

Livestock as a potential biological control agent for an invasive wetland plant

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ABSTRACT

Invasive species threaten biodiversity and incur costs exceeding billions of US\$. Eradication efforts, however, are nearly always unsuccessful. Throughout much of North America, land managers have used expensive, and ultimately ineffective, techniques to combat invasive *Phragmites australis* in marshes. Here, we reveal that *Phragmites* may potentially be controlled by employing an affordable measure from its native European range: livestock grazing. Experimental field tests demonstrate that rotational goat grazing (where goats have no choice but to graze *Phragmites*) can reduce *Phragmites* cover from 100 to 20% and that cows and horses also readily consume this plant. These results, combined with the fact that Europeans have suppressed *Phragmites* through seasonal livestock grazing for 6,000 years, suggest *Phragmites* management can shift to include more economical and effective top-down control strategies. More generally, these findings support an emerging paradigm shift in conservation from high-cost eradication to economically sustainable control of dominant invasive species.

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INTRODUCTION

Invasive species globally threaten biodiversity and, in the United States alone, incur costs to human economies estimated to exceed 120 billion US\$ each year (*Pimentel, Zuniga & Morrison, 2005*). Controlling the spread and reducing the impacts of invasive species are therefore foundational objectives of conservation science and policy (*Kareiva & Marvier, 2010*). Historically, eradication of invasive species has been an ideal goal of management programs, but this has rarely been achieved on ecologically relevant spatial

or temporal scales (*Kettenring & Adams, 2011*). In the majority of cases, complete and permanent removal of these species is simply unrealistic (*Sax, Stachowicz & Gaines, 2005*). Consequently, the objectives of invasive species' management are being recast to prioritize control and mitigation, rather than elimination, of invasive species' impacts. In addition, conservation groups are becoming increasingly focused on finding solutions that not only achieve their goals but also bolster local economies (*Kareiva & Marvier, 2010*). Win-win synergisms of this type, however, are rare. To ensure long-term efficacy of control-oriented programs, management strategies should be tailored to both local habitat requirements (i.e., duration, frequency and intensity of control measures) and community needs.

Invasive plants that form expansive monocultures are often key targets for management due to the direct, and usually negative, impact they have on ecosystem structure, function, and services (e.g., cordgrass: *Neira et al., 2006*, crested wheatgrass: *Christian & Wilson, 1999*, reed canary grass: *Lavergne & Molofsky, 2004*, Japanese stiltgrass: *Flory & Clay, 2010*). In the United States, control of invasive plants has been attempted through herbicide application, mechanical removal (e.g., mowing, burning, excavation), or biological control programs that are often costly to implement, difficult to sustain over sufficient timescales, or may result in unintended, harmful consequences (e.g., spillover of herbicides, non-target impacts of arthropod control agents, landscape damage; see *Kettenring & Adams, 2011* for review). In Europe, however, farmers have been culling (whether intentionally or not) similarly 'invasive' plants long before such modern control techniques by deploying livestock to feed on dense vegetation. Grazing by large-bodied domestic herbivores, such as cows, horses, sheep, and goats, cannot only be effective in suppressing dominant plants (*Esselink et al., 2000*), but can also result in reciprocal positive effects for humans by generating valuable goods, including meat, milk, leather, and wool to support local economies. In the United States, use of livestock to control invasive species has been largely restricted to terrestrial grasslands where this method has met with mixed success (e.g., *DiTomaso, 2000*; *Reiner & Craig, 2011*, but see *Marty, 2005*; *Tesauro & Ehrenfeld, 2007* for wetlands). Low species richness and discrete plant zonation common in wetland ecosystems may allow for greater success and targeted control of invasive plants by livestock. New evidence from North American and European marshes, which we document below, coupled with prior research on long-term grazing impacts on plant distribution in Europe, suggests that livestock can be a cost-effective tool for managing the impacts and spread of monoculture-forming invasive plants in wetlands, where monoculture-forming invasive plant species are common and drive large-scale ecosystem change (*Zedler & Kercher, 2004*).

Under natural field settings, there is broad support for the ability of herbivores to suppress invasive plant success. Specifically, herbivores can reduce invasion success by limiting both invasive plant establishment and performance (*Levine, Adler & Yelenik, 2004*), with generalist native herbivores strongly suppressing invasive plants (*Parker, Burkepile & Hayt, 2006*; *Morrison & Hay, 2011*). Invasive herbivores, on the other hand, have opposite effects and can facilitate invasions by reducing the abundance of native species (*Parker, Burkepile & Hayt, 2006*). This suggests that co-evolution/exposure of both

herbivore and autotroph are important considerations when choosing an appropriate biocontrol agent. A novel management approach to maximize potential for success would attempt to control an invasive plant with an introduced herbivore (e.g., domestic livestock) that has a demonstrated effect in controlling the plants in their native range.

In Eastern North American marshes, the common reed, *Phragmites australis* (Cav.) Trin. ex Steud. has invaded with unrelenting success since its cryptic introduction during the 18th century from Europe (Chambers, Meyerson & Saltonstall, 1999; Saltonstall, 2002). *Phragmites*, which reaches average heights of >3 m and generates dense layers of lignified litter, outcompetes native plants for light and often forms expansive, towering monocultures (Bertness, Ewanchuk & Silliman, 2002; Silliman & Bertness, 2004; Meyerson, Saltonstall & Chambers, 2009). Introduced *Phragmites* is particularly successful in marshes along developed shorelines (Bertness, Ewanchuk & Silliman, 2002; Silliman & Bertness, 2004; King et al., 2007), and its rate of invasion is likely to increase in the future with predicted increases in anthropogenic N pollution and rising CO₂ concentrations (Bertness, Ewanchuk & Silliman, 2002; Mozdzer & Zieman, 2010; Mozdzer & Megonigal, 2012; Mozdzer, Brisson & Hazelton, 2013). Where *Phragmites* has become established, native plant diversity declines precipitously (Silliman & Bertness, 2004), ecosystem processes such as nitrogen cycling, methane emissions, and accretion change (e.g., Rooth, Stevenson & Cornwell, 2003; Windham & Ehrenfeld, 2003; Mozdzer & Megonigal, 2013), and once-expansive marsh vistas become obfuscated by this impenetrable grass.

The control of *Phragmites* has dominated marsh conservation efforts in the Northeastern US for the past 30 years (Hazelton et al., 2014). During this time, no cost-effective, long-term control measures have been found. For example, land managers and private organizations have treated over 80,000 hectares of marsh with herbicide over the past five years with limited success, despite costs that exceed \$4.6 million per year (Martin & Blossey, 2013). Similarly, mechanical removal techniques, such as mowing and burning, have proven to be uneconomical, given their high labor costs, and ineffective (Lee, 1990; Cowie et al., 1992). While insect biocontrol has been investigated (Tewksbury et al., 2002; Van Driesche et al., 2002), and specific biocontrol agents tested in laboratory conditions (e.g., Lambert, Winiarski & Casagrande, 2007), it is currently not an option available to land managers, in part because some prospective control agents do greater damage to native strains of *Phragmites* than the invasive (e.g., Lambert, Winiarski & Casagrande, 2007).

As a facultative halophyte, *Phragmites* distribution is largely restricted by salinity in Europe and North America. *Phragmites* salinity tolerance may exceed normal seawater (~33ppt) (Chambers et al., 2003), but its competitiveness increases with decreasing salinity in the high marsh elevations associated with freshwater seepage or in naturally brackish wetlands (Minchinton & Bertness, 2003). However, within brackish marshes in Europe, *Phragmites* abundance is markedly limited. For instance, in the 400-km² salt marshes of the Wadden Sea, *Phragmites* accounts for only 2.5% of vegetation coverage (Esselink et al., 2009), a significant decrease from its historical extent. Although nutrient pollution has been attributed to *Phragmites* die back in Europe (Van Der Putten, 1997), two primary reasons likely account for *Phragmites*' reduced distribution in European brackish marshes: (1)

seawall construction and land reclamation during the Middle Ages (c. A.D. 1000–1300), which caused major loss of brackish marshes with *Phragmites* and (2) an extended history of livestock grazing in these marshes. For example, along the brackish coast of the microtidal Baltic, reed beds dominated by *Phragmites* were transformed into salt meadows from c. 4000 B.C. onwards, when human exploitation started (Vestergaard, 1998). Likewise, in the Netherlands, marshes have been used as rangelands since Late Neolithic, i.e., 3500 B.C. In these heavily grazed European marshes, *Phragmites* is rare, but in areas where livestock grazing has been abandoned in recent times, *Phragmites* has become dominant again (Dijkema, 1990; Jutila, 1999; Esselink, Fresco & Dijkema, 2002; Esselink et al., 2009). A recent study confirmed these observational results: specifically, in marshes still grazed by livestock (Sammul, Kauer & Koster, 2012), *Phragmites* has increased in relative abundance inside grazer exclusion cages but not in control, grazed areas (Milotic et al., 2010; Esselink, 2008, unpublished data). It is unclear whether similar top-down control methods would be effective in mitigating the impacts of introduced *Phragmites* in North America, and, if so, which large grazers and deployment strategy would result in an ecologically effective and economically sustainable solution for both land managers and farmers.

Based on observational and experimental evidence revealing that top-down forces limit *Phragmites* in its native range in Europe, we explored the potential for livestock control of invasive *Phragmites* in North America, and, reciprocally, the nutritional benefit of *Phragmites* to livestock consuming it. Our specific objectives were: (1) to evaluate the ability of caged livestock to control invasive *Phragmites* and increase plant biodiversity in a small-scale experimental setting, (2) to test if various commercially reared livestock breeds will readily consume *Phragmites*, and (3) to investigate the nutritional value of *Phragmites* as livestock forage.

We tested the potential for livestock to control introduced *Phragmites* with a goat inclusion field experiment in a *Phragmites*-invaded North American marsh. As top-down control of invasive plants by free-ranging livestock can be hindered by alternative grazing options that could be superior in quality (Belovsky, 1986; Vulink & Drost, 1991a; Vulink, 2001), we chose to conduct a pressed, grazing experiment in a marsh already dominated by *Phragmites*. To evaluate the potential of additional top-down control agents besides goats and the nutritional benefits of *Phragmites* to livestock, we conducted no-choice feeding trials with cows and horses and, using data from past, unpublished studies, assessed whether livestock can digest *Phragmites* effectively and if *Phragmites* nutritional quality varies significantly over a growing season.

METHODS

To experimentally test the hypothesis that livestock can suppress *Phragmites* monocultures in North American marshes and promote the recovery of native plants, we established randomly located replicated ($n = 4$) goat enclosures (8.5 m × 40 m) made out of wire fencing, a single-strand of electrical wire, and metal stakes in a *Phragmites*-dominated freshwater marsh in the Beltsville Agricultural Research Center in Beltsville, Maryland, USA. Two domestic goats (IACUC number 103453) were stocked in each of the enclosures

(a stocking rate of 58.8 goats/ha), which were paired with ungrazed control plots (also 8.5 m × 40 m) for three treatment periods of 2–4 weeks, beginning mid September 2008, late May 2009, and late August 2009. Two goats per enclosure were used because, first, goats are social animals and solitary confinement might alter their well-being and grazing behavior, and second, because >2 goats would result in too rapid consumption of available plants for grazing (W Hare, pers. comm., 2012, Veterinarian, USDA Beltsville Agricultural Research Center). Goats were left in enclosures until *Phragmites* was completely consumed within at least one of the four enclosures to maximize the duration of grazing and preventing starvation of goats. *Phragmites* was allowed to re-sprout and grow to a height of about 1.5 m before applying the next round of grazing. This approach was implemented to allow potential colonization of native plants and maximize depletion of belowground resources of *Phragmites* (i.e., a level of grazing pressure in excess of that typical of standard rotational grazing practices was intended). *Phragmites* stem density and height of the five tallest stems were measured before and after each grazing period in four 1-m² quadrats spaced systematically at 4, 8, 12, and 16 m along the center line of each grazed and control plot (total = 32 quadrats). Percent cover and plant species richness and diversity were determined in a 100-m² (5 m × 20 m) “module” centered within each grazed and control plot (Peet, Wentworth & White, 1998; Gurevich, Scheiner & Fox, 2006). Plants that were seedlings or lacking flowering or fruiting material, but that could be distinguished as separate species, were identified as “morphospecies” and used in calculations of richness and diversity. Nativity (native or introduced) was determined using the USDA PLANTS database for plants identified to species level, or for taxa where all species were native or introduced. Comparisons between grazed and ungrazed plots were made using mixed model repeated measures ANOVA ($n = 4$) using the MIXED procedure of SAS 9.2 (SAS Institute, Cary, North Carolina), with each pair of grazed and ungrazed plots treated as a block, after checking assumptions of homogeneity of variances and normality. Stem density and stem height from the four 1-m² quadrats within each plot were averaged to generate a single value for each grazed and control plot prior to analysis. Simple effect slices were used to test for significant grazing effects for each sampling date, and the Kenward–Roger method used to calculate denominator degrees of freedom (which can result in fractional ddf).

To explore whether *Phragmites* could be restricted by other domestic livestock species in addition to goats, we conducted no-choice feeding trials in August of 2011 with freshly cut 30-cm sections of *Phragmites* stems and leaves from established reed stands (>2 m in diameter). To do so, we offered a 30-cm section of *Phragmites* stem to 20 individual horses and cows and counted the number of individuals who fully consumed the section after 30 s.

To explore the nutritional quality of *Phragmites* relative to five other common marsh plants, we present data here from a past, unpublished study that asked this question and measured temporal variation in leaf quality of six marsh plants over a growing season. Specifically, both crude-protein and energy content were assessed based on sampling of the top five leaves of each species at each sampling date. Samples with a fresh weight of >200–300 g were collected every 3–4 weeks throughout the 1992 grazing season (~end

of May–mid September) from a brackish salt marsh in Dollard Bay (53° 16'N, 7° 10'E), the Netherlands (Marsh Section 3 of the study area in [Esselink et al., 2000](#)). Crude protein content was calculated by multiplying the nitrogen content by a factor 6.25 ([Allen, 1989](#)). The *in-vitro* digestible dry matter content (DDM) was used as parameter for the energy content of the plant material from the animal perspective. DDM was measured according to [Tilley & Terry \(1963\)](#).

RESULTS

By the end of our livestock enclosure experiment, goats had strongly suppressed *Phragmites* growth, reducing stem density by ~50% (29–14 stems m⁻²; [Fig. 1A](#)), stem height by ~60% (3.9–1.4 m; [Fig. 1B](#)), and percent cover five-fold (from 94% to 21%; [Fig. 1C](#)). In grazed plots, *Phragmites* resprouted from rhizomes or colonized from adjacent ungrazed plots, but never attained the stem density, height, or cover of that in ungrazed plots, except in spring of the second growing season during early shoot emergence ([Fig. 1](#)). Concomitant with the strong decline of *Phragmites* was a marked and significant increase in plant species richness and Shannon–Weiner diversity index by the end of the experiment (~100% & 400% respectively, [Figs. 2A](#) and [2B](#)). At the end of the experiment, goat-grazed plots contained a total of 36 taxa (22 confirmed native species, 8 indeterminate, and 6 introduced taxa), while ungrazed plots contained only 20 taxa (12 native species, 2 indeterminate, and 6 introduced taxa), all of which were at low abundance relative to *Phragmites*. Important native species that colonized (from seeds in the seed bank or dispersed to the site) included *Alisma subcordatum*, *Epilobium coloratum*, *Leersia oryzoides*, *Mimulus ringens*, *Penthorum sedoides*, *Polygonum punctatum*, and *Ranunculus sceleratus*.

In our no-choice feeding trials, both horses and cows readily ate *Phragmites* (20 out of 20 individuals for both species ate the 30 cm stem section offered). In comparison with five other marsh plant species, nutritional quality of *Phragmites* had lower digestible matter content but higher protein content ([Fig. 3A](#)). It must be noted that North American introduced *Phragmites* was introduced from Europe, and we do not expect there to be any differences in tissue quality. Plants described in [Fig. 3A](#) have congeneric representation in North American wetlands, and serve as our proxy for comparable nutritional quality. Throughout the 4-month grazing season in the Dollard salt marshes, crude-protein content in *Phragmites* leaves was surprisingly high. The energy content of *Phragmites* leaves, on the contrary, was lower than in other common plant species, and dropped markedly during the course of the grazing season ([Fig. 3B](#)); after ~mid-July it fell below the level of maintenance requirement for cattle ([Australian Research Council, 1980](#); [Van Soest, 1982](#)). These values for digestible dry matter were within the range found by a larger survey of Dutch plant species palatability to cattle, which found *Phragmites* to be an important natural forage species ([Bokdam & Wallis de Vries, 1992](#)).

DISCUSSION

Our results and those of others in Europe indicate that controlling invasive *Phragmites* in North America via purposeful livestock grazing has a high potential to suppress its impact on native plant communities. Our feeding trial from The Netherlands, together

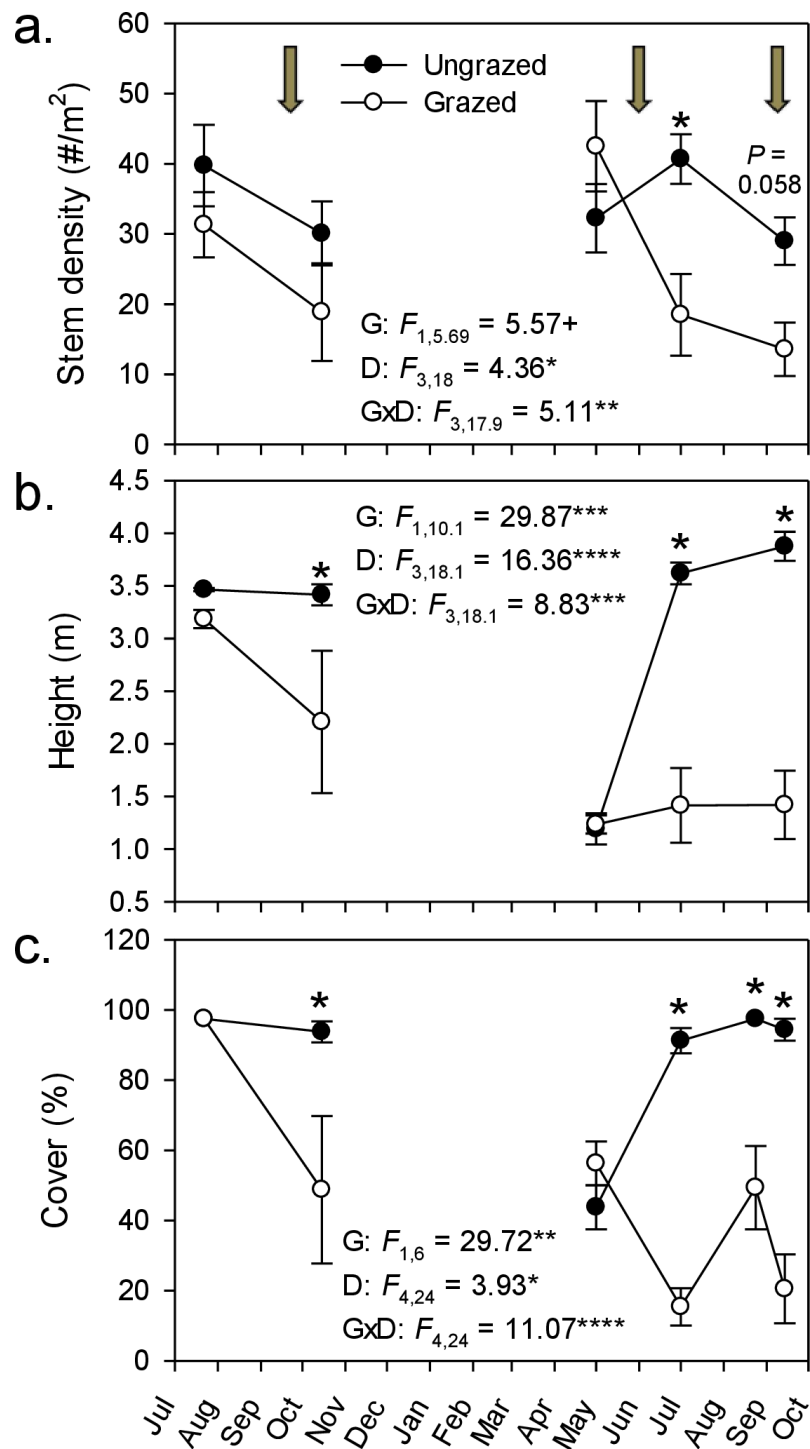


Figure 1 Goat grazing impacts on *Phragmites*. Effect of goats on three measures of *Phragmites australis* abundance from July 2008–October 2009. Values are mean \pm SE for 4 replicate enclosures (grazed) and control (ungrazed) plots. Stem density and height were determined in 1-m² plots and percent cover was determined in 100-m² plots. Arrows indicate the initiation of grazing periods; (continued on next page...)

Figure 1 (...continued)

for cover (C), the third grazing period falls between the last two measurement points. Results of repeated measures ANOVA are given within each panel for effects of grazing (G), date (D), and their interaction (G × D). + $P < 0.1$, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$. Asterisks above plotted points denote a significant grazing effect for that sampling date ($P < 0.05$, simple effect of grazing by date); P -value given for the last stem density comparison, where P was between 0.05 and 0.1.

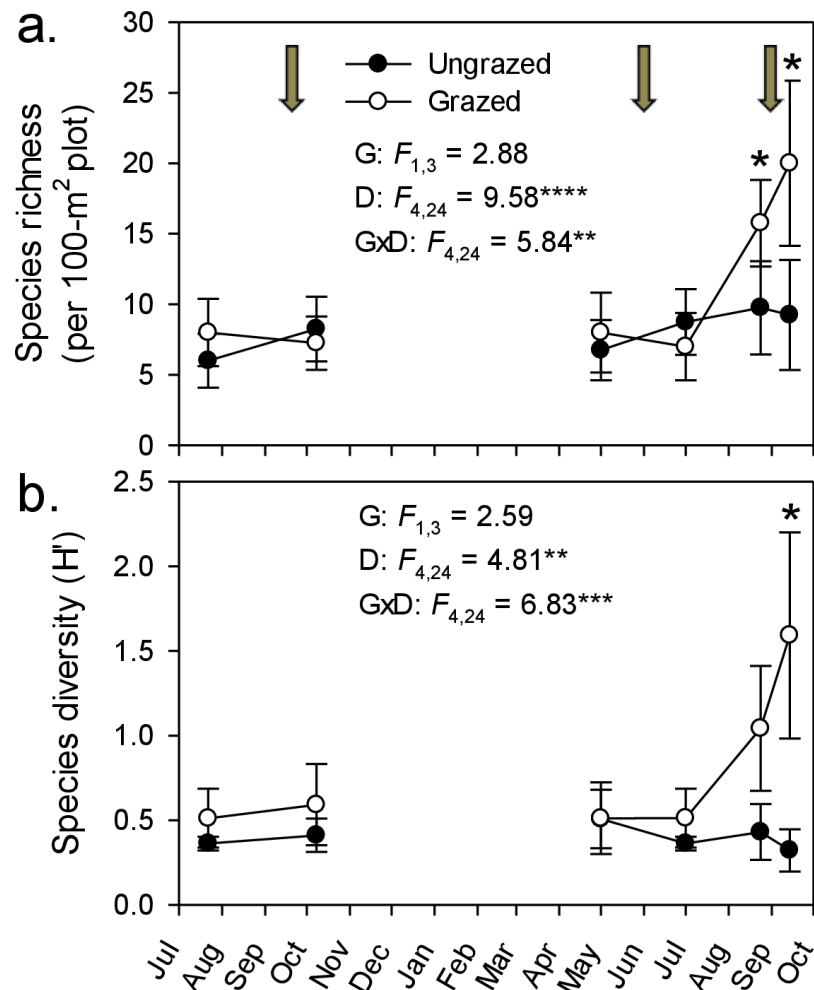


Figure 2 Goat grazing impacts on plant diversity. Changes in plant species richness (A) and Shannon–Weaver diversity (B) throughout the experiment in response to rotational goat grazing. Values are means ± SE for 4 replicate enclosures (grazed) and controls (ungrazed). Arrows indicate grazing period between sampling events; the third grazing period falls between the last two measurement points. An asterisk indicates a significant difference between grazed and ungrazed plots on a particular date ($P < 0.05$, simple effect of grazing by date). Also presented are results of repeated measures ANOVA for effects of grazing (G), date (D), and their interaction (G × D). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$.

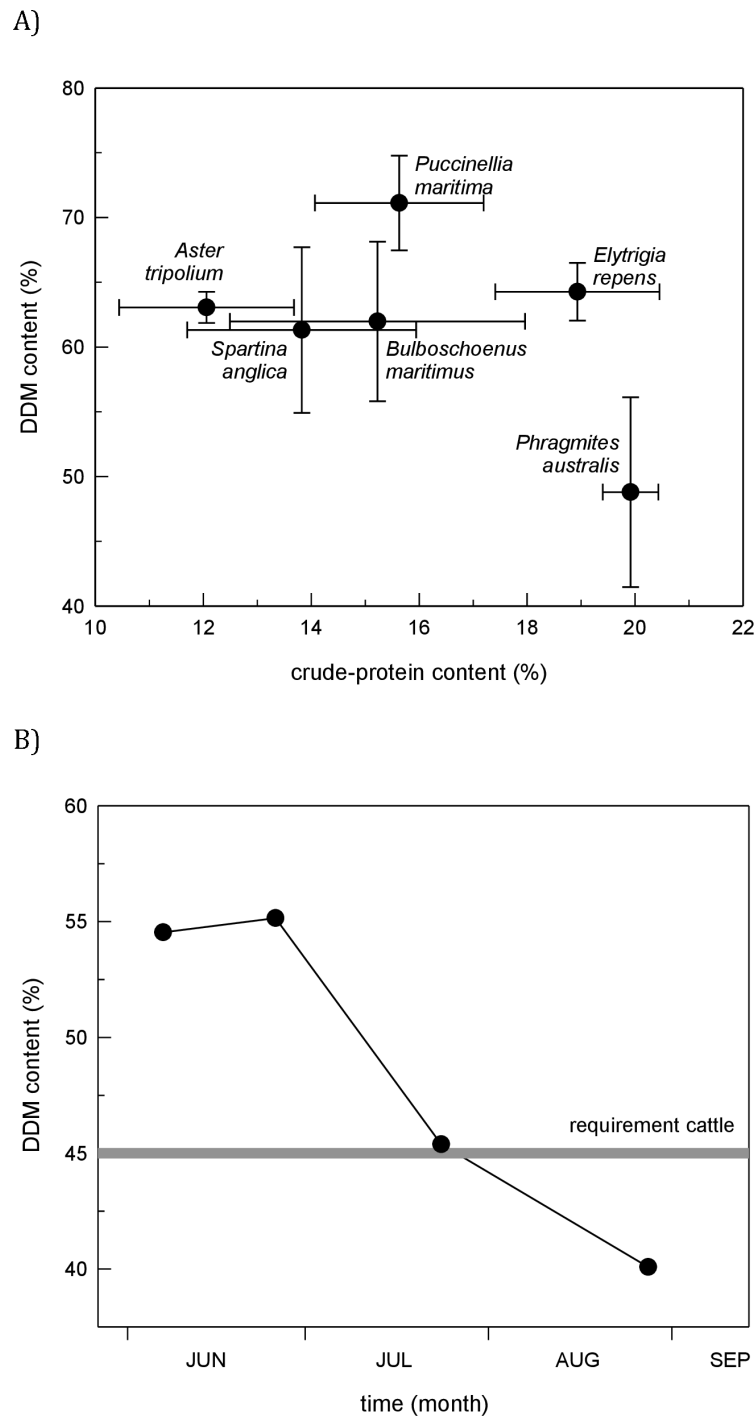


Figure 3 Marsh plant nutritional value. (A) Comparison of nutritional quality among six potential food plants in the cattle-grazed Dollard salt marshes, Netherlands. Figure shows the *in vitro* digestible dry-matter (DDM) content plotted with the crude-protein content (mean \pm SD) in young leaf tissue (five top leaves) during the grazing season (3rd decade of May–mid September). (B) Fall of forage quality (i.e., energy content) in leaf tissue of primary shoots of *Phragmites* in the Dollard salt marshes, Netherlands, during the grazing season based on the *in vitro* digestible dry-matter content in leaf tissue and compared with the level that cattle require for maintenance (after [Australian Research Council, 1980](#); [Van Soest, 1982](#)).

with evidence from livestock-removal and comparative studies in European marshes (Esselink, Fresco & Dijkema, 2002), suggest that livestock strongly restrict *Phragmites* distribution and facilitate the growth of shorter grasses and forbs in its native habitat. These results, in combination with our goat enclosure experiment in the U.S., indicate *Phragmites* is also likely susceptible to top-down control by livestock in Eastern North America. Furthermore, the short-term duration of our goat inclusion periods (3, <1 month deployments over 1 year), affordable infrastructure (wire fences), and limited number of animals (2 goats per 340-m² plot) needed to reduce *Phragmites* cover, imply that livestock has the potential to offer an effective, pesticide-free solution for managers trying to regulate this invasive plant, and likely other invasive plants that form vast monocultures. The conclusion that goat grazing could be an economically sustainable, win-win invasive plant control solution is also supported by the fact that livestock can persist over short time periods (i.e., weeks to months) on *Phragmites*-based diets without detriment to their health.

In invaded areas, *Phragmites* outcompetes native plants for light and space due to its height, dense canopy, thick litter, and rapidly growing rhizomes, and these advantages drive its rapid expansion and dominance across marshes (Bertness, Ewanchuk & Silliman, 2002; Silliman & Bertness, 2004; Mozdzer & Zieman, 2010; Holdredge et al., 2010). Our results and prior studies from Europe indicate that domestic livestock can reduce the competitive advantage of *Phragmites* through a combination of eating down or trampling live stems, breaking up the litter mat, and severing rhizomes with their hooves (Turner, 1987). Combined, these activities can increase the light availability to native plants, reduce belowground competition for nutrients, and thus provide opportunities for recolonization of native plants, estuarine nekton, and even endangered turtles (Angradi, Hagan & Able, 2001; Tesauero, 2002; Hunter et al., 2006; Tesauero & Ehrenfeld, 2007; Tesauero, 2002). In disrupting *Phragmites* growth, livestock also have the potential to reduce seed set, an important mechanism of expansion of *Phragmites* in North America (McCormick et al., 2010). By removing the primary mechanisms of *Phragmites* competitive exclusion (i.e., its height and litter), livestock may not only facilitate recovery of native plants and dependent faunal communities (e.g., invertebrates, arthropod herbivores), but also restore coastal ecosystem services. However, we must caution that introduction of livestock to invaded marshes in North America will not lead to a complete return to the pre-invasion marsh structure. Instead, we suspect that an alternative state will be induced (Hobbs & Norton, 1996). Such an alternative state may be characterized by a reduced *Phragmites* dominance and an increased abundance of native plants and fauna. Livestock grazing, however, is not without its own effects on ecosystem characteristics, affecting soil bulk density, soil organic matter, mineralization rate (Schrama et al., 2013), invertebrate abundance, among others. Comparative, multi-year trials are needed to assess grazing impacts and to determine the best regimen of grazers for *Phragmites* control, ecosystem integrity, and livestock production.

Context-dependency of grazer control and next steps

Evidence from our study coupled with other livestock and large grazer manipulative experiments (Tesauro, 2002; Sturm, 2007; URS, 2005) suggest that the efficacy of livestock control of *Phragmites* in North America will be context-dependent and contingent both on the grazing regime and the background cover of *Phragmites*. Specifically, these studies suggest that livestock can control *Phragmites* when its cover is high and livestock are forced to graze in those areas (Fig. 4). For example, when *Phragmites* is dominant and grazers are enclosed in these areas as in our experiment and a 2-year study in New Jersey, USA that manipulated goats and sheep in small (0.8 ha), un-replicated pens (Tesauro, 2002), livestock were effective at reducing *Phragmites* from ~100% to <50% cover. In contrast, when *Phragmites* is uncommon and livestock are free-roaming (i.e., grazers not forced to eat *Phragmites* only), horses and deer in Maryland increased *Phragmites* abundance relative to grazer exclusion plots (Sturm, 2007). Similarly, goats released into larger *Phragmites*-invaded tidal marsh in New Jersey did not reduce *Phragmites* cover and consumed other marsh plants to a greater degree (URS, 2005). These findings suggest that if livestock are released into mixed marsh plant communities where alternative food choices are abundant (i.e., *Phragmites* is uncommon) large grazers have the potential to facilitate *Phragmites* invasion, and thus be counterproductive to management objectives. This conclusion is supported by our nutritional content study (Fig. 3) and those of others (e.g., Vulink & Drost, 1991b) that indicate *Phragmites* has lower digestible matter content (although higher protein content) than other common European salt marsh plants relative and thus would not likely be preferred by grazers if given a choice.

Based on these conclusions, we suggest preliminary guidance for applications of livestock for invasive plant control (Table 1) and recommend future directions for research. Our grazing experiment, in which grazing had stronger effects in early summer than late summer, as well as our assessment of a decline in *Phragmites*' nutritional value through time, indicate that the timing of grazer implementation may be critical for the success of livestock control programs as young stems have higher nutritional quality (Fig. 3) and grazing on young *Phragmites*' stems in early spring is more effective at reducing future regrowth and promoting native plant recovery (Karunaratne, Asaeda & Yutani, 2004). Future research should address whether springtime or early summer grazing has stronger impact on *Phragmites* and other monoculture forming invasive plants. Looking forward, the next step in determining the potential for livestock to control *Phragmites* and facilitate the recovery of native plants, animals and pre-invasion soil properties is to test these ideas at larger scales and over multiple years to compare to reference wetlands without grazer control of invasive plants and those using other invasive species control techniques.

Finally, prior to application, it is critical to investigate the potential for livestock grazing impacts on non-target organisms and ecosystem processes. Decisions about the placement and timing of grazers should incorporate local site knowledge to avoid priority seasons and habitat areas for nesting birds and other possibly sensitive taxa or conservation targets. Further research could also identify the effects of short periods of grazing on critical wetland ecosystem processes such as soil compaction and surface elevation accretion,



Figure 4 Images of goat grazing impacts. Pictures of impacts of no-choice goat grazing in the *Phragmites*-dominated experimental wetland.

and examine the possibility that invasive plant seeds remain viable during livestock gut passage and are unwittingly dispersed to other sites.

Although inter-site variation and inter-annual differences make the synthesis of experimental findings from different decades and continents difficult, we find the consistent palatability of *Phragmites* to a diverse set of commercially important grazers in Europe and North America inspiring to pursue livestock grazing as a invasive species management tool. Other effective methods may be found by looking across ecosystems and

Table 1 Management considerations.

Based on our experimental findings, we find that livestock grazing for control of invasive plants holds great potential to reduce invasive plant biomass, increase plant diversity, and support livestock production. For effective control and to avoid negative impacts of over-grazing, we recommend:

1. High-intensity, short-duration, rotational grazing. Grazers will be most effective in dense, monotypic stands that are common in the establishment and spread phases of invasions. Periods without grazers are likely very important in allowing native plants to establish (Fig. 2) and for the health of grazing livestock (Fig. 3B).
2. Small scale enclosures to concentrate feeding on the dominant, invasive plant (Fig. 4). In the case of *Phragmites*, the high digestible dry matter content of other wetland plant species (Fig. 3A) suggests that livestock permitted to graze freely might prefer other available plants.
3. The incorporation of grazing into a long-term management scheme. Grazing will not eradicate an invasive plant, but will release native plants from invasive dominance temporarily. Therefore, grazing may have to occur throughout many years, and possibly indefinitely.
4. Species-specific grazing windows. Time grazing events to suppress dominant plant invaders (i.e., early in the growing season) and limit clonal regrowth while providing adequate windows for native plants recolonization.
5. Landscape considerations. Grazing is unlikely to be effective in soft-sediment environments, such as low elevation marshes, where trampling effects may overwhelm native plant recovery. Grazers will be most effective on firm soils, such as those in the high marsh and at the upland marsh ecotone, where *Phragmites* invasions begin.

continents for scenarios where dominant plants, whether purposefully or not, are being controlled using measures that involve and benefit local communities.

For monoculture-forming plants invading softer, lower elevations of marshes, such as *Spartina alterniflora*, on the West Coast of the US and China or *Cueerpa* in soft-sediment intertidal expanses throughout the world, domestic livestock are not likely an option for management. However, other economically sustainable, but currently overlooked, rotational top-down control methods may work for these species, such as systematic human harvesting of invasive plants to be used as livestock feed or biofuel.

Evidence from European marshes that a top-down restoration strategy will work

Restoration of coastal marshes presently dominated by *Phragmites* has not been practiced widely in Europe, except for in the Baltics. There, a comparison of uninterruptedly managed (seasonal summer grazing), abandoned (no grazing) and restored (i.e., summer grazing re-introduced after abandonment) sites in coastal marshes revealed that plant biomass in restored sites rapidly changed back to the level of managed marshes (Sammul, Kauer & Koster, 2012) and *Phragmites* cover decreased significantly. Plant species composition remained different, but typical coastal grassland species colonized and increased in abundance in restored sites (Sammul, Kauer & Koster, 2012). The response of soil properties to the re-introduction of grazing evolved more slowly. In abandoned sites for instance, organic matter content and C/N ratio were significantly higher and bulk density significantly lower than in continuously managed sites. In the five-year-old restored sites, however, all soil variables still did not differ from abandoned sites, implying

that the results of grazer-driven restoration may be slow for some variables. In addition, return of tall-growing *Phragmites* is likely if management intensity wanes. [Sammul, Kauer & Koster \(2012\)](#) conclude that *Phragmites* can indeed be suppressed in sites where it is dominant, but considering the slow response of soil properties, long-lasting periods of livestock-enhanced restoration should be planned in order to reach pre-abandonment environmental conditions.

Further incentives for integrating top-down control into invasive species management

While we have specifically focused on the control of an invasive plant as a management objective, this control has ancillary benefits and indirectly addresses multiple conservation targets. In addition to offering a solution for management of invasive plants that form expansive, hard-to-eradicate monocultures, livestock control programs can have reciprocal, positive impacts on local economies. Specifically, as is done in Europe, farmers could potentially receive payment for their services in controlling invasive species, and resources (e.g., fencing, transportation of livestock) to engage in such programs. At the same time, conservation groups and government organizations will receive more cost-effective and ecologically friendly tools to manage problematic invasive plants. Several instances of companies offering services of goats and other livestock to control *Phragmites* in urban wetlands in New York City, tidal wetlands in the Chesapeake Bay (e.g., Eco-Goats), and riverbank wetlands in the U.S. mid-west suggest that these ventures are marketable and can suppress *Phragmites* success (A Deer, pers. comm., 2014). More data are needed to confirm the short- and long-term sustainability of these business models. In many arenas, win-win solutions of economic gains in controlling invasives are often criticized with the argument that at some point the invasive species is going to be needed to maintain the economic model based upon it. In our proposed scenario using goat control of an invasive plant, however, we do not believe this will ever be the case as goats will likely always have more invasive to graze in the area (e.g., Kudzu) and, even when invasive plants have been locally suppressed, goats can still provide numerous benefits to their owners (e.g., dairy and meat production).

Beyond the target site where grazing is implemented, control of *Phragmites* reduces propagule pressure ([McCormick et al., 2010](#)) and interrupts positive feedback reducing spread to un-invaded sites ([Hazelton et al., 2014](#)). In addition, this approach provides an alternative treatment option when herbicide use is unacceptable or infeasible or where reduced *Phragmites* biomass and some native cover is an acceptable goal.

This general framework, designed to link invasive species management with the production of useable goods and benefit of local economies can also be applied to other systems where invasive species threaten ecosystem services ([Tulbure, Ghioca & Whigham, 2007](#); [Levin, 2006](#)). By identifying, and then harnessing the positive effects of grazers, coastal managers could potentially fulfill their conservation goals with significant reduction in cost. Overall, a shift in invasive species management from eradication to mitigation of invasive species impacts is creating opportunities for the implementation of new strategies, including the use of atypical top-down control agents.

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Author Contributions

- Brian R. Silliman analyzed the data, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Thomas Mozdzer, Christine Angelini, Jan P. Bakker and Johan van de Koppel wrote the paper, reviewed drafts of the paper.
- Jennifer E. Brundage, Peter Esselink and Andrew H. Baldwin conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Keryn B. Gedan analyzed the data, wrote the paper, reviewed drafts of the paper.

Animal Ethics

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REFERENCES

- Allen SE. 1989. *Chemical analysis of ecological materials*. 2nd edition. Oxford: Blackwell Scientific Publications, 368.
- Angradi TR, Hagan SM, Able KW. 2001. Vegetation type and the intertidal macroinvertebrate fauna of a brackish marsh: *Phragmites* vs. *Spartina*. *Wetlands* 21:75–92
[DOI 10.1672/0277-5212\(2001\)021\[0075:VTATIM\]2.0.CO;2](https://doi.org/10.1672/0277-5212(2001)021[0075:VTATIM]2.0.CO;2).

- Australian Research Council. 1980.** *The nutrient requirements of ruminant livestock*. Farnham Royal: Commonwealth Agricultural Bureaux.
- Belovsky GE. 1986.** Optimal foraging and community structure—implications for a guild of generalist grassland herbivores. *Oecologia* **70**:35–52 DOI [10.1007/BF00377109](https://doi.org/10.1007/BF00377109).
- Bertness MD, Ewanchuk PJ, Silliman BR. 2002.** Anthropogenic modification of New England salt marsh landscapes. *Proceedings of the National Academy of Sciences of the United States of America* **99**:1395–1398 DOI [10.1073/pnas.022447299](https://doi.org/10.1073/pnas.022447299).
- Bokdam J, Wallis de Vries MF. 1992.** Forage quality as a limiting factor for cattle grazing in isolated Dutch nature reserves. *Conservation Biology* **6**:399–408 DOI [10.1046/j.1523-1739.1992.06030399.x](https://doi.org/10.1046/j.1523-1739.1992.06030399.x).
- Chambers RM, Meyerson LA, Saltonstall K. 1999.** Expansion of *Phragmites australis* into tidal wetlands of North America. *Aquatic Botany* **64**:261–273 DOI [10.1016/S0304-3770\(99\)00055-8](https://doi.org/10.1016/S0304-3770(99)00055-8).
- Chambers RM, Osgood DT, Bart DJ, Montalto F. 2003.** *Phragmites australis* invasion and expansion in tidal wetlands: interactions among salinity, sulfide, and hydrology. *Estuaries* **26**:398–406 DOI [10.1007/BF02823716](https://doi.org/10.1007/BF02823716).
- Christian JM, Wilson SD. 1999.** Long-term ecosystem impacts of an introduced grass in the northern Great Plains. *Ecology* **80**:2397–2407 DOI [10.1890/0012-9658\(1999\)080\[2397:LTEIOA\]2.0.CO;2](https://doi.org/10.1890/0012-9658(1999)080[2397:LTEIOA]2.0.CO;2).
- Cowie NR, Sutherland WJ, Dithlago MKM, James R. 1992.** The effects of conservation management of reed beds II. The flora and litter disappearance. *Journal of Applied Ecology* **29**:277–284 DOI [10.2307/2404496](https://doi.org/10.2307/2404496).
- Dijkema KS. 1990.** Salt and brackish marshes around the Baltic Sea and adjacent parts of the North-Sea—their vegetation and management. *Biological Conservation* **51**:191–209 DOI [10.1016/0006-3207\(90\)90151-E](https://doi.org/10.1016/0006-3207(90)90151-E).
- DiTomaso JM. 2000.** Invasive weeds in rangelands: species, impacts, and management. *Weed Science* **48**:255–265 DOI [10.1614/0043-1745\(2000\)048\[0255:IWIRSI\]2.0.CO;2](https://doi.org/10.1614/0043-1745(2000)048[0255:IWIRSI]2.0.CO;2).
- Esselink P, Fresco LFM, Dijkema KS. 2002.** Vegetation change in a man-made salt marsh affected by a reduction in both grazing and drainage. *Applied Vegetation Science* **5**:17–32.
- Esselink P, Petersen J, Arens S, Bakker JP, Bunje J, Dijkema KS, Hecker N, Hellwig U, Jensen AV, Kers AS, Körber P, Lammerts EJ, Lüerssen G, Marencic H, Stock M, Veeneklaas RM, Vreeken M, Wolters M. 2009.** Salt marshes. In: Marencic H, de Vlas J, eds. *Quality status report 2009, Wadden sea ecosystems*, vol. 25. 1–54.
- Esselink P, Zijlstra W, Dijkema KS, Van Diggelen R. 2000.** The effects of decreased management on plant-species distribution patterns in a salt marsh nature reserve in the Wadden Sea. *Biological Conservation* **93**:61–76 DOI [10.1016/S0006-3207\(99\)00095-6](https://doi.org/10.1016/S0006-3207(99)00095-6).
- Flory SL, Clay K. 2010.** Non-native grass invasion alters native plant composition in experimental communities. *Biological Invasions* **12**:1285–1294 DOI [10.1007/s10530-009-9546-9](https://doi.org/10.1007/s10530-009-9546-9).
- Gurevich J, Scheiner SM, Fox GA. 2006.** *The ecology of plants*. Sunderland, MA: Sinauer.
- Hazelton ELG, Mozdzer TJ, Burdick DM, Kettenring KM, Whigham DF. 2014.** *Phragmites australis* management in the United States: 40 years of methods and outcomes. *AoB Plants* **6**:plu001 DOI [10.1093/aobpla/plu001](https://doi.org/10.1093/aobpla/plu001).
- Hobbs RJ, Norton DA. 1996.** Towards a conceptual framework for restoration ecology. *Restoration Ecology* **4**:93–110 DOI [10.1111/j.1526-100X.1996.tb00112.x](https://doi.org/10.1111/j.1526-100X.1996.tb00112.x).
- Holdredge C, Bertness MD, von Wettberg E, Silliman BR. 2010.** Nutrient enrichment enhances hidden differences in phenotype to drive a cryptic plant invasion. *Oikos* **119**:1776–1784 DOI [10.1111/j.1600-0706.2010.18647.x](https://doi.org/10.1111/j.1600-0706.2010.18647.x).

- Hunter KL, Fox DA, Brown LM, Able KW. 2006.** Responses of resident marsh fishes to stages of *Phragmites australis* invasion in three mid Atlantic estuaries. *Estuaries and Coasts* **29**:487–498.
- Jutila H. 1999.** Effect of grazing on the vegetation of shore meadows along the Bothnian Sea, Finland. *Plant Ecology* **140**:77–88 DOI [10.1023/A:1009744117329](https://doi.org/10.1023/A:1009744117329).
- Kareiva P, Marvier M. 2010.** *Conservation science: balancing the needs of people and nature*. Greenwood Village, Colorado: Roberts Publishers.
- Karunaratne S, Asaeda T, Yutani K. 2004.** Shoot regrowth and age-specific rhizome storage dynamics of *Phragmites australis* subjected to summer harvesting. *Ecological Engineering* **22**:99–111 DOI [10.1016/j.ecoleng.2004.02.006](https://doi.org/10.1016/j.ecoleng.2004.02.006).
- Kettenring KM, Adams CR. 2011.** Lessons learned from invasive plant control experiments: a systematic review and meta-analysis. *Journal of Applied Ecology* **48**:970–979 DOI [10.1111/j.1365-2664.2011.01979.x](https://doi.org/10.1111/j.1365-2664.2011.01979.x).
- King RS, Deluca WV, Whigham DF, Marra PP. 2007.** Threshold effects of coastal urbanization on *Phragmites australis* (common reed) abundance and foliar nitrogen in Chesapeake Bay. *Estuaries and Coasts* **30**:469–481 DOI [10.1007/BF02819393](https://doi.org/10.1007/BF02819393).
- Lambert AM, Winiarski K, Casagrande RA. 2007.** Distribution and impact of exotic gall flies (*Lipara* sp.) on native and exotic *Phragmites australis*. *Aquatic Botany* **86**:163–170 DOI [10.1016/j.aquabot.2006.09.017](https://doi.org/10.1016/j.aquabot.2006.09.017).
- Lavergne S, Molofsky J. 2004.** Reed canary grass (*Phalaris arundinacea*) as a biological model in the study of plant invasions. *Critical Reviews in Plant Sciences* **23**:415–429 DOI [10.1080/07352680490505934](https://doi.org/10.1080/07352680490505934).
- Lee SY. 1990.** Net aerial primary productivity, litter production and decomposition of the reed *Phragmites communis* in a nature-reserve in Hong-Kong—management implications. *Marine Ecology Progress Series* **66**:161–173 DOI [10.3354/meps066161](https://doi.org/10.3354/meps066161).
- Levin SA. 2006.** Learning to live in a global commons: socioeconomic challenges for a sustainable environment. *Ecological Research* **21**:328–333 DOI [10.1007/s11284-006-0162-1](https://doi.org/10.1007/s11284-006-0162-1).
- Levine JM, Adler PB, Yelenik SG. 2004.** A meta-analysis of biotic resistance to exotic plant invasions. *Ecology Letters* **7**(10):975–989 DOI [10.1111/j.1461-0248.2004.00657.x](https://doi.org/10.1111/j.1461-0248.2004.00657.x).
- Martin LJ, Blossey B. 2013.** The runaway weed: costs and failures of *Phragmites australis* management in the USA. *Estuaries and Coasts* **36**:626–632 DOI [10.1007/s12237-013-9593-4](https://doi.org/10.1007/s12237-013-9593-4).
- Marty JT. 2005.** Effects of cattle grazing on diversity in ephemeral wetlands. *Conservation Biology* **19**:1626–1632 DOI [10.1111/j.1523-1739.2005.00198.x](https://doi.org/10.1111/j.1523-1739.2005.00198.x).
- Meyerson L, Saltonstall K, Chambers R. 2009.** *Phragmites australis* in eastern North America: a historical and ecological perspective. In: Silliman BR, Gorsholz T, Bertness M, eds. *Human impacts in salt marshes: a global perspective*. California Press.
- Milotic T, Erfanzadeh R, Petillon J, Maelfait JP, Hoffmann M. 2010.** Short-term impact of grazing by sheep on vegetation dynamics in a newly created salt-marsh site. *Grass and Forage Science* **65**:121–132 DOI [10.1111/j.1365-2494.2009.00725.x](https://doi.org/10.1111/j.1365-2494.2009.00725.x).
- Minchinton TE, Bertness MD. 2003.** Disturbance-mediated competition and the spread of *Phragmites australis* in a coastal marsh. *Ecological Applications* **13**:1400–1416 DOI [10.1890/02-5136](https://doi.org/10.1890/02-5136).
- Morrison WE, Hay ME. 2011.** Herbivore preference for native vs. exotic plants: generalist herbivores from multiple continents prefer exotic plants that are evolutionarily Naïve. *PLoS ONE* **6**(3):e17227 DOI [10.1371/journal.pone.0017227](https://doi.org/10.1371/journal.pone.0017227).

- Mozdzer TJ, Brisson J, Hazelton ELG. 2013.** Physiological ecology and functional traits of North American native and Eurasian introduced *Phragmites australis* lineages. *AoB Plants* 5:plt048 DOI 10.1093/aobpla/plt048.
- Mozdzer TJ, Megonigal JP. 2012.** Jack-and-master trait responses to elevated CO₂ and N: a comparison of native and introduced *Phragmites australis*. *PLoS ONE* 7(10):e42794 DOI 10.1371/journal.pone.0042794.
- Mozdzer TJ, Megonigal JP. 2013.** Increased methane emissions by an introduced *Phragmites australis* lineage under global change. *Wetlands* 33(4):609–615 DOI 10.1007/s13157-013-0417-x.
- Mozdzer TJ, Zieman JC. 2010.** Ecophysiological differences between genetic lineages facilitate the invasion of non-native *Phragmites australis* in North American Atlantic coast wetlands. *Journal of Ecology* 98(2):451–458 DOI 10.1111/j.1365-2745.2009.01625.x.
- McCormick MK, Kettenring KM, Baron HM, Whigham DF. 2010.** Extent and reproductive mechanisms of *Phragmites australis* spread in brackish wetlands in Chesapeake Bay, Maryland (USA). *Wetlands* 30:67–74 DOI 10.1007/s13157-009-0007-0.
- Neira C, Grosholz ED, Levin LA, Blake R. 2006.** Mechanisms generating modification of benthos following tidal flat invasion by a *Spartina* hybrid. *Ecological Applications* 16:1391–1404 DOI 10.1890/1051-0761(2006)016[1391:MGMBOBF]2.0.CO;2.
- Parker JD, Burkepile DE, Hayt ME. 2006.** Opposing effects of native and exotic herbivores on plant invasions. *Science* 311(5766):1459–1461 DOI 10.1126/science.1121407.
- Peet RK, Wentworth TR, White PS. 1998.** A flexible, multi-purpose method for recording vegetation composition and structure. *Castanea* 63:262–274.
- Pimentel D, Zuniga R, Morrison D. 2005.** Update on the environmental and economic costs associated with alien-invasive species in the United States. *Ecological Economics* 52:273–288 DOI 10.1016/j.ecolecon.2004.10.002.
- Reiner R, Craig A. 2011.** Conservation easements in California blue oak woodlands: testing the assumption of livestock grazing as a compatible use. *Natural Areas Journal* 31:408–413 DOI 10.3375/043.031.0411.
- Rooth J, Stevenson JC, Cornwell JC. 2003.** Increased sediment accretion rates following invasion by *Phragmites australis*: the role of litter. *Estuaries* 26:475–483 DOI 10.1007/BF02823724.
- Saltonstall K. 2002.** Cryptic invasion by a non-native genotype of the common reed, *Phragmites australis*, into North America. *Proceedings of the National Academy of Sciences of the United States of America* 99:2445–2449 DOI 10.1073/pnas.032477999.
- Sammul M, Kauer K, Koster T. 2012.** Biomass accumulation during reed encroachment reduces efficiency of restoration of Baltic coastal grasslands. *Applied Vegetation Science* 15:219–230 DOI 10.1111/j.1654-109X.2011.01167.x.
- Sax DF, Stachowicz JJ, Gaines SD (eds.) 2005.** *Species invasions: insights into ecology, evolution and biogeography*. Sunderland, MA: Sinauer.
- Schrama M, Heijning P, Bakker JP, Berg MP, Olf H. 2013.** Herbivore trampling as an alternative pathway for explaining differences in nitrogen mineralization in moist grasslands. *Oecologia* 172:231–243 DOI 10.1007/s00442-012-2484-8.
- Silliman BR, Bertness MD. 2004.** Shoreline development drives invasion of *Phragmites australis* and the loss of plant diversity on New England salt marshes. *Conservation Biology* 18:1424–1434 DOI 10.1111/j.1523-1739.2004.00112.x.
- Sturm M. 2007.** *Assessment of the effects of feral horses, sika deer and white-tailed deer on Assateague Island's forest and shrub habitats*. Berlin, MD: Assateague Island National Seashore.

- Tesauro J. 2002.** *The effects of livestock grazing on the bog turtle (Clemmys muhlenbergii)*. New Brunswick, NJ: Rutgers University.
- Tesauro J, Ehrenfeld D. 2007.** The effects of livestock grazing on the bog turtle [*Glyptemys* (= *Clemmys*) *muhlenbergii*]. *Herpetologica* **63**:293–300
DOI [10.1655/0018-0831\(2007\)63\[293:TEOLGO\]2.0.CO;2](https://doi.org/10.1655/0018-0831(2007)63[293:TEOLGO]2.0.CO;2).
- Tewksbury L, Casagrande R, Blossey B, Häfliger P, Schärzlander M. 2002.** Potential for biological control of *Phragmites australis* in North America. *Biological Control* **23**:191–212
DOI [10.1006/bcon.2001.0994](https://doi.org/10.1006/bcon.2001.0994).
- Tilley JM, Terry RA. 1963.** A two stage technique for the *in vitro* digestion of forage crops. *Journal of the British Grassland Society* **18**:104–111 DOI [10.1111/j.1365-2494.1963.tb00335.x](https://doi.org/10.1111/j.1365-2494.1963.tb00335.x).
- Tulbure MT, Ghioca DM, Whigham CA. 2007.** *Comparative ecology of native and non-native Phragmites australis (common reed) genotypes*. Sacramento, CA: Society of Wetland Scientists.
- Turner MG. 1987.** Effects of grazing by feral horses, clipping, trampling, and burning on a Georgia salt-marsh. *Estuaries* **10**:54–60 DOI [10.2307/1352025](https://doi.org/10.2307/1352025).
- URS. 2005.** *Phragmites control alternatives assessment report*. Wayne, New Jersey: Prepared for PSEG Services Corporation Estuary Enhancement Program.
- Van der Putten WH. 1997.** Die-back of *Phragmites australis* in European wetlands: an overview of the European Research Programme on Reed Die-Back and Progression (1993–1994). *Aquatic Botany* **59**(3–4):263–275 DOI [10.1016/S0304-3770\(97\)00060-0](https://doi.org/10.1016/S0304-3770(97)00060-0).
- Van Driesche R, Blossey B, Hoddle M, Lyon S, Reardon R. 2002.** Biological control of invasive plants in the eastern United States. In: *USDA forest service publication FHTET-2002-04*. 413.
- Van Soest PJ. 1982.** *Nutritional ecology of the ruminant*. Corvallis, Oregon: O&B Books Inc.
- Vestergaard P. 1998.** Vegetation ecology of coastal meadows in Southeastern Denmark. *Opera Botanica* **134**:5–69.
- Vulink JT. 2001.** Hungry herds. Management of temperate lowland wetlands by grazing. *Van Zee tot Land* **66**:1–385.
- Vulink JT, Drost HJ. 1991a.** Nutritional characteristics of cattle forage plants in the eutrophic nature reserve Oostvaardersplassen, the Netherlands. *Netherlands Journal of Agricultural Science* **39**:263–272.
- Vulink JT, Drost HJ. 1991b.** A causal-analysis of diet composition in free ranging cattle in reed-dominated vegetation. *Oecologia* **88**:167–172 DOI [10.1007/BF00320807](https://doi.org/10.1007/BF00320807).
- Windham L, Ehrenfeld JG. 2003.** Net impact of a plant invasion on nitrogen-cycling processes within a brackish tidal marsh. *Ecological Applications* **13**:883–896 DOI [10.1890/02-5005](https://doi.org/10.1890/02-5005).
- Zedler JB, Kercher S. 2004.** Causes and consequences of invasive plants in wetlands: opportunities, opportunists, and outcomes. *Critical Reviews in Plant Sciences* **23**(5):431–452
DOI [10.1080/07352680490514673](https://doi.org/10.1080/07352680490514673).