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Review article

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The role of lncRNA in the pathogenesis of chronic obstructive pulmonary disease

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ABSTRACT

Chronic obstructive pulmonary disease (COPD) is characterized by progressive and irreversible airflow obstruction with abnormal lung function. Because its pathogenesis involves multiple aspects of oxidative stress, immunity and inflammation, apoptosis, airway and lung repair and destruction, the clinical approach to COPD treatment is not further updated. Therefore, it is crucial to discover a new means of COPD diagnosis and treatment. COPD etiology is associated with complex interactions between environmental and genetic determinants. Numerous genes are involved in the pathogenic process of this illness in research samples exposed to hazardous environmental conditions. Among them, Long non-coding RNAs (IncRNAs) have been reported to be involved in the molecular mechanisms of COPD development induced by different environmental exposures and genetic susceptibility encounters, and some potential lncRNA biomarkers have been identified as early diagnostic, disease course determination, and therapeutic targets for COPD. In this review, we summarize the expression profiles of the reported lncRNAs that have been reported in COPD studies related to environmental risk factors such as smoking and air pollution exposure and provided an overview of the roles of those lncRNAs in the pathogenesis of the disease.

1. Introduction

With the climate crisis, aging population, deepening industrialization, and changes in lifestyles around the world, people in both developing and developed countries are plagued by chronic respiratory diseases [1,2]. One of the most frequent chronic respiratory

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Fig. 1. Diagrammatic representation of COPD etiology and pathogenesis. Risk factors for COPD include smoking and air pollution, and so on. When irritants and toxicants are inhaled, the structural cells of the lung, such as epithelial cells, fibroblasts, and endothelial cells, get activated. When bronchial epithelial cells are damaged, alarm molecules (TSLP, IL-33, IL-25) are released, activating a variety of immune cells, endothelial cells, and fibroblasts. These cells release inflammatory mediators, which attract other inflammatory cells such as neutrophils, macrophages, and lymphocytes to the exposed area, resulting in persistent airway inflammation.

Table 1

Activation of immune cells in COPD patients.

Inflammatory Cells	Expression	Location	Downstream Targets	References
Neutrophils	+	COPD patients' bronchoalveolar lavage (BAL) fluid and sputum	CXCL8	[17]
Macrophages	+	COPD patients' sputum, BAL fluid, and lung parenchyma	cytokines (TNF- α), chemotactic factors [CXCL1, CXCL8, CCL2, LTB4] and ROS	[30]
Lymphocytes (CD8 ⁺ and CD4 ⁺)	+	COPD patients' lung parenchyma, bronchi, and bronchioles	IL-17A and IL-22	[31,32]
Mast Cells	+	human lung parenchyma	VEGF-A and FGF-2	[33,34]
Eosinophils	+	Stable COPD patients' airways and BAL fluid	VEGF-A, FGF-2, TNF- α , GM-CSF, nerve growth factor (NGF), and CXCL8	[35,36]
Basophils	+	COPD patient's lung tissue	VEGF-A and ANGPT1	[35,37]

disorders is chronic obstructive pulmonary disease (COPD). As of 2019, researchers predicted that over 391 million persons aged 30–79 globally have COPD, with a global incidence of 10.3 % [3]. In 2019, COPD was the third leading cause of death globally [4]. In China, the incidence and mortality rates of COPD are also high, making further research into COPD an urgent task for researchers [5,6].

Long non-coding RNAs (lncRNAs) are a class of linear RNA molecules that are longer than 200 nucleotides and have little to no capacity to code for proteins. Their structure is comparable to that of mRNA (with a 5' cap structure and a polyA tail at the 3' end in certain lncRNAs) [7,8]. Researchers' understanding of lncRNAs has increasingly expanded as high-throughput sequencing technologies and bioinformatics platforms have advanced. Growing evidence reveals that lncRNAs can influence gene expression at several levels, including epigenetic, transcriptional, and post-transcriptional regulation [7-11].

With the deepening of research, the presence of lncRNAs has been found in the plasma, serum, sputum, bronchial lavage fluid, and lung tissue of COPD patients [12–15]. Increasing evidence suggests that lncRNAs may become a new generation of biomarkers for COPD and have profound research significance. In this paper, we evaluated the lncRNAs engaged in COPD-related research that were produced by environmental variables such as smoking and air pollution, and we discussed their implications in the pathogenesis of COPD.

2. Overview and clinical management of COPD

Smoking and air pollution are the primary environmental causes of COPD, which is a disease that is brought on by environmental and genetic risk factors [3]. Simply calling it COPD may seem too general, as the abnormalities exhibited by COPD may be due to multiple pathophysiological factors [16] and have multiple endogenous phenotypes and clinical presentations [17]. It is currently



Fig. 2. Heat maps displaying the differential expression and hierarchical clustering of lncRNAs in (A) smokers with no COPD (SNC) compared with non-smokers without COPD (NS) and in (B) COPD patients compared with SNC. Reprinted with permission from Ref. [39]. Copyright (2015) Springer Link.

believed that at least several mechanisms are involved in the pathogenesis of COPD, including inflammatory responses, oxidative stress, and abnormal apoptosis of lung structural cells [18]. (Fig. 1). The activation of immune cells (Table 1) observed in various clinical trials of COPD and the abnormal apoptosis of lung structural cells all confirm this point. These mechanisms involve multiple signaling pathways, accelerating the natural physiological decline of the lungs that should occur with age, leading to persistent and irreversible airway obstruction [19].

Despite the heterogeneity of COPD, progressive and irreversible airflow limitation with abnormal lung function is a common feature of COPD [20,21].Clinical diagnosis approaches have not altered considerably as a result of this trait, and still rely nearly exclusively on post-bronchodilator spirometry, which is the most often used diagnostic strategy [22]. Although widely used, this approach also has its limitations, including insensitivity to early pathological changes, underutilization, frequent misinterpretation, and inability to predict symptoms. In addition, most of the treatment strategies for COPD use palliative care, that is, to directly improve airflow limitation. Inhaled medications, which include bronchodilators and corticosteroids, are the backbone of COPD management [23], especially in the acute exacerbation of COPD [24]. However, relieving airway limitation in AECOPD patients by drugs remains difficult. Furthermore, the side effects of these treatments must not be overlooked, such as the frequent use of inhaled corticosteroids, which increases the risk of osteoporosis, pneumonia, immunosuppression, and infection [25,26], considerably lowering patients' quality of life.

Alpha-1 antitrypsin deficiency which is the only known genetic cause of COPD [27]. It is worth mentioning that AAT intensive therapy is also a treatment method for COPD. Currently available intensive AAT therapies include lifestyle modification, asthma and COPD drug management, oxygen therapy, influenza vaccine and pneumococcal vaccine combination, and polysaccharides [28]. The only particular medication that has been approved for AAT-deficient patients who have symptoms of respiratory illness is augmentation therapy with intravenous pure AAT [29]. However, considering the cost of treatment and the limitations of the applicable population, intensive treatment with AAT has not been widely used.

Therefore, we need new biomarkers to assess the progression of COPD, identify and diagnose high-risk patients for treatment, and develop specific targeted drugs to improve the condition of COPD patients. Fortunately, with the development of molecular biology and the deepening understanding of lncRNAs, developing targeted drugs for treating COPD from the perspective of lncRNAs is no longer just a pipe dream.

3. The expression of lncRNA in COPD samples under different environmental risk factors

COPD is a complex disease characterized by hereditary and environmental risk factors. Cigarette smoking (CS) and air pollution are the most prevalent environmental pathogenic causes among the multiple environmental impacts [38]. Exposure to Cigarette smoke (CSE) is the major risk factor for induction of COPD. In a genome-wide lncRNA expression analysis study [39,40], RNA was extracted from lung tissues (all with lung cancer) from nonsmokers without COPD(NS), smokers without COPD(SNC), and smokers with COPD, and compared using microarray analysis, and the results were compared. It was found that compared to non-COPD smokers, COPD patients had 120 lncRNAs with high expression and 43 lncRNAs with low expression, with some lncRNAs possibly implicated in the control of the expression of cellular mediators and signaling pathways relevant to immune system alterations in COPD. (Fig. 2).

Smoking causes COPD in most patients, but it is now recognized that around 30 % of all COPD cases in the world occur in neversmokers [41]. The impact of air pollution cannot be ignored. Similar to CS-induced lung disease, air pollution also causes inflammation and oxidative stress. However, air pollution can cause more bronchial involvement, with less change in emphysema [42], whereas CS particles have a higher deposition rate in the lungs, which also explains why smoking is associated with emphysema [43]. In response to air pollution, researchers explored the effect of traffic-related PM2.5 (TAPM2.5) and wood smoke particles (WSPM2.5) on the expression of lncRNA in human bronchial epithelial cells (HBECs) and performed microarray analysis on the HBECs [44]. They



Fig. 3. Heat maps displaying the differential expression and hierarchical clustering of lncRNAs in (A) WSPM2.5 and (B) TAPM2.5 concentrations stimulation. Reprinted with permission from Ref. [44]. Copyright (2018) Portland Press.

discovered that in cells treated with WSPM2.5 and TAPM2.5, respectively, 94 and 1292 lncRNAs were upregulated and 203 and 1362 lncRNAs were downregulated in comparison to the control group. (Fig. 3). By comparing the differences in lncRNA expression between the two, we may be able to identify more different pathological responses in COPD patients under different environmental exposure conditions, which may lead to better-targeted treatments.

4. The role of lncRNA in COPD pathology under different environmental risk factors

Whether in vitro or clinical studies, researchers have found that many functional lncRNAs interact with COPD-related genes and perform multiple physiological functions involved in the pathogenesis of COPD through multiple pathways, including inflammatory response [45–49], oxidative stress [48,50,51] and apoptosis [45,48,52–54], etc. However, their functions in COPD disease progression under different environmental risk factors remain to be elucidated.

4.1. The role of lncRNA in the pathogenesis of CS-induced COPD

4.1.1. Cs-induced COPD inflammatory response and the role of lncRNA in it

COPD is a classic inflammatory disease, and as mentioned earlier, one of its pathogenetic features is the activation of inflammatory cells in the alveoli, including increased numbers of macrophages, neutrophils, and eosinophils. Typically, when inhaling harmful particles such as CS, inflammatory cells will enter the respiratory tract and lungs, leading to varying degrees of inflammation [55]. Although it is unclear how CSE affects the mechanisms of inflammation generation in COPD and what associations exist, CSE-induced changes in the expression of some lncRNAs have been found to be associated with the dysregulation of various immune cells in the lung, suggesting that CSE can act as a driving factor for lncRNA regulation of inflammation and play an indelible role.

LASI lncRNA expression is upregulated in the airways of CSE macaques and COPD smokers, which is associated with high mucus expression and mucus cell proliferation in severe COPD [56]. Mucus is the reaction product of innate and adaptive inflammatory responses made by the organism at the airway level to combat toxic gases and particles and is a complex viscoelastic gel [57]. The number and size of bronchial mucus glands, the amount of mucus in the airway epithelium, the proliferative activity of epithelial cells, and the development of squamous cell chemosis are all increased as a result of the infiltration of inflammatory cells into lung tissue and their involvement in the damage and repair of lung structural cells in COPD. The inflammatory exudate containing mucus obstructs the airway lumen and the repair process of the tissue thickens the wall, thus narrowing the diameter of the conducting airway [58], which is consistent with airflow limitation in COPD patients. Inc-ILTR expression is reduced in association with airway epithelial cell inflammation, suggesting impaired lung function, the pathogenesis of COPD, and predicting COPD progression [59].

It has now been shown that LINC00612 and HOXA-AS2 are significantly downregulated in COPD lung tissue. Regulation of HPMECs proliferation through Notch1 is involved in the pathogenesis of COPD, including CSE-induced apoptosis, inflammation, and oxidative stress in endothelial cells [48,60]. Most critically, Notch signaling has been linked to smoking and COPD and is essential for lung homeostasis, damage, and repair [61].

It is well documented that the lncRNA-miRNA-mRNA axis is important in the development of COPD. LncRNAs act as sponges for

Table 2 IncRNAs associated with COPD.

LncRNA	Expression	Location	Action	Targeted Pathway	Downstream Targets	References
ANRIL	Down	plasma of AECOPD patients	Anti-inflammatory	-	TNF, IL1B, IL17A, CXCL8	[12]
CCAT1	Up	lungs of COPD patients and CSE- treated HBE cells	MircoRNA sponging and epigenetic modification	miR-152-3p	-	[13]
GAS5	Up	Human lung fibroblast cell line	Involved in the regulation of pyroptosis	miR-223-3p	NLRP3	[15]
	Down	lung of mice exposed to CS	Suppress airway inflammation and fibroblast activation	miR-217-5p	PTEN	[64]
H19	Up	quadriceps of FFMI patients with COPD	Susceptibility to low FFMI	-	MYOD1	[65,66]
HOTAIR	Up	lungs of COPD patients and CSE- treated HPVEC	aggravate the apoptosis of CSE HPVEC	-	DNMT1	[67]
	Up	male BALB/c mice exposed to CS and CSE-treated HBE cells	-	-	IL6, CXCL8, CDH2, VIM, ACTA2, CDH1	[68]
IL7R	Down	serum from COPD patients	Protected against cellular senescence and apoptosis	-	EZH2 recruitment	[52,59, 69]
LASI	Up	CS-exposed macaques and airways of COPD smokers	Increase the mucoinflammatory response	_	secretory mucin MUC5AC. and inflammatory factors, ICAM-1, and IL-6	[56,70]
LUCAT1	Up	serum from COPD patients	Promoted CSE-induced 16HBE cells apoptosis and inhibited proliferation	miR- 181a- 5p	Wnt/β-catenin	[71,72]
MEG3	Up	lungs of COPD patients and CSE- treated HBE cells	Induces apoptosis and inflammation	Apoptosis	IL1B, IL6, TNF	[73]
MHC-R	Up	lungs of rats exposed to air pollution PM	Regulate the immune activities of DCs	-	GATA3	[74]
MIAT	Up	lung of mice exposed to CS	Knockdown of MIAT attenuates CS- induced airway remodeling	miR-29c-3p	HIF3A	[53]
TUG1	Up	lungs and sputum from smokers and non-smokers with COPD	Inhibits inflammation and airway remodeling	-	DUSP6	[75]
		TGFB1 treated BEAS-2B and HFL1 cells	Inhibits cell proliferation	-	ACTA2, FN1	[14]
SNHG5	Down	COPD tissues and CSE-treated HBE cells	Repress cell apoptosis, proliferation, and inflammation	miR-132	PTEN	[76]
LINC00612	Down	COPD tissues and CSE-treated HBE cells	Repress cell apoptosis, inflammation, and oxidative stress	miR-31-5p	Notch1	[48]
HOXA-AS2	Down	COPD tissues and CSE-treated HBE cells	Repress cell apoptosis	-	Notch1	[60]
MIR155HG	Up	Lung tissues of smokers without or with COPD	Contributed to the apoptosis and inflammation	miR-128-5p	BRD4	[54]
Nqo1-AS1	Up	The cytoplasm of mouse alveolar epithelium	Attenuated CS-induced oxidative stress	-	Nqo1	[50]
LINC00987	Up	COPD tissues	Repress cell apoptosis, inflammation, and oxidative stress	let-7b-5p	SIRT1	[51]
PVT1	Up	peripheral blood mononuclear cells (PBMCs), and serum of COPD patients	-	miR-146a	-	[77]
RP11- 86H7.1	Up	TRAPM2.5 exposed HBE cells	-	miR-9-5p	NFKB1/NF-ĸB	[47]
NEAT1	Up	plasma of COPD patients	-	miR-193a	-	[78]

miRNAs and influence gene expression by competitively binding them. As a ceRNA for miR18a-5p, serum CASC2 was much lower in COPD patients than in non-COPD smokers, and in 16HBE cells, overexpression of miR-18a-5p reversed CASC2's effects on apoptosis and inflammation [62]. Meanwhile, the expression of CCAT1 was significantly increased in CS-induced HBE cells [63], which has piqued the interest of researchers to explore deeper mechanisms between CCAT1 and COPD, CCAT1 was highly expressed in HBE cells from COPD patients treated with CSE, and CCAT1 knockdown alleviated the inflammatory response of HBE cells [13]. An abnormal increase of MIR155HG was also found in lung tissues of smokers. In smoking-associated COPD HPMECs, MIR155HG upregulates the miR-128-5p/BRD4 axis to promote apoptosis and inflammatory responses [54]. There are many more examples of the same, which are summarized in Table 2.

4.1.2. Cs-induced COPD oxidative stress and the role of lncRNA in it

CS contains a large number of harmful chemicals, including high concentrations of free radicals and oxidants [79,80]. These high concentrations of free radicals and exogenous oxidants are released in the respiratory tract and bronchial epithelium, producing large

amounts of reactive oxygen species (ROS). While this chemical has the advantage of being involved as a signaling component in maintaining intracellular homeostasis [81] but excessive intracellular accumulation may lead to oxidative imbalance, altered dynamic homeostasis, and subsequent cell death. Oxidative stress occurs when the production of ROS exceeds the levels allowed by the body's antioxidant defense mechanisms [82]. An association between oxidative stress and the skeletal muscle dysfunction common in COPD patients was proposed in an earlier study [79], that is, it may be caused by an imbalance in the redox environment of skeletal muscle. This suggests, in part, that the imbalance in the redox environment in COPD patients extends beyond the diseased lung and is likely to extend throughout the body.

Vascular endothelial growth factor (VEGF) may also be downregulated by oxidative stress [83]. Notably, it is frequently overexpressed in chronic inflammatory conditions [84], which is connected to a decline in lung function and the death of structural lung cells such alveolar endothelial cells [85].

As research progresses, the role of lncRNAs in oxidative stress in COPD is gradually being discussed. Among them, lncRNA Nqo1-AS1 was shown to lessen CS-induced oxidative stress by enhancing Nqo1 expression and boosting Nqo1 mRNA stability through antisense pairing with Nqo1 3'UTR because of its particular relationship with Nqo1, a multifunctional antioxidant enzyme that is essential for preventing oxidative damage to cells [50]. It has also been mentioned that LINC00612 [48], and LINC00987 [51]. However, unfortunately, there is a lack of data on the effects of COPD pathogenesis in vivo, which needs to be explored in more depth.

4.1.3. Cs-induced COPD in apoptosis and remodeling of lung structural cells and role of lncRNA in it

Among the diseases related to the lung, apoptosis is often considered to be related to emphysema and plays a crucial role in the course of COPD patients. It is inextricably linked to inflammation and oxidative stress. In CS-induced COPD samples, the initiation of apoptosis in lung structures is often accompanied by an inflammatory response and oxidative stress [67,86]. Low or short exposure to CS induces apoptosis, while high or prolonged exposure can lead to necrosis [87]. Although it is unclear from current research what role apoptosis has in the pathophysiology of COPD, it has been demonstrated that several lncRNAs are crucial in the imbalance between apoptosis and remodeling of lung structural cells in COPD samples exposed to CS. Bcl-2 is a widely accepted anti-apoptotic regulator, and it has been shown that reduced Bcl-2/Bax induces apoptosis through the release of cytoplasmic C from mitochondria release of cytoplasmic cytosolic C causes apoptosis [88]. Researchers discovered apoptosis in HPVEC cells from emphysema mice models and HPVEC from CSE-induced cells that had their expression of Bcl-2 downregulated and their levels of DNMT1, Bax, and cleaved cysteinase 3 upregulated [67]. Meanwhile, HOTAIR expression levels were upregulated in the lungs of COPD patients and CSE-induced HPVEC. In mice, HOTAIR knockdown could reduce emphysema and apoptosis through DNMT1-mediated hypermethylation of the Bcl-2 promoter.

The long non-coding RNA (lncRNA) growth arrest specific 5 (GAS5) is a well-known tumor suppressor linked to cell development and demise in a number of disorders. lncRNA GAS5 stimulates COPD inflammation by concentrating on the miR-223-3p/NLRP3 axis [15]. MIAT binds to miR-29c-3p and upregulates hypoxia-inducible factor 3α (HIF3A), which is involved in CS-induced EMT and COPD airway remodeling [53]. Knockdown of OIP5-AS1 promotes cell recovery from CSE injury by stimulating cell viability, regulating apoptosis, and alleviating inflammatory conditions via miR-410-3p and IL-13 [86]. By targeting the miR-18a-5p/IGF1 axis [62], overexpression of CASC2 may reduce inflammation and apoptosis in bronchial epithelial cells. These results suggest, at least in part, that CSE-induced dysregulation of lncRNAs contributes to the development of COPD by aberrantly regulating downstream targets, leading to an imbalance between apoptosis and replenishment of lung structural cells.

4.2. The role of lncRNA in the pathogenesis of air pollution-induced COPD

However, not all smokers develop COPD, and at least 20–30 % of COPD patients never smoke [89], which may be due to a variety of factors, but it is undeniable that air pollution must account for a large part of them. The pathogenic consequences of air pollution on the lung are comparable to those of CS in that they are mediated by inflammatory pathways and include oxidative stress and apoptosis of lung tissues once more [90]. Harmful airborne particles or gases are a major culprit in inducing COPD, especially when exposed to higher concentrations of particulate matter (PM), especially PM2.5, and are strongly associated with increased COPD prevalence and respiratory dysfunction [91].

As hazardous particles enter the airway, they initially make contact with bronchial epithelial cells. This causes alterations in the bronchial epithelial phenotype and prompts the release of significant amounts of cytokines and inflammatory mediators from these cells. This release sets off a chain of inflammatory events that worsens lung damage and airway inflammation. lncRNA RP11-86H7.1 promotes TRAPM2's inflammatory response. Since forced expression of lncRNARP11-86H7.1 decreases the production of miR-9-5p, which releases NFKB1 and maintains the activation of NF–B, 5 stimulated bronchial epithelial cells by functioning as a ceRNA for miR-9-5p [47]. With the upregulation of PM2.5 concentration in the air, lncRNA IL7R is exposed in normal cells, and although it is not known whether the persistent oxidative stress in COPD patients is involved in this response, it is possible to obtain the result that its expression is negatively correlated with the concentration of emphysema and PM2.5 [52]. Additionally, air pollution-exposed rats' lung tissue had considerably higher levels of the lncRNA MHC-R, with particular expression in immune-related cells, antigen presentation, and adaptive immune response [74].

Given the limited number of studies on the association between air pollution-induced lncRNAs and COPD, the mechanisms by which the environment affects differential lncRNA expression in COPD patients are unclear. However, as demonstrated above by the differential expression profiles of lncRNAs in air pollution³⁴, this is a neglected area that needs to be explored by more researchers.



Fig. 4. Triggers of COPD exacerbations and associated pathophysiological changes leading to increased exacerbation symptoms.

4.3. The role of lncRNA in AECOPD

Acute exacerbations of COPD(AECOPD) are known as exacerbations, and such episodes account for a large proportion of patient morbidity and mortality [92]. (Fig. 4). They have a prognosis that is comparable to many stage III or IV solid organ cancers and are linked to rapid loss of lung function, long-term quality of life degradation, and these symptoms [93]. However, the definition of acute exacerbations is sometimes ambiguous, and their intensity is determined more by the location of therapy than by the severity of a physiologic illness that underlies it. Chronic airway respiratory diseases, COPD and asthma have some degree of commonality in their course progression. And in terms of clinical practice, lnc-PVT1 has an excellent ability to differentiate between patients with severe asthma and non-severe asthma [94] The researchers studied lnc-PVT1 in relation to the commonalities between asthma and COPD and found that it can also differentiate COPD patients from HCs (healthy controls) and AECOPD patients from stable COPD patients [77].

In addition, similar to lnc-PVT1, lncRNAs involved in COPD disease course by targeting mRNAs are lnc-NEAT1 [78], whose expression is positively correlated with disease severity, while the expression of the miRNAs they target is inversely associated to the GOLD stage and inflammatory cytokine levels. Although LncRNA ANRIL was also used to predict disease susceptibility and risk of acute exacerbation in COPD, the difference was that lncRNA ANRIL expression was inversely linked with pro-inflammatory cytokines in AECOPD patients and stable COPD patients, and with GOLD stage in AECOPD patients, but not in stable COPD patients [12]. These findings suggest that these reported lncRNAs may be used not only to detect the severity of COPD but also as possible therapeutic targets for COPD.

5. The role of lncRNA in genetic susceptibility to COPD

The chronic obstructive pulmonary disease itself is a highly heterogeneous disease with complex and diverse etiologies. Studies have found that approximately 40 % of the variability in airflow limitation and up to 60 % of the risk of smoking-related COPD is associated with genetic variation [95]. Numerous genetic variations that affect COPD risk, decreased lung function, and other COPD-related traits have been discovered through genetic association studies [96]. Other pulmonary features are linked to these genetic variations. These genetic variations have been linked to various pulmonary and non-pulmonary features, indicating a genetic foundation for part of the COPD heterogeneity that significantly affects the overall risk of developing the disease [97].

Alpha-1 antitrypsin deficiency (AATD) was the first genetic risk factor for chronic obstructive pulmonary disease (COPD) described [98] and is inherited in an autosomal co-dominant fashion. The alpha-1 antitrypsin molecule is a serine protease inhibitor that is predominantly produced in the liver. Its most important physiologic functions are the protection of pulmonary tissue from aggressive proteolytic enzymes and regulation of pulmonary immune processes [99]. At present, some researchers have reviewed the role of



Fig. 5. MicroRNAs may be captured by lncRNAs like a sponge. By attaching to these microRNAs, they stop microRNAs from attaching to their target mRNAs, eliminating post-transcriptional control.

IncRNA before and after intensive treatment of AATD [100].

However, as research has progressed, COPD has been considered to be a self-induced disease resulting from external stimuli in genetically susceptible individuals. Recent studies have shown the upregulation of p16 expression, a marker of cellular senescence, in CSE-stimulated endothelial progenitor cells (EPC) and in COPD patients [101]. Cellular senescence is involved in the development of COPD, i.e. we can deduce the role of lncRNA in genetic susceptibility to COPD by reversing the genes associated with cellular senescence. IL7R is associated with cellular senescence-associated genes p16^{INK4a} and p21^{CIP1/WAF1}, and surprisingly Lnc-IL7R levels are decreased in COPD patients, whereas higher lnc-IL7R expression can prevent cellular senescence and apoptosis, while lower lnc-IL7R expression exacerbated PM2.5-treated cellular damage [69]. (Fig. 5).

6. Summary

A complex illness, COPD is brought on by hereditary and environmental risk factors. Few studies, however, have been able to identify the pathophysiology and susceptibility of COPD. The diagnosis of lung illness is receiving more attention as a result of COVID-19's continued and widespread occurrence. One of the most common comorbidities of COVID-19 disease progression is COPD [102]. Clinical symptoms of COPD vary, and it might be challenging to identify any apparent patterns. Determining the pathophysiology of COPD and developing precise diagnosis techniques and preventative plans are therefore crucial and urgent concerns for the present and the future.

Numerous in vitro and in vivo studies have demonstrated that environmental exposures like CS and air pollutants can disrupt the expression of lncRNAs in various samples from COPD and non-COPD patients, interfering with their regulation of COPD-related genes in crucial signaling pathways in COPD pathogenesis, including the imbalance between inflammatory cell entry into the lung, oxidative stress, and apoptosis as well as remodeling of lung structures. Yet, while having comparable environmental risk factors, each individual may experience clinical symptoms differently because of their individuality and genetic predisposition.

In conclusion, lncRNA is a molecule with far-reaching effects and has an important position in transcriptional regulation. Unfortunately, however, their relationship with COPD lacks more literature, and more studies revealing the biological functions of lncRNAs in COPD or revealing the pathogenesis of COPD would generate more scientific interest among researchers.

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Data availability statement

No data was used for the research described in the article.

CRediT authorship contribution statement

Jing Xie: Writing – review & editing, Writing – original draft. Yongkang Wu: Visualization, Investigation. Qing Tao: Investigation. Hua Liu: Supervision. Jingjing Wang: Investigation. Chunwei Zhang: Investigation. Yuanzhi Zhou: Investigation. Chengyan Wei: Investigation. Yan Chang: Supervision, Resources. Yong Jin: Writing – review & editing, Supervision, Resources. Zhen Ding: Writing – review & editing, Supervision, Resources, Project administration.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- [1] P. Martins, J. Rosado-Pinto, M. do Céu Teixeira, N. Neuparth, O. Silva, H. Tavares, J.L. Spencer, D. Mascarenhas, A.L. Papoila, N. Khaltaev, I. Annesi-Maesano, Under-report and underdiagnosis of chronic respiratory diseases in an African country, Allergy 64 (2009) 1061–1067, https://doi.org/10.1111/j.1398-9995.2009.01956.x.
- [2] J. Bai, Y. Zhao, D. Yang, Y. Ma, C. Yu, Secular trends in chronic respiratory diseases mortality in Brazil, Russia, China, and South Africa: a comparative study across main BRICS countries from 1990 to 2019, BMC Publ. Health 22 (2022) 91, https://doi.org/10.1186/s12889-021-12484-z.
- [3] D. Adeloye, P. Song, Y. Zhu, H. Campbell, A. Sheikh, I. Rudan, NIHR RESPIRE Global Respiratory Health Unit, Global, regional, and national prevalence of, and risk factors for, chronic obstructive pulmonary disease (COPD) in 2019: a systematic review and modelling analysis, Lancet Respir. Med. 10 (2022) 447–458, https://doi.org/10.1016/S2213-2600(21)00511-7.
- [4] Global health estimates, Leading causes of death. https://www.who.int/data/gho/data/themes/mortality-and-global-health-estimates/ghe-leading-causes-ofdeath, 2023. (Accessed 19 September 2023).
- [5] C. Wang, J. Xu, L. Yang, Y. Xu, X. Zhang, C. Bai, J. Kang, P. Ran, H. Shen, F. Wen, K. Huang, W. Yao, T. Sun, G. Shan, T. Yang, Y. Lin, S. Wu, J. Zhu, R. Wang, Z. Shi, J. Zhao, X. Ye, Y. Song, Q. Wang, Y. Zhou, L. Ding, T. Yang, Y. Chen, Y. Guo, F. Xiao, Y. Lu, X. Peng, B. Zhang, D. Xiao, C.-S. Chen, Z. Wang, H. Zhang, X. Bu, X. Zhang, L. An, S. Zhang, Z. Cao, Q. Zhan, Y. Yang, B. Cao, H. Dai, L. Liang, J. He, China Pulmonary Health Study Group, Prevalence and risk factors of chronic obstructive pulmonary disease in China (the China Pulmonary Health [CPH] study): a national cross-sectional study, Lancet 391 (2018) 1706–1717, https://doi.org/10.1016/S0140-6736(18)30841-9.
- [6] M. Zhou, H. Wang, X. Zeng, P. Yin, J. Zhu, W. Chen, X. Li, L. Wang, L. Wang, Y. Liu, J. Liu, M. Zhang, J. Qi, S. Yu, A. Afshin, E. Gakidou, S. Glenn, V.S. Krish, M. K. Miller-Petrie, W.C. Mountjoy-Venning, E.C. Mullany, S.B. Redford, H. Liu, M. Naghavi, S.I. Hay, L. Wang, C.J.L. Murray, X. Liang, Mortality, morbidity, and risk factors in China and its provinces, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017, Lancet 394 (2019) 1145–1158, https://doi.org/10.1016/S0140-6736(19)30427-1.
- [7] L. Statello, C.-J. Guo, L.-L. Chen, M. Huarte, Gene regulation by long non-coding RNAs and its biological functions, Nat. Rev. Mol. Cell Biol. 22 (2021) 96–118, https://doi.org/10.1038/s41580-020-00315-9.
- [8] M.C. Bridges, A.C. Daulagala, A. Kourtidis, LNCcation: lncRNA localization and function, J. Cell Biol. 220 (2021), e202009045, https://doi.org/10.1083/ jcb.202009045.
- [9] F. K, Jt M, Functional classification and experimental dissection of long noncoding RNAs, Cell 172 (2018), https://doi.org/10.1016/j.cell.2018.01.011.
- [10] N. Mukherjee, L. Calviello, A. Hirsekorn, S. de Pretis, M. Pelizzola, U. Ohler, Integrative classification of human coding and noncoding genes through RNA metabolism profiles, Nat. Struct. Mol. Biol. 24 (2017) 86–96, https://doi.org/10.1038/nsmb.3325.
- [11] K. Arman, Z. Dalloul, E. Bozgeyik, Emerging role of microRNAs and long non-coding RNAs in COVID-19 with implications to therapeutics, Gene 861 (2023), 147232, https://doi.org/10.1016/j.gene.2023.147232.
- [12] J. Ge, S. Geng, H. Jiang, Long noncoding RNAs antisense noncoding RNA in the INK4 locus (ANRIL) correlates with lower acute exacerbation risk, decreased inflammatory cytokines, and mild GOLD stage in patients with chronic obstructive pulmonary disease, J. Clin. Lab. Anal. 33 (2019), e22678, https://doi.org/ 10.1002/jcla.22678.
- [13] D. Zong, X. Liu, J. Li, Y. Long, R. Ouyang, Y. Chen, LncRNA-CCAT1/miR-152-3p is involved in CSE-induced inflammation in HBE cells via regulating ERK signaling pathway, Int Immunopharmacol 109 (2022), 108818, https://doi.org/10.1016/j.intimp.2022.108818.
- [14] W. Tang, Z. Shen, J. Guo, S. Sun, Screening of long non-coding RNA and TUG1 inhibits proliferation with TGF-β induction in patients with COPD, Int. J. Chronic Obstr. Pulm. Dis. 11 (2016) 2951–2964, https://doi.org/10.2147/COPD.S109570.
- [15] R. Mo, J. Li, Y. Chen, Y. Ding, IncRNA GAS5 promotes pyroptosis in COPD by functioning as a ceRNA to regulate the miR-223-3p/NLRP3 axis, Mol. Med. Rep. 26 (2022) 219, https://doi.org/10.3892/mmr.2022.12735.
- [16] K.F. Rabe, H. Watz, Chronic obstructive pulmonary disease, Lancet 389 (2017) 1931–1940, https://doi.org/10.1016/S0140-6736(17)31222-9.
- [17] Ae J, Wj M, E. S, Gm W, Understanding the role of neutrophils in chronic inflammatory airway disease, F1000Research 8 (2019), https://doi.org/10.12688/ f1000research.18411.1.
- [18] I.K. Demedts, T. Demoor, K.R. Bracke, G.F. Joos, G.G. Brusselle, Role of apoptosis in the pathogenesis of COPD and pulmonary emphysema, Respir. Res. 7 (2006) 53, https://doi.org/10.1186/1465-9921-7-53.
- [19] C.F. Vogelmeier, G.J. Criner, F.J. Martinez, A. Anzueto, P.J. Barnes, J. Bourbeau, B.R. Celli, R. Chen, M. Decramer, L.M. Fabbri, P. Frith, D.M.G. Halpin, M. V. López Varela, M. Nishimura, N. Roche, R. Rodriguez-Roisin, D.D. Sin, D. Singh, R. Stockley, J. Vestbo, J.A. Wedzicha, A. Agusti, Global strategy for the diagnosis, management, and prevention of chronic obstructive lung disease 2017 report: GOLD executive summary, Eur. Respir. J. 49 (2017), 1700214, https://doi.org/10.1183/13993003.00214-2017.
- [20] R.A. Holloway, L.E. Donnelly, Immunopathogenesis of chronic obstructive pulmonary disease, Curr. Opin. Pulm. Med. 19 (2013) 95–102, https://doi.org/ 10.1097/MCP.0b013e32835cfff5.
- [21] G.G. Brusselle, G.F. Joos, K.R. Bracke, New insights into the immunology of chronic obstructive pulmonary disease, Lancet 378 (2011) 1015–1026, https://doi. org/10.1016/S0140-6736(11)60988-4.
- [22] İ. Şerifoğlu, G. Ulubay, The methods other than spirometry in the early diagnosis of COPD, Tuberk Toraks 67 (2019) 63–70, https://doi.org/10.5578/tt.68162.
- [23] W.W. Labaki, S.R. Rosenberg, Chronic obstructive pulmonary disease, Ann. Intern. Med. 173 (2020) ITC17–ITC32, https://doi.org/10.7326/AITC202008040.
- [24] P. Guo, R. Li, T.H. Piao, C.L. Wang, X.L. Wu, H.Y. Cai, Pathological mechanism and targeted drugs of COPD, Int. J. Chronic Obstr. Pulm. Dis. 17 (2022) 1565–1575, https://doi.org/10.2147/COPD.S366126.
- [25] C. Janson, K. Lisspers, B. Ställberg, G. Johansson, F.S. Gutzwiller, K. Mezzi, L. Mindeholm, B.K. Bjerregaard, L. Jorgensen, K. Larsson, Osteoporosis and fracture risk associated with inhaled corticosteroid use among Swedish COPD patients: the ARCTIC study, Eur. Respir. J. 57 (2021), 2000515, https://doi.org/ 10.1183/13993003.00515-2020.
- [26] D. Price, B. Yawn, G. Brusselle, A. Rossi, Risk-to-benefit ratio of inhaled corticosteroids in patients with COPD, Prim. Care Respir. J. 22 (2013) 92–100, https:// doi.org/10.4104/pcrj.2012.00092.

- [27] S.G. Pillai, D. Ge, G. Zhu, X. Kong, K.V. Shianna, A.C. Need, S. Feng, C.P. Hersh, P. Bakke, A. Gulsvik, A. Ruppert, K.C. Lødrup Carlsen, A. Roses, W. Anderson, S.I. Rennard, D.A. Lomas, E.K. Silverman, D.B. Goldstein, ICGN Investigators, A genome-wide association study in chronic obstructive pulmonary disease (COPD): identification of two major susceptibility loci, PLoS Genet. 5 (2009), e1000421, https://doi.org/10.1371/journal.pgen.1000421.
- [28] M. Cazzola, D. Stolz, P. Rogliani, M.G. Matera, α1-Antitrypsin deficiency and chronic respiratory disorders, Eur. Respir. Rev. 29 (2020), 190073, https://doi. org/10.1183/16000617.0073-2019.
- [29] A.R. Tonelli, M.L. Brantly, Augmentation therapy in alpha-1 antitrypsin deficiency: advances and controversies, Ther. Adv. Respir. Dis. 4 (2010) 289–312, https://doi.org/10.1177/1753465810373911.
- [30] K. Yamasaki, S.F. van Eeden, Lung macrophage phenotypes and functional responses: role in the pathogenesis of COPD, Int. J. Mol. Sci. 19 (2018) E582, https://doi.org/10.3390/ijms19020582.
- [31] M. Williams, I. Todd, L.C. Fairclough, The role of CD8 + T lymphocytes in chronic obstructive pulmonary disease: a systematic review, Inflamm. Res. 70 (2021) 11–18, https://doi.org/10.1007/s00011-020-01408-z.
- [32] M. Saetta, A. Di Stefano, G. Turato, F.M. Facchini, L. Corbino, C.E. Mapp, P. Maestrelli, A. Ciaccia, L.M. Fabbri, CD8+ T-lymphocytes in peripheral airways of smokers with chronic obstructive pulmonary disease, Am. J. Respir. Crit. Care Med. 157 (1998) 822–826, https://doi.org/10.1164/ajrccm.157.3.9709027.
- [33] A. Soltani, Y.P. Ewe, Z.S. Lim, S.S. Sohal, D. Reid, S. Weston, R. Wood-Baker, E.H. Walters, Mast cells in COPD airways: relationship to bronchodilator responsiveness and angiogenesis, Eur. Respir. J. 39 (2012) 1361–1367, https://doi.org/10.1183/09031936.00084411.
- [34] J. Boesiger, M. Tsai, M. Maurer, M. Yamaguchi, L.F. Brown, K.P. Claffey, H.F. Dvorak, S.J. Galli, Mast cells can secrete vascular permeability factor/vascular endothelial cell growth factor and exhibit enhanced release after immunoglobulin E-dependent upregulation of fc epsilon receptor I expression, J. Exp. Med. 188 (1998) 1135–1145, https://doi.org/10.1084/jem.188.6.1135.
- [35] P. Jogdand, P. Siddhuraj, M. Mori, C. Sanden, J. Jönsson, A.F. Walls, J. Kearley, A.A. Humbles, R. Kolbeck, L. Bjermer, P. Newbold, J.S. Erjefält, Eosinophils, basophils and type 2 immune microenvironments in COPD-affected lung tissue, Eur. Respir. J. 55 (2020), 1900110, https://doi.org/10.1183/ 13993003.00110-2019.
- [36] D.P. Tashkin, M.E. Wechsler, Role of eosinophils in airway inflammation of chronic obstructive pulmonary disease, Int. J. Chronic Obstr. Pulm. Dis. 13 (2018) 335–349, https://doi.org/10.2147/COPD.S152291.
- [37] A. de Paulis, N. Prevete, I. Fiorentino, F.W. Rossi, S. Staibano, N. Montuori, P. Ragno, A. Longobardi, B. Liccardo, A. Genovese, D. Ribatti, A.F. Walls, G. Marone, Expression and functions of the vascular endothelial growth factors and their receptors in human basophils, J. Immunol. 177 (2006) 7322–7331, https://doi.org/10.4049/jimmunol.177.10.7322.
- [38] D.M. Mannino, A.S. Buist, Global burden of COPD: risk factors, prevalence, and future trends, Lancet 370 (2007) 765–773, https://doi.org/10.1016/S0140-6736(07)61380-4.
- [39] H. Bi, J. Zhou, D. Wu, W. Gao, L. Li, L. Yu, F. Liu, M. Huang, I.M. Adcock, P.J. Barnes, X. Yao, Microarray analysis of long non-coding RNAs in COPD lung tissue, Inflamm. Res. 64 (2015) 119–126, https://doi.org/10.1007/s00011-014-0790-9.
- [40] H. Zhang, D. Sun, D. Li, Z. Zheng, J. Xu, X. Liang, C. Zhang, S. Wang, J. Wang, W. Lu, Long non-coding RNA expression patterns in lung tissues of chronic cigarette smoke induced COPD mouse model, Sci. Rep. 8 (2018) 7609, https://doi.org/10.1038/s41598-018-25702-3.
- [41] C. Rodríguez García, A. Ruano-Ravina, M. Pérez Ríos, L. Martín Gisbert, L. Varela-Lema, C. Candal-Pedreira, C. Represas-Represas, J. Rey-Brandariz, L. Valdés-Cuadrado, A. Agustí, Clinical characteristics of chronic obstructive pulmonary disease in never-smokers: a systematic review, Respir. Med. 214 (2023), 107284, https://doi.org/10.1016/j.rmed.2023.107284.
- [42] F. Aghaeimeybodi, G. Samadzadeh, Z. Haji Safari, S. Nouri, H.R. Talebi, S.H. Shahcheraghi, Comparison of chronic obstructive pulmonary diseases induced by wood smoke and tobacco smoke, Tanaffos 20 (2021) 268–276.
- [43] L. Nicolaou, W. Checkley, Differences between cigarette smoking and biomass smoke exposure: an in silico comparative assessment of particulate deposition in the lungs, Environ. Res. 197 (2021), 111116, https://doi.org/10.1016/j.envres.2021.111116.
- [44] X. Li, M. Zheng, J. Pu, Y. Zhou, W. Hong, X. Fu, Y. Peng, W. Zhou, H. Pan, B. Li, P. Ran, Identification of abnormally expressed lncRNAs induced by PM2.5 in human bronchial epithelial cells, Biosci. Rep. 38 (2018), BSR20171577, https://doi.org/10.1042/BSR20171577.
- [45] P. Chen, P. Jiang, J. Chen, Y. Yang, X. Guo, XIST promotes apoptosis and the inflammatory response in CSE-stimulated cells via the miR-200c-3p/EGR3 axis, BMC Pulm. Med. 21 (2021) 215, https://doi.org/10.1186/s12890-021-01582-8.
- [46] M. Wang, Y. Liu, Y. Zhang, L. Zhang, LncRNA LOC729178 acts as a sponge of miR-144-3p to mitigate cigarette smoke extract-induced inflammatory injury via regulating PHLPP2 in 16HBE cells, J. Mol. Histol. 52 (2021) 437–447, https://doi.org/10.1007/s10735-021-09972-2.
- [47] J. Zhao, J. Pu, B. Hao, L. Huang, J. Chen, W. Hong, Y. Zhou, B. Li, P. Ran, LncRNA RP11-86H7.1 promotes airway inflammation induced by TRAPM2.5 by acting as a ceRNA of miRNA-9-5p to regulate NFKB1 in HBECS, Sci. Rep. 10 (2020), 11587, https://doi.org/10.1038/s41598-020-68327-1.
- [48] J. Luo, L. Li, D. Hu, X. Zhang, LINC00612/miR-31-5p/Notch1 Axis regulates apoptosis, inflammation, and oxidative stress in human pulmonary microvascular endothelial cells induced by cigarette smoke extract, Int. J. Chronic Obstr. Pulm. Dis. 15 (2020) 2049–2060, https://doi.org/10.2147/COPD.S255696.
- [49] J. Mei, Y. Zhang, S. Lu, J. Wang, Long non-coding RNA NNT-AS1 regulates proliferation, apoptosis, inflammation and airway remodeling of chronic obstructive pulmonary disease via targeting miR-582-5p/FBXO11 axis, Biomed. Pharmacother. 129 (2020), 110326, https://doi.org/10.1016/j. biopha.2020.110326.
- [50] H. Zhang, R. Guan, Z. Zhang, D. Li, J. Xu, Y. Gong, X. Chen, W. Lu, LncRNA nqo1-AS1 attenuates cigarette smoke-induced oxidative stress by upregulating its natural antisense transcript Nqo1, Front. Pharmacol. 12 (2021), 729062, https://doi.org/10.3389/fphar.2021.729062.
- [51] Y. Wang, J. Chen, W. Chen, L. Liu, M. Dong, J. Ji, D. Hu, N. Zhang, LINC00987 ameliorates COPD by regulating LPS-induced cell apoptosis, oxidative stress, inflammation and autophagy through let-7b-5p/SIRT1 Axis, Int. J. Chronic Obstr. Pulm. Dis. 15 (2020) 3213–3225, https://doi.org/10.2147/COPD.S276429.
- [52] K.-Y. Lee, S.-C. Ho, W.-L. Sun, P.-H. Feng, C.-W. Lin, K.-Y. Chen, H.-C. Chuang, C.-H. Tseng, T.-T. Chen, S.-M. Wu, Lnc-IL7R alleviates PM2.5-mediated cellular senescence and apoptosis through EZH2 recruitment in chronic obstructive pulmonary disease, Cell Biol. Toxicol. (2022), https://doi.org/10.1007/s10565-022-09709-1.
- [53] W. Gu, L. Wang, G. Deng, X. Gu, Z. Tang, S. Li, W. Jin, J. Yang, X. Guo, Q. Li, Knockdown of long noncoding RNA MIAT attenuates cigarette smoke-induced airway remodeling by downregulating miR-29c-3p-HIF3A axis, Toxicol. Lett. 357 (2022) 11–19, https://doi.org/10.1016/j.toxlet.2021.12.014.
- [54] J. Song, Q. Wang, L. Zong, LnCRNA MIR155HG contributes to smoke-related chronic obstructive pulmonary disease by targeting miR-128-5p/BRD4 axis, Biosci. Rep. 40 (2020), BSR20192567, https://doi.org/10.1042/BSR20192567.
- [55] P.J. Barnes, Cellular and molecular mechanisms of asthma and COPD, Clin. Sci. 131 (2017) 1541–1558, https://doi.org/10.1042/CS20160487.
- [56] M. Manevski, D. Devadoss, C. Long, S.P. Singh, M.W. Nasser, G.M. Borchert, M.N. Nair, I. Rahman, M. Sopori, H.S. Chand, Increased expression of LASI IncRNA regulates the cigarette smoke and COPD associated airway inflammation and mucous cell hyperplasia, Front. Immunol. 13 (2022), 803362, https://doi.org/ 10.3389/fimmu.2022.803362.
- [57] R. Pangeni, T. Meng, S. Poudel, D. Sharma, H. Hutsell, J. Ma, B.K. Rubin, W. Longest, M. Hindle, Q. Xu, Airway mucus in pulmonary diseases: muco-adhesive and muco-penetrating particles to overcome the airway mucus barriers, Int. J. Pharm. 634 (2023), 122661, https://doi.org/10.1016/j.ijpharm.2023.122661.
- [58] I. Cerveri, V. Brusasco, Revisited role for mucus hypersecretion in the pathogenesis of COPD, Eur. Respir. Rev. 19 (2010) 109–112, https://doi.org/10.1183/ 09059180.00002710.
- [59] O.A. Bamodu, S.-M. Wu, P.-H. Feng, W.-L. Sun, C.-W. Lin, H.-C. Chuang, S.-C. Ho, K.-Y. Chen, T.-T. Chen, C.-H. Tseng, W.-T. Liu, K.-Y. Lee, Inc-IL7R expression reflects physiological pulmonary function and its aberration is a putative indicator of COPD, Biomedicines 10 (2022) 786, https://doi.org/10.3390/ biomedicines10040786.
- [60] A.-Y. Zhou, Y.-Y. Zhao, Z.-J. Zhou, J.-X. Duan, Y.-Z. Zhu, S. Cai, P. Chen, Microarray analysis of long non-coding RNAs in lung tissues of patients with COPD and HOXA-AS2 promotes HPMECs proliferation via Notch1, Int. J. Chronic Obstr. Pulm. Dis. 15 (2020) 2449–2460, https://doi.org/10.2147/COPD.S259601.
- [61] D. Zong, R. Ouyang, J. Li, Y. Chen, P. Chen, Notch signaling in lung diseases: focus on Notch1 and Notch3, Ther. Adv. Respir. Dis. 10 (2016) 468–484, https:// doi.org/10.1177/1753465816654873.

- [62] P. Liu, H. Zhang, H. Zeng, Y. Meng, H. Gao, M. Zhang, L. Zhao, LncRNA CASC2 is involved in the development of chronic obstructive pulmonary disease via targeting miR-18a-5p/IGF1 axis, Ther. Adv. Respir. Dis. 15 (2021), 17534666211028072, https://doi.org/10.1177/17534666211028072.
- [63] L. Lu, H. Xu, F. Luo, X. Liu, X. Lu, Q. Yang, J. Xue, C. Chen, L. Shi, Q. Liu, Epigenetic silencing of miR-218 by the lncRNA CCAT1, acting via BMI1, promotes an altered cell cycle transition in the malignant transformation of HBE cells induced by cigarette smoke extract, Toxicol. Appl. Pharmacol. 304 (2016) 30–41, https://doi.org/10.1016/j.taap.2016.05.012.
- [64] Y. Du, Y. Ding, T. Shi, W. He, J. Feng, Z. Mei, X. Chen, X. Feng, X. Zhang, Z. Jie, Long noncoding RNA GAS5 attenuates cigarette smoke-induced airway remodeling by regulating miR-217-5p/PTEN axis, Acta Biochim. Biophys. Sin. 54 (2022) 1–9, https://doi.org/10.3724/abbs.2022074.
- [65] A. Lewis, J.Y. Lee, A.V. Donaldson, S.A. Natanek, S. Vaidyanathan, W.D.-C. Man, N.S. Hopkinson, A.A. Sayer, H.P. Patel, C. Cooper, H. Syddall, M.I. Polkey, P. R. Kemp, Increased expression of H19/miR-675 is associated with a low fat-free mass index in patients with COPD, J Cachexia Sarcopenia Muscle 7 (2016) 330–344, https://doi.org/10.1002/jcsm.12078.
- [66] Z.-H. Zheng, D.-M. Wu, S.-H. Fan, Z.-F. Zhang, G.-Q. Chen, J. Lu, Upregulation of miR-675-5p induced by lncRNA H19 was associated with tumor progression and development by targeting tumor suppressor p53 in non-small cell lung cancer, J. Cell. Biochem. 120 (2019) 18724–18735, https://doi.org/10.1002/ jcb.29182.
- [67] Z. Dai, X. Liu, H. Zeng, Y. Chen, Long noncoding RNA HOTAIR facilitates pulmonary vascular endothelial cell apoptosis via DNMT1 mediated hypermethylation of Bcl-2 promoter in COPD, Respir. Res. 23 (2022) 356, https://doi.org/10.1186/s12931-022-02234-z.
- [68] H. Xia, J. Xue, H. Xu, M. Lin, M. Shi, Q. Sun, T. Xiao, X. Dai, L. Wu, J. Li, Q. Xiang, H. Tang, Q. Bian, Q. Liu, Andrographolide antagonizes the cigarette smokeinduced epithelial-mesenchymal transition and pulmonary dysfunction through anti-inflammatory inhibiting HOTAIR, Toxicology 422 (2019) 84–94, https:// doi.org/10.1016/j.tox.2019.05.009.
- [69] S.-M. Wu, P.-H. Feng, H.-C. Chuang, S.-C. Ho, K. Fan Chung, K.-Y. Chen, G.-S. Wu, T.-T. Chen, C.-H. Tseng, W.-T. Liu, K.-Y. Lee, Impaired Inc-ILTR modulatory mechanism of Toll-like receptors is associated with an exacerbator phenotype of chronic obstructive pulmonary disease, Faseb. J. 34 (2020) 13317–13332, https://doi.org/10.1096/fj.202000632R.
- [70] M. Manevski, D. Devadoss, C. Long, S.P. Singh, M.W. Nasser, G.M. Borchert, M.N. Nair, I. Rahman, M. Sopori, H.S. Chand, Corrigendum: increased expression of LASI LncRNA regulates the cigarette smoke and COPD associated airway inflammation and mucous cell hyperplasia, Front. Immunol. 13 (2022), 988069, https://doi.org/10.3389/fimmu.2022.988069.
- [71] T. Vierbuchen, S. Agarwal, J.L. Johnson, L. Galia, X. Lei, K. Stein, D. Olagnier, K.I. Gaede, C. Herzmann, C.K. Holm, H. Heine, A. Pai, A. O'Hara Hall, K. Hoebe, K.A. Fitzgerald, The lncRNA LUCAT1 is elevated in inflammatory disease and restrains inflammation by regulating the splicing and stability of NR4A2, Proc. Natl. Acad. Sci. U. S. A. 120 (2023), e2213715120, https://doi.org/10.1073/pnas.2213715120.
- [72] S. Zhao, C. Lin, T. Yang, X. Qian, J. Lu, J. Cheng, Expression of long non-coding RNA LUCAT1 in patients with chronic obstructive pulmonary disease and its potential functions in regulating cigarette smoke extract-induced 16HBE cell proliferation and apoptosis, J. Clin. Lab. Anal. 35 (2021), e23823, https://doi. org/10.1002/jcla.23823.
- [73] B. Song, L. Ye, S. Wu, Z. Jing, Long non-coding RNA MEG3 regulates CSE-induced apoptosis and inflammation via regulating miR-218 in 16HBE cells, Biochem. Biophys. Res. Commun. 521 (2020) 368–374, https://doi.org/10.1016/j.bbrc.2019.10.135.
- [74] F. He, N. Wang, X. Yu, Y. Zheng, Q. Liu, Q. Chen, J. Pu, N. Li, W. Zou, B. Li, P. Ran, GATA3/long noncoding RNA MHC-R regulates the immune activity of dendritic cells in chronic obstructive pulmonary disease induced by air pollution particulate matter, J. Hazard Mater. 438 (2022), 129459, https://doi.org/ 10.1016/j.jhazmat.2022.129459.
- [75] W. Gu, Y. Yuan, L. Wang, H. Yang, S. Li, Z. Tang, Q. Li, Long non-coding RNA TUG1 promotes airway remodelling by suppressing the miR-145-5p/DUSP6 axis in cigarette smoke-induced COPD, J. Cell Mol. Med. 23 (2019) 7200–7209, https://doi.org/10.1111/jcmm.14389.
- [76] Q. Shen, J. Zheng, X. Wang, W. Hu, Y. Jiang, Y. Jiang, LncRNA SNHG5 regulates cell apoptosis and inflammation by miR-132/PTEN axis in COPD, Biomed. Pharmacother. 126 (2020), 110016, https://doi.org/10.1016/j.biopha.2020.110016.
- [77] Y. Wang, X. Lyu, X. Wu, L. Yu, K. Hu, Long non-coding RNA PVT1, a novel biomarker for chronic obstructive pulmonary disease progression surveillance and acute exacerbation prediction potentially through interaction with microRNA-146a, J. Clin. Lab. Anal. 34 (2020), e23346, https://doi.org/10.1002/ icla 23346
- [78] X. Ming, W. Duan, W. Yi, Long non-coding RNA NEAT1 predicts elevated chronic obstructive pulmonary disease (COPD) susceptibility and acute exacerbation risk, and correlates with higher disease severity, inflammation, and lower miR-193a in COPD patients, Int. J. Clin. Exp. Pathol. 12 (2019) 2837–2848.
- [79] R.C.J. Langen, S.H. Korn, E.F.M. Wouters, ROS in the local and systemic pathogenesis of COPD, Free Radic. Biol. Med. 35 (2003) 226–235, https://doi.org/ 10.1016/s0891-5849(03)00316-2.
- [80] G.F. Korytina, I.Z. Akhmadishina, Y.G. Aznabaeva, O.V. Kochetova, N.Sh Zagidullin, J.G. Kzhyshkowska, S.Z. Zagidullin, T.V. Viktorova, Associations of the NRF2/KEAP1 pathway and antioxidant defense gene polymorphisms with chronic obstructive pulmonary disease, Gene 692 (2019) 102–112, https://doi.org/ 10.1016/j.gene.2018.12.061.
- [81] A.B. Fisher, Redox signaling across cell membranes, Antioxidants Redox Signal. 11 (2009) 1349–1356, https://doi.org/10.1089/ars.2008.2378.
- [82] H. Sies, C. Berndt, D.P. Jones, Oxidative stress, Annu. Rev. Biochem. 86 (2017) 715–748, https://doi.org/10.1146/annurev-biochem-061516-045037.
 [83] H. Kanazawa, J. Yoshikawa, Elevated oxidative stress and reciprocal reduction of vascular endothelial growth factor levels with severity of COPD, Chest 128 (2005) 3191–3197, https://doi.org/10.1378/chest.128.5.3191.
- [84] D.S. Postma, W. Timens, Remodeling in asthma and chronic obstructive pulmonary disease, Proc. Am. Thorac. Soc. 3 (2006) 434–439, https://doi.org/ 10.1513/nats.200601-006AW.
- [85] R.M. Tuder, L. Zhen, C.Y. Cho, L. Taraseviciene-Stewart, Y. Kasahara, D. Salvemini, N.F. Voelkel, S.C. Flores, Oxidative stress and apoptosis interact and cause emphysema due to vascular endothelial growth factor receptor blockade, Am. J. Respir. Cell Mol. Biol. 29 (2003) 88–97, https://doi.org/10.1165/rcmb.2002-02280C.
- [86] W. Hao, F. Lin, H. Shi, Z. Guan, Y. Jiang, Long non-coding RNA OIP5-AS1 regulates smoke-related chronic obstructive pulmonary disease via targeting micro RNA -410-3p/IL-13, Bioengineered 12 (2021) 11664–11676, https://doi.org/10.1080/21655979.2021.2000199.
- [87] Y. Zeng, Y. Zhao, Y. Chen, S. Cai, P. Chen, PECAM EMPs regulate apoptosis in pulmonary microvascular endothelial cells in COPD by activating the Akt signaling pathway, Tob. Induc. Dis. 20 (2022) 40, https://doi.org/10.18332/tid/146959.
- [88] C. Wang, R.J. Youle, The role of mitochondria in apoptosis, Annu. Rev. Genet. 43 (2009) 95–118, https://doi.org/10.1146/annurev-genet-102108-134850.
 [89] I.A. Yang, C.R. Jenkins, S.S. Salvi, Chronic obstructive pulmonary disease in never-smokers: risk factors, pathogenesis, and implications for prevention and treatment, Lancet Respir. Med. 10 (2022) 497–511, https://doi.org/10.1016/S2213-2600(21)00506-3.
- [90] N. Berend, Contribution of air pollution to COPD and small airway dysfunction, Respirology 21 (2016) 237-244, https://doi.org/10.1111/resp.12644.
- [91] S. Liu, Y. Zhou, S. Liu, X. Chen, W. Zou, D. Zhao, X. Li, J. Pu, L. Huang, J. Chen, B. Li, S. Liu, P. Ran, Association between exposure to ambient particulate matter and chronic obstructive pulmonary disease: results from a cross-sectional study in China, Thorax 72 (2017) 788–795, https://doi.org/10.1136/ thoraxjnl-2016-208910.
- [92] F.W. Ko, K.P. Chan, D.S. Hui, J.R. Goddard, J.G. Shaw, D.W. Reid, I.A. Yang, Acute exacerbation of COPD, Respirology 21 (2016) 1152–1165, https://doi.org/ 10.1111/resp.12780.
- [93] B.R. Celli, L.M. Fabbri, S.D. Aaron, A. Agusti, R. Brook, G.J. Criner, F.M.E. Franssen, M. Humbert, J.R. Hurst, D. O'Donnell, L. Pantoni, A. Papi, R. Rodriguez-Roisin, S. Sethi, A. Torres, C.F. Vogelmeier, J.A. Wedzicha, An updated definition and severity classification of chronic obstructive pulmonary disease exacerbations: the Rome proposal, Am. J. Respir. Crit. Care Med. 204 (2021) 1251–1258, https://doi.org/10.1164/rccm.202108-1819PP.
- [94] P.J. Austin, E. Tsitsiou, C. Boardman, S.W. Jones, M.A. Lindsay, I.M. Adcock, K.F. Chung, M.M. Perry, Transcriptional profiling identifies the long noncoding RNA plasmacytoma variant translocation (PVT1) as a novel regulator of the asthmatic phenotype in human airway smooth muscle, J. Allergy Clin. Immunol. 139 (2017) 780–789, https://doi.org/10.1016/j.jaci.2016.06.014.
- [95] A. Agustí, E. Melén, D.L. DeMeo, R. Breyer-Kohansal, R. Faner, Pathogenesis of chronic obstructive pulmonary disease: understanding the contributions of gene-environment interactions across the lifespan, Lancet Respir. Med. 10 (2022) 512–524, https://doi.org/10.1016/S2213-2600(21)00555-5.

- [96] M.H. Cho, B.D. Hobbs, E.K. Silverman, Genetics of chronic obstructive pulmonary disease: understanding the pathobiology and heterogeneity of a complex disorder, Lancet Respir. Med. 10 (2022) 485–496, https://doi.org/10.1016/S2213-2600(21)00510-5.
- [97] E.K. Silverman, Genetics of COPD, Annu. Rev. Physiol. 82 (2020) 413–431, https://doi.org/10.1146/annurev-physiol-021317-121224.
- [98] C. Strange, Alpha-1 antitrypsin deficiency associated COPD, Clin. Chest Med. 41 (2020) 339–345, https://doi.org/10.1016/j.ccm.2020.05.003.
- [99] T. Köhnlein, T. Welte, Alpha-1 antitrypsin deficiency: pathogenesis, clinical presentation, diagnosis, and treatment, Am. J. Med. 121 (2008) 3–9, https://doi. org/10.1016/j.amjmed.2007.07.025.
- [100] S.G.J. Smith, C.M. Greene, Long non-coding RNA expression in alpha-1 antitrypsin deficient monocytes pre- and post-AAT augmentation therapy, Noncoding RNA 9 (2023) 6, https://doi.org/10.3390/ncrna9010006.
- [101] Z. He, H. Peng, M. Gao, G. Liang, M. Zeng, X. Zhang, p300/Sp1-Mediated high expression of p16 promotes endothelial progenitor cell senescence leading to the occurrence of chronic obstructive pulmonary disease, Mediat. Inflamm. 2021 (2021), 5599364, https://doi.org/10.1155/2021/5599364.
- [102] S. Chattopadhyay, L. Malayil, S. Kaukab, Z. Merenstein, A.R. Sapkota, The predisposition of smokers to COVID-19 infection: a mini-review of global perspectives, Heliyon 9 (2023), https://doi.org/10.1016/j.heliyon.2023.e17783.