

REVIEW

The SARS-CoV-2 main protease (M^{Pro}): Structure, function, and emerging therapies for COVID-19

Qing Hu^{1,2,#} | Yuan Xiong^{1,#} | Guang-Hao Zhu¹ | Ya-Ni Zhang¹ | Yi-Wen Zhang² | Ping Huang^{2,*} | Guang-Bo Ge^{1,*}

¹Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai, China

²Clinical Pharmacy Center, Cancer Center, Department of Pharmacy, Zhejiang Provincial People's Hospital, Affiliated People's Hospital, Hangzhou Medical College, Hangzhou, Zhejiang, China

*Correspondence

Guang-Bo Ge, Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China.

Email: geguangbo@dicp.ac.cn

Ping Huang, Clinical Pharmacy Center, Cancer Center, Department of Pharmacy, Zhejiang Provincial People's Hospital, Affiliated People's Hospital, Hangzhou Medical College, Hangzhou, Zhejiang 310014, China.

Email: huangping@hmc.edu.cn

[#]Qing Hu and Yuan Xiong contributed equally.

Funding information

Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine, Grant/Award Number: ZYYCXTD-D-202004; Three-year Action Plan for Shanghai TCM Development and Inheritance Program, Grant/Award Number: ZY(2021-2023)-0401; The Basic Public Welfare Research Program of Zhejiang Province, Grant/Award Number: LGF22H280012; Shanghai Science and Technology Innovation Action Plans, Grant/Award Numbers: 20S21901500,

Abstract

The main proteases (M^{Pro}), also termed 3-chymotrypsin-like proteases (3CL^{Pro}), are a class of highly conserved cysteine hydrolases in β -coronaviruses. Increasing evidence has demonstrated that 3CL^{Pro}s play an indispensable role in viral replication and have been recognized as key targets for preventing and treating coronavirus-caused infectious diseases, including COVID-19. This review is focused on the structural features and biological function of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) main protease M^{Pro} (also known as 3CL^{Pro}), as well as recent advances in discovering and developing SARS-CoV-2 3CL^{Pro} inhibitors. To better understand the characteristics of SARS-CoV-2 3CL^{Pro} inhibitors, the inhibition activities, inhibitory mechanisms, and key structural features of various 3CL^{Pro} inhibitors (including marketed drugs, peptidomimetic, and non-peptidomimetic synthetic compounds, as well as natural compounds and their derivatives) are summarized comprehensively. Meanwhile, the challenges in this field are highlighted, while future directions for designing and developing efficacious 3CL^{Pro} inhibitors as novel anti-coronavirus therapies are also proposed. Collectively, all information and knowledge presented here are very helpful for understanding the structural features and inhibitory mechanisms of SARS-CoV-2 3CL^{Pro} inhibitors, which offers new insights or inspiration to medicinal chemists for designing and developing more efficacious 3CL^{Pro} inhibitors as novel anti-coronavirus agents.

KEYWORDS

3-chymotrypsin-like protease (3CL^{Pro}), broad-spectrum anti-coronavirus agents, SARS-CoV-2, β -coronavirus 3CL^{Pro} inhibitor

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. *MedComm* published by Sichuan International Medical Exchange & Promotion Association (SCIMEA) and John Wiley & Sons Australia, Ltd.

20S21900900; Shanghai Science and Technology Committee, Zhejiang Provincial Medical and Health Science and Technology Program, Grant/Award Number: 2022495401; Hangzhou Medical College Basic Research Program, Grant/Award Number: KYQN202124; Chinese Medicine Research Program of Zhejiang Province, Grant/Award Numbers: 2020ZZ003, 2021ZZ001; “10000 Talents Plan” of Zhejiang Province, Grant/Award Number: 2020R52029; National Natural Science Foundation of China, Grant/Award Numbers: 82141203, 81922070, 81973286; Zhejiang Provincial Program for the Cultivation of New Health Talents to Yiwen Zhang

1 | INTRODUCTION

Coronaviruses (CoVs) are single-stranded positive-sense ribonucleic acid (RNA) enveloped viruses with a 5'-cap and 3'-poly-A tail that can be classified into four subgroups: α , β , γ , and δ . The hosts of CoVs are vertebrates that range from human beings to birds, generally causing respiratory and gastrointestinal tract disorders.¹⁻⁶ Seven human coronaviruses have emerged, including three fatal β -CoVs (severe acute respiratory syndrome coronavirus [SARS-CoV], Middle-East respiratory syndrome coronavirus [MERS-CoV], and severe acute respiratory syndrome coronavirus 2 [SARS-CoV-2]).⁷⁻¹⁰ Among them, SARS-CoV-2 and SARS-CoV belong to the subgenus *Sarbecovirus* of β -CoVs according to the latest release of the International Committee on Taxonomy of Viruses (<https://talk.ictvonline.org/>). Particularly, SARS-CoV-2, the pathogen for coronavirus disease 2019 (COVID-19), has taken millions of lives, generating a huge negative impact on the public.¹¹⁻¹⁴ To combat this epidemic effectively, scientists have made great efforts in drug repurposing, vaccine development, and novel medication discovery. To date, several effective vaccines that mainly target the viral spike (S) protein can be used for the preliminary prevention of COVID-19 by eliciting an immune response.^{15,16} Newly emerging variants (e.g., Delta and Omicron) of SARS-CoV-2 have generated high-frequency mutations in the S protein, including nucleic acid mutations and amino acid mutations, which present potential hazards for the effectiveness of vaccines and mutation-mediated resistance.¹⁷⁻²⁰

In the process of virus multiplication, the main proteases (M^{pro} , also known as $3CL^{pro}$), a class of highly conserved cysteine hydrolases from CoVs, are capable of cleaving polyproteins at multiple sites to yield multiple functional

proteins.²¹ Considering that $3CL^{pro}$ s play a vital role in CoV replication, especially in the two of the most serious pandemics of the 21st century caused by SARS-CoV-2 and SARS-CoV, these key hydrolases have been validated as promising targets for developing broad-spectrum anti-CoV agents.²²⁻²⁶ Because no homolog of $3CL^{pro}$ has been identified in humans, it is feasible to develop efficacious and specific $3CL^{pro}$ inhibitors with extremely weak inhibitory effects on human proteases, thereby reducing the side effects caused by $3CL^{pro}$ inhibitors. As shown in Figure 1, the phylogenetic relationships of 14 kinds of $3CL^{pro}$ s from coronaviruses show that the relatedness of the $3CL^{pro}$ s for SARS-CoV-2 and SARS-CoV are extremely close²⁷⁻³⁷; thus, most attempts to develop new SARS-CoV-2 $3CL^{pro}$ inhibitors are based on previously reported SARS-CoV $3CL^{pro}$ inhibitor. As an attractive target for combating viral replication and pathogenesis to control various CoVs, $3CL^{pro}$ has drawn much interest from both academics and industry.

In recent years, multiple drug discovery strategies have been utilized to find or develop a number of $3CL^{pro}$ inhibitors against SARS-CoV-2, such as drug repurposing, virtual screening coupled with high-throughput screening (HTS), and structure-based drug design.³⁸⁻⁴⁰ Moreover, the discovery of active compounds from natural products remains one of the most important sources for developing novel anti-CoV agents.⁴¹⁻⁴⁷ Therefore, many research groups have devoted their efforts to finding anti-CoV agents in naturally occurring compounds.⁴⁸⁻⁵² To date, a variety of marketed drugs and other structurally diverse synthetic compounds, as well as a number of natural compounds, have been found to be efficacious inhibitors of SARS-CoV-2 $3CL^{pro}$, showing great potential for developing novel broad-spectrum anti-CoV agents.⁵³⁻⁵⁹ Thus, this review focuses on the structural features and

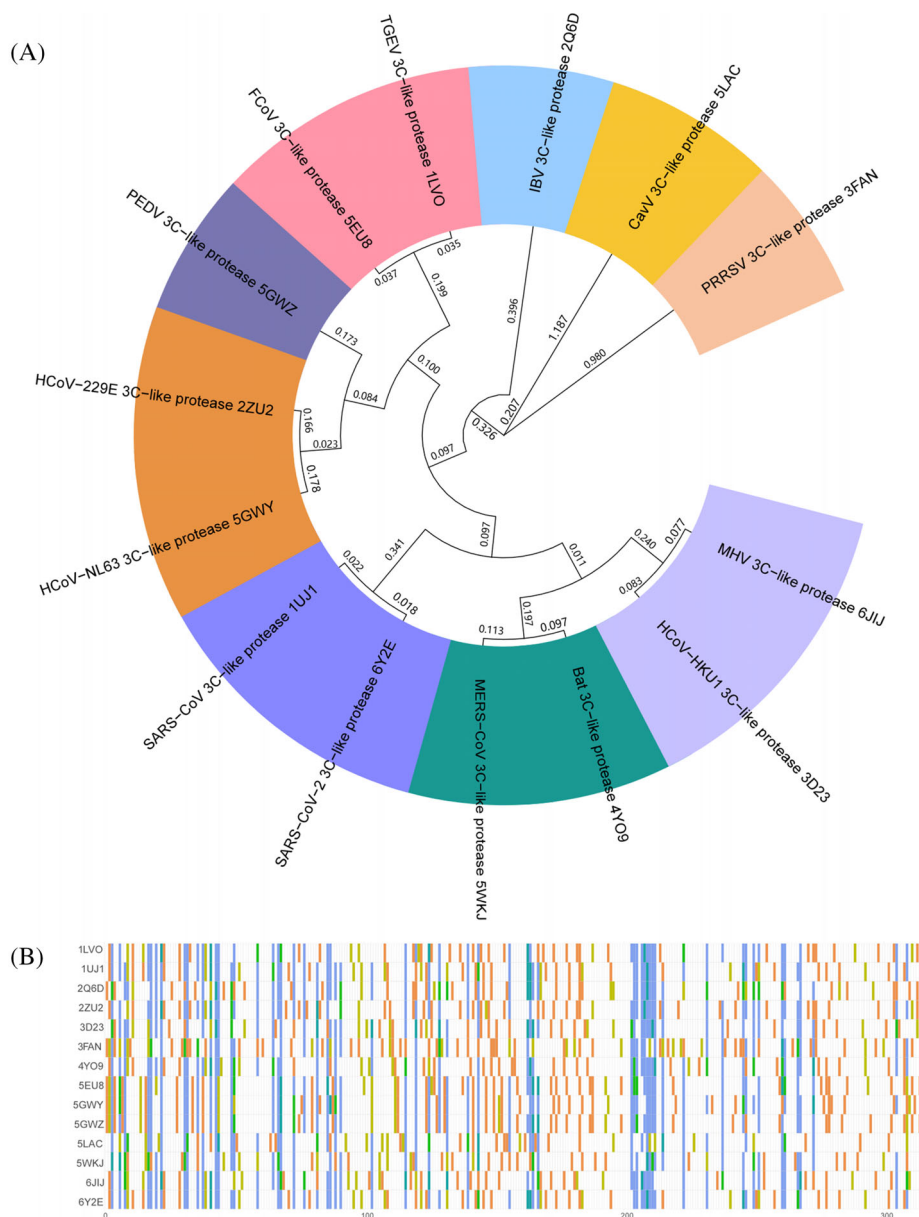


FIGURE 1 Phylogenetic relationships for 14 reported 3-chymotrypsin-like proteases (3CL^{PRO}s) in *Nidovirus*. (A) The evolutionary distances (genetic variations) of 3CL^{PRO}s are presented on branches. (B) Amino acid homologous sequence alignment of 3CL^{PRO}s

function of 3CL^{PRO} and recent advances in the discovery of SARS-CoV-2 3CL^{PRO} inhibitors, aiming to provide a SARS-CoV-2 3CL^{PRO} inhibitor library for medicinal chemists to design and develop more efficacious anti-CoV agents in the future.

2 | STRUCTURAL FEATURES AND FUNCTION OF 3CL^{PRO}

A total of 432 structures of SARS-CoV-2 3CL^{PRO} are currently uploaded to the PDB database, containing 54 apoprotein structures and 378 liganded protein structures (Supporting Information). The available structures

of SARS-CoV-2 3CL^{PRO} were crystallized at temperatures ranging from 277 to 300 K and refined at resolutions ranging from 1.2 to 2.98 Å. 3CL^{PRO} is approximately 34.21 kDa per monomer (average molecular weight of monomeric deposited models). 3CL^{PRO} is matured in a dimeric form, and the individual monomers are enzymatically less active, where the monomers consist of three domains, including domain I, domain II, and domain III.^{60,61} Among them, domain III is an extra helix domain, whose aggregation initiates the dimerization of 3CL^{PRO}.^{16,22,36,62} Generally, the monomer of 3CL^{PRO} is a transient state that proved to be enzymatically less active, while the dimeric form acts as a functional unit with the highest hydrolytic activity (Table 1).^{63–65} The firm binding between the N-finger and

TABLE 1 Molecular features of 3-chymotrypsin-like proteases (3CL^{PRO}) from severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and severe acute respiratory syndrome coronavirus (SARS-CoV)

Property	SARS-CoV-2 3CL ^{PRO}	SARS-CoV 3CL ^{PRO}
Molecular weight (kDa)	34	34
Isoelectric point	6.0	6.2
Optimal pH	7.5	7.0
Length of monomer (residue)	306	306
Mature form	Homodimer	Homodimer
Catalytic residues	His41, Cys145	His41, Cys145

C-terminus is one of the key conditions for the formation of dimeric 3CL^{PRO}, especially the salt bridge between Arg4 and Arg298.^{66–72}

The catalytic site of 3CL^{PRO} is located at the intersection of domains I and II, which can be divided into mainly five sub-pockets, including S1, S2, S3, S4, and S5 (Figure 2).^{36,73} The key facial residues of five sub-pockets are listed in Table S1, whose dimensional chemical environment matches five specific substrate-binding positions.^{64,74,75} P1, P2, and P1' positions mainly determine the substrate specificity of 3CL^{PRO}, while P4, P3, and P3' boost the recognition and stable binding of substrates.^{64,76} The O^β atom of glutamine could bind to the oxyanion hole (residues 143–145) of S1, and then the thiol of Cys145 could attack the C atom of glutamine as a nucleophile.^{64,77} Therefore, P1 almost always requires glutamine or lactam warhead.^{78–80} Notably, only one of the catalytic sites possesses hydrolytic function in the homodimer.^{16,78,81}

Different from the catalytic triad of 3-chymotrypsin, the catalytic dyad of 3CL^{PRO} is formed by Cys145 and His41.^{82,83} The zwitter catalytic dyad Cys⁻145–His⁺41 needs to be activated by energetical water that is maintained by His164 and Asp187.^{63,84–87} Cleavage of the large polyprotein chains by 3CL^{PRO} occurs at the glutamine residue in the P1 position of the substrate via a Cys145–His41 dyad, in which the cysteine thiol functions as the nucleophile in the proteolytic process. The cleavage of polyproteins by 3CL^{PRO} using a universal nucleophilic-type reaction mechanism is as follows (Figure 2E). Initially, the Cys145-thiol on the catalytic dyad is deprotonated with the help of nearby His41, where the anionic sulfur attacks the C-terminal C atom of the specially recognized Gln as a nucleophile.⁸⁸ Then, after spitting the amide bond, the histidine restores the deprotonated form, and the generated thioester is attacked in an identical fashion, with water acting as the nucleophile leading to the release of the hydrolyzed C-terminal, thus resetting the catalytic dyad.^{89,90} The mutant experiment proved that the catalytic cysteine is essential to 3CL^{PRO}, as replacing cysteine with serine would result in a covalent product–enzyme complex or a covalent

Ser145O^γ–Gln306C bond, fatally blocking the self-cleavage process.⁶⁴

As a key cysteine protease, SARS-CoV-2 3CL^{PRO} has 12 cysteine residues, but only three of them (Cys85, Cys145, and Cys156) are exposed to solvent.⁹¹ Catalytic Cys145 is the most important cysteine located in the catalytic site. Myricetin can bind to Cys145 in its oxidized form.⁹² Ebse-len and its derivatives can modify Cys145 by forming a Se–S bond.⁹³ Peptidomimetic α -acyloxymethylketone warheads can react with Cys145 through a structure-based selectivity mechanism.⁹⁴ However, few covalent inhibitors have been discovered to be accessible to Cys85 and Cys156. Cys156 is only profiled using *N*-ethylmaleimide, a small-molecule electrophile that engages cysteine side-chain thiolates by creating a covalent bond.⁶³ The inconducive spatial environments enclosing Cys85 and Cys156 are speculated to be the cause. Although the bulk of the 3CL^{PRO} cysteines is buried inside the protein, several cysteines are tested and predicted to be reactive. Cys22 and Cys44 are two conserved deprotonated cysteines in 3CL^{PRO}. Constant-pH molecular dynamics (CpHMD) titration revealed that Cys22 and Cys44 are more nucleophilic than catalytic Cys145.⁹⁵ Cys44 is largely inclined to be modified by flavonoids because the pocket encircled Cys44 is compatible with flavonoids, such as baicalein and covalent-binding myricetin.^{95,96} Cys300 is proven to be an allosteric site of 3CL^{PRO}.⁹⁵ Myricetin and colloidal bismuth subcitrate (CBS) can bind to Cys300 of 3CL^{PRO} and inhibit 3CL^{PRO} as allosteric inhibitors.^{96,97} Notably, a prominent dissociation of 3CL^{PRO} occurs after incubation of CBS with 3CL^{PRO}, resulting in degradation of 3CL^{PRO} and collapse of the active site.⁹⁷

3 | SYNTHETIC COMPOUNDS

With the help of the high-resolution crystal structures of both SARS-CoV-2 3CL^{PRO} and its homolog SARS-CoV 3CL^{PRO}, a panel of computer-aided drug design and crystallography-guided fragment-based drug

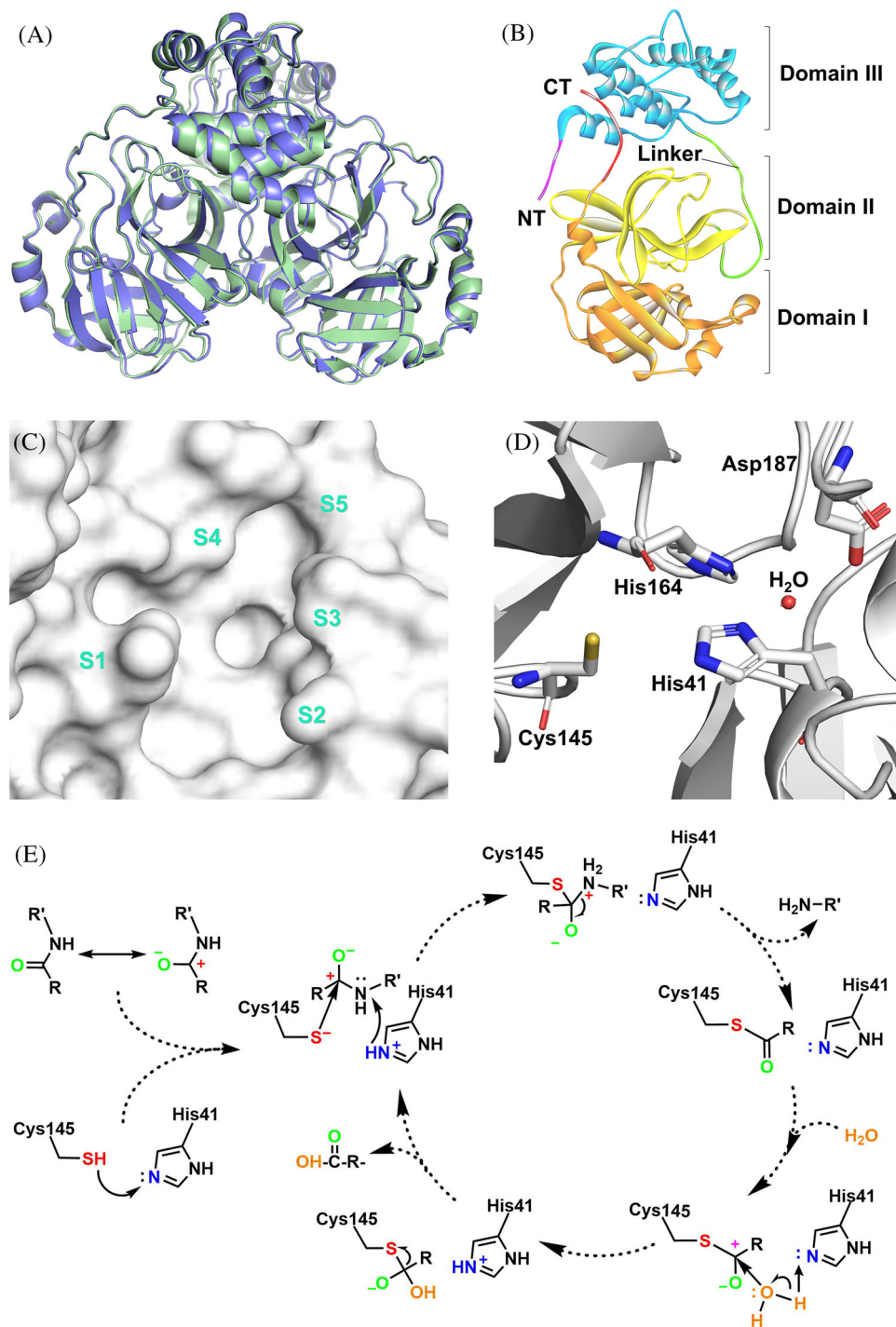


FIGURE 2 (A) The 3D structure of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) 3CL^{pro} (pale green, PDB: 6XHU) and severe acute respiratory syndrome coronavirus (SARS-CoV) 3CL^{pro} (slate, PDB: 1UJ1). (B) Three structural domains (domain I: orange, domain II: yellow, domain III: blue) of SARS-CoV-2 3CL^{pro} monomer. (C) The surface representation for the catalytic pocket (sub-pockets: S1–S5) of SARS-CoV-2 3CL^{pro}. (D) The amino acid residues in the active site of SARS-CoV-2 3CL^{pro}. (E) The catalytic mechanism of 3CL^{pro} on the hydrolysis of amide substrate

discovery approach have been widely used to screen and design novel inhibitors against SARS-CoV 3CL^{pro}.⁹⁸ A majority of synthetic SARS-CoV-2 3CL^{pro} inhibitors are designed based on the 3D structure of the active pocket and substrate preferences of the target enzyme, which could

be structurally categorized into peptidomimetics and non-peptidomimetics (small molecules).^{40,99} Currently, a number of synthetic compounds (including peptidomimetics and non-peptidomimetics) have been found with strong to extremely potent SARS-CoV-2 3CL^{pro} inhibitors, which

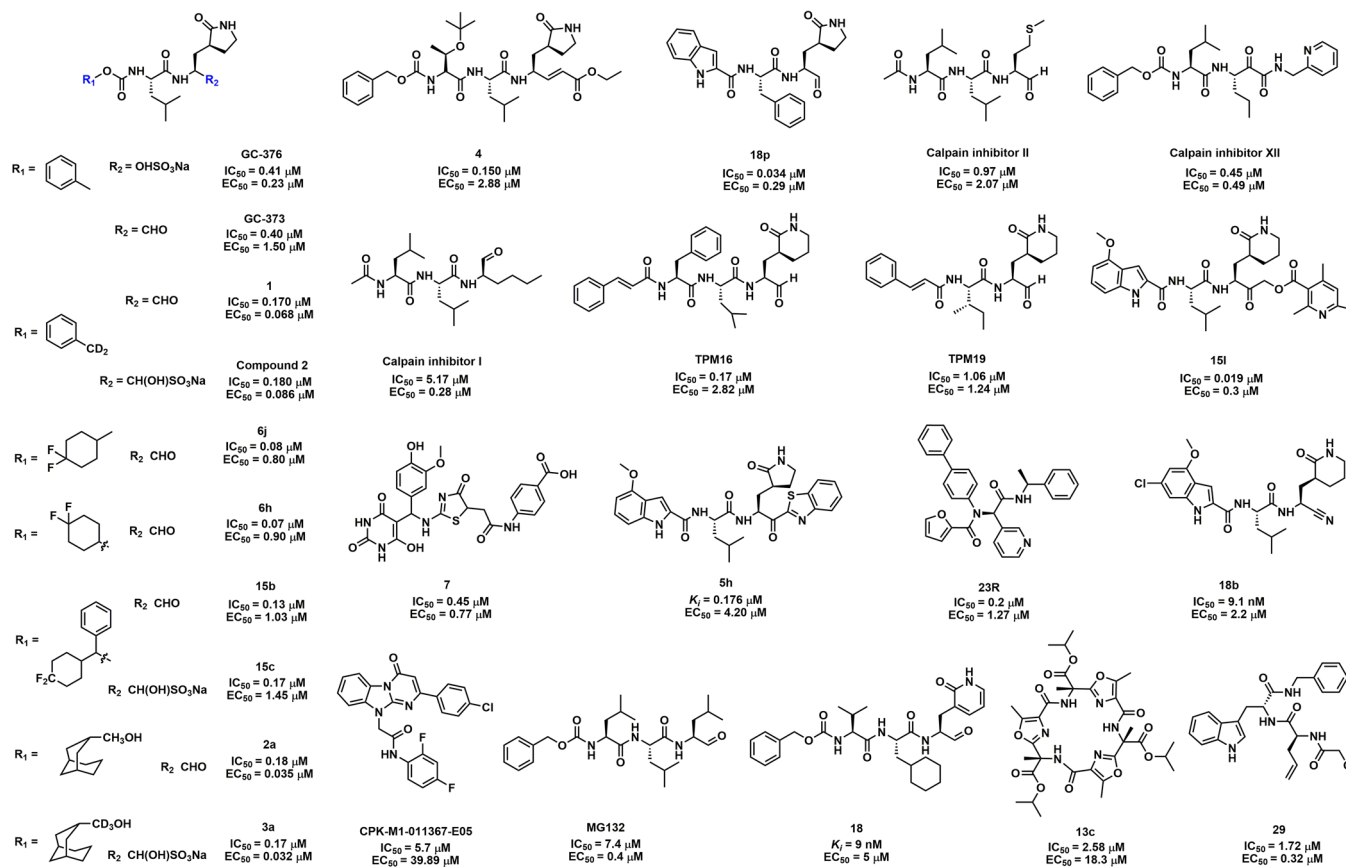


FIGURE 3 The chemical structures and half-maximal inhibitory concentration (IC₅₀) values for representative peptidomimetic SARS-CoV-2 3CL^{pro} inhibitors, as well as their half-maximal effect concentration (EC₅₀) values for anti-SARS-CoV-2

have aroused significant interest in the pharmaceutical industry to develop more efficacious antiviral drug candidates with satisfying drug-likeness properties and safety profiles for combating COVID-19.¹⁰⁰

3.1 | Peptidomimetic SARS-CoV-2 3CL^{pro} inhibitors

Peptidomimetics have been widely used for the development of antiviral drugs, offering numerous properties, such as superior efficiency and safety, as well as less accumulation within the body.¹⁰¹ Thus, a series of peptidomimetic antiviral drugs have been rationally designed for the treatment of COVID-19. As shown in Figure 3, GC376, a peptidomimetic antiviral protease inhibitor for the treatment of cats infected with feline infectious peritonitis virus, showed strong inhibition of SARS-CoV-2 3CL^{pro} and SARS-CoV-2 replication with the half-maximal inhibitory concentration (IC₅₀) of 26.4 nM and 0.91 μM, respectively.^{102–104} Vuong et al.¹⁰⁵ also reported that this drug and its parent GC373 were potent inhibitors of 3CL^{pro} of SARS-CoV and SARS-CoV-2 with IC₅₀ values in the nanomolar range. Nuclear magnetic resonance

(NMR) analysis showed that these inhibitors covalently modified Cys145 to reversibly form a hemithioacetal. Meanwhile, GC376 and compound 4 were found to be covalent inhibitors of SARS-CoV-2 3CL^{pro}.¹⁰⁶ Furthermore, Dampalla et al.¹⁰⁷ synthesized a series of deuterated derivatives of GC376 and determined the therapeutic efficacy in a lethal mouse model. In the co-crystal structure of SARS-CoV 3CL^{pro}, a novel stereocenter formed by compound 2 covalently attached to Cys145 with nearly the same hydrogen bonding interactions as SARS-CoV-2 3CL^{pro}. As a result of the multiple advantages of introducing deuterium into the drug, the deuterated variants at the R-site exhibited a significant increase in the anti-3CL^{pro} and cell-based assays, as well as improved pharmacokinetics and reduced toxicity.^{107,108} Moreover, by using the fluorine-walk approach to explore the binding modes of the F-substituted phenyl ring, they found that compounds 15b and 15c were the most effective against SARS-CoV-2 3CL^{pro}, with IC₅₀ values of 0.13 and 0.17 μM, respectively.¹⁰⁹ Compounds 6j and 6h inhibited SARS-CoV-2 3CL^{pro} with significant efficacy, while administration of compound 6j significantly improved survival, reductions in lung virus titers, and lung histopathology throughout the day in a mouse model of MERS-CoV

infection.⁸⁰ Several non-deuterated and deuterated compounds containing a conformationally constrained cyclohexane moiety were synthesized, of which compound **2a/3a** displayed high potency in biochemical assays with IC_{50} values in the submicromolar range. Importantly, the half-maximal effect concentration (EC_{50}) values of compounds **2a** and **3a** against SARS-CoV-2 in Vero E6 cells were 0.035 and 0.032 μM , respectively.¹¹⁰ All of the above compounds contain GC376 variants with potent biological activity, making them potential anti-COVID-19 candidates.

A series of novel protease inhibitors with an aldehyde warhead targeting the 3C protease of enterovirus 71 was designed and synthesized, especially compound **18p**, which showed potent enzyme inhibitory activity and broad-spectrum antiviral activity against a group of enteroviruses and rhinoviruses.¹¹¹ Notably, compound **18p** showed strong replication inhibition against SARS-CoV-2 3CL^{pro} ($IC_{50} = 0.034 \mu\text{M}$, $EC_{50} = 0.29 \mu\text{M}$), making it a further potential candidate for the treatment of COVID-19. In particular, calpain inhibitors I, II, and XII were identified as SARS-CoV-2 3CL^{pro} inhibitors in vitro and in vivo.^{112,113} Moreover, five tetrapeptidomimetic anti-3CL^{pro} inhibitors, similar to the backbone of **13a**, were successfully involved in the design of the catalytic dyad histidine residue (His41) of 3CL^{pro}. Among them, TPM16 and TPM19 exhibited nanomolar inhibition and attenuated the cellular viral loads of SARS-CoV-2.¹¹⁴ Compound **15l** with novel α -acyloxymethylketone warhead mimetic compounds was described by Bai et al.,⁹⁴ which was identified to have potent SARS-CoV-2 3CL^{pro} and viral replication inhibition in vitro. Moreover, co-crystallization of **15l** with SARS-CoV-2 3CL^{pro} confirmed the formation of covalent adducts. Compound **7** showed inhibition activity against 3CL^{pro}, papain-like protease (PL^{pro}), and furin protease at IC_{50} values of 0.45, 0.085, and 0.29 μM , respectively. Moreover, compound **7** has a higher inhibitory effect on the virus and is nontoxic to mammalian cells, making it a powerful dual inhibitory activity against SARS-CoV-2.¹¹⁵ Using a covalent DNA-encoded library screening platform, Ge et al.¹¹⁶ found that compound **1e** showed potent inhibition of SARS-CoV-2 3CL^{pro} (Table S2).

A small-molecule compound **5h** containing an indole moiety was characterized against SARS-CoV-2 3CL^{pro} (inhibition constant, $K_i = 17.6 \text{ nM}$) via reversible covalent interactions. Based on Vero E6 cell assays, **5h** blocked the infectivity of SARS-CoV-2 with an EC_{50} value of 4.2 μM .^{117,118} One novel SARS-CoV-2 3CL^{pro} inhibitor, compound **23R**, was highly selective compared to covalent inhibitors. The co-crystal structure of SARS-CoV-2 3CL^{pro} with **23R** reveals a previously unexplored binding site located between the S2 and S4 pockets.¹¹⁹ Bai et al.¹²⁰ described that compound **18b** bearing nitrile warheads displays good SARS-CoV-2 3CL^{pro} inhibition activity, which

could reduce SARS-CoV-2 plaques in Vero E6 host cells ($EC_{50} = 2.2 \mu\text{M}$), and showed a better selectivity than the aldehyde warhead peptidomimetics for human cysteine proteases (cathepsins B, S, and L). Seven peptidomimetic SARS-CoV-2 3CL^{pro} inhibitors were identified from the Korea Chemical Bank library. Among these agents, CPK-M1-011367-E05 showed strong anti-3CL^{pro} activity and anti-SARS-CoV-2 activity.¹²¹

For CoVs to successfully invade the host cell, the S protein of CoVs needs to be cleaved and activated by some host cell proteases, such as furin and transmembrane protease serine 2 (TMPRSS2). Cathepsin L is a lysosomal cysteine protease in the host that is closely related to the membrane fusion of SARS-CoV.¹²² A clinical study suggested that cathepsin L level in COVID-19 patients was positively correlated with disease course and severity.¹²³ Specifically, the secondary cleavage of the S protein by cathepsin L promotes the cell-cell fusion of SARS-CoV-2, indicating that cathepsin L is a promising target for anti-COVID-19.¹²⁴ Recently, a proteasome inhibitor, MG132, was identified as a dual inhibitor for SARS-CoV-2 3CL^{pro} and cathepsin L, which could covalently and reversibly bind to Cys145 of 3CL^{pro}.¹²⁵ A novel class of self-masked aldehyde inhibitors for cruzain was developed, in which compound **18** showed extremely potent 3CL^{pro} inhibitory activity ($K_i = 9 \text{ nM}$) and good anti-SARS-CoV-2 activity ($EC_{50} = 5 \mu\text{M}$) in A549/angiotensin-converting enzyme 2 (ACE2) cells.¹²⁶ Macrocyclic peptides are known for their higher membrane permeability, superior selectivity, and stability, making them a promising privileged structure in drug discovery. Macrocyclic peptide **13c** has been found to have significant inhibitory activity against SARS-CoV-2 3CL^{pro} ($IC_{50} = 2.58 \mu\text{M}$).¹²⁷ Moreover, Johansen-Leete et al.¹²⁸ reported several high-affinity thioether-linked cyclic peptide inhibitors of SARS-CoV-2 3CL^{pro}, and several inhibitors exhibited in vitro anti-SARS-CoV-2 activity with EC_{50} values in the low micromolar range. Compound **29** was identified as a dual-action inhibitor of SARS-CoV-2 proteases that inhibits 3CL^{pro} at a micromolar level ($IC_{50} = 1.72 \mu\text{M}$) while inhibiting PL^{pro} at a submicromolar level ($IC_{50} = 0.67 \mu\text{M}$).¹²⁹

As shown in Table S2, a set of submicromolar covalent inhibitors with warheads were screened by Stille et al.,¹³⁰ and compounds **16a** and **14a** significantly inhibited the catalytic activity of SARS-CoV-2 3CL^{pro}. Breidenbach et al.¹³¹ identified and optimized two classes of protease inhibitors (azanitrile and pyridyl esters), of which azanitrile **8** ($K_i = 24 \text{ nM}$), equipped with a unique azanitrile warhead, was an irreversible inhibitor of SARS-CoV-2 3CL^{pro}. SDZ224015, a promising clinical caspase-1 inhibitor, was identified as a SARS-CoV-2 3CL^{pro} inhibitor ($IC_{50} = 30 \text{ nM}$) and might form an irreversible covalent adduct with the target enzyme.¹³² MPI3 and MPI8

displayed high potency against SARS-CoV-2 3CL^{pro}, with MPI8 showing the best selectivity toward host cathepsin L, reducing the potential toxicity toward host cells and high antiviral potency.^{133,134} Using chlorofluoroacetamide as a reactive warhead, Yamane et al.¹³⁵ have developed an irreversible inhibitor of SARS-CoV-2 3CL^{pro}. Among them, the inhibitory activity of (*R,R*)-18 against 3CL^{pro} was significantly higher than that of the other isomers.

3.2 | Non-peptidomimetic SARS-CoV-2 3CL^{pro} inhibitors

To meet the urgent requirements for anti-SARS-CoV-2 agents, scientists have made great efforts to discover anti-SARS-CoV-2 agents from in-house compound libraries or commercially available compounds via virtual screening coupled with experimental validation in early studies. The activities of non-peptidomimetic SARS-CoV-2 3CL^{pro} inhibitors are summarized in Figure 4 and Table S2. For instance, Yang et al.¹³⁶ adopted a multiple conformational-based virtual screening strategy and surface plasmon resonance assay for SARS-CoV-2 3CL^{pro} inhibitors from a protein mimetics library. Six compounds presented inhibitory effects against 3CL^{pro} both in vitro and in HEK293T cells, and Z1759961356 hindered viral replication in Vero E6 cells with an EC₅₀ value of 8.52 μ M. Four isoquinolone-based compounds from a database were reported as SARS-CoV-2 3CL^{pro} inhibitors with IC₅₀ values of approximately 1 μ M.¹³⁷

In fact, some inhibitors have a potential impact on several 3CL^{pro}s due to the high conservation of this protease among CoVs.¹³⁸ For example, ML188 inhibited the 3CL^{pro} of SARS-CoV, SARS-CoV-2, and porcine epidemic diarrhea virus, making it a promising broad-spectrum antiviral agent.⁷⁴ Moreover, structural optimization based on the reported inhibitor was a feasible strategy to develop SARS-CoV-2 3CL^{pro} inhibitors. CCF0058981, a novel compound derived from ML300 (SARS-CoV 3CL^{pro} inhibitor),¹³⁹ exerted a nanomolar level of activity (IC₅₀ = 68 nM) against SARS-CoV-2 3CL^{pro}, as well as superior anti-SARS-CoV-2 activities in both cytopathic effect inhibition assays (EC₅₀ = 0.497 μ M) and plaque reduction assays (EC₅₀ = 0.558 μ M).¹⁴⁰ Except for viral proteases, host proteases related to viral infection are also promising targets for fighting COVID-19. In this context, Elseginy et al.¹¹⁵ focused on multi-target inhibitors for fighting SARS-CoV-2. They demonstrated that not only compound **13d** inhibited 3CL^{pro} and PL^{pro} in SARS-CoV-2 but also furin protease in the host. Meanwhile, this compound could significantly inhibit SARS-CoV-2 (IC₅₀ = 0.11 μ M) in vitro.

Perampanel, an antiepileptic drug, showed weak inhibitory activity against SARS-CoV-2 3CL^{pro}, while its cloverleaf motif could occupy three sub-pockets with a high docking score, thus proposing it as a promising skeleton for novel 3CL^{pro} inhibitors.¹⁴¹ To validate this hypothesis, Zhang et al.¹⁴² put forward a useful strategy to guide the rational design of perampanel-derived inhibitors, which was a combination of several methodologies, including the free-energy perturbation calculation, structural analysis, biochemistry assessments, and X-ray crystallography. Among the 27 analogs, compound **21b** exhibited a potent inhibitory effect against SARS-CoV-2 3CL^{pro} (IC₅₀ = 18 nM) but with the greatest cytotoxicity. In particular, the combined use of compound **5** and remdesivir was initially predicted to have a synergistic effect on antiviral activity. To elevate the inhibitory activity and safety, they conducted crystallographic studies for further refinements.¹⁴³ In a follow-up study, this team found that 13 uracyl-containing compounds presented strong activity. Among 13 newly designed compounds, compound **19a** had potent enzyme inhibitory activities, good anti-SARS-CoV-2 effects, good aqueous solubility, and low toxicity, suggesting that it is a promising compound for anti-COVID-19.¹⁴⁴ According to the effective pharmacophores, some inhibitors were designed and synthesized to optimize the valuable interactions that could perfectly fit the enzymatic active pockets. Lutens et al.¹⁴⁵ carried out several screening cycles and combinations of promising scaffolds for SARS-CoV-2 3CL^{pro} inhibitors. Compound **19b** was optimized as a noncovalent broad-spectrum 3CL^{pro} inhibitor and showed promising antiviral activity, as well as good metabolic stability and plasma protein binding in humans.

In addition, some synthetic compounds bear at least reactive groups (such as Michael receptors and α,β -unsaturated carbonyl) that can covalently bind to crucial residuals (e.g., Cys145) of SARS-CoV-2 3CL^{pro}, giving rise to irreversible inactivation of the target enzyme.¹⁴⁶ Such inhibitors generated long-lasting and efficient inhibitory effects against SARS-CoV-2 3CL^{pro}, implying a promising strategy for the development of anti-COVID-19 agents.^{16,147,148} Jin et al.¹⁶ screened six inhibitors against SARS-CoV-2 3CL^{pro} from a library of 3CL^{pro} containing over 10,000 compounds, and ebselen and PX-12 could covalently bind to Cys145 of 3CL^{pro}, and ebselen might also noncovalently bind to 3CL^{pro} simultaneously. Beyond that, ebselen could react with cysteine residues of several viral proteases, such as SARS-CoV-2 PL^{pro} and 3C^{pro} of enterovirus A71 and enterovirus D68, which was suggested as a multi-target antiviral agent.¹⁴⁹ Some ebselen and ebsulfur derivatives were synthesized as SARS-CoV-2 3CL^{pro} inhibitors, and **1i** and **2k** were proved as potent

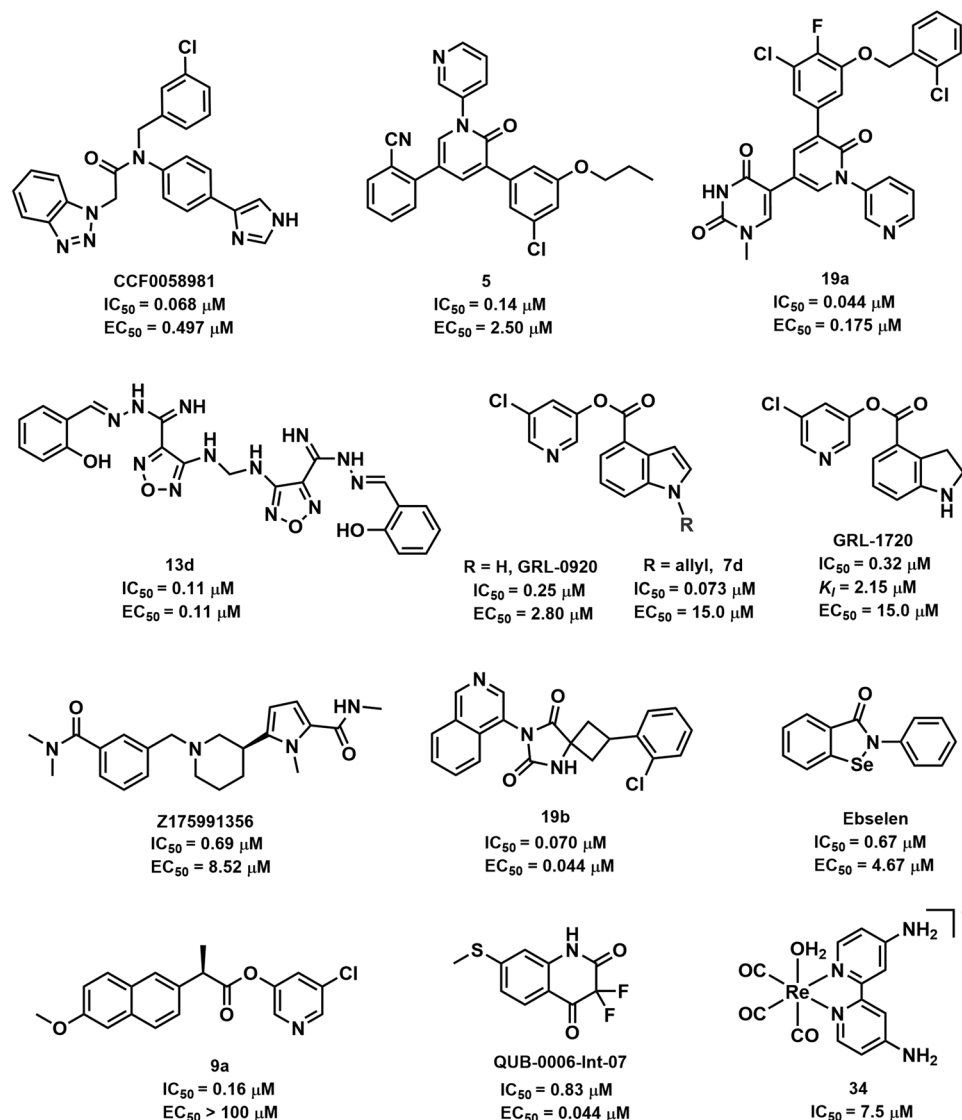


FIGURE 4 The structures and half-maximal inhibitory concentration (IC_{50}) values for representative non-peptidomimetic SARS-CoV-2 3CL^{pro} inhibitors, as well as their half-maximal effect concentration (EC_{50}) values for anti-SARS-CoV-2

and covalent inhibitors, with K_i values of 0.031 and 0.078 μM , respectively.^{150,151}

Mitsuya and coworkers have been devoted to developing effective 3CL^{pro} inhibitors for fighting SARS-CoV and SARS-CoV-2. In particular, they suggested that carbonyl indole could function as a warhead to modify Cys145 of 3CL^{pro}, and several carbonyl-indole-containing compounds were identified as covalent inhibitors, such as GRL-0920.^{147,148,152,153} Another compound, GRL-1720, an inhibitor of SARS-CoV 3CL^{pro} ($IC_{50} = 30 \text{ nM}$), could also irreversibly inhibit SARS-CoV-2 3CL^{pro} ($K_{\text{inact}} = 2.53 \text{ min}^{-1}$, $K_i = 2.15 \mu\text{M}$). Meanwhile, this compound could also block the infectivity of SARS-CoV-2^{WK-521} (SARS-CoV-2 JPN/TY/WK-521 strain) in Vero E6 cells, with an EC_{50} value of 15 μM and an apparent half-maximal cytotoxicity concentration (CC_{50}) value more

than 100 μM .^{117,148} Recently, a group of 5-chloropyridinyl indole carboxylate derivatives was designed for inhibiting SARS-CoV-2 3CL^{pro}. Among all tested compounds, **7d** was a potent SARS-CoV-2 3CL^{pro} inhibitor ($IC_{50} = 73 \text{ nM}$) that blocked viral infection in vitro ($EC_{50} = 15 \mu\text{M}$). The detailed structure-activity relationship (SAR) analysis revealed that the 5-chloropyridinyl ester was crucial for inhibitory activity. On the indole ring, the N-allyl substituent could significantly improve the activity, while the incorporation of the methyl group at position-5 and fluorine at position-6 generated a declining potency. The X-ray crystal structure of **7b** and SARS-CoV-2 3CL^{pro} presented a covalent binding mechanism.¹⁵² Subsequently, compound **9a** was identified as a SARS-CoV-2 3CL^{pro} inhibitor from another series of new 5-chloropyridinyl ester analogs, with a calculated IC_{50} value of 160 nM.¹⁵³

Recently, the SAR study of a group of benzoisothiazolone-containing SARS-CoV-2 3CL^{PRO} demonstrated that the phenyl group was optimized as the best group at the tail benzene ring, and the acetamide group in the linker was essential to the inhibitory activity. In particular, the crucial benzoisothiazolone that could function as a warhead for covalently binding to Cys145 of 3CL^{PRO} should avoid adverse steric hindrance. As shown in Table S2, **16b-3** was a promising lead compound for novel anti-COVID-19 agents.¹⁵⁴ Beyond that, the thiazolidinone derivatives (k3), QUB-00006-Int-07, VS10, and VS12 were promising inhibitors for SARS-CoV-2 3CL^{PRO}.^{155–157} Additionally, it has been reported that metal-based (such as Au, Pt, and Re) coordination compounds could form metal complexes with cysteine proteases via coordinate covalent bonds.^{158–160} Karges et al.⁹¹ preliminarily suggested that the [Re(2,2'-bipyridine)(CO)₃]⁺ fragment could bind to Cys145 of SARS-CoV-2 3CL^{PRO} with the lowest energy. Then, a series of Re(I) tricarbonyl complexes were synthesized for SARS-CoV-2 3CL^{PRO} inhibitor evaluation. Both compounds **34** and **22** formed a metal—Cys145 covalent bond with SARS-CoV-2 3CL^{PRO} in the axial position. Preliminary studies indicate that this compound was a selective inhibitor of 3CL^{PRO}, which exhibited poor inhibitory effects against several human proteases at 50 μM, including human serine protease dipeptidyl peptidase-4, aspartate protease β-secretase 1, and cysteine protease cathepsin B.

4 | NATURALLY DERIVED SARS-COV-2 3CL^{PRO} INHIBITORS

It is well known that natural compounds are still the major sources for the identification of drug lead compounds.^{161–165} Over the past few years, a number of structurally diverse natural products and their derivatives (such as flavonoids, phenolic acids, tannins, and quinones) have been found to have anti-SARS-CoV-2 3CL^{PRO} effects, and some of them have been identified as covalent 3CL^{PRO} inhibitors.^{96,166} In this review, the reported naturally derived SARS-CoV-2 3CL^{PRO} inhibitors, accompanied by their inhibitory effects and inhibitory mechanisms, were well summarized, which well explained the anti-COVID-19 effects of some herbal medicines and provided new inspiration to medicinal chemists for designing and developing novel anti-COVID-19 agents by targeting 3CL^{PRO}.

4.1 | Flavonoids and their derivatives

Flavonoids are secondary metabolites that widely exist in edible and medicinal plants and usually comprise sev-

eral subclasses, such as flavanones, flavones, flavonols, and biflavones.^{167,168} This class of polyphenol compounds is known for good safety profiles and multiple health benefits, including antioxidative, anti-inflammatory, anti-cancer, antiviral, and immunomodulatory effects.^{169–174} Recently, many flavonoids have shown strong inhibitory effects against SARS-CoV-2 3CL^{PRO}, and their inhibitory effects are listed in Figure 5 and Table S3.

Scutellaria baicalensis is a traditional Chinese medicine used for upper respiratory tract infections and possesses wide antiviral activity, including against SARS-CoV-2 (EC₅₀ = 0.74 μg/ml).^{175,176} Recently, some flavonoids in *S. baicalensis* were reported to be SARS-CoV-2 3CL^{PRO} inhibitors, such as baicalein and scutellarein.¹⁷⁷ It is well known that phenolic groups can transform into orthoquinone under oxidizing conditions, which can easily be attacked by nucleophiles (such as thiol).¹⁷⁸ According to a new study, six scutellarein-methylated derivatives were synthesized as novel 3CL^{PRO} inhibitors. 4'-O-methylscutellarein was characterized as a potent noncovalent 3CL^{PRO} inhibitor (IC₅₀ = 0.40 μM). Further SAR study demonstrated that the replacement of hydroxyl groups at the A-ring was indispensable, and hydrophobicity of the B-ring might be beneficial to inhibitory activity.¹⁷⁹ Xiong et al.⁹⁶ identified myricetin, dihydromyricetin, and isodihydromyricetin as covalent inhibitors for SARS-CoV-2 3CL^{PRO}, whose orthoquinone form could modify the key cysteines near the catalytic site (Cys145) or dimeric interface (Cys300) of the target enzyme. Meanwhile, Su et al.¹⁸⁰ also demonstrated that myricetin could covalently bind to Cys145 of SARS-CoV-2 3CL^{PRO} by using crystal structure analysis of the complex. To gain more ideal SARS-CoV-2 3CL^{PRO} inhibitors, several analogs were designed based on myricetin, among which 7-O-methyl-dihydropopulins showed the highest inhibitory effect with an IC₅₀ value of 0.26 μM. Moreover, this study revealed that the pyrogallol group could be used as an alternative electrophile warhead to develop covalent inhibitors for 3CL^{PRO}. In contrast, another study revealed that baicalein was a noncovalent inhibitor that could act as a “shield” to prevent the substrate from entering the catalytic pocket.¹⁸¹ Even though these compounds bear a pyrogallol group, the different conformations of myricetin and baicalein in the 3CL^{PRO} catalytic site generate different action modes.

Ugonin J is a flavonoid isolated from the Rhizome of *Helminthostachys zeylanica*, which has an ethyl-(2,2-dimethyl-6-methylenecyclohexyl) moiety at the C-6 position and possessed a potent inhibitory effect on SARS-CoV-2 3CL^{PRO} (IC₅₀ = 0.94 μM). Furthermore, the anti-SARS-CoV-2 activity and anti-inflammatory activity of this inhibitor have been proven in vitro, suggesting that ugonin J could be used as a leading compound to fight COVID-19.¹⁸² Quercetin has a broad spectrum

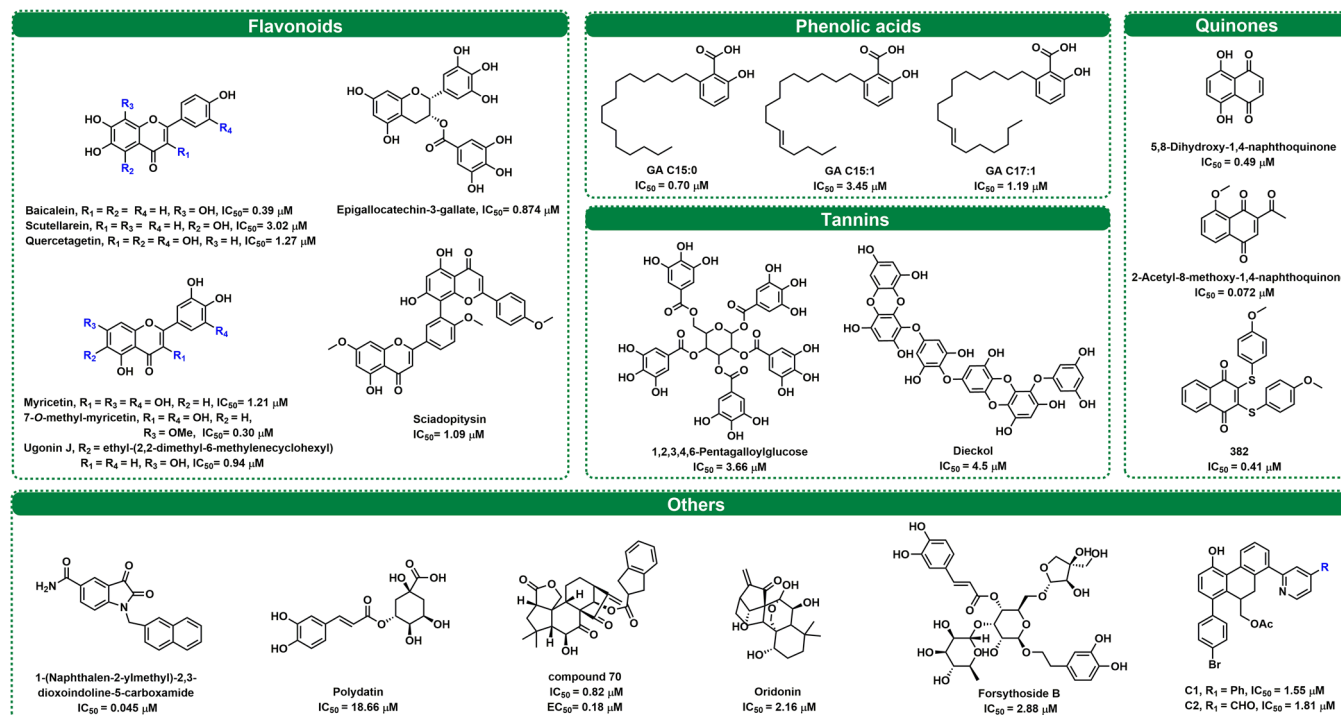


FIGURE 5 The structures and half-maximal inhibitory concentration (IC_{50}) values for representative naturally derived SARS-CoV-2 3CL^{pro} inhibitors

of antiviral activities (including poliovirus type 1, herpes simplex virus type 1 [HSV-1], HSV-2, respiratory syncytial virus, and influenza A subtypes).³⁹ Regarding anti-3CL^{pro} activity, quercetin exhibited a better inhibitory effect against SARS-CoV-2 3CL^{pro} than SARS-CoV 3CL^{pro}.^{166,183–186} Recently, Mangiavacchi et al.¹⁸⁷ synthesized and evaluated a series of compounds based on the skeletons of quercetin and chrysin. The SAR analysis suggested that the phenylselenyl moiety at the C-8 position was highly effective, while the double substitution resulted in a drop in the activity. 2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-8-(p-tolylselenanyl)-4H-chromen-4-one was a strong ($IC_{50} = 11 \mu M$) and reversible ($K_i = 3.8 \mu M$) 3CL^{pro} inhibitor, presenting a safe and effective inhibition activity ($IC_{50} = 8 \mu M$) against the replication of SARS-CoV-2 in a Vero cell model. The major tea catechins, including epigallocatechin-3-gallate, (-)-epicatechin 3-O-caffeate, as well as etc-pyrrolidinone C and D, possessed strong inhibitory effects toward 3CL^{pro}.^{188–192} In addition, some flavonoids, such as kaempferol, luteolin, genkwanin, and isorhamnetin, displayed various degrees of inhibitory activities toward 3CL^{pro}.^{192,193}

However, the bioavailability of naturally occurring flavonoids is generally poor, which are easily glycosylated by UDP-glucuronosyltransferases in vivo.¹⁹⁴ The metabolites (glycosyl flavonoids) have better solubility, stability, and bioavailability properties than their aglycones.^{193,195,196} Some molecular docking and simula-

tion studies found that glycosyl flavonoids calculated a high-affinity score for binding to 3CL^{pro}.^{197–200} By using experimental (spectroscopy and calorimetry) and simulation techniques (docking and molecular dynamics simulations), Rizzuti et al.²⁰² revealed that rutin was a promising inhibitor of SARS-CoV-2 3CL^{pro}.^{201,202} However, most studies found that the introduction of glycosides on flavonoids would weaken the inhibitory effects on 3CL^{pro}. For example, baicalin, narcissoside, and kaempferol-3-O-gentiobioside presented relatively poorer inhibitory effects than their aglycones.^{181,203} Further SAR studies demonstrated that glycosylation on the 7-hydroxy of quercetin could be allowed, but the acetoxylation of the glycosyl was adverse.²⁰⁴

Biflavones are a class of compounds with a dimer of flavonoid structure, some of which have been validated as 3CL^{pro} inhibitors.²⁰⁵ Xiong et al.¹⁶⁶ found that *Ginkgo biloba* leaves showed strong inhibitory activity against SARS-CoV-2 3CL^{pro} via a scale screening of herbal extracts, while 20 major constituents (including five biflavones) isolated from this herb were collected for SARS-CoV-2 3CL^{pro} inhibition assays. Further kinetic analyses and molecular docking suggested that sciadopitysin could strongly inhibit SARS-CoV-2 3CL^{pro} in a mixed manner, with a K_i value of $2.96 \mu M$. Other biflavones, including ginkgetin, isoginkgetin, amentoflavone, and bilobetin, could also dose dependently inhibit SARS-CoV-2 3CL^{pro}, with IC_{50} values ranging from 2.33 to $11.19 \mu M$.

4.2 | Phenolic acids

Phenolic acids are a group of secondary metabolites and bioactive compounds produced by plants.^{206,207} Xiong et al.¹⁶⁶ pointed out that four ginkgolic acids (GAs) from *Folium ginkgo* showed relatively potent SARS-CoV-2 3CL^{pro} inhibitory activity ($IC_{50} < 5 \mu M$). Further inhibition kinetic studies and docking simulations clearly showed that GAs **C15:0** and **C17:1** strongly inhibited SARS-CoV-2 3CL^{pro} in a mixed manner (Figure 5 and Table S4). Moreover, GAs (**C15:0** and **C15:1**) were identified as dual inhibitors targeting both 3CL^{pro} and PL^{pro} of SARS-CoV-2 at nontoxic concentrations by Chen et al.²⁰⁸ However, allergenic GAs are severely restricted in commercially available *G. biloba* products.²⁰⁹ There is growing evidence that GA has broad antiviral effects by interfering with viral replication.^{210,211} Additionally, Nguyen et al.²¹² reported the inhibitory activity of different phenolic acids from black garlic on SARS-CoV-2 3CL^{pro}, including gallic acid, caffeic acid, vanillic acid, ferulic acid, and chlorogenic acid. The above results suggest that these phenolic acids are worth exploring as potential new therapeutics for COVID-19.

4.3 | Tannins

Tannins displayed potent inhibitory effects against SARS-CoV-2 3CL^{pro} and SARS-CoV 3CL^{pro} with IC_{50} values at the micromolar level. Wang et al.²¹³ found tannic acid to be a potent inhibitor of SARS-CoV-2 3CL^{pro} and TMPRSS2, with an IC_{50} of $13.4 \mu M$ for SARS-CoV-2 3CL^{pro}. Consistently, tannic acid could also target the mechanisms governing virus entry.²¹³ As early as 2005, Chen et al.²¹⁵ demonstrated a significant inhibitory effect of tannic acid on SARS-CoV 3CL^{pro} ($IC_{50} = 3 \mu M$).^{190,214,215} Therefore, the above results suggest that tannic acid has a high potential for the development of anti-coronavirus therapeutics as a broad-spectrum inhibitor. As shown in Table S5, 1,2,3,4,6-pentagalloylglucose is a hydrolysable tannin that has been reported to inhibit a variety of viruses.²¹⁶ In terms of anti-3CL^{pro} activity, Chiou et al.¹⁸⁸ found that 1,2,3,4,6-pentagalloylglucose inhibited 50% of SARS-CoV-2 3CL^{pro} and SARS-CoV 3CL^{pro} at 3.66 and $6.89 \mu M$, respectively.

Additionally, Park et al.²¹⁷ evaluated the biological activity of nine phlorotannins, dieckol ($IC_{50} = 2.7 \mu M$), which possesses two eckol groups linked through a diphenyl ether and showed the most potent SARS-CoV 3CL^{pro} inhibitory activity. Up to now, Yan et al.²¹⁸ developed a novel screening method combining fluorescence polarization technology with a biotin-avidin system and identified dieckol as a new competitive inhibitor against SARS-CoV-2 3CL^{pro} with an IC_{50} value of $4.5 \mu M$. Recently,

Du et al.²¹⁹ demonstrated that chebulagic acid and punicalagin, which have been recognized as broad-spectrum antiviral agents, inhibited SARS-CoV-2 plaque formation in a dose-dependent manner, indicating that they exhibit antiviral activity in vitro. Furthermore, chebulagic acid and punicalagin exhibited reversible inhibitory effects against SARS-CoV-2 3CL^{pro} via noncompetitive modes.

4.4 | Quinones and their derivatives

Quinones are a class of cyclohexadienedione-containing or cyclohexadiene dimethylene-containing organic compounds that are usually divided into benzoquinones, naphthoquinones, phenanthraquinones, and anthraquinones.^{220,221} As shown in Figure 5 and Table S6, quinones and their derivatives provide several promising leading compounds for the development of anti-COVID-19 agents by targeting SARS-CoV-2 3CL^{pro}. It has been reported that tanshinones isolated from *Salvia miltiorrhiza* are inhibitors of SARS-CoV cysteine proteases (including 3CL^{pro} and PL^{pro}).²²² Recently, tanshinone I was shown to inhibit SARS-CoV-2 at the cellular level with an EC_{50} value of $2.26 \mu M$. These results substantiate the use of tanshinone derivatives as antiviral agents. In the meantime, tanshinone I and tanshinone IIA were identified as SARS-CoV-2 PL^{pro} inhibitors.^{223–225} Jin et al.¹⁶ found that shikonin exhibits potent inhibition of SARS-CoV-2 3CL^{pro} activity with an IC_{50} of $15.75 \mu M$. Moreover, shikonin presented a noncovalent binding configuration with multiple interactions at the S1–S4 subsites of the binding pocket and occupied the space of one water molecule of 3CL^{pro}.⁵⁶

It is well known that the specific chemical structure of quinone confers oxidative and electrophilic properties.^{220,226–228} Wang et al.²²⁹ screened vitamin K3 as a time-dependent SARS-CoV-2 3CL^{pro} inhibitor ($IC_{50} = 4.78 \mu M$ at 60 min preincubation) from Food and Drug Administration (FDA)-approved drug library. Based on this finding, a set of vitamin K3 analogs was collected for SAR analysis. The results showed that 5,8-dihydroxy-1,4-naphthoquinone could strongly and time dependently inhibit SARS-CoV-2 3CL^{pro} and covalently bind to the target enzyme. However, the high electrophilicity of quinones may lead to cytotoxicity.²³⁰ To discover safe and effective quinone-derived inhibitors against SARS-CoV-2 3CL^{pro}, Cui and Jia²³¹ designed a set of juglone-like compounds by using a simple skeleton. The results suggested that the interaction between the acetyl substituent on the quinone ring and the methyl group attached to the phenolic hydroxyl group of juglone was crucial for the inhibitory effects. Further cytotoxicity and

antiviral assays demonstrated that 2-acetyl-8-methoxy-1,4-naphthoquinone exhibited a low cytotoxic profile and good anti-SARS-CoV-2 in Vero E6 cells ($EC_{50} = 4.55 \mu\text{M}$). This study demonstrated the possibility of quinone being developed as a safe antiviral agent. Recently, four compounds were screened to inhibit SARS-CoV-2 3CL^{pro} from a compound library, with IC_{50} values ranging from 0.41 to 66 μM . Further studies suggested that compound **382** was a reversible SARS-CoV-2 3CL^{pro} inhibitor, while compound **415** might form a covalent bond with Cys145 of 3CL^{pro}.²³² Similarly, aloesin was identified as a SARS-CoV-2 3CL^{pro} inhibitor from a fluorescence resonance energy transfer (FRET)-based THS assessment.²³³

4.5 | Others

Beyond the abovementioned classes of natural compounds, other compounds derived from natural compounds were also found to have SARS-CoV-2 3CL^{pro} inhibition activity, such as alkaloids, terpenoids, and theaflavins, as well as phenylethanol glycosides.^{181,214,234,235} The compound information alongside their SARS-CoV-2 3CL^{pro} inhibitory effects are shown in Figure 5 and Table S7. Some isatin derivatives exhibited strong inhibition effects against 3CL^{pro}, such as 1-(naphthalen-2-ylmethyl)-2,3-dioxindoline-5-carboxamide, which was a promising compound for developing broad-spectrum anti-coronavirus agents.^{236,237} Zhong et al.²³⁸ identified that oridonin displayed effective inhibition of SARS-CoV-2 3CL^{pro} activity and bound to 3CL^{pro} via covalent bonding, while inhibiting SARS-CoV-2 in Vero E6 cells with an IC_{50} of 4.95 μM , above indicating that oridonin prevented SARS-CoV-2 replication by inhibiting 3CL^{pro}.²³⁹ Most recently, by pharmacophore-oriented semisynthesis combining the pharmacophore of oridonin and a novel scaffold (maoelactone A), Zhou et al.²⁴⁰ created a series of compounds with anti-SARS-CoV-2 activity, where compound **70** inhibited the replication of SARS-CoV-2-affected Vero E6 cells with low EC_{50} values.

One newly reported study indicated that six phenylethanol glycosides (including forsythoside A, B, E, H, I, and isoforsythiaside) isolated from *Forsythia suspensa* were strong inhibitors of SARS-CoV-2 3CL^{pro} (IC_{50} values range from 2.88 to 10.17 μM), which contributed to the excellent anti-SARS-CoV-2 activity of Shuanghuanglian preparation (a traditional proprietary Chinese medicine).¹⁸¹ Some other compounds, including polydatin, resveratrol, and all-*trans* retinoic acid, generated inhibitory effects against SARS-CoV-2 3CL^{pro}.^{241,242} A series of 9,10-dihydrophenanthrene derivatives were synthesized to discover strong SARS-CoV-2 3CL^{pro}

inhibitors.²⁴³ The preliminary SAR suggested that a suitable bulkier group at the C-8 position displayed good inhibitory activities. Among all derivatives, compound **C1** could dose dependently inhibit the target enzyme in a mixed manner, with an IC_{50} value of 1.55 μM and a K_i value of 6.09 μM . Further study suggested that this inhibitor had the potential to be a novel orally administered and broad-spectrum antiviral agent.²⁴³

5 | ANTI-COV CLINICAL CANDIDATES TARGETING SARS-COV-2 3CL^{pro}

5.1 | SARS-CoV-2 3CL^{pro} inhibitors under clinical trials

In view of the ongoing mutation of SARS-CoV-2 (such as Omicron), clinical studies of some antibody drugs have stagnated.^{17,244} Small-molecule anti-CoV drugs, however, have great potential to combat new CoV variants, as the convenience and flexibility of oral administration, along with the large production capacity, provide good conditions to achieve a global fight against COVID-19.²⁴⁵ The following are some clinical advances in the development of small-molecule drugs targeting SARS-CoV-2 3CL^{pro} (Figure 6 and Table 2).^{25,78,246–255}

Recently, paxlovid, a novel orally available agent combining a 3CL^{pro} inhibitor (PF-07321332) with ritonavir, has been approved by the FDA for the treatment of patients with moderate or severe COVID-19.^{247,248,256–258} PF-07321332 exhibited potent inhibition against 3CL^{pro} from various CoV types known to infect humans, as well as significant SARS-CoV-2 antiviral activity in Vero E6 cells ($EC_{50} = 74.5 \text{ nM}$). In the meantime, PF-07321332 (100 μM) showed no inhibitory activity against caspase-2, cathepsin B/D/L, chymotrypsin, elastase, thrombin, and HIV-1 protease, indicating a high selectivity for CoV proteases. A phase II/III clinical trial of PF-07321332/ritonavir (ID: NCT04960202) assessed its safety and effectiveness for treating COVID-19 in adults who did not require hospitalization. The data showed that the patients treated with this drug experienced an 89% reduction in the risk of hospitalization or death, which was highly effective. More recently, the clinical trial results from the treatment of hospitalized patients with paxlovid oral agents during the Omicron BA.2 outbreak in Hong Kong showed significantly lower disease progression composite outcomes (hazard ratio [HR] = 0.33, $p < 0.001$), significantly lower all-cause mortality (HR = 0.32, $p < 0.001$), and faster reduction in viral load (HR = 1.25, $p = 0.015$).²⁵⁹ Additionally, the novel phosphate prodrug PF-07304814, which can be rapidly converted in vivo to the active moiety of PF-00835231, has broad-spectrum inhibitory activity against a

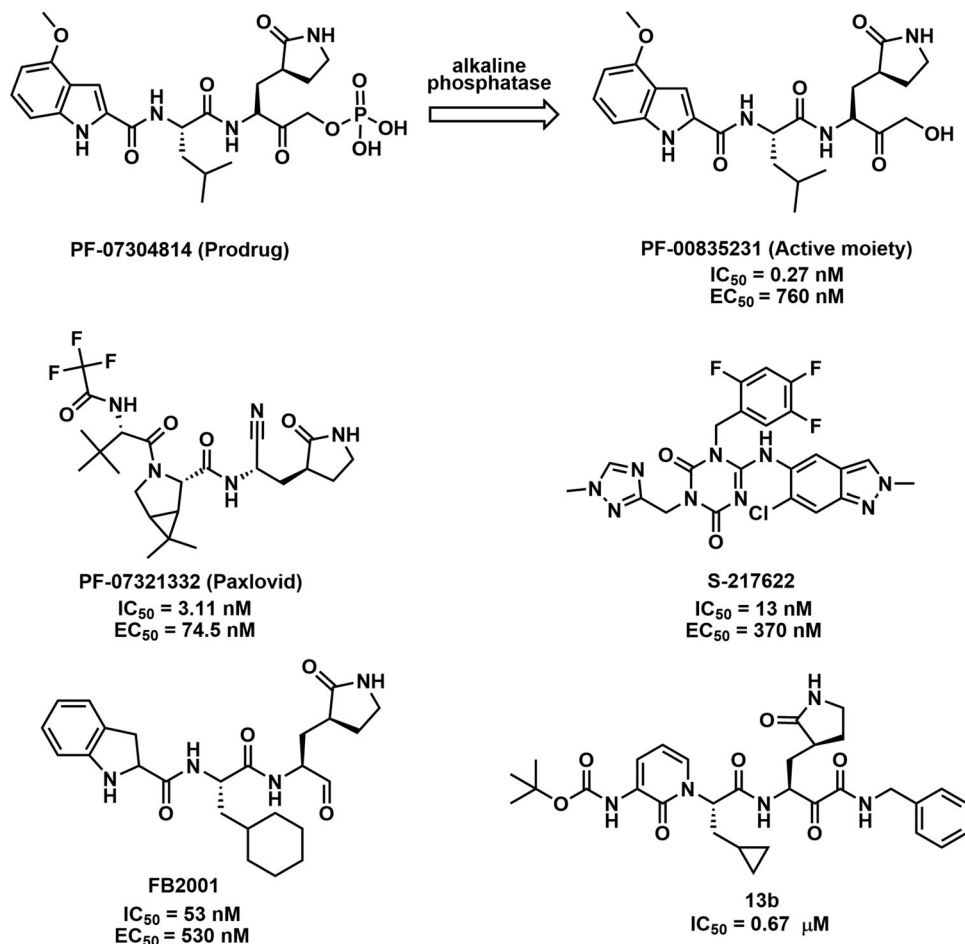


FIGURE 6 The structures and half-maximal inhibitory concentration (IC_{50}) values for representative clinical candidates SARS-CoV-2 3CL^{pro} inhibitors, as well as their half-maximal effect concentration (EC_{50}) values for anti-SARS-CoV-2

TABLE 2 Representative clinical candidates for SARS-CoV-2 3CL^{pro} inhibitors

Drug	Company	Delivery	States	IC_{50} (nM)	EC_{50} (nM)	CT.GOV ID/Ref.
PF-07321332 (Paxlovir)	Pfizer	Oral	Proved	3.11	74.5	NCT04960202 ²⁵⁸
s-217622	Shionogi	Oral	Phase III	13	370	NCT05305547 ²⁶⁰
PF-07304814	Pfizer	IV	Phase I	0.27	760	NCT05050682
FB2001/11a (DC402234)	Frontier	IV	Phase I	53	530	NCT05197179
EDP-235	Enanta	Oral	Phase I	5.8	5.1	NCT05246878
SIM0417 (SSD8432)	Simcere	Oral	Phase II	–	–	NCT05373433
PBI-0451	Pardes	Oral	Phase I	–	–	NCT05011812
13b	University of Lübeck	Inhaled	Preclinical	670	4–5 μM (Calu-3 cell)	²⁵
ALG-097111	Aligos	–	Preclinical	7	200 (A549 cell)	²⁵¹
MPI8	Sorrento	–	Preclinical	105	30	¹³⁴
ASC11	Ascleptis	Oral	Preclinical	–	–	²⁶³
EDDC-2214	Everest	Oral	Preclinical	–	–	²⁶⁴
RAY003	Zhongsheng	Oral	Preclinical	–	–	¹⁷⁹

Abbreviations: EC_{50} , half-maximal effect concentration; IC_{50} , half-maximal inhibitory concentration.

panel of 3CL^{pro} and potent antiviral activity in vivo.^{253,254} Furthermore, clinical trials in phase Ib of PF-07304814 (ID: NCT05050682) evaluated its safety, metabolism, and pharmacokinetics in patients with SARS-CoV-2 infection.

After virtual screening and SAR analysis of the hit compounds, S-217622 displayed potent inhibition activity against SARS-CoV-2 3CL^{pro} and exhibited in vitro antiviral activity against a range of CoVs, including more aggressive SARS-CoV-2 variants.^{249,250,252} The clinical trial of S-217622 (ID: NCT05305547) was a randomized, placebo-controlled, double-blind study with Japanese adults, which evaluated the antiviral effects and safety of this drug once daily for 5 days. New data indicated that the proportion of patients with positive viral titers was decreased by approximately 90% versus placebo on the fourth day of treatment.²⁶⁰ Dai et al.⁷⁸ based on the crystal structure of 3CL^{pro} designed and synthesized peptidomimetic compounds **11a** and **11b**, which possess potent antiviral activity with EC₅₀ values of 530 and 720 nM against SARS-CoV-2, respectively. Of these, FB2001 (**11a**) was an anti-CoV candidate for reaching clinical trials. In addition, FB2001 (ID: NCT05197179) demonstrated excellent safety and tolerability in the first human clinical trial conducted in the United States, for which its pharmacokinetics and safety will subsequently be evaluated in healthy Chinese populations.

At the 2022 Annual Meeting of American Society for Biochemistry and Molecular Biology, Enanta noted that EDP-235 potently inhibited the SARS-CoV-2 3CL^{pro} protease and effectively blocked the replication of SARS-CoV-2 in multiple cellular models. In addition, EDP-235 was shown to have good in vivo penetration into a variety of target tissues.²⁴⁶ Furthermore, EDP-235 (ID: NCT05246878) was evaluated in the first in-human phase I study in healthy volunteers for safety, tolerability, and pharmacokinetics. PBI-0451 (ID: NCT05011812), administered twice daily as a stand-alone agent, has shown good tolerability in the ongoing phase I clinical trial, at >20-fold single- and >14-fold multiple-total daily dosage.²⁶¹ SSD8432 (ID: NCT05373433) was the first oral SARS-CoV-2 3CL^{pro} drug approved for clinical trials in China, and its phase II clinical trial evaluated its efficacy and safety in combination with ritonavir in asymptomatic infections or mild/general safety studies in adult subjects with COVID-19. The role of an α -ketoamide inhibitor was explored, and it was found that compound **13b** could inhibit 3CL^{pro} from SARS-CoV-2, SARS-CoV, and MERS-CoV, with IC₅₀ values of 0.67, 0.90, and 0.58 μ M, respectively.²⁵ The inhibitory effect of compound **13b** on human Calu-3 cells infected with SARS-CoV-2 (EC₅₀ = 4–5 μ M). Furthermore, the pharmacokinetic profile of the optimized inhibitor revealed a clear pulmonary propensity and was suitable for administration via the inhalation route.

In brief, 3CL^{pro} inhibitor therapy is an attractive and effective pharmacotherapy for treating CoV-associated

infectious diseases, owing to its broad spectrum of antiviral activities and ability to prevent the posttranslational processing of SARS-CoV-2 polypeptides as well as reduce the risk of mutation-mediated resistance to drug therapy.²⁶²

5.2 | Old drugs as SARS-CoV-2 3CL^{pro} inhibitors

It is well known that the development of a novel drug generally takes a long time. Comprehensive clinical studies of approved drugs promote drug repurposing a shortcut for the discovery of safe and effective anti-COVID-19 agents, which can bypass animal safety studies and directly enter clinical phase II or III to ensure supply. Many approved drugs have been identified as SARS-CoV-2 3CL^{pro} inhibitors by using computational and experimental studies, such as teicoplanin, dipyridamole, hydroxychloroquine, and chloroquine,^{265,266} and their inhibitory effects are listed in Table 3.

According to the predicted poses and docking scores of complexes, 17 agents were predicted as potential inhibitors for SARS-CoV-2 3CL^{pro}, five of which could inhibit the hydrolysis of SARS-CoV-2 3CL^{pro}-catalyzed fluorescent peptide substrate.¹⁴¹ Chiou et al.²⁶⁷ identified 20 drugs as SARS-CoV-2 3CL^{pro} inhibitors in silico and in vitro. Among them, ethacrynic acid was the strongest inhibitor, with an IC₅₀ value of 1.11 μ M, while the anti-inflammatory and immunosuppressive activities of naproxen (IC₅₀ = 3.45 μ M) might be advantageous in COVID-19 treatment. A large-scale screening campaign was conducted for the anti-OC43 (one β -CoV) effects in vitro. Twenty drugs were screened out for anti-SARS-CoV-2 infection assays in A549 cells and enzyme measurements in green fluorescent protein (GFP)-expressing 293T cells. Masitinib competitively inhibited SARS-CoV-2 3CL^{pro}, both in vitro and in live cells. Moreover, this agent also significantly reduced the SARS-CoV-2 viral load in mice and inflammatory cytokines in the lungs. The clinical combination of masitinib and isoquercetin suggested that masitinib was a promising agent for the early treatment of COVID-19.²⁶⁸

CBS is a metallodrug usually used for duodenal ulcer treatment. A newly reported study found that CBS remarkably inhibited SARS-CoV-2 3CL^{pro} activity in vitro and in cellulo. Rather than the active residual (Cys145), CBS bound to the allosteric site (Cys300) and caused dimeric enzyme to dissociate into monomers.⁹⁵ Additionally, CBS exhibited potent anti-SARS-CoV-2 activity both in the cells and golden Syrian hamster model, which also inactivated SARS-CoV-2 helicase by displacing the zinc(II) ions in helicase by bismuth(III) ions. All these findings suggested that CBS was a promising agent for anti-COVID-19.²⁶⁹ Recently, merbromin was identified as a mixed inhibitor

TABLE 3 The current indications of clinical drugs and their inhibitory activities against SARS-CoV-2 3CL^{PRO}

Compound	Pharmacological activities	IC ₅₀ /K _i (μM)	Ref.
Teicoplanin	Antibacteria	1.61	265,273
Dipyridamole	Antiplatelet	0.04	265
Hydroxychloroquine	Antimalarial and anti-inflammatory	0.36	
Chloroquine	Antimalarial and anti-inflammatory	0.56	
Manidipine	Anti-hypertension	4.81	141
Lercanidipine	Anti-hypertension	16.2	
Efonidipine	Anti-hypertension	38.5	
Bedaquiline	Antituberculosis	18.7	
Ethacrynic acid	Hydragogue for treating chronic heart failure	1.11	267,274
Naproxen	Nonsteroidal anti-inflammatory drug for treating mild-to-moderate pain and arthritis	3.45	267,275
Allopurinol	Treat gout, hyperuricemia, and kidney stones	3.77	267,276
Butenafine hydrochloride	Antifungal	5.40	267,277
Raloxifene hydrochloride	Prevent osteoporosis	5.61	267,278
Tranylcypromine hydrochloride	Antidepressant and antianxiety	8.64	267,279
Saquinavir mesylate	Anti-HIV	9.92	267,280
Triptorelin acetate	Anti-prostate cancer	10.12	267,281
Goserelin acetate	Anti-prostate and breast cancer	12.02	267,282
Rocuronium bromide	Muscle relaxant	17.47	267,283
Bisacodyl	Treat constipation	17.51	267,284
Armodafinil	Promotes wakefulness	17.87	267,285
Clobetasol propionate	Treat skin conditions	18.09	267,286
Sirolimus (Rapamycin)	An immunosuppressant drug for allografting rejection therapy	22.30	267,287
Colistin sulfate	Antibacteria	23.20	267,273
Cetirizine	Relieve allergy	25.58	267,288
Bexarotene	Treat cutaneous T-cell lymphoma	26.49	267,289
Cefpodoxime proxetil	Antibacteria	32.43	267,290
Clindamycin palmitate hydrochloride	Antibacteria	33.21	267,291
Oxaliplatin	Anti-colorectal cancer	47.31	267,292
Masitinib	Inhibit tyrosine kinase	2.5/2.6	268,293
Colloidal bismuth subcitrate	Anti- <i>Helicobacter pylori</i> , anti-duodenal ulcer	0.93	97,294
Merbromin	Antibacteria	2.7	270,295
Tolcapone	Treat Parkinson's disease	7.9	271
Levothyroxine	Thyroid hormone	19.2	
Manidipine-2HCl	Anti-hypertension	10.4	
Disulfiram	Alcohol aversion	9.35	16,296
Carmofur	Antitumors	1.82	16,297
Tideglusib	Anti-Alzheimer disease	1.55	16,298
Z-FA-FMK	Inhibit cysteine proteases irreversibly	26.3	141,272
Boceprevir	Anti-HCV protease	5.40	272,299

Abbreviations: HCV, hepatitis C virus; HIV, human immunodeficiency virus; IC₅₀, half-maximal inhibitory concentration; K_i, inhibition constant.

against SARS-CoV-2 3CL^{pro}.²⁷⁰ Manidipine-2HCl served as a dual inhibitor for SARS-CoV-2 3CL^{pro} (IC₅₀ = 7.90 μM) and PL^{pro} (IC₅₀ = 14.20 μM), showing effective anti-SARS-CoV-2 activity with an EC₅₀ value of 14.5 μM.²⁷¹ However, due to the rigorous experimental conditions of cell-based assays, the majority of reported SARS-CoV-2 3CL^{pro} inhibitors were restricted to *in vitro* effects. In a new study, a novel cell-based luciferase complementation reporter assay was reported for the discovery of SARS-CoV-2 3CL^{pro} inhibitors, which could readily differentiate false positives caused by cytotoxicity. The method was further applied to the validation of cell-based inhibitory effects and cytotoxicity for 22 reported SARS-CoV-2 3CL^{pro} inhibitors.²⁷²

6 | CONCLUSIONS AND PERSPECTIVES

The ongoing COVID-19 pandemic has created a serious threat to human health and life safety worldwide, thus, there is an urgent medical need to find more effective therapeutic strategies for combating COVID-19.^{300–303} Among all validated targets for fighting CoVs, including SARS-CoV-2, the highly conserved 3D structure of 3CL^{pro} plays an essential role in CoV replication, and no known human protease possesses a similar cleavage specificity, making 3CL^{pro} an ideal target for developing clinically effective anti-SARS-CoV-2 agents.^{21,60,109,304,305} It is worth noting that a wide range of compounds have been found to have strong to moderate SARS-CoV-2 3CL^{pro} inhibitory effects in the past few years. To better understand the structural features of SARS-CoV-2 3CL^{pro} inhibitors and their inhibitory mechanisms, this study systematically summarized the reported structurally diverse SARS-CoV-2 3CL^{pro} inhibitors (including marketed drugs and other synthetic compounds, herbal constituents, and their derivatives), as well as their inhibition potentials and mechanisms of action. The information and knowledge presented here offer a basic reference for medicinal chemists to design and develop more effective 3CL^{pro} inhibitors as novel anti-SARS-CoV-2 agents.

Targeting the key amino acids surrounding the catalytic site to block the hydrolytic process of 3CL^{pro} is one of the practical strategies for developing efficacious 3CL^{pro} inhibitors.^{30,31,34,36,37,133} According to the different inhibitory mechanisms, all reported SARS-CoV-2 3CL^{pro} inhibitors can be divided into reversible inhibitors and covalent inhibitors. Most reversible inhibitors of SARS-CoV-2 3CL^{pro} exhibit micromolar activity, and these agents have difficulty blocking the hydrolytic activity of 3CL^{pro} *in vivo*. In contrast, covalent SARS-CoV-2 3CL^{pro} inhibitors bear at least one of the warheads (such as pyrogallol groups, quinones, or Michael receptors), which are capable of inhibiting viral replication by forming a covalent

bond with the thiol of key cysteines on the catalytic site and dimeric interface of 3CL^{pro} (such as Cys145, Cys300, and Cys44). Compared to reversible inhibitors, these covalent inhibitors tend to show stronger and prolonged activities, which motivates medicinal chemists to develop more potent covalent inhibitors against 3CL^{pro}. Although a number of synthetic compounds and natural compounds have been identified as SARS-CoV-2 3CL^{pro} covalent inhibitors, most of them show poor bioavailability, poor metabolic stability, and poor aqueous solubility. In the future, the anti-SARS-CoV-2 3CL^{pro} potency and drug-likeness properties should be improved simultaneously to overcome these limitations.³⁰⁶

Another alternative potential strategy for designing SARS-CoV-2 3CL^{pro} inhibitors is to block the formation of 3CL^{pro} dimers, the active form of this key enzyme.^{307,308} Considering that the hydrolytic activity of 3CL^{pro} relies on its dimeric form, inhibitors targeting protein self-association that disturb dimerization formation and stabilization by destroying the key interactions essential for 3CL^{pro} are also highly desirable. Such agents can prevent virus replication and proliferation in the invisible battlefield against this enigmatic and rapidly evolving virus. It has been reported that some known SARS-CoV-2 3CL^{pro} inhibitors can bind to either the catalytic site or the allosteric sites (especially the dimer interface) via different binding modes, including competitive, noncompetitive, and mixed manners. Theoretically, it is more likely to block SARS-CoV-2 3CL^{pro} by using combinations of various 3CL^{pro} inhibitors that target different ligand-binding sites, which may display synergistic 3CL^{pro} inhibitory effects via different inhibitory modes (such as occupying the catalytic domain and blocking dimerization formation).

In addition to 3CL^{pro} inhibition activity, many synthetic agents and herbal constituents (including flavonoids, alkaloids, and polyphenols) have been found to have strong inhibitory or modulatory effects on other key targets for treating CoVs (such as PL^{pro}, RNA-dependent RNA polymerase [RdRp], and TMPRSS2).^{125,173,309–312} It is well known that herbal medicines contain numerous compounds, while various constituents may interact with different anti-CoV targets or different ligand-binding sites.^{313,314} In these cases, the synergetic effects of multiple components from herbal medicines should be carefully investigated, which may partially explain the excellent anti-COVID-19 activities of some marketed Chinese medicines.³¹⁴ Furthermore, cathepsin L (a lysosomal cysteine protease in the host that cleaves furin-induced SARS-CoV-2 S protein into smaller fragments and activates its membrane fusion) has also been identified as a key target participating in SARS-CoV-2 infection.^{112,123,315,316} Thus, it is highly recommended to develop more efficacious dual inhibitors by targeting both viral protease and host cathepsin L to combat COVID-19.¹²⁵

COVID-19 is a complex, multi-organ, and heterogeneous illness, and severe disease cases are frequently accompanied by a hypercoagulable inflammatory state.^{317,318} Thus, an ideal anti-COVID-19 medication should have multiple pharmacological activities, such as anti-inflammatory, anticoagulant, anti-CoV, and immunomodulatory activities. Numerous studies have confirmed that several marketed Chinese medicines display significant anti-inflammatory and immunomodulatory effects in vivo, achieving good protective effects on the organs as well as inhibiting viral replication.^{319,320} For example, Qingfei Paidu Decoction, a widely used Chinese medicine prescription for the treatment of COVID-19 in China, has been found to have multiple pharmacological activities, including anti-inflammatory, immunomodulatory, and antiviral effects.³²¹⁻³²⁵ In the future, to obtain better therapeutic effects, the clinically used Chinese medicine prescriptions for treating COVID-19 can be used in combination with marketed anti-CoV agents for clinical observations in a reasonable dose range,³⁰¹ which may be beneficial to COVID-19 patients with pre-existing diseases (e.g., cardiovascular disease, diabetes, and pulmonary disease).

ACKNOWLEDGMENTS

This work was supported by the National Natural Science Foundation of China (82141203, 81922070, 81973286), Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine (ZYYCXTD-D-202004), Three-year Action Plan for Shanghai TCM Development and Inheritance Program (ZY(2021-2023)-0401), The Basic Public Welfare Research Program of Zhejiang Province (LGF22H280012), Shanghai Science and Technology Innovation Action Plans (20S21901500 and 20S21900900), Shanghai Science and Technology Committee, Zhejiang Provincial Medical and Health Science and Technology Program (2022495401), Hangzhou Medical College Basic Research Program (KYQN202124), Chinese Medicine Research Program of Zhejiang Province (2020ZZ003 and 2021ZZ001), “10000 Talents Plan” of Zhejiang Province (2020R52029), and Zhejiang Provincial Program for the Cultivation of New Health Talents to Yiwen Zhang.

CONFLICTS OF INTEREST

The authors declare they have no conflicts of interest.

AUTHOR CONTRIBUTIONS

Q.H., Y.X., and G.H.Z. drafted this manuscript and prepared the figures. Y.N.Z. and Y.W.Z. participated in the collection of the related literature. P.H. and G.B.G. supervised the review process. All authors have read and approved the final manuscript.

ETHICS STATEMENT

No ethical approval is required.

DATA AVAILABILITY STATEMENTS

All data are freely available from the corresponding author upon request.

REFERENCES

1. Artika IM, Dewantari AK, Wiyatno A. Molecular biology of coronaviruses: current knowledge. *Heliyon*. 2020;6(8):e04743.
2. Gorbalenya AE, Enjuanes L, Ziebuhr J, et al. Nidovirales: evolving the largest RNA virus genome. *Virus Res*. 2006;117(1):17-37.
3. Mirza MU, Froeyen M. Structural elucidation of SARS-CoV-2 vital proteins: computational methods reveal potential drug candidates against main protease, Nsp12 polymerase and Nsp13 helicase. *J Pharm Anal*. 2020;10(4):320-328.
4. Raj VS, Mou H, Smits SL, et al. Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature*. 2013;495(7440):251-254.
5. Wang Y, Grunewald M, Perlman S. Coronaviruses: an updated overview of their replication and pathogenesis. *Methods Mol Biol*. 2020;2203:1-29.
6. Zuniga S, Sola I, Alonso S, et al. Sequence motifs involved in the regulation of discontinuous coronavirus subgenomic RNA synthesis. *J Virol*. 2004;78(2):980-994.
7. Li Q, Guan X, Wu P, et al. Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. *N Engl J Med*. 2020;382(13):1199-1207.
8. Mitsuya H, Kokudo N. Sustaining containment of COVID-19: global sharing for pandemic response. *Glob Health Med*. 2020;2(2):53-55.
9. Tu H, Tu S, Gao S, et al. Current epidemiological and clinical features of COVID-19; a global perspective from China. *J Infect*. 2020;81(1):1-9.
10. Zhu N, Zhang D, Wang W, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med*. 2020;382(8):727-733.
11. Alderson MR, Arkwright PD, Bai X, et al. Surveillance and control of meningococcal disease in the COVID-19 era: a global meningococcal initiative review. *J Infect*. 2022;84(3):289-296.
12. Li X, Geng M, Peng Y, et al. Molecular immune pathogenesis and diagnosis of COVID-19. *J Pharm Anal*. 2020;10(2):102-108.
13. Ramos-Guzman CA, Ruiz-Pernia JJ, Tunon I. Multiscale simulations of SARS-CoV-2 3CL protease inhibition with aldehyde derivatives. Role of protein and inhibitor conformational changes in the reaction mechanism. *ACS Catal*. 2021;11(7):4157-4168.
14. Xie X, Hu L, Xue H, et al. Prognosis and treatment of complications associated with COVID-19: a systematic review and meta-analysis. *Acta Mater Med*. 2022;1(1).
15. Gurung AB, Ali MA, Lee J, et al. Structural and functional insights into the major mutations of SARS-CoV-2 spike RBD and its interaction with human ACE2 receptor. *J King Saud Univ Sci*. 2022;34(2):101773.
16. Jin Z, Du X, Xu Y, et al. Structure of M(pro) from SARS-CoV-2 and discovery of its inhibitors. *Nature*. 2020;582(7811):289-293.
17. Berkhout B, Herrera-Carrillo E. SARS-CoV-2 evolution: on the sudden appearance of the Omicron variant. *J Virol*. 2022;96(7):e0009022.

18. del Rio C, Omer SB, Malani PN. Winter of Omicron—the evolving COVID-19 pandemic. *JAMA*. 2022;327(4):319-320.
19. Li Q, Kang C. Progress in developing inhibitors of SARS-CoV-2 3C-like protease. *Microorganisms*. 2020;8(8):1250.
20. Tian D, Sun YH, Zhou JM, et al. The global epidemic of SARS-CoV-2 variants and their mutational immune escape. *J Med Virol*. 2022;94(3):847-857.
21. Qiao J, Li YS, Zeng R, et al. SARS-CoV-2 M(pro) inhibitors with antiviral activity in a transgenic mouse model. *Science*. 2021;371(6536):1374-1378.
22. Anand K, Palm GJ, Mesters JR, et al. Structure of coronavirus main proteinase reveals combination of a chymotrypsin fold with an extra alpha-helical domain. *EMBO J*. 2002;21(13):3213-3224.
23. He J, Hu L, Huang X, et al. Potential of coronavirus 3C-like protease inhibitors for the development of new anti-SARS-CoV-2 drugs: insights from structures of protease and inhibitors. *Int J Antimicrob Agents*. 2020;56(2):106055.
24. Pillaiyar T, Manickam M, Namasivayam V, et al. An overview of severe acute respiratory syndrome-coronavirus (SARS-CoV) 3CL protease inhibitors: peptidomimetics and small molecule chemotherapy. *J Med Chem*. 2016;59(14):6595-6628.
25. Zhang L, Lin D, Sun X, et al. Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved alpha-ketoamide inhibitors. *Science*. 2020;368(6489):409-412.
26. Ziebuhr J, Snijder EJ, Gorbalenya AE. Virus-encoded proteinases and proteolytic processing in the Nidovirales. *J Gen Virol*. 2000;81(Pt 4):853-879.
27. Cui W, Cui S, Chen C, et al. The crystal structure of main protease from mouse hepatitis virus A59 in complex with an inhibitor. *Biochem Biophys Res Commun*. 2019;511(4):794-799.
28. Galasiti Kankanamalage AC, Kim Y, Damalanka VC, et al. Structure-guided design of potent and permeable inhibitors of MERS coronavirus 3CL protease that utilize a piperidine moiety as a novel design element. *Eur J Med Chem*. 2018;150:334-346.
29. Kanitz M, Blanck S, Heine A, et al. Structural basis for catalysis and substrate specificity of a 3C-like cysteine protease from a mosquito mesonivirus. *Virology*. 2019;533:21-33.
30. Lee C-C, Kuo C-J, Ko T-P, et al. Structural basis of inhibition specificities of 3C and 3C-like proteases by zinc-coordinating and peptidomimetic compounds. *J Biol Chem*. 2009;284(12):7646-7655.
31. St John SE, Tomar S, Stauffer SR, et al. Targeting zoonotic viruses: structure-based inhibition of the 3C-like protease from bat coronavirus HKU4 – the likely reservoir host to the human coronavirus that causes Middle East Respiratory Syndrome (MERS). *Bioorg Med Chem*. 2015;23(17):6036-6048.
32. Tian X, Lu G, Gao F, et al. Structure and cleavage specificity of the chymotrypsin-like serine protease (3CLSP/nsp4) of porcine reproductive and respiratory syndrome virus (PRRSV). *J Mol Biol*. 2009;392(4):977-993.
33. Wang F, Chen C, Liu X, et al. Crystal structure of feline infectious peritonitis virus main protease in complex with synergetic dual inhibitors. *J Virol*. 2016;90(4):1910-1917.
34. Wang F, Chen C, Yang K, et al. Michael acceptor-based peptidomimetic inhibitor of main protease from porcine epidemic diarrhea virus. *J Med Chem*. 2017;60(7):3212-3216.
35. Xue X, Yu H, Yang H, et al. Structures of two coronavirus main proteases: implications for substrate binding and antiviral drug design. *J Virol*. 2008;82(5):2515-2527.
36. Yang H, Yang M, Ding Y, et al. The crystal structures of severe acute respiratory syndrome virus main protease and its complex with an inhibitor. *Proc Natl Acad Sci U S A*. 2003;100(23):13190-13195.
37. Zhao Q, Li S, Xue F, et al. Structure of the main protease from a global infectious human coronavirus, HCoV-HKU1. *J Virol*. 2008;82(17):8647-8655.
38. Shagufta Ahmad I. The race to treat COVID-19: potential therapeutic agents for the prevention and treatment of SARS-CoV-2. *Eur J Med Chem*. 2021;213:113157.
39. Wu C, Liu Y, Yang Y, et al. Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods. *Acta Pharm Sin B*. 2020;10(5):766-788.
40. Liu Y, Liang C, Xin L, et al. The development of coronavirus 3C-Like protease (3CL(pro)) inhibitors from 2010 to 2020. *Eur J Med Chem*. 2020;206:112711.
41. Boozari M, Hosseinzadeh H. Natural products for COVID-19 prevention and treatment regarding to previous coronavirus infections and novel studies. *Phytother Res*. 2021;35(2):864-876.
42. Gowrishankar S, Muthumanickam S, Kamaladevi A, et al. Promising phytochemicals of traditional Indian herbal steam inhalation therapy to combat COVID-19—an in silico study. *Food Chem Toxicol*. 2021;148:111966.
43. Luo L, Jiang J, Wang C, et al. Analysis on herbal medicines utilized for treatment of COVID-19. *Acta Pharm Sin B*. 2020;10(7):1192-1204.
44. Meng L, Hong G. Application of traditional Chinese medicine in treating COVID-19. *Chin Med Cult*. 2021;4(1).
45. Mirzaie A, Halaji M, Dehkordi FS, et al. A narrative literature review on traditional medicine options for treatment of corona virus disease 2019 (COVID-19). *Complement Ther Clin Pract*. 2020;40:101214.
46. Ren W, Liang P, Ma Y, et al. Research progress of traditional Chinese medicine against COVID-19. *Biomed Pharmacother*. 2021;137:111310.
47. Tassakka A, Sumule O, Massi MN, et al. Potential bioactive compounds as SARS-CoV-2 inhibitors from extracts of the marine red alga *Halymenia durvillei* (Rhodophyta)—a computational study. *Arab J Chem*. 2021;14(11):103393.
48. Huang K, Zhang P, Zhang Z, et al. Traditional Chinese Medicine (TCM) in the treatment of COVID-19 and other viral infections: efficacies and mechanisms. *Pharmacol Ther*. 2021;225:107843.
49. Lyu M, Fan G, Xiao G, et al. Traditional Chinese medicine in COVID-19. *Acta Pharm Sin B*. 2021;11(11):3337-3363.
50. Ni L, Chen L, Huang X, et al. Combating COVID-19 with integrated traditional Chinese and Western medicine in China. *Acta Pharm Sin B*. 2020;10(7):1149-1162.
51. Ren JL, Zhang AH, Wang XJ. Traditional Chinese medicine for COVID-19 treatment. *Pharmacol Res*. 2020;155:104743.
52. Yang Y, Islam MS, Wang J, et al. Traditional Chinese medicine in the treatment of patients infected with 2019-new coronavirus (SARS-CoV-2): a review and perspective. *Int J Biol Sci*. 2020;16(10):1708-1717.
53. Dogan K, Erol E, Didem Orhan M, et al. Instant determination of the artemisinin from various *Artemisia annua* L. extracts by LC-ESI-MS/MS and their in-silico modelling and in vitro antiviral activity studies against SARS-CoV-2. *Phytochem Anal*. 2022;33(2):303-319.
54. Fayyazi N, Mostashari-Rad T, Ghasemi JB, et al. Molecular dynamics simulation, 3D-pharmacophore and scaffold

- hopping analysis in the design of multi-target drugs to inhibit potential targets of COVID-19. *J Biomol Struct Dyn*. 2021;1:22.
55. Guijarro-Real C, Plazas M, Rodriguez-Burruezo A, et al. Potential in vitro inhibition of selected plant extracts against SARS-CoV-2 chymotrypsin-like protease (3CL(Pro)) activity. *Foods*. 2021;10(7):1503.
 56. Li J, Zhou X, Zhang Y, et al. Crystal structure of SARS-CoV-2 main protease in complex with the natural product inhibitor shikonin illuminates a unique binding mode. *Sci Bull*. 2021;66(7):661-663.
 57. Mandal A, Jha AK, Hazra B. Plant products as inhibitors of coronavirus 3CL protease. *Front Pharmacol*. 2021;12:583387.
 58. Rizzuti B, Ceballos-Laita L, Ortega-Alarcon D, et al. Submicromolar inhibition of SARS-CoV-2 3CLpro by natural compounds. *Pharmaceuticals*. 2021;14(9):892.
 59. Tahir Ul Qamar M, Alqahtani SM, Alamri MA, et al. Structural basis of SARS-CoV-2 3CL(pro) and anti-COVID-19 drug discovery from medicinal plants. *J Pharm Anal*. 2020;10(4):313-319.
 60. Abe K, Kabe Y, Uchiyama S, et al. Pro108Ser mutation of SARS-CoV-2 3CL(pro) reduces the enzyme activity and ameliorates the clinical severity of COVID-19. *Sci Rep*. 2022;12(1):1299.
 61. Denesyuk AI, Permyakov EA, Johnson MS, et al. Structural and functional significance of the amino acid differences Val35Thr, Ser46Ala, Asn65Ser, and Ala94Ser in 3C-like proteinases from SARS-CoV-2 and SARS-CoV. *Int J Biol Macromol*. 2021;193(Pt B):2113-2120.
 62. Feng J, Li D, Zhang J, et al. Crystal structure of SARS-CoV 3C-like protease with baicalein. *Biochem Biophys Res Commun*. 2022;611:190-194.
 63. Kneller DW, Phillips G, O'Neill HM, et al. Room-temperature X-ray crystallography reveals the oxidation and reactivity of cysteine residues in SARS-CoV-2 3CL M(pro): insights into enzyme mechanism and drug design. *IUCrJ*. 2020;7(Pt 6):1028-1035.
 64. Noske GD, Nakamura AM, Gawriljuk VO, et al. A crystallographic snapshot of SARS-CoV-2 main protease maturation process. *J Mol Biol*. 2021;433(18):167118.
 65. Pablos I, Machado Y, de Jesus HCR, et al. Mechanistic insights into COVID-19 by global analysis of the SARS-CoV-2 3CL(pro) substrate degradome. *Cell Rep*. 2021;37(4):109892.
 66. Barrila J, Gabelli SB, Bacha U, et al. Mutation of Asn28 disrupts the dimerization and enzymatic activity of SARS 3CL(pro). *Biochemistry*. 2010;49(20):4308-4317.
 67. Chen H, Wei P, Huang C, et al. Only one protomer is active in the dimer of SARS 3C-like proteinase. *J Biol Chem*. 2006;281(20):13894-13898.
 68. Chou CY, Chang HC, Hsu WC, et al. Quaternary structure of the severe acute respiratory syndrome (SARS) coronavirus main protease. *Biochemistry*. 2004;43(47):14958-14970.
 69. Hsu MF, Kuo CJ, Chang KT, et al. Mechanism of the maturation process of SARS-CoV 3CL protease. *J Biol Chem*. 2005;280(35):31257-31266.
 70. Kidera A, Moritsugu K, Ekimoto T, et al. Allosteric regulation of 3CL protease of SARS-CoV-2 and SARS-CoV observed in the crystal structure ensemble. *J Mol Biol*. 2021;433(24):167324.
 71. Tomar S, Johnston ML, St John SE, et al. Ligand-induced dimerization of Middle East respiratory syndrome (MERS) coronavirus nsp5 protease (3CLpro): implications for nsp5 regulation and the development of antivirals. *J Biol Chem*. 2015;290(32):19403-19422.
 72. Wei P, Fan K, Chen H, et al. The N-terminal octapeptide acts as a dimerization inhibitor of SARS coronavirus 3C-like proteinase. *Biochem Biophys Res Commun*. 2006;339(3):865-872.
 73. Akbulut E. Investigation of changes in protein stability and substrate affinity of 3CL-protease of SARS-CoV-2 caused by mutations. *Genet Mol Biol*. 2022;45(2):e20210404.
 74. Lockbaum GJ, Reyes AC, Lee JM, et al. Crystal structure of SARS-CoV-2 main protease in complex with the non-covalent inhibitor ML188. *Viruses*. 2021;13(2):174.
 75. Douangamath A, Fearon D, Gehrtz P, et al. Crystallographic and electrophilic fragment screening of the SARS-CoV-2 main protease. *Nat Commun*. 2020;11(1):5047.
 76. Greasley SE, Noell S, Plotnikova O, et al. Structural basis for the in vitro efficacy of nirmatrelvir against SARS-CoV-2 variants. *J Biol Chem*. 2022;298(6):101972.
 77. Mishra B, Ballaney P, Saha G, et al. An in silico discovery of potential 3CL protease inhibitors of SARS-CoV-2 based upon inactivation of the cysteine 145-Histidine 41 catalytic dyad. *J Biomol Struct Dyn*. 2022;1-20.
 78. Dai W, Zhang B, Jiang XM, et al. Structure-based design of antiviral drug candidates targeting the SARS-CoV-2 main protease. *Science*. 2020;368(6497):1331-1335.
 79. Lu IL, Mahindroo N, Liang PH, et al. Structure-based drug design and structural biology study of novel nonpeptide inhibitors of severe acute respiratory syndrome coronavirus main protease. *J Med Chem*. 2006;49(17):5154-5161.
 80. Rathnayake AD, Zheng J, Kim Y, et al. 3C-like protease inhibitors block coronavirus replication in vitro and improve survival in MERS-CoV-infected mice. *Sci Transl Med*. 2020;12(557):eabc5332.
 81. Lee CC, Kuo CJ, Hsu MF, et al. Structural basis of mercury- and zinc-conjugated complexes as SARS-CoV 3C-like protease inhibitors. *FEBS Lett*. 2007;581(28):5454-5458.
 82. Webber SE, Okano K, Little TL, et al. Tripeptide aldehyde inhibitors of human rhinovirus 3C protease: design, synthesis, biological evaluation, and cocrystal structure solution of PI glutamine isosteric replacements. *J Med Chem*. 1998;41(15):2786-2805.
 83. Chen S, Zhang J, Hu T, et al. Residues on the dimer interface of SARS coronavirus 3C-like protease: dimer stability characterization and enzyme catalytic activity analysis. *J Biochem*. 2008;143(4):525-536.
 84. Needle D, Lountos GT, Waugh DS. Structures of the Middle East respiratory syndrome coronavirus 3C-like protease reveal insights into substrate specificity. *Acta Crystallogr D Biol Crystallogr*. 2015;71(Pt 5):1102-1111.
 85. Shi J, Song J. The catalysis of the SARS 3C-like protease is under extensive regulation by its extra domain. *FEBS J*. 2006;273(5):1035-1045.
 86. Shi J, Wei Z, Song J. Dissection study on the severe acute respiratory syndrome 3C-like protease reveals the critical role of the extra domain in dimerization of the enzyme: defining the extra domain as a new target for design of highly specific protease inhibitors. *J Biol Chem*. 2004;279(23):24765-24773.
 87. Yin J, Niu C, Cherney MM, et al. A mechanistic view of enzyme inhibition and peptide hydrolysis in the active site of the SARS-CoV 3C-like peptidase. *J Mol Biol*. 2007;371(4):1060-1074.
 88. Kneller DW, Zhang Q, Coates L, et al. Michaelis-like complex of SARS-CoV-2 main protease visualized by room-temperature X-ray crystallography. *IUCrJ*. 2021;8(Pt 6):973-979.

89. Achutha AS, Pushpa VL, Suchitra S. Theoretical insights into the anti-SARS-CoV-2 activity of chloroquine and its analogs and in silico screening of main protease inhibitors. *J Proteome Res.* 2020;19(11):4706-4717.
90. Xiong M, Su H, Zhao W, et al. What coronavirus 3C-like protease tells us: from structure, substrate selectivity, to inhibitor design. *Med Res Rev.* 2021;41(4):1965-1998.
91. Karges J, Kalaj M, Gembicky M, et al. Re(I) tricarbonyl complexes as coordinate covalent inhibitors for the SARS-CoV-2 main cysteine protease. *Angew Chem Int Ed Engl.* 2021;60(19):10716-10723.
92. Kuzikov M, Costanzi E, Reinshagen J, et al. Identification of inhibitors of SARS-CoV-2 3CL-pro enzymatic activity using a small molecule in vitro repurposing screen. *ACS Pharmacol Transl Sci.* 2021;4(3):1096-1110.
93. Ampornnanai K, Meng X, Shang W, et al. Inhibition mechanism of SARS-CoV-2 main protease by ebsele and its derivatives. *Nat Commun.* 2021;12(1):3061.
94. Bai B, Belovodskiy A, Hena M, et al. Peptidomimetic alpha-acetyloxymethylketone warheads with six-membered lactam P1 glutamine mimic: SARS-CoV-2 3CL protease inhibition, coronavirus antiviral activity, and in vitro biological stability. *J Med Chem.* 2022;65(4):2905-2925.
95. Verma N, Henderson JA, Shen J. Proton-coupled conformational activation of SARS coronavirus main proteases and opportunity for designing small-molecule broad-spectrum targeted covalent inhibitors. *J Am Chem Soc.* 2020;142(52):21883-21890.
96. Xiong Y, Zhu GH, Zhang YN, et al. Flavonoids in *Ampelopsis grossedentata* as covalent inhibitors of SARS-CoV-2 3CL(pro): inhibition potentials, covalent binding sites and inhibitory mechanisms. *Int J Biol Macromol.* 2021;187:976-987.
97. Tao X, Zhang L, Du L, et al. Allosteric inhibition of SARS-CoV-2 3CL protease by colloidal bismuth subcitrate. *Chem Sci.* 2021;12(42):14098-14102.
98. Ullrich S, Nitsche C. The SARS-CoV-2 main protease as drug target. *Bioorg Med Chem Lett.* 2020;30(17):127377.
99. Citarella A, Scala A, Piperno A, et al. SARS-CoV-2 M(pro): a potential target for peptidomimetics and small-molecule inhibitors. *Biomolecules.* 2021;11(4):607.
100. Mengist HM, Mekonnen D, Mohammed A, et al. Potency, safety, and pharmacokinetic profiles of potential inhibitors targeting SARS-CoV-2 main protease. *Front Pharmacol.* 2020;11:630500.
101. Gentilucci L, Tolomelli A, Squassabia F. Peptides and peptidomimetics in medicine, surgery and biotechnology. *Curr Med Chem.* 2006;13(20):2449-2466.
102. Kim Y, Lovell S, Tiew KC, et al. Broad-spectrum antivirals against 3C or 3C-like proteases of picornaviruses, noroviruses, and coronaviruses. *J Virol.* 2012;86(21):11754-11762.
103. Hung HC, Ke YY, Huang SY, et al. Discovery of M protease inhibitors encoded by SARS-CoV-2. *Antimicrob Agents Chemother.* 2020;64(9):e00872-20.
104. Caceres CJ, Cardenas-Garcia S, Carnaccini S, et al. Efficacy of GC-376 against SARS-CoV-2 virus infection in the K18 hACE2 transgenic mouse model. *Sci Rep.* 2021;11(1):9609.
105. Vuong W, Khan MB, Fischer C, et al. Feline coronavirus drug inhibits the main protease of SARS-CoV-2 and blocks virus replication. *Nat Commun.* 2020;11(1):4282.
106. Iketani S, Forouhar F, Liu H, et al. Author correction: lead compounds for the development of SARS-CoV-2 3CL protease inhibitors. *Nat Commun.* 2021;12(1):2708.
107. Dampalla CS, Zheng J, Perera KD, et al. Postinfection treatment with a protease inhibitor increases survival of mice with a fatal SARS-CoV-2 infection. *Proc Natl Acad Sci U S A.* 2021;118(29):e2101555118.
108. Gant TG. Using deuterium in drug discovery: leaving the label in the drug. *J Med Chem.* 2014;57(9):3595-3611.
109. Dampalla CS, Rathnayake AD, Perera KD, et al. Structure-guided design of potent inhibitors of SARS-CoV-2 3CL protease: structural, biochemical, and cell-based studies. *J Med Chem.* 2021;64(24):17846-17865.
110. Dampalla CS, Kim Y, Bickmeier N, et al. Structure-guided design of conformationally constrained cyclohexane inhibitors of severe acute respiratory syndrome coronavirus-2 3CL protease. *J Med Chem.* 2021;64(14):10047-10058.
111. Dai W, Jochmans D, Xie H, et al. Design, synthesis, and biological evaluation of peptidomimetic aldehydes as broad-spectrum inhibitors against enterovirus and SARS-CoV-2. *J Med Chem.* 2022;65(4):2794-2808.
112. Ma C, Sacco MD, Hurst B, et al. Boceprevir, GC-376, and calpain inhibitors II, XII inhibit SARS-CoV-2 viral replication by targeting the viral main protease. *Cell Res.* 2020;30(8):678-692.
113. Milligan JC, Zeisner TU, Papageorgiou G, et al. Identifying SARS-CoV-2 antiviral compounds by screening for small molecule inhibitors of Nsp5 main protease. *Biochem J.* 2021;478(13):2499-2515.
114. Wang Y, Xu B, Ma S, et al. Discovery of SARS-CoV-2 3CL(Pro) peptidomimetic inhibitors through the catalytic dyad histidine-specific protein-ligand interactions. *Int J Mol Sci.* 2022;23(4):2392.
115. Elseginy SA, Fayed B, Hamdy R, et al. Promising anti-SARS-CoV-2 drugs by effective dual targeting against the viral and host proteases. *Bioorg Med Chem Lett.* 2021;43:128099.
116. Ge R, Shen Z, Yin J, et al. Discovery of SARS-CoV-2 main protease covalent inhibitors from a DNA-encoded library selection. *SLAS Discov.* 2022;27(2):79-85.
117. Hattori SI, Higashi-Kuwata N, Hayashi H, et al. A small molecule compound with an indole moiety inhibits the main protease of SARS-CoV-2 and blocks virus replication. *Nat Commun.* 2021;12(1):668.
118. Konno S, Kobayashi K, Senda M, et al. 3CL protease inhibitors with an electrophilic arylketone moiety as anti-SARS-CoV-2 agents. *J Med Chem.* 2022;65(4):2926-2939.
119. Kitamura N, Sacco MD, Ma C, et al. Expedited approach toward the rational design of noncovalent SARS-CoV-2 main protease inhibitors. *J Med Chem.* 2022;65(4):2848-2865.
120. Bai B, Arutyunova E, Khan MB, et al. Peptidomimetic nitrile warheads as SARS-CoV-2 3CL protease inhibitors. *RSC Med Chem.* 2021;12(10):1722-1730.
121. Lee JY, Kuo CJ, Shin JS, et al. Identification of non-covalent 3C-like protease inhibitors against severe acute respiratory syndrome coronavirus-2 via virtual screening of a Korean compound library. *Bioorg Med Chem Lett.* 2021;42:128067.
122. Simmons G, Gosalia DN, Rennekamp AJ, et al. Inhibitors of cathepsin L prevent severe acute respiratory syndrome coronavirus entry. *Proc Natl Acad Sci U S A.* 2005;102(33):11876-11881.

123. Gomes CP, Fernandes DE, Casimiro F, et al. Cathepsin L in COVID-19: from pharmacological evidences to genetics. *Front Cell Infect Microbiol.* 2020;10:589505.
124. Shang J, Wan Y, Luo C, et al. Cell entry mechanisms of SARS-CoV-2. *Proc Natl Acad Sci U S A.* 2020;117(21):11727-11734.
125. Costanzi E, Kuzikov M, Esposito F, et al. Structural and biochemical analysis of the dual inhibition of MG-132 against SARS-CoV-2 main protease (Mpro/3CLpro) and human cathepsin-L. *Int J Mol Sci.* 2021;22(21):11779.
126. Li L, Chenna BC, Yang KS, et al. Self-masked aldehyde inhibitors: a novel strategy for inhibiting cysteine proteases. *J Med Chem.* 2021;64(15):11267-11287.
127. Al-Wahaibi LH, Mostafa A, Mostafa YA, et al. Discovery of novel oxazole-based macrocycles as anti-coronaviral agents targeting SARS-CoV-2 main protease. *Bioorg Chem.* 2021;116:105363.
128. Johansen-Leete J, Ullrich S, Fry SE, et al. Antiviral cyclic peptides targeting the main protease of SARS-CoV-2. *Chem Sci.* 2022;13(13):3826-3836.
129. Di Sarno V, Lauro G, Musella S, et al. Identification of a dual acting SARS-CoV-2 proteases inhibitor through in silico design and step-by-step biological characterization. *Eur J Med Chem.* 2021;226:113863.
130. Stille JK, Tjuttrins J, Wang G, et al. Design, synthesis and in vitro evaluation of novel SARS-CoV-2 3CL(pro) covalent inhibitors. *Eur J Med Chem.* 2022;229:114046.
131. Breidenbach J, Lemke C, Pillaiyar T, et al. Targeting the main protease of SARS-CoV-2: from the establishment of high throughput screening to the design of tailored inhibitors. *Angew Chem Int Ed Engl.* 2021;60(18):10423-10429.
132. Redhead MA, Owen CD, Brewitz L, et al. Bispecific repurposed medicines targeting the viral and immunological arms of COVID-19. *Sci Rep.* 2021;11(1):13208.
133. Yang KS, Ma XR, Ma Y, et al. A quick route to multiple highly potent SARS-CoV-2 main protease inhibitors. *ChemMedChem.* 2021;16(6):942-948.
134. Ma XR, Alugubelli YR, Ma Y, et al. MPI8 is potent against SARS-CoV-2 by inhibiting dually and selectively the SARS-CoV-2 main protease and the host cathepsin L. *ChemMedChem.* 2022;17(1):e202100456.
135. Yamane D, Onitsuka S, Re S, et al. Selective covalent targeting of SARS-CoV-2 main protease by enantiopure chlorofluoroacetamide. *Chem Sci.* 2022;13(10):3027-3034.
136. Yang J, Lin X, Xing N, et al. Structure-based discovery of novel nonpeptide inhibitors targeting SARS-CoV-2 M(pro). *J Chem Inf Model.* 2021;61(8):3917-3926.
137. Glaser J, Sedova A, Galanie S, et al. Hit expansion of a non-covalent SARS-CoV-2 main protease inhibitor. *ACS Pharmacol Transl Sci.* 2022;5(4):255-265.
138. Chen CC, Yu X, Kuo CJ, et al. Overview of antiviral drug candidates targeting coronaviral 3C-like main proteases. *FEBS J.* 2021;288(17):5089-5121.
139. Turlington M, Chun A, Tomar S, et al. Discovery of N-(benzo[1,2,3]triazol-1-yl)-N-(benzyl)acetamido)phenyl) carboxamides as severe acute respiratory syndrome coronavirus (SARS-CoV) 3CLpro inhibitors: identification of ML300 and noncovalent nanomolar inhibitors with an induced-fit binding. *Bioorg Med Chem Lett.* 2013;23(22):6172-6177.
140. Han SH, Goins CM, Arya T, et al. Structure-based optimization of ML300-derived, noncovalent inhibitors targeting the severe acute respiratory syndrome coronavirus 3CL protease (SARS-CoV-2 3CL(pro)). *J Med Chem.* 2022;65(4):2880-2904.
141. Ghahremanpour MM, Tirado-Rives J, Deshmukh M, et al. Identification of 14 known drugs as inhibitors of the main protease of SARS-CoV-2. *ACS Med Chem Lett.* 2020;11(12):2526-2533.
142. Zhang CH, Stone EA, Deshmukh M, et al. Potent noncovalent inhibitors of the main protease of SARS-CoV-2 from molecular sculpting of the drug peramppanel guided by free energy perturbation calculations. *ACS Cent Sci.* 2021;7(3):467-475.
143. Deshmukh MG, Ippolito JA, Zhang CH, et al. Structure-guided design of a peramppanel-derived pharmacophore targeting the SARS-CoV-2 main protease. *Structure.* 2021;29(8):823-833.e5.
144. Zhang CH, Spasov KA, Reilly RA, et al. Optimization of triarylpyridinone inhibitors of the main protease of SARS-CoV-2 to low-nanomolar antiviral potency. *ACS Med Chem Lett.* 2021;12(8):1325-1332.
145. Lutgens A, Gullberg H, Abdurakhmanov E, et al. Ultralarge virtual screening identifies SARS-CoV-2 main protease inhibitors with broad-spectrum activity against coronaviruses. *J Am Chem Soc.* 2022;144(7):2905-2920.
146. Ramos-Guzman CA, Ruiz-Pernia JJ, Tunon I. Inhibition mechanism of SARS-CoV-2 main protease with ketone-based inhibitors unveiled by multiscale simulations: insights for improved designs. *Angew Chem Int Ed Engl.* 2021;60(49):25933-25941.
147. Hattori SI, Higshi-Kuwata N, Raghavaiah J, et al. GRL-0920, an indole chloropyridinyl ester, completely blocks SARS-CoV-2 infection. *mBio.* 2020;11(4):e01833-20.
148. Ghosh AK, Gong G, Grum-Tokars V, et al. Design, synthesis and antiviral efficacy of a series of potent chloropyridyl ester-derived SARS-CoV 3CLpro inhibitors. *Bioorg Med Chem Lett.* 2008;18(20):5684-5688.
149. Ma C, Hu Y, Townsend JA, et al. Ebselen, disulfiram, carmofur, PX-12, tideglusib, and shikonin are nonspecific promiscuous SARS-CoV-2 main protease inhibitors. *ACS Pharmacol Transl Sci.* 2020;3(6):1265-1277.
150. Qiao Z, Wei N, Jin L, et al. The Mpro structure-based modifications of ebselen derivatives for improved antiviral activity against SARS-CoV-2 virus. *Bioorg Chem.* 2021;117:105455.
151. Sun LY, Chen C, Su J, et al. Ebsulfur and Ebselen as highly potent scaffolds for the development of potential SARS-CoV-2 antivirals. *Bioorg Chem.* 2021;112:104889.
152. Ghosh AK, Raghavaiah J, Shahabi D, et al. Indole chloropyridinyl ester-derived SARS-CoV-2 3CLpro inhibitors: enzyme inhibition, antiviral efficacy, structure-activity relationship, and X-ray structural studies. *J Med Chem.* 2021;64(19):14702-14714.
153. Ghosh AK, Shahabi D, Yadav M, et al. Chloropyridinyl esters of nonsteroidal anti-inflammatory agents and related derivatives as potent SARS-CoV-2 3CL protease inhibitors. *Molecules.* 2021;26(19):5782.
154. Chen W, Feng B, Han S, et al. Discovery of highly potent SARS-CoV-2 M(pro) inhibitors based on benzoisothiazolone scaffold. *Bioorg Med Chem Lett.* 2022;58:128526.
155. Petrou A, Zagaliotis P, Theodoroula NF, et al. Thiazole/thiadiazole/benzothiazole based thiazolidin-4-one

- derivatives as potential inhibitors of main protease of SARS-CoV-2. *Molecules*. 2022;27(7):2180.
156. El Khoury L, Jing Z, Cuzzolin A, et al. Computationally driven discovery of SARS-CoV-2 M(pro) inhibitors: from design to experimental validation. *Chem Sci*. 2022;13(13):3674-3687.
157. Guo S, Xie H, Lei Y, et al. Discovery of novel inhibitors against main protease (Mpro) of SARS-CoV-2 via virtual screening and biochemical evaluation. *Bioorg Chem*. 2021;110:104767.
158. Fricker SP, Mosi RM, Cameron BR, et al. Metal compounds for the treatment of parasitic diseases. *J Inorg Biochem*. 2008;102(10):1839-1845.
159. Fricker SP. Cysteine proteases as targets for metal-based drugs. *Metallomics*. 2010;2(6):366-377.
160. Karges J, Cohen SM. Metal complexes as antiviral agents for SARS-CoV-2. *ChemBioChem*. 2021;22(16):2600-2607.
161. Qin XY, Hou XD, Zhu GH, et al. Discovery and characterization of the naturally occurring inhibitors against human pancreatic lipase in *Ampelopsis grossedentata*. *Front Nutr*. 2022;9:844195.
162. Huo PC, Hu Q, Shu S, et al. Design, synthesis and biological evaluation of novel chalcone-like compounds as potent and reversible pancreatic lipase inhibitors. *Bioorg Med Chem*. 2021;29:115853.
163. Li C-Y, Wang H-N, Zhu G-H, et al. Discovery and characterization of naturally occurring chalcones as potent inhibitors of bile salt hydrolases. *Acta Mater Med*. 2022;1(2).
164. Hou XD, Guan XQ, Cao YF, et al. Inhibition of pancreatic lipase by the constituents in St. John's Wort: in vitro and in silico investigations. *Int J Biol Macromol*. 2020;145:620-633.
165. Song YQ, Guan XQ, Weng ZM, et al. Discovery of a highly specific and efficacious inhibitor of human carboxylesterase 2 by large-scale screening. *Int J Biol Macromol*. 2019;137:261-269.
166. Xiong Y, Zhu GH, Wang HN, et al. Discovery of naturally occurring inhibitors against SARS-CoV-2 3CL(pro) from *Ginkgo biloba* leaves via large-scale screening. *Fitoterapia*. 2021;152:104909.
167. Shen N, Wang T, Gan Q, et al. Plant flavonoids: classification, distribution, biosynthesis, and antioxidant activity. *Food Chem*. 2022;383:132531.
168. Ververidis F, Trantas E, Douglas C, et al. Biotechnology of flavonoids and other phenylpropanoid-derived natural products. Part I: chemical diversity, impacts on plant biology and human health. *Biotechnol J*. 2007;2(10):1214-1234.
169. D'Arcy MS. A review of biologically active flavonoids as inducers of autophagy and apoptosis in neoplastic cells and as cytoprotective agents in non-neoplastic cells. *Cell Biol Int*. 2022.
170. Fraga CG, Croft KD, Kennedy DO, et al. The effects of polyphenols and other bioactives on human health. *Food Funct*. 2019;10(2):514-528.
171. Zaragoza C, Villaescusa L, Monserrat J, et al. Potential therapeutic anti-inflammatory and immunomodulatory effects of dihydroflavones, flavones, and flavonols. *Molecules*. 2020;25(4):1107.
172. Shaito A, Thuan DTB, Phu HT, et al. Herbal medicine for cardiovascular diseases: efficacy, mechanisms, and safety. *Front Pharmacol*. 2020;11:422.
173. Mouffouk C, Mouffouk S, Mouffouk S, et al. Flavonols as potential antiviral drugs targeting SARS-CoV-2 proteases (3CL(pro) and PL(pro)), spike protein, RNA-dependent RNA polymerase (RdRp) and angiotensin-converting enzyme II receptor (ACE2). *Eur J Pharmacol*. 2021;891:173759.
174. Liskova A, Samec M, Koklesova L, et al. Flavonoids against the SARS-CoV-2 induced inflammatory storm. *Biomed Pharmacother*. 2021;138:111430.
175. Song JW, Long JY, Xie L, et al. Applications, phytochemistry, pharmacological effects, pharmacokinetics, toxicity of *Scutellaria baicalensis* Georgi. and its probably potential therapeutic effects on COVID-19: a review. *Chin Med*. 2020;15:102.
176. Li K, Liang Y, Cheng A, et al. Antiviral properties of Baicalin: a concise review. *Rev Bras Farmacogn*. 2021;31(4):408-419.
177. Liu H, Ye F, Sun Q, et al. *Scutellaria baicalensis* extract and baicalein inhibit replication of SARS-CoV-2 and its 3C-like protease in vitro. *J Enzyme Inhib Med Chem*. 2021;36(1):497-503.
178. Bolton JL, Dunlap TL, Dietz BM. Formation and biological targets of botanical o-quinones. *Food Chem Toxicol*. 2018;120:700-707.
179. Wu Q, Yan S, Wang Y, et al. Discovery of 4'-O-methylscutellarein as a potent SARS-CoV-2 main protease inhibitor. *Biochem Biophys Res Commun*. 2022;604:76-82.
180. Su H, Yao S, Zhao W, et al. Identification of pyrogallol as a warhead in design of covalent inhibitors for the SARS-CoV-2 3CL protease. *Nat Commun*. 2021;12(1):3623.
181. Su HX, Yao S, Zhao WF, et al. Anti-SARS-CoV-2 activities in vitro of Shuanghuanglian preparations and bioactive ingredients. *Acta Pharmacol Sin*. 2020;41(9):1167-1177.
182. Chiou WC, Lu HF, Hsu NY, et al. Ugonin J acts as a SARS-CoV-2 3C-like protease inhibitor and exhibits anti-inflammatory properties. *Front Pharmacol*. 2021;12:720018.
183. Park JY, Yuk HJ, Ryu HW, et al. Evaluation of polyphenols from *Broussonetia papyrifera* as coronavirus protease inhibitors. *J Enzyme Inhib Med Chem*. 2017;32(1):504-515.
184. Ryu YB, Jeong HJ, Kim JH, et al. Biflavonoids from *Torreya nucifera* displaying SARS-CoV 3CL(pro) inhibition. *Bioorg Med Chem*. 2010;18(22):7940-7947.
185. Abian O, Ortega-Alarcon D, Jimenez-Alesanco A, et al. Structural stability of SARS-CoV-2 3CLpro and identification of quercetin as an inhibitor by experimental screening. *Int J Biol Macromol*. 2020;164:1693-1703.
186. Saakre M, Mathew D, Ravisankar V. Perspectives on plant flavonoid quercetin-based drugs for novel SARS-CoV-2. *Beni Suef Univ J Basic Appl Sci*. 2021;10(1):21.
187. Mangiavacchi F, Botwina P, Menichetti E, et al. Selenofunctionalization of quercetin improves the non-covalent inhibition of M(pro) and its antiviral activity in cells against SARS-CoV-2. *Int J Mol Sci*. 2021;22(13):7048.
188. Chiou WC, Chen JC, Chen YT, et al. The inhibitory effects of PGG and EGCG against the SARS-CoV-2 3C-like protease. *Biochem Biophys Res Commun*. 2022;591:130-136.
189. Du A, Zheng R, Disoma C, et al. Epigallocatechin-3-gallate, an active ingredient of Traditional Chinese Medicines, inhibits the 3CLpro activity of SARS-CoV-2. *Int J Biol Macromol*. 2021;176:1-12.
190. Nguyen TT, Woo HJ, Kang HK, et al. Flavonoid-mediated inhibition of SARS coronavirus 3C-like protease expressed in *Pichia pastoris*. *Biotechnol Lett*. 2012;34(5):831-838.
191. Liu SY, Wang W, Ke JP, et al. Discovery of *Camellia sinensis* catechins as SARS-CoV-2 3CL protease inhibitors through

- molecular docking, intra and extra cellular assays. *Phytomedicine*. 2022;96:153853.
192. Park J, Park R, Jang M, et al. Therapeutic potential of EGCG, a green tea polyphenol, for treatment of coronavirus diseases. *Life*. 2021;11(3):197.
 193. Cianciosi D, Forbes-Hernandez TY, Regolo L, et al. The reciprocal interaction between polyphenols and other dietary compounds: impact on bioavailability, antioxidant capacity and other physico-chemical and nutritional parameters. *Food Chem*. 2022;375:131904.
 194. Le Roy J, Huss B, Creach A, et al. Glycosylation is a major regulator of phenylpropanoid availability and biological activity in plants. *Front Plant Sci*. 2016;7:735.
 195. Plaza M, Pozzo T, Liu J, et al. Substituent effects on in vitro antioxidantizing properties, stability, and solubility in flavonoids. *J Agric Food Chem*. 2014;62(15):3321-3333.
 196. Sun W, Liang L, Meng X, et al. Biochemical and molecular characterization of a flavonoid 3-o-glycosyltransferase responsible for anthocyanins and flavonols biosynthesis in *Freesia hybrida*. *Front Plant Sci*. 2016;7:410.
 197. Cherrak SA, Merzouk H, Mokhtari-Soulimane N. Potential bioactive glycosylated flavonoids as SARS-CoV-2 main protease inhibitors: a molecular docking and simulation studies. *PLoS One*. 2020;15(10):e0240653.
 198. Jo S, Kim S, Shin DH, et al. Inhibition of SARS-CoV 3CL protease by flavonoids. *J Enzyme Inhib Med Chem*. 2020;35(1):145-151.
 199. da Silva FMA, da Silva KPA, de Oliveira LPM, et al. Flavonoid glycosides and their putative human metabolites as potential inhibitors of the SARS-CoV-2 main protease (Mpro) and RNA-dependent RNA polymerase (RdRp). *Mem Inst Oswaldo Cruz*. 2020;115:e200207.
 200. Godinho PIC, Soengas RG, Silva VLM. Therapeutic potential of glycosyl flavonoids as anti-coronaviral agents. *Pharmaceuticals*. 2021;14(6):546.
 201. Refaat H, Mady FM, Sarhan HA, et al. Optimization and evaluation of propolis liposomes as a promising therapeutic approach for COVID-19. *Int J Pharm*. 2021;592:120028.
 202. Rizzuti B, Grande F, Conforti F, et al. Rutin is a low micromolar inhibitor of SARS-CoV-2 main protease 3CLpro: implications for drug design of quercetin analogs. *Biomedicines*. 2021;9(4):375.
 203. Liao Q, Chen Z, Tao Y, et al. An integrated method for optimized identification of effective natural inhibitors against SARS-CoV-2 3CLpro. *Sci Rep*. 2021;11(1):22796.
 204. Chen L, Li J, Luo C, et al. Binding interaction of quercetin-3-beta-galactoside and its synthetic derivatives with SARS-CoV 3CL(pro): structure-activity relationship studies reveal salient pharmacophore features. *Bioorg Med Chem*. 2006;14(24):8295-8306.
 205. Menezes J, Campos VR. Natural biflavonoids as potential therapeutic agents against microbial diseases. *Sci Total Environ*. 2021;769:145168.
 206. Heleno SA, Martins A, Queiroz MJ, et al. Bioactivity of phenolic acids: metabolites versus parent compounds: a review. *Food Chem*. 2015;173:501-513.
 207. Kumar N, Goel N. Phenolic acids: natural versatile molecules with promising therapeutic applications. *Biotechnol Rep*. 2019;24:e00370.
 208. Chen Z, Cui Q, Cooper L, et al. Ginkgolic acid and anacardic acid are specific covalent inhibitors of SARS-CoV-2 cysteine proteases. *Cell Biosci*. 2021;11(1):45.
 209. Yao Q-Q, Li L, Xu M-C, et al. The metabolism and hepatotoxicity of ginkgolic acid (17:1) in vitro. *Chin J Natural Med*. 2018;16(11):829-837.
 210. Borenstein R, Hanson BA, Markosyan RM, et al. Ginkgolic acid inhibits fusion of enveloped viruses. *Sci Rep*. 2020;10(1):4746.
 211. Lu JM, Yan S, Jamaluddin S, et al. Ginkgolic acid inhibits HIV protease activity and HIV infection in vitro. *Med Sci Monit*. 2012;18(8):BR293-BR298.
 212. Nguyen TTH, Jung JH, Kim MK, et al. The inhibitory effects of plant derivate polyphenols on the main protease of SARS coronavirus 2 and their structure-activity relationship. *Molecules*. 2021;26(7):1924.
 213. Wang S-C, Chen Y, Wang Y-C, et al. Tannic acid suppresses SARS-CoV-2 as a dual inhibitor of the viral main protease and the cellular TMPRSS2 protease. *Am J Cancer Res*. 2020;10(12):4538-4546.
 214. Jang M, Park YI, Cha YE, et al. Tea polyphenols EGCG and theaflavin inhibit the activity of SARS-CoV-2 3CL-protease in vitro. *Evid Based Complement Alternat Med*. 2020;2020:5630838.
 215. Chen CN, Lin CP, Huang KK, et al. Inhibition of SARS-CoV 3C-like protease activity by theaflavin-3,3'-digallate (TF3). *Evid Based Complement Alternat Med*. 2005;2(2):209-215.
 216. Torres-León C, Ventura-Sobrevilla J, Serna-Cock L, et al. Pentagalloylglucose (PGG): a valuable phenolic compound with functional properties. *J Funct Foods*. 2017;37:176-189.
 217. Park JY, Kim JH, Kwon JM, et al. Dieckol, a SARS-CoV 3CL(pro) inhibitor, isolated from the edible brown algae *Ecklonia cava*. *Bioorg Med Chem*. 2013;21(13):3730-3737.
 218. Yan G, Li D, Lin Y, et al. Development of a simple and miniaturized sandwich-like fluorescence polarization assay for rapid screening of SARS-CoV-2 main protease inhibitors. *Cell Biosci*. 2021;11(1):199.
 219. Du R, Cooper L, Chen Z, et al. Discovery of chebulagic acid and punicalagin as novel allosteric inhibitors of SARS-CoV-2 3CL(pro). *Antiviral Res*. 2021;190:105075.
 220. Monks TJ, Jones DC. The metabolism and toxicity of quinones, quinonimines, quinone methides, and quinone-thioethers. *Curr Drug Metab*. 2002;3(4):425-438.
 221. Ali K, Mishra P, Kumar A, et al. Reactivity vs. selectivity of quinone methides: synthesis of pharmaceutically important molecules, toxicity and biological applications. *Chem Commun*. 2022;58(42):6160-6175.
 222. Park JY, Kim JH, Kim YM, et al. Tanshinones as selective and slow-binding inhibitors for SARS-CoV cysteine proteases. *Bioorg Med Chem*. 2012;20(19):5928-5935.
 223. Zhao Y, Du X, Duan Y, et al. High-throughput screening identifies established drugs as SARS-CoV-2 PLpro inhibitors. *Protein Cell*. 2021;12(11):877-888.
 224. Ma C, Wang J. Validation and invalidation of SARS-CoV-2 papain-like protease inhibitors. *ACS Pharmacol Transl Sci*. 2022;5(2):102-109.
 225. Lim CT, Tan KW, Wu M, et al. Identifying SARS-CoV-2 antiviral compounds by screening for small molecule inhibitors of Nsp3 papain-like protease. *Biochem J*. 2021;478(13):2517-2531.
 226. Elgawish MS, Kishikawa N, Helal MA, et al. Molecular modeling and spectroscopic study of quinone-protein adducts:

- insight into toxicity, selectivity, and reversibility. *Toxicol Res.* 2015;4(4):843-847.
227. Su C, Liu Z, Wang Y, et al. The electrophilic character of quinones is essential for the suppression of Bach1. *Toxicology.* 2017;387:17-26.
 228. O'Brien PJ. Molecular mechanisms of quinone cytotoxicity. *Chem Biol Interact.* 1991;1(80):1-41.
 229. Wang R, Hu Q, Wang H, et al. Identification of vitamin K3 and its analogues as covalent inhibitors of SARS-CoV-2 3CL(pro). *Int J Biol Macromol.* 2021;183:182-192.
 230. Borges RS, Carneiro AS, Barros TG, et al. Understanding the cytotoxicity or cytoprotective effects of biological and synthetic quinone derivatives by redox mechanism. *J Mol Model.* 2014;20(12):2541.
 231. Cui J, Jia J. Discovery of juglone and its derivatives as potent SARS-CoV-2 main proteinase inhibitors. *Eur J Med Chem.* 2021;225:113789.
 232. Santos LH, Kronenberger T, Almeida RG, et al. Structure-based identification of naphthoquinones and derivatives as novel inhibitors of main protease Mpro and papain-like protease PLpro of SARS-CoV-2. *bioRxiv.* 2022.
 233. Hicks EG, Kandel SE, Lampe JN. Identification of Aloe-derived natural products as prospective lead scaffolds for SARS-CoV-2 main protease (M(pro)) inhibitors. *Bioorg Med Chem Lett.* 2022;66:128732.
 234. Gyebi GA, Ogunro OB, Adegunloye AP, et al. Potential inhibitors of coronavirus 3-chymotrypsin-like protease (3CL(pro)): an in silico screening of alkaloids and terpenoids from African medicinal plants. *J Biomol Struct Dyn.* 2021;39(9):3396-3408.
 235. Ryu YB, Park SJ, Kim YM, et al. SARS-CoV 3CLpro inhibitory effects of quinone-methide triterpenes from *Tripterygium regelii*. *Bioorg Med Chem Lett.* 2010;20(6):1873-1876.
 236. Chen LR, Wang YC, Lin YW, et al. Synthesis and evaluation of isatin derivatives as effective SARS coronavirus 3CL protease inhibitors. *Bioorg Med Chem Lett.* 2005;15(12):3058-3062.
 237. Liu P, Liu H, Sun Q, et al. Potent inhibitors of SARS-CoV-2 3C-like protease derived from N-substituted isatin compounds. *Eur J Med Chem.* 2020;206:112702.
 238. Zhong B, Peng W, Du S, et al. Oridonin inhibits SARS-CoV-2 by targeting its 3C-like protease. *Small Sci.* 2022:2100124.
 239. Hasegawa T, Imamura RM, Suzuki T, et al. Application of acoustic ejection MS system to high-throughput screening for SARS-CoV-2 3CL protease inhibitors. *Chem Pharm Bull.* 2022;70(3):199-201.
 240. Zhou YF, Yan BC, Yang Q, et al. Harnessing natural products by a pharmacophore-oriented semisynthesis approach for the discovery of potential anti-SARS-CoV-2 agents. *Angew Chem Int Ed Engl.* 2022:e202201684.
 241. Xu H, Li J, Song S, et al. Effective inhibition of coronavirus replication by *Polygonum cuspidatum*. *Front Biosci.* 2021;26(10):789-798.
 242. Morita T, Miyakawa K, Jeremiah SS, et al. All-trans retinoic acid exhibits antiviral effect against SARS-CoV-2 by inhibiting 3CLpro activity. *Viruses.* 2021;13(8):1669.
 243. Zhang JW, Xiong Y, Wang F, et al. Discovery of 9,10-dihydrophenanthrene derivatives as SARS-CoV-2 3CL(pro) inhibitors for treating COVID-19. *Eur J Med Chem.* 2022;228:114030.
 244. Khan NA, Al-Thani H, El-Menyar A. The emergence of new SARS-CoV-2 variant (Omicron) and increasing calls for COVID-19 vaccine boosters—the debate continues. *Travel Med Infect Dis.* 2022;45:102246.
 245. Harrison C. COVID-19 antiviral pills raise hopes for curbing pandemic. *Nat Biotechnol.* 2021.
 246. EDP-235-ASBMB-Annual-Meeting-Seminar_Final.pdf.
 247. Li J, Lin C, Zhou X, et al. Structural basis of the main proteases of coronavirus bound to drug candidate PF-07321332. *J Virol.* 2022;96(8):e0201321.
 248. Owen DR, Allerton CMN, Anderson AS, et al. An oral SARS-CoV-2 M(pro) inhibitor clinical candidate for the treatment of COVID-19. *Science.* 2021;374(6575):1586-1593.
 249. Sasaki M, Tabata K, Kishimoto M, et al. Oral administration of S-217622, a SARS-CoV-2 main protease inhibitor, decreases 1 viral load and accelerates recovery from clinical aspects of COVID-19. 2022.
 250. Tyndall JDA. S-217622, a 3CL protease inhibitor and clinical candidate for SARS-CoV-2. *J Med Chem.* 2022;65(9):6496-6498.
 251. Vandyck K, Abdelnabi R, Gupta K, et al. ALG-097111, a potent and selective SARS-CoV-2 3-chymotrypsin-like cysteine protease inhibitor exhibits in vivo efficacy in a Syrian Hamster model. *Biochem Biophys Res Commun.* 2021;555:134-139.
 252. Unoh Y, Uehara S, Nakahara K, et al. Discovery of S-217622, a noncovalent oral SARS-CoV-2 3CL protease inhibitor clinical candidate for treating COVID-19. *J Med Chem.* 2022;65(9):6499-6512.
 253. Boras B, Jones RM, Anson BJ, et al. Preclinical characterization of an intravenous coronavirus 3CL protease inhibitor for the potential treatment of COVID19. *Nat Commun.* 2021;12(1):6055.
 254. Hoffman RL, Kania RS, Brothers MA, et al. Discovery of ketone-based covalent inhibitors of coronavirus 3CL proteases for the potential therapeutic treatment of COVID-19. *J Med Chem.* 2020;63(21):12725-12747.
 255. Shcherbakov D, Baev D, Kalinin M, et al. Design and evaluation of bispidine-based SARS-CoV-2 main protease inhibitors. *ACS Med Chem Lett.* 2022;13(1):140-147.
 256. Eng H, Dantonio AL, Kadar EP, et al. Disposition of nirmatrelvir, an orally bioavailable inhibitor of SARS-CoV-2 3C-like protease, across animals and humans. *Drug Metab Dispos.* 2022;50(5):576-590.
 257. Li H, Yang L, Liu FF, et al. Overview of therapeutic drug research for COVID-19 in China. *Acta Pharmacol Sin.* 2020;41(9):1133-1140.
 258. Abdelnabi R, Foo CS, Jochmans D, et al. The oral protease inhibitor (PF-07321332) protects Syrian hamsters against infection with SARS-CoV-2 variants of concern. *Nat Commun.* 2022;13(1):719.
 259. Wong CKH, Au ICH, Lau KTK, et al. Real-world effectiveness of molnupiravir and nirmatrelvir/ritonavir among COVID-19 inpatients during Hong Kong's Omicron BA.2 wave: an observational study. *medRxiv.* 2022. 2022.05.19.22275291.
 260. s-217622. Available from: <https://www.shionogi.com/us/en/news/2022/04/new-data-for-shionogis-covid-19-once-dailyoral-antiviral-s-217622-show-rapid-virus-clearance.html>
 261. PBI-0451-ZW. Available from: <https://ir.pardesbio.com/news-releases/news-release-details/pardes-biosciences-presents-interim-clinical-data-ongoing-pbi>

262. Mengist HM, Fan X, Jin T. Designing of improved drugs for COVID-19: crystal structure of SARS-CoV-2 main protease M(pro). *Signal Transduct Target Ther*. 2020;5(1):67.
263. ASC11. Available from: <https://www.prnewswire.com/news-releases/ascl1-announces-3clpro-inhibitor-ascl1-demonstrated-potential-to-be-effective-treatment-for-covid-19-301527722.html>
264. EDDC-2214. Available from: <https://www.a-star.edu.sg/News/a-star-news/news/press-releases/everest-medicines-to-develop-commercialise-eddc-s-small-molecules>
265. Tripathi PK, Upadhyay S, Singh M, et al. Screening and evaluation of approved drugs as inhibitors of main protease of SARS-CoV-2. *Int J Biol Macromol*. 2020;164:2622-2631.
266. Li Z, Li X, Huang YY, et al. Identify potent SARS-CoV-2 main protease inhibitors via accelerated free energy perturbation-based virtual screening of existing drugs. *Proc Natl Acad Sci U S A*. 2020;117(44):27381-27387.
267. Chiou WC, Hsu MS, Chen YT, et al. Repurposing existing drugs: identification of SARS-CoV-2 3C-like protease inhibitors. *J Enzyme Inhib Med Chem*. 2021;36(1):147-153.
268. Drayman N, DeMarco JK, Jones KA, et al. Masitinib is a broad coronavirus 3CL inhibitor that blocks replication of SARS-CoV-2. *Science*. 2021;373(6557):931-936.
269. Yuan S, Wang R, Chan JF, et al. Metallo drug ranitidine bismuth citrate suppresses SARS-CoV-2 replication and relieves virus-associated pneumonia in Syrian hamsters. *Nat Microbiol*. 2020;5(11):1439-1448.
270. Chen J, Zhang Y, Zeng D, et al. Merbromin is a mixed-type inhibitor of 3-chymotrypsin like protease of SARS-CoV-2. *Biochem Biophys Res Commun*. 2022;591:118-123.
271. Kuo CJ, Chao TL, Kao HC, et al. Kinetic characterization and inhibitor screening for the proteases leading to identification of drugs against SARS-CoV-2. *Antimicrob Agents Chemother*. 2021;65(4):e02577-20.
272. Rawson JMO, Duchon A, Nikolaitchik OA, et al. Development of a cell-based luciferase complementation assay for identification of SARS-CoV-2 3CL(pro) inhibitors. *Viruses*. 2021;13(2):173.
273. Chen X, Liu M, Zhang P, et al. Phage-derived depolymerase as an antibiotic adjuvant against multidrug-resistant *Acinetobacter baumannii*. *Front Microbiol*. 2022;13:845500.
274. Yu L, Kim HJ, Park MK, et al. Ethacrynic acid, a loop diuretic, suppresses epithelial-mesenchymal transition of A549 lung cancer cells via blocking of NDP-induced WNT signaling. *Biochem Pharmacol*. 2021;183:114339.
275. Angiolillo DJ, Weisman SM. Clinical pharmacology and cardiovascular safety of naproxen. *Am J Cardiovasc Drugs*. 2017;17(2):97-107.
276. Si K, Wei C, Xu L, et al. Hyperuricemia and the risk of heart failure: pathophysiology and therapeutic implications. *Front Endocrinol*. 2021;12:770815.
277. Porras AMG, Terra BS, Braga TC, et al. Butenafine and analogues: an expeditious synthesis and cytotoxicity and antifungal activities. *J Adv Res*. 2018;14:81-91.
278. Ha J, Kim J, Jeong C, et al. Effect of follow-up raloxifene therapy after denosumab discontinuation in postmenopausal women. *Osteoporos Int*. 2022;33(7):1591-1599.
279. McGrath PJ, Stewart JW, Fava M, et al. Tranylcypromine versus venlafaxine plus mirtazapine following three failed antidepressant medication trials for depression: a STAR*D report. *Am J Psychiatry*. 2006;163(9):1531-1541. quiz 666.
280. Mahdi M, Motyan JA, Szojka ZI, et al. Analysis of the efficacy of HIV protease inhibitors against SARS-CoV-2's main protease. *Virology*. 2020;17(1):190.
281. Ren T, Zhang Z, Fawcett JP, et al. Micro-solid phase extraction and LC-MS(3) for the determination of triptorelin in rat plasma and application to a pharmacokinetic study. *J Pharm Biomed Anal*. 2019;166:13-19.
282. Zhao Y, Sun H, Zheng J, et al. Identification of predictors based on drug targets highlights accurate treatment of goserelin in breast and prostate cancer. *Cell Biosci*. 2021;11(1):5.
283. Couto M, Vide S, Marco-Arino N, et al. Comparison of two pharmacokinetic-pharmacodynamic models of rocuronium bromide during profound neuromuscular block: analysis of estimated and measured post-tetanic count effect. *Br J Anaesth*. 2022;128(3):473-481.
284. Kongdang P, Pruksakorn D, Koonrunsesomboon N. Preclinical experimental models for assessing laxative activities of substances-products under investigation-a scoping review of the literature. *Am J Transl Res*. 2022;14(2):698-717.
285. Dodet P, Sanapo F, Leu-Semenescu S, et al. Sleep disorders in adults with Prader-Willi syndrome: review of the literature and clinical recommendations based on the experience of the French reference centre. *J Clin Med*. 2022;11(7):1986.
286. Nair AB, Kumar S, Dalal P, et al. Novel dermal delivery cargos of clobetasol propionate: an update. *Pharmaceutics*. 2022;14(2):383.
287. Sehgal SN. Rapamune (Sirolimus, rapamycin): an overview and mechanism of action. *Ther Drug Monit*. 1995;17(6):660-665.
288. Liu A, Lin W, Ping S, et al. Analysis of degradation and pathways of three common antihistamine drugs by NaClO, UV, and UV-NaClO methods. *Environ Sci Pollut Res Int*. 2022.
289. Elsayad K, Rolf D, Sunderkotter C, et al. Low-dose total skin electron beam therapy plus oral bexarotene maintenance therapy for cutaneous T-cell lymphoma. *J Dtsch Dermatol Ges*. 2022;20(3):279-285.
290. Mujtaba A, Kohli K. In vitro/in vivo evaluation of HPMC/alginate based extended-release matrix tablets of cefpodoxime proxetil. *Int J Biol Macromol*. 2016;89:434-441.
291. Bharathi C, Jayaram P, Sunder Raj J, et al. Identification, isolation and characterization of impurities of clindamycin palmitate hydrochloride. *J Pharm Biomed Anal*. 2008;48(4):1211-1218.
292. Julio C, Benoist S, Allard MA, et al. Treatment strategies to resectable metachronous colorectal liver metastases after adjuvant oxaliplatin-based chemotherapy for primary colorectal cancer. *J Surg Oncol*. 2022.
293. Vermersch P, Brieva-Ruiz L, Fox RJ, et al. Efficacy and safety of masitinib in progressive forms of multiple sclerosis: a randomized, phase 3, clinical trial. *Neurol Neuroimmunol Neuroinflamm*. 2022;9(3).
294. Wang C, Li X, Cheng T, et al. Eradication of porphyromonas gingivalis persists through colloidal bismuth subcitrate synergistically combined with metronidazole. *Front Microbiol*. 2021;12:748121.
295. Chander J, Maini S, Subrahmanyam S, et al. Otomycosis—a clinico-mycological study and efficacy of mercurochrome in its treatment. *Mycopathologia*. 1996;135(1):9-12.

296. Lovestone S, Boada M, Dubois B, et al. A phase II trial of tideglusib in Alzheimer's disease. *J Alzheimers Dis.* 2015;45(1):75-88.
297. Comlekci E, Kutlu HM, Vejselova Sezer C. Toward stimulating apoptosis in human lung adenocarcinoma cells by novel nano-carmofur compound treatment. *Anticancer Drugs.* 2021;32(6):657-663.
298. Wang L, Bharti Kumar R, et al. Small molecule therapeutics for tauopathy in Alzheimer's disease: walking on the path of most resistance. *Eur J Med Chem.* 2021;209:112915.
299. Das D, Pandya M. Recent advancement of direct-acting antiviral agents (DAAs) in hepatitis C therapy. *Mini Rev Med Chem.* 2018;18(7):584-596.
300. Islam MT, Sarkar C, El-Kersh DM, et al. Natural products and their derivatives against coronavirus: a review of the non-clinical and pre-clinical data. *Phytother Res.* 2020;34(10):2471-2492.
301. Zhang F, Huang J, Liu W, et al. Inhibition of drug-metabolizing enzymes by Qingfei Paidu decoction: implication of herb-drug interactions in COVID-19 pharmacotherapy. *Food Chem Toxicol.* 2021;149:111998.
302. He X, Hong W, Pan X, et al. SARS-CoV-2 Omicron variant: characteristics and prevention. *MedComm (2020).* 2021;2(4):838-845.
303. Silva RC, Freitas HF, Campos JM, et al. Natural products-based drug design against SARS-CoV-2 Mpro 3CLpro. *Int J Mol Sci.* 2021;22(21):11739.
304. Cantrelle FX, Boll E, Brier L, et al. NMR spectroscopy of the main protease of SARS-CoV-2 and fragment-based screening identify three protein hotspots and an antiviral fragment. *Angew Chem Int Ed Engl.* 2021;60(48):25428-25435.
305. Hariono M, Hariyono P, Dwiastuti R, et al. Potential SARS-CoV-2 3CLpro inhibitors from chromene, flavonoid and hydroxamic acid compound based on FRET assay, docking and pharmacophore studies. *Results Chem.* 2021;3:100195.
306. DI Pierro F, Khan A, Bertuccioli A, et al. Quercetin Phytosome(R) as a potential candidate for managing COVID-19. *Minerva Gastroenterol.* 2021;67(2):190-195.
307. Thabault L, Liberelle M, Frederick R. Targeting protein self-association in drug design. *Drug Discov Today.* 2021;26(5):1148-1163.
308. Goyal B, Goyal D. Targeting the dimerization of the main protease of coronaviruses: a potential broad-spectrum therapeutic strategy. *ACS Comb Sci.* 2020;22(6):297-305.
309. Xian Y, Zhang J, Bian Z, et al. Bioactive natural compounds against human coronaviruses: a review and perspective. *Acta Pharm Sin B.* 2020;10(7):1163-1174.
310. Huang F, Li Y, Leung EL, et al. A review of therapeutic agents and Chinese herbal medicines against SARS-COV-2 (COVID-19). *Pharmacol Res.* 2020;158:104929.
311. Bahun M, Jukic M, Oblak D, et al. Inhibition of the SARS-CoV-2 3CL(pro) main protease by plant polyphenols. *Food Chem.* 2022;373(Pt B):131594.
312. Kaul R, Paul P, Kumar S, et al. Promising antiviral activities of natural flavonoids against SARS-CoV-2 targets: systematic review. *Int J Mol Sci.* 2021;22(20):11069.
313. Chen X, Wu Y, Chen C, et al. Identifying potential anti-COVID-19 pharmacological components of traditional Chinese medicine Lianhuaqingwen capsule based on human exposure and ACE2 biochromatography screening. *Acta Pharm Sin B.* 2021;11(1):222-236.
314. Leung EL, Pan HD, Huang YF, et al. The scientific foundation of Chinese herbal medicine against COVID-19. *Engineering.* 2020;6(10):1099-1107.
315. Ashhurst AS, Tang AH, Fajtova P, et al. Potent anti-SARS-CoV-2 activity by the natural product gallinamide A and analogues via inhibition of cathepsin L. *J Med Chem.* 2022;65(4):2956-2970.
316. Sacco MD, Ma C, Lagarias P, et al. Structure and inhibition of the SARS-CoV-2 main protease reveal strategy for developing dual inhibitors against M(pro) and cathepsin L. *Sci Adv.* 2020;6(50):eabe0751.
317. Zaim S, Chong JH, Sankaranarayanan V, et al. COVID-19 and multiorgan response. *Curr Probl Cardiol.* 2020;45(8):100618.
318. Abou-Ismaïl MY, Diamond A, Kapoor S, et al. The hypercoagulable state in COVID-19: incidence, pathophysiology, and management. *Thromb Res.* 2020;194:101-115.
319. Zhao Z, Li Y, Zhou L, et al. Prevention and treatment of COVID-19 using Traditional Chinese Medicine: a review. *Phytomedicine.* 2021;85:153308.
320. Lau KM, Lee KM, Koon CM, et al. Immunomodulatory and anti-SARS activities of *Houttuynia cordata*. *J Ethnopharmacol.* 2008;118(1):79-85.
321. Chen J, Wang YK, Gao Y, et al. Protection against COVID-19 injury by qingfei paidu decoction via anti-viral, anti-inflammatory activity and metabolic programming. *Biomed Pharmacother.* 2020;129:110281.
322. Wu Y, Xu L, Cao G, et al. Effect and mechanism of qingfei paidu decoction in the management of pulmonary fibrosis and COVID-19. *Am J Chin Med.* 2022;50(1):33-51.
323. Li Y, Li B, Wang P, et al. Traditional chinese medicine, qingfei paidu decoction and xuanfei baidu decoction, inhibited cytokine production via NF-kappaB signaling pathway in macrophages: implications for coronavirus disease 2019 (COVID-19) therapy. *Front Pharmacol.* 2021;12:722126.
324. Lee DYW, Li QY, Liu J, et al. Traditional Chinese herbal medicine at the forefront battle against COVID-19: clinical experience and scientific basis. *Phytomedicine.* 2021;80:153337.
325. Paraiso IL, Revel JS, Stevens JF. Potential use of polyphenols in the battle against COVID-19. *Curr Opin Food Sci.* 2020;32:149-155.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Hu Q, Xiong Y, Zhu G-H, et al. The SARS-CoV-2 main protease (M^{PRO}): Structure, function, and emerging therapies for COVID-19. *MedComm.* 2022;3:e151.
<https://doi.org/10.1002/mco2.151>