# Comparison of the Efficacy and Safety of Cell-Assisted Lipotransfer and Platelet-Rich Plasma Assisted Lipotransfer: What Should We Expect from a Systematic Review with Meta-Analysis?

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### Abstract

Due to the high absorption rate of traditional autologous fat grafting, cell-assisted lipotransfer (CAL) and platelet-rich plasma (PRP)-assisted lipotransfer were developed. The purpose of this article was to evaluate the efficacy and safety of CAL and PRP in promoting the survival of autologous fat grafting through systematic review and meta-analysis. We searched Pubmed, Cochrane Library, Web of Science, and EMBASE for clinical studies on CAL and PRP-assisted lipotransfer published from January 2010 to January 2020. Then a meta-analysis was performed to assess the efficacy of CAL and PRP-assisted lipotransfer through data analysis of fat survival rate. We also assessed the incidence of complications and multiple operations to analyze their safety. A total of 36 studies (1697 patients) were included in this review. Regardless of the recipient area, CAL and PRP-assisted lipotransfer significantly improved the fat survival rate (CAL vs non-CAL: 71% vs 48%, P < 0.0001; PRP vs non-PRP: 70% vs 40%, P < 0.0001; CAL vs PRP: 71% vs 70%, P = 0.7175). However, in large-volume fat grafting, such as breast reconstruction, both increased the incidence of complications and did not decrease the frequency of multiple operations after lipotransfer. Further prospective studies are needed to evaluate the clinical benefits of CAL and PRP-assisted lipotransfer.

### **Keywords**

cell-assisted lipotransfer, platelet-rich plasma, stromal vascular fraction, autologous fat grafting, meta-analysis

### Introduction

Autologous fat grafting is a revolution in the field of soft tissue reconstruction and augmentation, and is mainly used to fill up the congenital deficiencies and soft tissue defects in plastic and reconstructive surgery, such as: filling of the face, breast, and buttocks, wound repair and breast reconstruction after breast cancer, etc.<sup>1</sup> Compared with other filling materials, autologous fat grafting has the advantages of good biocompatibility, easy access, abundant sources, good filling effect, and small trauma. So it is recognized as a safe and effective method<sup>2,3</sup>. Traditional autologous fat grafting can obtain satisfactory results early after surgery, but the long-term results are not satisfactory. The main reason is the high absorption rate (20-80%) and low survival rate after fat grafting<sup>2,4</sup>. Therefore, how to improve the survival rate of autologous fat grafting has become the critical factor. In order to improve the survival rate of fat graft, many methods have been proposed, including the application of cell-assisted lipotransfer (CAL) and platelet-rich plasma (PRP)-assisted lipotransfer.

CAL was first proposed by Matsumoto et  $al^5$  in 2006, which referred to co-transplantation of aspirated fat with

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stromal vascular fraction (SVF) or aspirated fat with enrichment of adipose-derived stem cells (ADSCs), and was subsequently applied to the human body for the first time by Yoshimura et al.<sup>6</sup> Matsumoto and Yoshimura demonstrated that CAL could be used to improve the efficacy of autologous fat grafting in the preclinical and clinical trials. In this technique, the liposuction fluid is rich in SVF. Freshly isolated SVF cells contain multiple cell types, such as endothelial cells, pericytes, fibroblasts, macrophages, and preadipocytes<sup>7</sup>. In particular, adipose SVF provides a rich source of ADSCs. Multiple studies have shown that cytokines and growth factors secreted by ADSCs stimulate tissue repair in a paracrine manner<sup>8</sup>. At the same time, ADSCs promote angiogenesis of endothelial progenitor cells in SVF<sup>9,10</sup>. The advantages of SVF components and its ease of use have facilitated the development of the SVF-enriched ADSC techniques<sup>11</sup>.

PRP is a kind of autologous plasma with red blood cells removed by centrifugation. It contains platelets and other cellular components. It also has various biologically active molecules, including growth factors (such as transforming growth factor- $\beta$ , platelet-derived growth factors, epidermal growth factor, vascular endothelial growth factor, basic fibroblast growth factor, insulin-like growth factor-1), cytokines, chemokines, and so on<sup>12,13</sup>. Because PRP is rich in growth factors and plasma components, it provides nutritional support, increases angiogenesis and the survival of fat grafts, promotes the proliferation of preadipocytes and adipose-derived mesenchymal stem cells. Its accessibility has led to the active components in autologous blood into the eyes of researchers<sup>14</sup>.

So far, there is no consensus in the literature on the best way to handle fat to ensure the maximum fat survival rate and the viability of grafted fat. CAL and PRP-assisted lipotransfer are the two most promising options currently. Therefore, we conducted a meta-analysis of clinical trials, cohort studies and case series of CAL and PRP-assisted lipotransfer to analyze and summarize their clinical efficacy and safety to better guide clinical practice.

### **Materials and Methods**

### Protocol

The research methodology of this study followed the PRISMA 2009 guidelines<sup>15</sup>.

### Data Sources and Search Strategy

Two independent authors identified a systematic review of relevant articles published between January 2010 and January 2020 in PubMed, Cochrane Library, Web of Science and EMBASE. The search terms included keywords related to "fat graft" or "autologous fat" or "fat transplantation" or "lipofilling" or "lipotransfer" or "lipograft" in combination with "stromal vascular fraction" or "SVF" or "stem cell" or "cell assisted" or "ADSC" or "ADRC" or "ASC" or in combination with "platelet rich plasma" or "PRP." The keyword search strategy for PubMed is shown in Appendix 1. The retrieval of articles and the selection of titles and abstracts were carried out independently by the two authors. Full text evaluation of qualified articles were according to inclusion criteria. Any discrepancies were resolved by discussions among authors.

### Literature Selection

The eligible articles must meet the following criteria: (a) autologous fat for soft tissue reconstruction or augmentation; (b) evaluating the effect of CAL or PRP-assisted lipotransfer in human; (c) with at least 3 months' follow-up period; (d) the number of cases included shall be no less than 5; (e) articles written in English.

The exclusion criteria were listed as: (a) the articles did not meet the inclusion criteria. (b) other diseases did not associate with soft tissue reconstruction or augmentation. (c) there were other interventions that interfered with the results. (d) abstracts, letters, preclinical articles, reviews.

### Data Collection Process and Quality Assessment

Data extraction was performed by two independent reviewers (A.C. and L.Z.) according to a pre-designed data extraction method. The following information was included: (a) Identity: authors, years. (b) Patients included in each study: number, age, BMI. (c) Treatments: intervention factor, stem cell isolation method, injected PRP/fat graft, recipient sites, number of operations, injected volume, volumetric measurement methods, follow-up period. (d) Outcomes: gained volume, fat survival rate, postoperative complication rate, patients Satisfaction. (e) Study design, level of evidence. For articles reporting incomplete data, we contacted the relevant authors.

Two reviewers (A.C. and L.Z.) independently scored the quality of the research and the level of evidence for each study in accordance with the Oxford center for evidence-based medicine 2011 guidelines<sup>16</sup>. Disagreements among reviewers were resolved through discussion.

According to the Cochrane collaboration's tool for assessing risk of bias in randomised trials<sup>17</sup>, a biased risk map and a biased risk summary chart were drawn for bias risk assessment.

### Types of Outcome Indicators

The efficacy of intervention measures were evaluated by fat survival rate. In the light of the incidence of complications and the frequency of multiple operations to evaluate the safety. Complications were assessed including cysts, calcification, fat necrosis, nodules, and fibrosis. Fat necrosis is the initial manifestation of ischemia and hypoxia of the grafted fat particles. Cysts, nodules and calcifications are different manifestations of the further progress of fat necrosis. Redness, swelling and subcutaneous ecchymosis that appear immediately after surgery, local infection after surgery recovering within one to two weeks, undercorrection or overcorrection, asymmetry which are not considered complications. The frequency of multiple operations was defined as repeating the same type of fat grafting surgery at the same recipient site.

### Statistical Analysis

Cochrane collaboration software (RevMan 5.2) was used for meta-analysis. A total of 23 eligible articles were included to evaluate the efficacy of CAL technology, PRP-assisted lipotransfer technology and traditional fat grafting. In order to include articles that included traditional fat grafting as a control and enrich the sample data in this paper, we used a meta-analysis of the non-controlled dichotomy data, namely a meta-analysis of the single rate, to calculate the pooled estimates and the 95% fat survival confidence interval for each group using the generic inverse variance and random effect models. For evaluating the heterogeneity of the results of the included studies and exploring the differences in the results of the studies in different recipient sites, subgroup analyses were performed to calculate the inconsistent statistics (I2) of the totals and subtotals. Funnel plot was used to show publication bias.

In addition, 30 articles were included to evaluate the safety of CAL technology, PRP-assisted lipotransfer technology and traditional fat grafting. Chi-square test with Graphpad Prism 8.0 was used to evaluate whether different interventions had statistical differences in the incidence of complications and multiple surgical operations in different recipient sites.

### Results

### Literature Search

A total of 2,639 articles were retrieved using the predefined keywords, and another 37 were included from the references of other articles. We eliminated 1,835 duplicate articles and included 112 potential full-text articles by reading the title and abstract. After a careful review of the full text, this review finally included 36 articles. The selecting process is shown in Fig. 1.

### Characteristics and Quality of the Included Studies

We included 36 studies with 1,697 cases<sup>2,4,18–51</sup>. The basic characteristics of 36 studies are summarized in Table 1 . The included studies included randomized controlled trials, cohort studies (single-arm or double-arm), and case series. The meta analysis included 24 articles containing the survival rate of grafted fat, including 19 studies about CAL with 453 cases<sup>2,4,18,22,23,26,28–37,40,41</sup> and 7 studies about PRP-assisted lipotransfer with 359 cases<sup>31,34,44–47,49</sup> (there were 2 studies that included both cell-assisted and PRP-assisted lipotransfer<sup>31,34</sup>) (Table 1). For safety evaluation, we assessed the incidence of fat grafting complications. A total of 31 studies reported

postoperative complications<sup>2,4,18,19,21,23–40,43,45–49,51</sup>. We evaluated the quality of study design and evidence levels for all included studies based on the Oxford centre for evidence-based medicine 2011 guidelines (Table 2). Of these, 8 studies had evidence levels of II, 21 had evidence levels of IV.

### Methodological Quality of Included Studies

The risk of bias was assessed according to the Cochrane collaboration group's risk of bias tool manual<sup>17</sup> for eight included randomized controlled trials<sup>2,20,23,30,38,39,47,49</sup> (Figs. 2, 3). In the eight randomized controlled trials, the risk of each biased item was assessed by percentages. More than 25% risk of allocation concealment bias, performance bias, and reporting bias existed in the eight studies. All the included studies showed a low risk of attrition bias. To sum up, the methodological quality of included studies was feasible.

### Efficacy of Cell-Assisted and PRP-Assisted Lipotransfer

According to the survival rate of grafted fat to determine the efficacy of fat grafting (the percentage of obtained fat volume to injected fat volume). In order to study the efficacy of fat grafting in different recipient areas, subgroup analyses were performed on different recipient areas (Figs. 4–7). For heterogeneity assessment in different recipient areas, the inconsistency value (I2) of CAL is 39.4%, non-CAL is 73.1%, and PRP-assisted lipotransfer are 0%.

Figs. 4 and 5 show that the fat survival rate of the CAL group was significantly higher than that of the non-CAL group (71%), 95% CI [67, 75] vs 48%, 95% CI [38, 58], P < 0.0001). In the subgroup analysis of the recipient areas, the facial fat survival rate of the CAL group was significantly higher than that of the non-CAL group (77%, 95% CI [69, 86] vs 51%, 95% CI [38, 64], P < 0.0001). Similarly, the breast fat survival rate of the CAL group was higher than the non-CAL group (69%, 95% CI [64, 74] vs 51%, 95% CI [36, 65], P < 0.0001). The fat survival rate of CAL group was significantly higher than non-CAL group in the only arm study 2 (81% vs 16%, P < 0.0001). As can be seen from Figs. 6 and 7, the fat survival rate of the PRP group was significantly higher than that of the non-PRP group (70%, 95% CI [65, 75] vs 40%, 95% CI [33, 47], *P* < 0.0001). There was no difference in subgroup analysis in different recipient areas (P > 0.5, I2 = 0%). In Figs. 4 and 6, we compared the ability of CAL group with PRP group about promoting the survival of fat grafting, and found no statistical difference between the two groups(71%, 95% CI [67, 75] vs 70%, 95% CI [65, 75], P = 0.7175).

CAL researches include enzymatic separation of SVF (including automatic separation and manual separation), non-enzymatic separation of SVF (i.e., mechanical separation), and *in vitro* culture of ADSCs. In order to compare the differences in fat survival rates of SVF treated by different methods, the heterogeneity assessment by subgroup analysis demonstrated that inconsistency value (I2) was 63.2%

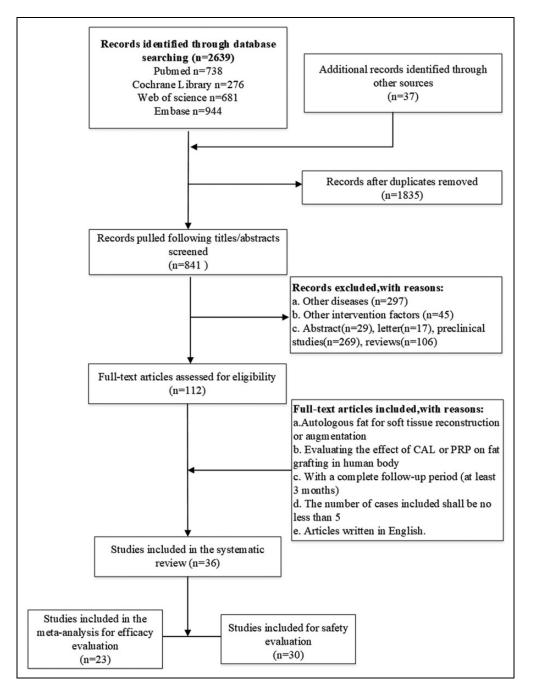


Figure 1. Flow diagram of the study selection. CAL: cell-assisted lipotransfer; PRP: platelet-rich plasma.

(Fig. 8). There was no statistical difference in the survival rate of fat between automatic separation and manual separation (70%, 95% CI [64,76] and 68% [62,75], P = 0.6072). The mechanically separated fat and ADSCs with *in vitro* culture have higher survival rate (90%, 95% CI [76,105] and (80% [60,100], P = 0.5628).

## Safety of Cell-Assisted and PRP-Assisted Lipotransfer

For safety evaluation, we assessed the incidence of fat grafting complications, including: cysts, calcification, fat necrosis, nodules, and fibrosis. A total of 31 studies reported postoperative complications<sup>2,4,18,19,21,23-40,43,45-49,51</sup>. The total average follow-up time was  $12.2 \pm 8.2$  months. The total complication rate was 8.7% (125/1429), of which the cysts had the highest rate (6.0%, 86/1429), followed by fat necrosis (2.0%, 28/1429), calcification (0.6%, 9/1429) and nodule (0.1%, 2/1429). We found that all complications occurred in breast surgery, and no complications occurred in the rest of the body. Therefore, we specifically analyzed the incidence of breast complications in different interventions (Fig. 9A). The incidence

Reference	Intervention	Study size	Age, Yr (mean ± SD, range)	BMI Kg/m <sup>2</sup> (mean $\pm$ SD, range) Recipient	Recipient sites	No. of operations	Injected volume, ml (mean ± SD, range)	Volumetric methods	Gained volume, ml (mean ± SD, range)	Fat survival rate,% (mean ± SD, range)	Postoperative complications	Follow-up, Mo (range)	Patients satisfaction	tion tion method or injected PRP/fat graft
Tanikawa et al.,	SVF + Fat	7	$12.1 \pm 2.2$	2 patients <20; 5 patients 20–25	Face	-	$27 \pm 7$	Photography+ 3D CT	NA	88 ± 13	No cases of surgical complications	9	All patients were satisfied	Manual
5102	Fat	7	18.7 ± 7.6	3 patients <20; 4 patients 20–25	Face	-	$29\pm6$		٩N	$54 \pm 20$	No cases of surgical	9	42.9% (3/7)	
Sterodimas et al., 2011 <sup>20</sup>	SVF + Fat	0	$43.9\ \pm\ 16.9\ (22{-72})$	21.5 (17.4-25.2)	Face tissue defects	_	74.3 ± 47 (8–155)	Satisfaction	NA	NA	NA	8	76%	Manual
107	Fat	0	$46.2 \pm 17.7 \ (25{-70})$	21.6 (17.1–25.6)	Face tissue defects	2 in 30 % (3/10); 3 in 40% (4/10)	78.5 ± 49.4; (I2–I65)		AN	NA	NA	8	80% (after several operations)	
Doi et al., 2012 <sup>27</sup>	SVF + Fat	4	<b>43.2</b> ± 13.0	200 ± 2.4	<ol> <li>Faces rejuvenation</li> <li>Faces reconstruction</li> <li>Breasts</li> <li>Breasts</li> </ol>	-	16.8 ± 3.1 29.4 ± 9.4 217 ± 60 225 ± 48	Physical + photography	۲ ۲	42	Without any serious side- effects or complications	38	Sausfied	Automatic
Lee et al., (2012 <sup>24</sup>	SVF + Fat	6	43.3 ± 14.7 (29–68)	NA	Left face	-	NA	Clinical examination + photography	AN	NA	No significant adverse effect	m	75%	Manual
	Fat	6	43.3 ± 14.7 (29–68)	NA	Right face	-	NA	- L- 0 L	NA	NA	No significant adverse	m	62%	
Peltoniemi et al., 2013 <sup>29</sup>	t SVF + Fat	2	48.5 ± 7.9 (29–58)	$24.8 \pm 3.9 \; (20.3 – 32.5)$	Breast augmentation	-	Left: 289.6 $\pm$ 88.8; Right: 279.3 $\pm$ 72.1	MRI	Left: 147.7 ± 70.5; Right: 147.1 ± 573	Left: 49.2 $\pm$ 10; Right: 51.8 $\pm$ 9.5	enect I in few small oil cysts	Q	۹Z	Automatic
	Fat	æ	$43.4 \pm 11.1 \ (33-63)$	23.5 ± 2 (20.3–25.9)	Breast augmentation	-	Left: 303.1 ± 27.6; Right: 297.5 ± 27.6		26.9; Right: 26.9; Right: 163.5 + 24	Left: 52 $\pm$ 10.3; Right: 55.1 $\pm$ 7.6	I in few small oil cysts	Q		
Gentile et al., 2015 <sup>33</sup>	SVF + Fat	10 (Celution) 10 (Fatstem) 10 (Mystem) 10 (Medikhan)	19-65	NA	Breast soft tissue defects	7	187 (110–250)	MRI + US	A N	63 ± 6.2 52 ± 4.6 43 ± 3.8 39 + 3.5	Oily cysts in 45.83% (23/ 50) cytosteatonecrotic in 12.5% (6/50)	2	All patients were satisfied (Celution and Fatstem)	Automatic
	Fat	10 (Manual)	21-65	NA	Breast soft tissue	2	187 (110–250)		NA	39 ± 4.4		12		
Jung et al., 2016 <sup>36</sup>	SVF + Fat	5 (10breasts)	34.4 ± 9.15 (19–41)	20.18 ± 2.84	Breast augmentation	-	Left: 196.2 ± 49.69; Right: 246.2 ± 62.99	MRI	Left: 103.6 ± 65.10 Right: 102.3 ± 35.91	47%	3 in cysts	12	۹Z	Automatic
Perez-Cano et al., 2012 <sup>25</sup>	SVF + Fat	67	52 (37–68)	24.5 (17–31)	Partial mastectomy	l in 64.2% (43/67); 2 in 35.8% (24/67)	140 (35–250)	MRI	NA	٩٧	10 in cysts and no reported local cancer recurrences	13	74.6% (50/67)	Automatic
Li et al., 2013 <sup>28</sup>	SVF + Fat	26	29.5 ± 6.8	NA	Face	] –	17.5 ± 7.3	CT Scan + photography	$11.5 \pm 5.3$	<b>64.8</b> ± 10.2	No complications	9	٩N	Manual
Gentile et al., 2012 <sup>22</sup>	Fat , SVF + Fat	10	29.1 ± 6.0 (19–60)	NA	Face Breast reconstruction		16.2 土 6.3 395.4 (80-600)	MRI + US	7.6 ± 3.3 NA	46.4 ± 9.3 63	No complications NA	9	NA All satisfied	Automatic
	Fat	0	21-65	NA	Breast	-	395.4 ( 80–600)		NA	39	ΝA	12		
Kamakura et al., 2011 <sup>19</sup>	SVF + Fat	20	35.6 ± 8.8 (21–52)	NA	Breast augmentation	-	Right: 244.9 ± 36.1; Left: 235.1 ± 34	MRI	AN	NA	2 in cysts	6	75% satisfaction	Automatic
Schendel et al., 2015 <sup>35</sup>	SVF + Fat	0	51.6 ± 9.57 (36–71)	NA	Face	-	18.4 ± 15.34	3D scan	AN	89	No complications	12.6	Ч	Manual
Dos Anjos et al., 2015 <sup>32</sup>	t H-SVF + Fat	28	<b>39.36</b> ± 12.83	21.57 ± 1.75	Breast reconstruction	2 in 3.9% (2/56)	<b>270.74</b> ± <b>55.6</b>	3D scan + MRI	AN	75	14 in cysts	8	ЧN	Automatic
	L-SVF + Fat	21	37.8 土 18.89	$21.59 \pm 2.09$	Breast	-	$229.09 \pm 63.42$		NA	50		8	NA	Automatic
Tissiani et al., 2016 <sup>37</sup>	, SVF + Fat	=	49.63 ± 5.66 (41–58)	26.26 ± 2.38 (23.5–30.9)	Breast reconstruction	-	134.3 ± 40.8 (45–180)	MRI scan	116.23 ± 108.49 (−12.3– 406.48)	$\begin{array}{l} \textbf{78.85} \pm \textbf{74.87} \\ \textbf{(?27.33-276.51)} \end{array}$	4 in fat necrosis	36 (20–49)	50%	Manual
	Fat	ω	49.75 ± 11.46 (36–69)	25.87 ± 3.51 (20.8–32.4)	Breast reconstruction	_	III.5 ± 22.8 (96–147)		$55.4 \pm 16.51$ $(31.73-$ $76.09)$	$51.39 \pm 18.53 \\ (27.65-72.86)$	No complications	16 (8–30)	80%	

# Table 1. Overview of the Characteristics of Included Studies.

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(continued)	
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Table	

tion method or injected Patients satisfaction PRP/fat graft	NA Manual	NA	Satisfied Manual	Satisfied	Satisfied Automatic	Satisfied	All patients Manual were satisfied	92%(11/12) Manual	All satisfied Automatic in 18 cases: Manual in 11	NA Mechanical dissociation	NA NA Mechanical dissociation	NA Manual	AN	90% NA	62%	I he same as placebo	The same as placebo	Satisfied Manual	Satisfied 5/10	90–95% Automatic	90–95% 2/10	sausned NA 2–5/10 NA 2–5/10	AN
Follow-up, Mo (range) Pa	9	9	13.4 土 1.6	$15.8 \pm 2.7$	over12	over12	2	6	0	12	12	4	4	15	5	71	12	12	2 2	12	12	2 2	12
Postoperative complications	No complications	No complications			necrotic cysts (3.6%) 4 in recurrences(3.3%)	5 in recurrences(10%)	No serious side-effects have observed	2 in cysts and nodules	No complications	No major complications	No major complications NA	No serious adverse events	No serious adverse events	No complications	No complications	No complications	No complications	No complications	No complications	INO COMPICATIONS NA	AA	۲ ۲ ۲	AN
Fat survival rate, % (mean ± SD, range)	<b>68.3</b> ± 1.7	58.5 ± 1.3	68.7	67.9	70.8	41.4	57	$\textbf{48.2}~\pm~\textbf{16.7}$	NA	90.4	76 NA	80.9 ± 6 (76.6–85.2)	16.3 ± 7.2 (11.1–21.4)	79.41	53.19	Uid not significantly improve volume	retention NA	63	69 39	57 72.9 ± 50 (49–79)	68.5 ± 39.5	38.3 土 12.9 (18–56) 0.4–0.5/1m1?70% 0.3/1m1?62% 0.3/1m1?62%	31
Gained volume, ml (mean 土 SD, range)	NA	NA	228.6	211.1	NA	NA	Right: 155 ± 50 Left: 143 ± 80	125.35 ± 45.49	NA	NA	A A NA	23 ± 3.2 (20.57–	25.43) 4.66 土 2.1 2.1	(3.16–6.16) 18.1 ± 5.2	4.4 ± 1.9	AN	NA	NA	¥ ₹	A N	NA	A N N	٩
Volumetric methods	CT Scan		US+3D laser	scanning	MRI + US		MRI + 3D measurements	MRI	Physical examination +	pnorography Photography + CT	Palpation + photography	MRI		3D CT + 3D	camera	Skin messurement +	photography Skin messurement +	photography Photography + Mol. 4 15	50 + MH	3D Imaging		Photography+ MRI + US	
Injected volume, ml (mean ± SD, range)	34.4 土 13.7	38.2 土 14.9	334 土 44	310 ± 36	۸A	AN	Right: 268 ± 29 Left: 259 ± 39	256 ± 36.27	(10–390)	58 ± 3.52I (45–65)	53 ± 5.385 (40-70) 10-100	28.2 ± 3.2 (25.94–30.46)	$28.6\ \pm\ 3.3\ (26.26{-}30.94)$	29.9 ± 6.7	12.5 ± 3.4	AN	AN	NA	A N N A	9 ± 2 (9–11)	8.6 ± 2.0 (8–10)	8.5 ± 1 (8-10) Scars: 10-80 Romberg syndrome: 60-140	Scars: 10-80 Romberg syndrome: 60-141
No. of operations	1 in 50% 2 in 30%, 3 in 30%	3 in 20% 1 in 50% 2 in 30%,	2 in 19.8%	(20/101) 2 in 3.8%	(c01%)	_	-	_	2 in 13.8% (4/29)	_		_	_	_		_	-	2 in 40% (4/10)	2 in 40% 2 in 40% (4/10)	(01/ <del>1-</del> ) % 0+ 111 7	-		-
Recipient sites	Progressive hemifacial	arropny Progressive hemifacial	Breast	augmentation Breast	augmentation Breast	Breast reconstruction	Breast augmentation	Breast	augmentation Breast: 17. Face: 5. Others: 7	Face	Face Facial contouring	Upper arm	Upper arm	Face	Face	Facial filling	Facial filling	Face	Face	Face	Face	Face Soft tissue defects	Soft tissue defects
BMI Kg/n $^2$ (mean $\pm$ SD, range) Recipient sites	AN	AN	$20.3 \pm 2.4$	18.8 土 1.6	ΥN	NA	<b>19.5</b> ± <b>1.4</b>	$22.1 \pm 3.24$	Ч Ч	29.89 ± 0.659 (25.6–33.2)	27.89 ± 0.539 (24.6–30.9) NA	24.7 ± 2 (23.3–26.1)	24.7 ± 2 (23.3–26.1)	AN	AN N	NA	NA	NA	AN MA	22 (21–30.8)	22.3 (16.9–29.5)	22 (19:9–24.2) NA	AN
Age, Yr (mean ± SD, range) E	27.5 (19–35)	27.5 (19–35)	37 土 7.4	33 土 9.4	56.24 ± 11.44 (25–85)	56.24 ± 11.44 (25–86)	<b>37.I</b> ± 12.5	$32\pm7.3(28{-}56)$	٩N	42.40 ± 2.636 (27–60)	39.67 ± 2.681 (20–55) NA	$28.4 \pm 8.9 \; (22 – 34.8)$	$28.4 \pm 8.9 \; (22 – 34.8)$	28	28	(38−62) ± 6.7 (38−62)	52.5 ± 7.1 (42–63)	(23–67)	21–69 073–67)	(22-01) 65.5 (52-77)	62.1(19–77)	60.5 (58–63) 36.6 (18–75)	36.6 (18–75)
Study size	01	0	101	105	121	50	15	12	29	15	15 25	0	0	5	<u>،</u> د	3	12	0	<u>o</u> <u>o</u>	2 6	901	92 136	0
Intervention	SVF + Fat	Fat	SVF + Fat	Fat	SVF + Fat	Fat	SVF + Fat	SVF + Fat	SVF + Fat	SVF + Fat	Fat SVF + Fat	ADSCs + Fat	Fat	ADSCs + Fat	Fat	PKP + Fat	Fat	SVF + Fat	PRP + Fat Eat	rat SVF + Fat	PRP + Fat	Fat PRP + Fat	Fat
Reference	Chang et al. 2013 <sup>26</sup>		Chiu et al.,	- 8107	Gentile et al., 2019 <sup>41</sup>	107	Yoshimura et al., 2010 <sup>18</sup>	Wang et al.	Z013 Tiryaki et al., 2011 <sup>21</sup>	Gontijo-de- Amorim et al., 2017 <sup>38</sup>	Gontijo-de- Amorim et al.,	Kolle et al.,	-5107	Koh et al.,	7107	Willemsen et al,	2018 <sup>39</sup>	Gentile et al.,	+ 107	Sasaki et al., 2015 <sup>34</sup>	6107	Cervelli et al., 2013 <sup>44</sup>	
öZ	15		9		17		8	6	50	21	22	23		24		จ		26		27		28	

ö Z	Reference	Reference Intervention	Study size	Age, Yr (mean ± SD, range)	BMI $kg/m^2$ (mean $\pm$ SD, range) Recipient sites	Recipient sites	No. of operations	Injected volume, ml (mean ± SD, range)	Volumetric methods	Gained volume, ml (mean ± SD, range)	Fat survival rate, $\%$ (mean $\pm$ SD, range)	Postoperative complications	Follow-up, Mo (range)	Patients satisfaction	Stem cell isola- tion method or injected PRP/fat graft
29	Keyhan et al., 2013 <sup>47</sup>	PRP + Fat	25	45 (24-69)	NA	Facial lipostructure	-	8 ml per cheekbone and 7 ml per cheek	Clinical examination +	AA	82	No major complications	12	96% Satisfied (1/25)	01/1-6/1
		PRF + Fat	25	45 (24–69)	NA	Facial lipostructure	_	8 ml per cheekbone and 7	photography	Υ	87	No major complications	12	Satisfied	01/1
30	Fontdevila et al., 2014 <sup>49</sup>	PRP + Fat	11	<b>47.3</b> ± <b>8.0</b>	$\textbf{24.5}\pm\textbf{2.9}$	Human immuno- deficiency virus facial	-	ти рег спеек 17.2 ± 4.8	Photography + CT	NA	51.6	No major complications	2	AN	2.5/10
		Fat	28	45.6 ± 6.9	<b>24.2</b> ± <b>3.5</b>	IIpoatrophy Human immuno- deficiency virus facial	-	22.5 ± 6.2		NA	ΥN	No major complications	2	Ϋ́	
E	Willemsen et al., 2014 <sup>50</sup>	PRP + Fat	8	35-65	NA	Inpoartopriy Facial rejuvenation	-	I5 (13–23 in each side of Photography the face)	Photography	NA	Better than control	٩Z	m	ΥN	01/1
		Fat	25	3565	NA	Facial rejuvenation	_	15 (13–23 in each side of the face)		AN	ΔN	٩N	ĸ	NA	
32	Gentile et al., PRP + Fat 2013 <sup>46</sup>	PRP + Fat	50	1960	ΔN	Breast reconstruction	-	120 for each breast	Photography + MRI + US	ΥN	69	I in cystic formation and macrocalcification	12	Satisfied	5/10
		Fat	50	1960	NA	Breast reconstruction	-	120 for each breast		AN	39	٩N	12	AN	
e e	Salgarello et al., 2011 <sup>43</sup>	PRP + Fat	5	47 (27–68)	NA	Breast lipofilling	3 in 12% (2/17) 2 in 59% (10/17) 1 in 29% (5/17)	120 (25–231)	SU	AN	No statistical difference compared with the control group	4/17 in oily cysts 7/17 in fat necrosis	9 (3–16)	24% Satisfied (4/17)	01/10
		Fat	25	47 (27–68)	NA	Breast lipofilling	3 in 16% (4/25) 2 in 56% (14/25) 1 in 28% (7/25)	115 (21–169)		AN	AN	4/25 in oily cysts 7/25 in fat necrosis	9 (3–24)	28% Satisfied (7/25)	
34	Ozer et al., 2019 <sup>51</sup>	PRP + Fat	4	$44.9 \pm 11.9 \; (33-65)$	AA	Facial filling	_	35.6	Photography	AN	AN	No major complications	6≕<	Satisfied (87.6 + 16.8)	4/10
35	Fiaschetti et al., 2013 <sup>45</sup>	PRP + Fat	I5 (24 breasts)	46.27	22.458	Breast lipofilling	2	93.54 (50–150)	MRI + US	AN	71.77	11/24 in oily cysts 4/24 in cytosteatonecrosis 8/ 24 in calcification	12	٩Z	
36	Willemsen et PRP + Fat al., 2013 <sup>48</sup>	PRP + Fat	24	42.8 (21–69)	$24.7\pm0.9$	Buttock augmentation	-	481	Post-operative questionnaire	NA	AN	No major complications	4	Slightly higher at 3 months after	01/1
														the operation than	

BMI: body mass index; SVF: stromal vascular fraction; CT: computed tomography; NA: not available. MRI: magnetic resonance imaging; US: ultrasound; ADSCs: adipose-derived stem cells; PRP: platelet-rich plasma.

Table 2. Study Design and Level of Evidence.

No	Reference	Study design	Level of evidence
I	Tanikawa et al., 2013 <sup>30</sup>	Randomized clinical trial	II
2	Sterodimas et al., 2011 <sup>20</sup>	Randomized clinical trial	П
3	Doi et al., 2012 <sup>27</sup>	Retrospective cohort	IV
4	Lee et al., 2012 <sup>24</sup>	prospective cohort	111
5	Peltoniemi et al., 2013 <sup>29</sup>	Prospective cohort	III
6	Gentile et al., 2015 <sup>33</sup>	Prospective cohort	111
7	Jung et al., 2016 <sup>36</sup>	Prospective cohort	111
8	Perez-Cano et al., 2012 <sup>25</sup>	prospective clinical trial	111
9	Li et al., 2013 <sup>28</sup>	Retrospective cohort	111
10	Gentile et al., 2012 <sup>22</sup>	Retrospective cohort	111
11	Kamakura et al., 2011 <sup>19</sup>	Single-arm clinical trial	III
12	Schendel et al., 2015 <sup>35</sup>	Prospective cohort	III
13	Dos Anjos et al., 2015 <sup>32</sup>	Retrospective clinical trial	111
14	Tissiani et al., 2016 <sup>37</sup>	Prospective clinical trial	111
15	Chang et al., 2013 <sup>26</sup>	Retrospective cohort	111
16	Chiu et al., 2018 <sup>40</sup>	Retrospective clinical trial	III
17	Gentile et al., 2019 <sup>41</sup>	Retrospective cohort	111
18	Yoshimura et al., 2010 <sup>18</sup>	case series	IV
19	Wang et al., 201 <sup>54</sup>	Case series	IV
20	Tiryaki et al., 2011 <sup>21</sup>	Case series	IV
21	Gontijo-de-Amorim et al., 2017 <sup>38</sup>	Randomized clinical trial	11
22	Gontijo-de-Amorim et al., 2020 <sup>42</sup>	Case series	IV
23	Kolle et al., 201 <sup>32</sup>	Randomized clinical trial	11
24	Koh et al., 2012 <sup>23</sup>	Randomized clinical trial	11
25	Willemsen et al., 2018 <sup>39</sup>	Randomized clinical trial	11
26	Gentile et al., 2014 <sup>31</sup>	Retrospective clinical trial	111
27	Sasaki et al., 2015 <sup>34</sup>	Prospective clinical trial	111
28	Cervelli et al., 2013 <sup>44</sup>	Prospective clinical trial	111
29	Keyhan et al., 2013 <sup>47</sup>	Randomized controlled trial	11
30	Fontdevila et al., 2014 <sup>49</sup>	Randomized clinical trial	11
31	Willemsen et al., 2014 <sup>50</sup>	Retrospective cohort	111
32	Gentile et al., 2013 <sup>46</sup>	Clinical trial	111
33	Salgarello et al., 2011 <sup>43</sup>	Retrospective clinical trial	Ш
34	Ozer et al., 2019 <sup>51</sup>	Case series	IV
35	Fiaschetti et al., 2013 <sup>45</sup>	Case series	IV
36	Willemsen et al., 2013 <sup>48</sup>	Case series	IV

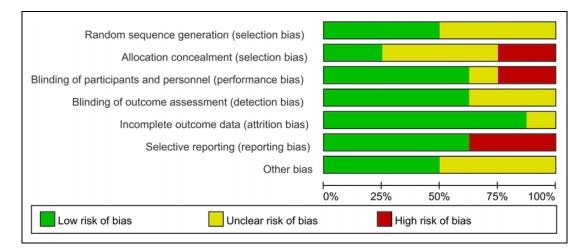
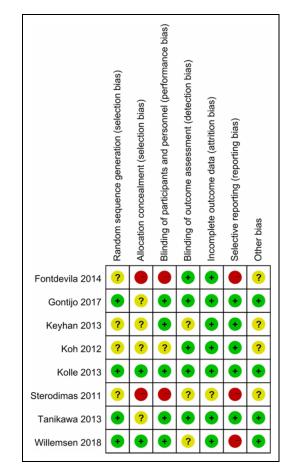


Figure 2. Bias risk graph: a review of the authors' estimates of bias risk in the seven randomized controlled trials included.



**Figure 3.** Risk of bias summary: review the author's decisions on each bias risk item for each randomized controlled trial. Red (-): high risk of bias; Yellow (?): unclear risk of bias; Green (+): low risk of bias.

of CAL complications was higher than non-CAL (12.81% [67/523] vs 6.08% [11/181], P = 0.0129), PRP group was higher than non-PRP group (36% [36/100] vs 14.67% [11/75], P = 0.0016).

Another indicator for safety assessment is the frequency of multiple operations. All the studies in Table 1 were included. For the CAL group and the non-CAL group, both the overall and breast surgery statistics, the incidence of multiple operations of the CAL group was higher than the non-CAL group (the overall: 14.45% [99/685] vs 8.09% [30/371], P = 0.0026; breast: 16.7% [90/539] vs 7.33% [14/191], P = 0.0015), but in terms of the frequency of facial operations, there was no statistical difference between CAL and non-CAL (6.62% [9/136] vs 9.41% [16/170], P = 0.3752) (Fig. 9B–D).

Between the PRP-assisted lipotransfer group and the non-PRP-assisted lipotransfer group, regardless of the overall, face, or breast, there was no statistical difference in the incidence of multiple operations (the overall: 6.97% [31/445] vs 8.73% [22/252], P = 0.3986; breast: 32.93% [27/82] vs 24% [18/75], P = 0.2166; face: 1.18% [4/339] vs 2.26% [4/177], P = 0.3459;) (Fig. 9B–D).

We also compared the relationship between manual and automatic separation of SVF and the incidence of multiple operations, and found no significant correlation between the two interventions (12.93% [30/232] vs 10.33% [38/368], P = 0.3270).

### Publication Bias

The funnel charts (Fig. 10) show that the four groups of CAL, non-CAL, PRP-assisted lipotransfer, and non-PRP-assisted lipotransfer have no published bias intuitively.

### Discussion

With the development of autologous fat grafting technology, researchers tried to find a way to maximize the survival of grafted fat and ensure its safety. In the past 10 years, we can see that cell-assisted fat grafting and the addition of active ingredients from the blood have become the two methods that were recognized by experts in the industry most, and related research articles have also appeared continuously. Most studies indicate that both can promote the survival of grafted fat to a certain extent, but there are a few articles think that both have not improved the survival rate<sup>4,29,36,39,49,52</sup>, and may even increase the incidence of complications<sup>40,43</sup>. Therefore, we conducted a meta-analysis to determine the efficacy and safety of CAL and PRP-assisted lipotransfer compared to traditional fat grafting.

### Meta-Analysis Outcomes

The results of the cell-assisted and PRP-assisted lipotransfer studies indicate that regardless of the location of the recipient areas, both cell-assisted and PRP-assisted lipotransfer can increase the survival rate of grafted fat, and there is no difference in the degree of survival rate between the two (Fig. 4). However, both increased the incidence rate of complications in the recipient area. And we found that all complications only occurred in the breast, and no related complications occurred on the face (Fig. 9A). Therefore, we think that one of the main reasons is that the volume of grafted fat leads to complications. According to Yoshimura "three-zone theory"<sup>53</sup>, the evolution of the fat from the edge to the center after grafting, respectively were: survival zone (fat cells survived), regeneration zone (fat cells died, adipose stromal cells survived and dead fat cells were replaced by new ones), and necrotic zone (both fat cells and adipose stromal cells died). The breast surgery requires a larger graft volume than facial surgery, so ischemic necrosis and reabsorption are more likely to occur in the center of breast grafts, which can lead to calcification, oil cysts, and masses<sup>54</sup>. The current cell-assisted and PRP-assisted lipotransfer technology is not enough to reverse the ischemic state and cannot reduce the complications of fat grafting in breast surgery. In addition, it can be seen from

				Rate Difference	Rate Dif	ference
Study or Subgroup	Rate Difference	SE	Weight	IV, Random, 95% CI	IV, Rando	m, 95% Cl
1.2.1 Face						
Chang 2013	0.683	0.147	2.1%	0.68 [0.39, 0.97]		
Gentile 2014	0.63	0.153	1.9%	0.63 [0.33, 0.93]		
Gontijo 2017	0.904	0.076	7.6%	0.90 [0.76, 1.05]		
Koh 2012	0.794	0.181	1.4%	0.79 [0.44, 1.15]		
Li 2013	0.648	0.094	5.0%	0.65 [0.46, 0.83]		
Sasaki 2015	0.729	0.148	2.1%	0.73 [0.44, 1.02]		
Schendel 2015	0.68	0.148	2.1%	0.68 [0.39, 0.97]		
Tanikawa 2013	0.88	0.123	3.0%	0.88 [0.64, 1.12]		
Subtotal (95% CI)			25.1%	0.77 [0.69, 0.86]		•
Heterogeneity: Tau <sup>2</sup> = 0	0.00; Chi <sup>2</sup> = 7.24, c	lf = 7 (P	= 0.40); I	<sup>2</sup> = 3%		
Test for overall effect: 2	Z = 17.84 (P < 0.00	001)				
1.2.2 Breast						
Chiu 2018	0.687	0.046	20.0%	0.69 [0.60, 0.78]		-
Dos Anjos 2015	0.75	0.058	12.9%	0.75 [0.64, 0.86]		
Gentile 2012	0.63	0.153	1.9%	0.63 [0.33, 0.93]		
Gentile 2015	0.63	0.153	1.9%	0.63 [0.33, 0.93]		
Gentile 2019	0.708	0.041	24.7%	0.71 [0.63, 0.79]		-
Jung 2016	0.47	0.223	0.9%	0.47 [0.03, 0.91]		
Peltoniemi 2013	0.505	0.158	1.8%	0.51 [0.20, 0.81]		· · ·
Tissiani 2016	0.79	0.123	3.0%	0.79 [0.55, 1.03]		
Wang 2015	0.482	0.144	2.2%	0.48 [0.20, 0.76]		
Yoshimura 2010	0.57	0.128	2.7%	0.57 [0.32, 0.82]		
Subtotal (95% CI)			72.0%	0.69 [0.64, 0.74]		•
Heterogeneity: Tau <sup>2</sup> = 0	0.00; Chi <sup>2</sup> = 7.55, c	lf = 9 (P	= 0.58); I	<sup>2</sup> = 0%		
Test for overall effect: 2	Z = 28.32 (P < 0.00	001)				
1.2.3 Arm						
Kolle 2013	0.809	0.124	2.9%	0.81 [0.57, 1.05]		
Subtotal (95% CI)			2.9%	0.81 [0.57, 1.05]		
Heterogeneity: Not app						
Test for overall effect: 2	Z = 6.52 (P < 0.000	01)				
			100.0%	0 74 [0 67 0 75]		•
Total (95% CI)	0.00.062-40.00		100.0%	0.71 [0.67, 0.75]		<b>•</b>
Heterogeneity: Tau <sup>2</sup> = 0			(P = 0.44	); 1~ = 2%	-1 -0.5 0	0.5 1
Test for overall effect: 2			(D - 0.40	12 - 20 40/		
Test for subgroup differ	rences: $Chi^2 = 3.30$	, ar = 2	(P = 0.19	), 1- = 39.4%		

Figure 4. Pooled estimates of fat survival rate with cell-assisted lipotransfer at different recipient areas.

the study of Fiaschetti et al.<sup>45</sup> that different detection methods (mammography, ultrasound, MRI, etc.) have significant differences in the detection rate of complications. Ultrasound had a higher detection rate of cysts than mammography and MRI, and the detection rate of mammography for calcification was higher than that of ultrasound. However, the objective detection methods for postoperative follow-up in most literatures mainly focus on the final fat volume, so MRI, 3D scanning were more used, while ignoring its impact on the rate of missed diagnosis of complications.

Our original conjecture was that after CAL and PRPassisted lipotransfer, it should reduce the occurrence of multiple operations, thereby reducing the cost of multiple treatments. However, the study found that there was no difference in the incidence of multiple operations between PRP-assisted lipotransfer and non-PRP-assisted lipotransfer. In addition, in breast surgery, the incidence of multiple operations for CAL is actually higher than non-CAL. Analyzing specific data sources, we believe that this result is mainly due to the difference in the number of samples in the CAL group and the non-CAL group. For example, In the study of Gentile<sup>33</sup>, all cases were performed twice, but there were 40 cases in the CAL group, while the number in the control group was only 10. In Chiu's study<sup>40</sup>, CAL group has a higher rate of multiple operations (19.8% vs

				Rate Difference		Difference
Study or Subgroup	Rate Difference	SE	Weight	IV, Random, 95% CI	IV, Ran	idom, 95% Cl
1.3.1 Face						
Chang 2013		0.156	6.0%	0.58 [0.28, 0.89]		
Gentile 2014		0.154	6.0%	0.39 [0.09, 0.69]		
Gontijo 2017	0.76		8.3%	0.76 [0.54, 0.98]		
Koh 2012		0.223	3.8%	0.53 [0.09, 0.97]		
Li 2013		0.144	6.5%	0.46 [0.18, 0.75]		-
Sasaki 2015	0.383	0.051	11.8%	0.38 [0.28, 0.48]		-
Tanikawa 2013	0.54	0.188	4.8%	0.54 [0.17, 0.91]		
Subtotal (95% CI)			47.0%	0.51 [0.38, 0.64]		-
Heterogeneity: Tau <sup>2</sup> =	0.01; Chi <sup>2</sup> = 10.85,	df = 6 (	P = 0.09);	l <sup>2</sup> = 45%		
Test for overall effect:	Z = 7.76 (P < 0.000	001)				
1.3.2 Breast						
Chiu 2018	0.679	0.046	12.0%	0.68 [0.59, 0.77]		-
Gentile 2012	0.39	0.154	6.0%	0.39 [0.09, 0.69]		
Gentile 2015		0.154	6.0%	0.39 [0.09, 0.69]		
Gentile 2019	0.414	0.07	10.7%	0.41 [0.28, 0.55]		
Peltoniemi 2013		0.176	5.2%	0.54 [0.20, 0.88]		
Tissiani 2016		0.177	5.1%	0.51 [0.17, 0.86]		
Subtotal (95% CI)			45.1%	0.51 [0.36, 0.65]		•
Heterogeneity: Tau <sup>2</sup> =	0.02: Chi <sup>2</sup> = 13.44.	df = 5 (	P = 0.02):			
Test for overall effect:			0.02),			
1.3.3 Arm						
Kolle 2013	0.163	0.117	7.9%	0.16 [-0.07, 0.39]		+
Subtotal (95% CI)	0.100	5.117	7.9%	0.16 [-0.07, 0.39]		<b> </b>
Heterogeneity: Not app	olicable					
Test for overall effect:						
Total (95% CI)			100.0%	0.48 [0.38, 0.58]		•
Heterogeneity: Tau <sup>2</sup> =	0 02: Chi <sup>2</sup> - 37 74	df - 12		. , .		
Test for overall effect:			(1 0.000	00), 1 = 00 /0	-1 -0.5	0 0.5 1
			(D = 0.02)	12 - 72 10/		
Test for subgroup diffe	rences: $Chr = 7.42$	., ui = 2	(P = 0.02)	), 1 - 73.1%		

Figure 5. Pooled estimates of fat survival rate with non-cell-assisted lipotransfer at different recipient areas.

3.8%), which was explained that the overall economic situation of the CAL group is better and can bear the cost of the second operation. In addition, the perez-cano study<sup>25</sup> did not set a control, and the rate of patients undergoing secondary surgery reached 35.8% (24/67), which seriously affected the actual results of this index. After excluding these three studies, it was found that there was no difference in the incidence of multiple surgeries between the CAL group and the non-CAL group (1.85% [6/325] vs 0% [0/76], P = 0.5997).

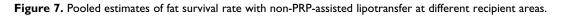
Our results indicated that the SVF separation method (mainly referring to automatic separation and manual separation) does not affect the fat survival rate (Fig. 8), which is consistent with Doi<sup>27</sup> and Laloze, J<sup>55</sup>. It showed that the automatic separation system is a reliable method, and it is expected that clinical trials based on enzymatically hydrolyzed fat particles will be carried out directly in small,

sterile facilities without the need for advanced cell laboratories. In addition, non-enzymatic mechanical separation of SVF and in vitro cultured ADSCs seemed to be a good choice to improve the survival of grafted fat. According to the included articles<sup>2,23,38,42</sup>, fat survival rates of adding with non-enzymatic mechanical separation of SVF and in vitro cultured ADSCs were higher than the average level of SVF in automatic and manual separation. No complications were observed and only underwent a single operation. Studies have analyzed the SVF obtained by enzymatic and nonenzymatic treatment, and found that they have similar cell yield, cell survival rate, and SVF composition, The nonenzymatic method consumes less time and lower cost than the enzymatic method, which is more suitable for clinical application<sup>56,57</sup>. However, only a small sample size was used to confirm this processing technique. In this sense, we hope to further evaluate the clinical safety of this processing

Study or Subgroup	Rate Difference	SF	Weight	Rate Difference IV, Random, 95% CI		fference om, 95% Cl
1.4.1 Face	Hute Billerende	02	meight		IV, Runde	
Cervelli 2013	0.7	0.039	37.7%	0.70 [0.62, 0.78]		-
Fontdevila 2014		0.121	3.9%	0.52 [0.28, 0.75]		
Gentile 2014		0.146		0.69 [0.40, 0.98]		
Keyhan 2013		0.077		• • •		
Sasaki 2015	0.685	0.045	28.3%			-
Subtotal (95% CI)			82.2%	0.70 [0.64, 0.76]		♦
1.4.2 Breast						
	0.740	0.440	4.00/	0.70 (0.40, 0.05)		
Fiaschetti 2013 Gentile 2013		0.116	4.3%			
Subtotal (95% CI)	0.69	0.065	13.6% <b>17.8%</b>	0.69 [0.56, 0.82] 0.70 [0.59, 0.81]		•
Heterogeneity: Tau <sup>2</sup> = Test for overall effect:		•				•
Total (95% CI)			100.0%	0.70 [0.65, 0.75]		•
Heterogeneity: Tau <sup>2</sup> = Test for overall effect:	0.00; Chi <sup>2</sup> = 4.90, d Z = 29.22 (P < 0.00		= 0.56); l	2 = 0%	-1 -0.5 (	0 0.5 1

Figure 6. Pooled estimates of fat survival rate with PRP-assisted lipotransfer at different recipient areas.

Study or Subgroup	Rate Difference	SE	Weight	Rate Difference IV, Random, 95% CI	Rate Difference IV. Random. 95% Cl
1.5.1 Face					
Cervelli 2013	0.31	0.146	5.9%	0.31 [0.02, 0.60]	
Fontdevila 2014	0.516	0.094	14.2%	0.52 [0.33, 0.70]	
Gentile 2014	0.39	0.154	5.3%	0.39 [0.09, 0.69]	
Sasaki 2015	0.383	0.051	48.3%	0.38 [0.28, 0.48]	
Subtotal (95% CI)			73.6%	0.40 [0.32, 0.48]	•
Heterogeneity: Tau <sup>2</sup> =	0.00; Chi <sup>2</sup> = 2.01, d	lf = 3 (P	= 0.57); I	<sup>2</sup> = 0%	
Test for overall effect: 2	Z = 9.77 (P < 0.000	001)			
1.5.2 Breast					
Gentile 2013	0.39	0.069	26.4%	0.39 [0.25, 0.53]	
Subtotal (95% CI)			26.4%	0.39 [0.25, 0.53]	•
Heterogeneity: Not app	olicable				
Test for overall effect: 2	Z = 5.65 (P < 0.000	001)			
Total (95% CI)			100.0%	0.40 [0.33, 0.47]	•
Heterogeneity: Tau <sup>2</sup> =	0.00; Chi <sup>2</sup> = 2.04, d	lf = 4 (P	= 0.73); I	<sup>2</sup> = 0%	-1 -0.5 0 0.5 1
Test for overall effect: 2	Z = 11.29 (P < 0.00	0001)			-1 -0.5 0 0.5 1
Test for subgroup diffe	rences: Chi <sup>2</sup> = 0.03	s, df = 1	(P = 0.87	), $I^2 = 0\%$	



technology. If the outcome is good, we will not need to use enzymatic hydrolysis or *in vitro* amplified stem cells. And the non-enzymatic technology will greatly facilitate the implementation of such surgery, improve the survival of grafted fat and reduce the cost of the operation.

# Which One Is Better, Cell-Assisted or PRP-Assisted Lipotransfer?

In order to better promote the development of clinical autologous fat grafting surgery, improve its survival rate, reduce

				Rate Difference	Rate Difference
Study or Subgroup	Rate Difference	SE	Weight	IV, Random, 95% CI	IV, Random, 95% CI
1.6.1 Manual					
Chang 2013	0.683	0.147	2.1%	0.68 [0.39, 0.97]	
Chiu 2018	0.687	0.046	20.0%	0.69 [0.60, 0.78]	-
Gentile 2014	0.63	0.153	1.9%	0.63 [0.33, 0.93]	
Li 2013	0.648	0.094	5.0%	0.65 [0.46, 0.83]	
Schendel 2015	0.68	0.148	2.1%	0.68 [0.39, 0.97]	
Tanikawa 2013	0.88	0.123	3.0%	0.88 [0.64, 1.12]	
Tissiani 2016	0.79	0.123	3.0%	0.79 [0.55, 1.03]	
Wang 2015	0.482	0.144	2.2%	0.48 [0.20, 0.76]	
Yoshimura 2010	0.57	0.128	2.7%	0.57 [0.32, 0.82]	
Subtotal (95% CI)			41.9%	0.68 [0.62, 0.75]	● ●
Heterogeneity: Tau <sup>2</sup> =	0.00; Chi <sup>2</sup> = 6.32, c	lf = 8 (P	= 0.61); I	<sup>2</sup> = 0%	
Test for overall effect:	Z = 21.15 (P < 0.00	001)			
1.6.2 Automatic					
Dos Anjos 2015	0.75	0.058	12.9%	0.75 [0.64, 0.86]	-
Gentile 2012	0.63	0.153	1.9%	0.63 [0.33, 0.93]	
Gentile 2015	0.63	0.153	1.9%	0.63 [0.33, 0.93]	
Gentile 2019	0.708	0.041	24.7%	0.71 [0.63, 0.79]	
Jung 2016	0.47	0.223	0.9%	0.47 [0.03, 0.91]	
Peltoniemi 2013	0.505	0.158	1.8%	0.51 [0.20, 0.81]	· · · · ·
Sasaki 2015	0.729	0.148	2.1%	0.73 [0.44, 1.02]	
Subtotal (95% CI)			46.2%	0.70 [0.64, 0.76]	♦
Heterogeneity: Tau <sup>2</sup> = Test for overall effect:		•	= 0.70); l	<sup>2</sup> = 0%	
1.6.3 Mechanical					
Gontijo 2017	0.904	0.076	7.6%	0.90 [0.76, 1.05]	
Subtotal (95% CI)			7.6%	0.90 [0.76, 1.05]	•
Heterogeneity: Not app	olicable				
Test for overall effect:	Z = 11.89 (P < 0.00	001)			
1.6.4 ADSCs					
Koh 2012	0.794	0.181	1.4%	0.79 [0.44, 1.15]	
Kolle 2013		0.124	2.9%	0.81 [0.57, 1.05]	
Subtotal (95% CI)			4.3%	0.80 [0.60, 1.00]	
Heterogeneity: Tau <sup>2</sup> =	0.00; Chi <sup>2</sup> = 0.00, c	lf = 1 (P	= 0.95); 1		
Test for overall effect:					
Total (95% CI)			100.0%	0.71 [0.67, 0.75]	•
Heterogeneity: Tau <sup>2</sup> =	0.00; Chi <sup>2</sup> = 18.30.	df = 18	(P = 0.44)	): $ ^2 = 2\%$ —	
Test for overall effect:			,		1 -0.5 0 0.5 1
Test for subaroup diffe		,	(D - 0.04)	12 - 62 20/	

Figure 8. Pooled estimates of fat survival rates for different handling methods of cell-assisted lipotransfer.

its complications and the number of multiple operations. We tried to analyze related articles containing CAL and PRPassisted lipotransfer for more accurately apply related assisted fat grafting technology in the future.

Many studies have confirmed that traditional autologous fat grafting is safe for breast reconstruction after breast cancer surgery<sup>58,59</sup>, but adipose-derived stem cells have been shown to promote cancer progression in pre-clinical

studies<sup>60,61</sup>. Which has caused concern about the safety of cell-assisted fat grafting in breast reconstruction after breast cancer surgery. Most recently Simon Gebremeskel et al.<sup>62</sup> observed that simple adipose-derived stem cells can promote the proliferation and invasiveness of breast cancer cells *in vivo* and *in vitro*, but cell-assisted fat grafting will not promote the development of breast cancer. In this review, only two articles reported breast cancer recurrence after breast reconstruction

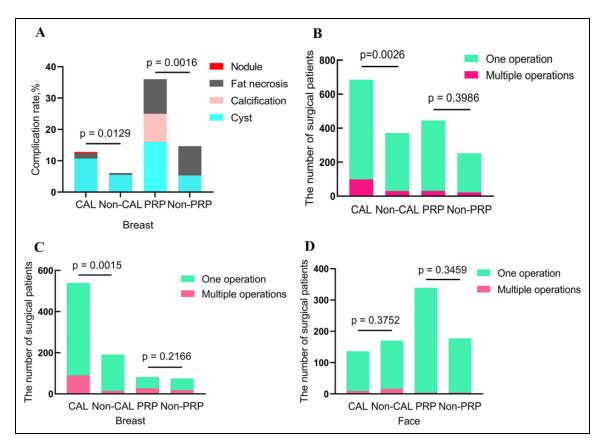


Figure 9. (A) Complication rates of different interventions; (B) All patients were summarized for having complications or not; (C) A summary of all patients undergo facial surgery with single and multiple operations; (D) A summary of all patients undergo breast surgery with single and multiple operations.

surgery. In the study of Perez-Cano et al.<sup>25</sup>, there was a patient with pelvic bone metastasis after fat grafting, which was considered to be a natural progression of the disease and had nothing to do with the use of CAL. In the study by Gentile et al.<sup>41</sup>, there was no statistical difference between CAL group and the traditional fat grafting group in terms of local recurrence and systemic recurrence and think that cell-assisted or traditional fat grafting is not a risk factor for recurrence. Therefore, there is currently no definite evidence that performing cell-assisted fat grafting on breast cancer patients will increase the risk of breast cancer recurrence. Instead, more clinical data support that the technology is safe and effective<sup>63,64</sup>.

PRP which is easy to implement, does not affect the total operation time, does not require special laboratory equipment and haven't side effects been reported, so its role in fat grafting is getting more and more attention<sup>43</sup>. Several articles in the included studies mentioned that PRP-assisted lipotransfer shortened postoperative recovery time<sup>39,47,48,50</sup>, which may be due to the increased amount of PDGFs. These cytokines and growth factors are involved in homing, migration, proliferation and differentiation of various cells. Its highly concentrated growth factors can promote the production of collagen and fibronectin, promote angiogenesis, accelerate wound healing and shorten recovery time<sup>65</sup>.

Based on our meta-analysis results and other data from the 36 studies we included, we recommend to use the CAL techniques in large-volume fat grafting, such as breasts and buttocks fat grafting. Although the forest plot results proved that there was no statistical difference between the CAL and PRP-assisted lipotransfer in promoting the survival of grafted fat, no matter whether it was applied to the largevolume or the small-volume fat grafting, we mainly consider the problem of the excessive volume of autologous whole blood required due to the demand for PRP in large-volume fat grafting. We summarized data from 11 included studies that included the ratio of whole blood to PRP or the ratio of PRP to grafted fat (Appendix 2). In addition, we calculated the average value of the injected fat volume of the unilateral breast in the included breast fat grafting articles, which was 219 ml. It can be seen from Appendix 2 that the ratio of whole blood to PRP or the ratio of PRP to grafted fat was different in different articles. Therefore, the proportional relationship between PRP and grafted fat has led to the need for total blood volume as an issue of concern. We choose a moderate ratio from Appendix 2, assuming that the ratios of whole blood to PRP and PRP to grafted fat are 10:1 and 1:5, respectively, and which is the most appropriate ratio. For a patient who wants a breast filling, supposing 200 ml of fat is needed for one-side and 400 ml for both sides. Then 80 ml of

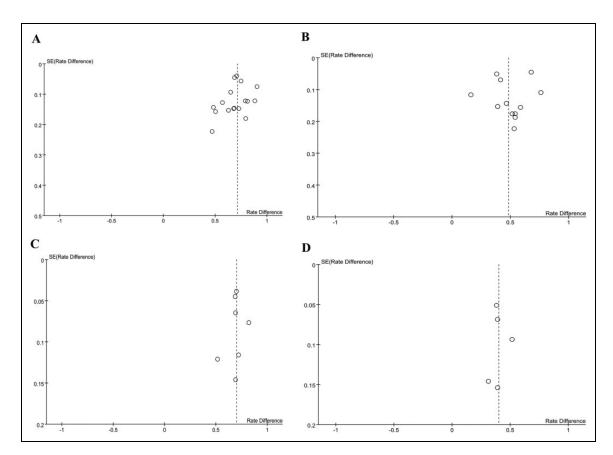


Figure 10. Funnel plot analysis of publication bias for studies on (A) CAL; (B) non-CAL; (C) PRP; (D) non-PRP.

PRP would be required. Which means it is necessary to draw 800 ml of whole blood<sup>34,44,49</sup>. This will limit the application of PRP in fat grafting because it will pose a threat to the health of patients. For small-volume fat grafting areas such as face and arm (grafted fat volume <100 ml), we recommend to use PRP-assisted lipotransfer technology. Because based on the above ratio, the amount of whole blood we need will not exceed 200 ml. Furthermore, CAL consumes longer operation time than PRP because SVF requires enzymatic separation or mechanical separation. In addition, compared to SVF, PRP has the advantages of low operational difficulty, short operation time and postoperative recovery time<sup>39,47,48,50,65</sup>. So PRP is superior to SVF in small volume fat grafting.

### Limitation

Our study has the following limitations: First, the objective measurement methods for postoperative volume measurement and follow-up of complications were not thorough. Although some studies have confirmed that there was no difference among different objective measurement methods for measuring fat volume<sup>32,66</sup>, the fat volume retention rate obtained by palpation, comparison with immediate post-operative photos and follow-up photos were somewhat subjective<sup>47</sup>. And different objective measurement methods have differences in the detection rate of complications<sup>45</sup>.

Therefore, it is hoped that in the future, the objective measurement methods related to the measurement of fat survival volume and the detection of complications will be unified as much as possible, so as to make different studies more comparable. Second, the optimal concentration of enriched SVF cells/ADSCs and PRP in improving the survival of grafted fat cannot be obtained. Because the preparation equipment and methods used in different studies are not uniform, and the information provided is insufficient. So it is hard to compare each other between different studies. Therefore, in order to further improve the research of cell-assisted and PRP-assisted lipotransfer, in the future, we need more complete large samples, objective volume and complication measurement methods, standard and unified preparation equipments and procedures, complete data information recording and follow-up plans in randomized controlled clinical research to further determine the true level of various indicators under clinical conditions and to obtain the optimal use concentration of cell-assisted and PRP-assisted lipotransfer, which have great significance for its conversion into clinical practice.

### Conclusions

In this study, we confirmed that both CAL and PRP-assisted lipotransfer can significantly improve the survival rate of grafted fat. But in large-volume fat grafting, such as breast reconstruction, both increased the incidence of complications to a certain extent. We recommend to use CAL for breasts, buttocks and other parts that require large-volume fat grafting, and PRP-assisted lipotransfer for parts that require small-volume fat grafting such as face and arm. Finally, the optimal concentration of SVF/ADSCs and PRP is still a very important and urgent issue. Future studies should address these issues through high-quality multicenter randomized controlled clinical studies and use objective measurements to evaluate the results.

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### **Supplemental Material**

Supplemental material for this article is available online.

### References

- Abboud MH, Dibo SA. Immediate large-volume grafting of autologous fat to the breast following implant removal. Aesthet Surg J. 2015;35(7):819–829.
- Kølle S-FT, Fischer-Nielsen A, Mathiasen AB, Elberg JJ, Oliveri RS, Glovinski PV, Kastrup J, Kirchhoff M, Rasmussen BS, Talman M-LMThomsen C, et al. Enrichment of autologous fat grafts with ex-vivo expanded adipose tissue-derived stem cells for graft survival: a randomised placebo-controlled trial. The Lancet. 2013;382(9898):1113–1120.
- O'Halloran N, Courtney D, Kerin MJ, Lowery AJ. Adiposederived stem cells in novel approaches to breast reconstruction: their suitability for tissue engineering and oncological safety. Breast Cancer (Auckl). 2017;11:1178223417726777.
- Wang L, Luo X, Lu Y, Fan ZH, Hu X. Is the resorption of grafted fat reduced in cell-assisted lipotransfer for breast augmentation? Ann Plast Surg. 2015;75(2):128–134.
- Matsumoto D, Sato K, Gonda K, Takaki Y, Shigeura T, Sato T, Aiba-Kojima E, Iizuka F, Inoue K, Suga H, Yoshimura K. Cellassisted lipotransfer: supportive use of human adipose-derived cells for soft tissue augmentation with lipoinjection. Tissue Eng. 2006;12(12):3375–3382.
- 6. Yoshimura K, Sato K, Aoi N, Kurita M, Hirohi T, Harii K. Cell-assisted lipotransfer for cosmetic breast augmentation:

supportive use of adipose-derived stem/stromal cells. Aesthetic Plast Surg. 2008;32(1):48–55; discussion 56–57.

- Gentile P, Orlandi A, Scioli MG, Di Pasquali C, Bocchini I, Cervelli V. Concise review: adipose-derived stromal vascular fraction cells and platelet-rich plasma: basic and clinical implications for tissue engineering therapies in regenerative surgery. Stem Cells Transl Med. 2012;1(3):230–236.
- Gimble JM, Katz AJ, Bunnell BA. Adipose-derived stem cells for regenerative medicine. Circ Res. 2007;100(9):1249–1260.
- Suga H, Eto H, Shigeura T, Inoue K, Aoi N, Kato H, Nishimura S, Manabe I, Gonda K, Yoshimura K. IFATS collection: Fibroblast growth factor-2-induced hepatocyte growth factor secretion by adipose-derived stromal cells inhibits postinjury fibrogenesis through a c-Jun N-terminal kinase-dependent mechanism. Stem Cells. 2009;27(1):238–249.
- Liu ZJ, Zhuge Y, Velazquez OC. Trafficking and differentiation of mesenchymal stem cells. J Cell Biochem. 2009;106(6): 984–991.
- 11. Bora P, Majumdar AS. Adipose tissue-derived stromal vascular fraction in regenerative medicine: a brief review on biology and translation. Stem Cell Res Ther. 2017;8(1):145.
- Blanton MW, Hadad I, Johnstone BH, Mund JA, Rogers PI, Eppley BL, March KL. Adipose stromal cells and platelet-rich plasma therapies synergistically increase revascularization during wound healing. Plast Reconstr Surg. 2009;123(2 Suppl): 56S–64S.
- Fukaya Y, Kuroda M, Aoyagi Y, Asada S, Kubota Y, Okamoto Y, Nakayama T, Saito Y, Satoh K, Bujo H. Platelet-rich plasma inhibits the apoptosis of highly adipogenic homogeneous preadipocytes in an *in vitro* culture system. Exp Mol Med. 2012; 44(5):330–339.
- Lynch MD, Bashir S.Applications of platelet-rich plasma in dermatology: a critical appraisal of the literature. J Dermatolog Treat. 2016;27(3):285–289.
- Moher D, Liberati A, Tetzlaff J, Altman DG, Group TP. Preferred reporting items for systematic reviews and metaanalyses: The PRISMA statement. PLoS Med. 2009;62(10): 1006–1012.
- The Oxford 2011 Levels of Evidence. Oxford centre for evidence-based medicine [Internet]. 2009. [cited 2020 Feb 6]. Available from http://www.cebm.net/index.aspxo=5653.
- Higgins JPT, Altman DG, Gotzsche PC, Juni P, Moher D, Oxman AD, Savovic J, Schulz KF, Weeks L, Sterne JACCochrane Bias Methods Group. The Cochrane Collaboration's tool for assessing risk of bias in randomised trials. BMJ. 2011;343:d5928–d5928.
- Yoshimura K, Asano Y, Aoi N, Kurita M, Oshima Y, Sato K, Inoue K, Suga H, Eto H, Kato H, Harii K. Progenitor-enriched adipose tissue transplantation as rescue for breast implant complications. Breast J. 2010;16(2):169–175.
- Kamakura T, Ito K. Autologous cell-enriched fat grafting for breast augmentation. Aesthet Plast Surg. 2011;35(6): 1022–1030.
- Sterodimas A, de Faria J, Nicaretta B, Boriani F. Autologous fat transplantation versus adipose-derived stem cell-enriched lipografts: a study. Aesthet Surg J. 2011;31(6):682–693.

- Tiryaki T, Findikli N, Tiryaki D. Staged stem cell-enriched tissue (set) injections for soft tissue augmentation in hostile recipient areas: a preliminary report. Aesthet Plast Surg. 2011;35(6):965–971.
- 22. Gentile P, Orlandi A, Scioli MG, Di Pasquali C, Bocchini I, Curcio CB, Floris M, Fiaschetti V, Floris R, Cervell V. A comparative translational study: the combined use of enhanced stromal vascular fraction and platelet-rich plasma improves fat grafting maintenance in breast reconstruction. Stem Cells Transl Med. 2012;1(4):341–351.
- 23. Koh KS, Oh TS, Kim H, Chung IW, Lee KW, Lee HB, Park EJ, Jung JS, Shin IS, Ra JC, Choi JW. Clinical application of human adipose tissue-derived mesenchymal stem cells in progressive hemifacial atrophy (Parry-Romberg disease) with microfat grafting techniques using 3-dimensional computed tomography and 3-dimensional camera. Ann Plast Surg. 2012;69(3):331–337.
- Lee SK, Kim DW, Dhong ES, Park SH, Yoon ES. Facial Soft Tissue Augmentation using Autologous Fat Mixed with Stromal Vascular Fraction. Arch Plast Surg. 2012;39(5):534–539.
- 25. Pérez-Cano R, Vranckx JJ, Lasso JM, Calabrese C, Merck B, Milstein AM, Sassoon E, Delay E, Weiler-Mithoff EM. Prospective trial of Adipose-Derived Regenerative Cell (ADRC)enriched fat grafting for partial mastectomy defects: The RESTORE-2 trial. Eur J Surg Oncol (EJSO). 2012;38(5): 382–389.
- Chang Q, Li J, Dong Z, Liu L, Lu F. Quantitative volumetric analysis of progressive hemifacial atrophy corrected using stromal vascular fraction-supplemented autologous fat grafts. Dermatol Surg. 2013;39(10):1465–1473.
- Doi K, Tanaka S, Iida H, Eto H, Kato H, Aoi N, Kuno S, Hirohi T, Yoshimura K. Stromal vascular fraction isolated from lipoaspirates using an automated processing system: bench and bed analysis. J Tissue Eng Regen Med. 2013;7(11):864–870.
- Li J, Gao J, Cha P, Chang Q, Liao Y, Liu C, Li K, Lu F. Supplementing fat grafts with adipose stromal cells for cosmetic facial contouring. Dermatol Surg. 2013;39(3 Pt 1): 449–456.
- Peltoniemi HH, Salmi A, Miettinen S, Mannerstrom B, Saariniemi K, Mikkonen R, Kuokkanen H, Herold C. Stem cell enrichment does not warrant a higher graft survival in lipofilling of the breast: a prospective comparative study. J Plast Reconstr Aesthet Surg. 2013;66(11):1494–1503.
- Tanikawa DYS, Aguena M, Bueno DF, Passos-Bueno MR, Alonso N. Fat Grafts Supplemented with Adipose-Derived Stromal Cells in the Rehabilitation of Patients with Craniofacial Microsomia. Plast Reconstr Surg. 2013;132(1):141–152.
- 31. Gentile P, De Angelis B, Pasin M, Cervelli G, Curcio CB, Floris M, Di Pasquali C, Bocchini I, Balzani A, Nicoli F, Insalaco C, et al. Adipose-derived stromal vascular fraction cells and platelet-rich plasma: basic and clinical evaluation for cell-based therapies in patients with scars on the face. J Craniofac Surg. 2014;25(1):267–272.
- 32. Dos Anjos S, Matas-Palau A, Mercader J, Katz AJ, Llull R. Reproducible volume restoration and efficient long-term volume retention after point-of-care standardized cell-enhanced

fat grafting in breast surgery. Plast Reconstr Surg Glob Open. 2015;3(10):e547.

- Gentile P, Scioli MG, Orlandi A, Cervelli V. Breast reconstruction with enhanced stromal vascular fraction fat grafting: what is the best method? Plast Reconstr Surg Glob Open. 2015;3(6): e406.
- Sasaki GH.The safety and efficacy of cell-assisted fat grafting to traditional fat grafting in the anterior mid-face: an indirect assessment by 3d imaging. Aesthetic Plast Surg. 2015;39(6): 833–846.
- Schendel SA. Enriched autologous facial fat grafts in aesthetic surgery: 3d volumetric results. Aesthet Surg J. 2015;35(8): 913–919.
- Jung HK, Kim CH, Song SY. Prospective 1-year follow-up study of breast augmentation by cell-assisted lipotransfer. Aesthet Surg J. 2016;36(2):179–190.
- Tissiani LA, Alonso N. A prospective and controlled clinical trial on stromal vascular fraction enriched fat grafts in secondary breast reconstruction. Stem Cells Int. 2016;2016:2636454.
- Gontijo-de-Amorim NF, Charles-de-Sa L, Rigotti G. Mechanical supplementation with the stromal vascular fraction yields improved volume retention in facial lipotransfer: a 1-year comparative study. Aesthet Surg J. 2017;37(9):975–985.
- Willemsen JCN, Van Dongen J, Spiekman M, Vermeulen KM, Harmsen MC, van der Lei B, Stevens HPJ. The addition of platelet-rich plasma to facial lipofilling: a double-blind, placebo-controlled, randomized trial. Plast Reconstr Surg. 2018;141(2):331–343.
- 40. Chiu CH. Does stromal vascular fraction ensure a higher survival in autologous fat grafting for breast augmentation? A volumetric study using 3-dimensional laser scanning. Aesthet Surg J. 2019;39(1):41–52.
- 41. Gentile P, Casella D, Palma E, Calabrese C. Engineered fat graft enhanced with adipose-derived stromal vascular fraction cells for regenerative medicine: clinical, histological and instrumental evaluation in breast reconstruction. J Clin Med. 2019;8(4):504.
- Gontijo-de-Amorim NF, Charles-de-Sa L, Rigotti G. Fat grafting for facial contouring using mechanically stromal vascular fraction-enriched lipotransfer. Clin Plast Surg. 2020;47(1): 99–109.
- 43. Salgarello M, Visconti G, Rusciani A. Breast fat grafting with platelet-rich plasma: a comparative clinical study and current state of the art. Plast Reconstr Surg. 2011;127(6):2176–2185.
- 44. Cervelli V, Bocchini I, Di Pasquali C, De Angelis B, Cervelli G, Curcio CB, Orlandi A, Scioli MG, Tati E, Delogu P, Gentile P. P.R.L. platelet rich lipotransfert: our experience and current state of art in the combined use of fat and PRP. Biomed Res Int. 2013;2013:434191.
- 45. Fiaschetti V, Pistolese CA, Fornari M, Liberto V, Cama V, Gentile P, Floris M, Floris R, Cervelli V, Simonetti G. Magnetic resonance imaging and ultrasound evaluation after breast autologous fat grafting combined with platelet-rich plasma. Plast Reconstr Surg. 2013;132(4):498e–509e.
- 46. Gentile P, Di Pasquali C, Bocchini I, Floris M, Eleonora T, Fiaschetti V, Floris R, Cervelli V. Breast reconstruction with

autologous fat graft mixed with platelet-rich plasma. Surg Innov. 2013;20(4):370-376.

- Keyhan SO, Hemmat S, Badri AA, Abdeshahzadeh A, Khiabani K. Use of platelet-rich fibrin and platelet-rich plasma in combination with fat graft: which is more effective during facial lipostructure? J Oral Maxillofac Surg. 2013;71(3): 610–621.
- Willemsen JC, Lindenblatt N, Stevens HP. Results and longterm patient satisfaction after gluteal augmentation with platelet-rich plasma-enriched autologous fat. Eur J Plast Surg. 2013;36(12):777–782.
- Fontdevila J, Guisantes E, Martínez E, Prades E, Berenguer J. Double-blind clinical trial to compare autologous fat grafts versus autologous fat grafts with PDGF. Plast Reconstr Surg. 2014;134(2):219e–230e.
- 50. Willemsen JC, van der Lei B, Vermeulen KM, Stevens HP. The effects of platelet-rich plasma on recovery time and aesthetic outcome in facial rejuvenation: preliminary retrospective observations. Aesthet Plast Surg. 2014;38(5):1057–1063.
- Ozer K, Colak O. Micro-autologous fat transplantation combined with platelet-rich plasma for facial filling and regeneration: a clinical perspective in the shadow of evidence-based medicine. J Craniofac Surg. 2019;30(3):672–677.
- 52. Gentile P, Piccinno MS, Calabrese C. Characteristics and potentiality of human adipose-derived stem cells (hascs) obtained from enzymatic digestion of fat graft. Cells. 2019; 8(3):282.
- 53. Yoshimura K, Eto H, Kato H, Suga H, Aoi N, Doi K, Kuno S. The fate of adipocytes after nonvascularized fat grafting: evidence of early death and replacement of adipocytes. Plast Reconstr Surg. 2012;129(5):1081–1092.
- 54. Wei H, Gu S-X, Liang Y-D, ZhuXu F-T, He N, Liang Z-J, Chen H, Wei X-J, Li H-M. Nanofat-derived stem cells with platelet-rich fibrin improve facial contour remodeling and skin rejuvenation after autologous structural fat transplantation. Oncotarget. 2017;8(40):68542–68556.
- 55. Laloze J, Varin A, Gilhodes J, Bertheuil N, Grolleau JL, Brie J, Usseglio J, Sensebe L, Filleron T, Chaput B.Cell-assisted lipotransfer: friend or foe in fat grafting? Systematic review and metaanalysis. J Tissue Eng Regen Med. 2018;12(2):e1237–e1250.
- Mashiko T, Wu SH, Feng J, Kanayama K, Kinoshita K, Sunaga A, Narushima M, Yoshimura K. Mechanical micronization of lipoaspirates: squeeze and emulsification techniques. Plast Reconstr Surg. 2017;139(1):79–90.

- 57. van Dongen JA, Tuin AJ, Spiekman M, Jansma J, van der Lei B, Harmsen MC. Comparison of intraoperative procedures for isolation of clinical grade stromal vascular fraction for regenerative purposes: a systematic review. J Tissue Eng Regen Med. 2018;12(1):e261–e274.
- Kronowitz SJ, Mandujano CC, Liu J, Kuerer HM, Smith B, Garvey P, Jagsi R, Hsu L, Hanson S, Valero V. Lipofilling of the breast does not increase the risk of recurrence of breast cancer: a matched controlled study. Plast Reconstr Surg. 2016;137(2):385–393.
- Waked K, Colle J, Doornaert M, Cocquyt V, Blondeel P.Systematic review: The oncological safety of adipose fat transfer after breast cancer surgery. The Breast. 2017;31:128–136.
- 60. Zhang Y, Daquinag AC, Amaya-Manzanares F, Sirin O, Tseng C, Kolonin MG. Stromal progenitor cells from endogenous adipose tissue contribute to pericytes and adipocytes that populate the tumor microenvironment. Cancer Res. 2012;72(20): 5198–5208.
- Kamat P, Schweizer R, Kaenel P, Salemi S, Calcagni M, Giovanoli P, Gorantla VS, Eberli D, Andres AC, Plock JA. Human adipose-derived mesenchymal stromal cells may promote breast cancer progression and metastatic spread. Plast Reconstr Surg. 2015;136(1):76–84.
- 62. Gebremeskel S, Gencarelli J, Gareau AJ, Levatte T, Dugandzic A, Johnston B, Bezuhly M. Promotion of primary murine breast cancer growth and metastasis by adipose-derived stem cells is reduced in the presence of autologous fat graft. Plast Reconstr Surg. 2019;143(1):137–147.
- Bateman ME, Strong AL, Gimble JM, Bunnell BA. Concise review: using fat to fight disease: a systematic review of nonhomologous adipose-derived stromal/stem cell therapies. Stem Cells. 2018;36(9):1311–1328.
- Knackstedt RW, Gatherwright J, Ataya D, Duraes EFR, Schwarz GS. Fat grafting and the palpable breast mass in implant-based breast reconstruction. Plast Reconstr Surg. 2019;144(2):265–275.
- El-Sharkawy H, Kantarci A, Deady J, Hasturk H, Liu H, Alshahat M, Van Dyke TE. Platelet-rich plasma: growth factors and pro- and anti-inflammatory properties. J Periodontol. 2007; 78(4):661–669.
- Zhou Y, Wang J, Li H, Liang X, Bae J, Huang X, Li Q. Efficacy and safety of cell-assisted lipotransfer. Plast Reconstr Surg. 2016;137(1):44e–57e.