

# Complete Genome Sequence of *Enterobacter cloacae* UW5, a Rhizobacterium Capable of High Levels of Indole-3-Acetic Acid Production

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**We report the complete genome sequence of *Enterobacter cloacae* UW5, an indole-3-acetic acid-producing rhizobacterium originally isolated from the rhizosphere of grass. The 4.9-Mbp genome has a G+C content of 54% and contains 4,496 protein-coding sequences.**

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Members of the *Enterobacter* genus are Gram-negative facultative anaerobic bacteria commonly found inhabiting the rhizosphere of plants, and they may also be members of the intestinal microflora of humans. *Enterobacter cloacae* strains are often endophytes capable of biocontrol of plant infectious diseases or production of plant growth-stimulating compounds (1). *E. cloacae* UW5 was originally isolated from the rhizosphere of reeds in Waterloo, Ontario, Canada (2, 3), and has been shown to produce high levels of indole-3-acetic acid (IAA) (4), a signaling molecule that acts as a bacterial stress response regulator, antimicrobial metabolite, and plant growth stimulant (5). Regulation of IAA by the transcription factor TyrR has been characterized in this strain (4, 6). The genome of *E. cloacae* UW5 was sequenced to facilitate further insight into the TyrR regulon and the function of IAA in bacterial physiology and bacterial-plant interactions.

Genomic DNA was extracted from an overnight culture grown at 30°C, using the Wizard Genomic DNA purification kit (Promega). A 100-bp paired-end TruSeq library was prepared with an average fragment size of 444 bp and sequenced in a single Illumina HiSeq 2500 lane (McGill University and Génome Québec Innovation Center, Montreal, Canada). A total of 251,715,967 reads were generated, totaling over 51 billion bases. Seqtk (<https://github.com/lh3/seqtk>) was used to randomly extract a subset of 10 million forward and reverse paired reads, which were then trimmed using Trimmomatic (7) to remove adapters and low-quality sequences. The resulting reads were assessed for sequence quality using FastQC version 0.11.2 (8) before assembly. Genome assembly was performed using a combination of *de novo* assembly and reference-guided read mapping to a closely related reference genome. An optimal *k*-mer of 71 was chosen using VelvetOptimiser, which was used for *de novo* assembly with Velvet version 1.2.10 (9). A total of 83 contigs were generated, with an average of 330-fold coverage and an  $N_{50}$  length of 2,836,112 bp. The same read data were then mapped to the *E. cloacae* EcWSU1 reference genome (NC\_016514) (10) using CLC Genomics Workbench version 8.0.1 (Qiagen) at both low (80% identity and 50% length fraction) and high (90% identity

and 90% length fraction) specificity, and the consensus sequences extracted to produce the reference-guided contigs. Contigs from both *de novo* and reference-guided assemblies were then joined using the CLC Microbial Genome Finishing module to merge overlapping sequences into a single contig. Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) ([http://www.ncbi.nlm.nih.gov/genome/annotation\\_prok](http://www.ncbi.nlm.nih.gov/genome/annotation_prok)).

The *E. cloacae* UW5 genome consists of a single chromosome that is 4,904,981 bp in length with a G+C content of 54.46%. The genome contains 4,496 genes assigned with protein-encoding function, 77 tRNA genes, and 25 rRNA genes organized into eight rRNA operons.

**Nucleotide sequence accession number.** The complete *E. cloacae* UW5 genome sequence has been deposited in GenBank under the accession number CP011798. The version reported in this paper is the first version.

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## REFERENCES

1. Nguyen MT, Ranamukhaarachchi SL. 2010. Soil-borne antagonists for biological control of bacterial wilt disease caused by *Ralstonia solanacearum* in tomato and pepper. *J Plant Pathol* 92:395–406.
2. Glick BR, Karaturovic DM, Newell PC. 1995. A novel procedure for rapid isolation of plant growth promoting pseudomonads. *Can J Microbiol* 41:533–536. <http://dx.doi.org/10.1139/m95-070>.
3. Shah S, Li J, Moffatt BM, Glick BR. 1997. ACC deaminase genes from plant growth-promoting bacteria, p 320–324. In Ogoshi A, Kobayashi K, Homma Y, Kodama F, Kondo N, Akino S (ed), *Plant growth promoting bacteria: present status and future prospects*. Organization for Economic Cooperation and Development, Paris, France.
4. Ryu RJ, Patten CL. 2008. Aromatic amino acid-dependent expression of indole-3-pyruvate decarboxylase is regulated by TyrR in *Enterobacter cloacae* UW5. *J Bacteriol* 190:7200–7208. <http://dx.doi.org/10.1128/JB.00804-08>.

5. Patten CL, Blakney AJ, Coulson TJ. 2013. Activity, distribution and function of indole-3-acetic acid biosynthetic pathways in bacteria. *Crit Rev Microbiol* 39:395–415. <http://dx.doi.org/10.3109/1040841X.2012.716819>.
6. Coulson TJ, Patten CL. 2015. The TyrR transcription factor regulates the divergent *akr-ipdC* operons of *Enterobacter cloacae* UW5. *PLoS One* 10: e0121241. <http://dx.doi.org/10.1371/journal.pone.0121241>.
7. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <http://dx.doi.org/10.1093/bioinformatics/btu170>.
8. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
9. Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
10. Humann JL, Wildung M, Cheng CH, Lee T, Stewart JE, Drew JC, Triplett EW, Main D, Schroeder BK. 2011. Complete genome of the onion pathogen *Enterobacter cloacae* EcWSU1. *Stand Genomic Sci* 5:279–286.