsume T, Takayama S, Ikeda K and Watanabe K: A novel ubiquitin-binding protein ZNF216 func-tioning in muscle atrophy. EMBO J 25: 554-564, 2006.
- 34. Dolcet X, Llobet D, Pallares J and Matias-Guiu X: NF-kB in development and progression of human cancer. Virchows Arch 446: 475-482, 2005.
- 35. Marx J: Cancer research. Inflammation and cancer: The link grows stronger. Science 306: 966-968, 2004.
- 36. Karin M: NF-kappaB as a critical link between inflammation and cancer. Cold Spring Harb Perspect Biol 1: a000141, 2009.
- 37. Karin M: Nuclear factor-kappaB in cancer development and progression. Nature 441: 431-436, 2006.
- 38. Huang J, Teng L, Li L, Liu T, Li L, Chen D, Xu LG, Zhai Z and Shu HB: ZNF216 is an A20-like and IkappaB kinase gamma-interacting inhibitor of NFkappaB activation. J Biol Chem 279: 16847-16853, 2004.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.