# Functional Implications of Ubiquitous Semicircular Canal Non-Orthogonality in Mammals 

Jeri C. Berlin ${ }^{1 *}$, E. Christopher Kirk ${ }^{1,2}$, Timothy B. Rowe ${ }^{1,3}$<br>1 Vertebrate Paleontology Laboratory, University of Texas at Austin, Austin, Texas, United States of America, 2 Department of Anthropology, University of Texas at Austin, Austin, Texas, United States of America, $\mathbf{3}$ Jackson School of Geosciences, University of Texas at Austin, Austin, Texas, United States of America


#### Abstract

The 'canonical model' of semicircular canal orientation in mammals assumes that 1) the three ipsilateral canals of an inner ear exist in orthogonal planes (i.e., orthogonality), 2) corresponding left and right canal pairs have equivalent angles (i.e., angle symmetry), and 3) contralateral synergistic canals occupy parallel planes (i.e., coplanarity). However, descriptions of vestibular anatomy that quantify semicircular canal orientation in single species often diverge substantially from this model. Data for primates further suggest that semicircular canal orthogonality varies predictably with the angular head velocities encountered in locomotion. These observations raise the possibility that orthogonality, symmetry, and coplanarity are misleading descriptors of semicircular canal orientation in mammals, and that deviations from these norms could have significant functional consequences. Here we critically assess the canonical model of semicircular canal orientation using high-resolution X-ray computed tomography scans of 39 mammal species. We find that substantial deviations from orthogonality, angle symmetry, and coplanarity are the rule for the mammals in our comparative sample. Furthermore, the degree to which the semicircular canals of a given species deviate from orthogonality is negatively correlated with estimated vestibular sensitivity. We conclude that the available comparative morphometric data do not support the canonical model and that its overemphasis as a heuristic generalization obscures a large amount of functionally relevant variation in semicircular canal orientation between species.


Citation: Berlin JC, Kirk EC, Rowe TB (2013) Functional Implications of Ubiquitous Semicircular Canal Non-Orthogonality in Mammals. PLoS ONE 8(11): e79585. doi:10.1371/journal.pone. 0079585

Editor: Andrew A. Farke, Raymond M. Alf Museum of Paleontology, United States of America
Received August 8, 2013; Accepted October 2, 2013; Published November 19, 2013
Copyright: © 2013 Berlin et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: Funding for this research was provided by grants NSFIIS-0208675 (http://www.nsf.gov/cise/iis/hcc_pgm.jsp), and EAR-0948842 (http://www.nsf.gov/ awards/award_visualization.jsp?org = EAR). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.
Competing Interests: The authors have declared that no competing interests exist.

* E-mail: JeriCR55@aol.com


## Introduction

Detection of angular head accelerations is mediated by the semicircular canals of the inner ear. Each semicircular canal consists of a toroidal bony passage within the petrous portion of the temporal bone and contains an endolymph-filled duct. When the head rotates, inertial drag of endolymph within the duct acts upon sensory hair cells that modulate the firing rates of primary vestibular afferent neurons. Firing rates are either increased or decreased depending on the direction of head rotation [1]. The excitatory and inhibitory signals from all six semicircular canals are combined in the brain to generate reflexive movements that help to stabilize the eyes and head when the body is in motion [2].
In contrast with more readily accessible peripheral sense organs like the eye, the fact that the inner ear is encased within dense bone has hampered the comparative study of semicircular canal anatomy. This limitation, combined with the assumption that the three canals in each inner ear evolved to optimally detect rotations in each of the three orthogonal spatial dimensions, led researchers to rely on a series of simplifying assumptions about semicircular canal anatomy that are seldom critically examined. According to this "canonical model" of semicircular canal morphology, the plane of each canal is orthogonal to the planes of the other two ipsilateral canals so that all three canals in a single inner ear intersect at $90^{\circ}$ angles [3-9]. Furthermore, contralateral semicircular
canals are assumed to be essentially identical in dimension and orientation $[4,10]$. As a result, corresponding left and right canal pairs are expected to have equivalent angles and contralateral synergistic canals are expected to occupy parallel planes (Figure 1). These three basic components of the model, including orthogonality, angle symmetry, and coplanarity, are stated explicitly or implicitly in nearly every textbook or academic review covering the vestibular system [11-16]. Nonetheless, some empirical studies that measured semicircular canal orientation in a limited range of species reported results that are considerably divergent from the canonical model [e.g. 17,18-23]. Humans, for example, are reported to have ipsilateral canal pairs that differ by as much as $22^{\circ}$ from orthogonality [17].

The comparative morphology of semicircular canals is important because canal orientation ostensibly influences vestibular function [24-26]. However, most comparative analyses have examined the relationship between semicircular canal radius of curvature and locomotor agility [27-31]. Although radius of curvature is a major determinant of the sensitivity in each canal, the orientations of all six canals also help determine the relative sensitivity of the vestibular system to angular accelerations in three dimensions [15,24-26,32].

As a result, some authors have incorporated canal orientation in their calculations of vestibular sensitivity to angular accelerations $[25,33,34]$. To date, the largest comparative analysis of


Figure 1. Bony inner ear endocast of Petauroides volans (AMNH 150055) showing embedded head-centered reference planes and SCC canal pair types. A, dorsal view showing $X, Y$, and $Z$ axes. Center point occurs at intersection of all three planes. Axial plane $=Y Z$ reference plane passing through the interaural line; frontal plane $=X Y$ reference plane defined by Reid's Plane (perpendicular to viewer); sagittal plane $=X Z$ reference plane passing through central features. B, Left lateral view with sagittal plane perpendicular to viewer. Abbreviations: A, ampulla; C, cochlea; CC, common crus; LASC, left anterior semicircular canal; LLSC, left lateral semicircular canal; LPSC, left posterior semicircular canal; R, arc radius of curvature of the left anterior semicircular canal; $S$, saccule; $U$, utricle. doi:10.1371/journal.pone.0079585.g001
three-dimensional vestibular sensitivity focused on semicircular canal morphology and locomotor kinetics in 11 species of strepsirrhine primates [35]. This study found substantial variation between species in the homogeneity of three dimensional sensitivity maps, and demonstrated that strepsirrhines with more orthogonal canals tend to encounter higher angular head velocities during locomotion. More broadly, this analysis also provided evidence that deviations from canal orthogonality have important consequences for vestibular function.

In contrast to early studies of vestibular anatomy that relied on gross dissection or histology (11, 12, 19, 22, 28-30), computed tomography is now the standard for studies of semicircular canal morphology because it is nondestructive, quantitative, and can provide excellent resolution of internal cranial spaces [36,37]. We used high-resolution x-ray computed tomography scans of bilateral inner ear labyrinths to quantify semicircular canal size and orientation in 39 extant species from 11 mammalian orders (Table 1). For each taxon in our sample, these data were used to quantify mean deviations of ipsilateral semicircular canal pairs
from orthogonality $\left(90_{\mathrm{var}}\right)$, the degree to which corresponding contralateral canal pair angles differ (Angle Symmetry dev ), and the degree to which synergistic canal pairs deviate from coplanarity (Coplanarity ${ }_{\text {dev }}$ ). The term $90_{\text {var }}$ was introduced by Malinzak et al. as the sum of the absolute value of the difference between each of three unilateral ipsilateral canal pair angle and $90^{\circ}$ [35]. Here we calculate $90_{\text {var }}$ by summing the absolute value of the difference between all six ipsilateral semicircular canal pair angles and $90^{\circ}$, and taking the mean (see below).

We also used bilateral measurements of the size and orientation of all six semicircular canals to estimate the maximum (Sensitivity ${ }_{\text {max }}$ ) and average (Sensitivityave) sensitivity of the vestibular system to angular accelerations in three dimensions. These data for a large and taxonomically diverse sample allowed us to examine the degree to which orthogonality, symmetry, and coplanarity are characteristic of mammalian semicircular canals and to determine whether deviations from these norms are correlated with interspecific differences in estimated vestibular sensitivity.

Table 1. Taxa, museum specimen number, High Resolution X-ray Computed Tomography image slices used for skull images, spacing between image slices in image stack, Field of Reconstruction, and image slice pixel size.

| Taxon | Common Name | Museum Specimen Number | Number of slices | Interslice Spacing (mm) | Field of Reconstruction (mm) | File size (pixels) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Acrobates pygmaeus | Pygmy Gliding Possum | AMNH 155057 | 406 | 0.03 | 28.0 | $1024 \times 1024$ |
| Allactaga major | Five-toed Jerboa | AMNH 178795 | 1170 | 0.04 | 37.0 | $1024 \times 1024$ |
| Anomalurus beecrofti | Scaly-tailed Flying Squirrel | AMNH 50483 | 1270 | 0.04 | 39.0 | $1024 \times 1024$ |
| Caluromys sp. | Woolly Opossum | AMNH 95526 | 746 | 0.08 | 38.0 | $1024 \times 1024$ |
| Cavia porcellus | Guinea Pig | TMM M-7283 | 1524 | 0.04 | 42.0 | $1024 \times 1024$ |
| Cercartetus caudatus | Dormouse Possum | AMNH 155090 | 705 | 0.04 | 18.0 | $1024 \times 1024$ |
| Chinchilla laniger | Chinchilla | Hullar | 1887 | 0.04 | 40 | $1024 \times 1024$ |
| Chironectes minimus | Water Opossum | AMNH 129701 | 1522 | 0.04 | 33.0 | $1024 \times 1024$ |
| Chrysochloris sp. | Golden Mole | AMNH 82372 | 513 | 0.05 | 31.0 | $1024 \times 1024$ |
| Crocuta crocuta | Hyena | UCMVZ 184551 | 528 | 0.50 | 166.0 | $512 \times 512$ |
| Dactylopsila trivirgata | Striped Possum | AMNH 104040 | 1301 | 0.05 | 45.0 | $1024 \times 1024$ |
| Dolichotis patagonum | Patagonian Hare | AMNH 80078 | 1705 | 0.07 | 56.2 | $1024 \times 1024$ |
| Dromiciops gliroides | Monito del Monte | FMNH 127463 | 711 | 0.04 | 16.6 | $1024 \times 1024$ |
| Enhydra lutris | Sea Otter | SO 2853-97 | 645 | 0.22 | 106.0 | $1024 \times 1024$ |
| Felis catus | Domestic Cat | TMM M-628 | 606 | 0.15 | 70.0 | $1024 \times 1024$ |
| Glaucomys volans | Eastern Flying Squirrel | TMM M-6332 | 474 | 0.08 | 22.9 | $522 \times 522$ |
| Hemibelideus lemuroides | Brush-tipped Ring-tailed Possum | AMNH 154375 | 1207 | 0.05 | 40.0 | $1024 \times 1024$ |
| Heterocephalus glaber | Naked Mole Rat | AMNH 113974 | 1050 | 0.02 | 21.0 | $1024 \times 1024$ |
| Lepus californicus | Hare | TMM M-7500 | 660 | 0.14 | 67.0 | $1024 \times 1024$ |
| Meriones unguiculatus | Gerbil | TMM M-05306 | 1394 | 0.02 | 23.0 | $1024 \times 1024$ |
| Monodelphis domestica | Short-tailed Opossum | TMM M-9039 | 885 | 0.14 | 21.0 | $1024 \times 1024$ |
| Mus musculus | House Mouse | TMM M-3196 | 737 | 0.03 | 13.5 | $1024 \times 1024$ |
| Notoryctes typhlops | Marsupial Mole | AMNH 202107 | 705 | 0.04 | 18.0 | $1024 \times 1024$ |
| Ornithorhynchus anatinus | Duck-billed Platypus | TMM M-5899 | 1998 | 0.05 | 43.0 | $1024 \times 1024$ |
| Pedetes capensis | Springhare | AMNH 42016 | 1145 | 0.07 | 67.0 | $1024 \times 1024$ |
| Petauroides volans | Greater Gliding Possum | AMNH 150055 | 1251 | 0.05 | 48.0 | $1024 \times 1024$ |
| Petaurus breviceps | Sugar Glider | TMM M-8226 | 555 | 0.07 | 33.0 | $1024 \times 1024$ |
| Petropseudes dahli | Rock Possum | AMNH 183391 | 1424 | 0.05 | 46.0 | $1024 \times 1024$ |
| Potorous tridacty/us | Long-nosed Potoroo | AMNH 65337 | 915 | 0.01 | 48.0 | $1024 \times 1024$ |
| Pseudocheirus peregrinus | Common Ring-tailed Possum | TMM M-847 | 795 | 0.09 | 43.0 | $1024 \times 1024$ |
| Pseudochirops cupreus | Coppery Ring-tailed Possum | AMNH 151829 | 1289 | 0.05 | 49.5 | $1024 \times 1024$ |
| Pseudochirulus forbesi | New Guinean Ring-tailed Possum | AMNH 104136 | 1339 | 0.03 | 33.0 | $1024 \times 1024$ |
| Rattus norvegicus | Brown Rat | TMM M-2272 | 1571 | 0.03 | 28.0 | $1024 \times 1024$ |
| Saimiri sciureus | Squirrel Monkey | NSm7 | 310 | 0.07 | 64.0 | $1024 \times 1024$ |
| Sciurus niger | Fox Squirrel | UMMZ 123729 | 450 | 0.16 | 44.4 | $512 \times 512$ |
| Talpa europaea | Old World Mole | UCLGMZ 5437 | 585 | 0.06 | 18.5 | $1024 \times 1024$ |
| Tarsipes rostratus | Honey Possum | AMNH 119717 | 921 | 0.03 | 13.0 | $1024 \times 1024$ |
| Vulpes vulpes | Red Fox | UCLA 13112 | 825 | 0.17 | 80.0 | $1024 \times 1024$ |
| Wallabia bicolor | Swamp Wallaby | TMM M-4169 | 885 | 0.16 | 74.5 | $1024 \times 1024$ |

Museum Abbreviations: AMNH, American Museum of Natural History, New York; FMNH, Field Museum of Natural History, Chicago; TMM, Texas Natural Science Centers, Vertebrate Paleontology, Austin; UCL GMZ, University College, London Grant Museum of Zoology; SO, University of California Los Angeles Museum; UCLA, University of California Los Angeles; UCMVZ, University of California Museum of Vertebrate Zoology, Berkeley; UMMZ, University of Michigan Museum of Zoology, Ann Arbor.
doi:10.1371/journal.pone.0079585.t001

## Materials and Methods

The crania of 39 extant mammals, each representing a different genus, were scanned at the University of Texas High-Resolution

X-ray Computed Tomography Facility (Austin, Texas). This facility maintains an archive of all scans used in this analysis. Taxon, museum specimen number, and scan parameters for each cranium used in this study are listed in Table 1. With the
Table 2. Summary of semicircular canal angle relationships.

| Taxon | LASC $\times$ LLSC IPS | LASC $\Varangle$ LPSC IPS | LLSC $\Varangle$ LPSC IPS | RASC $\Varangle$ RLSC IPS | RASC $\Varangle$ RPSC IPS | RLSC $\times$ RPSC IPS | LASC $\Varangle$ RPSC SYN | LPSC $\Varangle$ RASC SYN | LLSC $\times$ RLSC SYN |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Acrobates | 103.86 | 90.54 | 81.01 | 98.35 | 89.17 | 86.43 | 7.63 | 9.86 | 8.95 |
| Allactaga | 72.26 | 82.03 | 86.86 | 71.09 | 83.50 | 84.69 | 19.58 | 19.55 | 10.55 |
| Anomalurus | 86.65 | 104.52 | 97.34 | 88.10 | 102.05 | 97.77 | 4.39 | 7.52 | 1.44 |
| Caluromys | 83.19 | 96.48 | 89.63 | 77.14 | 97.75 | 87.54 | 10.36 | 10.03 | 26.72 |
| Cavia | 82.29 | 86.47 | 85.12 | 88.99 | 87.10 | 84.44 | 7.11 | 6.69 | 1.19 |
| Cercartetus | 84.15 | 91.73 | 87.35 | 81.53 | 87.41 | 84.80 | 9.97 | 7.99 | 25.90 |
| Chinchilla | 86.19 | 84.87 | 83.58 | 74.51 | 87.09 | 81.58 |  |  |  |
| Chironectes | 82.09 | 104.33 | 91.91 | 77.84 | 93.55 | 93.34 | 19.10 | 11.69 | 20.25 |
| Chrysochloris | 58.82 | 86.41 | 103.87 | 71.65 | 84.12 | 91.58 | 15.48 | 10.06 | 14.71 |
| Crocuta | 80.54 | 95.71 | 99.05 | 81.39 | 94.32 | 90.01 | 3.59 | 8.48 | 13.20 |
| Dactylopsila | 82.82 | 89.83 | 94.77 | 88.04 | 95.08 | 96.03 | 9.25 | 2.69 | 7.16 |
| Dolichotis | 88.13 | 89.25 | 88.36 | 74.61 | 75.16 | 89.79 | 14.26 | 1.58 | 17.54 |
| Dromiciops | 92.87 | 89.71 | 92.39 | 88.75 | 101.19 | 94.69 | 19.17 | 14.68 | 15.72 |
| Enhydra | 89.35 | 98.42 | 91.64 | 91.16 | 102.79 | 83.18 | 3.91 | 0.51 | 8.55 |
| Felis | 87.27 | 83.62 | 81.94 | 83.79 | 84.62 | 81.65 | 12.57 | 11.53 | 10.75 |
| Glaucomys | 86.71 | 90.28 | 94.48 | 82.19 | 87.17 | 89.96 | 5.66 | 12.71 | 18.26 |
| Hemibelideus | 83.86 | 105.74 | 91.00 | 78.14 | 102.18 | 85.44 | 10.91 | 9.13 | 3.27 |
| Heterocephalus | 79.61 | 96.08 | 96.45 | 77.65 | 90.61 | 97.21 | 10.84 | 10.96 | 20.34 |
| Lepus | 87.92 | 93.87 | 87.13 | 87.87 | 95.84 | 87.45 | 4.60 | 5.19 | 0.61 |
| Meriones | 88.92 | 87.66 | 82.46 | 84.03 | 89.03 | 86.05 | 1.66 | 2.55 | 4.49 |
| Monodelphis | 83.90 | 95.99 | 91.06 | 81.31 | 99.56 | 89.32 | 15.05 | 17.04 | 24.19 |
| Mus | 85.82 | 78.08 | 85.21 | 85.84 | 84.89 | 85.26 | 6.09 | 10.45 | 9.91 |
| Notoryctes | 78.61 | 121.21 | 93.47 | 68.23 | 107.69 | 82.78 | 6.74 | 9.29 | 17.44 |
| Ornithorhynchus | 82.12 | 80.10 | 82.98 | 75.70 | 82.23 | 88.63 | 4.56 | 6.70 | 21.67 |
| Pedetes | 94.52 | 93.63 | 89.23 | 91.36 | 93.12 | 89.25 | 6.64 | 2.63 | 3.00 |
| Petauroides | 82.95 | 98.23 | 85.29 | 79.67 | 93.42 | 86.31 | 11.04 | 15.03 | 22.29 |
| Petaurus | 80.42 | 89.18 | 98.58 | 86.01 | 91.33 | 102.68 | 2.00 | 4.85 | 3.21 |
| Petropseudes | 76.77 | 89.07 | 93.20 | 89.48 | 89.09 | 90.86 | 15.74 | 14.14 | 3.61 |
| Potorous | 91.64 | 88.35 | 88.41 | 94.76 | 87.65 | 88.32 | 5.74 | 4.63 | 5.01 |
| Pseudocheirus | 84.86 | 93.67 | 92.90 | 84.10 | 98.52 | 90.41 | 19.13 | 24.15 | 6.42 |
| Pseudochirops | 93.28 | 83.83 | 104.81 | 87.93 | 87.37 | 85.92 | 11.87 | 15.29 | 9.53 |
| Pseudochirulus | 92.18 | 104.50 | 98.10 | 89.71 | 101.19 | 103.14 | 9.93 | 2.32 | 7.64 |
| Rattus | 88.64 | 87.63 | 79.72 | 84.10 | 87.33 | 81.97 | 11.65 | 9.96 | 8.22 |
| Saimiri | 85.97 | 79.77 | 89.55 | 87.26 | 76.24 | 88.10 | 7.97 | 12.29 | 5.52 |
| Sciurus | 79.26 | 86.88 | 90.60 | 84.04 | 82.88 | 91.93 | 5.54 | 1.71 | 6.10 |

Table 2. Cont.


[^0]exception of Chinchilla, for which a preexisting scan was made available by Dr. Timothy Hullar, all cranial specimens used in this analysis were scanned with the permission of the museums listed in Table 1. All crania were scanned bilaterally, ensuring that both the right and left semicircular canals were scanned in situ. This bilateral scanning protocol allowed measurements of contralateral canal pairs, a parameter that is rarely measured. Bilateral scanning also permitted the calculation of the vestibular sensitivity of each specimen to head rotations in three dimensions. The resulting image stacks were imported into VGStudioMax (Versions 1.2 and 2.0; Volume Graphics GmbH, 2004 and 2007) for 3D imaging and analysis.

For the present study, canal angle comparisons required stable head-centered reference planes, especially for angle comparison of contralateral canals. Three reference planes were determined and segmented into the 3D digital images before any other analysis was undertaken. The terminology follows that of vestibular researchers [e.g., 22,24,26]. Approximately eight small reference segments along the median sutures of the skull images (e.g., nasals, nasion, bregma, and medial palatine sutures) were aligned in a best-fit plane to define the vertical sagittal (XZ) plane. Numerous previous authors assumed that the LSC represents the horizontal plane of a live animal's head orientation, thus the alternative designation of the canal as the horizontal semicircular canal [e.g., 26,38,39], but that also assumed that both the left and right lateral canals lie within the same horizontal plane. Because this assumption is not correct (see below), we used bilateral measurements of Reid's line (the line extending from the lower edge of the orbit to the center of the aperture of the external auditory canal [40] to define the horizontal/frontal plane (XY). The axial (YZ) plane contained the line connecting the two external auditory meatuses (interaural line) perpendicular to the frontal and sagittal reference planes. The positive X axis of the resultant head-centered reference system passed through the rostrum, the positive Y axis passed through the left meatus, and the positive Z axis passed dorsally through the skull. Such a coordinate system was fitted successfully to all specimens except a Thylacinus, which was discarded for phylogenetic purposes.

Images were thresholded in VGStudioMax based on the density of the petrosal using the VGStudio density averaging tool. The selected region of the bony labyrinth was subsequently outlined for each CT slice image in VGStudioMax and added together to produce segments representing endocasts of both bony labyrinths in each specimen. A resulting file of the reference planes and bony labyrinth endocasts for Petauroides volans is shown in Figure 1. For our determination of canal orientations with regard to the reference planes in VGStudioMax, we used a measurement tool to select an array of points ( $\sim 60-100$ per canal) representing the lumen centers of a canal from the end of the ampulla, around the canal and including the common crus. A circle circumference for each canal was calculated by a linear regression best-fit of the selected lumen points. The radius to the semicircular canal circumference ( R , in mm ) was recorded for use in sensitivity calculations. The fitpoints were imported into a VGStudioMax best-fit calculation to obtain the plane containing that semicircular canal [19], defined by coordinates of the unit normal axis perpendicular to that plane. A plane's normal line has no polarity, but each semicircular canal can be rotated in a direction that provides an increase in afferent neuron firing rate (excitatory direction) or it may be rotated in the opposite direction to produce a decrease in afferent neuron firing rate (inhibitory direction). To express this additional information, the normal line, serving as an axis of rotation, was polarized to give a vector ( $V$ ) showing excitatory sensitivity direction according to the right-hand rule as

Table 3. Summary data for the 39 mammalian species in the comparative sample.

|  | ASC $\Varangle$ LSC IPS | ASC $\Varangle$ PSC IPS | LSC $\chi_{4} \mathbf{P S C}$ IPS | ASC $\Varangle$ PSC SYN | LSC $\Varangle$ LSC SYN |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Arithmetic Mean | 84.49 | 91.87 | 90.32 | 9.52 | 11.22 |
| Arithmetic Standard Deviation | 7.33 | 8.14 | 5.91 | 5.19 | 7.51 |
| Mean Direction | 84.50 | 91.85 | 90.32 | 9.52 | 11.21 |
| Circular Standard Deviation | 4.80 | 5.32 | 3.87 | 3.40 | 4.88 |

described by Ezure and Graf [19] and utilized by Calabrese and Hullar [22]. Mathematical calculation of angles between all canals was performed in VGStudioMax, with corrections to ensure all angles are internal (in lateral direction of skull) [see 19,41]. Naming convention of the angles closely follows that of Spoor and Zonneveld [42]. For example, LASC $\Varangle$ LLSC refers to the angle between the left anterior semicircular canal and the left lateral semicircular canal. All angles measured for each species are listed in Table 2, and summary data for ipsilateral canal angles and synergistic contralateral canals are provided in Table 3. The summary angular data in Table 3 includes both arithmetic means with standard deviations, as well as mean directions with circular standard deviations calculated by treating our data as vectors [43]. The arithmetic mean and mean direction for these data demonstrated negligible differences (i.e., $\leqq 0.02^{\circ}$ ), while the circular standard deviation is less than the arithmetic standard deviation (Table 3). In all analyses, our angular measurements were quantified as the absolute value of the deviation from an expected value (either $90^{\circ}$ or $180^{\circ}$; see below). Although these measurements are expressed in degrees, the data used in all analyses are scalar and do not require the use of circular statistics.

## Orthogonality, Symmetry, and Coplanarity Calculations

We quantified semicircular canal orientation by comparing the deviations of canal pair angles from the expected normative values. Deviation from orthogonality $\left(90_{\mathrm{var}}\right)$ [35] was calculated by taking the absolute value of the difference between each canal pair angle and $90^{\circ}$, adding those difference for all six ipsilateral canal pairs, and dividing by six. Deviation from side to side semicircular canal angle symmetry (Angle Symmetry ${ }_{\text {dev }}$ ) was calculated as the absolute value of the difference between the left canal pair angle and the corresponding right canal pair angle. To quantify deviation from coplanarity (Coplanarity dev ) we first subtracted the angle between each synergistic contralateral canal pair from $180^{\circ}$. We then summed the absolute value of this difference for each of the three synergistic canal pairs and divided by 3 . Values for these variables are given in Table 4.

## Sensitivity Calculations

A rotation of the head in the plane of a given semicircular canal (i.e. around V ) increases the firing rate of the vestibular nerve cells in that canal above a resting rate (in spikes $\cdot \sec ^{-1}$ ), or decreases the firing rate (axis opposite to V ). The rate of nerve cell firing change is proportional to R and the speed of rotation (in degrees • $\sec ^{-1}$ ), and is referred to as the sensitivity of the canal to rotation (in spikes $\cdot \sec ^{-1} /$ degrees $\cdot \sec ^{-1}$ ) $[16,24,25,32]$. A head rotation around an axis with orientation X changes the sensitivity of the canal nerve responses in proportion to the cosine of the angle between X and V . The responses of all six individual semicircular canals to rotation along X can be determined with R and V for each canal, and the orientation of X . Therefore, for a head
rotation along any head-centered axis X , the sensitivities of all six canals can be summed to provide a global sensitivity to the rotation ( S , in spikes $\cdot \mathrm{sec}^{-1} /$ degrees $\cdot \mathrm{sec}^{-1}$ ). By calculating such global sensitivities for a high number of rotational axes, an axis of rotation resulting in the maximum summed sensitivity of all six canals can be determined (Sensitivity ${ }_{\text {max }}$ ). The average sensitivity for a high number of rotational axes can also be calculated (Sensitivity ave ). The Sensitivity ${ }_{\text {max }}$ and Sensitivity ${ }_{\text {ave }}$ values calculated for specimens used in this study are listed in Table 4. Both sensitivity values were calculated using Bubbles.mat [35] software, which is described in detail by Yang and Hullar [25], Rodgers [32], and Malinzak et al. [32]. Bubbles.mat uses both the orientation and radius of curvature of the six semicircular canals to calculate estimated sensitivity of the vestibular system to angular accelerations in three dimensions. According to the Bubbles.mat results, the calculated Sensitivity max $^{\text {a }}$ is a function of both canal orientations and canal radii but Sensitivity ave is entirely determined by canal radii. These effects are illustrated in Figure 2, which shows the relationship between $90_{\mathrm{var}}$ and the ratio of our observed sensitivities to sensitivity calculated with canals constrained to strict orthogonality (OSensitivity ${ }_{\text {max }}$ and OSensitivity${ }_{\mathrm{min}}$ ) through setting all ipsilateral canal angles as $90^{\circ}$ bilaterally, and setting ASC and PSC $45^{\circ}$ away from the sagittal plane for Bubbles.mat calculations. In Figure 2, the ratio of Sensitivity ave $^{\text {to }}$ OSensitivity $_{\text {ave }}$ is always 1.0 across a range of $90_{\text {var }}$ values which indicates that Sensitivityave is solely a function of R. By comparison, the ratio of Sensitivity $y_{\text {max }}$ to OSensitivity max is always greater than 1.0 and tends to increase with increases in $90_{\text {var }}$ (Figure 2). This ratio of Sensitivity max : OSensitivity max $^{\text {max }}$ reflects the fact that artificially constraining strict orthogonality leads to a decrease in the maximum estimated sensitivity of the vestibular system according to the Bubbles.mat calculations. The implications of these effects for our analyses of canal orientation an estimated sensitivity are discussed below.

## Statistical Analysis

All statistical tests were performed in R using the ape and nlme packages [44-46]. A normal distribution for all continuous data was confirmed using Shapiro-Wilk tests for normality. Raw data for $90_{\text {var }}$ and Angle Symmetry ${ }_{\text {dev }}$ deviated significantly from normality, so these data were $\log 10$ transformed to satisfy the assumptions of parametric statistical tests. A normal distribution for both $90_{\text {var }}$ and Angle Symmetry $y_{\text {dev }}$ following $\log 10$ transformation was confirmed with a Shapiro-Wilk test. Data for Coplanarity $_{\text {dev }}$, Sensitivity ${ }_{\text {max }}$, and Sensitivity ${ }_{\text {ave }}$ did not deviate significantly from normality and were therefore included in analyses without transformation.

Two types of statistical tests were used assess the relationship between the morphology and estimated sensitivity of the semicircular canals. First, Pearson product-moment correlations were calculated for our three measures of canal morphology $\left(90_{\mathrm{var}}\right.$,

Table 4. Deviations from orthogonality ( $90_{\text {var }}$ ), side-to-side angle symmetry (Angle Symmetry ${ }_{\text {dev }}$ ), and synergistic canal coplanarity (Coplanarity ${ }_{\text {dev }}$ ).

| Taxon | $\mathbf{9 0}_{\text {var }}$ | Angle Symmetry ${ }_{\text {dev }}$ | Coplanarity ${ }_{\text {dev }}$ | $\mathbf{S}_{\text {max }}$ | Save | $\mathbf{O S}_{\text {max }}$ | $\mathrm{OS}_{\text {ave }}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Acrobates | 6.02 | 4.10 | 8.83 | 0.42 | 0.34 | 0.39 | 0.34 |
| Allactaga | 9.93 | 1.60 | 16.56 | 1.35 | 1.05 | 1.22 | 1.05 |
| Anomalurus | 7.82 | 1.45 | 4.44 | 1.23 | 0.99 | 1.14 | 0.99 |
| Caluromys | 6.12 | 3.13 | 15.70 | 0.68 | 0.56 | 0.65 | 0.56 |
| Cavia | 4.27 | 2.67 | 5.00 | 1.17 | 0.97 | 1.12 | 0.97 |
| Cercartetus | 4.42 | 3.16 | 14.32 | 0.49 | 0.34 | 0.40 | 0.34 |
| Chinchilla | 7.03 | 5.30 |  | 1.39 | 1.10 | 1.27 | 1.1 |
| Chironectes | 7.20 | 5.49 | 17.01 | 0.71 | 0.59 | 0.68 | 0.59 |
| Chrysochloris | 12.41 | 9.13 | 13.41 | 0.38 | 0.27 | 0.32 | 0.27 |
| Crocuta | 6.20 | 3.76 | 8.41 | 2.17 | 1.78 | 2.07 | 1.78 |
| Dactylopsila | 4.20 | 3.91 | 6.40 | 0.87 | 0.72 | 0.84 | 0.72 |
| Dolichotis | 5.78 | 9.68 | 11.13 | 1.62 | 1.30 | 1.50 | 1.3 |
| Dromiciops | 3.78 | 5.97 | 16.53 | 0.38 | 0.32 | 0.37 | 0.32 |
| Enhydra | 5.25 | 4.88 | 4.36 | 1.62 | 1.30 | 1.50 | 1.3 |
| Felis | 6.19 | 1.59 | 11.62 | 1.42 | 1.07 | 1.24 | 1.07 |
| Glaucomys | 3.12 | 4.05 | 12.21 | 0.88 | 0.74 | 0.85 | 0.74 |
| Hemibelideus | 8.58 | 4.95 | 7.79 | 1.52 | 1.17 | 1.37 | 1.17 |
| Heterocephalus | 7.18 | 2.73 | 14.04 | 0.41 | 0.33 | 0.39 | 0.33 |
| Lepus | 3.22 | 0.78 | 3.48 | 1.35 | 1.13 | 1.31 | 1.13 |
| Meriones | 3.64 | 3.28 | 2.90 | 0.83 | 0.68 | 0.78 | 0.68 |
| Monodelphis | 5.35 | 2.64 | 18.76 | 0.46 | 0.37 | 0.44 | 0.37 |
| Mus | 5.82 | 2.29 | 8.82 | 0.26 | 0.21 | 0.25 | 0.21 |
| Notoryctes | 15.45 | 11.55 | 11.14 | 0.38 | 0.27 | 0.32 | 0.27 |
| Ornithorhynchus | 8.04 | 4.73 | 10.98 | 1.12 | 0.90 | 1.05 | 0.9 |
| Pedetes | 2.36 | 1.23 | 4.05 | 1.58 | 1.32 | 1.53 | 1.32 |
| Petauroides | 6.24 | 3.04 | 16.12 | 1.11 | 0.92 | 1.07 | 0.92 |
| Petaurus | 6.16 | 3.95 | 3.39 | 0.73 | 0.58 | 0.67 | 0.58 |
| Petropseudes | 3.27 | 5.02 | 11.16 | 1.16 | 0.93 | 1.08 | 0.93 |
| Potorous | 2.28 | 1.31 | 5.11 | 1.20 | 0.99 | 1.15 | 0.99 |
| Pseudocheirus | 4.42 | 2.70 | 16.57 | 1.16 | 0.95 | 1.11 | 0.95 |
| Pseudochirops | 5.51 | 9.26 | 12.23 | 1.11 | 0.92 | 1.07 | 0.92 |
| Pseudochirulus | 8.24 | 3.61 | 6.63 | 1.00 | 0.79 | 0.92 | 0.79 |
| Rattus | 5.10 | 2.36 | 9.94 | 0.75 | 0.63 | 0.73 | 0.63 |
| Saimiri | 5.52 | 2.09 | 10.13 | 1.32 | 1.05 | 1.21 | 1.05 |
| Sciurus | 4.91 | 3.37 | 4.47 | 1.65 | 1.32 | 1.53 | 1.32 |
| Talpa | 8.78 | 8.73 | 7.57 | 0.59 | 0.48 | 0.56 | 0.48 |
| Tarsipes | 7.41 | 8.86 | 8.15 | 0.26 | 0.20 | 0.23 | 0.2 |
| Vulpes | 5.32 | 4.74 | 8.72 | 1.62 | 1.30 | 1.51 | 1.3 |
| Wallabia | 2.41 | 3.47 | 16.66 | 1.81 | 1.55 | 1.81 | 1.55 |

Sensitivities calculated from canal dimensions [32] and angles from Table 2. Hypothetical 'Orthogonal Sensitivity' calculated by forcing all canal angles to be orthogonal and symmetrical, but keeping all $R$ dimensions as originally measured.
doi:10.1371/journal.pone.0079585.t004

Angle Symmetry ${ }_{\text {dev }}$, and Coplanarity $y_{\text {dev }}$ ) and our two estimates of sensitivity to angular accelerations in three dimensions (Sensitivi$t_{\text {max }}$ and Sensitivityave). (Table 5). Because we expect deviations from orthogonality (i.e., greater $90_{\text {var }}$ values), deviations from equality in corresponding contralateral angle pairs (i.e., greater Angle Symmetry ${ }_{\text {dev }}$ values), and deviations from coplanarity in synergistic canal pairs (i.e., greater Coplanarity ${ }_{\text {dev }}$ values) to be negatively correlated with vestibular sensitivity, all correlations
were one-tailed. Second, phylogenetic generalized least-squares regression (PGLS) [47] was used to examine the relationships between canal morphology and estimated sensitivity while controlling for phylogenetic relationships. Tree topology and branch lengths for the included taxa follow Bininda-Emonds et al. $[48,49]$. The strength of the phylogenetic signal (i.e., the degree to which data approximate a Brownian-motion model of evolution)


Figure 2. Effect of constraining semicircular canals to strict orthogonality. This plot shows the relationship between $90_{\text {var }}$ ( $x$-axis) and two ratios (y-axis): (1) Maximum observed sensitivity ( $\mathrm{S}_{\max }$ ) : Maximum sensitivity with orthogonality constrained ( $\mathrm{OS}_{\max }$ ) (blue diamonds), and (2) Average observed sensitivity ( $\mathrm{S}_{\text {ave }}$ ) : Average sensitivity with orthogonality constrained ( $\mathrm{OS}_{\text {ave }}$ ) (green squares). Data from Table 4.
doi:10.1371/journal.pone.0079585.g002
in each PGLS analysis was quantified using Pagel's lambda ( $\lambda$ ) [50].

## Results

## Deviations from orthogonality, angle symmetry, and coplanarity

We find that the angle between two ipsilateral canals ranges between $58.8^{\circ}$ and $121.2^{\circ}$ for the species in our sample (Table 2; Figure 3). The smallest canal pair angle in our sample (LASC $\Varangle$ LLSC in Chrysochloris) is $31.2^{\circ}$ less than $90^{\circ}$, while the largest canal pair angle in our sample (LASC $\Varangle$ LPSC in Notoryctes) is $31.2^{\circ}$ greater than $90^{\circ}$. The mean angle for all ipsilateral canal pairs across all taxa is $88.9^{\circ}$ (st. dev. $=7.8^{\circ}$ ) and the average deviation from orthogonality for all ipsilateral canal pairs is $6.0^{\circ}$ (st. dev. $=5.1^{\circ}$ ). Similarly, $90_{\text {var }}$ ranges from a low value of $2.3^{\circ}$ in Potorous to a high value of $15.5^{\circ}$ in Notoryctes (mean $=6.0^{\circ}$, st. dev. $=2.6^{\circ}$ ). Of the ipsilateral canal pairs, ASC $\Varangle$ LSC is smallest, with a mean angle of $84.5^{\circ}$ (st. dev. $=7.3^{\circ}$ ) and a $95 \%$ mean confidence interval of $83^{\circ}-86^{\circ}$ (i.e., excluding orthogonality). By
comparison, mean $\operatorname{ASC} \Varangle \mathrm{PSC}$ is $91.9^{\circ}$ (st. dev. $=8.1^{\circ}$ ) and mean LSC $\Varangle$ PSC is $90.3^{\circ}$ (st. dev. $=5.9^{\circ}$ ) (Table 3).

In addition to these deviations from orthogonality, our data demonstrate that ipsilateral canal pair angles differ by an average of $4.3^{\circ}$ (st. dev. $=2.6^{\circ}$ ) between the right and left sides of individual specimens. Mean Angle Symmetry ${ }_{\text {dev }}$ values range from a low of $0.8^{\circ}$ in Lepus to a high of $11.6^{\circ}$ in Notoryctes. Angles between synergistic canal pairs range from $0.5^{\circ}$ (Enhydra LPSC $\Varangle$ RASC) to $27.7^{\circ}$ (Caluromys LLSC $\Varangle$ RLSC). The mean deviation from coplanarity is $9.5^{\circ}$ (st. dev. $=5.2^{\circ}$ ) for the two ASC $\Varangle$ PSC pairs and $11.2^{\circ}$ (st. dev. $=7.5^{\circ}$ ) for the LSC $\Varangle$ LSC pair (Table 3). The mean deviation of all three synergistic canal pair angles from coplanarity is $10.1^{\circ}\left(\right.$ st. dev. $\left.=6.1^{\circ}\right)$.

## Canal orientation and vestibular sensitivity

As expected, there is a negative correlation between the two estimates of vestibular sensitivity and the three measures of semicircular canal orientation (Table 5). This relationship is significant at $\mathrm{P}<0.05$ for all comparisons except that of Angle Symmetry $_{\text {dev }}$ and Sensitivity ${ }_{\text {max }}$, which are negatively correlated at $\mathrm{P}=0.054$. These data demonstrate that species with lower

Table 5. Results of Statistical Tests.

|  | $\mathbf{9 0}_{\text {var }}$ | Angle Symmetry ${ }_{\text {dev }}$ | Coplanarity ${ }_{\text {dev }}$ |
| :---: | :---: | :---: | :---: |
| Save | Pearson: $\mathrm{P}=\mathbf{0 . 0 1 9 *}$; $\mathrm{r}=-0.335$ PGLS: $P=\mathbf{0 . 0 2 9 * ; ~} \lambda=0.648$ | Pearson: $\mathrm{P}=\mathbf{0 . 0 4 7 *}$; $r=-0.272$ PGLS: $P=0.236 ; \lambda=0.636$ | Pearson: $P=\mathbf{0 . 0 4 1}$ * $r=-0.286$ PGLS: $P=0.363 ; \lambda=0.762$ |
| $\mathrm{s}_{\text {max }}$ | Pearson: $\mathrm{P}=\mathbf{0 . 0 3 6}$ *; $\mathrm{r}=-0.292$ PGLS: $\mathrm{P}=\mathbf{0 . 0 6 0} ; \lambda=0.682$ | Pearson: $P=\mathbf{0 . 0 5 4} ; r=-0.262$ PGLS: $P=0.273 ; \lambda=0.684$ | $\begin{aligned} & \text { Pearson: } P=\mathbf{0 . 0 4 6} * ; r=-0.276 \\ & \text { PGLS: } P=0.372 ; \lambda=0.791 \end{aligned}$ |

[^1]

Figure 3. Comparison of ipsilateral canal pair angles from left and right inner ears. Data from Table 2. doi:10.1371/journal.pone.0079585.g003
estimated sensitivity to angular accelerations tend to have semicircular canals that deviate more from orthogonality, angle symmetry, and coplanarity. However, the strength of these negative correlations is relatively modest, with correlation coefficients ranging between -0.262 and -0.335 (Table 5). Furthermore, when these relationships are analyzed using PGLS regression to control for phylogenetic non-independence, the results for Angle Symmetry ${ }_{\mathrm{dev}}$ and Coplanarity ${ }_{\mathrm{dev}}$ are nonsignificant. By contrast, the PGLS regression of $90_{\text {var }}$ and Sensitivity $_{\text {ave }}$ remains significant at $\mathrm{P}=0.029$ (Figure 4) and the PGLS regression of $90_{\text {var }}$ and Sensitivity max $_{\text {m }}$ is near significance at $\mathrm{P}=0.060$. Lambda values for all six comparisons demonstrate that the relationships between these variables do not follow a strict Brownian motion model of evolution, nor are they completely free of the influence of phylogeny (Table 5). According to these results, phylogenetic proximity has the smallest influence on the relationship between Angle Symmetry ${ }_{\text {dev }}$ and vestibular sensitivity ( $\lambda \sim 0.64-0.68$ ) and the greatest influence on the relationship between Coplanarity ${ }_{\mathrm{dev}}$ and vestibular sensitivity $(\lambda \sim 0.76-0.79)$.

## Discussion

Our results reveal that the canonical model of semicircular canal orientation is not strictly correct for a wide range of mammalian species. In our comparative sample, angles between
pairs of ipsilateral semicircular canals deviate from orthogonality by an average of $6.0^{\circ}$, corresponding left-right canal pair angles differ by an average of $4.3^{\circ}$, and synergistic canals deviate from coplanarity by an average of $10.1^{\circ}$. Although the angle between any two ipsilateral semicircular canals does approach $90^{\circ}$ when data are averaged for all canal pairs across all taxa, it is nevertheless clear that all mammals deviate from canal orthogonality to some degree. For example, Potorous, Pedetes, and Wallabia all have ipsilateral canal pair angles that diverge from $90^{\circ}$ by an average of between only $2^{\circ}$ and $3^{\circ}$. By contrast, Notoryctes, Chrysochloris, and Allactaga all have ipsilateral canal pair angles that diverge from $90^{\circ}$ by an average of nearly $10^{\circ}$ or more. These data further underscore the fact that none of the taxa considered in this analysis has truly orthogonal semicircular canals, and that substantial deviations from orthogonality, symmetry, and coplanarity appear to be a common feature of vestibular anatomy in mammals. However, it is also important to acknowledge that our comparative sample does not address questions related to intraspecific variation in canal orientation, so it is currently unclear how representative the values reported here are for each species in our dataset (Tables 1, 2). Our results also demonstrate that the mean angle between the anterior and lateral canals is considerably less than $90^{\circ}$ and is $6.4^{\circ}-7.4^{\circ}$ lower than the mean angle between the other two ipsilateral canal pairs. In a functional


Figure 4. Regressions of mean estimated semicircular canal sensitivity (Sensitivity ${ }_{\text {ave }}$ ) on $\mathbf{L o g}_{10} \mathbf{9 0}_{\text {var }}$. doi:10.1371/journal.pone.0079585.g004
context, is not presently clear why the angle between ipsilateral anterior and lateral canals is systematically lower than that for other ipsilateral canal pairs, but this finding further reinforces the inaccuracy of general characterizations of semicircular canals as orthogonal [3-16].

Previously published studies of semicircular canal morphology in 43 living and fossil species support our general finding that substantial deviations from the canonical model are the norm for mammals (Table 6). When unilateral measurements are considered, these analyses document a range of ipsilateral canal pair angles between $52.2^{\circ}$ and $117.0^{\circ}$ and an average $90_{\text {var }}$ for all ipsilateral canal pairs of $7.9^{\circ}$ (st. dev. $=4.4^{\circ}$ ). As in the species we examined, the published data also show that across taxa the angle between the anterior and lateral canals (ASC $\Varangle \mathrm{LSC}$ mean $=82.6^{\circ}$, st. dev. $\left.=11.3^{\circ}\right)$ is considerably smaller than the angle between the other two ipsilateral canal pairs (ASC $\Varangle$ PSC mean $=93.6^{\circ}$, st. dev. $9.7^{\circ} ; \operatorname{LSC} \Varangle$ PSC mean $=91.4^{\circ}$, st. dev. $\left.=7.4^{\circ}\right)$. The published taxa also show a mean deviation from coplanarity of $16.9^{\circ}$ (st. dev. $=6.7^{\circ}$ ) for both ASC $\Varangle$ PSC pairs and $13.3^{\circ}$ (st. dev. $=6.9^{\circ}$ ) for the LSC $\Varangle$ LSC pair. The mean deviation of all three synergistic canal pair angles from coplanarity is $15.8^{\circ}$ (st. $\mathrm{dev} .=6.9^{\circ}$.
From a practical standpoint, these data have important implications for the use of lateral canal orientation as an indicator of the horizontal plane in reconstructions of head posture in fossil mammals [31,39,51-53]. In our comparative sample, 7 of the 39 species have right and left lateral canals that deviate from coplanarity by more than $20^{\circ}$ (Table 2). This large amount of bilateral variation in lateral canal orientation within individual specimens suggests that the lateral canal is an imprecise indicator of habitual head orientation (resting or active) in fossil species,
particularly if reconstructions are based on unilateral measurements of semicircular canals [54].

Our data also generally confirm the expectation that there are important functional consequences of the degree to which a species' vestibular anatomy deviates from the canonical model. In particular, deviations from canal orthogonality (as measured by $90_{\mathrm{var}}$ ) are negatively correlated with both of our estimates of vestibular sensitivity (Table 5). This result is most pronounced for the relationship between $90_{\text {var }}$ and mean sensitivity (Sensitivity ${ }_{\text {ave }}$, Figure 4). As noted previously, the values for Sensitivity ${ }_{\text {ave }}$ reported here are determined entirely by canal radii of curvature (Figure 2), so the significant negative relationship between $90_{\text {var }}$ and Sensitivity ${ }_{\text {ave }}$ is unrelated to our methods for estimating canal sensitivity. By the same token, deviations from orthogonality tend to increase the maximum vestibular sensitivity (i.e., result in higher Senstivity $_{\text {max }}$ : OSensitivity max $^{\text {ratio; Figure 2) according to the }}$ methods employed here. In other words, based on our estimates of canal sensitivity, constraining canals to be perfectly orthogonal always decreases Senstivity ${ }_{\text {max }}$ (Figure 2, Table 4). Accordingly, our results for $90_{\text {var }}$ and Sensitivity max $_{\text {max }}$ (Table 5) should be interpreted with caution because estimated maximum vestibular sensitivity is determined both by the radii and orientations of canals. Nevertheless, our analysis demonstrates that as the average deviation from canal orthogonality in our interspecific comparative sample increases (i.e., higher $90_{\text {var }}$ ), the mean estimated sensitivity to angular head accelerations tends to decrease. As a result, species with more orthogonal semicircular canals tend to have higher mean vestibular sensitivity than species with less orthogonal semicircular canals (Figure 4). Although canal radius of curvature remains a major determinant of semicircular canal sensitivity, these findings imply that selection for greater sensitivity
Table 6. Previous semicircular canal pair angle research results.

| Taxon | LASC ŁLLSC IPS | LASC ŁLPSC IPS | LLSC $\Varangle$ LPSC IPS | RASC ŁRLSC IPS | RASC $\Varangle$ RPSC IPS | RLSC $\Varangle$ RPSC IPS | LASC $\Varangle$ RASC CON | LPSC $\Varangle$ RPSC CON | LASC $\Varangle$ RPSC SYN | RASC ¢LPSC SYN | LLSC $\Varangle$ RLSC SYN | n | Sources and Notes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Atelerix albiventris | 82.20 | 91.70 | 92.10 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Balaenopteridae $\dagger$ | 71.60 | 105.00 | 75.60 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Bathygenys reevesi† | 86.00 | 99.60 | 91.30 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Canis familiaris | 80.40 | 101.00 | 89.10 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Cavia porcellus | 57.85 | 76.71 | 82.36 | 57.85 | 76.71 | 82.36 |  |  | 32.17 | 36.16 | 30.82 | 10 | Curthoys et al (1975)*[1]a |
|  | $\pm 6.13$ | $\pm 5.49$ | $\pm 4.74$ | $\pm 6.13$ | $\pm 5.49$ | $\pm 4.74$ |  |  | $\pm 4.42$ | $\pm 4.86$ | $\pm 10.05$ |  |  |
|  | 10.6 | 7.16 | 5.76 | 10.60 | 7.16 | 5.76 |  |  | 13.74 | 13.44 | 32.61 |  |  |
| Cavia porcellus |  |  |  | 91.22 | 91.20 | 85.88 |  |  |  |  |  | 8 | Cox and |
|  |  |  |  | $\pm 6.97$ | $\pm 5.78$ | $\pm 5.3$ |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 7.64 | 6.34 | 6.17 |  |  |  |  |  |  | (2008) [2]a |
| Cavia porcellus | 77.20 | 105.00 | 85.50 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Chinchilla | 76.30 | 90.90 | 103.00 | 78.70 | 91.00 | 101.00 |  |  | 15.60 | 14.80 | 14.60 | 3 | Hullar and Williams (2006)*a |
| lanigera | $\pm 2.7$ | $\pm 1.7$ | $\pm 5.7$ | $\pm 5.7$ | $\pm 2.3$ | $\pm 6.9$ |  |  | $\pm 1.3$ | $\pm 0.3$ | $\pm 12$ |  |  |
|  | 3.54 | 1.87 | 5.53 | 7.24 | 2.53 | 6.83 |  |  | 8.33 | 2.03 | 82.19 |  |  |
| Chrysochloris sp. | 65.60 | 86.90 | 96.70 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Cynocephalus volans | 92.20 | 90.00 | 91.80 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Dasypus novemcinctus | 62.40 | 67.70 | 87.30 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Didelphis virginiana | 109.00 | 102.00 | 104.00 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Elephas $\dagger$ | 66.30 | 73.70 | 96.70 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Equus caballus | 84.70 | 93.30 | 90.10 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Eumetopias jubatus | 79.70 | 105.00 | 90.60 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Felis catus | 95.60 | 87.90 | 94.10 | 95.60 | 87.90 | 94.10 | 81.10 | 103.80 | 13.90 | 13.90 | 14.40 | 3 | Ezure \& Graf (1984) <br> [1]a |
| Felis catus | 89.62 | 90.21 | 94.23 | 89.62 | 90.21 | 94.23 |  |  | 13.92 | 14.49 | 12.49 | 7 | Blanks et al. |
|  | $\pm 8.71$ | $\pm 4.05$ | $\pm 3.84$ | $\pm 8.71$ | $\pm 4.05$ | $\pm 3.84$ |  |  | $\pm 3.99$ | $\pm 4.52$ | $\pm 9.21$ |  | (1972) [1]a |
|  | 9.72 | 4.49 | 4.08 | 9.72 | 4.49 | 4.08 |  |  | 28.66 | 31.19 | 73.74 |  |  |
| Felis catus |  |  |  | 78.36 | 103.34 | 89.49 |  |  |  |  |  | 8 | Cox and |
|  |  |  |  | $\pm 10.05$ | $\pm 9.7$ | $\pm 6.94$ |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 12.83 | 9.39 | 7.76 |  |  |  |  |  |  | (2008) [2]a |
| Felis catus | 76.80 | 91.40 | 96.70 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Hemicentetes semispinosum | 85.40 | 117.00 | 92.60 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Homo sapiens | 90.60 | 94.40 | 90.40 | 90.60 | 94.40 | 90.40 | 103.40 | 83.20 | 15.30 | 15.30 | 11.30 | 10 | Della |

Table 6. Cont.

| Taxon | LASC xLLSC IPS | LASC ŁLPSC IPS | LLSC ŁLPSC IPS | RASC <br> $\Varangle$ RLSC <br> IPS | RASC $\Varangle$ RPSC IPS | RLSC $\Varangle$ RPSC IPS | LASC <br> $\Varangle$ RASC <br> CON | LPSC $\Varangle$ RPSC CON | LASC $\Varangle$ RPSC SYN | RASC <br> \& LPSC <br> SYN | LLSC <br> $\Varangle$ RLSC <br> SYN | n | Sources and Notes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\pm 6.2$ | $\pm 1.85$ | $\pm 4.9$ | $\pm 6.2$ | $\pm 4$ | $\pm 4.9$ | $\pm 9.5$ | $\pm 9.7$ | $\pm 7.2$ | $\pm 7.2$ | $\pm 6.9$ |  | Santina et |
|  | 6.84 | 2.02 | 5.42 | 6.84 | 4.24 | 5.42 | 9.19 | 11.66 | 47.06 | 47.06 | 61.06 |  | al. (2005) |
|  |  |  |  |  |  |  |  |  |  |  |  |  | ${ }^{\text {[1]c }}$ |
| Homo sapiens | 68.24 | 86.16 | 95.75 | 68.24 | 86.16 | 95.75 |  |  | 24.56 | 23.73 | 19.82 | 10 | Blanks et al. |
|  | $\pm 7.55$ | $\pm 4.72$ | $\pm 4.66$ | $\pm 7.55$ | $\pm 4.72$ | $\pm 4.66$ |  |  | $\pm 7.19$ | $\pm 6.71$ | $\pm 14.93$ |  | (1975)* [1]a |
|  | 11.06 | 5.48 | 4.87 | 11.06 | 5.48 | 4.87 |  |  | 29.28 | 28.28 | 75.33 |  |  |
| Homo sapiens | 90.50 | 91.70 | 94.52 | 90.50 | 91.70 | 94.52 |  |  |  |  |  | 7 | Hashimoto |
|  | $\pm 2.98$ | $\pm 1.85$ | $\pm 3.32$ | $\pm 2.98$ | $\pm 1.85$ | $\pm 3.32$ |  |  |  |  |  |  | et al. 2005a |
|  | 3.29 | 2.02 | 3.51 | 3.29 | 2.02 | 3.51 |  |  |  |  |  |  |  |
| Homo sapiens |  |  |  | 85.30 | 97.14 | 88.96 |  |  |  |  |  | 6 | Cox and |
|  |  |  |  | $\pm 5.81$ | $\pm 4.82$ | $\pm 6.33$ |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 6.81 | 4.96 | 7.12 |  |  |  |  |  |  | (2008) [2]a |
| Homo sapiens | 98.90 | 100.00 | 89.80 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Kulbeckia kulbecke ${ }^{\dagger}$ | 79.90 | 79.90 | 89.60 |  |  |  |  |  |  |  |  | 4 | Ekdale (2009) [1] |
| Lepus californicus | 84.20 | 94.00 | 88.60 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Macaca mulatta | 98.73 | 86.48 | 88.49 | 98.73 | 86.48 | 88.49 |  |  | 10.67 | 11.18 | 2.24 | 10 | Blanks et al. |
|  | $\pm 5.39$ | $\pm 3.43$ | $\pm 3.91$ | $\pm 5.39$ | $\pm 3.43$ | $\pm 3.91$ |  |  | $\pm 3.55$ | $\pm 3.12$ | $\pm 0.77$ |  | (1985) [1]a |
|  | 5.46 | 3.97 | 4.42 | 5.46 | 3.97 | 4.42 |  |  | 33.27 | 27.91 | 34.38 |  |  |
| Macaca mulatta | 83.10 | 100.00 | 89.00 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Macroscelides proboscideus | 100.00 | 90.70 | 73.50 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Manis tricuspis | 77.00 | 84.80 | 88.60 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Mus C57BL/6J | 92.56 | 99.02 | 101.17 | 91.55 | 99.26 | 101.26 |  |  | 17.61 | 17.66 | 9.14 | 4 | Calabrese |
|  | $\pm 1.93$ | $\pm 1.46$ | $\pm 0.97$ | $\pm 1.02$ | $\pm 1.29$ | $\pm 0.96$ |  |  | $\pm 2.73$ | $\pm 1.43$ | $\pm 0.98$ |  | and Hullar |
|  | 2.09 | 1.47 | 0.96 | 1.11 | 1.30 | 0.95 |  |  | 15.50 | 8.10 | 10.72 |  | (2006)a |
| Mus CBA/CaJ | 96.82 | 89.65 | 102.29 | 95.47 | 88.94 | 102.05 |  |  | 11.00 | 14.79 | 10.42 | 4 | Calabrese |
|  | $\pm 5.73$ | $\pm 2.51$ | $\pm 1.86$ | $\pm 1.94$ | $\pm 1.98$ | $\pm 2.14$ |  |  | $\pm 1.24$ | $\pm 2.11$ | $\pm 3.8$ |  | and Hullar |
|  | 5.92 | 2.80 | 1.82 | 2.03 | 2.23 | 2.10 |  |  | 11.27 | 14.27 | 36.47 |  | (2006)a |
| Mus musculus |  |  |  | 76.63 | 101.54 | 96.08 |  |  |  |  |  | 9 | Cox and |
|  |  |  |  | $\pm 6.02$ | $\pm 6.32$ | $\pm 6.06$ |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 7.86 | 6.22 | 6.31 |  |  |  |  |  |  | (2008) [2]a |
| Mus musculus | 88.80 | 94.40 | 95.60 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Nycteris grandis | 85.90 | 112.00 | 94.90 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Nycticebus coucang |  | 88.60 |  |  | 88.60 |  | 68.20 | 114.40 | 23.10 | 23.10 |  | 3 | Matano et al. (1985) [1] |

Table 6. Cont.

| Taxon | LASC \& LLSC IPS | LASC «LPSC IPS | LLSC ŁLPSC IPS | RASC $\Varangle$ RLSC IPS | RASC $\Varangle$ RPSC IPS | RLSC $\Varangle$ RPSC IPS | LASC $\Varangle$ RASC CON | LPSC $\Varangle$ RPSC CON | LASC $\Varangle$ RPSC SYN | RASC <br> ŁLPSC <br> SYN | LLSC <br> $\Varangle$ RLSC <br> SYN | n | Sources and Notes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Orycteropus afer | 78.50 | 91.90 | 95.70 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Oryctolagus cuniculus | 79.80 | 76.60 | 75.50 | 79.80 | 76.60 | 75.50 | 90.50 | 116.50 | 13.60 | 13.60 | 8.60 | 3 | Ezure \& Graf (1984) <br> [1]a |
| Oryctolagus | 79.36 | 71.36 | 75.85 | 79.36 | 71.36 | 75.85 | 85.76 | 47.54 | 26.78 | 26.78 | 15.32 | 7 | Mazza and |
| cuniculus | $\pm 9.4$ | $\pm 4.4$ | $\pm 6.7$ | $\pm 9.4$ | $\pm 4.4$ | $\pm 6.7$ | $\pm 5.6$ | $\pm 5.3$ | $\pm 6.8$ | $\pm 6.8$ | $\pm 7.2$ |  | Winterson |
|  | 11.84 | 6.17 | 8.83 | 11.84 | 6.17 | 8.83 | 6.53 | 11.15 | 25.39 | 25.39 | 47.00 |  | (1984)* [1]a |
| Oryctolagus |  |  |  | 81.73 | 97.05 | 97.52 |  |  |  |  |  | 9 | Cox and |
| cuniculus |  |  |  | $\pm 11$ | $\pm 5.6$ | $\pm 9.82$ |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 13.46 | 5.77 | 10.07 |  |  |  |  |  |  | (2008) [2]a |
| Procavia capensis | 87.40 | 112.00 | 87.40 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Pteropus lyelli | 84.90 | 98.30 | 90.40 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Rattus norvegicus |  |  |  | 73.35 | 97.57 | 98.12 |  |  |  |  |  | 8 | Cox and |
|  |  |  |  | 6.37 | 4.80 | 10.89 |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 8.68 | 4.92 | 11.10 |  |  |  |  |  |  | (2008) [2]a |
| Rattus norvegicus | 97.60 | 94.20 | 93.50 | 97.60 | 94.20 | 93.50 |  |  | 9.90 | 9.90 | 8.00 | 14 | Blanks and Torigoe (1989) [1]b |
| Rhinolophus ferrumequinum | 79.90 | 104.00 | 87.90 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Saimiri sciureus | 90.43 | 87.02 | 89.95 | 90.43 | 87.02 | 89.95 |  |  | 12.53 | 14.80 | 15.45 | 10 | Blanks et al. |
|  | $\pm 6.94$ | $\pm 4.22$ | $\pm 5.08$ | $\pm 6.94$ | $\pm 4.22$ | $\pm 5.08$ |  |  | $\pm 5.55$ | $\pm 5.37$ | $\pm 5.98$ |  | (1985) [1]a |
|  | 7.67 | 4.85 | 5.65 | 7.67 | 4.85 | 5.65 |  |  | 44.29 | 36.28 | 38.71 |  |  |
| Sciurus |  |  |  | 78.97 | 89.52 | 104.41 |  |  |  |  |  | 5 | Cox and |
| carolinensis |  |  |  | $\pm 6.61$ | $\pm 4.4$ | $\pm 9.64$ |  |  |  |  |  |  | Jeffery |
|  |  |  |  | 8.37 | 4.92 | 9.23 |  |  |  |  |  |  | (2008) [2]a |
| Sorex monticolus | 75.30 | 89.60 | 89.30 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Sus scrofa | 82.80 | 96.00 | 87.90 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Sylvilagus floridanus | 92.70 | 97.50 | 77.90 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Tadarida brasiliensis | 74.70 | 98.40 | 98.40 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Tarsius bancanus |  | 91.80 |  |  | 91.80 |  | 73.80 | 102.80 | 14.50 | 14.50 |  | 3 | Matano et al. (1985) [1] |
| Trichechus manatus | 52.20 | 84.90 | 86.30 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Tupaia glis | 82.30 | 106.00 | 102.00 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Tursiops truncatus | 52.20 | 84.90 | 77.50 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Ukhaatherium gobiensis ${ }^{\dagger}$ | ${ }^{\dagger} 88.80$ | 105.00 | 88.40 |  |  |  |  |  |  |  |  | 1 | Ekdale (2009) [3]c |
| Zalambdalestes lecheit | 81.00 | 93.60 | 85.60 |  |  |  |  |  |  |  |  | 4 | Ekdale (2009) [3]c |
| Zhelestid ${ }^{\dagger}$ | 88.80 | 96.80 | 93.10 |  |  |  |  |  |  |  |  | 7 | Ekdale (2009) [3]c |

Table 6. Cont.

| Taxon | LASC xLLSC IPS | LASC ŁLPSC IPS | LLSC xLPSC IPS | RASC $\Varangle$ RLSC IPS | RASC $\Varangle$ RPSC IPS | $\begin{aligned} & \text { RLSC } \\ & \Varangle \text { RPSC } \end{aligned}$ IPS | LASC ŁRASC CON | $\begin{aligned} & \text { LPSC } \\ & \times \text { RPSC } \end{aligned}$ CON | LASC $\Varangle$ RPSC SYN | RASC ŁLPSC SYN | $\begin{aligned} & \text { LLSC } \\ & \Varangle \text { RLSC } \end{aligned}$ SYN | n | Sources and Notes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Mean | 82.46 | 93.21 | 90.37 | 84.27 | 90.42 | 92.35 | 83.79 | 94.71 | 17.01 | 17.65 | 13.28 |  |  |
| Standard Dev. | $\pm 11.87$ | $\pm 10.14$ | $\pm 7.19$ | $\pm 10.41$ | $\pm 7.82$ | $\pm 7.85$ | $\pm 12.52$ | $\pm 25.95$ | $\pm 6.59$ | $\pm 6.94$ | $\pm 6.85$ |  |  |
| Coeff. Variation | 14.39 | 10.88 | 7.96 | 12.35 | 8.64 | 8.50 | 14.94 | 27.40 | 38.77 | 39.35 | 51.63 |  |  |
| 95\% Confidence | 80.78 | 91.80 | 89.35 | 82.00 | 88.79 | 90.64 | 78.68 | 84.11 | 15.31 | 15.85 | 11.38 |  |  |
| Interval of Mean | 84.13 | 94.61 | 91.39 | 86.54 | 92.05 | 94.07 | 88.90 | 105.30 | 18.71 | 19.44 | 15.88 |  |  |

 different species reported. Abbreviations for notes: [1] sides averaged, [2] MRI study based on one side only, [3] mostly left labyrinths used, but specimens with right sides indeterminate, CON contralateral canals, IPS ipsilateral canals, $\mathbf{n}$ number of specimens examined, SYN synergistic canals, ${ }^{*}$ ASC $\Varangle$ LSC corrected for internal angles. doi:10.1371/journal.pone.0079585.t006
to angular head accelerations may influence semicircular canal orthogonality.

We also find that deviations from canal angle symmetry and coplanarity are negatively correlated with estimated vestibular sensitivity. However, these correlations are weaker and more strongly influenced by phylogeny compared to the results for orthogonality. As a result, phylogenetically controlled analyses of the relationship between estimated vestibular sensitivity and both angle symmetry and coplanarity are not significant (Table 5). These results do not necessarily imply the absence of a functional relationship between canal angle symmetry or coplanarity and vestibular sensitivity, but they do indicate that there is a strong phylogenetically correlated influence on these relationships.

In this context, it is also noteworthy that the obligate fossorial genera in our analysis (Notoryctes, Chrysochloris, Talpa, and Heterocephalus) show greater deviations from canal orthogonality than most non-fossorial genera (Table 2). Indeed, the average $90_{\text {var }}$ value for the 4 fossorial taxa in our sample (mean $=11.0^{\circ}$; st. dev. $=3.2^{\circ}$ ) is twice that of non-fossorial taxa (mean $=5.5^{\circ}$; st. dev. $=1.8^{\circ}$ ). Notoryctes, Chrysochloris, Talpa, and Heterocephalus also share comparatively low estimates of semicircular canal sensitivity (fossorial taxa: Sensitivity ${ }_{\text {ave }}=0.27-0.48 ;$ Sensitivity $_{\max }=0.38-$ 0.59; non-fossorial taxa: Sensitivity ${ }_{\text {ave }}$ mean $=0.89$, st. dev. $=0.38 ;$ Sensitivity $\max ^{\operatorname{mean}}=1.10$, st. dev. $=0.47$; Table 4). While this sample of fossorial genera is small, it is also taxonomically diverse, including a marsupial (Notoryctes), an afrothere (Chrysochloris), a eulipotyphlan (Talpa), and a rodent (Heterocephalus). These data therefore suggest that low degrees of semicircular canal orthogonality and relatively low sensitivity to angular accelerations may have evolved concurrently with a fossorial lifestyle at least 4 times in mammals. Nonetheless, it is not functionally clear why lower degrees of canal orthogonality would be associated with a burrowing lifestyle.

Our findings are consistent with those of Billet et al. [55], who report highly variable and non-orthogonal ipsilateral canal pair angles in three-toed sloths (Bradypus variegatus). Billet et al. suggested that such high variability is the result of diminished selection pressure in slower-moving mammals to maintain orthogonal semicircular canals. Furthermore, the negative relationship between angular head velocities and $90_{\text {var }}$ observed by Malinzak et al. [33] accords well with our finding of a negative relationship between mean vestibular sensitivity and $90_{\text {var }}$ (Figure 4). The combined results of both studies thus show that species with the greatest deviations from canal orthogonality tend to experience slower head rotations during locomotion [33] and to have less sensitive semicircular canals (Figure 4). Although Malinzak et al. [33] based their conclusions on a smaller sample of 11 primate species, their analysis is the only comparative study to date that directly measured angular head velocities produced during locomotion. These authors further concluded that species which regularly encounter higher angular head accelerations during locomotion require more orthogonal canals in order to have more uniform sensitivity to angular accelerations in three dimensions. Here we have shown that the degree to which semicircular canals approach orthogonality is correlated with mean estimated sensitivity to angular accelerations, and that mean sensitivity in turn is solely determined by canal radius of curvature. These findings reinforce the conclusion that both the radii and orientations of the semicircular canals may be influenced by selection related to forces generated during locomotion.

## Acknowledgments

This research was greatly facilitated by Chris Bell, James Sprinkle and Larry Witmer who reviewed it in dissertation form. Special thanks are due
to Tim Hullar, who helped immensely in providing and implementing Bubbles.mat, and for conversations describing calculation of sensitivity. We thank editor Andrew Farke and reviewers Justin Georgi and Rich Kay for their assistance in revising the first submission of this manuscript. Thanks also to Charlie Nunn, Denné Reed, and Sally Amen for statistical advice. Scanning help was graciously provided by Jessie Maisano, Matt Colbert, Rich Ketcham and the staff of the UTCT lab. Mark Coleman, Tim Hullar, Ted Macrini, and Blaire Van Valkenburgh gave permission to use

## References

1. Hullar TE (2006) Semicircular canal geometry, afferent sensitivity, and animal behavior. Anatomical Record Part A 288A: 466-472.
2. Graf W (1988) Motion detection in physical space and its peripheral and central representation. Annals of the New York Academy of Sciences 545: 154-169.
3. Agur AM (1999) Grant's Atlas of Anatomy. Baltimore, MD: Lippincott, Williams and Wilkins. 760 p.
4. Gray H (1918) Anatomy of the Human Body. Philadelphia, PA: Lea \& Febiger. 1396 p.
5. Gulya AJ (2007) Gulya and Schuknecht's Anatomy of the Temporal Bone with Surgical Implications. New York, NY: Informa Healthcare. 356 p.
6. Clarke AH (2005) On the vestibular labyrinth of Brachiosaurus brancai. Journal of Vestibular Research 15: 65-71.
7. Kaas J (2009) Evolutionary Neuroscience. Oxford, UK: Academic Press. 1038 p.
8. Marugán-Lobón J, Chiappe LM, Farke AA (2013) The variability of inner ear orientation in saurischian dinosaur: testing the use of semicircular canals as a reference system for comparative anatomy. PeerJ 1: 16 pp .
9. Romer AS, Parsons TS (1986) The Vertebrate Body. Saunders: Philadelphia. 679 p.
10. Spoor F, Zonneveld F (1995) Morphometry of the primate bony labyrinth: a new method based on high-resolution computed tomography. Journal of Anatomy 186: 271-286.
11. Oman CM, Marcus EN, Curthoys IS (1987) The influence of semicircular canal morphology on endolymph flow dynamics. Acta Otolaryngolica 103: 1-13.
12. Spoor F (2003) The semicircular canal system and locomotor behaviour, with special reference to hominin evolution. Courier Forschungsinstitut Senckenberg 243: 93-104.
13. Haines DE (2012) Fundamental Neuroscience for Basic and Clinical Applications. Philadelphia: Elsevier. 492 p.
14. Lessard C (2009) Basic Feedback Controls in Biomedicine; Enderle JD, editor. Connecticut: Morgan and Claypool Publishers. 209 p.
15. Highsteen SM, Rabbitt RD, Holstein GR, Boyle RD (2005) Determinants of spatial and temporal coding by semicircular canal afferents. Journal of Neurophysiology 93: 2359-2370.
16. Rabbitt RD, Damiano ER, Grant JW (2004) Biomechanics of the vestibular semicircular canals and otolith organs. In: Highstein SM, Popper A, Fay R, editors. The Vestibular System. Berlin, Germany: Springer-Verlag. pp. 153201.
17. Blanks RHI, Curthoys IS, Markham CH (1975) Planar relationships of the semicircular canals in man. Acta Otolaryngolica 80: 185-196.
18. Curthoys IS, Curthoys EJ, Blanks RHI, Markham CH (1975) The orientation of the semicircular canals in the guinea pig. Acta Otolaryngolica 80: 197-205.
19. Ezure K, Graf W (1984) A quantitative analysis of the spatial organization of the vestibulo-ocular reflexes in lateral- and frontal-eyed animals - I. Orientation of semicircular canals and extraocular muscles. Neuroscience 12: 85-93.
20. Mazza D, Winterson BJ (1984) Semicircular canal orientation in the adult resting rabbit. Acta Otolaryngolica 98: 472-480.
21. Blanks RHI, Torigoe Y (1989) Orientation of the semicircular canals in the rat. Brain Research 487: 278-287.
22. Calabrese DR, Hullar TE (2006) Planar relationships of the semicircular canals in two strains of mice. Journal of the Association for Research in Otolaryngology 7: 151-159.
23. Ekdale EG (2009) Variation within the bony labyrinth of mammals PhD Dissertation]. Austin: The University of Texas. 439 p.
24. Ifediba MA, Rajguru SM, Hullar TE, Rabbitt RD (2007) The role of 3-canal biomechanics in angular motion transduction by the human vestibular labyrinth. Annals of Biomedical Engineering 35: 1247-1263.
25. Yang A, Hullar TE (2007) Relationship of semicircular canal size to vestibularnerve afferent sensitivity in mammals. Journal of Neurophysiology 98: 31973205.
26. Rabbitt RD (1999) Directional coding of three-dimensional movements by the vestibular semicircular canals. Biological Cybernetics 80: 417-431.
27. Jones GM, Spells KE (1963) A theoretical and comparative study of the functional dependence of the semicircular canal upon its physical dimensions. Proceedings of the Royal Society 157: 403-419.
28. Silcox MT, Bloch JI, Boyer DM, Godinot M, Ryan TM, et al. (2009) Semicircular canal system in early primates. Journal of Human Evolution 56: 315-327.
29. Spoor F, Garland T, Krovitz G, Ryan TM, Silcox MT, et al. (2007) The primate semicircular canal system and locomotion. Proceedings of the National Academy of Science of the United States of America 104: 10808-10812.
high resolution X-ray computed tomographic scans of their specimens for this study.

## Author Contributions

Conceived and designed the experiments: JCB ECK TBR. Performed the experiments: JCB. Analyzed the data: JCB ECK. Contributed reagents/ materials/analysis tools: TBR. Wrote the paper: JCB ECK TBR.
30. ten Kate JH, van Barneveld HH, Kuiper JW (1970) The dimensions and sensitivities of semicircular canals. Journal of Experimental Biology 53: 501-514.
31. Spoor F, Wood BA, Zonneveld F (1994) Implications of early hominid labyrinthine morphology for evolution of human bipedal locomotion. Nature 369: 645-648.
32. Rodgers J (2011) Comparative morphology of the metatherian and eutherian bony labyrinth. Austin: University of Texas at Austin. 400 p.
33. Malinzak MD (2010) Experimental analyses of the relationship between semicircular canal morphology and locomotor head rotations in primates [Dissertation]. Durham, North Carolina: Duke University. 236 p.
34. Malinzak M, Kay RF, Hullar TE (2011) Semicircular canal orthogonality, not radius, best predicts mean speed of locomotor head rotation: a new hypothesis with implications for reconstructing behaviors in extinct species. American Journal of Physical Anthropology 144: 204.
35. Malinzak MD, Kay RF, Hullar TE (2012) Locomotor head movements and semicircular canal morphology in primates. Proceedings of the National Academy of Science 109: 17914-17919.
36. Rowe T, Kappelman J, Carlson WD, Ketcham RA, Denison C (1997) HighResolution Computed Tomography: a breakthrough technology for Earth scientists. Geotimes 42: 23-27.
37. Carlson WD, Rowe T, Ketcham RA, Colbert MW (2003) Geological applications of high-resolution X-ray computed tomography in petrology, meteoritics and paleontology. In: Mees F, Swennen R, Van Geet M, Jacobs P, editors. Applications of X-ray computed tomography in the geosciences. London: Geological Society. pp. 7-22.
38. Romer AS (1962) The Vertebrate Body. Philadelphia, Pennsylvania: W. B. Saunders Company. 627 p.
39. David R, Droulez J, Allain R, Berthoz A, Janvier P, et al. (2010) Motion from the past. A new method to infer vestibular capacities of extinct species. Comptes Rendus Palevol 9: 397-410.
40. Venes D, editor (2005) Taber's Cyclopedic Medical Dictionary, 20th Edition. 20 ed. Philadelphia, Pennsylvania: F. A. Davis Company. 2788 p.
41. Cox PG, Jeffery N (2008) Geometry of the semicircular canals and extraocular muscles in rodents, lagomorphs, felids, and modern humans. Journal of Anatomy 213: 583-596.
42. Spoor F, Zonneveld F (1998) Comparative review of the human bony labyrinth. Yearbook of Physical Anthropology 41: 211-251.
43. Fisher NI (1995) Statistical analysis of circular data. Melbourne, Australia: University of Cambridge. 277 p.
44. Paradis E, Claude J, Strimmer K (2004) APE: analyses of phylogenetics and evolution in R language. Bioinformatics 20: 289-290.
45. R-Development-Core-Team (2011) R: A language and environment for statistical computing. Vienna, Austria.
46. Pinheiro J, Bates D, DebRoy S, Sarkar D, Team RDC (2011) nlme: Linear and Nonlinear Mixed Effects Models. R package version 3:1-101.
47. Freckelton RP, Harvey PH, Pagel M (2002) Phylogenetic analysis and comparative data: a test and review of evidence. American Naturalist 160: 712-726.
48. Bininda-Emonds ORP, Cardillo M, Jones KE, MacPhee RDE, Beck RMD, et al. (2007) The delayed rise of present-day mammals. Nature 446: 507-512.
49. Bininda-Emonds ORP, Cardillo M, Jones KE, MacPhee RDE, Beck RMD, et al. (2008) Corrigendum. The delayed rise of present-day mammals. Nature 256: 274.
50. Pagel M (1999) Inferring the historical patterns of biological evolution. Nature 401: 877-844.
51. Ryan TM, Silcox MT, Walker A, Mao X, Begun DR, et al. (2012) Evolution of locomotion in Anthropoidea: the semicircular canal evidence. Proceedings of the Royal Society B 279: 3467-3475.
52. Sereno PC, Wilson JA, Witmer LM, Whitlock JA, Maga A, et al. (2007) Structural extremes in a Cretaceous dinosaur. PLoS ONE 11: 1-9.
53. Witmer LM, Chatterjee S, Franzosa J, Rowe T (2003) Neuroanatomy of flying reptiles and implications for flight, posture and behaviour. Nature 425: 950-953.
54. Davies KTJ, Bates PJJ, Maryanto I, Cotton JA, Rossiter SJ (2013) The evolution of bat vestibular systems in the face of potential antagonistic selection pressures for flight and echolocation. PLoS One 8: e61998.
55. Billet G, Hautier L, Asher RJ, Schwarz C, Crumpton N, et al. (2012) High morphological variation of vestibular system accompanies slow and infrequent locomotion in three-toed sloths. Proceedings of the Royal Society B 279: 29322939.


[^0]:     doi:10.1371/journal.pone.0079585.t002

[^1]:    P-values for significant results and non-significant trends shown in bold; Results significant at $\mathrm{P}<0.05$ marked with an asterisk. "Pearson" $=$ one-tailed Pearson productmoment correlation, "PGLS" = phylogenetic generalized least-squares regression.
    doi:10.1371/journal.pone.0079585.t005

