



The Role of the Effects of Endoplasmic Reticulum Stress on NLRP3 Inflammasome in Diabetes

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Endoplasmic reticulum (ER) is an important organelle for the protein synthesis, modification, folding, assembly, and the transport of new peptide chains. When the folding ability of ER proteins is impaired, the accumulation of unfolded or misfolded proteins in ER leads to endoplasmic reticulum stress (ERS). The nucleotidebinding oligomerization domain-like receptor family, pyrin domain-containing 3 (NLRP3) inflammasome, can induce the maturation and secretion of interleukin-1beta (IL-1 β) and IL-18 through activating caspase-1. It is associated with many diseases. Studies have shown that ERS can regulate NLRP3 inflammasome in many diseases including diabetes. However, the mechanism of the effects of ERS on NLRP3 inflammasome in diabetes has not been fully understood. This review summarizes the recent researches about the effects of ERS on NLRP3 inflammasome and the related mechanism in diabetes to provide ideas for the relevant basic research in the future.

OPEN ACCESS

Edited by:

Giuseppe Calamita, University of Bari Aldo Moro, Italy

Reviewed by:

Tiziano Verri, University of Salento, Italy Onofrio Laselva, University of Foggia, Italy

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Specialty section:

This article was submitted to Cellular Biochemistry, a section of the journal Frontiers in Cell and Developmental Biology

> Received: 04 February 2021 Accepted: 15 March 2021 Published: 14 April 2021

Citation:

Lv S, Li X and Wang H (2021) The Role of the Effects of Endoplasmic Reticulum Stress on NLRP3 Inflammasome in Diabetes. Front. Cell Dev. Biol. 9:663528. doi: 10.3389/fcell.2021.663528 Keywords: endoplasmic reticulum stress, NLRP3 inflammasome, diabetes, diabetic encephalopathy, diabetic nephropathy

INTRODUCTION

Diabetes is characterized by the destruction of glucose homeostasis and the deficiency of insulin effect on the liver, muscle, pancreas, and fat (Sarwar et al., 2010; Defronzo et al., 2015). Diabetes can be divided into type 1 and type 2 diabetes. Type 1 diabetes accounts for about 5–10% of all diabetics, which is due to β -cell dysfunction, decreased insulin release, and decreased circulating insulin level. Type 2 diabetes is the most common type of diabetes, accounting for about 90–95% of all diabetics. It is mainly related to the insufficiencies of insulin response and insulin resistance of surrounding tissues (Amer Diabet, 2014). There were 366 million diabetics in 2011, and it is estimated that the total number of diabetics will increase to 552 million by 2030 (Whiting et al., 2011). Long-term diabetes can lead to multiple organ dysfunction, including kidney disease, retinopathy, neuropathy, atherosclerosis, and heart disease (Zheng et al., 2018; Inzucchi et al., 2019). Therefore, it is particularly urgent to find ways to treat and prevent diabetes complications.

Inflammasome, composed of many proteins, is an important part of the innate immune system and used to detect whether there are infections, pathogens, and metabolic alarms in cells (Elliott and Sutterwala, 2015; Jo et al., 2016). Inflammasomes have been identified to include NLRP1, NLRC4, RIG-I, AIM2, and NLRP3 (Barbe et al., 2014). NLRP3 inflammasome, the most deeply studied one, is composed of NLRP3, the adaptor protein ASC, and pre-caspase-1. It is mainly expressed in bone marrow cells, such as macrophages (O'connor et al., 2003). When the host is stimulated by the exogenous or endogenous stimuli, NLRP3 inflammasome is activated, leading to the recruitment of pre-caspase-1 and the adaptor protein ASC in macrophages. Stimulated NLRP3 interacts with pre-caspase-1 and the adaptor protein ASC to form a large cytoplasmic complex, thus activating caspase-1. Activated caspase-1 induces inflammation by cleaving pre-interleukin (IL)-1β and pre-IL-18 into IL-1β and IL-18 (Prochnicki et al., 2016). NLRP3 inflammasome is activated in two steps: the first signal (signal 1) indicating infection or tissue damage includes toll-like receptor 4, a pattern recognition receptor that can recognize lipopolysaccharide (LPS) and a series of endogenous risk signals, which activate nuclear factor kappa B (NF-κB) to increase the protein expression of NLRP3, pro-IL-1β, and pro-IL-18. Then, the second signal (signal 2), indicating cell damage, includes extracellular adenosine triphosphate (ATP), urate, and cholesterol crystals. It induces the assembly of inflammasome, activates caspase-1, and cleaves pre-IL-18 and pre-IL-1β into their active forms (Sokolova et al., 2019). The abnormal activation of NLRP3 inflammasome can lead to a variety of diseases including sterile inflammatory diseases (Abderrazak et al., 2015), diabetes (Luo et al., 2014), and non-alcoholic fatty liver diseases (Lv et al., 2019). In the hyperglycemic environment, the increase in NLRP3 inflammasome-dependent IL-1ß secretion leads to the dysfunction of insulin secretion of β cells, promotes obesity and insulin resistance, and eventually leads to diabetes (Iannantuoni et al., 2019).

Endoplasmic reticulum (ER) is an important organelle, responsible for the protein synthesis, modification, folding, assembly, and transport (Han and Kaufman, 2016). In various stress states (including glucose deficiency, environmental toxins, viral infection, Ca²⁺ level changes, hypoxia, inflammation, and oxidative stress), ER homeostasis is destroyed, which triggers ER stress (ERS). ERS refers to the dysfunction of ER, which interferes with the protein folding, posttranslational modification, and secretion. The unfolded protein reaction (UPR) is initiated by the accumulation of unfolded protein in ER (Liu et al., 2017; Lan et al., 2019), which improves protein folding, promotes quality control mechanisms and degradation pathways, or activates apoptosis when the damage is irreversible (Hetz and Saxena, 2017). In addition to UPR, the ubiquitin-proteasome system (UPS) is another intracellular machinery for the unfolded protein degradation, in which the target proteins are first polyubiquitinated before being degraded (Caldeira et al., 2014). Under ERS, there are three parallel signaling pathways in UPR: pancreatic endoplasmic reticulum kinase (PERK)-mediated pathway, inositol-dependent enzyme 1 (IRE1)-mediated pathway, and activating transcription factor 6 (ATF6)-mediated pathway. Moderately activated ERS and UPR can promote the steady-state recovery of ER and help cells adapt to environmental changes. Under excessive and sustained stress, ERS and UPR can activate caspase-12 and CCAAT/enhancer-binding protein homologous protein (CHOP)-dependent apoptosis pathways and promote cell death, thus participating in the occurrence of various diseases (Ji et al., 2019). Studies have shown that ERS is closely related to NLRP3 inflammasome in many diseases including diabetes (Ji et al., 2019). However, the mechanism of the effects of ERS on NLRP3

inflammasome in diabetes has not been fully understood. In this review, the effects of ERS on NLRP3 inflammasome and its mechanism are explored in diabetes to provide ideas for the relevant basic research in the future.

THE ROLE OF THE EFFECTS OF ERS ON NLRP3 INFLAMMASOME IN ENDOTHELIAL DYSFUNCTION OF DIABETES

Vascular endothelial dysfunction is the main pathophysiological cause of diabetic vascular complications. Glucose toxicity plays a key role in the occurrence of diabetic endothelial dysfunction (Giaccari et al., 2009). Mangiferin, a xanthic acid, is found in mangoes and Anemarrhena asphodeloides Bunge. It has been widely used in traditional Chinese medicine for the treatment of diabetes by reducing blood sugar and improving blood lipid in patients with diabetes (Muruganandan et al., 2005; Marquez et al., 2012; Gong et al., 2013; Wang et al., 2014). Song et al. (2015) established ERS-related endothelial dysfunction model by using high glucose (HG) to stimulate endothelial cells and conducted a series of experiments. The results showed that in endothelial cells, the level of AMP-activated protein kinase (AMPK) phosphorylation was increased by Mangiferin. Mangiferin and AMPK activator (AICAR) inhibited IRE1a-mediated ERS and ROS production induced by HG, while Compound C, an AMPK inhibitor, abolished Mangiferin effects, suggesting that Mangiferin suppressed HG-induced ERS and oxidative stress by activating AMPK. Moreover, Mangiferin suppressed apoptosis, NLRP3 inflammasome activation, and the resultant increased level of IL-1ß induced by HG, while compound C reversed the changes, indicating that Mangiferin suppressed HG-induced NLRP3 inflammasome activation and apoptosis of endothelial cells by activating AMPK (Song et al., 2015). Thioredoxininteracting protein (TXNIP) connects ERS with NLRP3 inflammasome activation (Oslowski et al., 2012). Mangiferin and tauroursodeoxycholic acid (TUDCA, an ER stress inhibitor) suppressed HG-induced TXNIP expression, while Compound C reduced the inhibitory effects of Mangiferin on TXNIP expression. Collectively, it can be inferred that Mangiferin inhibited HG-induced TXNIP/NLRP3 inflammasome activation through suppressing ERS by activating AMPK to improve endothelial dysfunction (Song et al., 2015). In the above studies, the relationship among ERS, NLRP3 inflammasome, TXNIP, reactive oxygen species (ROS), and apoptosis needs to be further clarified.

THE ROLE OF THE EFFECTS OF ERS ON NLRP3 INFLAMMASOME IN ADIPOSE DYSFUNCTION OF DIABETES

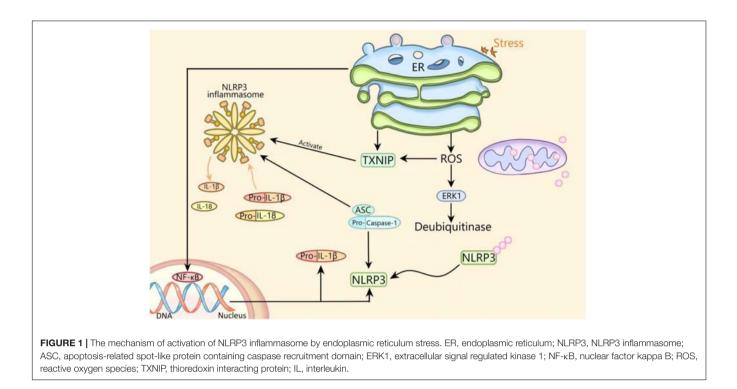
In diabetes, hyperglycemia usually leads to chronic inflammation by increasing macrophage infiltration and oxidative stress in adipose tissue (Zeyda and Stulnig, 2007; Kawasaki et al., 2012). ERS can induce inflammation mediated by NLRP3 inflammasome (Lerner et al., 2012) and link inflammation and oxidative stress in the adipose tissue (Hummasti and Hotamisligil, 2010). Metformin, which can suppress inflammation, is an oral hypoglycemic agent for type 2 diabetes (Hattori et al., 2006). Resveratrol has strong antihyperglycemia, antioxidant, and anti-inflammatory effects (Chang et al., 2012). Both metformin and resveratrol improve adipose dysfunction by inhibiting inflammation and activating AMPK (Ashabi et al., 2015). In view of the above, in order to study whether metformin and resveratrol can improve adipose dysfunction by regulating ERS-induced NLRP3 inflammasome, Li and colleagues conducted a series of studies. The studies showed that metformin and resveratrol, as well as TUDCA, inhibited ROS generation and ERS induced by HG and improved glucose homeostasis in the adipose tissue of diabetic mice, indicating that ERS was involved in the inhibitory effects of metformin and resveratrol on HG-induced oxidative stress (Li et al., 2016). HG-induced oxidative stress could promote mitochondrial fission by activating dynamin-related protein1 (Drp1), a mitochondrial fission regulator (Smirnova et al., 2001), which was reversed by DPI (a cellular ROS inhibitor) and mitoTEMPO (a mitochondrial ROS inhibitor) in adipocytes or adipose tissue of diabetic mice, indicating that ROS mediated the mitochondrial fission induced by HG-induced oxidative stress. In addition, compound C and AMPK smallinterfering RNA (siRNA) diminished the inhibitory effects of metformin and resveratrol on Drp1, suggesting that AMPK activation mediated the suppression of Drp1 by metformin and resveratrol. Metformin, resveratrol, and TUDCA also suppressed TXNIP/NLRP3 inflammasome by decreasing the induction of TXNIP, NLRP3, and cleaved caspase-1 and the IL-1β secretion in the adipose tissue of diabetic mice, suggesting that ERS mediated the inhibitory effects of metformin and resveratrol on HGinduced TXNIP/NLRP3 inflammasome. Moreover, adipocyte apoptosis was inhibited by metformin, resveratrol, TUDCA, and the anti-IL-1ß antibody. Collectively, it can be inferred that metformin and resveratrol ameliorate adipose dysfunction through inhibiting ERS-induced NLRP3 inflammasome and Drp1-mediated mitochondrial fission, which may be an effective treatment strategy for adipose dysfunction in diabetes (Li et al., 2016). Enhancing AMPK could inhibit ERS (Song et al., 2015); therefore, whether AMPK mediates the effects of metformin and resveratrol remains to be studied. The interplay between Drp1-mediated mitochondrial fission and ERS also remains to be elucidated.

THE ROLE OF THE EFFECTS OF ERS ON NLRP3 INFLAMMASOME IN DIABETIC ENCEPHALOPATHY

Diabetic encephalopathy including depression is one of the major complications of diabetes (Elamoshy et al., 2018; Han et al., 2018). Oxidative stress, ERS, and inflammation are involved in diabetic encephalopathy (Pei and Sun, 2018). Previous studies have shown that NLRP3 inflammasome is activated in the hippocampus of diabetic mice (Zhai et al., 2018) and is induced by ERS in obese rats fed with high fat diet (Cai et al., 2016). However, the link between NLRP3 inflammasome and ERS in diabetic encephalopathy is not clear. Ye et al. found that Gastrodin (Gas), one of the main bioactive components of rhizome of Gastrodia elata, could improve diabetes through reducing body weight, fasting blood glucose, dyslipidemia, and insulin resistance in diabetic mice. Gas also ameliorated depression-like behaviors by improving damaged neurons of diabetic mice. Mechanistic studies revealed that Gas decreased the expression levels of ERS-related proteins, NLRP3 inflammasome, and TXNIP and increased Bcl-2/Bax ratio in the hippocampus of diabetic mice, suggesting that Gas alleviated diabetic depression-like behaviors via inhibiting ERS-induced TXNIP/NLRP3 inflammasome, which need to be further studied (Ye et al., 2018a,b). Similar to the above, recent evidence suggests that hyperglycemia promotes oxidative stress and inflammation in the development of memory and cognitive impairment caused by diabetes (Miguel Roman-Pintos et al., 2016), while the inhibition of HG-induced NLRP3 inflammasome has protective effects on memory and cognitive impairment (Ye et al., 2018a; Zhai et al., 2018). Neferine (NE), suggesting that NLRP3 inflammasome is involved in diabetesinduced memory and cognitive impairment. Neferine (NE) is a unique dibenzylisoquinoline alkaloid from the seed embryo of lotus root and has a long history of application as a folk Chinese herbal medicine in China. NE has been reported to have anti-inflammatory effect (Jung et al., 2010; Asokan et al., 2018). The research of Wu et al. showed that NE ameliorated glucose tolerance, insulin sensitivity, dyslipidemia, memory impairment, and cognitive dysfunction in diabetic mice. Mechanistic studies revealed that NE inhibited oxidative stress and decreased the expression levels of TXNIP, NLRP3 inflammasomes, and ERSrelated proteins in the hippocampus of diabetic mice (Wu et al., 2020). It can be inferred from the above that NE may ameliorate dyslipidemia, memory impairment, and cognitive dysfunction in diabetic mice by inhibiting ERS-induced TXNIP/NLRP3 inflammasome, which provides a treatment strategy for diabetic encephalopathy. The above inference needs to be further studied by using ERS activator and NLRP3 inflammasome inducer.

THE ROLE OF THE EFFECTS OF ERS ON NLRP3 INFLAMMASOME IN DIABETIC RETINAL INJURY

Retinal injury, which is characterized by chronic low-grade inflammation, is the most common complication of diabetes (Kern, 2007; Rossi et al., 2016). It has been reported that intact retina expresses hydroxycarboxylic acid receptor 2 (HCA2), which is activated by β -hydroxybutyrate (BHB). BHB, an endogenous ketone body, is produced by the oxidation of fatty acids in liver mitochondria when carbohydrate supply is insufficient (Martin et al., 2009; Gambhir et al., 2012; Dedkova and Blatter, 2014). The evidence demonstrate that HCA2 receptor has anti-inflammatory properties (Lukasova et al., 2011; Wanders et al., 2013). Trotta et al. (2019) found that the levels of retinal HCA2 receptor protein and apoptosis of retinal cells



were increased in in diabetic mice, while the treatment with BHB activated HCA2 and reduced apoptosis, which alleviated retinal damage. Treatment with BHB inhibited retinal ERS by decreasing the levels of the pPERK, pIRE1, and ATF-6 α and suppressed NLRP3 inflammasome activation by reducing the levels of NLRP3 inflammasome, IL-1 β , and IL-18 in diabetic mice (Trotta et al., 2019). Moreover, NLRP3 inflammasome is stimulated by ERS and involved in retinal damage in diabetes (Rathinam et al., 2012; Chen et al., 2018). Therefore, it can be inferred that the activation of HCA2 receptor by treatment with BHB improves retinal damage through inhibiting ERS-induced NLRP3 inflammasome, which needs ERS and NLRP3 inflammasome activator to further prove. HCA2 activation may offer a new therapeutic target for treating inflammation in diabetic retinal damage.

THE ROLE OF THE EFFECTS OF ERS ON NLRP3 INFLAMMASOME IN DIABETIC NEPHROPATHY

Diabetic nephropathy (DN), which is the main cause of endstage renal disease (Liu L. et al., 2019), is characterized by progressive renal fibrosis, decreased renal function, and irreversible tissue loss (Liu, 2011; Duffield, 2014). Pyroptosis is an inflammatory programmed cell death induced by inflammatory caspase-1 and proinflammatory mediators (Dong et al., 2015; Jorgensen et al., 2016; Man et al., 2017). The activation of NLRP3 inflammasome is closely related to the pyrocytosis, which is involved in diabetes (Wu et al., 2018; Yu et al., 2020). In addition, TXNIP can regulate pyrocytosis through regulating NLRP (Liu X. et al., 2019). Therefore, the inhibition of inflammasome/TXNIP may be a potential treatment for DN. Ke et al. (2020) found that in DN rats, the levels of NLRP3 inflammasome, TXNIP, the pyroptosis-related protein were increased, while silencing TXNIP by lentivirus vector reversed the changes, suggesting that the inhibition of TXNIP/NLRP3 improved renal injury through inhibiting pyroptosis in DN rats. Evidence indicated that ERS regulate pyroptosis through TXNIP/NLRP3 (Chen et al., 2018; Dong et al., 2020). In DN rats, ERS was activated by increasing the levels of CHOP and ATF4 and the phosphorylation of IRE1a. Inhibition of IRE1a could increase miR-200a expression, which downregulated TXNIP levels, indicating that IRE1a mediated the regulatory role of ERS on pyroptosis through TXNIP/NLRP3. The similar results were obtained in HG-induced NRK-52E cell. In sum, the inhibition of IRE1a-mediated ERS can improve DN by downregulating TXNIP/NLRP3 inflammasome-mediated pyroptosis (Ke et al., 2020). In addition to IRE1a, PERK is also an ERS-related protein. Whether the suppression of PERK can downregulate TXNIP/NLRP3 inflammasome-mediated pyroptosis needs to be elucidated. The mechanisms of the relationship between ERS and pyroptosis are complex, which deserves to be further studied.

CONCLUSION

There is an interaction between ERS and NLRP3 inflammasome. Previous studies have shown that ERS can activate NLRP3 inflammasome through ROS/ERK1 pathway, ROS/TXNIP pathway and NF- κ B pathway (**Figure 1**; Ji et al., 2019). In this review, ERS activates NLRP3 inflammasome through the ROS/TXNIP pathway in diabetes. Whether ERS can activate

NLRP3 inflammasome through the ROS/ERK1 pathway or NF- κ B pathway and whether NLRP3 inflammasome can regulate ERS in diabetes deserved to be studied. In addition, whether ERS can regulate NLRP3 inflammasome through other ways remains to be further elucidated.

It has been reported that ERS and NLRP3 inflammasome are both the regulative targets of H_2S (Wang et al., 2020a,b) and involved in diabetes (Ji et al., 2019). Therefore, whether H_2S can improve diabetes through regulating ERS/NLRP3 inflammasome is a subject worth studying. It is believed that the in-depth study of the relationship between ERS and NLRP3 in diabetes will provide a new strategy for the treatment of diabetes.

REFERENCES

- Abderrazak, A., Syrovets, T., Couchie, D., El Hadri, K., Friguet, B., Simmet, T., et al. (2015). NLRP3 inflammasome: from a danger signal sensor to a regulatory node of oxidative stress and inflammatory diseases. *Redox Biol.* 4, 296–307. doi: 10.1016/j.redox.2015.01.008
- Amer Diabet, A. (2014). Diagnosis and classification of diabetes mellitus. *Diabetes Care* 37, S81–S90.
- Ashabi, G., Khalaj, L., Khodagholi, F., Goudarzvand, M., and Sarkaki, A. (2015). Pre-treatment with metformin activates Nrf2 antioxidant pathways and inhibits inflammatory responses through induction of AMPK after transient global cerebral ischemia. *Metab. Brain Dis.* 30, 747–754. doi: 10.1007/s11011-014-9632-2
- Asokan, S. M., Mariappan, R., Muthusamy, S., and Velmurugan, B. K. (2018). Pharmacological benefits of neferine – A comprehensive review. *Life Sci.* 199, 60–70. doi: 10.1016/j.lfs.2018.02.032
- Barbe, F., Douglas, T., and Saleh, M. (2014). Advances in nod-like receptors (NLR) biology. *Cytokine Growth Factor Rev.* 25, 681–697. doi: 10.1016/j.cytogfr.2014. 07.001
- Cai, M., Wang, H., Li, J.-J., Zhang, Y.-L., Xin, L., Li, F., et al. (2016). The signaling mechanisms of hippocampal endoplasmic reticulum stress affecting neuronal plasticity-related protein levels in high fat diet-induced obese rats and the regulation of aerobic exercise. *Brain Behav. Immun.* 57, 347–359. doi: 10.1016/ j.bbi.2016.05.010
- Caldeira, M. V., Salazar, I. L., Curcio, M., Canzoniero, L. M. T., and Duarte, C. B. (2014). Role of the ubiquitin-proteasome system in brain ischemia: friend or foe? *Prog. Neurobiol.* 112, 50–69. doi: 10.1016/j.pneurobio.2013.10.003
- Chang, C.-C., Chang, C.-Y., Huang, J.-P., and Hung, L.-M. (2012). Effect of resveratrol on oxidative and inflammatory stress in liver and spleen of streptozotocin-induced type 1 diabetic rats. *Chin. J. Physiol.* 55, 192–201. doi: 10.4077/cjp.2012.baa012
- Chen, D., Dixon, B. J., Doycheva, D. M., Li, B., Zhang, Y., Hu, Q., et al. (2018). IRE1 alpha inhibition decreased TXNIP/NLRP3 inflammasome activation through miR-17-5p after neonatal hypoxic-ischemic brain injury in rats. *J. Neuroinflamm.* 15:32.
- Chen, H., Zhang, X., Liao, N., Mi, L., Peng, Y., Liu, B., et al. (2018). Enhanced expression of NLRP3 inflammasome-related inflammation in diabetic retinopathy. *Invest. Ophthalmol. Vis. Sci.* 59, 978–985. doi: 10.1167/ iovs.17-22816
- Dedkova, E. N., and Blatter, L. A. (2014). Role of beta-hydroxybutyrate, its polymer poly-beta-hydroxybutyrate and inorganic polyphosphate in mammalian health and disease. *Front. Physiol.* 5:260. doi: 10.3389/fphys.2014.00260
- Defronzo, R. A., Ferrannini, E., Groop, L., Henry, R. R., Herman, W. H., Holst, J. J., et al. (2015). Type 2 diabetes mellitus. *Nat. Rev. Dis. Primers* 1:15019.
- Dong, F., Dong, S., Liang, Y., Wang, K., Qin, Y., and Zhao, X. (2020). miR-20b inhibits the senescence of human umbilical vein endothelial cells through regulating the Wnt/beta-catenin pathway via the TXNIP/NLRP3 axis. *Int. J. Mol. Med.* 45, 847–857.
- Dong, T., Liao, D., Liu, X., and Lei, X. (2015). Using small molecules to dissect non-apoptotic programmed cell death: necroptosis, ferroptosis, and pyroptosis. *Chembiochem* 16, 2557–2561. doi: 10.1002/cbic.201500422

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

This work was supported by grants from the key scientific and technological projects in Henan Province, China (Grant No. 202102310153) and the National Natural Science Foundation of China (Grant No. 81971280).

- Duffield, J. S. (2014). Cellular and molecular mechanisms in kidney fibrosis. J. Clin. Invest. 124, 2299–2306. doi: 10.1172/jci72267
- Elamoshy, R., Bird, Y., Thorpe, L., and Moraros, J. (2018). Risk of depression and suicidality among diabetic patients: a systematic review and meta-analysis. *J. Clin. Med.* 7:445. doi: 10.3390/jcm7110445
- Elliott, E. I., and Sutterwala, F. S. (2015). Initiation and perpetuation of NLRP3 inflammasome activation and assembly. *Immunol. Rev.* 265, 35–52. doi: 10. 1111/imr.12286
- Gambhir, D., Ananth, S., Veeranan-Karmegam, R., Elangovan, S., Hester, S., Jennings, E., et al. (2012). GPR109A as an anti-inflammatory receptor in retinal pigment epithelial cells and its relevance to diabetic retinopathy. *Invest. Ophthalmol. Vis. Sci.* 53, 2208–2217. doi: 10.1167/iovs.11-8447
- Giaccari, A., Sorice, G., and Muscogiuri, G. (2009). Glucose toxicity: the leading actor in the pathogenesis and clinical history of type 2 diabetes - mechanisms and potentials for treatment. *Nutr. Metab. Cardiovasc. Dis.* 19, 365–377. doi: 10.1016/j.numecd.2009.03.018
- Gong, X., Zhang, L., Jiang, R., Ye, M., Yin, X., and Wan, J. (2013). Antiinflammatory effects of mangiferin on sepsis-induced lung injury in mice via up-regulation of heme oxygenase-1. *J. Nutr. Biochem.* 24, 1173–1181. doi: 10.1016/j.jnutbio.2012.09.003
- Han, J., and Kaufman, R. J. (2016). Thematic review series: lipotoxicity: many roads to cell dysfunction and cell death the role of ER stress in lipid metabolism and lipotoxicity. J. Lipid Res. 57, 1329–1338. doi: 10.1194/jlr.r067595
- Han, X., Min, M., Wang, J., Bao, Z., Fan, H., Li, X., et al. (2018). Quantitative profiling of neurotransmitter abnormalities in brain, cerebrospinal fluid, and serum of experimental diabetic encephalopathy male rat. *J. Neurosci. Res.* 96, 138–150. doi: 10.1002/jnr.24098
- Hattori, Y., Suzuki, K., Hattori, S., and Kasai, K. (2006). Metformin inhibits cytokine-induced nuclear factor kappa B activation via AMP-activated protein kinase activation in vascular endothelial cells. *Hypertension* 47, 1183–1188. doi: 10.1161/01.hyp.0000221429.94591.72
- Hetz, C., and Saxena, S. (2017). ER stress and the unfolded protein response in neurodegeneration. *Nat. Rev. Neurol.* 13, 477–491. doi: 10.1038/nrneurol. 2017.99
- Hummasti, S., and Hotamisligil, G. S. (2010). Endoplasmic reticulum stress and inflammation in obesity and diabetes. *Circ. Res.* 107, 579–591. doi: 10.1161/ circresaha.110.225698
- Iannantuoni, F., Diaz-Morales, N., Escribano-Lopez, I., Sola, E., Roldan-Torres, I., Apostolova, N., et al. (2019). Does glycemic control modulate the impairment of NLRP3 inflammasome activation in type 2 diabetes? *Antioxid. Redox Signal.* 30, 232–240. doi: 10.1089/ars.2018.7582
- Inzucchi, S. E., Viscoli, C. M., Young, L. H., and Kernan, W. N. (2019). Diabetes prevention and cardiovascular complications. *Diabetologia* 62, 2161–2162.
- Ji, T., Han, Y., Yang, W., Xu, B., Sun, M., Jiang, S., et al. (2019). Endoplasmic reticulum stress and NLRP3 inflammasome: crosstalk in cardiovascular and metabolic disorders. J. Cell. Physiol. 234, 14773–14782. doi: 10.1002/jcp.28275
- Jo, E.-K., Kim, J. K., Shin, D.-M., and Sasakawa, C. (2016). Molecular mechanisms regulating NLRP3 inflammasome activation. *Cell. Mol. Immunol.* 13, 148–159. doi: 10.1038/cmi.2015.95
- Jorgensen, I., Lopez, J. P., Laufer, S. A., and Miao, E. A. (2016). IL-1 beta, IL-18, and eicosanoids promote neutrophil recruitment to pore-induced intracellular

traps following pyroptosis. Eur. J. Immunol. 46, 2761–2766. doi: 10.1002/eji. 201646647

- Jung, H. A., Jin, S. E., Choi, R. J., Kim, D. H., Kim, Y. S., Ryu, J. H., et al. (2010). Anti-amnesic activity of neferine with antioxidant and anti-inflammatory capacities, as well as inhibition of ChEs and BACE1. *Life Sci.* 87, 420–430. doi: 10.1016/j.lfs.2010.08.005
- Kawasaki, N., Asada, R., Saito, A., Kanemoto, S., and Imaizumi, K. (2012). Obesityinduced endoplasmic reticulum stress causes chronic inflammation in adipose tissue. *Sci. Rep.* 2:799.
- Ke, R., Wang, Y., Hong, S., and Xiao, L. (2020). Endoplasmic reticulum stress related factor IRE1 alpha regulates TXNIP/NLRP3-mediated pyroptosis in diabetic nephropathy. *Exp. Cell Res.* 396:112293. doi: 10.1016/j.yexcr.2020. 112293
- Kern, T. S. (2007). Contributions of inflammatory processes to the development of the early stages of diabetic retinopathy. *Exp. Diabetes Res.* 2007:95103.
- Lan, B., He, Y., Sun, H., Zheng, X., Gao, Y., and Li, N. (2019). The roles of mitochondria-associated membranes in mitochondrial quality control under endoplasmic reticulum stress. *Life Sci.* 231:116587. doi: 10.1016/j.lfs.2019. 116587
- Lerner, A. G., Upton, J.-P., Praveen, P. V. K., Ghosh, R., Nakagawa, Y., Igbaria, A., et al. (2012). IRE1 alpha induces thioredoxin-interacting protein to activate the NLRP3 inflammasome and promote programmed cell death under irremediable ER stress. *Cell Metab.* 16, 250–264. doi: 10.1016/j.cmet.2012.07. 007
- Li, A., Zhang, S., Li, J., Liu, K., Huang, F., and Liu, B. (2016). Metformin and resveratrol inhibit Drp1-mediated mitochondrial fission and prevent ER stressassociated NLRP3 inflammasome activation in the adipose tissue of diabetic mice. *Mol. Cell. Endocrinol.* 434, 36–47. doi: 10.1016/j.mce.2016.06.008
- Liu, L., Wang, Y., Yan, R., Liang, L., Zhou, X., Liu, H., et al. (2019). BMP-7 inhibits renal fibrosis in diabetic nephropathy via miR-21 downregulation. *Life Sci.* 238:116957. doi: 10.1016/j.lfs.2019.116957
- Liu, S., Xin, D., Wang, L., Zhang, T., Bai, X., Li, T., et al. (2017). Therapeutic effects of L-cysteine in newborn mice subjected to hypoxia-ischemia brain injury via the CBS/H2S system: role of oxidative stress and endoplasmic reticulum stress. *Redox Biol.* 13, 528–540. doi: 10.1016/j.redox.2017.06.007
- Liu, X., Zhang, Y.-R., Cai, C., Ni, X.-Q., Zhu, Q., Ren, J.-L., et al. (2019). Taurine alleviates schistosoma-induced liver injury by inhibiting the TXNIP/NLRP3 inflammasome signal pathway and pyroptosis. *Infect. Immun.* 87:e00732-19.
- Liu, Y. (2011). Cellular and molecular mechanisms of renal fibrosis. Nat. Rev. Nephrol. 7, 684–696. doi: 10.1038/nrneph.2011.149
- Lukasova, M., Malaval, C., Gille, A., Kero, J., and Offermanns, S. (2011). Nicotinic acid inhibits progression of atherosclerosis in mice through its receptor GPR109A expressed by immune cells. J. Clin. Invest. 121, 1163–1173. doi: 10.1172/jci41651
- Luo, B., Li, B., Wang, W., Liu, X., Liu, X., Xia, Y., et al. (2014). Rosuvastatin alleviates diabetic cardiomyopathy by inhibiting NLRP3 inflammasome and MAPK pathways in a type 2 diabetes rat model. *Cardiovasc. Drugs Ther.* 28, 33–43. doi: 10.1007/s10557-013-6498-1
- Lv, Y., Gao, X., Luo, Y., Fan, W., Shen, T., Ding, C., et al. (2019). Apigenin ameliorates HFD-induced NAFLD through regulation of the XO/NLRP3 pathways. J. Nutr. Biochem. 71, 110–121. doi: 10.1016/j.jnutbio.2019.05.015
- Man, S. M., Karki, R., and Kanneganti, T.-D. (2017). Molecular mechanisms and functions of pyroptosis, inflammatory caspases and inflammasomes in infectious diseases. *Immunol. Rev.* 277, 61–75. doi: 10.1111/imr.12534
- Marquez, L., Garcia-Bueno, B., Madrigal, J. L. M., and Leza, J. C. (2012). Mangiferin decreases inflammation and oxidative damage in rat brain after stress. *Eur. J. Nutr.* 51, 729–739. doi: 10.1007/s00394-011-0252-x
- Martin, P. M., Ananth, S., Cresci, G., Roon, P., Smith, S., and Ganapathy, V. (2009). Expression and localization of GPR109A (PUMA-G/HM74A) mRNA and protein in mammalian retinal pigment epithelium. *Mol. Vis.* 15, 362–372.
- Miguel Roman-Pintos, L., Villegas-Rivera, G., Daniel Rodriguez-Carrizalez, A., Guillermina Miranda-Diaz, A., and German Cardona-Munoz, E. (2016). Diabetic polyneuropathy in type 2 diabetes mellitus: inflammation, oxidative stress, and mitochondrial function. *J. Diabetes Res.* 2016:3425617.
- Muruganandan, S., Srinivasan, K., Gupta, S., Gupta, P. K., and Lal, J. (2005). Effect of mangiferin on hyperglycemia and atherogenicity in streptozotocin diabetic rats. *J. Ethnopharmacol.* 97, 497–501. doi: 10.1016/j.jep.2004.12.010

- O'connor, W., Harton, J. A., Zhu, X. S., Linhoff, M. W., and Ting, J. P. Y. (2003). Cutting edge: CIAS1/cryopyrin/PYPAF1/NALP3/CATERPILLER 1.1 is an inducible inflammatory mediator with NF-kappa B suppressive properties. *J. Immunol.* 171, 6329–6333. doi: 10.4049/jimmunol.171.12.6329
- Oslowski, C. M., Hara, T., O'sullivan-Murphy, B., Kanekura, K., Lu, S., Hara, M., et al. (2012). Thioredoxin-interacting protein mediates ER stress-induced beta cell death through initiation of the inflammasome. *Cell Metab.* 16, 265–273. doi: 10.1016/j.cmet.2012.07.005
- Pei, B., and Sun, J. (2018). Pinocembrin alleviates cognition deficits by inhibiting inflammation in diabetic mice. J. Neuroimmunol. 314, 42–49. doi: 10.1016/j. jneuroim.2017.11.006
- Prochnicki, T., Mangan, M. S., and Latz, E. (2016). Recent insights into the molecular mechanisms of the NLRP3 inflammasome activation. *F1000Res* 5:F1000.
- Rathinam, V. A. K., Vanaja, S. K., and Fitzgerald, K. A. (2012). Regulation of inflammasome signaling. Nat. Immunol. 13, 333–342. doi: 10.1038/ni.2237
- Rossi, S., Maisto, R., Gesualdo, C., Trotta, M. C., Ferraraccio, F., Kaneva, M. K., et al. (2016). Activation of melanocortin receptors MC1 and MC5 attenuates retinal damage in experimental diabetic retinopathy. *Mediat. Inflamm.* 2016:7368389.
- Sarwar, N., Gao, P., Seshasai, S. R. K., Gobin, R., Kaptoge, S., Di Angelantonio, E., et al. (2010). Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective studies. *Lancet* 375, 2215–2222. doi: 10.1016/s0140-6736(10)60484-9
- Smirnova, E., Griparic, L., Shurland, D. L., and Van Der Bliek, A. M. (2001). Dynamin-related protein Drp1 is required for mitochondrial division in mammalian cells. *Mol. Biol. Cell* 12, 2245–2256. doi: 10.1091/mbc.12.8. 2245
- Sokolova, M., Ranheim, T., Louwe, M. C., Halvorsen, B., Yndestad, A., and Aukrust, P. (2019). NLRP3 inflammasome: a novel player in metabolically induced inflammation-potential influence on the myocardium. J. Cardiovasc. Pharmacol. 74, 276–284. doi: 10.1097/fjc.000000000000704
- Song, J., Li, J., Hou, F., Wang, X., and Liu, B. (2015). Mangiferin inhibits endoplasmic reticulum stress-associated thioredoxin-interacting protein/NLRP3 inflammasome activation with regulation of AMPK in endothelial cells. *Metab. Clin. Exp.* 64, 428–437. doi: 10.1016/j.metabol.2014. 11.008
- Trotta, M. C., Maisto, R., Guida, F., Boccella, S., Luongo, L., Balta, C., et al. (2019). The activation of retinal HCA2 receptors by systemic beta-hydroxybutyrate inhibits diabetic retinal damage through reduction of endoplasmic reticulum stress and the NLRP3 inflammasome. *PLoS One* 14:e0211005. doi: 10.1371/ journal.pone.0211005
- Wanders, D., Graff, E. C., White, B. D., and Judd, R. L. (2013). Niacin increases adiponectin and decreases adipose tissue inflammation in high fat diet-fed mice. *PLoS One* 8:e71285. doi: 10.1371/journal.pone.0071285
- Wang, F., Yan, J., Niu, Y., Li, Y., Lin, H., Liu, X., et al. (2014). Mangiferin and its aglycone, norathyriol, improve glucose metabolism by activation of AMPactivated protein kinase. *Pharm. Biol.* 52, 68–73. doi: 10.3109/13880209.2013. 814691
- Wang, H., Shi, X., Qiu, M., Lv, S., and Liu, H. (2020a). Hydrogen sulfide plays an important protective role through influencing endoplasmic reticulum stress in diseases. *Int. J. Biol. Sci.* 16, 264–271. doi: 10.7150/ijbs.38143
- Wang, H., Shi, X., Qiu, M., Lv, S., Zheng, H., Niu, B., et al. (2020b). Hydrogen sulfide plays an important role by influencing NLRP3 inflammasome. *Int. J. Biol. Sci.* 16, 2752–2760. doi: 10.7150/ijbs.47595
- Whiting, D. R., Guariguata, L., Weil, C., and Shaw, J. (2011). IDF diabetes atlas: global estimates of the prevalence of diabetes for 2011 and 2030. *Diabetes Res. Clin. Pract.* 94, 311–321. doi: 10.1016/j.diabres.2011.10.029
- Wu, M., Han, W., Song, S., Du, Y., Liu, C., Chen, N., et al. (2018). NLRP3 deficiency ameliorates renal inflammation and fibrosis in diabetic mice. *Mol. Cell. Endocrinol.* 478, 115–125. doi: 10.1016/j.mce.2018. 08.002
- Wu, X.-L., Deng, M.-Z., Gao, Z.-J., Dang, Y.-Y., Li, Y.-C., and Li, C.-W. (2020). Neferine alleviates memory and cognitive dysfunction in diabetic mice through modulation of the NLRP3 inflammasome pathway and alleviation of endoplasmic-reticulum stress. *Int. Immunopharmacol.* 84:106559. doi: 10.1016/ j.intimp.2020.106559

- Ye, T., Meng, X., Wang, R., Zhang, C., He, S., Sun, G., et al. (2018a). Gastrodin alleviates cognitive dysfunction and depressive-like behaviors by inhibiting ER stress and NLRP3 inflammasome activation in db/db mice. *Int. J. Mol. Sci.* 19:3977. doi: 10.3390/ijms19123977
- Ye, T., Meng, X., Zhai, Y., Xie, W., Wang, R., Sun, G., et al. (2018b). Gastrodin ameliorates cognitive dysfunction in diabetes rat model via the suppression of endoplasmic reticulum stress and NLRP3 inflammasome activation. *Front. Pharmacol.* 9:1346. doi: 10.3389/fphar.2018.01346
- Yu, Z.-W., Zhang, J., Li, X., Wang, Y., Fu, Y.-H., and Gao, X.-Y. (2020). A new research hot spot: the role of NLRP3 inflammasome activation, a key step in pyroptosis, in diabetes and diabetic complications. *Life Sci.* 240:117138. doi: 10.1016/j.lfs.2019.117138
- Zeyda, M., and Stulnig, T. M. (2007). Adipose tissue macrophages. *Immunol. Lett.* 112, 61–67. doi: 10.1016/j.imlet.2007.07.003
- Zhai, Y., Meng, X., Luo, Y., Wu, Y., Ye, T., Zhou, P., et al. (2018). Notoginsenoside R1 ameliorates diabetic encephalopathy by activating the Nrf2 pathway and

inhibiting NLRP3 inflammasome activation. Oncotarget 9, 9344-9363. doi: 10.18632/oncotarget.24295

Zheng, Y., Ley, S. H., and Hu, F. B. (2018). Global aetiology and epidemiology of type 2 diabetes mellitus and its complications. *Nat. Rev. Endocrinol.* 14, 88–98. doi: 10.1038/nrendo.2017.151

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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